

The *Arabidopsis* IDD14, IDD15, and IDD16 Cooperatively Regulate Lateral Organ Morphogenesis and Gravitropism by Promoting Auxin Biosynthesis and Transport

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Abstract

The plant hormone auxin plays a critical role in regulating various aspects of plant growth and development, and the spatial accumulation of auxin within organs, which is primarily attributable to local auxin biosynthesis and polar transport, is largely responsible for lateral organ morphogenesis and the establishment of plant architecture. Here, we show that three *Arabidopsis* INDETERMINATE DOMAIN (IDD) transcription factors, IDD14, IDD15, and IDD16, cooperatively regulate auxin biosynthesis and transport and thus aerial organ morphogenesis and gravitropic responses. Gain-of-function of each *IDD* gene in *Arabidopsis* results in small and transversally down-curved leaves, whereas loss-of-function of these *IDD* genes causes pleiotropic phenotypes in aerial organs and defects in gravitropic responses, including altered leaf shape, flower development, fertility, and plant architecture. Further analyses indicate that these *IDD* genes regulate spatial auxin accumulation by directly targeting *YUCCA5* (*YUC5*), *TRYPTOPHAN AMINOTRANSFERASE of ARABIDOPSIS1* (*TAA1*), and *PIN-FORMED1* (*PIN1*) to promote auxin biosynthesis and transport. Moreover, mutation or ectopic expression of *YUC* suppresses the organ morphogenic phenotype and partially restores the gravitropic responses in gain- or loss-of-function *idd* mutants, respectively. Taken together, our results reveal that a subfamily of IDD transcription factors plays a critical role in the regulation of spatial auxin accumulation, thereby controlling organ morphogenesis and gravitropic responses in plants.

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Introduction

Auxin is a key plant hormone that plays critical roles in the regulation of plant growth and development. A combination of physiological, genetic, biochemical, and molecular studies has greatly enriched our understanding of auxin biosynthesis, transport, and signal transduction [1–3]. Increasing evidence indicates that auxin is essential for nearly all developmental processes, including gametogenesis, embryogenesis, lateral organ formation and patterning, branching, and tropic responses [4,5]. It is generally believed that most auxin-mediated developmental events are highly dependent on the differential accumulation of auxin within plant organs (auxin gradients), which are mainly attributable to both local auxin biosynthesis and the intercellular polar transport of auxin [1,5].

Direct evidence that local auxin biosynthesis is involved in the regulation of plant organogenesis comes from studies of several genes in the auxin biosynthetic pathway of *Arabidopsis*, including *YUCCA* (*YUC*) and *TRYPTOPHAN AMINOTRANSFERASE of ARABIDOPSIS* (*TAA*). *YUCs* encode the flavin monooxygenases that catalyze a key step in converting tryptophan into IAA, a main auxin in plants [6]. Overexpression of *YUC* genes in *Arabidopsis*

substantially elevates the endogenous IAA level and causes distinct phenotypes such as epinastic cotyledons, elongated hypocotyls, and narrow and curly leaves [6–8]. Although a *yuc* single mutant in *Arabidopsis* does not show an obvious phenotype, the mutation of multiple *YUC* genes leads to a diversity of auxin-related phenotypes, including reduced apical dominance, crinkled leaves, simple venation, and abnormal flower development, demonstrating that *YUC*-modulated local auxin biosynthesis is critical for plant morphogenesis and architecture formation [9,10]. Consistently, such developmental defects in *yuc* mutants can be rescued by local expression of *iaaM*, a bacterial auxin biosynthetic gene, but not by the application of exogenous auxin [9]. The expressions of *YUC* genes are overlapping and spatiotemporally regulated in various organs [9,10], suggesting that *YUCs* function redundantly and cooperatively in different organs. Further studies demonstrate that *TAA1* and its homologs function in auxin biosynthesis in response to environmental and developmental signals in *Arabidopsis*. The *taa1* plant has a decreased level of endogenous IAA and displays defects in shade avoidance and root-specific ethylene sensitivity, and the simultaneous mutation of *TAA1* and its close homologs (*TAR1* and *TAR2*) results in phenotypes that are obviously auxin-related, such as reduced gravitropic response of

Author Summary

Auxin is a key plant hormone and the spatial accumulation of auxin is essential for lateral organ morphogenesis and gravitropic responses in higher plants. However, the various mechanisms through which spatial auxin accumulation is regulated remain to be fully elucidated. Here, we identify a gain-of-function mutant of *Arabidopsis* *IDD14* that exhibits small and transversally down-curved leaves. Further characterization of both gain- and loss-of-function mutants in *IDD14* and its close homologs, *IDD15* and *IDD16*, reveals that these three IDD transcription factors function redundantly and cooperatively in the regulation of multiple aspects of lateral organ morphogenesis and gravitropic responses. We further demonstrate that these IDD transcription factors influence the spatial accumulation of auxin by directly targeting auxin biosynthetic and transport genes to activate their expression. These findings identify a subfamily of IDD transcription factors that coordinates spatial auxin gradients and thus directs lateral organ morphogenesis and gravitropic responses in plants.

roots, shortened hypocotyls, and *monopteros*-like seedlings with a single cotyledon [11–13]. Recently, YUCs and TAAs have been shown to function in a key two-step auxin biosynthetic pathway [14–16]. Therefore, the regulation of YUCs and TAAs may represent an important mechanism to alter auxin gradients and thus modify plant morphogenesis and responses to environmental cues. Indeed, several transcription factors have been shown to participate in the regulation of *YUC* expression. *STYLISH1* (*STY1*), one of the *SHORT INTERNODES* (*SHI*) transcription factors and *NGATHA3* (*NGA3*), a member of the B3 transcription factor family, are both proposed to act cooperatively to direct style development, partially through the activation of *YUC2*- and *YUC4*-mediated auxin biosynthesis in the apex of the gynoecium [17,18]. *LEAFY COTYLEDON2* (*LEC2*), a B3 domain transcription factor acting as a central regulator of embryogenesis, has been found to be involved in modulating *de novo* auxin biosynthesis via the activation of *YUC2* and *YUC4* [19]. Recent studies have also revealed that *PIF4*-activated *YUC8* expression is required for hypocotyl growth under high temperature conditions [20], and a putative transcription factor *SPOROCTELESS/NOZZLE* (*SPL/NZZ*) has been found to be involved in the regulation of lateral organ morphogenesis by repressing transcription of *YUC2* and *YUC6* [21].

In addition to local auxin biosynthesis, polar auxin transport also contributes greatly to auxin redistribution. The polar auxin transport is mediated by *AUX1/LIKE AUX1* (*AUX1/LAX*), *PIN-FORMED* (*PIN*), and ATP-binding cassette class B (*ABCB*) transporters [2,22–24]. Mutation in *AUX1/LAX*, which encode auxin influx carriers, influences lateral root formation, root hair development, phyllotaxy, and tropic responses [22,24–28]. Similarly, single or multiple mutations in auxin efflux carrier genes, *PINs* and *ABCBs*, lead to severe defects in multiple developmental processes including embryogenesis, lateral organ formation and patterning, vascular development, and tropic responses [29–33], demonstrating that polar auxin transport is also essential for plant morphogenesis and tropic responses.

The *INDETERMINATE DOMAIN* (*IDD*) transcription factors belong to a plant-specific transcription factor family that contains a conserved *ID* domain with four zinc finger motifs [34,35]. The founding member of this family, maize *INDETERMINATE1* (*ID1*), is a key regulator of flowering transition [36]. The *Arabidopsis* genome contains 16 *IDD* members, and the

characterization of several *IDD* members in *Arabidopsis* has indicated that *IDD* genes are involved in regulation of multiple developmental processes. For example, *IDD8/NUTCRACKER* (*NUC*) is involved in the regulation of flowering time through modulation of sugar metabolism [37], and *IDD14* regulates starch metabolism in response to cold stimulus [38]. *IDD15/SHOOT GRAVITROPISM 5* (*SGR5*) is involved in the gravitropic response of inflorescence stems, possibly through alteration of gravity sensing [39–41]. *IDD8/NUC*, *IDD3/MAGPIE* (*MAG*), and *IDD10/JACKDAW* (*JKD*) regulate root development and patterning [42,43], while *IDD1/ENHYDROUS* (*EN1*) participates in seed maturation and germination [44]. Recent studies have demonstrated that a rice *ID1* homolog is required for flowering induction [45–47], and that the rice *Loose Plant Architecture1* (*LPA1*), an ortholog of *Arabidopsis* *IDD15*, influences shoot gravitropism and architecture [48].

Here, we identified a gain-of-function mutant of *Arabidopsis* *IDD14*, *idd14-1D*, which exhibited small and transversally down-curved leaves. Further characterization of loss- and gain-of-function mutants of *IDD14* and its close homologs, *IDD15* and *IDD16*, revealed that these three *IDD* genes redundantly but differentially regulate multiple aspects of organ development and gravitropic responses. Moreover, we provide evidence that these *IDD* transcription factors can directly target auxin biosynthetic and transport genes and alter auxin accumulation in multiple organs. These results reveal that *IDD*-mediated auxin biosynthesis and transport are critical for lateral organ morphogenesis and gravitropic responses in plants.

