

Review



Recent Advances in Toll Like Receptor-Targeting Glycoconjugate Vaccines

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Abstract: Many malignant cell surface carbohydrates resulting from abnormal glycosylation patterns of certain diseases can serve as antigens for the development of vaccines against these diseases. However, carbohydrate antigens are usually poorly immunogenic by themselves, thus they need to be covalently coupled with immunologically active carrier molecules to be functional. The most well established and commonly used carriers are proteins. In recent years, the use of toll-like receptor (TLR) ligands to formulate glycoconjugate vaccines has gained significant attention because TLR ligands can serve not only as carrier molecules but also as built-in adjuvants to form fully synthetic and self-adjuvanting conjugate vaccines, which have several advantages over carbohydrate-protein conjugates and formulated mixtures with external adjuvants. This article reviews recent progresses in the development of conjugate vaccines based on TLR ligands. Two major classes of TLR ligands, lipopeptides and lipid A derivatives will be covered with more focus on monophosohoryl lipid A (MPLA) and related analogs, which are TLR4 ligands demonstrated to be able to provoke T cell-dependent, adaptive immune responses. Corresponding conjugate vaccines have shown promising application potentials to multiple diseases including cancer.

Keywords: conjugate vaccine; carrier molecule; adjuvant; toll-like receptor ligand; lipopeptide; monophosphoryl lipid A

1. Introduction

Cells are covered by a dense glycocalyx that is mainly comprised of glycoproteins and glycolipids. The carbohydrate motifs, carrying signaling information, are involved in cell recognition and responses. Each cell uses these signals to mediate communication, proliferation, survival and various other cellular activities [1–3]. The human immune system also relies on the recognition of these signaling molecules to differentiate invasive cells from native ones. When presented to the immune system in the proper ways, surface carbohydrates of foreign cells can serve as antigens to induce immune responses that eradicate the invasive cells. Thus, cell surface carbohydrates are ideal candidate antigens for vaccine development. In this context, however, a major caveat is that carbohydrate antigens are typically weakly immunogenic [4–6]. In addition, unlike protein antigens, which usually elicit immune responses via T cell-dependent pathways [7], carbohydrate antigens alone can only induce innate immune responses and fail to induce T cell-dependent immunity [4,8]. Thus, an ultimate goal of carbohydrate-based vaccine development is to achieve optimized presentation of carbohydrate antigens and switch of innate immunity against them to adaptive immunity. Covalent coupling of carbohydrates to carrier molecules or adjuvants, which are proteins, lipids or other molecules having immunological stimulating activities, to form functional glycoconjugate vaccines has proved to be a solution to the problem. For example, conjugation with an adjuvant can alter the immune responses

to carbohydrates into T cell-dependent immunities that can have improved therapeutic efficacy and lasting immunological memory [9–11].

1.1. Conjugate Vaccines

The first generation of vaccines were attenuated live viruses and bacteria, which were often either insufficient or still infectious/inflammatory due to inherit difficulties in the attenuation process. Vaccine formulations consisting of purified ingredients, including antigens and adjuvants, have provided a better solution since these vaccines can induce more specific immune responses and reduce undesired activity. Accordingly, many external adjuvants, including mineral salts, liposome emulsions, Freund's adjuvant, protein toxins or small molecules such as toll-like receptor (TLR) ligands, have been developed to probe and augment the activities of experimental and clinical vaccines. Although various adjuvants may have different mechanisms of action, their general objective aims at increasing the uptake of target antigens by antigen presenting cells (APCs), such as dendritic cells (DCs) and macrophages, activating APCs, and thereby triggering immunoregulatory cytokine production and a cascade of immune responses including T cell recruitment. However, many adjuvants are derived from bacteria/virus and may bring about side effects and toxicity similar to that of attenuated live cell vaccines.

An alternative approach is to covalently link antigens to an immunostimulating carrier molecule or adjuvant to construct conjugate vaccines that have proved to have advantages over heterogeneous antigen/adjuvant mixtures [12,13]. First, by incorporating an immunostimulating moiety, the resultant conjugate vaccines become more immunogenic. Second, conjugate vaccines have relatively defined compositions. Third, the toxicity of carrier molecules or adjuvants derived from natural sources may be reduced upon conjugation with large hydrophilic carbohydrate antigens.

Originally proteins were used as carrier molecules in conjugate vaccine development. In 1929, Avery et al. conjugated a polysaccharide to a protein carrier to create the first conjugate vaccine with enhanced immunogenicity [14–16]. Since then, numerous bacterium-derived proteins, such as tetanus toxoid (TT), diphtheria toxoid (DT) and meningococcus B outer membrane protein complex (OMPC), and other proteins, such as keyhole limpet hemocyanin (KLH), have been used to formulate carbohydrate-based conjugate vaccines [17,18]. These protein carriers have witnessed great success. For example, a series of glycoconjugate vaccines against various infectious diseases, such as H. influenzae type B, Neisseria meningitidis and pneumonia, have been licensed for clinical use [19] and several anticancer glycoconjugates vaccines have entered clinical trials. Mechanistic studies revealed that T cells can recognize the sugar epitopes of glycoconjugate vaccines in the context of major histocompatibility complex (MHC), allowing the binding of sugar epitopes to MHC and activation of the adaptive immune system [20]. However, most protein carriers are very immunogenic themselves, thereby complicating the evaluation of immunogenicity against the antigen. In addition, conjugation sites and equivalents of carbohydrate antigen to carrier protein are random and uncontrollable, resulting in inconsistent efficacy from batch to batch. This has led to the exploration of conjugate vaccines having small molecule carriers or adjuvants, such as ligands to specific receptors like TLRs, mannose receptors and lectin receptors on dendritic cells (DCs) and other APCs.

1.2. Toll Like Receptors

The TLR family is a class of pattern recognition receptors (PRRs) of mammalian species, which are considered the bridge between innate and adaptive immune responses [21]. Facing the daunting task of fending off ever evolving microbes with tremendous molecular diversity and heterogeneity, mammalian species have developed the PRR-centered pattern-recognition strategy to detect the conserved molecular patterns of characteristic microbes. As the best representative of PRR functions, TLRs detect pathogen-associated molecular patterns (PAMPs) of invading cells and trigger the first level of defense, the innate immunity. TLRs on dendritic cells are also involved in the initiation of adaptive immune responses and play a key role in bridging innate and adaptive immunities [21].

Thus far, ten human TLRs and two TLR signaling pathways have been identified. The predominant myeloid differentiation primary response 88 (MyD88)-dependent pathway is used by TLR1, 2 and 4–9 [22], while the MyD88-independent pathway is only used by TLR3 and 4 [23,24]. TLR1, 2, 4, 5 and 6 are expressed on the cell surface whereas TLR3, 4, 7, 8 and 9 are in endosomal compartments. Some of them form complexes when binding to ligands to be activated. TLR2 can form complexes with either TLR1 or TLR6, and TLR4 associates with myeloid differentiation factor 2 (MD2) [25] after being activated by lipopolysaccharide (LPS) ligand [26]. Each of these TLRs or their complexes recognize and are activated by multiple molecular patterns characteristic of bacteria or viruses. For example, TLR1 can be activated by multiple triacyl lipopeptides; TLR2 recognizes various glycolipids, lipopeptides and lipoteichoic acid (LTA); TLR3 recognizes double strand RNA; TLR4 can be activated by multiple small organic molecules and single strand RNAs.

The ligands of all ten human TLRs have been identified and extensively examined, paving the path for their applications to immunological studies and vaccine development. The well-defined activation mechanisms and signaling pathways of TLRs represent a major advantage of TLR ligands over proteins as carrier molecules for formulating glycoconjugate vaccines as this allows for rational design and precise structural modification towards optimized immunostimulating property. As a result, many TLR ligands, such as tripalmitoylated/dipalmitoylated cysteine (Pam₃Cys/Pam₂Cys), lipid A analogues, recombinant flagellin and imidazoquinoline analogues, have been covalently conjugated with antigens and evaluated as vaccine carriers. In fact, the ligands of all 9 TLRs, except for TLR10 which is believed to be a negative regulator of TLR signaling [27], have been probed as conjugate vaccine carriers/adjuvants. It has been demonstrated that the defined structures of conjugate vaccines with TLR ligands as carrier molecules have facilitated structure-activity relationship studies, which is critical for tuning and optimizing the structure of carrier molecules. In addition, the on-target delivery of antigens to dendritic cells can further enhance their uptake and avoid adjuvant overuse and thereby minimize the endotoxicity.

Although numerous TLR ligands have been identified as immunostimulants, some of which were discovered even long before TLRs, only a handful of them have been successfully applied to conjugate vaccine development. Some ligands, such as lipoteichoic acids, are limited by the accessibility due to their complex structures. Some ligands, such as small heterocycles, are excluded because the target TLRs locate in cell compartments. Others, such as RNAs, although commercially available, are limited by the intrinsically poor drug properties, including difficult characterization and analysis after conjugation. Two classes of TLR ligands, lipopeptides and lipooligosaccharides, stand out from the rest because of their outstanding potential as built-in adjuvants for carbohydrate-based conjugate vaccines. Aside from these two major classes of ILR7/8) and unmethylated cytidine-phosphate-guanosine (CpG) oligodeoxynucleotides (agonists of TLR9) have also been studied as built-in vaccine adjuvants. Most of these compounds are, however, linked to protein or peptide antigens and have been reviewed elsewhere [26,28], and conjugation of these ligands with carbohydrate antigen has not been reported.

