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Advanced function, design and application of skin substitutes for skin regeneration

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ABSTRACT

The development of skin substitutes aims to replace, mimic, or improve the functions of human skin, regenerate damaged skin tissue, and replace or enhance skin function. This includes artificial skin, scaffolds or devices designed for treatment, imitation, or improvement of skin function in wounds and injuries. Therefore, tremendous efforts have been made to develop functional skin substitutes. However, there is still few reports systematically discuss the relationship between the advanced function and design requirements. In this paper, we review the classification, functions, and design requirements of artificial skin or skin substitutes. Different manufacturing strategies for skin substitutes such as hydrogels, 3D/4D printing, electrospinning, microfluidics are summarized. This review also introduces currently available skin substitutes in clinical trials and on the market and the related regulatory requirements. Finally, the prospects and challenges of skin substitutes in the field of tissue engineering are discussed.

1. Introduction

The skin is vulnerable to frequent damages. Minor superficial skin injuries are capable of healing by epithelialization without any particular treatment. Severe injuries may delay recovery time and cause a permanent loss of skin function or life-threatening. Skin diseases are the fourth most common non-fatal disease, affecting one-third of the global population [1]. Caring for skin diseases requires time, medical care, and financial investment, placing a heavy burden on governments [2]. Skin diseases also have a significant impact on people's mental health, which is an urgent problem that needs to be addressed [3]. Therefore, clinical intervention is critical for wound healing. The most effective strategy is to use an artificial skin substitute, such as a wound dressing or skin graft, as a temporary matrix to facilitate the healing process [4].

Treating wounds has always been a challenge for humans. Suitable skin substitutes are necessary for skin tissue engineering, so it is important for to choose appropriate materials and study their characteristics in skin tissue engineering [5]. With advances in technology, researchers began to explore various materials and preparation methods, gradually expanding the application of skin substitutes to a wider range of clinical fields.

In addition to basic protective functions such as water retention, antibacterial, and adhesion, skin substitutes are also designed for advanced functions such as mechanical strength, scar resistance, and intelligent skin to simulate normal skin. Cells are incorporated in skin substitutes for secreting proteins, cytokines, and growth factors used for wound healing. Different cells are selected to simulate the epidermis, dermis, and dermo-epidermal composite to simulate normal skin structure. More and more multifunctional skin substitutes are being developed.

Time doesn't cure everything, but skin substitutes is making an effort to achieve this. In this review, the function and design requirements of artificial skin substitutes are summarized, and the development status of artificial skin with different functions is classified and introduced. Additionally, representative manufacturing methods and research results to realize the advanced functions are discussed. Although many biologically engineered methods, such as decellularization and cellcontaining scaffolds, have been explored to treat wounds, many of

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these methods are still in the preclinical stage. This review also introduced the current research progress on clinical and commercial products and the related regulatory requirements (Fig. 1).

2. Wounds and skin substitutes

2.1. Skin and wound management

Human skin (Fig. 2A) consists of several types of cells distributed across the skin layers, including the epidermis, dermis, and subcutaneous tissue [12]. The **epidermis**, which is attached to the dermis's extracellular matrix with a thickness of $10-30 \mu$ m, is primarily composed of keratinocytes and other cells, such as Melanocytes, Langerhans cells, and Merkel cells [13]. Keratinocytes can form numerous tight junctions between cells and are capable of unlimited proliferation [14]. Melanocytes are responsible for producing and transferring melanin to the Keratinocytes and aesthetically harmonize the color [15]. Langerhans cells are involved in antigen presentation during the

immune process, while Merkel cells are responsible for tactile responses [16].

The dermis connects the epidermis and the subcutaneous tissue, mainly consisting of fibroblasts and extracellular matrix (ECM). Fibroblasts are the primary cellular components of the dermis, capable of supporting the formation of ECM by secreting proteins, cytokines, and growth factors used for wound healing. Fibroblasts can also migrate to the wound site and differentiate into myofibroblasts, making them attractive candidates as carrier cells for designing skin substitutes [17]. The ECM represents the largest component of skin and contains a complex collection of fibrous structural proteins (mainly collagen) and polysaccharides. The ECM provides dermal skin with strength and elasticity, allowing the cells to be suspended and form 3D structures [18, 19]. Besides, the dermis contains abundant blood vessels to create a microvascular network, angiogenesis are important to skin regeneration as vascular defect can hinder the supply of nutrition, oxygen, and growth factors [20,21]. Hypodermis is mainly composed of adipose tissue cells and are the primary storage for triglyceride fat and will be



Fig. 1. Scheme of the properties for skin substitutes. Skin substitutes can be prepared from autologous or allogeneic cell cultures for wound coverage and treatment. Technologies such as hydrogels, 3D/4D bioprinting, *in-situ* bioprinting, electrospinning, or microfluidics can be employed to manufacture these substitutes. Skin substitutes are characterized by their properties of providing (A) protection [6], (B) antibacterial activity, (C) adhesion and hemostasis [7], (D) mechanical strength, (E) anti-scarring ability, (F) drug loading [8], (G) detection [9], (H) perception [10], and (I) wearability [11].



Fig. 2. Human skin structure and skin substitutes. A. The structure of human skin comprises three main layers, including epidermis (keratinocytes, melanocytes, Langerhans cells), dermis (with fibroblasts and ECM components), and the hypodermis. B. Cell types in skin substitutes. The skin substitutes can be divided into acellular substitutes and cellular substitutes classified by their composition.

involved in the regeneration of the skin and skin accessories [22,23]. The three layers of skin work together to create an effective barrier against the external environment while allowing the transmission of sensory information, playing a vital role in maintaining the body's internal balance [24].

2.2. Wounds and skin substitutes

The wound can be divided into acute wound and chronic wound based on the time needed of wound healing. Acute wound healing follows the four processes of hemostasis, proliferation, maturation, and remodeling. The total healing time is typically about 8–12 weeks. However, if not treated properly, acute wounds can become chronic wounds, leading to long-term inflammation. Chronic wounds usually damage the inner lining of the skin and are associated with delayed or inadequate healing. In diabetic foot ulcer, the healing process is interrupted and replaced by a persistent state of inflammation and longterm microcirculatory deficiencies [25].

Moreover, diabetic wounds or burn wounds are vulnerable to bacteria and other pathogens due to impaired integrity and immune responses, resulting increased reactive oxygen species (ROS) production and worsening of the inflammatory response [26]. Severe burns can also impair organ function and lead to multiple organ dysfunction syndrome. The recruited inflammatory cells at the wound produce massive amounts of ROS, which result in oxidative stress [27]. Inflammatory factors such as tumor necrosis factor α (TNF- α) and interleukins 6, 8, and 1 β (IL-6, IL-8, and IL-1 β) are over produced and will increase thus prevent transition to the proliferative phase and disrupts the processes of angiogenesis, ECM remodeling, and re-epithelialization for tissue repair [26].

Wound dressings play a crucial role in the healing of complex acute and chronic wounds by absorbing wound exudate, maintaining a moist environment, and promoting accelerated wound healing. Furthermore, wound dressings can provide controlled release and spatio-temporal transmission of drugs or growth factors during wound healing to regulate cell proliferation and differentiation [28]. However, passive dressings such as gauze can only provide coverage and absorb exudates from the wound surface. These dressings may also attach to newly formed granulation tissue and cause pain when removed.

Although auto transplantation is considered the gold standard for skin regeneration, it has some disadvantages, such as inadequate skin supply, secondary damage, and unsuitability for diabetic patients. Additionally, there is a shortage of donors and a risk of hyperplastic scars or keloids for donors. Allotransplantation and xenotransplantation both carry the risk of virus transmission and immune rejection, which can affect the survival rate of transplanted skin [29].

With the development in tissue engineering and biomaterial manufacturing, a variety of advanced artificial skin substitutes have been developed and are providing clinical solutions for patients who need tissue replacement, ultimately improving their quality of life [30]. In this review, **skin substitutes** are defined as a material or device that can be applied to a patient's damaged skin, with the aim of replacing, imitating, or improving any skin function. This encompasses a diverse range of wound dressings that can be placed at the wound site to temporarily or permanently replace the skin's functionality, including acellular skin substitutes, such as acellular matrix, as well as skin substitutes, which are materials and scaffolds specifically designed for wound management.

2.3. Cells in skin substitutes

The category of skin substitutes can be divided into several types, including acellular substitutes and cellular substitutes classified by their composition. In terms of skin structure, skin substitutes can be applied as epidermal, dermal, or dermo-epidermal (composite). Based on the type of biomaterials used, skin substitutes can be classified into biological (autologous, allogeneic, xenogeneic) or synthetic (biodegradable, non-biodegradable) materials [31].