Results

cuf1-D/idd14-1D Displays Pleiotropic Leaf Phenotypes

To gain insight into how lateral organ morphogenesis is controlled in plants, we screened for mutants with altered aerial organ morphology in a transgenic *Arabidopsis* population harboring a T-DNA activation-tagging plasmid (*pSKI015*) [49]. A semi-dominant mutant was identified by its smaller and dramatically down-curling leaf phenotype, and thus designated as *curlyfolia1-D* (*cuf1-D*) and subsequently as *idd14-1D* (see below) (Figure 1A). To examine the genetic nature of *cuf1-D*, we backcrossed *cuf1-D* with wild type (*WT*) plants. All *F1* plants exhibited an intermediate phenotype as *cuf1-D/+*, and *F2* plants displayed a phenotypic segregation of *WT:cuf1-D/+:cuf1-D* as 1:2:1 (82:173:87, $P=0.89$, χ^2 -test), in which all *WT* plants were *BASTA* sensitive. This observation suggests that *cuf1-D* results from a semi-dominant mutation of a single gene that is likely to co-segregate with a T-DNA insertion event. The most striking phenotype in *cuf1-D* was the leaf size and shape. Detailed quantification showed that the average areas of fully-expanded leaf blades in heterozygous and homozygous *cuf1-D* were only about 70% and 47% of that in *WT*, respectively (Figure 1B). The leaves of *cuf1-D* were also dramatically curled downward in a transverse direction. The leaf transverse curvature (*TC*) index in *WT* was about 0.08, whereas the leaf *TC* index in heterozygous and homozygous *cuf1-D* reached about 0.29 and 0.36, respectively (Figure 1C). Moreover, the lamina of a mature *WT* rosette leaf displayed an elliptical shape with a leaf length/width index at about 1.5, while the heterozygous and homozygous *cuf1-D* leaves were comparatively narrow and their leaf indices were about 1.9 (Figure 1D). In addition to the morphological changes in leaves, *cuf1-D* was also late flowering and dwarfed (Figure 1A, Table S1). These observations demonstrate that the dominant mutation in *cuf1-D* has pleiotropic effects on lateral organ development and plant architecture.

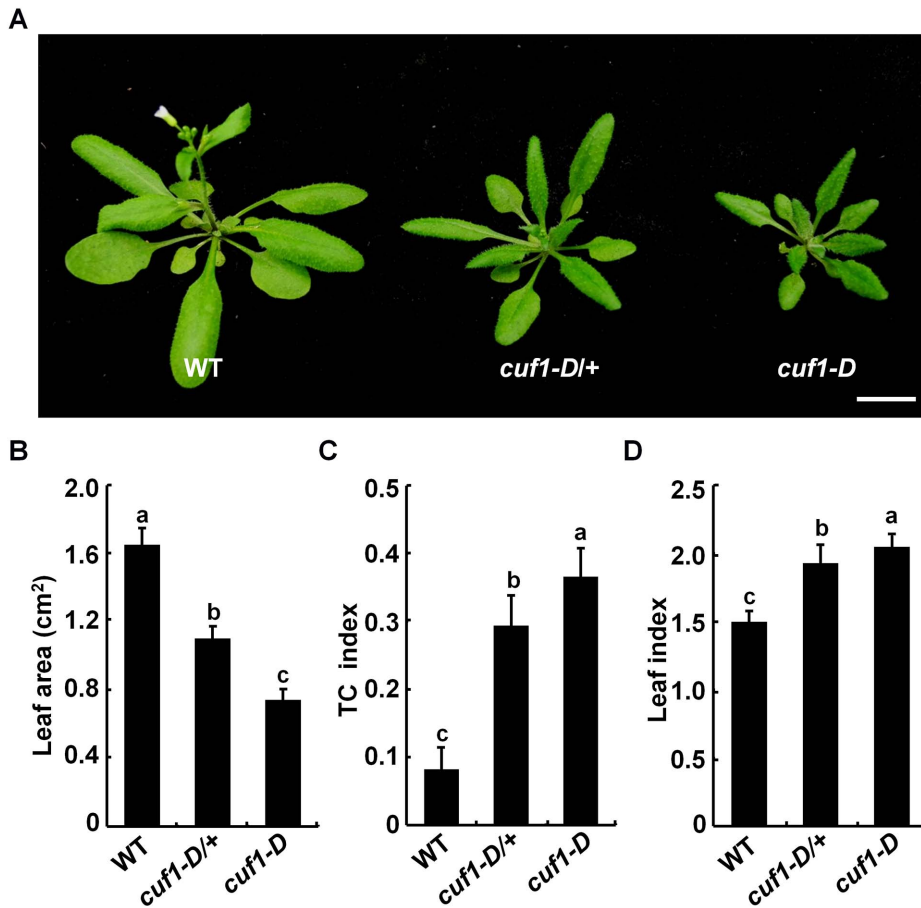


Figure 1. Phenotype of *cuf1-D* plants. (A) Morphology of 4-week-old wild-type (WT), *cuf1-D/+*, and *cuf1-D* plants. The scale bar represents 1 cm. (B) Blade areas of WT, *cuf1-D/+*, and *cuf1-D* leaves. (C) Transverse curvature (TC) index of WT, *cuf1-D/+*, and *cuf1-D* leaves. (D) Leaf index (ratio of length to width) in WT, *cuf1-D/+*, and *cuf1-D* plants. At least 10 sixth leaves from each genotype were used for determination of the leaf area, TC index, and leaf index as described in the methods section, respectively. Data are shown as mean values \pm one SD, and the letters (a to c) indicate the significance ($P < 0.05$) among the genotypes according to one-way ANOVA test (SPSS 13.0, Chicago, IL, USA). doi:10.1371/journal.pgen.1003759.g001

CUF1 Encodes the INDETERMINATE DOMAIN 14 Transcription Factor

Since *cuf1-D* is a single gene mutation that is likely to cosegregate with a T-DNA insertion event, we amplified the genomic DNA adjacent to the left border of the T-DNA by thermal asymmetric interlaced PCR (TAIL-PCR). The sequencing analysis indicated that a T-DNA was inserted in the intergenic region between At1g68120 and At1g68130 (Figure 2A). Genotyping analysis showed that the T-DNA insertion cosegregated with the leaf phenotype in heterozygous and homozygous *cuf1-D* mutants (Figure 2B), suggesting that *cuf1-D* was caused by the T-DNA insertion event. Because *cuf1-D* was a dominant mutant, we monitored the transcripts of genes flanking the T-DNA insertion using semi-quantitative reverse transcription PCR (RT-PCR) analysis. Compared to those in the WT, the transcripts of At1g68130 (*IDD14*) were dramatically elevated, while At1g68140 mRNA levels were slightly decreased in *cuf1-D* (Figure 2C). To determine whether the elevated levels of *IDD14* transcripts are responsible for the *cuf1-D* phenotype, we introduced a *p35S::anti-IDD14* construct into *cuf1-D* and a *p35S::IDD14* construct into WT plants, respectively. Transgenic plants overexpressing *IDD14* fully recapitulated the phenotype of *cuf1-D*, while the expression level of At1g68140 in these plants was comparable with that in WT plants (Figure 2D, S1B). Moreover, introduction of *p35S::anti-IDD14* into

cuf1-D restored the *cuf1-D* to WT morphology (Figure 2D). These results demonstrate that *cuf1-D* results from the ectopic expression of *IDD14*, and accordingly, the *cuf1-D* was thus re-designated as *idd14-1D*.

In *Arabidopsis*, the IDD14 transcription factor is phylogenetically sub-grouped with two close homologs: IDD15 (SGR5) and IDD16, to form a small subfamily that is distinct from the other IDD family members (Figure S1A) [34,39]. IDD14, IDD15 and IDD16 share 52%–62% amino acid identity, and their ID domains are highly conserved with 89%–95% amino acid identity (Figure 2E). To investigate the possible redundancy of *IDD15* and *IDD16* with *IDD14*, we generated transgenic *Arabidopsis* plants overexpressing *IDD15* or *IDD16*, respectively. The ectopic expression of either *IDD15* or *IDD16* resulted in a similar leaf phenotype as observed in *idd14-1D* (Figure 2F, S1B), suggesting that the three IDD members may have redundant function during plant development.

The Differential and Overlapping Expression of *IDD14*, *IDD15*, and *IDD16* in Multiple Organs

To explore the functions of this *IDD* subfamily, we examined the tissue-specific expression patterns of these genes in multiple organs of transgenic plants harboring a *pIDD::GUS* (β -Glucuronidase) construct. As shown in Figure 3A–3C, *IDD14* was mainly