This mini-review will only cover TLR ligands employed in conjugating with carbohydrate antigens to formulate glycoconjugate vaccines, with an emphasis on the optimization of monophosphoryl lipid A (MPLA) as a vaccine carrier. Moreover, we choose to limit its scope to carbohydrate-TLR ligand conjugates, TLR ligand conjugates with peptide or protein antigens are not discussed either.

2. Lipopeptides as Carrier Molecules or Adjuvants for Vaccine Development

Lipopeptides as agonists of TLR1, 2, 6 and 10 were the first and most studied TLR ligands applied to glycoconjugate vaccine development. Pam₃Cys and its tetralysine analog Pam₃CSK₄ were demonstrated to activate TLR1/TLR2 heterodimers [29], and Pam₂Cys and Pam₂CSK₄ were shown to be potent agonists of TLR2/TLR6 heterodimers [30]. Other ligands of TLR2 or TLR2/TLR6 heterodimers include lipoaminoacids and macrophage-activating lipopeptide 2 (MALP-2). All of

these lipopeptides and lipoaminoacids have been exploited as built-in adjuvants to conjugate with carbohydrate or glycopeptide antigens for conjugate vaccine development.

Pam₂Cys, introduced by Hopp even before the discovery of TLRs [31], was the first lipopeptide used for conjugate vaccine development, while the report by Jung et al. [32] had described both Pam₃Cys and Pam₂Cys. Since then, Pam₃Cys, Pam₂Cys, and their derivatives have become the most studied lipopeptide vaccine carrier because of their defined structures and commercial availability. In 1994, tumor-associated carbohydrate antigen (TACA) Tn was covalently conjugated to Pam₃Cys by the Toyokuni group to construct dimeric and trimeric vaccine 6 (Scheme 1) [33,34]. First, glycopeptide 3 containing three Tn antigens was assembled by standard peptide synthesis method, and its terminal acid was then converted into an active ester to be linked to Pam₃Cys 5 with an amide bond in a 57% yield. A robust immune response against this conjugate was observed without an external adjuvant. IgG antibody was observed, although in low concentration, indicating the recruitment of T cells and initiation of adaptive immune response. This was the first experimental evidence and proof-of-concept for the self-adjuvanting property of glycoconjugate vaccines with TLR ligands as carrier molecules. Inspired by this research, Danishefsky and coworkers synthesized blood type antigen Le^y, a hexasaccharide, and linked it to the same carrier to obtain conjugate 7 (Scheme 1), which also elicited strong immunoglobulin M (IgM) type of immune response [35,36]. Using the same strategy, the Danishefsky group synthesized a pentavalent vaccine, with each of Tn, sTn, TF, Le^Y and Globo H antigen conjugated to Pam₃Cys [37]. However, no immunological data was reported for this amazingly complex glycoconjugate.



Scheme 1. Synthesis of trimeric glycopeptide conjugate vaccines with Pam₃Cys as the carrier molecule.

Unfortunately, none of the above glycoconjugates of Pam₃Cys elicited robust IgG antibody response, which is considered the character of adaptive immunity. To achieve immunity class switch and induce IgG antibody production, the Boons group designed and synthesized a three-component conjugate vaccine **14** (Scheme 2) [38]. In addition to the TLR1/2 ligand Pam₃Cys as built-in adjuvant and Tn antigen as the B cell epitope, a helper T cell epitope derived from *Neisseria meningitis* was also incorporated in the conjugates as the third component. A helper T cell epitope was believed to activate helper T cells and achieve class switch of antibodies [39]. As shown in Scheme 2, T-helper epitope

peptide **8** was coupled with Pam₂Cys **9** on a solid support. Conversion of Pam₂Cys to Pam₃Cys was performed after coupling because the low reactivity of Pam₃Cys, due to its high hydrophobicity, with peptides. After introduction of the third lipid chain and dissociation from the beads, Tn antigen **13** with terminal amine was linked to lipopeptide **12** to complete the synthesis. Although only low to moderate IgG antibody titer was elicited in the subsequent immunological studies of **14** in mice, this is the first proof-of-concept that incorporation of T-helper epitope to facilitate the antibody class switch in conjugate vaccines.



Scheme 2. Synthesis of a three-component conjugate vaccine containing a Tn antigen, the Pam₃Cys epitope, and a helper T cell epitope.

Despite the promising immunological results of the above three-component conjugate vaccine, new synthetic challenges of covalently linking glycopeptide to Pam₃Cys cast a shadow over the application of this strategy. In the synthesis of 6 and 7, Pam₃Cys could be directly linked to the glycopeptide antigen N-terminus. However, with the increase of complexity of glycopeptide moieties by introducing the helper T cell epitope, their coupling with the lipopeptide carrier has become more difficult. Many approaches to achieve glycopeptides conjugation with Pam₃Cys, including preinstallation of molecular handles such as alkyne or active ester, in the Pam₃Cys moiety have been described [40–45]. In the Boons group, native chemical ligation (NCL) was brought into the synthesis of three-component vaccine 19 (Scheme 3) [46,47]. The reaction between the thioester of helper T cell epitope 15 from polio virus and tumor-associate antigen 16 derived from cell surface associated Mucin 1 (MUC1) with a terminal cysteine in the presence of tris(2-carboxyethyl)phosphine (TCEP) efficiently constructed the amide linkage between the two moieties. The N-terminal of helper T cell epitope 15 was also modified with a cysteine. Similarly, the reaction between the thioester of TLR1/2 ligand Pam₃CSK₄ 18 with the terminal cysteine in 17 furnished the three-component conjugate vaccine 19. It was later discovered that the NCL step could be further improved by embedding the thioester of Pam_3CSK_4 in liposomes [48]. In addition to the improvement from a synthetic perspective, conjugate **19** also induced much improved immune response with over 100 folds of IgG antibody titers against MUC1 as compared to **14**. The increased potency of **19** in the immunological study undoubtedly demonstrated the importance of vaccine design. It is noteworthy that enantiomerically pure glycerol was used to prepare Pam_3CSK_4 in contrast to previous examples. However, a later study by Shi et al. [49] demonstrated that the absolute configuration had no significant impact on the immunogenicity, which is in consistent with the earlier study on their peptide conjugates [50].



Scheme 3. Synthesis of MUC1 glycopetide-based three-component conjugate vaccine **19** containing the Pam₃CSK₄ epitope and a helper T cell epitope.

In pursuing a better presentation of antigens to the immune system, Cai et al. [51] synthesized cluster conjugate vaccines that had four MUC1 glycopeptide antigens linked to Pam₃CSK₄ (Scheme 4). A cluster effect showing positive correlation between the antigen valency and the antibody production was clearly demonstrated. In addition, stronger binding and higher cytotoxicity of the antiserum to cancer cells were also observed with the tetravalent glycopeptide antigen vaccines **22a** and **22b** as compared to the mono- and divalent counterparts.

In another report, the Dumy group [52,53] loaded multiple Tn antigens onto a cyclic decapeptide scaffold using a strategy called regioselectively addressable functionalized template (RAFT) developed by the same group earlier [54]. When constructing this peptide scaffold, versatile handles were introduced to lysine so that subsequent conjugation with the Tn antigen could be achieved in aqueous environment (Scheme 5). After excellent IgG immune response was observed for conjugate **25**, a systematic biological study was performed. Two conjugates with either linear or branched palmitoyl-peptide were evaluated in several aspects including the uptake, processing, cross-presentation pathways and how the B and T cell immunogenicity were affected. While both conjugates inhibit tumor growth, they appear to undergo different cross-presentation pathways. The linear molecule proved to be superior in terms of activating dendritic cells and inducing potent TLR2-dependent T cell activation.



Scheme 4. Synthesis of tetravalent MUC1 glycopeptide-based vaccines with Pam_3CSK_4 as the carrier molecule or adjuvant.



Scheme 5. Synthesis of tetravalent glycopeptide conjugate vaccine 25 based on RAFT.

Direct linkage of oligosaccharide antigens to Pam3Cys to form synthetic vaccines was also reported. In 2001, the Seeberger group [55] conjugated a leishmaniasis tetrasaccharide antigen with Pam₃Cys via an amide bond (Scheme 6). However, no immunological data was provided, thus we have no idea about the efficiency of this vaccine to induce immune responses in vivo.



Scheme 6. Synthesis of a carbohydrate-based Leishmaniasis vaccine with Pam₃Cys as the carrier molecule.

The less lipidated lipopeptide Pam₂Cys is a motif derived from MALP-2 of mycoplasma, which lacks the cell wall [56], and was shown to activate TLR2/6 heterodimer [30]. Compared to their Pam₃ counterparts, Pam₂Cys and Pam₂CSK₄ are more soluble in aqueous solutions, which is considered a significant pharmacological advantage as it facilitates the trafficking of vaccines. Moreover, the free amino group of Pam₂Cys allows for further modification, which may add unique functions to the conjugates. For example, in a recent study on cancer imaging conducted by the Vagner group, a fluorescent tag or metal chelate was attached to the amino group through a PEG spacer without affecting the potency of Pam₂Cys [57]. Despite these advantages, Pam₂Cys and Pam₂CSK₄ are believed to be more suitable for conjugation with peptide antigens because their conjugates with carbohydrate antigens were not satisfactory and less studied [46].

