Tumor promoting roles of IL-10, TGF- β , IL-4, and IL-35: Its implications in cancer immunotherapy

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Abstract

Cytokines play a critical role in regulating host immune response toward cancer and determining the overall fate of tumorigenesis. The tumor microenvironment is dominated mainly by immune-suppressive cytokines that control effector antitumor immunity and promote survival and the proliferation of cancer cells, which ultimately leads to enhanced tumor growth. In addition to tumor cells, the heterogeneous immune cells present within the tumor milieu are the significant source of immune-suppressive cytokines. These cytokines are classified into a broad range; however, in most tumor types, the interleukin-10, transforming growth factor- β , interleukin-4, and interleukin-35 are consistently reported as immunesuppressive cytokines that help tumor growth and metastasis. The most emerging concern in cancer treatment is hijacking and restraining the activity of antitumor immune cells in the tumor niche due to a highly immune-suppressive environment. This review summarizes the role and precise functions of interleukin-10, transforming growth factor- β , interleukin-4, and interleukin-35 in modulating tumor immune contexture and its implication in developing effective immune-therapeutic approaches.

Concise conclusion

Recent effort geared toward developing novel immune-therapeutic approaches faces significant challenges due to sustained mutations in tumor cells and a highly immune-suppressive microenvironment present within the tumor milieu. The cytokines play a crucial role in developing an immune-suppressive environment that ultimately dictates the fate of tumorigenesis. This review critically covers the novel aspects of predominant immune-suppressive cytokines such as interleukin-10, transforming growth factor- β , interleukin-4, and interleukin-35 in dictating the fate of tumorigenesis and how targeting these cytokines can help the development of better immune-therapeutic drug regimens for the treatment of cancer.

Keywords

Immune-suppressive cytokines, tumor microenvironment, cancer immunotherapy, antitumor immunity, tumor growth

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Introduction

Immune-suppressive cytokines are well known to play an essential role in controlling effector immune responses required to prevent autoimmune and inflammatory reactions.^{1,2} The functions and roles of immune-suppressive cytokines in autoimmune/infectious diseases are well characterized. These cytokines are beneficial in one arm and control the severity of autoimmune and inflammatory reactions; in the second arm, they play a negative role by damaging host effector antitumor immunity and making the environment favorable for tumor growth and metastasis.^{3,4} The altered epigenetic and growth kinetics in cancer cell leads to the extracellular secretion of several factors that

control the activity of immune cells and makes them favorable for tumor growth.5,6 Among these, secretion of immuneregulatory cytokines is the primary factor for recruiting immune-suppressive cells.⁷ The immune-regulatory cytokines in the tumor microenvironment recruit other immunesuppressive cells and are responsible for phenotypical and

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functional switch of effector immune cells, and make them supportive for tumor development.^{8,9} The phenotypically switched immune cells further elevate the level of immunesuppressive cytokines and make the tumor environment resistant to the activity of antitumor immune cells.^{10,11} These immune-suppressive cells mainly include MDSCs (myeloid-derived suppressor cells), TAMs (tumor-associated macrophages), regulatory T (Treg) cells, regulatory B (Breg) cells, and TANs (tumor-associated neutrophils).^{12,13} Although several other types of immune cells such as Th17, Th2, and dendritic cells may act as immune-suppressive in certain cancers, however, MDSCs, TAMs, Treg, and Breg cells are the prominent source of regulatory cytokines and dominate the tumor milieu.¹⁴⁻¹⁷ These cells exert their suppressive functions via several mechanisms, but the secretion of immune-regulatory cytokines is a significant contributor to immune suppression among several other mechanisms.^{2,9} These immune-regulatory cytokines show tumor-promoting characteristics by regulating intracellular signaling in cancer cells and immune cells present within the tumor milieu by an autocrine or positive feedback loop.^{13,18}