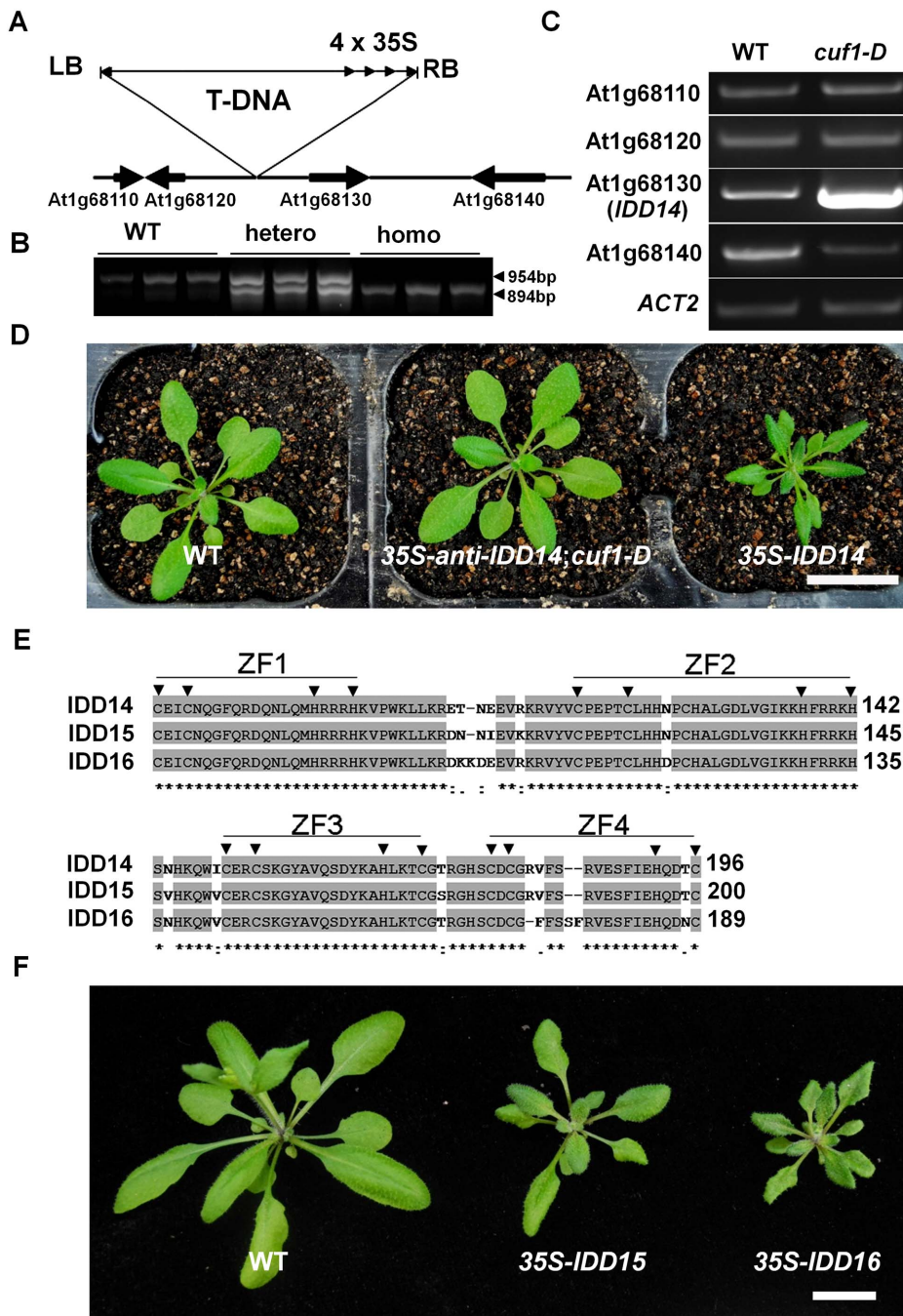


Figure 2. Molecular characterization of CUF1/IDD14. (A) Scheme of the genomic region flanking T-DNA insertion in *cuf1-D*. Genes are shown as thick arrows and intergenic regions are shown as lines. The orientation of T-DNA left border (LB), right border (RB), and the four CaMV 35S enhancers (4×35S) are indicated. (B) Linkage analysis of the T-DNA and *cuf1-D* phenotype. The 954-bp genomic fragments in WT and *cuf1-D*^{+/+} plants were amplified with primers in the genomic region flanking the T-DNA insertion site, and the 894-bp fragments in *cuf1-D*^{-/-} and *cuf1-D* were amplified with an LB primer and a downstream genomic primer. (C) Expression analysis of the genes flanking the T-DNA in WT and *cuf1-D* plants. *ACTIN2* (*ACT2*) was used as an internal control. Note that transcripts of *At1g68130* (*IDD14*) are highly elevated in *cuf1-D*. (D) Morphology of 25-days-old WT, *cuf1-D* carrying *p35S::anti-IDD14*, and *p35S::IDD14* plants. The scale bar represents 2 cm. (E) Alignment of the ID domains in *IDD14*, *IDD15*, and *IDD16*. ZF1–ZF4 represents the four C2H2-type zinc finger motifs. The arrowheads indicate the conserved cysteine and histidine residues. (F) Morphology of 4-week-old transgenic plants overexpressing *IDD15* or *IDD16*. The scale bar represents 1 cm. doi:10.1371/journal.pgen.1003759.g002

expressed in cotyledons and the vasculature of rosette leaves, and a weak level of expression was observed in hypocotyls and floral organs. However, the GUS signal was undetectable in roots and inflorescence stems. *IDD15* was highly expressed in petioles, hypocotyls, roots, floral organs, and especially in inflorescence

stems. In inflorescence stems, GUS staining was mainly present in the cortex, endodermis and vasculature tissues (Figure 3A–3C). *IDD16* was highly expressed in leaves, hypocotyls, roots, vasculature of cotyledons, floral organs, and in the endodermis and vasculature of inflorescence stems (Figure 3A–3C). RNA *in situ*

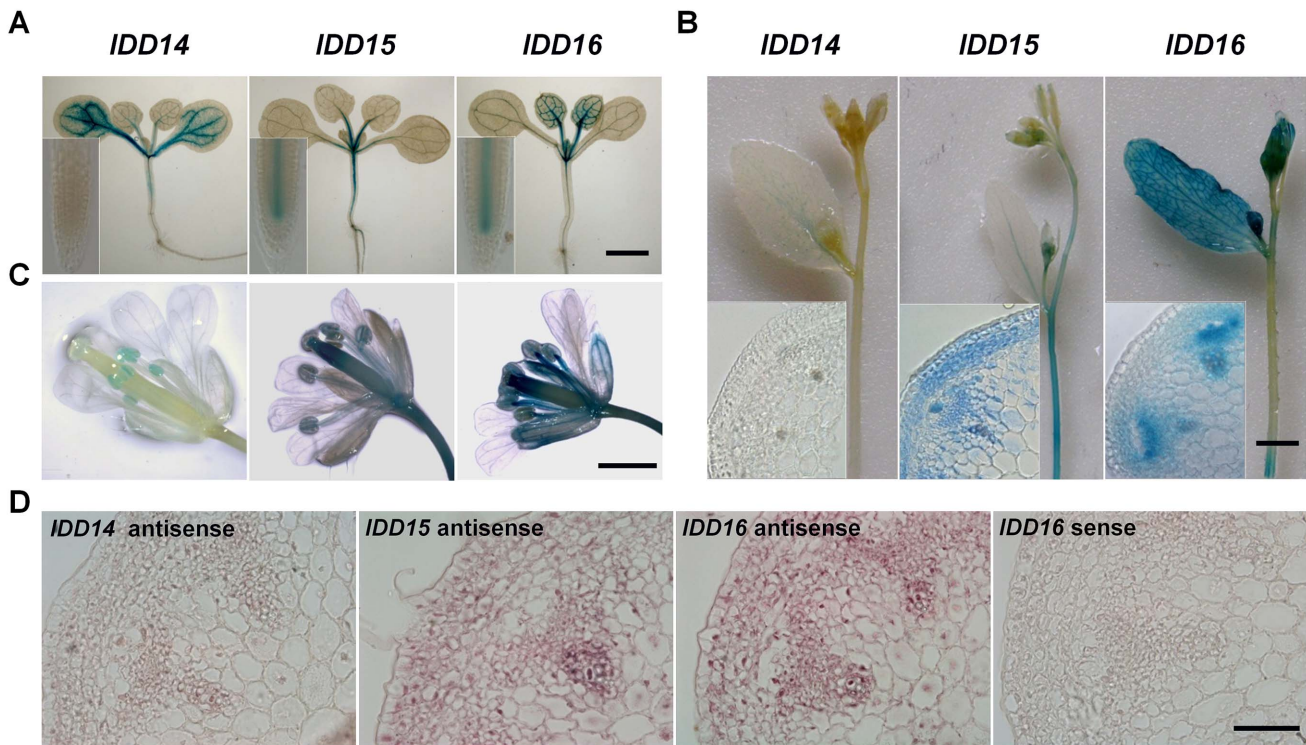


Figure 3. Differential and overlapping expression of *IDD14*, *IDD15*, and *IDD16*. (A) Expression of *IDD14*, *IDD15*, and *IDD16* in seedlings assayed by GUS staining. The 12-day-old transgenic seedlings carrying *pIDD14::GUS*, *pIDD15::GUS*, or *pIDD16::GUS* constructs were subjected to GUS staining assays. Insets show the primary roots. The scale bar represents 1 mm. (B) GUS staining of the florescence stems of *pIDD14::GUS*, *pIDD15::GUS*, and *pIDD16::GUS* transgenic plants. Insets show the transverse sections of inflorescence stems. The scale bar represents 2 mm. (C) GUS staining of the floral organs of *pIDD14::GUS*, *pIDD15::GUS*, and *pIDD16::GUS* transgenic plants. The scale bar represents 1 mm. (D) Expression of *IDD14*, *IDD15*, and *IDD16* in inflorescence stems assayed by RNA *in situ* hybridization. A section hybridized with an *IDD16* sense probe is shown as a control. The scale bar represents 50 μ m.

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hybridization assayed in the inflorescence stems validated the expressions of the three *IDD* genes detected by the GUS reporter (Figure 3D). Additionally, consistent with the previous finding that *IDD* members act as transcription factors [38,39], an *IDD14*-GFP fusion protein in transgenic plants carrying *p35S::IDD14-GFP*, which recapitulated the phenotype of *idd14-1D*, was found to be localized in nuclei (Figure S2A, S2B).

IDD14, *IDD15*, and *IDD16* Cooperatively Regulate Lateral Organ Morphogenesis and Gravitropic Responses

To gain further insight into the functions of this *IDD* subfamily, we obtained the T-DNA insertion mutant *idd14-1* (CS367164) and *idd15-5* (Salk_087765) from the Arabidopsis Biological Resource Center (ABRC), in which the T-DNA is inserted in an exon of *IDD14* or *IDD15*. Semi-quantitative RT-PCR analyses indicated that the transcripts of *IDD14* or *IDD15* were undetectable in *idd14-1* or *idd15-5*, respectively (Figure 4A, 4B). As no *idd16* mutant is publically available, we generated *IDD16-RNAi* transgenic plants with an *IDD16*-specific cDNA fragment (Figure 4A). Semi-quantitative RT-PCR analysis indicated that *IDD16* transcripts were dramatically reduced in transgenic *IDD16-RNAi* plants (Figure 4B). We then generated double and triple mutants of the three *IDD* genes, which allowed us to closely examine their differential and redundant functions during plant growth and development.

We first examined the aerial organ morphogenesis in these *idd* mutants. As shown in Figure 4C and 4D, none of the *idd* single mutants exhibited any obvious organ phenotype. However, the

leaves of *idd14-1 IDD16-RNAi* and *idd* triple mutants were not only downward-curved in the longitudinal direction, but were also more rotund when compared to WT leaves. Furthermore, *idd14-1 IDD16-RNAi* and the *idd* triple mutant had enlarged floral organs and infertile siliques. Careful examination showed that the infertile siliques resulted from the asynchronous elongation of stamen filaments and styles, and thus had poorly pollinated stigmas (Figure 4C, 4E). Manual pollination of styles in these mutants resulted in the development of normal siliques. In contrast to these dramatic phenotypes, the *idd15-5 IDD16-RNAi* plants had only slightly curled leaves (Figure 4C, 4D). These observations imply that *IDD14* and *IDD16* have redundant roles in directing leaf and floral organ morphogenesis.