Acellular substitutes are tissues that have been decellularized from cadaveric dermis, allogenes, or animals. Acellular skin substitutes have intact ECM components and can retain the primary functional structure of normal dermal tissue [32]. They are frequently utilized in a variety of pathologies, such as diabetic foot ulcer, rare skin conditions, and plastic surgery [33,34]. However, acellular skin substitutes have limitations, including poor barrier function and the risk of disease transmission. Moreover, the sterilization process such as irradiation, may damage or change the structural components of skin substitutes [35]. In addition to mammalian acellular grafts, acellular fish skin (AFS) xenografts can be used to treat superficial and partial-thickness burns [36]. AFS contains collagen, fibrin, proteoglycans and glycosaminoglycans, and offers advantages of accelerating wound healing, reducing treatment costs, and low immunogenicity [37].

Xenografts implants sometimes will be recognized by the host as "foreign bodies," triggering a complex signaling cascade known as the foreign body response (FBR) during wound healing [38]. The FBR involves four stages: protein adsorption, acute inflammation, chronic inflammation, and collagen encapsulation. Skin grafts typically trigger an acute inflammatory reaction in the host within a few hours of transplantation, causing inflammatory cells such as neutrophils and monocytes to migrate to the implantation site, release proinflammatory cytokines, recruit and activate macrophages, and trigger a cascade immune response [39]. The fibroblasts that migrate to the wound continually deposit collagen fibers around the implants, resulting in a highly fibrotic structure in the outer layer of the implant, which is the phrase of chronic inflammation [40]. While FBR is the natural protective mechanism of the body, both chronic inflammation and acute inflammation will bring strong discomfort to patients, greatly affecting the function of implanted materials. Therefore, immune regulation design is necessary

to reduce the inflammatory reaction of skin substitutes in order to alleviate immune rejection and improve the biocompatibility of skin substitutes.

The inclusion of cells within the skin substitutes will promote the regeneration of natural skin through the secretion of growth factors. Cell-based skin substitutes typically contain somatic cells or stem cells (Fig. 2B). Among the human somatic cells, keratinocytes and fibroblasts are the most commonly cells used to fabricate cellular skin substitutes. Many other skin cells such as melanocytes and adipocytes are also included to create functional skin substitutes. Keratinocytes are essential for the re-epithelialization process and provide barriers against the skin and environment. Monteiro et al. developed a polyelectrolyte multilayer film with hyaluronic acid and poly-L-lysine by spray-assisted layer-bylayer assemble with keratinocytes attached and proliferated on the coated porous scaffolds. This epidermal-dermal scaffold can promote cell adhesion and regeneration for full-thickness skin defects [41]. Fibroblasts are the most abundant cells in the dermis and synthesize collagen and differentiate into myofibroblast phenotype during the wound closure process [8]. The synthesis and deposition of collagen and other ECM components will lead to the formation of the granulation tissue, which is a symbol during wound healing. Skin pigmentation is also crucial for the recovery of skin function. Melanocytes are responsible for regulating skin color and can restore the pigmentation of skin substitutes. The combination of keratinocytes with in skin substitute can result in the color that close to the natural epidermis. This is essential for addressing aesthetic concerns and promoting the mental well-being of patients after healing [42]. Adipocytes, which are located around hair follicles, play a crucial role in fibroblast migration and participate in the regeneration of skin appendages during wound healing [23].

Skin cells are a good source of autologous cells as they can be directly used, but their applications are limited due to the limited availability of donor cells and potential damage to the donation sites. Stem cells offer a promising alternative cellular source for the fabrication of artificial skin substitutes. The most commonly studied stem cells in wound healing include human skin stem cells (hSSCs), mesenchymal stem cells (MSCs), and induced pluripotent stem cells (iPSCs) [43].

hSSCs, which are extracted from the skin, have similar differentiation ability to other tissue-derived stem cells [44]. hSSCs can be isolated from different parts of skin such as dermal papilla or hair follicles, named dermal papilla stem cells (DPSCs) and follicle stem cells (FSCs), respectively [45]. MSCs are found in various tissues, including human bone marrow-derived MSCs, human umbilical cord Wharton's jelly-derived MSCs, and human adipose tissue-derived MSCs, respectively [46]. MSCs has the ability to differentiate into cells of epidermal and dermal lineages and contribute to tissue regeneration by producing growth factors, cytokines, and chemokines [47]. Moreover, MSCs can be activated to initiate immunomodulatory functions by producing immunoregulatory factors, and thereby regulating inflammation and reduce inflammation-related scar formation [48]. iPSCs are stem cells generated from the reprogramming process of individual somatic cells and have similar characteristics to embryonic stem cells on morphology, self-renewal, and differentiation capacity [49]. iPSCs can be extracted from fibroblasts or cord blood mononuclear cells and specialized differentiation into fibroblasts and keratinocytes, respectively [50]. Bilousova's group reported the differentiation of mouse iPSCs into a keratinocyte lineage, and the keratinocyte iPSCs were able to regenerate the epidermis, hair follicles, and sebaceous glands in a graft assay in vivo [51]. iPSCs have also been reported to differentiate into other cell types, such as melanocytes, sensory neurons and Schwann cells [52].

Immune cells, such as neutrophils, macrophages, and mast cells, play essential roles in the wound healing process [53]. Therefore, incorporating human immune cells into skin substitutes may provide opportunities to enhance regeneration potential and develop more complex models of skin substitutes.

2.4. Skin substitutes simulate different skin structure

The skin substitutes can be classified anatomically as epidermal, dermal, or composite substitutes based on their structure and intended application.

Epidermal substitutes typically consist of a single layer of autologous keratinocytes isolated from a donor and expanded *in vitro*. Although cultured epidermal autografts have the advantage of autologous graft compatibility, they also have some drawbacks such as high cost, short shelf life, and poor infection tolerance [54]. To overcome these limitations, Zhao et al. developed a HaCaT cells-encapsulated photocrosslinking gelatin methyl acrylamide (GelMA) hydrogel (Fig. 3A) that can prolong degradation in collagenase solution, reduce water loss, and promote epidermal reconstruction [55].

The dermis, which constitutes the bulk of the skin, provides strength and nourishment to the epidermis. Loss of dermal tissue due to wounds can result in the loss of fibroblasts and delay the wound healing process [56]. Autologous skin transplantation or dermal substitutes are often used to treat severe burns, and one key factor in the development of dermal substitutes is the regeneration of scaffold elasticity. For example, a sodium alginate/gelatin composite dermal substitute scaffold was fabricated using a motor-assisted 3D extrusion system and showed good cell proliferation results [57]. Chong et al. developed a triple-polymer scaffold composed of collagen, elastin, and polycaprolactone (CEP) with enhanced mechanical properties and elasticity. The electrospun CEP scaffolds were observed to promote keratinocyte and fibroblast proliferation, tissue integration, and early-stage angiogenesis after subcutaneous implantation (Fig. 3B) [58].

Dermo-epidermal composite skin substitutes refer to skin substitutes that include both epidermal and dermal components. Lin et al. developed a tri-layered chitosan-based scaffold that replicates the structure of full-thickness skin. They used a compact layer on top to simulate the epidermis, a thin film to simulate the basement membrane at the dermal-epidermal junction, and a porous layer to simulate the dermis, allowing for dermal cell proliferation [59]. OrCel (Orate International, Inc., USA) is an FDA-approved substitute that is synthesized by culturing allogeneic neonatal keratinocytes and fibroblasts in a type I bovine collagen porous sponge. This bi-layered composite allograft can produce an array of growth factors such as vascular endothelial growth factor (VEGF), platelet-derived growth factor (PDGF), Fibroblast growth factor (FGF), keratinocyte growth factor-1, and transforming growth factor- α , as well as other cytokines, which are favorable for host cell proliferation, migration, and wound healing [60]. Peramo et al. developed a 3D tissue structure containing epidermal skin, vermillion, and oral mucosa in a continuous layer. The mucocutaneous equivalent was fabricated using full-thickness human skin and oral mucosa explants to simulate the morphological features of the lips. This technique can be used as a general basis for other mucocutaneous areas, such as the anus and vagina [61].

The dermis and epidermis play an essential role in protection, and the meeting point of these two layers is known as the dermal-epidermal junction (DEJ) [62]. In native skin, the DEJ structure is corrugated, forming rete ridges (50–400 μ m in width) in the epidermis projecting more deeply into the dermis and generating dermal papillae (50–200 μ m in depth) [63]. DEJ can maintenance of structural integrity and control of the cellular microenvironment, which are essential for the appropriate keratinocytes functioning within these areas and plays a pivotal role in dermal-epidermal homeostasis and adhesion.