McDonald et al. [58] synthesized the complete MALP-2 and conjugated it with the extracellular domain of MUC1 consisting of variable-number tandem repeat (VNTR) sequence with twenty amino acids and five O-glycans (Scheme 7). A fragment condensation strategy developed by Payne et al. [42,59,60] to construct MUC1-Pam₃Cys conjugates was adopted for the synthesis of target molecule 33. MUC1 glycopeptide 29 and active ester of PEG-peptide 30, which were both obtained from solid-phase peptide synthesis (SPPS), were coupled followed by conjugation with Pam₂Cys via active ester 32. Immunological studies of the resultant glycoconjugates 33a and 33b showed that they induced the production of several isotypes of IgG antibodies. The production of IgG antibody suggests that activation of TLRs by lipopeptides are sufficient and the T cell epitope may not always be necessary to initiate adaptive immune response [61]. It was thus possible that the failure of the aforementioned vaccines 6 and 7 to elicit IgG antibody production might be ascribed to other reasons, such as inefficient presentation of TLR ligands and lack of spacing between functional moieties. It is believed that an immunologically silent spacer like ethylene glycol is necessary to diminish the interplay between MUC1 antigen and peptide of Pam₃Cys analogs [62]. In fact, strong interactions between carbohydrate and peptide [63] affect the MUC1 antigen confirmation [64,65], resulting in a particular 3-D structure that can be recognized by immune cells as an entity. Disturbance from Pam₃Cys peptide may cause change in the confirmation of MUC1.



Scheme 7. Synthesis of MUC1 glycopeptide-MALP-2 conjugates 33a and 33b as vaccine candidates.

Pam₃Cys, Pam₂Cys, and MALP-2 are among the most studied TLR ligands as adjuvants and their structure-activity relationship (SAR) studies have been performed extensively in the attempt to improve their adjuvant properties. Optimization towards higher binding affinity with TLRs or their heterodimers and improvement of the pharmacodynamics properties are the two major directions of the SAR studies. The crystal structure of a TLR1/2 heterodimer has shown that the stereochemistry and orientation of acyl chains in the lipopeptide ligands are critical for their optimal fitting in the hydrophobic pocket of TLR1/2 heterodimer [66]. Therefore, the stereochemistry, length, and number of acyl chains in the lipopeptides were investigated in correlation with their agonist and adjuvant properties [61,67–69]. For example, it was disclosed that switching the amide linkage of lipid chains in Pam₃Cys to a urea linkage was beneficial for DC maturation [70] and replacing the two ester linkages with amide bonds could improve its stability towards esterases [71]. The position of Pam₃Cys in the conjugate vaccine was also found to be related to the biophysics of the conjugates [72]. When adjuvant was placed in the middle of B and T epitopes, the conjugate vaccine was found to be most soluble and immunogenic. In another study to develop new TLR ligand adjuvants, lipoamino acids with various lipid chains were synthesized and evaluated against Pam₃CSK₄ in three-component conjugate vaccines by the Toth group and they found that two repeating units of lipoamino acids with 16 carbon lipid chains were most immunostimulating by activating TLR2, although not as potent as Pam₃CSK₄ [73]. These investigations have been reviewed recently [74,75] and will not be elaborated here.

As amphiphiles, the above lipopeptides form micelles in aqueous environments. Micelle formation can improve the in vivo stability and presentation of antigens to B cells, and more importantly, the proper particle size (5–100 nm) is critical for passive drainage of micelles into lymph nodes where naïve T and B cells reside [12]. The assembly of three-component MUC1-T epitope-Pam₃Cys conjugates in an aqueous environment were evaluated by the Payne group [60], who found that a micelle size of 17–25 nm is ideal for passive drainage to lymph nodes.

3. MPLAs as Carrier Molecules or Adjuvants for Vaccine Development

3.1. TLR4 Ligands as Adjuvants

Among all TLRs, TLR4 stands out due to several unique features of its signaling pathway. It was the first human TLR discovered [76] and thus the most studied. It forms a complex with MD2 for action [25] and signals through the MyD88-dependent pathway on the cell surface. The complex can be internalized to endosome, creating an additional activation site for TLR4 signaling. Internalized TLR4-MD2 complex triggers the toll/interleukin-1 receptor (TIR) domain-containing adapter-inducing interferon- β (TRIF) pathway of signaling [77], adding another profile that can be targeted. These features allow for unusual flexibility in attenuating ligand structures towards better adjuvant properties in conjugate vaccine. This is supported by the variety of TLR4 ligands, which range from LPSs [76] to proteins (heat shock proteins [78]), small molecules (opioid drugs [79]) and even metals (nickel [80]).

Among all TLR4 ligands, lipid A is the most famous and studied. It is lipophilic anchor and the main biologically active motif of LPS, which is a constituent of gram-negative bacteria cell wall. Small amounts of LPS/lipid A released from invading bacteria can serve as a "danger signal" for host immune systems to initiate counteracts, which may be reflected as inflammation. If not controlled, this can lead to fatal septic shock.

Depending on the bacterial strain where LPS is isolated, lipid A may vary in structure, especially in terms of the length, saturation, and number of fatty acid chains. However, a shared core structure of β -D-glucosamine (GlcN)-(1 \rightarrow 6)- β -D-glucosamine disaccharide with 1,4'-di-O-phosphorylation and 2,2'-N-3,3'-O-acylation, is always observed with lipid A (Figure 1). Although lipid A has been known to be strongly immunostimulatory and evaluated as a vaccine adjuvant for decades, it's utility was limited by the endotoxicity related to septic shock mentioned above. Fortunately, it was found that mild hydrolytic treatment of LPS, e.g., that from *Salmonella minnesota* R595, gave rise to monophosohoryl lipid A (MPLA) [81], which was 1000-fold less toxic with retained immunostimulatory property. This discovery ensures the safe use of MPLA and led to the prosperity of lipid A-related research. For example, the combination of safety and efficacy with the identification of TLR4 as the signaling receptor of lipid A has finally paved the way for the application of MPLA as a vaccine carrier.



Figure 1. Conservative structure of lipid A as the core of LPS.

3.2. Use of Lipid A Derivatives for Vaccine Development and/or as External Adjuvants

Since then, much effort has been focused on the immunostimulatory activities of MPLAs as external adjuvants and extensive clinical studies have been reported. In combination with aluminum salts or QS21 (also known as AS04 and AS02, respectively), a heterogenous mixture of MPLAs having varied fatty acids in length and acylation pattern was administered together with multiple antigens against various diseases. Vaccines against hepatitis B [82], HIV [83], malaria [84], and multiple cancers [85–89] have entered clinical trials. Inspired by the success of MPLA mixtures, lipid A mimetics, such as aminoalkyl glucosaminide 4-phosphates (AGPs), have been developed and probed as external adjuvants. After being extensively evaluated in human clinical trials, a few APGs have demonstrated to be an efficacious adjuvant with an excellent safety profile [90]. An AGP-based vaccine against Hepatitis B has been approved for use in Argentina [91]. Compared to MPLA, AGPs have much simplified structures, rendering their synthesis easier and more efficient while exhibiting immunostimulatory activity comparable to or better than MPLA in preclinical studies.

3.3. MPLA Analogs as Carrier Molecules and Built-in Adjuvants for Conjugate Vaccine Development

Inspired by the success of Pam₃Cys conjugate vaccines, the Guo group designed and synthesized a series of oligosaccharide conjugate vaccines with MPLA as a vaccine carrier and built-in adjuvant (for lipid A synthesis, please refer to a recent review [92]). The synthesis of MPLA derived from *N. meningitidis* with symmetric lipid chain distribution and an aminoethyl group on the reducing-end GlcN unit is depicted in Scheme 8 [93]. After glycosylation of **36** with thioglycoside **35**, the designated amino and hydroxyl groups for lipidation were simultaneously exposed upon hydrazine treatment. Then, four long chain acyl groups were sequentially introduced onto the disaccharide to afford **40**. Regioselective reductive opening of the benzylidene ring exposed the 4'-hydroxyl group, which was phosphorylated to provide protected MPLA **42**.

To facilitate the coupling of carbohydrate antigens with MPLA, the azido group in **42** was reduced to free amine, which was then acylated with a bifunctional linker followed by conversion of the resultant acid into an active ester of *p*-nitrophenol (Scheme 9). Finally, the modified GM3 antigen **45** was coupled with **44** at ambient temperature, and the resulting conjugate was globally debenzylated via Pd-catalyzed hydrogenolysis to give vaccine candidate **46**. Immunological studies showed that this conjugate vaccine elicited strong immune responses without an external adjuvant, indicating its self-adjuvanting property. In addition, the conjugate induced mainly IgG3 antibody production, which suggested the elicitation of adaptive immune response with sustained immunological memory [94].



Scheme 8. Synthesis of *N. meningitidis* MPLA with an azidoethyl group on the reducing-end glucosamine unit.



Scheme 9. Conjugation of *N. meningitidis* MPLA with a modified GM3 antigen to form conjugate vaccine **46**.

Encouraged by the above discovery, the Guo group conducted a series of studies to optimize MPLA structure via SAR analysis and explore this new concept with other carbohydrate antigens. As reported by Park et al. [25], five of the six lipid chains of *E. coli* lipid A are deeply buried in the hydrophobic pocket of the TLR4/MD2 heterodimer. Any change on these lipid chains is deemed to have a significant impact on the binding affinity between the two parties. Accordingly, the Guo group investigated the influence of lipid chains on the adjuvanting property of MPLA in the context of conjugate vaccines [95]. Thus, a series of MPLAs with different lipids on the 3-*O*-positions of both glucosamine units were synthesized and then coupled with sTn antigen by procedures similar to that depicted above. The resulting conjugates **47a–d** (Figure 2) were immunologically evaluated in mice. It was demonstrated that **47a** derived from the natural form of *N. meningitides* lipid A was the

most effective in eliciting the production of both total and IgG3 antibodies. Moreover, additional or elongated lipid chains decreased the efficacy of antibody production. The results are useful for optimizing conjugate vaccine designs.



Figure 2. Conjugates of sTn antigen with MPLAs containing different lipid structures for SAR studies.