Consistent with the previous finding that *idd15* displays increased angles between inflorescence stems and siliques [40], we noticed that the orientation angles of both branches and siliques were obviously increased in *idd15-5* (Figure 4C). This phenotype was further enhanced in the *idd15-5 IDD16-RNAi* and *idd* triple mutants, but not in *idd14-1 idd15-5* plants (Figure 4C, 5A), indicating that *IDD15* and *IDD16* act cooperatively to control siliques and branch orientation. As *idd15* has a reduced gravitropic response in inflorescence stems [39–41], it is likely that the altered orientation of branches and siliques is related to the gravitropism defect in the *idd* mutants. To test this, we investigated the gravitropic responses in gain- and loss-of-function of *idd* plants. As expected, *idd15-5* inflorescence stems exhibited an obviously reduced gravitropic response, and this phenotype was greatly enhanced in *idd15-5 IDD16-RNAi* and the *idd* triple mutant

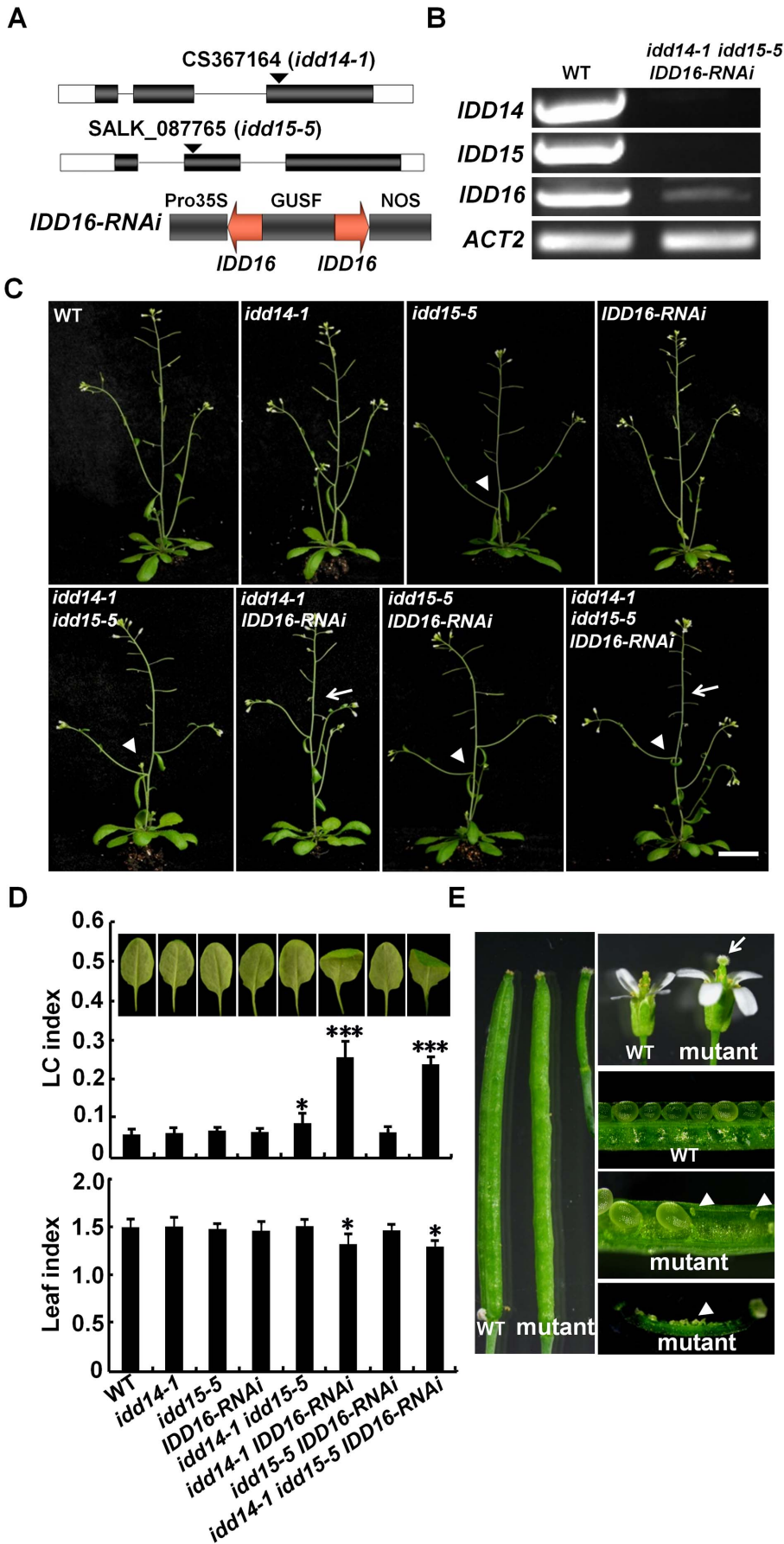


Figure 4. Pleiotropic organ phenotypes in loss-of-function *idd* mutants. (A) Schematic illustration of the *idd14-1*, *idd15-5*, and the *IDD16-RNAi* construct. A 248-bp specific *IDD16* cDNA fragment was used for construction of *p35S::IDD16-RNAi*. (B) Semi-quantitative RT-PCR analysis of *IDD* genes in WT and *idd* mutants. The transcripts of *IDD14*, *IDD15*, and *IDD16* in the *idd* triple mutant are shown as representatives. (C) 36-day-old plants of *idd* single, double, and triple mutants. Arrows indicate the infertile siliques. The arrowheads show the increased angles between the inflorescence stems and branches. The scale bar represents 2 cm. (D) Morphology of sixth leaves in 25-day-old *idd* mutants. The longitudinal curvature (LC) index and leaf index were determined from at least 10 leaves in each genotype. The data are shown as mean values \pm one SD (Student's *t*-test, **P*<0.05 and ****P*<0.001). (E) Enlarged floral organs and infertile siliques in the *idd* triple mutant. The arrow indicates the stigma lacking pollen. The arrowheads show the unfertilized ovules.
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(Figure 5B, S3A), demonstrating that *IDD15* and *IDD16* function coordinately in gravitropic responses. Interestingly, although *idd14-1* did not show any defect in gravitropic response, the inflorescence stems of *idd14-1D* were hypersensitive to gravistimu-

lation (Figure 5B, S3A), indicating that ectopic expression of *IDD14* also influences gravitropic responses.

Surprisingly, although the *IDD* genes were found to be expressed in hypocotyls and roots, we did not observe any obvious phenotype in hypocotyls or roots of either *idd14-1D* or the *idd* triple mutant, including gravitropic response phenotypes (Figure S3B, S3C). The only aberration we observed was a slightly waved primary root phenotype in the *idd* triple mutant when grown vertically (Figure S3D). These findings suggest that the three *IDDs* may primarily function in lateral aerial organs.

IDD Affects Auxin Accumulation by Modulating Auxin Homeostasis and Transport

Previous studies have suggested that *IDD14* and *IDD15* are involved in the regulation of starch metabolism [38,40]. However, the narrow, epinastic leaves in gain-of-function *IDD* mutant or transgenic plants seem to resemble, to some extent, those observed in auxin overproduction mutants or transgenic plants, such as *yucD* or *p35S::YUCs* transgenic plants [6,7,9]. By contrast, the rotund and curly rosette leaves, abnormal floral phenotype, and gravitropism defect in the loss-of-function *idd* mutants are also documented in the mutants defective in auxin biosynthesis or transport [2,9–11]. This led us speculate that auxin accumulation or signaling may also be involved in *IDD*-mediated organ development and/or gravitropic response. To test this, we used the *idd14-1D* and *idd* triple mutant plants as representatives of gain- and loss-of-function *idd* mutants for our further analysis. We first examined auxin accumulation in their leaf, inflorescence stem, and root, by monitoring the expression of *DR5::GUS* or *DR5::GFP*, a widely used auxin gradient reporter [50]. On day 3 after leaf initiation, an obvious GUS signal was observed in WT leaf distal tips. A stronger and spatially-expanded GUS signal was observed in *idd14-1D* leaf tips. No obvious GUS signal, and thus no localized auxin maxima, was observed in the leaf tips of *idd* triple mutant plants (Figure 6A). Similar differential GUS patterns were subsequently observed in 5-day-old expanding and 15-day-old expanded leaves (Figure 6A). In inflorescence stems, the GUS signal was mainly observed in vascular tissues in WT, whereas strong GUS staining was present in the cortex and endodermis tissues in *idd14-1D* and weak GUS expression without a tissue-specific pattern was observed in the *idd* triple mutant (Figure 6B). The increased or decreased auxin accumulation was also found in the meristem regions of primary roots in *idd14-1D* and the *idd* triple mutant, respectively (Figure 6C). Further quantification of the GUS activity in these organs confirmed the variations of *DR5::GUS* signals observed in *idd14-1D* and the *idd* triple mutant (Figure 6D). This observation strongly suggests that alteration of *IDD* expression affects auxin accumulation in multiple organs.