However, the interface between the epithelial layer and the underlying connective tissue in many skin substitutes are planar, causing weak adhesion between these layers [64]. So, mimicking the topographical and anatomical attributes of human skin mimic the skin microstructure is very important. It has been reported that after seeding keratinocytes onto decellularized dermis and reconstituting an epidermis, the dermal topography collapses and finally creates false rete ridges [65]. Ramos-Rodriguez et al. introduced complexity within electrospun membranes to mimic the morphology of the rete ridges in the skin [66]. Malara et al. developed a dermal template with stable dermal papillae through electrospinning collagen and seeding with human dermal fibroblasts using a combination of electrospinning and laser structuring (Fig. 3C). The ridged templates resulted in rete ridge was observed to formation in 2 weeks after grafting [67].

3. Functions of skin substitutes

3.1. Protective function as dressings

3.1.1. Water retention

Based on the concept of moist healing, which suggests that moisture benefits cells activities and promotes wound healing [68], water retention is an essential feature of wound dressing. While traditional wound dressings, such as gauze and cotton, insulate the injured tissue from the external environment, developing a wound dressing with good water retention properties is crucial. To enhance the freezing tolerance and moisture retention ability of organohydrogels, various agents can be added, such as ionic liquids, inorganic salts, zwitterionic osmolytes, and polyhydric alcohols [69]. For instance, Peng et al. developed a double network organohydrogel by incorporating ZnSO₄ into a polyvinyl alcohol-polyacrylamide (PPZE) hydrogel in a mixed solvent of ethylene glycol and water. After storage at ambient temperature for 31 days, the organohydrogel exhibited a moisture retaining rate of above 99.3 % (Fig. 4A) [70].

Polysaccharide-based wound dressings are effective strategies in the field of wound care, as they combine an ECM-like structure with excellent biomimicry. Chitosan, alginate [2], hyaluronic acid, konjac glucomannan [71] are common polysaccharides used in wound dressing. Hyaluronic acid and lactose-modified chitosan nanofibrous wound dressings were prepared by electrospinning, and their moisturizing ability helped to accelerate wound closure and have anti-scarring potential [72].

3.1.2. Antibacterial

Chronic wound infection is a serious problem during the healing process, as the skin cannot undergo self-repair under continuous bacterial infection. Antibiotics are a preferred strategy for the clinical treatment of infected wounds, and many antimicrobial drugs, such as tetracycline [73], are encapsulated in wound dressings. Inorganic metal nanoparticles, including silver [74], gold [75], zinc [76], photothermal therapy based on metal materials [77], carbon-based materials [78], nanozymes [79], or metal-organic frameworks (MOFs) [80], have been extensively studied. However, bacterial resistance caused by antibiotics and the potential for long-term retention of inorganic metal antimicrobials call for more effective antibacterial strategies. A multifunctional dressing (CHBC) was designed by the membrane-liquid interface multiple spraying method using the antibacterial agent hydroxypropyltrimethyl ammonium chloride chitosan, collagen I, and bacterial cellulose. The CHBC dressing had outstanding antibacterial properties and achieved wound healing for 8 days in a wound infection model (Fig. 4B) [81].

3.1.3. Adhesion and hemostasis

Hemostasis is the first stage of wound healing, and uncontrolled hemorrhage can lead to exsanguination. Therefore, achieving hemostasis is a basic and essential function in developing skin substitutes. Skin substitutes with adhesive properties can facilitate hemostasis by absorbing wound extract to congregate coagulation factors [82] or by sealing the wound though adhesiveness [83]. In addition to promoting hemostasis, wound dressings with adhesive properties can also help prevent infection by sealing the wound. Wang et al. developed a hydrogen-bond-based hydrogel with tough, wet, and underwater adhesion. The hydrogel was formed by polymerizing acrylic acid and 1-vinylimidazole to create hydrogen bonding and electrostatic



(caption on next page)

Fig. 3. Application of epidermal substitutes and dermal substitutes. A. Photocrosslinkable gelatin hydrogel for epidermal substitutes; (a) H&E stained sections of reconstructed epidermis on GelMA and control collagen scaffolds; (b) expression of proteins of reconstructed epidermis on hydrogel scaffolds [55]. Copyright 2016, Wiley. B. Electropsun collagen–elastin–PCL scaffolds as dermal substitutes; (a) SEM, H&E, and binary images of cross-linked CEP 1 and CEP 2 scaffolds; (b) H&E staining and SEM images of HDFs proliferation and infiltration on CEP 1 and CEP 2 scaffolds [58]. Copyright 2019, Elsevier. C. Skin substitutes mimic the dermal-epidermal junction; (a) H&E of CEAs (at *in vitro* culture day 20), and flat electrospun collagen dermal templates; (b) H&E stained cryosections of CEA-dermal template composite grafts 2 and 4 weeks post-grafting, demonstrating development of rete ridges when CEAs were grafted [67]. Copyright 2020, Mary Ann Liebert, Inc.



Fig. 4. Skin substitutes with water retention, antibacterial, adhesion, and hemostasis. A. Moisture retention PPZE hydrogel; (a) schematic illustration on the synthesis and structure of PPZE hydrogel; (b) SEM image of PPZE hydrogel; (c) effect of EG immersion on the weight-retention of PPZE-2 hydrogel with the storage time [70]. Copyright 2022, Elsevier. B. Schematic illustration of the preparation of a CHBC functional dressing via a novel membrane–liquid interface culture and the process of treating infected wounds *in vivo* [81]. Copyright 2022, Elsevier. C. Schematic illustration of PAAcVI hydrogel with tough underwater adhesion, Zn²⁺ responsiveness, and efficient hemostasis [84]. Copyright 2022, American Chemical Society.

interaction with the tissues. This hydrogel demonstrated excellent hemostatic ability in both a mice-tail docking model and a mice-liver hemorrhage model (Fig. 4C) [84]. Another approach to hemostasis is the application of hemostatic chitosan and kaolin in the preparation of chitosan/polyethylene oxide/kaolin nanofiber membranes using electrospinning technology. The hemostatic time of the nanofiber membranes was significantly lower (43 ± 1.4 s) than that of commercial QuikClot® Combat Gauze (55.7 ± 1.2 s) [85].

3.2. Advanced functions

3.2.1. Mechanical properties

The skin is a mechanically strong biological material, the collagen network and heterogeneous arrangement of hydrated matrix in the skin exhibit nonlinear stress-strain curve when subjected to external forces, redistributing pressure and absorbing strain to protect human organs from physical trauma and infection. Skin substitutes lie directly on the skin interface and will bear external stresses during application, so appropriate strength and softness (stiffness) are important factors for wound dressing. Strength refers to the maximum stress that a material can withstand when an external force is applied, and skin substitute needs to have a certain tensile strength to prevent rupture due to stretching during use. It also needs to have enough extensibility to be placed on joint areas such as knees, hips, and hands. Appropriate stiffness can ensure that the material is not easily deformed, in order to better adapt to the curvature and shape of the skin surface. In addition, stiffness can also affect the biomechanical properties of the material, such as sensation and comfort when in contact with the skin.

The skin substitutes will bear external stresses during application, so appropriate strength and softness are important factors for wound dressing. Silk fibroin and konjac glucomannan sponges in wet or dry state were reported to have similar compressive modulus with native skin tissue, the addition of konjac glucomannan can form interlocking entangled chains and intermolecular hydrogen bond to get tunable mechanical properties for wound dressing application [86].

The fatigue resistance refers to the stability of the material under multiple cyclic loading, which may be related to the service life of the material. For example, high fatigue resistance is required in joint areas such as the knee. To prevent failure after repeated stretching and to extend the service life without deformation or damage, the skin substitutes should have the ability to resist crack propagation during fatigue loads. Most fatigue resisting structure are produced by incorporating dynamic covalent or physical crosslinking in the network which rearrange through deformation [87]. Wang et al. reported a fatigue-free and self-healable hybrid ionic skin by polyurethane (PU) nano mesh and poly (acrylamide-*co*-acrylic acid) copolymer ionic matrix. The high-energy elastic PU nano mesh scaffold was embedded into the soft copolymer ionic matrix. Such a hybrid design leads to superhigh fracture energy (16.3 kJ m⁻²) and fatigue threshold (2950 J m⁻²) while maintaining skin-like self-healability, softness (modulus ~1.8 MPa), stretchability (680 %), and strain-stiffening response (37 times stiffness enhancement) (Fig. 5) [88].