The immune-suppressive interleukin (IL)-10, transforming growth factor- β (TGF- β), IL-4, and IL-35 have distinct physiological effects. These cytokines alter the fate of innate and adaptive immune cells and regulate the growth and proliferative capacity of cancer cells. Furthermore, the regulatory IL-10 and TGF-B drive and maintain the immune-suppressive phenotype of T cells by diminishing the effector Th1, Th2, and Th17 phenotypes,^{19,20} while IL-4 predominantly favors the Th2 phenotype of T cells.²¹ The IL-35 was shown to have major effects on T cells and B cells, where it converts conventional T cells into IL-35, producing immune-suppressive T cells by inhibiting Th1 and Th17, and effector phenotype of T cells.^{22,23} These cytokines are secreted by diverse immune populations, including cells from innate and adaptive immune systems. In certain scenarios, tumor cells might secret the IL-10, TGF- β , IL-4, and IL-35, and drive the immune-suppressive fate of tumor niche.^{24–26} Generally, IL-10, TGF-β, and IL-4 share similar receptor patterns across cancer cells, and immune cells, except IL-35 uses different receptor patterns depending on the cell type. IL-10 signals through IL-10R α and IL-10R β , TGF- β signals through TGF-BRI and TGF-BRII, IL-4 signals through IL-4R α , and IL-35 signals through IL-12R β 2 and gp130 in case of T cells and IL-12R β 2 and IL-27R α in case of B cells.^{27–31} The downstream signaling of these receptors activates phosphorylation of signal transducer and activator of transcription (STAT) proteins and different transcription factors, which further dictates immune suppression and tumorigenesis.

The immune-suppressive cytokines alter the epigenome and transcriptional network in naïve immune cells and favor immune-suppressive cell lineage commitment.^{8,17} Furthermore, these cytokines can directly act on cytotoxic CD8 T cells or natural killer (NK) cells and restrain their effector functions and proliferation into the tumor microenvironment.^{32,33} In addition, immune-suppressive cytokines directly modulate the tumor cell growth and favor their survival and proliferation.^{2,34,35} The immune-suppressive cytokines are classified into a broad range and secreted by various cells, including tumor cells, epithelial cells, innate immune cells, and adaptive immune cells. Among the wide

immune cells, and adaptive immune cells. Among the wide range of immune-suppressive cytokines, IL-10, TGF- β , IL-4, and IL-35 dominate the tumor environment in multiple cancers and are mainly responsible for immune suppression^{36–41} (Figure 1). Herein we reviewed the recent updates on essential functions of these cytokines in modulating tumor microenvironment and how targeting these cytokines can limit cancer cell survival and activate the antitumor immunity that helps to control tumor growth and metastasis.

Immune-suppressive cytokines IL-10, TGF- β , IL-4, and IL-35 in cancer and immunotherapy

IL-10

IL-10 is a multifaceted immune-suppressive cytokine and possesses immune-regulatory and angiogenic functions.⁴² IL-10 enhances tumor cell survival, proliferation, and metastasis by controlling antitumor immunity.43 IL-10 is secreted by cancer cells and several immune cells, including myeloid and lymphoid lineages. IL-10 exerts its suppressive effects on other effector immune cells, including potent antitumor cytotoxic NK and CD8 T cells.^{20,44} NK cells and CD8 T cells are essential for killing cancer cells and controlling tumor growth. Recent studies showed that the neutralization of the effects of IL-10 induces the tumorspecific cytotoxic immune response. The Cetuximab-based IL-10 fusion protein (CmAb-(IL10)₂) demonstrates potent antitumor effects by preventing dendritic cell-mediated apoptosis of tumor-infiltrating CD8 T cells. Combination of CmAb-(IL10), with immune checkpoint blockade showed improved antitumor immunity in mice with advanced tumors.45