Auxin homeostasis and transport are key factors that determine the accumulation of auxin in plant organs [1,2,4]. To assess the contributions of auxin homeostasis and transport to *IDD*-mediated auxin accumulation, we first quantified the endogenous free IAA levels in the *idd14-1D* and *idd* triple mutants. Consistent with enhanced or decreased expression of *DR5::GUS* reporter observed,

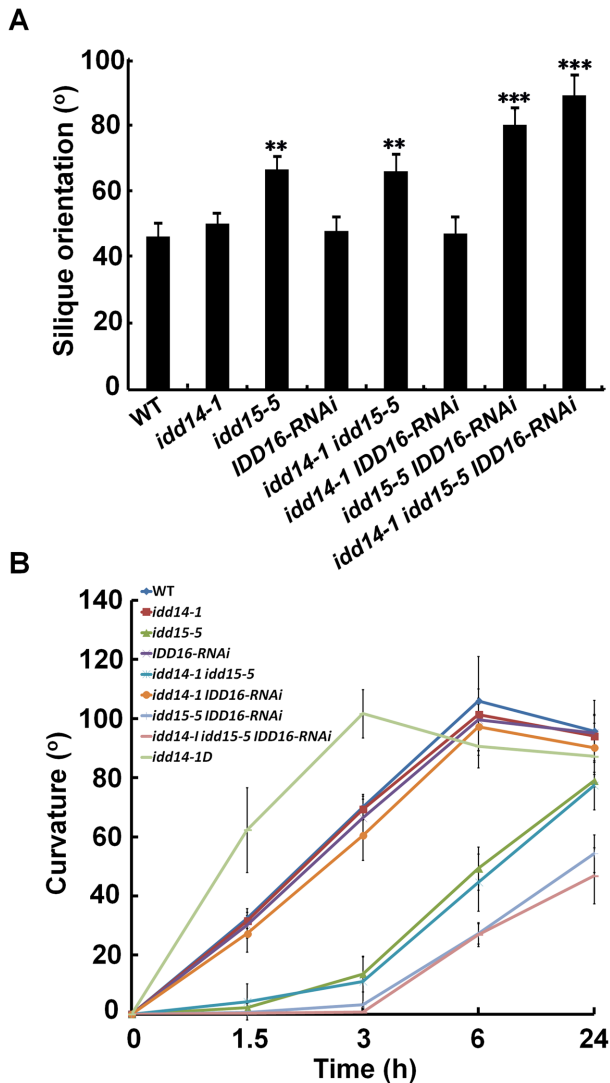


Figure 5. Silique orientation and gravitropic response in gain- and loss-of-function *idd* mutants. (A) Silique orientation in the *idd* single, double, and triple mutants. The angles between siliques and inflorescence stems were measured in 45-day-old WT and *idd* mutant plants. The data are shown as mean values \pm one SD (Student's *t*-test, ***P*<0.01 and ****P*<0.001). (B) Gravitropic responses of inflorescence stems in *idd14-1D* and loss-of-function *idd* mutants. The degree of curvature was determined from at least 18 inflorescence stems in each genotype after gravistimulation at the indicated times. The data are shown as mean values \pm one SE.
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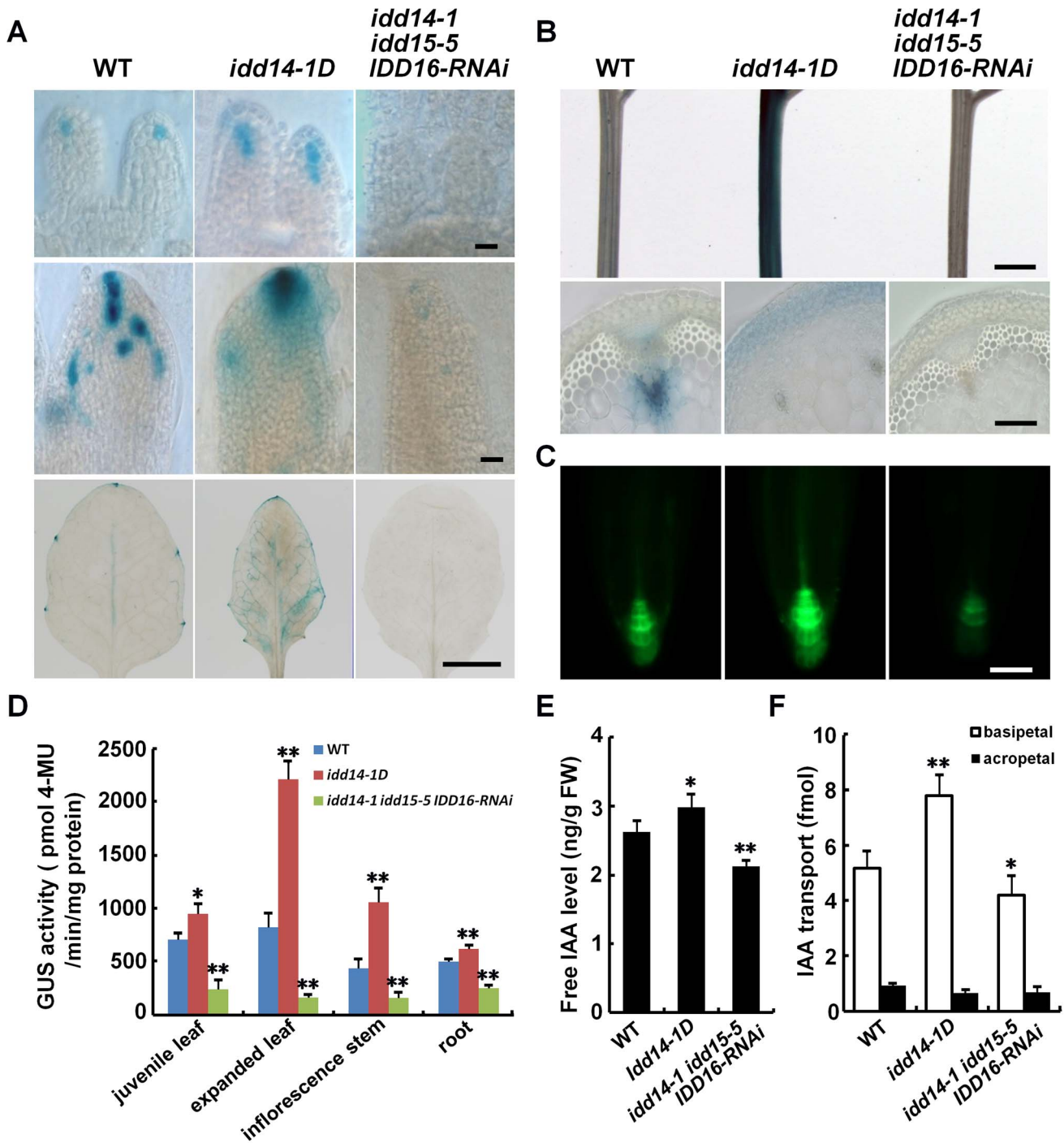


Figure 6. Altered auxin accumulation and transport in gain- and loss-of-function *idd* mutants. (A) Auxin accumulation assayed with the *DR5:GUS* reporter in WT, *idd14-1D*, and *idd* triple mutant leaves. The leaves at 3, 5, and 15 days after initiation (from top to bottom panels) were subjected to GUS staining. The scale bars represent 50 μ m in the top and middle panels and 5 mm in the bottom panel. (B) Expression of the *DR5:GUS* reporter in inflorescence stems of WT, *idd14-1D*, and *idd* triple mutant plants. The scale bars represent 2 mm in the top panel and 100 μ m in the bottom panel. (C) GFP fluorescence signals in the primary roots of WT, *idd14-1D*, and *idd* triple mutant carrying a *DR5:GFP* reporter. The scale bar represents 50 μ m. (D) Quantification of the GUS activity in the organs of the WT, *idd14-1D*, and *idd* triple mutant carrying a *DR5:GUS* reporter. Data are from three biological replicates and shown as mean values \pm one SD (Student's *t*-test, **P*<0.05 and ***P*<0.01). (E) Endogenous free IAA levels in WT, *idd14-1D*, and *idd* triple mutant plants. Aerial organs of 15-day-old plants were used for measurement of free IAA levels. Data are from four biological replicates and are shown as mean values \pm one SD (Student's *t*-test, **P*<0.05 and ***P*<0.01). (F) Polar auxin transport capability of WT, *idd14-1D*, and *idd* triple mutant stems. The inflorescence stem segments of 5-week-old plants were used to determine the basipetal IAA transport efficiency and the background acropetal movement. Data are from four biological replicates and shown as mean values \pm one SD (Student's *t*-test, **P*<0.05 and ***P*<0.01).

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the endogenous IAA level was increased by about 13% in *idd14-1D* but decreased by approximately 19% in *idd* triple mutant plants as compared to that in WT (Figure 6E). This result implied that the *IDD* genes may affect auxin biosynthesis. We then measured the auxin transport capability of inflorescence stems in *idd14-1D* and *idd* triple mutant. As shown in Figure 6F, the basipetal IAA transport efficiency was increased by over 50% in *idd14-1D* stems but decreased about 18% in *idd* triple mutant stems when compared with that in WT stem, demonstrating that *IDD* also modulates the auxin transport process. To further examine whether auxin signaling is affected in gain- and loss-of-function *idd* mutants, we monitored the expression of the *DR5:GUS* reporter, *IAA5*, and *IAA29* in response to exogenous IAA treatment in *idd14-1D* and the *idd* triple mutant, and observed that the expression of the *DR5:GUS* reporter, *IAA5*, and *IAA29* was normally induced following application of IAA as that in WT plants (Figure S4A, S4B), suggesting that *IDD* has no effect on auxin perception or signaling. We also observed that the expression of *IDD14*, *IDD15*, and *IDD16* transcripts was not modulated by auxin treatment (Figure S4C). Taken together, our results strongly suggest that the three *IDD* genes are involved in the establishment of auxin gradients through the regulation of auxin biosynthesis and transport.

IDD Directly Activates the Expression of *YUC5*, *TAA1*, and *PIN1*

To identify the genes downstream of *IDD*, we first carried out a real-time quantitative RT-PCR (qRT-PCR) analysis to examine the transcript abundances of genes known to function in auxin biosynthesis and transport in the *idd14-1D* and *idd* triple mutants. Among the *YUC* and *TAA* family genes, the transcription of *YUC1*, *YUC2*, *YUC3*, *YUC4*, *YUC5*, *YUC8*, and *TAA1* was found to be elevated in *idd14-1D*, and the transcription levels of *YUC2* and *YUC5* were decreased in *idd* triple mutant, when compared to those in WT plants (Figure 7A). Among 14 genes related to auxin transport, the transcription of *AUX1*, *PIN1*, *ABCB1*, *ABCB4* and *WAG1* were found to be elevated in *idd14-1D* but reduced in the *idd* triple mutant compared to those in WT plants. *PIN4* and *PINOID* (*PID*) transcripts were only elevated in *idd14-1D* or decreased in the *idd* triple mutant (Figure 7A). We further investigated the expression of 11 genes, which had the apparently altered expression in *idd14-1D* or the *idd* triple mutant (with a significance at $P < 0.01$), in the transgenic plants overexpressing *IDD15* or *IDD16* and the *idd15-5* *IDD16-RNAi* plants. As expected, their differential expressions in gain- and loss-of-function *IDD15* and *IDD16* plants were much similar to those observed in *idd14-1D* and the *idd* triple mutant (Figure S5A), demonstrating that these genes are also downstream of *IDD15* and *IDD16*.