MPLA from *E coli* lipid A, e.g., **48** (Scheme 10), has asymmetrically distributed lipid chains on each monosaccharide unit. Different from aforementioned synthesis, a highly convergent strategy featuring late stage coupling of two monosaccharides having preinstalled lipid chains was adopted for the synthesis of MPLA **29** by the Guo group [96]. This MPLA was furnished with an alkyne group at the reducing terminus, which served as the linker for direct coupling with a modified GM3 antigen **49** that had an azidoethyl group at the reducing end by click reaction. Under catalysis of copper (I) iodide, the two moieties were conjugated via a triazole linkage in a good yield (70%).



Scheme 10. Conjugation of E. coli MPLA 48 with a modified GM3 antigen by click reaction.

Globo H is a carbohydrate antigen found on multiple epithelial tumors including breast cancer, lung, colon and prostate cancers. Its conjugate with the KLH protein carrier showed potent immunogenicity against breast cancer and has reached phase III clinical trial [97]. In 2015, the Guo group synthesized the conjugate **51** of globo H with optimized *N. meningitidis* MPLA (Scheme 11) and compared it with the KLH-globo H conjugate **52** [98].



Scheme 11. Conjugation of globo H antigen with KLH and MPLA to formulate conjugate vaccines **52** and **53**.

The same linker was used for both KLH and MPLA conjugates to ensure unbiased comparison. Overall, the MPLA conjugate was found to induce IgG antibody titers 2-fold higher than the KLH conjugate. Moreover, the immune response towards MPLA-globo H conjugate was much faster. Considering that the failure of KLH-sTn vaccine in phase III clinical trials was ascribed to slow immune response that was overwhelmed by cancer progression [99], fast immune response may be one of the desirable features of the new cancer vaccine. A series of α -2,9-oligosialic acids varying in chain length were also conjugated with MPLA, and the resultant conjugates **54a–e** (Figure 3) as vaccine candidates against group C meningitis were compared to the corresponding KLH conjugates [100].



Figure 3. Conjugates of N. meningitidis MPLA with polysialic acids as group C meningitis vaccine candidates.

Both types of glycoconjugates showed comparable and robust activities to elicit IgG antibody induction. Further in-depth SAR study of the MPLA conjugates revealed that the hydroxyl group on the acyl lipid chains of MPLA was important for the vaccine, since **54c** elicited stronger immune response than **54e**, which were only different in the lipid structure. It was also concluded that the overall immunogenity of these conjugate vaccines decreased with elongation of the oligosialic acid chain and higher antibody titers were observed with **54a** than all other tested conjugates.

In natural LPSs, the polysaccharide chain is invariably linked to the 6'-O-position of lipid A. To avoid any potential influence of carbohydrate antigens attached to the MPLA disaccharide reducing end on the binding between MPLA and TLR4/MD-2 heterodimer, the Guo group also synthetized a MPLA derivative with modified 6'-O-position **56** (Scheme 12) and examined its activity as a vaccine carrier and built-in adjuvant [101]. In this context, **56** was conjugated with **55**, which is the non-reducing end cap oligosaccharide of lipoarabinomannan (LAM), a cell surface lipopolysaccharide of *Mycobacterium tuberculosis* essential for mycobacterial infections, by click reaction. The resultant

conjugate **57** was compared immunologically with conjugate **60** (Scheme 12) that had the same oligosaccharide antigen attached to the reducing end of the same MPLA. Immunological studies showed that although both conjugates induced strong IgG antibody responses, the 6'-O-linked MPLA conjugate **57** elicited significantly higher titers of both total antibodies and IgG antibodies as compared to the 1-O-linked analogue **60**. This may be attributed to the fact that the carbohydrate chain at the lipid A 6'-O-position in LPS is far away from the binding pocket and thus has less interactions with the TLR4/MD-2 heterodimers as demonstrated by Park et al. [25]. Consequently, having carbohydrate antigens linked to the MPLA 6'-O-position may be the optimal vaccine design.



Scheme 12. Conjugation of an upstream LAM oligosaccharide with MPLA at different positions.

Another TACA, the GM2 antigen, was also coupled with MPLA (Scheme 13) by a strategy similar to that used to synthesize the globo H conjugate (Scheme 11), and the resultant glycoconjugate **63** was evaluated [102]. Once again, immunological studies demonstrated the self-adjuvanting property of **63** as a vaccine candidate, since it elicited strong immune responses in mouse without combined usage with an external adjuvant, and the activity of **63** to elicit immune responses was only marginally weaker than that of the KLH-GM2 conjugate. More significantly, the antisera induced by glycoconjugate **63** showed antibody-mediated complement-dependent cytotoxicities to breast cancer cell line MCF-7, indicating the potential of **63** as a therapeutic cancer vaccine.



Scheme 13. Conjugation of GM2 antigen with MPLA to form conjugate vaccine 63.

Due to the structural complexity of MPLA, the scale-up of its chemical synthesis is a challenge. To overcome this problem, the Bedini group took a different approach to obtain selectively activated MPLA derivatives via regioselective modification of MPLA derived from fermentation [103]. This semisynthetic method started with selective oxidation of the 6'-hydroxyl in **64** (Scheme 14). The resulting 6'-carboxylic acid derivative was then coupled with linkers bearing various functionalities, including alkene, alkyne and azide, via an amido bond. This allowed for MPLA conjugation with carbohydrate antigens by diverse coupling methods. Eventually, TF antigen with a thiol group **67** was attached to the alkene group in **66a** in presence of a photoinitiator under UV light. Immunological studies showed that the resulting conjugate **68** could successfully induce cytokine production [104].



Scheme 14. Synthesis of a MPLA derivative **64** from naturally derived MPLA and its subsequent conjugation with the TF antigen.

3.4. Conjugate Vaccines with MPLA Mimics as Carrier Molecules or Built-In Adjuvants

Long before the total synthesis of natural lipid A and MPLA was pursued, researchers had foreseen the potential of lipid A in conjugate vaccine development from its success as external adjuvants. However, limited by the resources, they had to turn their attention to partial structures of lipid A. For example, in 1997, Achiwa et al. coupled Tn and sTn antigens to a partial structure of lipid A without the phosphoryl moiety [105]. They also synthesized a lipid A mimic 69 using L-serine to replace D-glucosamine (Scheme 15), which proved to be a potent mitogen [106]. This lipid A mimic was then coupled with Tn or sTn antigen bearing a diamine spacer to give conjugate **71**. Unfortunately, no immunological data was given regarding these conjugates.



Scheme 15. Conjugation of Tn and sTn antigens with MPLA mimic 69.

As a part of their efforts to simplify the structure of lipid A while maintaining its biological activities, the Jiang group designed and synthesized a series of mimics of *E. coli* MPLA. By screening their activities to elicit ICAM-1 and cytokine production in human THP-1 cells, **75** (Scheme 16) was found to be a potent immunostimulator [107]. TF antigen was covalently linked to this MPLA mimic via a linear spacer and amide bonds, followed by deprotection under hydrogenolysis conditions to afford conjugate vaccine **77**, setting the stage for subsequent immunological studies to be reported later [108].



Scheme 16. Conjugation of TF antigen with MPLA mimic 75 to prepare conjugate vaccine 77.

Compared to lipopeptides, lipooligosaccharides were less exploited as vaccine carriers with only a few successes exemplified by MPLA. However, it is noteworthy that even without a helper T epitope, MPLA conjugates can induce high titers of IgG antibodies, indicating the stronger immunostimulating activity of MPLA than lipopeptides such as Pam₂Cys/Pam₃Cys. It can be envisioned that, by implanting the innovative strategies exploited in Pam₂Cys/Pam₃Cys conjugates, the advantages of MPLA conjugate vaccines will be further explored in the future.

4. Other TLR Ligand Conjugates

Although less studied, other TLR ligands such as polyinosinic:polycytidylic acid (poly I:C, a RNA analog that activates TLR3) [109] and unmethylated oligodeoxy-nucleotides (CpG ODN) have been used together with MUC1 antigen. But due to their intrinsically poor drug properties and difficulties in characterizing the conjugates, utilization of DNA/RNA as built-in adjuvants is less attractive to researchers and only a few reports on their conjugation with glycopeptide antigens were found.

A multicomponent vaccine candidate comprised of CpG ODN, T-epitope and MUC1 antigen was first designed and synthesized by Boons et al. (Scheme 17, compound 78) [110] However, subsequent immunological study revealed that this vaccine candidate was less potent than Pam₃CSK₄ conjugates. Trials of incorporating CpG ODN by electrostatic interactions with T-epitope and MUC1 antigen have also been reported [111], but cannot be categorized as conjugate, therefore will not be elaborated in this review.



Scheme 17. Multicomponent vaccine candidate 78 consisting of CpG ODN, T-epitope and MUC1 antigen.

5. Conclusions

Melding two or more moieties with diverse functions into one multi-tasking conjugate to modulate the human immune system for fighting against cancer or infectious diseases is one of the most promising therapeutic strategies. Compared to well-established protein carriers for conjugate vaccines, TLR ligands such as lipopeptides and MPLA derivatives as vaccine carriers are still at their early stage. However, since their interactions with TLRs are well studied and understood, the prosperity of using TLR ligands for conjugate vaccine development can be envisioned, especially because these molecules are not only vaccine carriers but also have adjuvant property to generate self-adjuvanting conjugate vaccines. As a critical immunostimulatory component in all types of vaccine formulations, adjuvants are expected to stimulate T cell-mediated immune responses and antibody class switching for long-term immunity. In this respect, TLR ligands are ideal candidates as they help not only elicit immune responses but also switch innate immune responses to carbohydrate antigens to adaptive responses, allowing for robust and memorable immunity.