Further studies by Naing et al. showed that PEGylated IL-10 induces CD8 T-cell-mediated immune response in cancer patients by elevating their proliferation and tumor infiltration, and their interferon-gamma (IFN- γ) and granzyme B production. In addition, a combination of PEGylated IL-10 with immune checkpoint blockade enhances intratumoral expansion of CD8 T cells.⁴⁶ Similar studies confirmed that the PEGylated IL-10 treatment was shown to restore tumor-specific CD8 T cell response and attenuates tumor growth.⁴⁷ In agreement with the role of IL-10 in inhibiting tumor-specific cytotoxic immune



Figure 1. Immune-suppressive IL-10, TGF- β , IL-4, and IL-35 in promoting tumor growth. IL: interleukin; TGF- β : transforming growth factor- β .

The immune-suppressive cytokines secreted by immune cells or tumor cells inhibit antitumor immune cell activity in the tumor microenvironment by downregulating their proliferation, intratumoral infiltration, and effector functions. Furthermore, the augmented expression of the immune-suppressive cytokines in the tumor milieu converts the naïve or effector immune cell phenotype to immune-suppressive phenotype that restrains the activity of antitumor immunity and facilitates tumor growth.

response, therapeutic blockade of IL-10 has been shown to have a positive effect on antitumor immune response mediated by CD8 T cells and NK cells. In human breast cancer, therapeutic blockade of IL-10 receptor signaling augments CD8 T-cell-dependent primary tumor response to chemotherapeutic agents such as paclitaxel and carboplatin.⁴⁸ In addition, several studies have demonstrated the inverse correlation between CD8 T cells, poor prognosis, and elevated levels of IL-10 in the tumor microenvironment. In esophageal cancer patients, IL-10 suppresses activation of CD8 T cells, and the level of IL-10 in the tumor microenvironment is elevated in the advanced tumor stage.⁴⁹

In addition, B-cell-secreted IL-10 in patients with differentiated thyroid carcinoma (DTC) suppresses the effector functions of CD4 and CD8 T cells. It increases the frequency of tumor-infiltrating Treg cells.⁵⁰ Similar studies by Hu et al.⁵¹ demonstrated that the B-cell-secreted IL-10 reduces the effector CD4 and CD8 T-cell-mediated immunity by attenuating production of IFN- γ , TNF- α (tumor necrosis factor alpha), and IL-17 and converts these effector T cells into IL-10-producing T cells in patients with gastric carcinoma. In contrast, Zhang et al.⁵² showed the TAM-secreted IL-10 drives an immunoevasive microenvironment that causes poor prognosis and inferior therapeutic responsiveness to adjuvant chemotherapy in gastric cancer patients, indicating IL-10 TAMs can be used as a potential target for gastric cancer. In addition, IL-10 limits CD8 T cell activation and function via modulation of cell surface glycosylation.⁵³ Likewise, Li et al. reported the T follicular regulatory (Tfr) cell-secreted IL-10 was negatively correlated with tumor-infiltrating CD8⁺IFN- γ^+ T cells in ovarian cancer (OC) patients. The in vitro co-culture of CD8 and Tfr cells revealed Tfr cells suppress the activation of CD8 T cells in IL-10 but not TGF- β -dependent manner. The frequency of Tfr cells was enhanced in peripheral blood samples of OC patients.⁵⁴