As the *IDDs* are transcription factors, we speculated that some of the genes differentially expressed in gain- and loss-of-function *idd* mutants might be directly targeted by *IDD*. We first identified the genes whose expression was rapidly induced by the activation of *IDD*, using the chemically-inducible *IDD14* transgenic plants. After the transgenic seedlings were treated with the inducer, *IDD14* was dramatically induced by 0.5 h, and the expression of *YUC5*, *TAA1*, and *PIN1* was obviously elevated within 2 h (Figure S5B), suggesting that three genes might be direct targets of *IDDs*. To examine whether *IDD* can directly bind to the promoter regions of *YUC5*, *TAA1*, or *PIN1*, we performed chromatin immunoprecipitation (ChIP) assays with both *p35S::IDD16-GFP* and *p35S::IDD14-GFP* transgenic plants. It has been reported that the maize ID1 and ID domain proteins could bind to a specific 11 bp DNA consensus motif, T-T-T-G-T-C-G/C-T/C-T/a-T/a-T [35]. As such, we targeted similar possible *IDD*-binding motifs

in the promoter and/or upstream coding regions of *YUC5*, *TAA1*, *PIN1*, *YUC2*, and *YUC3*, and carried out the ChIP analysis (Figure 7B). As expected, we found that the three fragments containing a putative *IDD*-binding motif in *YUC5*, *TAA1*, or *PIN1* were greatly enriched by *IDD16* after GFP immunoprecipitation, whereas no binding activity was detected in the promoter regions of *YUC2* or *YUC3* (Figure 7B). Such enrichment was detected in a control DNA fragment of the *QUA-QUINE STARCH (QQS)* promoter, which was previously reported to be a target of *IDD14* [38], but no enrichment was detectable in a control DNA fragment in the *ACTIN2 (ACT2)* promoter which lacked the putative *IDD*-binding motif (Figure 7B). Likewise, similar enrichments of these DNA fragments were further confirmed by ChIP assayed with *IDD14* protein (Figure S5C). In addition, we indeed visualized the enhanced or attenuated *PIN1* accumulation in the roots of *idd14-1D* and *idd* triple mutant carrying a *pPIN1::PIN1-GFP* construct, respectively (Figure S6). These results illustrate that *IDD* can directly target *YUC5*, *TAA1*, and *PIN1*, to activate their expression.

Alteration of Auxin Accumulation Suppresses Organ Morphogenesis in *idd* Mutants

Since *idd14-1D* contains a high auxin level while the *idd* triple mutant has low endogenous auxin content, we attempted to genetically modify auxin biosynthesis to examine whether this could suppress or restore the phenotype observed in gain- and loss-of-function *idd* mutants. We first generated an *idd14-1D yuc2 yuc6* triple mutant through genetic crosses, and observed that loss-of-function *yuc2 yuc6* completely suppressed the leaf phenotypes of *idd14-1D* (Figure 8A, S7), and partially attenuated the hypersensitivity of *idd14-1D* inflorescence stems to gravistimulation (Figure 8B). Further, when we overexpressed *YUC2* in the *idd* triple mutant, the ectopic expression of *YUC2* fully rescued the infertile silique defect (Figure 8C), but only partially restored the silique orientation in the *idd* triple mutant (Figure 8D). These results provide further evidence that auxin biosynthesis is genetically downstream of this *IDD* subfamily.

As the amyloplast movement in endodermal cells of *idd15* stem has been reported to be defective under gravistimulation [39–41], we also investigated whether ectopic expression of *YUC2* has an effect on amyloplast movement in endodermal cells of the *idd* triple mutant stems. As shown in Figure S8, the retarded amyloplast movement in endodermal cells of the *idd* triple mutant stems under gravistimulation was not rescued by ectopically expressed *YUC2* (Figure S8), suggesting that altered auxin accumulation does not affect *IDD*-mediated amyloplast responsiveness to gravistimulation.

Discussion

The Distinct *IDD* Subfamily Is Critical for Organ Development and Plant Architecture Formation

The *IDD* family has been defined as a plant-specific transcription factor family [35,51], and previous characterizations of a few *IDD* members have indicated that *IDDs* are involved in the regulation of transition to flowering and starch metabolism [36–38]. *IDD14*, *IDD15*, and *IDD16* belong to a subfamily that is distinct from the other *IDD* members in *Arabidopsis*, rice, and maize [34]. The mutation of *IDD15* in *Arabidopsis* and rice reduces gravitropic response in inflorescence stems [39–41,48]. However, because the loss-of-function *idd14* mutant does not have obvious organ phenotype, and *IDD16* has not yet been characterized, the functions of this subfamily are still largely unknown.

In this study, we characterized both gain- and loss-of-function mutants of this *IDD* subfamily, and discovered that these three *IDD* genes function redundantly and cooperatively in regulating

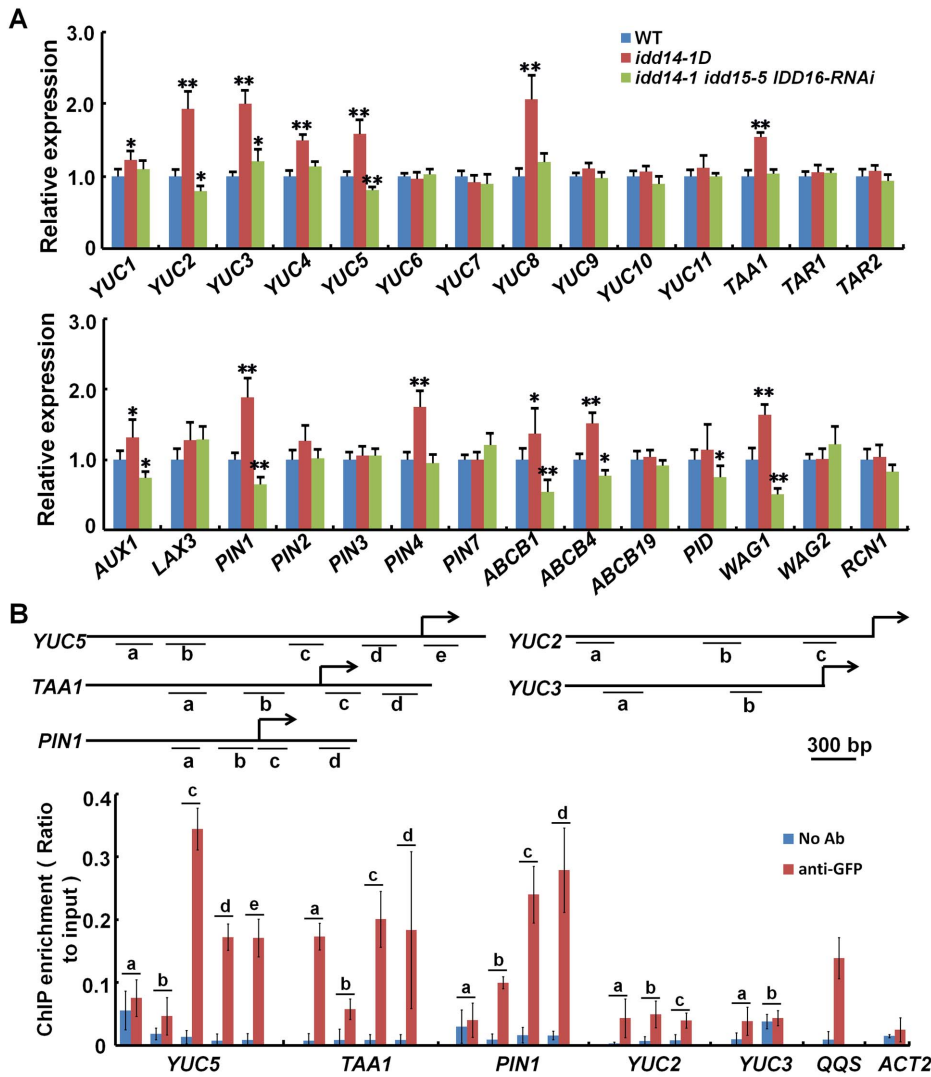


Figure 7. IDD Activates the expression of auxin biosynthetic and transport genes. (A) Relative expression levels of the genes involved in auxin biosynthesis (top panel) and transport (bottom panel) in WT, *idd14-1D*, and *idd14-1 idd15-5 IDD16-RNAi* plants. RNAs isolated from aerial organs of 4-week-old plants were subjected to qRT-PCR analysis, and the data are from three biological replicates and shown as mean values \pm one SD (Student's *t*-test, **P*<0.05 and ***P*<0.01). (B) ChIP assay performed with *p35S::IDD16-GFP* transgenic plants by anti-GFP antibody. The DNA fragments with a putative IDD-binding motif in the promoter and upstream regions of *YUC5*, *TAA1*, *PIN1*, *YUC2*, and *YUC3* are indicated as the letters a–e (top panel), and the enrichment values of their PCR products were quantified by qPCR (bottom panel). A reported IDD-targeted fragment in the *QQS* promoter and a fragment in the *ACT2* promoter were used as the positive and negative controls, respectively. The data are from three biological replicates and shown as mean values \pm one SD. doi:10.1371/journal.pgen.1003759.g007

organ morphogenesis and gravitropic responses. Gain-of-function of each *IDD* led to a small, narrow, and down-curved leaf phenotype. Although *idd* single mutants did not have obvious organ morphological phenotypes (except the reduced gravitropic response observed in *idd15*), our further characterization of the *idd* double and triple mutants clearly demonstrates that *IDD14* and *IDD16* act redundantly to regulate the morphology of aerial organs and affect fertility, while *IDD15* and *IDD16* cooperatively control the gravitropic responses and plant architecture.

Such redundant and cooperative roles of the three *IDD* genes in organ morphogenesis and gravitropism are consistent with their differential and overlapping expression patterns in particular organs. *IDD14* and *IDD16*, but not *IDD15*, were expressed in juvenile leaves, whereas *IDD15* and *IDD16*, but not *IDD14*, were highly expressed in inflorescence stems. The three *IDD* genes were

all expressed in floral organs, and the abnormal flower phenotype in *idd14-1 IDD16-RNAi* was enhanced in the *idd* triple mutant. Interestingly, although the *IDD* genes were also expressed in hypocotyls and roots, we could not observe obvious changes in their morphology or gravitropic response in either gain- or loss-of-function *idd* mutants, except the slightly waved roots in *idd* triple mutant seedlings. This may be attributable to decreased expression of *WAG1*, a gene that is an indirect target of IDD and has been identified as a suppressor of root waving (Figure 7A) [52].