Preliminary data of antibody induction by conjugate vaccines derived from TLR ligands, especially MPLA, have shown their unquestionable potential. These results, together with the long-lasting studies on MPLA as an external vaccine adjuvant, suggest very promising outlook of its application as a carrier molecule in conjugate vaccine development. Currently, one of the major challenges in this field is how to obtain sufficient quantities of the vaccine for clinical studies, as the structures of lipopeptides

and especially MPLA are complex and their synthesis and their conjugation with target antigens are not trivial. Semi-total synthesis and APGs synthesis are making significant input in solving this problem. The second challenge is how to reach the full potential of each moiety in a multi-component vaccine construct by rational considerations in vaccine design. As case study of vaccines compiling and crystal structures of TLR-ligand complexes elucidated, better vaccine designs may be in our reach through SAR analysis and molecular modeling. Furthermore, as suggested by the Pam₃Cys conjugates, the immunogenecity of conjugate vaccines can be highly dependent on the conjugation strategy [12]. Therefore, after structural optimization of MPLA, efforts aiming at further improvement of the properties of its conjugates should be focused on the ligation strategies and antigen presentation patterns. For example, multivalent vaccines may help improve the presentation of antigens to APCs, whereas hydrophilic linkers may help improve their pharmacologic properties.

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References

- Fuster, M.M.; Esko, J.D. The sweet and sour of cancer: Glycans as novel therapeutic targets. *Nat. Rev. Cancer* 2005, *5*, 526–542. [CrossRef] [PubMed]
- 2. Ohtsubo, K.; Marth, J.D. Glycosylation in cellular mechanisms of health and disease. *Cell* **2006**, *126*, 855–867. [CrossRef] [PubMed]
- 3. Stowell, S.R.; Ju, T.; Cummings, R.D. Protein Glycosylation in Cancer. *Annu. Rev. Pathol. Mech. Dis.* **2015**, *10*, 473–510. [CrossRef] [PubMed]
- 4. Kayhty, H.; Karanko, V.; Peltola, H.; Makela, P.H. Serum antibodies after vaccination with Haemophilus influenzae type b capsular polysaccharide and responses to reimmunization: No evidence of immunologic tolerance or memory. *Pediatrics* **1984**, *74*, 857–865. [PubMed]
- O'Brien, K.L.; Steinhoff, M.C.; Edwards, K.; Keyserling, H.; Thoms, M.L.; Madore, D. Immunologic priming of young children by pneumococcal glycoprotein conjugate, but not polysaccharide, vaccines. *Pediatr. Infect. Dis. J.* 1996, 15, 425–430. [CrossRef] [PubMed]
- Guttormsen, H.-K.; Sharpe, A.H.; Chandraker, A.K.; Brigtsen, A.K.; Sayegh, M.H.; Kasper, D.L. Cognate stimulatory B-cell-T-cell interactions are critical for T-cell help recruited by glycoconjugate vaccines. *Infect. Immun.* 1999, 67, 6375–6384. [PubMed]
- 7. Goldblatt, D. Conjugate vaccines. Clin. Exp. Immunol. 2000, 119, 1–3. [CrossRef] [PubMed]
- Gold, R.; Lepow, M.L.; Goldschneider, I.; Draper, T.L.; Gotschlich, E.C. Clinical evaluation of group A and group C meningococcal polysaccharide vaccines in infants. *J. Clin. Investig.* 1975, *56*, 1536–1547. [CrossRef] [PubMed]
- 9. Coffman, R.L.; Sher, A.; Seder, R.A. Vaccine Adjuvants: Putting Innate Immunity to Work. *Immunity* **2010**, *33*, 492–503. [CrossRef] [PubMed]
- 10. Awate, S.; Babiuk, L.A.; Mutwiri, G. Mechanisms of action of adjuvants. *Front. Immunol.* **2013**, *4*, 114. [CrossRef] [PubMed]
- 11. Avci, F.Y.; Kasper, D.L. How bacterial carbohydrates influence the adaptive immune system. *Annu. Rev. Immunol.* **2010**, *28*, 107–130. [CrossRef] [PubMed]
- 12. Liu, H.; Irvine, D.J. Guiding Principles in the Design of Molecular Bioconjugates for Vaccine Applications. *Bioconjug. Chem.* **2015**, *26*, 791–801. [CrossRef] [PubMed]
- 13. Durando, P.; Faust Saul, N.; Torres, A. Immunological Features and Clinical Benefits of Conjugate Vaccines against Bacteria. *J. Immunol. Res.* 2015, 2015, 934504. [CrossRef] [PubMed]
- 14. Goebel, W.F.; Avery, O.T. Chemo-immunological studies on conjugated carbohydrate-proteins. I. The synthesis of p-aminophenol β-glucoside, p-aminophenol β-galactoside, and their coupling with serum globulin. *J. Exp. Med.* **1929**, *50*, 521–531. [CrossRef] [PubMed]

- Tillett, W.S.; Avery, O.T.; Goebel, W.F. Chemo-immunological studies on conjugated carbohydrate-proteins. III. Active and passive anaphylaxis with synthetic sugar-proteins. *J. Exp. Med.* **1929**, *50*, 551–567. [CrossRef] [PubMed]
- Avery, O.T.; Goebel, W.F. Chemo-immunological studies on conjugated carbohydrate-proteins. II. Immunological specificity of synthetic sugar-protein antigens. *J. Exp. Med.* **1929**, *50*, 533–550. [CrossRef] [PubMed]
- 17. Pichichero, M.E. Protein carriers of conjugate vaccines: Characteristics, development, and clinical trials. *Hum. Vaccines Immunother.* **2013**, *9*, 2505–2523. [CrossRef] [PubMed]
- 18. Micoli, F.; Costantino, P.; Adamo, R. Potential targets for next generation antimicrobial glycoconjugate vaccines. *FEMS Microbiol. Rev.* **2018**, *42*, 388–423. [CrossRef] [PubMed]
- 19. Astronomo, R.D.; Burton, D.R. Carbohydrate vaccines: Developing sweet solutions to sticky situations? *Nat. Rev. Drug Discov.* **2010**, *9*, 308–324. [CrossRef] [PubMed]
- 20. Avci, F.Y.; Li, X.; Tsuji, M.; Kasper, D.L. A mechanism for glycoconjugate vaccine activation of the adaptive immune system and its implications for vaccine design. *Nat. Med.* **2011**, *17*, 1602–1609. [CrossRef] [PubMed]
- 21. Iwasaki, A.; Medzhitov, R. Toll-like receptor control of the adaptive immune responses. *Nat. Immunol.* **2004**, *5*, 987–995. [CrossRef] [PubMed]
- 22. Akira, S.; Takeda, K.; Kaisho, T. Toll-like receptors: Critical proteins linking innate and acquired immunity. *Nat. Immunol.* **2001**, *2*, 675–680. [CrossRef] [PubMed]
- Takeuchi, O.; Kaufmann, A.; Grote, K.; Kawai, T.; Hoshino, K.; Morr, M.; Muhlradt, P.F.; Akira, S. Cutting edge: Preferentially the R-stereoisomer of the mycoplasmal lipopeptide macrophage-activating lipopeptide-2 activates immune cells through a toll-like receptor 2- and MyD88-dependent signaling pathway. *J. Immunol.* 2000, 164, 554–557. [CrossRef] [PubMed]
- Kawai, T.; Takeuchi, O.; Fujita, T.; Inoue, J.-I.; Muhlradt, P.F.; Sato, S.; Hoshino, K.; Akira, S. Lipopolysaccharide stimulates the MyD88-independent pathway and results in activation of IFN-regulatory factor 3 and the expression of a subset of lipopolysaccharide-inducible genes. *J. Immunol.* 2001, 167, 5887–5894. [CrossRef] [PubMed]
- 25. Park, B.S.; Song, D.H.; Kim, H.M.; Choi, B.-S.; Lee, H.; Lee, J.-O. The structural basis of lipopolysaccharide recognition by the TLR4-MD-2 complex. *Nature* **2009**, *458*, 1191–1195. [CrossRef] [PubMed]
- 26. Ignacio, B.J.; Albin, T.J.; Esser-Kahn, A.P.; Verdoes, M. Toll-like Receptor Agonist Conjugation: A Chemical Perspective. *Bioconjug. Chem.* **2018**, *29*, 587–603. [CrossRef] [PubMed]
- Hess, N.J.; Felicelli, C.; Grage, J.; Tapping, R.I. TLR10 suppresses the activation and differentiation of monocytes with effects on DC-mediated adaptive immune responses. *J. Leukoc. Biol.* 2017, 101, 1245–1252. [CrossRef] [PubMed]
- 28. Fujita, Y.; Taguchi, H. Overview and outlook of Toll-like receptor ligand-antigen conjugate vaccines. *Ther. Deliv.* **2012**, *3*, 749–760. [CrossRef] [PubMed]
- Takeuchi, O.; Sato, S.; Horiuchi, T.; Hoshino, K.; Takeda, K.; Dong, Z.; Modlin, R.L.; Akira, S. Cutting edge: Role of toll-like receptor 1 in mediating immune response to microbial lipoproteins. *J. Immunol.* 2002, 169, 10–14. [CrossRef] [PubMed]
- Jackson, D.C.; Lau, Y.F.; Le, T.; Suhrbier, A.; Deliyannis, G.; Cheers, C.; Smith, C.; Zeng, W.; Brown, L.E. A totally synthetic vaccine of generic structure that targets toll-like receptor 2 on dendritic cells and promotes antibody or cytotoxic T cell responses. *Proc. Natl. Acad. Sci. USA* 2004, 101, 15440–15445. [CrossRef] [PubMed]
- 31. Hopp, T.P. Immunogenicity of a synthetic HBsAg peptide: Enhancement by conjugation to a fatty acid carrier. *Mol. Immunol.* **1984**, *21*, 13–16. [CrossRef]
- 32. Jung, G.; Wiesmueller, K.H.; Becker, G.; Buhring, H.J.; Bessler, W.G. Increased production of specific antibodies by presentation of antigen determinants with covalently bound lipopeptide mitogens. *Angew. Chem.* **1985**, *97*, 883–885. [CrossRef]
- 33. Toyokuni, T.; Hakomori, S.; Singhal, A.K. Synthetic carbohydrate vaccines: Synthesis and immunogenicity of Tn antigen conjugates. *Bioorg. Med. Chem.* **1994**, *2*, 1119–1132. [CrossRef]
- Toyokuni, T.; Dean, B.; Cai, S.; Boivin, D.; Hakomori, S.; Singhal, A.K. Synthetic vaccines: Synthesis of a dimeric Tn antigen-lipopeptide conjugate that elicits immune responses against Tn-expressing glycoproteins. *J. Am. Chem. Soc.* **1994**, *116*, 395–396. [CrossRef]