3.2.2. Anti-scarring

Early-stage human fetuses and oral mucosal wounds undergo much less scarring during skin healing than cutaneous wounds, possibly due to a weaker inflammatory response in these cases. This observation suggests that the inflammation in adult cutaneous wounds may have evolved as a compromise to combat microbial infection at the expense of scarring [89].



Fig. 5. Fatigue-free and self-healable hybrid ionic skin. A. Composite structure and wound-repairing mechanism. B. Schematic structural changes of self-healable nanofibrous hybrid ionic skin upon reversible stretching. C. Schematic illustrations of the hybrid structure and respective self-healing mechanisms of PU nano-mesh scaffold and ionic matrix. D. Nanofiber tension reversibly breathe moisture in air. E. A rough comparison of the sensing and mechanical performance between human skin and nanofibrous hybrid ionic skin [88]. Copyright 2022, Nature.

During the process of wound healing, fibrosis can occur as a result of the body's inflammatory and connective tissue repair response, which facilitates physiological repair. However, fibroproliferative disorders may cause hypertrophic scars and keloid formation, resulting in functional problems [90]. Deep dermal fibroblasts are known to be pro-fibrotic, so adjusting the degree of fibrosis is essential for anti-scar treatment [91]. Zhang et al. developed an anti-scar wound dressing consisting of carboxymethyl chitosan, poly-y-glutamic acid, and anti-fibrotic polypeptide (AF38Pep). The genipin crosslinked hydrogel network exhibited wound healing and scar prevention properties. The RNA-Seq data revealed that AF38Pep in the network suppressed genes related to the focal adhesion pathway in heterograft keloid fibroblasts and thus regulate the deposition and distribution of favorable collagen to prevent scar [92]. Many 3D printing [93], electrospinning [94], and hydrogel have been used to prepare artificial skin to prevent scar formation. Ni et al. developed Aloe vera-chitosan hydrogel, the cationicity of chitosan demonstrated potential to mitigate hypertrophic scar in wound healing by suppressing the expression of a-smooth muscle actin (a-SMA) and promoting secretion of type I matrix metalloproteinases (MMP-1) [95].

3.2.3. Regulation during wound healing process

The wound healing process involves a complex cascade of cellular and biochemical events, and chronic or non-healing wounds may result in the dysregulation of these biological activities [96]. Skin substitutes that combine exogenous growth factors and cytokines can serve as a motivating factor in coordinating the behavior of cells and promoting tissue repair. The TGF family acts as a chemotactic agent for inflammatory cells and fibroblasts, promoting fibroblast proliferation and differentiation into myofibroblasts [97,98]. FGF has the ability to facilitate mitogenic and proangiogenic activities, promoting granulation tissue formation and re-epithelialization [99]. VEGF promotes endothelial cell proliferation, while angiopoietins act as blood vessel stabilizers or participate in their remodeling [100]. Products containing growth factors such as PDGF, EGF, and FGF are already approved and available for human use. These growth factors play a crucial role in



Fig. 6. Oxygen-releasing Structures. A. Genetically modified microalgae to engineer photosynthetic scaffolds; (a) gene modified microalgae seeded into photosynthetic dermal scaffolds; (b) bioactivity of recombinant human VEGF expressed by *C. reinhardtii in vivo* (Microalgae, red, chlorophyll auto fluorescence) [108]. Copyright 2016, Elsevier. B. Black phosphorus-loaded separable microneedles as responsive oxygen delivery carriers for wound healing; (a) schematic illustrations of wound healing using NIR-responsive separable MNs which encapsulate BP QDs and oxygen-carrying Hb; (b) scheme and characterization of the design of responsive separable MNs; (c) thermal images of MNs applied to the rat dorsal skin before and after 2 min NIR irradiation; (d) oxygen releasing conditions of different groups [109]. Copyright 2020, American Chemical Society.

coordinating the cellular and biochemical events during wound healing, making them promising candidates for skin substitutes in tissue repair.

3.2.4. Oxygen-releasing structures

Oxygen is critical to the survival, function, and fate of mammalian cells. Oxygen tension controls cellular behavior through metabolic programming, which in turn controls tissue regeneration. Oxygen also acts as an essential substance and a signaling molecule involved in metabolism. Oxygen deficiency can lead to tissue necrosis and programmed cell death [101]. In some chronic wounds, the local blood supply is limited, which inhibits the migration of inflammatory cells and the proliferation of endothelial cells in wounds [102]. However, simple oxygen diffusion has a very limited range of 100–200 μ m [103], so the introduction of oxygen releasing structures can overcome this limitation by providing precise and on-demand oxygen supply. A variety of oxygen-releasing biomaterials have been developed and applied in for regenerative medicine. Some organic systems include haemoglobin, photoautotrophical Synechococcus elongatus, or Cyanobacteria [104, 105], inorganic systems include CaO₂, MgO₂, Na₂CO₃, and H₂O₂ et al. [106,107] are applied.

Chavez et al. incorporated *Chlamydomonas reinhardtii* microalgae into an integra dermal regeneration template for oxygenic photosynthesis (Fig. 6A). This led to significantly increased VEGF expression of genetically engineered algae 2 weeks after implantation into mice. The dermal regeneration template led to an increased recruitment of vascular endothelial cells and alpha smooth actin-positive vessels [108]. Zhang et al. developed black phosphorus (BP)-loaded separable responsive microneedles (MNs) with oxygen-carrying and controllable oxygen-delivering abilities for wound healing (Fig. 6B). The MNs are loaded with BP quantum dots (BP QDs) and hemoglobin. Due to the photothermal effect of BP QDs and the reversible oxygen binding properties of hemoglobin, the local temperature of the skin will increase after near-infrared irradiation, resulting in responsive oxygen release. The MNs exhibited practical wound healing ability in treating the full-thickness cutaneous wounds of a type I diabetes rat model [109].

3.3. Intelligent skins

In addition to their basic function as wound dressings, current skin substitutes are designed to replace or enhance the function of natural skin [110]. Advanced skins substitutes have been developed that can detect, perceive, and respond to changes in temperature or stains, while wearable devices that can interact with users and the environment for health monitoring are also being widely studied.

3.3.1. Detection and perception

The skin contains a variety of sensory receptors distributed throughout each layer, forming a vast nerve network. These receptors are responsible for specific stimuli, and through the use of electronics, the intelligent skins can process the data and provide artificial sensation and other functions [111,112]. Currently, pressure detection in electronic skin is achieved through two main approaches: resistive and capacitive sensing. Zhou et al. have developed a highly sensitive, mechanically compliant, and autonomously self-healable ionic skin. This hydrogel-based ionic skin exhibits high pressure sensitivity up to 1 kPa, enabling it to detect gentle finger touches, human motion, and even small water droplets (mimicking rain) (Fig. 7A) [9]. Some animals, such as pit vipers, possess sensitive membranes that enable them to locate prey. Raffaele et al. have prepared a pectin film that mimics the sensing mechanism of pit membranes. This film holds a sensitivity of at least 10 mK and can be integrated as a layer in skin substitutes [113].

3.3.2. Stimuli-responsive

The stimuli-responsive skin can respond to complex changes in the external environment, such as strain response, microenvironment response, photothermal response, and more. A chameleon-inspired strain-accommodating skin was developed (Fig. 7B), which can change its color in response to thermal heating, mechanical stretching, and illumination, including ambient sunlight. This strain-accommodating skin embeds responsive materials within a scaffolding polymer and provides a general framework to guide the future design of artificial intelligent skins [10].

Diabetic ulcers have a distinctive physiological microenvironment, including an acidic pH, high levels of glucose, proteases, and matrix metalloproteinases. Zhu et al. developed a pH/glucose dual-responsive injectable hydrogels through the covalent crosslinking of imine bonds and phenylboronate esters. During in situ crosslinking, insulin and fibroblasts were incorporated into the hydrogels. The decrease in pH and increase in glucose levels lead to the release of insulin. The fibroblastincorporated hydrogels exhibited accelerated wound healing in diabetic wounds [114]. Wang et al. developed an NIR-responsive nanocomposite membrane by incorporating Cu₂S nanoparticles into biopolymer fibers via a modified electrospinning method. The membranes exhibited excellent and controllable photothermal performance under near-infrared irradiation, effectively inhibiting tumor growth in mice (>90 %). Moreover, the angiogenic effects of Cu^{2+} released from the scaffolds improved skin tissue regeneration in tumor-induced wounds [115].