IDD14, IDD15, and IDD16 Regulate Auxin Gradients by Promoting Auxin Biosynthesis and Transport

Previous studies have shown that *IDD15* and *IDD14* are involved in the regulation of starch metabolism [38,40]. Here,

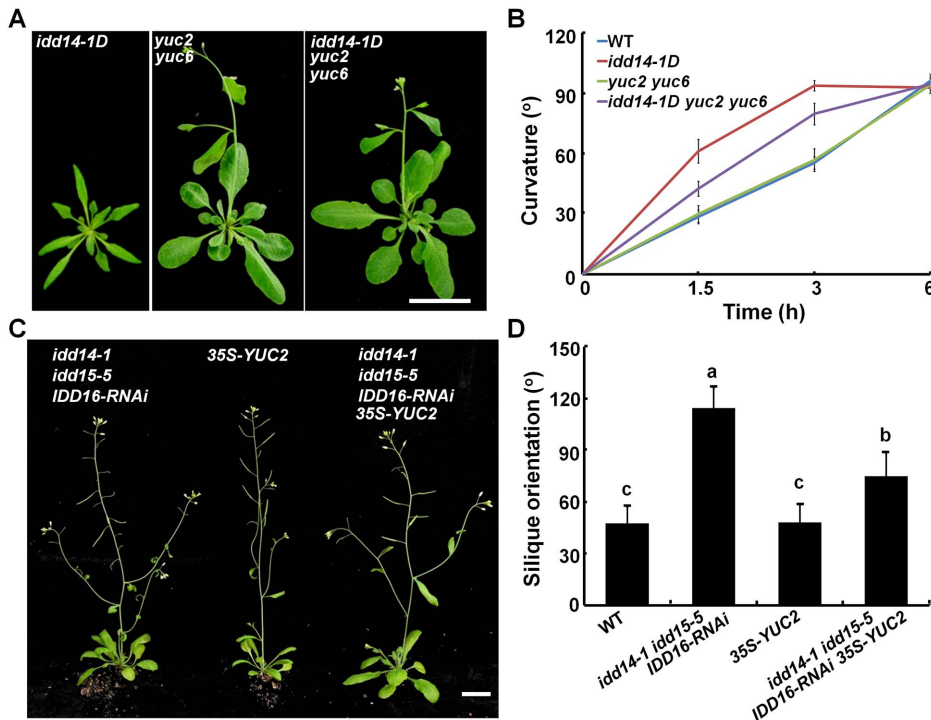


Figure 8. Genetic interaction of auxin biosynthesis and IDD-mediated organ morphogenesis and gravitropism. (A) Aerial organ morphology of 25-day-old *idd14-1D*, *yuc2 yuc6*, and *idd14-1D yuc2 yuc6* plants. The scale bar represents 2 cm. (B) Gravitropic responses of WT, *idd14-1D*, *yuc2 yuc6*, and *idd14-1D yuc2 yuc6* inflorescence stems. (C) 45-day-old plants of the *idd* triple mutant, *35S-YUC2*, and *idd* triple mutant carrying a *p35S::YUC2* construct. The scale bar represents 2 cm. (D) The silique orientation of WT, *idd* triple mutant, *35S-YUC2*, and *idd* triple mutant carrying a *p35S::YUC2* construct (one-way ANOVA test, $P < 0.05$). doi:10.1371/journal.pgen.1003759.g008

with a combination of phenotypic, genetic, and molecular approaches, we demonstrated that IDD14, IDD15, and IDD16 modulate auxin accumulation by affecting auxin biosynthesis and transport, thereby modifying organ morphogenesis and architecture formation. First, the organ phenotype in both gain- and loss-of-function *idd* mutants appears to be related to altered auxin homeostasis and distribution. For example, the narrow, epinastic leaves in the plants overexpressing *IDD* are similar to those in auxin overproduction mutants or transgenic plants, such as *yucca* (*yuc1D*), *yucca6-1D*, and *p35S::YUCs* transgenic plants [6,7,9], whereas the rotund and curly rosette leaves and abnormal floral phenotype in the loss-of-function *idd* mutants are similar to those observed in the mutants defective in auxin biosynthesis or transport [9,53]. Disruption of *IDD* genes also influenced the silique and branch angles, root waving, and gravitropism of inflorescence stems, which have been well documented to be related to polar auxin transport [2,54–57]. Second, expression analyses using *DR5::GUS* and *DR5::GFP* reporter clearly indicated that gain- or loss-of-function of *IDDs* enhanced or reduced auxin gradients, which was further confirmed by the increased or decreased endogenous auxin content and transport ability. Third, the expression of several genes involved in auxin biosynthesis and transport were altered in both gain- and loss-of-function *idd* mutants, and the *IDD* proteins could directly bind to the promoter regions of *YUC5*, *TAA1*, and *PIN1* to activate their expression. In addition, genetic manipulation of auxin biosynthesis could fully or partially restore the pleiotropic phenotypes in *idd14-1D* or the *idd* triple mutant. These results demonstrate that *IDD* indeed modulates auxin gradients by promoting auxin biosynthesis and transport.

IDD-Mediated Auxin Gradients and Starch Metabolism May Coordinate Gravitropic Response

The gravitropic response in plants requires a coordination of three sequential processes: gravity perception, signal transduction, and asymmetric growth response [58]. It is widely believed that the starch-filled amyloplasts (statoliths) within specific gravi-sensing cells (statocytes) perceive gravity stimulation [59,60]. Some other molecules, such as InsP_3 and Ca^{2+} have been found to be involved in gravity signaling [60]. A large body of evidence indicates that auxin plays a key role in gravitropic signaling and asymmetric organ growth, and that it may possibly be involved in gravi-sensing [2,60]. For example, many mutants related to auxin biosynthesis and especially transport such as *taa1*, *aux1*, *pin1*, and *pin2*, exhibit a defect in gravitropic responses [2,11,61]. Other mutants with altered silique or branch architecture, such as *plethora* (*pli*) in *Arabidopsis* and *lazy1* (*la1*) in rice, also show altered auxin accumulation or transport within their organs [54,56,62].

Among the three *IDD* members we characterized, *IDD15* and its rice ortholog, *LPA1*, have been previously reported to affect the gravitropic response by altering amyloplast sedimentation in the endodermis [39–41,48]. Recently, *IDD14* was also found to mediate starch degradation by directly activating the expression of *QQS* [38], suggesting that *IDD14* also participates in the regulation of starch metabolism. Our detailed characterization of gain- and loss-of-function of three *IDDs* provides substantial evidence that *IDD*-mediated auxin biosynthesis and transport contribute to the organ morphogenesis and also, to some extent, gravitropic responses, because the genetic manipulation of auxin biosynthesis does not alter the responsiveness of amyloplast to gravistimulation but partially restores the gravity sensitivity or defect in gain- or

loss-of-function *idd* mutants. Therefore, it is likely that IDD-mediated auxin accumulation and starch metabolism coordinately control the gravitropic responses. The IDD-regulated starch metabolism might be primarily involved in gravi-sensing while the IDD-regulated auxin gradient may be primarily involved in signaling and responses.

Materials and Methods

Plant Materials and Growth Conditions

The *Arabidopsis thaliana* accession Col-0 was used in this study. *idd14-1D* was isolated from a population generated by T-DNA activation-tagging mutagenesis. *idd14-1* (CS367164) and *idd15-5* (SALK_087765) were obtained from ABRC. All seeds were sterilized and germinated on 1/2 MS medium after vernalization for 2 days at 4°C, and the plants were grown in a culture room or growth chamber at 22±1°C with an illumination intensity of 80–90 μmol m⁻² s⁻¹ and a 16-h light/8-h dark photoperiod, as described previously [63].

Leaf Curvature and Gravitropism Assays

Leaf curvature was determined as described previously [64,65]. Briefly, the transverse curvature (TC) index was defined as TC = 1 - cw/pw, where cw and pw are the curved width and the pressed width of leaves, respectively. The longitudinal curvature (LC) index was defined as LC = 1 - cl/pl, where cl and pl are the curved length and the pressed length of leaves, respectively. To quantify the pressed width or length of leaves, the sixth leaf of 25-day-old plants was dissected transversely or longitudinally on a desk, and then the extreme distances between the margins of the leaf were measured before and after pressing.

To quantify the gravitropic responses of roots and hypocotyls, seedlings were grown vertically for 4 days and then turned horizontally, and curvature angles were measured [58]. For quantification of the gravitropic responses of inflorescence stems, 32-day-old plants with inflorescence stems of approximately 4–8 cm were gravistimulated by rotating them 90° in darkness, and stem curvatures were measured as the angles between the growing direction of apex and horizontal base line [39,58].

Genotyping of T-DNA Mutants

The T-DNA flanking sequence of *idd14-1D* was amplified by TAIL-PCR [66]. Three primers, P1, P2, and pSK-LB2 were used for co-segregation analysis. P1 and P2 were located in the *Arabidopsis* genome flanking the T-DNA insertion site, and pSK-LB2 was a primer corresponding to the left border of the T-DNA sequence. Similarly, *idd14-1* and *idd15-5* were genotyped with corresponding primers (Table S2), and *yuc2* and *yuc6* were genotyped according to the methods from a previous study [9].