- 35. Kudryashov, V.; Glunz, P.W.; Williams, L.J.; Hintermann, S.; Danishefsky, S.J.; Lloyd, K.O. Toward optimized carbohydrate-based anticancer vaccines: Epitope clustering, carrier structure, and adjuvant all influence antibody responses to Lewisy conjugates in mice. *Proc. Natl. Acad. Sci. USA* 2001, *98*, 3264–3269. [CrossRef] [PubMed]
- Glunz, P.W.; Hintermann, S.; Schwarz, J.B.; Kuduk, S.D.; Chen, X.-T.; Williams, L.J.; Sames, D.; Danishefsky, S.J.; Kudryashov, V.; Lloyd, K.O. Probing cell surface "Glyco-Architecture" through total synthesis. Immunological consequences of a human blood group determinant in a clustered mucin-like context. J. Am. Chem. Soc. 1999, 121, 10636–10637. [CrossRef]
- 37. Keding, S.J.; Danishefsky, S.J. Prospects for total synthesis: A vision for a totally synthetic vaccine targeting epithelial tumors. *Proc. Natl. Acad. Sci. USA* **2004**, *101*, 11937–11942. [CrossRef] [PubMed]
- 38. Buskas, T.; Ingale, S.; Boons, G.-J. Towards a fully synthetic carbohydrate-based anticancer vaccine: Synthesis and immunological evaluation of a lipidated glycopeptide containing the tumor-associated Tn antigen. *Angew. Chem. Int. Ed.* **2005**, *44*, 5985–5988. [CrossRef] [PubMed]
- 39. Reichel, F.; Ashton, P.R. Synthetic carbohydrate-based vaccines: Synthesis of an L-glycero-D-manno-heptose antigen-T-epitope-lipopeptide conjugate. *Chem. Commun.* **1997**, 2087–2088. [CrossRef]
- 40. Nalla, N.; Pallavi, P.; Reddy, B.S.; Miryala, S.; Naveen Kumar, V.; Mahboob, M.; Halmuthur, M.S.K. Design, synthesis and immunological evaluation of 1,2,3-triazole-tethered carbohydrate-Pam3Cys conjugates as TLR2 agonists. *Bioorg. Med. Chem.* **2015**, *23*, 5846–5855. [CrossRef] [PubMed]
- 41. Cai, H.; Huang, Z.-H.; Shi, L.; Zhao, Y.-F.; Kunz, H.; Li, Y.-M. Towards a Fully Synthetic MUC1-Based Anticancer Vaccine: Efficient Conjugation of Glycopeptides with Mono-, Di-, and Tetravalent Lipopeptides Using Click Chemistry. *Chem. Eur. J.* **2011**, *17*, 6396–6406. [CrossRef] [PubMed]
- 42. Wilkinson, B.L.; Malins, L.R.; Chun, C.K.Y.; Payne, R.J. Synthesis of MUC1-lipopeptide chimeras. *Chem. Commun.* **2010**, *46*, 6249–6251. [CrossRef] [PubMed]
- 43. Hossain, M.K.; Vartak, A.; Karmakar, P.; Sucheck, S.J.; Wall, K.A. Augmenting vaccine immunogenicity through the use of natural human anti-rhamnose antibodies. *ACS Chem. Biol.* **2018**. [CrossRef] [PubMed]
- Sarkar, S.; Salyer, A.C.D.; Wall, K.A.; Sucheck, S.J. Synthesis and Immunological Evaluation of a MUC1 Glycopeptide Incorporated into L-Rhamnose Displaying Liposomes. *Bioconjug. Chem.* 2013, 24, 363–375. [CrossRef] [PubMed]
- 45. Karmakar, P.; Lee, K.; Sarkar, S.; Wall, K.A.; Sucheck, S.J. Synthesis of a Liposomal MUC1 Glycopeptide-Based Immunotherapeutic and Evaluation of the Effect of L-Rhamnose Targeting on Cellular Immune Responses. *Bioconjug. Chem.* **2016**, *27*, 110–120. [CrossRef] [PubMed]
- 46. Ingale, S.; Wolfert, M.A.; Gaekwad, J.; Buskas, T.; Boons, G.-J. Robust immune responses elicited by a fully synthetic three-component vaccine. *Nat. Chem. Biol.* **2007**, *3*, 663–667. [CrossRef] [PubMed]
- 47. Lakshminarayanan, V.; Thompson, P.; Wolfert, M.A.; Buskas, T.; Bradley, J.M.; Pathangey, L.B.; Madsen, C.S.; Cohen, P.A.; Gendler, S.J.; Boons, G.-J. Immune recognition of tumor-associated mucin MUC1 is achieved by a fully synthetic aberrantly glycosylated MUC1 tripartite vaccine. *Proc. Natl. Acad. Sci. USA* **2012**, *109*, 261–266. [CrossRef] [PubMed]
- 48. Ingale, S.; Buskas, T.; Boons, G.-J. Synthesis of Glyco(lipo)peptides by Liposome-Mediated Native Chemical Ligation. *Org. Lett.* **2006**, *8*, 5785–5788. [CrossRef] [PubMed]
- 49. Shi, L.; Cai, H.; Huang, Z.-H.; Sun, Z.-Y.; Chen, Y.-X.; Zhao, Y.-F.; Kunz, H.; Li, Y.-M. Synthetic MUC1 Antitumor Vaccine Candidates with Varied Glycosylation Pattern Bearing R/S-configured Pam3CysSerLys4. *ChemBioChem* **2016**, *17*, 1412–1415. [CrossRef] [PubMed]
- Bessler, W.G.; Cox, M.; Lex, A.; Suhr, B.; Wiesmueller, K.H.; Jung, G. Synthetic lipopeptide analogs of bacterial lipoprotein are potent polyclonal activators for murine B lymphocytes. *J. Immunol.* 1985, 135, 1900–1905. [PubMed]
- Cai, H.; Sun, Z.-Y.; Chen, M.-S.; Zhao, Y.-F.; Kunz, H.; Li, Y.-M. Synthetic multivalent glycopeptide-lipopeptide antitumor vaccines: Impact of the cluster effect on the killing of tumor cells. *Angew. Chem. Int. Ed.* 2014, 53, 1699–1703. [CrossRef] [PubMed]
- 52. Renaudet, O.; Dasgupta, G.; Bettahi, I.; Shi, A.; Nesburn, A.B.; Dumy, P.; BenMohamed, L. Linear and branched glyco-lipopeptide vaccines follow distinct cross-presentation pathways and generate different magnitudes of antitumor immunity. *PLoS ONE* **2010**, *5*, e11216. [CrossRef] [PubMed]