3.3.3. Wearable products

Skin-like wearable devices are already being used for personal health monitoring, including the detection of glucose, uric acid, lactose, heart rate, blood pressure, ion levels, stress levels, strain, tactile input, temperature, and humidity [116,117]. These sensors are also being embedded into soft robots to enable intelligent interaction with users and the environment. The concentration of ions, small molecules, and proteins in perspiration can provide valuable health information. To enable *in situ* perspiration analysis, Javey and co-workers developed a flexible and wearable device that can measure and analyze perspiration metabolites, electrolytes, and skin temperature (Fig. 7C). An integrated sensing array was created by electrically decoupling the operating points of each sensor's interface. After strenuous exercise, dehydration was detected through sweat analysis [11].

4. Manufacturing methods

The physicochemical properties of skin substitutes and artificial skin are highly dependent on the materials and usage [118]. Traditional skin substitutes mainly include gauze, bandages dressings etc. They can absorb exudate and secretion from wound sites and protect wounds. However, they will dry out and cause secondary damage if prolonged used. Natural dermal substitutes, represented by "acellular substitutes", are prepared by removing cells from the epidermis and dermis, leaving intact ECM components such as collagen fibers and basement membranes to reduce rejection reactions. Integra® is commoditized acellular substitute consists of collagen crosslinked with glycosaminoglycan and function as a temporary epidermis [119]. Dermagraft contains human neonatal fibroblasts plated on biodegradable polylactic acid scaffolds to be more simulate natural skin [120]. These substitutes are more expensive. Modern approaches are applied in various clinical forms, such as hydrogels, films, sponges, and 3D skin scaffolds. These products can be prepared using methods like (in situ) 3D bioprinting, electrospinning, microfluidics, and spraying [121] (Table 1).

4.1. Hydrogel

The extracellular microenvironment of the skin is a highly hydrated network due to the presence of glycosaminoglycan chains, which make the structure of hydrogels highly similar to that of ECM. Polymer hydrogels provide an ideal moist environment for wound healing while protecting the wound. They have the advantage of providing a cooling effect and not adhering to the wound tissue. Polymer hydrogels can be



Fig. 7. Intelligent skin substitutes. A. Pressure Sensing electronic skin; (a) Schematic structure and (b) SEM image of the hydrogel; (c) the interfaces between the ionic conductive layers and the dielectric layer accumulate electric charge when a voltage is applied; photos of the self-healing hydrogel sensor attached to a (d) straight or (e) bent finger; (f) real-time capacitance signal recorded when the finger was bent cyclically [9]. Copyright 2017, Wiley. B. Chameleon-inspired strain-accommodating skin; (a) color change of chameleon; (b) photograph and schematic of the smart skin includes dimensions and chemical structure of polymers; (c) camouflaged "leaf" and positioned alongside real leaves before (left) and after (right) sunlight exposure for 10 min [10]. Copyright 2019, American Chemical Society. C. Health monitoring skin-like wearable devices; (a) photographs of the subject wearing a 'smart headband' and a 'smart wristband'; (b) data collection for each sensor from sweat sample [11]. Copyright 2016, Nature.

Table 1

Manufacturing methods for preparing skin substitutes.

Materials	Methods used	Active ingredient	Properties	Ref.
Cholesterol-modified hyaluronic acid micelles; Phenylboronic acid grafted alginate hydrogel	<i>In situ</i> injectable hydrogel	Antibiotic amikacin and anti-inflammatory drug naproxen	pH responsive; ROS responsive; antibacterial properties; accelerating bacterial infections wound healing	[123]
Chitosan-graft-aniline tetramer and dibenzaldehyde-terminated poly (ethylene glycol)	<i>In situ</i> injectable hydrogel	Exosomes	Regulation of macrophage polarization; enhance angiogenesis; diabetic wound healing	[124]
Dopamine-modified gelatin	<i>In situ</i> injectable hydrogel	Dopamine-modified gelatin@Ag nanoparticles	Injectable and self-healing performance; antioxidant activity; near-infrared (NIR) laser irradiation synergistic antibacterial behavior; accelerate wound healing	[125]
Gelatin/alginate/hyaluronic acid	3D bioprinting	Photoactive cationic conjugated poly (phenylene vinylene); laminin-derived peptide A5G81	Photodynamic therapy; cell adhesion, migration, and proliferation; anti-infection ability; wound repair properties	[131]
Plasma-derived fibrin matrix and fibrin scaffold	3D bioprinting	Human fibrinogen, primary human fibroblasts, and keratinocytes	Bilayered dermo-epidermal equivalents; structure similar to normal human skin	[133]
Platelet-rich plasma; alginate-gelatin	In situ extrusion bioprinting	Dermal fibroblasts and epidermal stem cells	Regulate the tube formation of vascular endothelial cells and macrophage polarization	[134]
Alginate and enzymatically cross-linkable proteins	Handheld skin printer	Dermal and epidermal cells	The capacity of depositing onto inclined and compliant wound surfaces that are subject to respiratory motion.	[135]
PCL and quaternized chitosan-graft- polyaniline	Electrospinning	Quaternized chitosan-graft-polyaniline	Antibacterial, anti-oxidant, and electroactive properties; full-thickness skin repair	[144]
PCL	Hand-held electrospinning	Aggregation-induced emission luminogens	Antibacterial activity; accelerated wound healing	[145]
Methacryloyl chondroitin sulfate	Microfluidics	VEGF	Wound healing applications	[148]
Alkylated chitosan/alginate microcapsules	Microfluidics	Polycationic surface of alkylated chitosan	Persistent antimicrobial activities hemostatic properties;	[149]
Alginate	Droplet microfluidics	Bubble-propelled nanomotors	Improve the healing effects of wounds in the type I diabetes rat models	[150]
Alginate	In situ spray-by-spray deposition	Tea tree oil microemulsion	Antimicrobial properties; Infected wounds	[153]

used as permanent or temporary dressings to support the regeneration and healing of injured epidermis or dermis. Developing advanced hydrogel dressings is an active research field aimed at improving skin regeneration and wound healing according to clinical needs, such as *in situ* injectable hydrogels and stimulus-responsive hydrogels [122].

The in situ injectable hydrogel can adapt to the exact shape of the defect and fit perfectly with the wound edge. Hu et al. developed a dualresponsive injectable hydrogel wherein the antibiotic amikacin and antiinflammatory drug naproxen were preloaded into the cholesterolmodified hyaluronic acid (HA-CHOL) micelles to prepare the phenylboronic acid grafted alginate hydrogel. The hydrogel showed dual responsiveness to low pH and high reactive oxygen species (ROS), and over 80 % amikacin and naproxen were released from the hydrogel at 24 h, effectively exhibiting antibacterial properties and accelerating bacterial infections wound healing (Fig. 8A) [123]. Wang et al. developed an exosome-loaded injectable hydrogel based on chitosan-graft-aniline tetramer (CS-AT) and dibenzaldehyde-terminated poly (ethylene glycol). The CS-AT hydrogel and exosomes work synergistically to induce local regulation of M2 macrophage polarization and enhance angiogenesis, resulting in improved diabetic wound healing in vivo (Fig. 8B) [124]. Zhang et al. developed dopamine-modified gelatin@Ag nanoparticles (Gel-DA@Ag NPs) in guar gum-based hydrogels for bacteria-derived wound infection and other tissue repair related to reactive oxygen species overexpression [125]. Conventional hydrogels are usually soft, weak, and have limited stretchability and adhesion [126]. Moreover, hydrogels are not stable under extreme conditions, and cannot maintain mechanical properties and electrical conductivity [127]. The toughness and self-repairing ability of hydrogels can be improved by using reversible covalent bonds in a double-network structure [128].

4.2. 3D bioprinting

3D bioprinting is a technique used for tissue engineering and woundhealing, which involves layer-by-layer deposition of biomaterials on a 3D controllable platform. This technique provides personalized shapes and tailored structures. To fabricate skin substitutes, many works have been done in the development of a suitable bioink, and using appropriate mechanisms and patterns based on inkjet, extrusion, laserassisted, and stereolithography [129]. The selection of appropriate materials and patterns depends on several factors, including printability, biocompatibility, degradation kinetics and by-products, and structural and mechanical properties [130]. For instance, Zhao et al. used a photoactive cationic conjugated poly (phenylene vinylene) (PPV) derivative as a photosensitizer and gelatin/alginate/hyaluronic acid as the ink to develop 3D printed skin substitutes patches. They further covalently modified the patches with laminin-derived peptide A5G81 to enhance cell adhesion, migration, and proliferation. The skin patch showed photodynamic therapy (PDT)-based anti-infection ability and in vitro and in vivo wound repair properties (Fig. 9A). [131].