Plasmid Construction and Plant Transformation

To generate *p35S::IDDs* and *p35S::anti-IDD14* transgenic plants, the *IDD* coding sequences were amplified by RT-PCR and ligated into the pGEM-T-Easy vector (Promega, USA), and then verified by sequencing. The resulting plasmids were digested with *EcoRI* and cloned into pVIP96 [67]. The *IDD14* cDNA was also cloned into pER8 to generate a chemically inducible *IDD14* construct. To investigate the tissue-specific expression of *IDD14*, *IDD15*, and *IDD16*, approximately 2-kb promoter fragments of *IDD* genes were amplified from genomic DNA and then fused with the β-glucuronidase (*GUS*) gene into pBI101. To generate the *p35S::IDD16-GFP* and *p35S::IDD14-GFP* constructs, an *IDD16* or *IDD14* coding sequence lacking a stop codon was amplified and then cloned in frame into pMDC83 (Invitrogen, USA). To

generate the *p35S::IDD16-RNAi* construct, a specific *IDD16* cDNA fragment was amplified and ligated inversely into pBluescript SK-GUSF [68], and then an *XbaI-BglII* digested fragment was subcloned into pVIP96. A *YUC2* cDNA fragment was amplified by RT-PCR and cloned into pVIP96myc to generate the *p35S::YUC2* construct. All the primers used are listed in Table S2.

All constructs were introduced into *Arabidopsis* by *Agrobacterium tumefaciens*-mediated transformation via the floral dip as described previously [69]. At least 20 independent lines harboring a single T-DNA insertion from each construct were generated, and 4–5 independent lines of T3 homozygous plants were used for detailed characterization.

Gene Expression Analysis

Total RNA was isolated using a guanidine thiocyanate extraction buffer [70]. For semi-quantitative RT-PCR or qRT-PCR analysis, cDNA was synthesized from 1 μg of total RNA using SuperScript III Reverse Transcriptase (Invitrogen, USA). qRT-PCR was performed with a Rotor-Gene 3000 thermocycler (Corbett Research, Australian) with a SYBR *Premix Ex Taq* II kit (Takara, Japan). The relative expression level for each gene was normalized to the *ACTIN2* and the data were collected from three biological replicates, as described previously [49].

The histochemical GUS assay was carried out according to previously described protocol [71]. The GUS activities were quantified by monitoring cleavage of the β-glucuronidase substrate 4-methylumbelliferyl β-D-glucuronide (MUG), as described [71].

For RNA *in situ* hybridization, a specific cDNA region of *IDD14*, *IDD15*, or *IDD16* was transcribed *in vitro* to generate sense and antisense probes using the Digoxigenin RNA labeling kit (Roche, Switzerland). The WT inflorescence stems were fixed and embedded in paraffin (Sigma-Aldrich, USA), and then sectioned to a 10 μm thickness. RNA *in situ* hybridization was performed according to a previously described method [54].

Free IAA Measurement

Aerial organs from 15-day-old plants of WT, *idd14-1D*, and *idd* triple mutant plants were used for measurement of free IAA content. The extraction, purification, and analysis of free IAA by gas chromatography-mass spectrometry was performed according to the methods described by Edlund *et al.* [72], except that an Agilent GC and a LECO Pegasus TOF mass spectrometer was used, with separation using a DB-5ht column (Agilent, USA) [63].

Auxin Transport Assays

Auxin transport in inflorescence stems was measured according to the methods of a previously published protocol [73]. 25-mm inflorescence segments were cut from 5-week-old plants, and the segments were submerged inversely into an auxin transport buffer (100 nM ³H-IAA, 0.05% MES, pH 5.5–5.7) in a 0.5-ml microcentrifuge tube. Control experiments were performed by submerging the base of inflorescence stems to measure acropetal IAA movement. After 12 h, 5-mm stem segments were dissected from the non-submerged ends and used to quantify the radiolabeled auxin using a scintillation counter.

Chromatin Immunoprecipitation (ChIP) Assays

ChIP assays were performed as described previously [74]. Briefly, 2 g of *p35S::IDD16-GFP* or *p35S::IDD14-GFP* transgenic plants grown on 1/2 MS plates for 16 days was harvested, and then submerged in 1% formaldehyde to crosslink the DNA with DNA-binding proteins. The chromatin pellets were extracted and sheared by sonication. 5 μl anti-GFP antibodies (Abcam, UK)

were used to immunoprecipitate the DNA-IDD16 or DNA-IDD14 complexes. DNA was released with proteinase K and then purified. The enrichment of DNA fragments was determined by quantitative PCR with the primers listed in Table S2.

Amyloplast Staining

Plants were grown in soil until the primary inflorescence stems bolted to a height of 4–9 cm, and then gravistimulated by rotating them upside down [40]. 1-cm-long inflorescence stems below the apex were fixed, embedded in paraffin, and sectioned to a 10- μ m thickness. A periodic acid-Schiff kit (Sigma-Aldrich, USA) was used for amyloplast staining, according to the manufacturer's instructions.

Supporting Information

Figure S1 IDD14, IDD15, and IDD16 belong to a distinct IDD subfamily. (A) Phylogeny of the *Arabidopsis* IDD family. A neighbor-joining tree of 16 IDD members was generated using CLUSTAL W based on the amino acid sequences of IDD proteins. Number of generations = 1000. (B) Transcriptional analysis of *IDD* genes in transgenic plants overexpressing *IDD14*, *IDD15*, or *IDD16*, respectively. Two independent lines of each construct were assayed by semi-quantitative RT-PCR and the expression of At1g68140 was determined in *35S-IDD14* plants. The *GLYCERALDEHYDE-3-PHOSPHATE DEHYDROGENASE C SUBUNIT (GAPC)* gene was used as an internal control. (TIF)

Figure S2 IDD14-GFP protein is localized in nucleus. (A) Phenotype of 25-day-old *p35S::IDD14-GFP* transgenic plant. The scale bar represents 1 cm. (B) Nuclear localization of the IDD14-GFP protein in transgenic plants. GFP fluorescence, DAPI staining, overlaid and bright fields (BF) are shown from left to right. The scale bar represents 10 μ m. (TIF)

Figure S3 Gravitropic response and root morphology of gain- and loss-of-function *idd* mutants. (A) Gravitropic responses of inflorescence stem in *idd* and *idd14-1D* plants. 32-day-old plants were gravistimulated by rotation by 90° in the dark for 0, 3, or 24 h. The scale bar represents 2 cm. (B–C) Kinetics of the gravitropic response of the hypocotyls (B) and primary roots (C) in WT, *idd14-1D*, and *idd* triple mutant plants. At least 20 seedlings from each genotype were used. The data are shown as mean values \pm one SE. (D) Morphology of primary roots of WT, *idd14-1D*, and *idd* triple mutant plants. Seedlings grown vertically for 6 days were photographed. Note that the primary roots in the *idd* triple mutant showed a slightly waving phenotype. The scale bar represents 0.5 cm. (TIF)

Figure S4 Auxin responses of *idd* mutants and *IDD* genes. (A) Expression of *DR5::GUS* in WT, *idd14-1D*, and *idd* triple mutant seedlings treated with/without auxin. The 5-day-old seedlings were treated with various concentrations of IAA for 6 h and subjected to GUS staining assays. The scale bar represents 2 mm. (B) Transcripts of *IAA5* and *IAA29* in WT, *idd14-1D*, and *idd* triple mutant before and after auxin treatment. (C) Transcript levels of *IDD14*, *IDD15*, and *IDD16* in WT plants before and after auxin treatment. Semi-quantitative RT-PCR was performed with the RNAs isolated from 10-day-old seedlings treated with 1 μ M IAA for the time durations indicated. *GAPC* was used as an internal control. (TIF)

Figure S5 Identification of IDD-regulated genes involved in auxin biosynthesis and transport. (A) Relative expression levels of the genes involved in auxin biosynthesis and transport in WT, *35S-IDD15*, *35S-IDD16*, and *idd15-5 IDD16-RNAi* mutant plants. RNAs isolated from aerial organs of 3-week-old plants were subjected to qRT-PCR analysis, and data are from three biological replicates and shown as mean values \pm one SD (Student's *t*-test, **P*<0.05 and ***P*<0.01). (B) Expression analyses of IDD-regulated genes in transgenic plants carrying an inducible *IDD14* construct. 15-day-old transgenic plants were transferred into a liquid medium containing DMSO or 10 μ M β -estradiol for the indicated time durations, and subjected to RNA isolation and qRT-PCR analysis. Data are from three biological replicates and shown as mean values \pm one SD. Note that *YUC5*, *TAA1*, and *PIN1* are rapidly induced by the activation of IDD14. (C) ChIP assay performed with *p35S::IDD14-GFP* transgenic plants by anti-GFP antibody. The DNA fragments with a possible IDD-binding motif in the promoter and upstream regions of *YUC5*, *TAA1*, *PIN1*, *YUC2*, and *YUC3* (a–e) were assayed by ChIP, and the enrichments of their qPCR products are shown as mean values \pm one SD from three biological replicates. An IDD-targeted fragment in the *QQS* promoter and a fragment in the *ACT2* promoter were used as the positive and negative controls, respectively. (TIF)

Figure S6 PIN1 accumulation in gain- and loss-of-function *idd* mutants. GFP fluorescent signals in primary roots of WT, *idd14-1D*, and *idd* triple mutant plants containing a *pPIN1::PIN1-GFP* construct. GFP fluorescence, bright fields (BF), and overlaid images are shown from left to right. The scale bar represents 50 μ m. (TIF)

Figure S7 Suppression of *idd14-1D* phenotypes by *yuc2 yuc6*. (A–C) The blade areas (A), transverse curvature (TC) index (B), and leaf index (C) of WT, *idd14-1D*, *yuc2 yuc6*, and *idd14-1D yuc2 yuc6* leaves. At least 10 sixth leaves from each genotype were used for determination of the leaf area, TC index, and leaf index, respectively. Data are shown as mean values \pm one SD (one-way ANOVA test, *P*<0.05). (TIF)

Figure S8 Alteration of auxin biosynthesis does not affect amyloplast sedimentation in *idd* mutants. Plants were gravistimulated by turning upside down for 0 or 20 min, and longitudinal sections of inflorescence stems were prepared and then stained with a periodic acid-Schiff kit. Arrowheads indicate the retarded movement of amyloplasts in the endodermal cells of the triple *idd* mutant and triple *idd* mutant carrying a *p35S::YUC2* construct. The scale bar represents 10 μ m. (TIF)

Table S1 Phenotypic characterization of *cuf1-D* plants. (DOC)

Table S2 Primers used in this study. (DOC)

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Conceived and designed the experiments: YH DC. Performed the experiments: DC JZ YJ MF JL ZW WX. Analyzed the data: DC YH. Wrote the paper: YH DC.

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