- 53. Grigalevicius, S.; Chierici, S.; Renaudet, O.; Lo-Man, R.; Deriaud, E.; Leclerc, C.; Dumy, P. Chemoselective Assembly and Immunological Evaluation of Multiepitopic Glycoconjugates Bearing Clustered Tn Antigen as Synthetic Anticancer Vaccines. *Bioconjug. Chem.* **2005**, *16*, 1149–1159. [CrossRef] [PubMed]
- Dumy, P.; Eggleston, I.M.; Cervigni, S.; Sila, U.; Sun, X.; Mutter, M. A convenient synthesis of cyclic peptides as regioselectively addressable functionalized templates (RAFT). *Tetrahedron Lett.* 1995, 36, 1255–1258. [CrossRef]
- 55. Hewitt, M.C.; Seeberger, P.H. Solution and Solid-Support Synthesis of a Potential Leishmaniasis Carbohydrate Vaccine. *J. Org. Chem.* **2001**, *66*, 4233–4243. [CrossRef] [PubMed]
- Muhlradt, P.F.; Kiess, M.; Meyer, H.; Sussmuth, R.; Jung, G. Structure and specific activity of macrophage-stimulating lipopeptides from Mycoplasma hyorhinis. *Infect. Immun.* 1998, 66, 4804–4810. [PubMed]
- Huynh, A.S.; Chung, W.J.; Cho, H.-I.; Moberg, V.E.; Celis, E.; Morse, D.L.; Vagner, J. Novel Toll-like Receptor 2 Ligands for Targeted Pancreatic Cancer Imaging and Immunotherapy. J. Med. Chem. 2012, 55, 9751–9762. [CrossRef] [PubMed]
- 58. McDonald, D.M.; Wilkinson, B.L.; Corcilius, L.; Thaysen-Andersen, M.; Byrne, S.N.; Payne, R.J. Synthesis and immunological evaluation of self-adjuvanting MUC1-macrophage activating lipopeptide 2 conjugate vaccine candidates. *Chem. Commun.* **2014**, *50*, 10273–10276. [CrossRef] [PubMed]
- Wilkinson, B.L.; Day, S.; Malins, L.R.; Apostolopoulos, V.; Payne, R.J. Self-Adjuvanting Multicomponent Cancer Vaccine Candidates Combining Per-Glycosylated MUC1 Glycopeptides and the Toll-like Receptor 2 Agonist Pam3CysSer. *Angew. Chem. Int. Ed.* 2011, 50, 1635–1639. [CrossRef] [PubMed]
- Wilkinson, B.L.; Day, S.; Chapman, R.; Perrier, S.; Apostolopoulos, V.; Payne, R.J. Synthesis and Immunological Evaluation of Self-Assembling and Self-Adjuvanting Tricomponent Glycopeptide Cancer-Vaccine Candidates. *Chem. Eur. J.* 2012, *18*, 16540–16548. [CrossRef] [PubMed]
- Zhu, X.; Ramos, T.V.; Gras-Masse, H.; Kaplan, B.E.; BenMohamed, L. Lipopeptide epitopes extended by an Nε-palmitoyl-lysine moiety increase uptake and maturation of dendritic cells through a Toll-like receptor-2 pathway and trigger a Th1-dependent protective immunity. *Eur. J. Immunol.* 2004, 34, 3102–3114. [CrossRef] [PubMed]
- 62. Kaiser, A.; Gaidzik, N.; Becker, T.; Menge, C.; Groh, K.; Cai, H.; Li, Y.-M.; Gerlitzki, B.; Schmitt, E.; Kunz, H. Fully Synthetic Vaccines Consisting of Tumor-Associated MUC1 Glycopeptides and a Lipopeptide Ligand of the Toll-like Receptor 2. *Angew. Chem. Int. Ed.* **2010**, *49*, 3688–3692. [CrossRef] [PubMed]
- Dziadek, S.; Griesinger, C.; Kunz, H.; Reinscheid, U.M. Synthesis and structural model of an α(2,6)-sialyl-T glycosylated MUC1 eicosapeptide under physiological conditions. *Chem. Eur. J.* 2006, *12*, 4981–4993. [CrossRef] [PubMed]
- 64. Fontenot, J.D.; Tjandra, N.; Bu, D.; Ho, C.; Montelaro, R.C.; Finn, O.J. Biophysical characterization of one-, two-, and three-tandem repeats of human mucin (muc-1) protein core. *Cancer Res.* **1993**, *53*, 5386–5394. [PubMed]
- 65. Mollick Joseph, A.; Hodi, F.S.; Soiffer Robert, J.; Nadler Lee, M.; Dranoff, G. MUC1-like tandem repeat proteins are broadly immunogenic in cancer patients. *Cancer Immun.* **2003**, *3*, 3.
- Jin, M.S.; Kim, S.E.; Heo, J.Y.; Lee, M.E.; Kim, H.M.; Paik, S.-G.; Lee, H.; Lee, J.-O. Crystal structure of the TLR1-TLR2 heterodimer induced by binding of a tri-acylated lipopeptide. *Cell* 2007, 130, 1071–1082. [CrossRef] [PubMed]
- 67. Seyberth, T.; Voss, S.; Brock, R.; Wiesmueller, K.-H.; Jung, G. Lipolanthionine Peptides Act as Inhibitors of TLR2-Mediated IL-8 Secretion. Synthesis and Structure-Activity Relationships. *J. Med. Chem.* **2006**, *49*, 1754–1765. [CrossRef] [PubMed]
- Wu, W.; Li, R.; Malladi, S.S.; Warshakoon, H.J.; Kimbrell, M.R.; Amolins, M.W.; Ukani, R.; Datta, A.; David, S.A. Structure-Activity Relationships in Toll-like Receptor-2 Agonistic Diacylthioglycerol Lipopeptides. J. Med. Chem. 2010, 53, 3198–3213. [CrossRef] [PubMed]
- 69. Zeng, W.-G.; Eriksson, E.; Chua, B.; Grollo, L.; Jackson, D.C. Structural requirement for the agonist activity of the TLR2 ligand Pam2Cys. *Amino Acids* **2010**, *39*, 471–480. [CrossRef] [PubMed]
- 70. Willems, M.M.J.H.P.; Zom, G.G.; Khan, S.; Meeuwenoord, N.; Melief, C.J.M.; van der Stelt, M.; Overkleeft, H.S.; Codee, J.D.C.; van der Marel, G.A.; Ossendorp, F.; et al. *N*-Tetradecylcarbamyl Lipopeptides as Novel Agonists for Toll-like Receptor 2. *J. Med. Chem.* **2014**, *57*, 6873–6878. [CrossRef] [PubMed]

- Guo, X.; Wu, N.; Liu, X.; Wu, T.; Zhou, Y.; Huang, J.; Wu, L.; Shang, Y.; Liu, X.; Liao, X. The Novel Toll-Like Receptor 2 Agonist SUP3 Enhances Antigen Presentation and T Cell Activation by Dendritic Cells. *Front. Immunol.* 2017, *8*, 158. [CrossRef] [PubMed]
- 72. Zeng, W.; Ghosh, S.; Lau Yuk, F.; Brown Lorena, E.; Jackson David, C. Highly immunogenic and totally synthetic lipopeptides as self-adjuvanting immunocontraceptive vaccines. *J. Immunol.* **2002**, *169*, 4905–4912. [CrossRef] [PubMed]
- Zaman, M.; Abdel-Aal, A.-B.M.; Phillipps, K.S.M.; Fujita, Y.; Good, M.F.; Toth, I. Structure-activity relationship of lipopeptide Group A streptococcus (GAS) vaccine candidates on toll-like receptor 2. *Vaccine* 2010, *28*, 2243–2248. [CrossRef] [PubMed]
- 74. McDonald, D.M.; Byrne, S.N.; Payne, R.J. Synthetic self-adjuvanting glycopeptide cancer vaccines. *Front. Chem.* **2015**, *3*, 60. [CrossRef] [PubMed]
- 75. Buskas, T.; Thompson, P.; Boons, G.-J. Immunotherapy for cancer: Synthetic carbohydrate-based vaccines. *Chem. Commun.* **2009**, 5335–5349. [CrossRef] [PubMed]
- 76. Poltorak, A.; He, X.; Smirnova, I.; Liu, M.-Y.; Van Huffel, C.; Du, X.; Birdwell, D.; Alejos, E.; Silva, M.; Galanos, C.; et al. Defective LPS signaling in C3H/HeJ and C57BL/10ScCr mice: Mutations in Tlr4 gene. *Science* 1998, 282, 2085–2088. [CrossRef] [PubMed]
- 77. Akira, S.; Takeda, K. Toll-like receptor signalling. Nat. Rev. Immunol. 2004, 4, 499–511. [CrossRef] [PubMed]
- Fang, H.; Wu, Y.; Huang, X.; Wang, W.; Bing, A.; Cao, X.; Wan, T. Toll-like Receptor 4 (TLR4) Is Essential for Hsp70-like Protein 1 (HSP70L1) to Activate Dendritic Cells and Induce Th1 Response. *J. Biol. Chem.* 2011, 286, 30393–30400. [CrossRef] [PubMed]
- Hutchinson, M.R.; Zhang, Y.; Shridhar, M.; Evans, J.H.; Buchanan, M.M.; Zhao, T.X.; Slivka, P.F.; Coats, B.D.; Rezvani, N.; Wieseler, J.; et al. Evidence that opioids may have toll-like receptor 4 and MD-2 effects. *Brain Behav. Immun.* 2010, 24, 83–95. [CrossRef] [PubMed]
- Peana, M.; Zdyb, K.; Medici, S.; Pelucelli, A.; Simula, G.; Gumienna-Kontecka, E.; Zoroddu, M.A. Ni(II) interaction with a peptide model of the human TLR4 ectodomain. *J. Trace Elem. Med. Biol.* 2017, 44, 151–160. [CrossRef] [PubMed]
- 81. Qureshi, N.; Takayama, K.; Ribi, E. Purification and structural determination of nontoxic lipid A obtained from the lipopolysaccharide of Salmonella typhimurium. *J. Biol. Chem.* **1982**, *257*, 11808–11815. [PubMed]
- Boland, G.; Beran, J.; Lievens, M.; Sasadeusz, J.; Dentico, P.; Nothdurft, H.; Zuckerman, J.N.; Genton, B.; Steffen, R.; Loutan, L.; et al. Safety and immunogenicity profile of an experimental hepatitis B vaccine adjuvanted with AS04. *Vaccine* 2004, 23, 316–320. [CrossRef] [PubMed]
- Goepfert, P.A.; Tomaras, G.D.; Horton, H.; Montefiori, D.; Ferrari, G.; Deers, M.; Voss, G.; Koutsoukos, M.; Pedneault, L.; Vandepapeliere, P.; et al. Durable HIV-1 antibody and T-cell responses elicited by an adjuvanted multi-protein recombinant vaccine in uninfected human volunteers. *Vaccine* 2007, 25, 510–518. [CrossRef] [PubMed]
- Bojang, K.A.; Milligan, P.J.M.; Pinder, M.; Vigneron, L.; Alloueche, A.; Kester, K.E.; Ballou, W.R.; Conway, D.J.; Reece, W.H.H.; Gothard, P.; et al. Efficacy of RTS,S/AS02 malaria vaccine against Plasmodium falciparum infection in semi-immune adult men in Gambia: A randomized trial. *Lancet* 2001, 358, 1927–1934. [CrossRef]
- 85. Elliott, G.T.; McLeod, R.A.; Perez, J.; Von Eschen, K.B. Interim results of a phase II multicenter clinical trial evaluating the activity of a therapeutic allogeneic melanoma vaccine (theraccine) in the treatment of disseminated malignant melanoma. *Semin. Surg. Oncol.* **1993**, *9*, 264–272. [PubMed]
- Sosman, J.A.; Unger, J.M.; Liu, P.Y.; Flaherty, L.E.; Park, M.S.; Kempf, R.A.; Thompson, J.A.; Terasaki, P.I.; Sondak, V.K. Adjuvant immunotherapy of resected, intermediate-thickness, node-negative melanoma with an allogeneic tumor vaccine: Impact of HLA class I antigen expression on outcome. *J. Clin. Oncol.* 2002, 20, 2067–2075. [CrossRef] [PubMed]
- Miles, D.W.; Towlson, K.E.; Graham, R.; Reddish, M.; Longenecker, B.M.; Taylor-Papadimitriou, J.; Rubens, R.D. A randomized phase II study of sialyl-Tn and DETOX-B adjuvant with or without cyclophosphamide pretreatment for the active specific immunotherapy of breast cancer. *Br. J. Cancer* 1996, 74, 1292–1296. [CrossRef] [PubMed]
- Butts, C.; Murray, N.; Maksymiuk, A.; Goss, G.; Marshall, E.; Soulieres, D.; Cormier, Y.; Ellis, P.; Price, A.; Sawhney, R.; et al. Randomized phase IIB trial of BLP25 liposome vaccine in stage IIIB and IV non-small-cell lung cancer. J. Clin. Oncol. 2005, 23, 6674–6681. [CrossRef] [PubMed]