It is relatively easy to incorporate a single cell type in skin substitutes, but due to limitations in current seeding methods, it is quite difficult to incorporate more than two cell types into one skin substitutes. 3D bioprinting shows potential for adding multiple cell lines to a skin substitute in a precise manner, resulting in a better resemblance to human skin [132]. Cubo et al. developed a human bilayered skin using bioinks containing human fibrinogen, primary human fibroblasts, and keratinocytes to imitate the dermis and epidermis structure (Fig. 9B). The regenerated human skin was then grafted onto the backs of immunodeficient mice, and histological analysis showed that it presented a structure similar to that of normal human skin [133].



Fig. 8. Injectable hydrogel as wound dressing. A. pH and ROS responsive injectable hydrogels encapsulating drug-loaded micelles; (a) schematic illustration for the formation of hydrogels; (b) the photograph of the hydrogel to pH and H_2O_2 variations; (c) the TEM images and (d) size distribution of HA-CHOL micelles; (e) the amikacin and (f) naproxen cumulative release of the hydrogel at different time points; the antibacterial ability for hydrogels on (g) CFU results and (h) live/dead staining of *S. aureus* (top) and *P. aeruginosa* (bottom) after treated with different hydrogels for 12 h; (i) schematic illustration for the mechanisms that anti-bacteria and wound healing [123]. Copyright 2020, Elsevier. B. Exosomes loaded self-healing injectable CS-AT-Exo hydrogel; (a) schematic description of the fabrication of *CS*-AT-Exo hydrogel and the application in the diabetic wound healing process; (b) representative images of diabetic wound healing in rats; (c) picrosirius red staining of collagen (yellow: type I collagen, green: type III collagen); (d) images of iNOS (red) and F4/80 (green) immunostaining showed accumulation of M2 macrophages at the wound bed on day 7 and day 14; (e) rate of the residual wound after day 3, day 7, and day 14 during the healing process; (f) quantification analysis of type I and type III collagen of wound area at day 14 [124]. Copyright 2022, Elsevier.

4.3. In-situ 3D printing and 4D printing

In situ 3D printing involves the direct printing of bio-ink to create or repair tissue or organs at defective sites in clinical [136]. Recent technological advancements have led to the development of handheld skin printers, which can be used to restore skin tissue *in situ* at the wound site. Zhao et al. utilized the extrusion strategy to fabricate 3D bioprinted constructs embedded with dermal fibroblasts and epidermal stem cells. The platelet-rich plasma (PRP) was integrated in the bioink and acted as

a source of patient-specific autologous growth factors. The bioprinting was utilized for *in-situ* extrusion bioprinting on full-thickness rat cutaneous defects (Fig. 9C) [134]. Hakimi et al. also developed a portable handheld extruded skin printer (<0.8 kg) capable of printing patches containing cells (Fig. 9D) [135]. These developments have made it possible to print on skin *in situ*, which is applicable for real-time treatment of burns, scalds, wars, and other emergencies [137]. However, the safety and sterility of the handheld 3D bioprinting technology need to be rigorously tested before it can be used clinically.



Fig. 9. 3D printing skin substitutes patch. A. Photodynamic therapy 3D printing skin substitutes patch; (a) schematic of the 3D printed antibacterial skin patches with high cell affinity for wound repair; (b) preparation scheme of the Gel/Alg/HA/PPV ink; FSEM images of the Ca^{2+} -crosslinked (c) Gel/Alg/HA and (d) Gel/Alg/HA/PPV ink; scale bar: 200 µm; (e) ROS-production ability of the skin patches; pattern diagram, digital and fluorescence images of 3D printed structures with (f) 30°, (g) 60° and (h) 90° pore geometry [131]. Copyright 2022, Royal Society of Chemistry. B. Bilayered skin containing primary human fibroblasts, and keratinocytes; (a) scheme of the bioprinting process; (b) picture of the head showing the tubes connection, (i) hFBs, (ii) plasma, (iii) CaCl₂, and (iv)hKCs; (c) visual appearance of the grafted human skin on the immunodeficient mice; H/E staining of the (d)regenerated human skin and (e) normal human skin [133]. Copyright 2016, IOP Science. C. Schematic illustration of PRP-integrated AG composite hydrogel bioinks. DFs and ESCs-laded 3D bioprinted constructs were fabricated according to a single-layer or double-layered structure [134]. Copyright 2022, Elsevier. D. Handheld skin printer; (a) schematic diagram illustrating working principle of handheld bioprinter; (b) rendered image of handheld bioprinter: ① handle, ② stepper motor, pulley and drive mechanism, ③ on-board syringe pump modules, ④ bioink, ⑤ cross-linker solution, ⑥ syringe holder, ⑦ 3D printed microfluidic cartridge; (c) photograph of 3D printed microfluidic cartridge, scale bar 10 mm; (d) schematic side-view image showing sheet formation between moving microfluidic cartridge and deposition surface or wound [135]. Copyright 2018, Royal Society of Chemistry.

4D printing is defined as multi-material printing with the ability to change over time, adding a temporal dimension to the existing 3D spatial dimension [138]. When exposed to predetermined stimuli such as light, heat, or magnetism, or based on identification of bio information such as body temperature, stress or sweat, the shape, property, or function of the material can change [139,140]. This technology can be used for the manufacture of wearable medical implants that are printed directly on the human body, allowing for clinical diagnosis and assistance in wound repair or tissue regeneration. 4D printing is an emerging area of research in biomedicine, with promising potential for tissue engineering techniques and biomedical devices [141].

4.4. Electrospinning

Electrospinning has recently attracted significant attention in the tissue engineering and healthcare fields. The electrospinning technique can be used to produce scaffolds/patches with controllable 3D fibrous structures and porosity. Materials such as fibrinogen, collagen, gelatin, chitosan, PU, poly (lactic acid), and PCL, either alone or in combination, have been used in electrospinning. Furthermore, drugs and other biologically active molecules can be added to the nanofibers to obtain a controlled release system with multifunctional or antibacterial activity [142]. The electrospinning device mainly consists of four parts: a high-voltage generator, fluid driver, spinneret, and collection device (Fig. 10A) [143]. The morphology and size of nanofibers can be regulated by adjusting the process parameters and raw materials, and



Fig. 10. Electrospinning and its application. A. Schematic of a conventional electrospinning device [146]. Copyright 2022, MDPI. B. Electrospinning PCL/QCSP nanofiber membrane with anti-bacteria, anti-oxidant, stretchability and electroactivity [144]. Copyright 2020, Elsevier. C. *in situ* deposited AIE nanofibrous electrospinning. (a) Schematic illustration of electrospinning AIEgen-incorporated antibacterial dressing. (b) Morphological changes of *S. aureus* and MRSA imaged by SEM [145]. Copyright 2022, Wiley.

nanofibers with suitable pore size, porosity, and ECM-like structure can be obtained. He et al. developed an electroactive nanofibrous membrane using electrospinning PCL and quaternized chitosan-graft-polyaniline (QCSP) polymer solutions. The nanofibrous wound dressings exhibited antibacterial, anti-oxidant, and electroactive properties. In a mouse full-thickness wound defect model, the wound dressing exhibited higher collagen deposition, granulation tissue thickness, and more angiogenesis compared to a commercial dressing (Tegaderm[™] Film) (Fig. 10B) [144]. Hand-held electrospinning devices have been designed to miniaturize electrospinning equipment for greater portability, convenience, and comfort of use. *In situ* electrospinning offers the potential to provide customized wound dressings that better cover the wound, reduce edema or pain, and promote faster healing [120]. Dong et al. have used a handheld electrospinning device to electrospin an aggregation-induced emission luminogens (AIEgens)-incorporated PCL solution into a nanofibrous dressing (Fig. 10C). This AIE nanofibrous dressing exhibited



Fig. 11. Applications of microfluidics. A. Schematic diagram of the basic principle of microfluidics [151]. Copyright 2014, Nature. B. Self-Assembled Alkylated Chitosan Microcapsules (SAAMs); (a) schematic illustration of the preparation of the SAAMs and the self-assembly of ACS and ALG; (b) design of SAAMs from all-aqueous microfluidics and their biomedical functions in the entire wound healing process [149]. Copyright 2022, American Chemical Society. C. Schematic illustrations of the generation process of PSNs and the porous microparticles and their applications [150]. Copyright 2022, Wiley.

excellent antibacterial activity and significantly accelerated wound healing. Hand-held *in-situ* deposited electrospinning could offer personalized therapies for emergency wounds [145].

4.5. Microfluidics

Microfluidics is a technique that employs channels ranging from tens to hundreds of microns to process small amounts of fluid, as shown in Fig. 11A. This method enables better monodispersity and higher reaction efficiency, along with continuous production and reduced byproduct formation. Therefore, microfluidics can achieve the production of materials that are difficult to complete using traditional processing methods [147]. Microfluidics is considered one of the most promising versatile technologies and for manufacturing high-performance functional dressings. It allows for the preparation of spherical (porous) particles, microspheres, microcapsules, microfibers, and nanofibers in common formats.

Zhang et al. have developed drug-loaded methacryloyl chondroitin sulfate (CSMA) microspheres to promote wound healing. The CSMA droplets were generated from microfluidic electrospray and polymerized using ultraviolet light. The drug-loaded CSMA microspheres exhibited excellent biocompatibility and wound healing properties [148]. Similarly, Gao et al. fabricated self-assembled, multifunctional alkylated chitosan/alginate microcapsules (SAAMs) using microfluidics. The polycationic surface and internal porous dextran-rich cores in SAAMs provide excellent hemostatic properties. The size-tailored soft microcapsules can realize time-sequential functions for tissue repair (Fig. 11B) [149]. Liu et al. developed novel droplet microfluidics for continuous and direct generation of porous particles by introducing bubble-propelled nanomotors into the system (Fig. 11C). The obtained platinum-coated silica nanoparticles (PSNs) porous microparticles can serve as microcarriers for 3D cell culture. Due to the existence of oxygen in these microparticles, they can be used to improve the healing effects of wounds in type I diabetes rat models [150].

4.6. Spray

Sprays are widely used in clinical practice due to their versatility and applicability in the treatment of both acute and chronic wounds, particularly those that are large or have irregular surfaces [152]. They can be categorized into two types based on their composition: acellular skin sprays and cellular skin sprays. Acellular skin sprays generally consist of hydrogels that act as a protective dressing to prevent wound infection and drying. For example, Catanzano et al. developed an in-situ forming alginate hydrogel for delivering tea tree oil microemulsion. This hydrogel were prepared via a spray-by-spray deposition method and exhibits antimicrobial properties against Escherichia coli and has potential for treating infected wounds [153]. On the other hand, cell spray autografting is an innovative technique that employs sprays as a delivery system for fibroblasts and keratinocytes. Cells are obtained from a biopsy of undamaged skin from the patient and can be sprayed directly onto the wound or cultured in vitro to obtain sufficient cell numbers before being sprayed. However, cell spray technology is not yet widely applicable or available for general use, as it is still under clinical evaluation [154].

5. Clinical/commercial progress and design requirements of skin substitutes

5.1. Clinical research of skin substitutes

In the last few years, various artificial skin or skin substitutes have shifted from cell or animal experiments to clinical trials to explore clinical applications and improve treatment efficacy. To show the recent clinical research advancements in this field, a search was conducted on the *ClinicalTrials.gov* website from January 2015 to January 2023, involving clinical studies on wound dressings and skin substitutes. The search terms used were "skin substitutes", "artificial skin", and "wound dressing". A preliminary screening yielded 400 results, including cell-containing, acellular, natural, and synthetic polymer-based substitutes (Table 2). Many decellularized skin substitutes and cellular skin substitutes have been adopted in clinical practice, primarily aimed at diabetic foot ulcers and burn wounds. CellMist System conducted by RenovaCare, is a stem cell-based skin substitutes treatment for burn patients (NCT04890574). Altrazeal (NCT05424354) aims to evaluate the effectiveness and safety of a composite material of hydroxyapatite and artificial dermis-based skin substitute for the treatment of chronic wounds. These new skin substitutes products are expected to improve treatment efficacy and comfort, and expand the applications of skin substitutes in the medical field.

5.2. Current research of commercial products

In the 1970s, Burke and Yannas developed the first biodegradable scaffold using collagen derived from cattle and pigs, and poly-saccharides derived from shark cartilage. The scaffold was inoculated with fibroblasts and named Apligraf®. Apligraf® was subsequently approved for sale by the US FDA for the treatment of diabetic foot ulcers and venous leg ulcers [155].

There are many artificial skin substitutes available on the market (Table 3). Acellular substitutes are typically derived from cadaveric or animal dermis, and the use of acellular cadaveric dermal matrix can preserve the ECM while reducing the risk of rejection and inflammation [34]. Examples of commercial allografts from cadaveric dermis include Allover[™] and Graf[®]. Allover[™] (LifeCell Corp., Branchburg, N.J.), extracted from acellular human dermis, is commonly used for burns and soft tissue defect repairs, but it may lead to scar formation due to skin contraction. Matriderm[®] (Dr. Suwelack Skin & Health Care AG), an animal-derived acellular skin substitute made from bovine type I collagen and elastin, is used for full-thickness burns and chronic wounds.

There are many other synthetic polymer-based skin substitutes available on the market, such as PLGA, polycaprolactone, polylactic acid, and hyaluronan. Laserskin® (Fidia Advanced Biopolymers Ltd, Abano Terme, Italy) is made from a benzyl-esterified hyaluronic acid derivative and is cultured with autologous keratinocytes for the treatment of burns or chronic full-thickness ulcers. Integra® (Ethicon, Inc., Somerville, N.J.) is perhaps the most established and promising artificial dermal template and is used in full-thickness burns as well as reconstruction of scar contractures. Integra® consists of a matrix of purified collagen from bovine tendon crosslinked with glycosaminoglycan obtained from shark cartilage, and may be supplied with a removable silicone layer that functions as a temporary epidermis [119]. However, there are risks associated with the use of bovine collagen-derived skin substitutes. Recently, many products have been developed and used in therapeutic applications to mimic natural skin structures and their microenvironmental properties.

5.3. The design requirements of skin substitutes

Skin substitutes play an indispensable role in saving patients' lives and improving their health with the continuous growth of medical demand. When designing skin substitutes, many factors should be taken into consideration, such as suitable mechanical properties, suitable skin anisotropy, suitable porosity and permeability, low rejection reaction and meet the comply with regulations.

5.3.1. Structural characteristics

Skin is anisotropic, viscoelastic, and relatively incompressible tissue with low modulus, self-stiffness, high toughness and tear resistance [156]. The Young's modulus of the human skin ranges from 4.5 to 8 kPa [157]. Therefore, skin substitutes should have rheology comparable to

Table 2

Clinical Trial of artificial skin or skin substitutes.

Product	NCT Number	Title	Conditions	Phase	State
Fitostimoline® hydrogel	NCT05661474	Fitostimoline® Hydrogel Versus Saline Gauze Dressing in Diabetic Foot Ulcers	Diabetic Foot	IV	Completed
Altrazeal Transforming Powder Dressing	NCT05424354	Acute Partial Thickness Burn Study Comparing Transforming Powder Dressing to Standard of Care Dressing	Wounds and Injuries	IV	Active, not recruiting
Novadress, Mepilex Ag, and Xeroform Occlusive dressings	NCT05499104	A Trial Comparing a Cellulose Dressing to Two Standard of Care Dressings in Treating Split Thickness Donor Sites in Burn and Wound Patients.	Donor Sites; Wound Drainage	III	Completed
hyaluronic acid (Gengiegel 0.2 % oral gel)	NCT04390100	Palatal Wound Healing Evaluation After Application of Platelet Rich Fibrin Versus 0.2 % Hyaluronic Acid Dressings	Free Gingival Graft	IV	Completed
Platelet-lysate loaded sustained release thermo-gelling formulation	NCT05671250	Bioactive Smart Dressings for Diabetic Foot Ulcers: Randomized Controlled Trial	Diabetic Foot Ulcer	II	Completed
Sulfadiazine Silver	NCT03592498	Study to Evaluate the Use of Tilapia Skin (Oreochromis Niloticus), in the Treatment of Burn Wounds	Burns	Π	Completed
Nile Tilapia Fish Skin	NCT04202289	Use of Nile Tilapia Fish Skin as a Xenograft for Burn Treatment: Phase III Study	Burns	III	Completed
Xeno-Skin™	NCT03695939	Evaluation of Safety, Tolerability and Efficacy of Xeno-Skin® for Temporary Closure of Severe Burn Wounds	Deep Full- thickness Burn	I/II	Completed
CellMist TM	NCT04890574	CellMist TM Autologous Cells to Treat Deep Second-Degree Burns	Second degree burns	Ι	Active, not recruiting
DenovoSkin	NCT03227146	Study With an Autologous Dermo-epidermal Skin Substitute for the Treatment of Burns in Adults and Adolescents	Burns	II/III	Active, not recruiting

the native skin and can withstand shear forces and mechanical tensions that are applied during the substitute placement. The arrangement of collagen in ECM not only supports the mechanical strength and elasticity of the skin, but also plays a key role in skin anisotropy [158]. Therefore, it is important to consider mimicking the arrangement of collagen tissue in order to reduce scarring when designing skin substitutes [159].