- 89. North, S.A.; Graham, K.; Bodnar, D.; Venner, P. A pilot study of the liposomal MUC1 vaccine BLP25 in prostate specific antigen failures after radical prostatectomy. *J. Urol.* **2006**, *176*, 91–95. [CrossRef]
- Persing, D.H.; Coler, R.N.; Lacy, M.J.; Johnson, D.A.; Baldridge, J.R.; Hershberg, R.M.; Reed, S.G. Taking toll: Lipid A mimetics as adjuvants and immunomodulators. *Trends Microbiol.* 2002, 10 (Suppl. 10), s32–s37. [CrossRef]
- 91. Dupont, J.; Altclas, J.; Lepetic, A.; Lombardo, M.; Vazquez, V.; Salgueira, C.; Seigelchifer, M.; Arndtz, N.; Antunez, E.; von Eschen, K.; et al. A controlled clinical trial comparing the safety and immunogenicity of a new adjuvanted hepatitis B vaccine with a standard hepatitis B vaccine. *Vaccine* **2006**, *24*, 7167–7174. [CrossRef] [PubMed]
- 92. Gao, J.; Guo, Z. Progress in the synthesis and biological evaluation of lipid A and its derivatives. *Med. Res. Rev.* **2018**, *38*, 556–601. [CrossRef] [PubMed]
- Wang, Q.; Xue, J.; Guo, Z. Synthesis of a monophosphoryl lipid A derivative and its conjugation to a modified form of a tumor-associated carbohydrate antigen GM3. *Chem. Commun.* 2009, 5536–5537. [CrossRef] [PubMed]
- Wang, Q.; Zhou, Z.; Tang, S.; Guo, Z. Carbohydrate-Monophosphoryl Lipid A Conjugates Are Fully Synthetic Self-Adjuvanting Cancer Vaccines Eliciting Robust Immune Responses in the Mouse. ACS Chem. Biol. 2012, 7, 235–240. [CrossRef] [PubMed]
- 95. Zhou, Z.; Mondal, M.; Liao, G.; Guo, Z. Synthesis and evaluation of monophosphoryl lipid A derivatives as fully synthetic self-adjuvanting glycoconjugate cancer vaccine carriers. *Org. Biomol. Chem.* **2014**, *12*, 3238–3245. [CrossRef] [PubMed]
- Tang, S.; Wang, Q.; Guo, Z. Synthesis of a Monophosphoryl Derivative of Escherichia coli Lipid A and Its Efficient Coupling to a Tumor-Associated Carbohydrate Antigen. *Chem. Eur. J.* 2010, *16*, 1319–1325. [CrossRef] [PubMed]
- 97. Danishefsky, S.J.; Shue, Y.-K.; Chang, M.N.; Wong, C.-H. Development of Globo-H cancer vaccine. *Acc. Chem. Res.* **2015**, *48*, 643–652. [CrossRef] [PubMed]
- Zhou, Z.; Liao, G.; Mandal, S.S.; Suryawanshi, S.; Guo, Z. A fully synthetic self-adjuvanting globo H-Based vaccine elicited strong T cell-mediated antitumor immunity. *Chem. Sci.* 2015, *6*, 7112–7121. [CrossRef] [PubMed]
- 99. Cluff Christopher, W. Monophosphoryl lipid A (MPL) as an adjuvant for anti-cancer vaccines: Clinical results. *Adv. Exp. Med. Biol.* **2010**, *667*, 111–123.
- 100. Liao, G.; Zhou, Z.; Suryawanshi, S.; Mondal, M.A.; Guo, Z. Fully Synthetic Self-Adjuvanting α-2,9-Oligosialic Acid Based Conjugate Vaccines against Group C Meningitis. ACS Cent. Sci. 2016, 2, 210–218. [CrossRef] [PubMed]
- Wang, L.; Feng, S.; Wang, S.; Li, H.; Gu, G.; Guo, Z. Synthesis and Immunological Comparison of Differently Linked Lipoarabinomannan Oligosaccharide-Monophosphoryl Lipid A Conjugates as Antituberculosis Vaccines. J. Org. Chem. 2017, 82, 12085–12096. [CrossRef] [PubMed]
- 102. Zhou, Z.; Mandal Satadru, S.; Liao, G.; Guo, J.; Guo, Z.; Liao, G.; Guo, J.; Guo, Z. Synthesis and Evaluation of GM2-Monophosphoryl Lipid A Conjugate as a Fully Synthetic Self-Adjuvant Cancer Vaccine. *Sci. Rep.* 2017, 7, 11403. [CrossRef] [PubMed]
- 103. D'Alonzo, D.; Cipolletti, M.; Tarantino, G.; Ziaco, M.; Pieretti, G.; Iadonisi, A.; Palumbo, G.; Alfano, A.; Giuliano, M.; De Rosa, M.; et al. A Semisynthetic Approach to New Immunoadjuvant Candidates: Site-Selective Chemical Manipulation of Escherichia coli Monophosphoryl Lipid A. *Chem. Eur. J.* 2016, 22, 11053–11063. [CrossRef] [PubMed]
- 104. Ziaco, M.; Gorska, S.; Traboni, S.; Razim, A.; Casillo, A.; Iadonisi, A.; Gamian, A.; Corsaro, M.M.; Bedini, E. Development of Clickable Monophosphoryl Lipid A Derivatives toward Semisynthetic Conjugates with Tumor-Associated Carbohydrate Antigens. J. Med. Chem. 2017, 60, 9757–9768. [CrossRef] [PubMed]
- 105. Miyajima, K.; Nekado, T.; Ikeda, K.; Achiwa, K. Synthesis of Tn and sialyl Tn antigen-lipid A analog conjugates for synthetic vaccines. *Chem. Pharm. Bull.* **1997**, *45*, 1544–1546. [CrossRef] [PubMed]
- 106. Miyajima, K.; Gomi, N.; Ikeda, K.; Achiwa, K. Lipid A and related compounds. XXXIII. Synthesis and structure-activity relationships of *N*-acylated L-serine or L-threonine-containing D-glucosamine derivatives as mimics of lipid A disaccharide. *Chem. Pharm. Bull.* **1997**, *45*, 1089–1093. [CrossRef]
- Lewicky, J.D.; Ulanova, M.; Jiang, Z.-H. Synthesis and immunostimulatory activity of diethanolamine-containing lipid A mimics. *RSC Adv.* 2012, 2, 1917–1926. [CrossRef]

- Lewicky, J.D.; Ulanova, M.; Jiang, Z.-H. Synthesis of a TLR4 Agonist-Carbohydrate Antigen Conjugate As A Self-Adjuvanting Cancer Vaccine. *ChemistrySelect* 2016, 1, 906–910. [CrossRef]
- 109. Glaffig, M.; Stergiou, N.; Schmitt, E.; Kunz, H. Immunogenicity of a fully synthetic MUC1 glycopeptide antitumor vaccine enhanced by poly(I:C) as a TLR3-activating adjuvant. *ChemMedChem* 2017, 12, 722–727. [CrossRef] [PubMed]
- 110. Abdel-Aal, A.-B.M.; Lakshminarayanan, V.; Thompson, P.; Supekar, N.; Bradley, J.M.; Wolfert, M.A.; Cohen, P.A.; Gendler, S.J.; Boons, G.-J. Immune and Anticancer Responses Elicited by Fully Synthetic Aberrantly Glycosylated MUC1 Tripartite Vaccines Modified by a TLR2 or TLR9 Agonist. *ChemBioChem* 2014, 15, 1508–1513. [CrossRef] [PubMed]
- 111. Hartmann, S.; Nuhn, L.; Palitzsch, B.; Glaffig, M.; Stergiou, N.; Gerlitzki, B.; Schmitt, E.; Kunz, H.; Zentel, R. CpG-Loaded Multifunctional Cationic Nanohydrogel Particles as Self-Adjuvanting Glycopeptide Antitumor Vaccines. *Adv. Healthc. Mater.* 2015, *4*, 522–527. [CrossRef] [PubMed]



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