The shape and structure of a skin substitute scaffold, including pore size and interconnectivity, can have a significant impact on cell function and the tissue healing process. Micron-sized pores can enhance cellular attachment and proliferation, while pores of around $100 \,\mu\text{m}$ or more can improve cell and capillary ingrowth [160]. Biomaterials with gradient or graded pores can further accelerate tissue reconstruction [161]. An ideal skin substitute needs to transmit water in a similar way to the normal skin, in order to avoid fluid loss or accumulation [162]. An appropriate water absorption ratio will enhance the biological activity of skin equivalents and contributes to the maintenance of 3D structure [163]. Scaffolds with high water absorption ratio are appropriate in excess exudates wounds such as the full-thickness wounds while scaffolds with a lower water absorption ratio are more suitable for partial-thickness wounds [164].

5.3.2. Performance and regulatory requirements

Implants are recognized by the host as "foreign bodies," triggering a complex signaling cascade known as the foreign body response (FBR) during wound healing [38]. The FBR involves four stages: protein adsorption, acute inflammation, chronic inflammation, and collagen encapsulation. Skin grafts typically trigger an acute inflammatory reaction in the host within a few hours of transplantation, causing inflammatory cells such as neutrophils and monocytes to migrate to the implantation site, release proinflammatory cytokines, recruit and activate macrophages, and trigger a cascade immune response [39]. The fibroblasts that migrate to the wound continually deposit collagen fibers around the implants, resulting in a highly fibrotic structure in the outer layer of the implant, which is the phrase of chronic inflammation [40]. While FBR is the natural protective mechanism of the body, both chronic inflammation and acute inflammation will bring strong discomfort to patients, greatly affecting the function of implanted materials. Therefore, immune regulation design is necessary to reduce the inflammatory reaction of skin substitutes in order to alleviate immune rejection and improve the biocompatibility of skin substitutes.

The International Organization of Standardization (ISO) has defined several tests to ensure the biocompatibility of skin substitutes (ISO 10 993; biological evaluation of medical devices), based on the type and length of body contact. Tests such as cytotoxicity, sensitization, irritation, systemic (acute) toxicity, subacute, and subchronic toxicity, genotoxicity, hemocompatibility, chronic toxicity, carcinogenicity, reproductive (developmental) toxicity, biodegradation—identification and toxicokinetic studies, physicochemical, morphological, and topographical characterization, and immunotoxicology should be evaluated [3]. The US Food and Drug Administration (FDA) approves a range of natural and synthetic biomaterials, such as collagen, silk, Pluronic F-127, and poly (ε -caprolactone) (PCL) used in biocompatible scaffold either individually or collectively. Clinical trials for the development of cell-containing skin substitutes should comply with Good Manufacturing Practice (GMP) standards, and quality items such as biological tests of cell components, microbiological examination, and environmental control are strictly controlled [165].

6. Conclusions

Skin substitutes have revolutionized wound management by shortening time to wound closure and decreasing wound-associated morbidities, ultimately resulting in overall cost reduction compared with standard-of-care therapy. Although skin substitutes may have high initial costs, their benefits are significant. These advancements in skin substitutes serve as a model for regenerative medicine, cosmetic, and pharmaceutical applications.

In addition to the development of new biomaterials and the use of more human skin phenotypes or cytokines, bioprinting *in situ* has also emerged as a promising approach for skin substitute fabrication. Despite the lack of a perfect skin substitute currently available, rapid advancements in our understanding of skin development and wound repair, as well as progress in stem cell and tissue bioengineering, provide hope that the development of such a product is a feasible goal in the future. Future research on skin substitutes will likely focus on simulating the functional and aesthetic characteristics of natural skin, including the introduction of sensory perception, skin pigments, and appendages, which will enable skin substitutes to be customized in terms of performance and appearance for more personalized treatment.

However, there are still many challenges that need to be addressed. Natural biomaterials have biocompatibility and similarity to the native skin while suffer from low mechanical properties and fast degradation. Synthetic biomaterials have controlled good mechanical and physical properties but with less ideal cell interaction and overlong degradation rate. Reasonable combine the natural and synthetic biomaterials can design the potential candidate for a successful and economical skin Commercially available skin substitutes used in the treatment of wounds.

Commercial Product	corporation	Composition	Cell type	Skin structure	Indications	Defect	Ref.
Integra®	Ethicon, Inc., Somerville, N.J.	Collagen from bovine tendon crosslinked with glycosaminoglycan obtained from shark cartilage	/	Dermal	Extensive burns and limited autotransplant donor area	Allergic to cow collagen, chondroitin, or silicone material.	[166]
Matriderm®	Dr. Suwelack Skin & Health Care AG	Type I collagen from bovine and elastin	/	Dermal	Full-thickness burns and chronic wounds	Scar formation due to skin contraction	[166]
Dermagraft	Smith and Nephew, Largo, FL	Human neonatal fibroblasts plated on biodegradable polylactic acid scaffolds	human dermal fibroblast	Dermal	Restore the dermal bed in a diabetic foot ulcer	Complications, such as infection	[120]
Biobrane®	Bertec Pharmaceuticals, Morgantown, WV, USA	Chemically adhering collagen to nylon fabric embedded in a silicone film	/	Dermo- epidermal	Partial and full thickness burns in children	Not for contaminate wounds	[31]
BioSeed®	BioTissue Technologies GmbH, Freiburg, Germany	Autologous keratinocytes cultured in fibrin sealant	Keratinocyte	Epidermal	Chronic leg ulcer	/	[167]
Laserskin®	Fidia Advanced Biopolymers Ltd, Abano Terme, Italy	Benzyl-esterified hyaluronic acid derivative with Keratinocyte on the surface	Keratinocyte	Epidermal	Burns or chronic full-thickness ulcers	Late graft rejection	[168]
AWBAT-D TM	Aubrey Inc., Carlsbad, CA, USA	A thin silicone membrane attached to nylon fabric and porcine collagen	/	Epidermal	Transplantation donor site	Non-adhesive and not elastic	[169]
Allover TM	LifeCell Corp., Branchburg, N.J.	Acellular human dermis	/	Dermal	Burns and other wounds; repair of soft tissue defects	Infection, seroma, dehiscence, hypersensitive, Allergic or other immune response	[170]
NovoSorb	BTM; PolyNovo Ltd, Port Melbourne, Victoria, Australia	Biodegradable polyurethane foam	/	Dermal	Reconstruction of deep burn and soft tissue defect	Potential failure to integrate especially in cases of borderline vascularity or infection	[119]
TransCyte®	Advanced Tissue Sciences, La Jolla, Calif	The scaffold made of type I collagen from porcine and human neonatal fibroblasts cultured on the scaffold coated with bioabsorbable polvglactin.	extracellular matrix of allogeneic human dermal fibroblasts	Dermo- epidermal	partial thickness burns	Infectious issues however, prohibitive in children.	[171]
Hyalomatrix®	Anika Therapeutics S. r.l., Abano Terme, Padova, Italy	Bilayer of an esterified hyaluronan scaffold beneath a silicone membrane	/	Dermal	partial thickness and full thickness burns	The silicone layer had to be removed as it peeled off	[172]
Epigard®	/	Polytetrafluorethylene, polyurethane	/	Epidermal	Preparation of wound bed before skin transplantation	/	[173]
MySkin™ Apligraf®	/ Organogenesis, Inc., Canton, MA	Silicone Bovine type I collagen	(keratinocytes) Fibroblasts, keratinocytes	Epidermal Dermo- epidermal	Diabetic foot ulcers Partial- and full- thickness burns, chronic ulcers	/ Alloantigens rejection	[174] [175]
MyDerm™ Cellular	Universiti Kebangsaan Malaysia	Fibrin	Fibroblasts, keratinocytes	Dermo- epidermal	Full-thickness wounds	Soft and fragile	[176]

substitute. Currently, most artificial skin substitutes are still in the early stages of development. In order to bring these substitutes to clinical application as soon as possible, the materials, cells, and equipment used in research must meet corresponding regulatory, clinical, and aseptic requirements. Furthermore, the cost of producing these substitutes remains a significant obstacle that needs to be overcome. To achieve this, it is essential for clinicians to collaborate with scientists to develop better skin substitutes and make appropriate improvements to existing products.

Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Data availability

Data will be made available on request.

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