Besides the effects of IL-10 in controlling CD8 T-cellmediated immune response, IL-10 can have tumor-promoting effects by augmenting regulatory CD4 and CD8 T cell phenotypes. Mechanistic studies in multiple myeloma patients showed IL-10 augments immune-suppressive activities by favoring generation and expansion of CD8⁺CD28⁻CD57⁺LFA-1^{high} Treg. These CD8 Treg cells contribute to the immune escape mechanism of tumors in multiple myeloma patients.⁵⁵ Furthermore, IL-10 was also shown to regulate negative receptor expression on CD8 T cells. Liu et al. observed the high infiltration of T cell immunoglobulin and immunoreceptor tyrosine-based inhibitory motif (ITIM) domain (TIGIT) expressing CD8 T cells resulted in poor survival of muscle-invasive bladder cancer (MIBC) patients. Mechanistically, TIGIT⁺CD8⁺ T cells impaired cytotoxic activity of effector CD8 T cells by producing immune-suppressive cytokine IL-10 and overall promotes suppressive tumor immune contexture.⁵⁶ Likewise, IL-10 was shown to upregulate the expression of lymphocyte activation gene-3 (LAG-3) and programmed cell death protein-1 (PD-1) on CD8 T cells and impaired the production of effector cytokines IFN- γ and TNF- α in patients with epithelial OC. Dual blockade of IL-10 with LAG-3 and PD-1 induces the proliferation and cytokine production of tumor-specific CD8 T cells.⁵⁷ In addition, Tannir et al.58,59 revealed pegilodecakin (pegylated recombinant human IL-10) with immune checkpoint blockade

induces intratumoral effector IFN- γ^+ CD8 T cell response and showed promising clinical activity in patients with metastatic renal cell carcinoma. This study demonstrates the combinational treatment targeting IL-10 with immune checkpoint blockade has notable clinical benefits.

Recently, the immune-modulatory effects of IL-10 on antitumor NK cells were well characterized. In pancreatic ductal adenocarcinoma (PDAC) patients, CD16^{hi}CD57^{hi} NK cells produce high-level IL-10, hindering their cytotoxic activity and IFN-y expression. The NK cells secreting IL-10 were abundantly found in the blood of PDAC patients, indicating possible options for NK-based immunotherapy for the treatment of PDAC.⁶⁰ Sun et al. observed that in patients with hepatocellular carcinoma (HCC), IL-10 enhances expression of NKG2A in NK cells and the NKG2A ligand, HLA-E, in intratumor HCC tissues. These NKG2A expressing NK cells were functionally exhausted and associated with a poor prognosis. Moreover, IL-10 blockade specifically inhibits NKG2A expression in NK cells and restores immune response against tumor cells by reversing NK-cell exhaustion.⁶¹ Similar studies in HCC patients showed CD96⁺ NK cells were functionally exhausted with diminished IFN- γ and TNF- α secretion due to high expression of IL-10 and exhibit poorer clinical outcomes.⁶² Ostapchuk et al.⁶³ showed IL-10 activates immune-regulatory functions of NK cells, which impair the efficiency of antitumor immunity in breast cancer patients. These observations indicate IL-10 has diverse suppressive effects on antitumor immune cells, required for tumor cell clearance. Therefore, targeting IL-10 in the tumor microenvironment alone or combined with conventional chemotherapeutic or immunotherapeutic agents has substantial clinical benefits for cancer treatment.

$TGF-\beta$

The TGF- β is an immune-suppressive cytokine that mainly exerts tumor immune-suppressive effects, resulting in tumor development.^{28,64} TGF- β is required for the development and lineage-specific differentiation of Treg cells that ultimately leads to developing a highly immune-suppressive tumor environment.65,66 Recent reports showed the different mechanisms of TGF- β overexpression in the tumor environment that restrains effector antitumor immunity. By blocking the activity of histone demethylase LSD1 (lysine-specific demethylase) in tumor cells, Sheng et al. found that the intensity and percentage of TGF- β were significantly higher, inhibiting the cytotoxic activity of intratumoral CD8 T cells. Next, the synergistic depletion of LSD1 and TGF- β in combination with immune checkpoint blockade ameliorates intratumoral CD8 T cell infiltration and effector functions, resulting in eradication of tumor and long-term protection from tumor re-challenge.67

In parallel, Marwitz et al. showed the TGF-B downstream-activated SMAD3 phosphorylation in CD8 T cells and Treg cells negatively impacts the overall survival of lung cancer patients. In a rapidly progressing group of lung cancer patients, the Treg cells expressing high pSMAD3 are near CD8 T cells, which indicates TGF-β pathway activation in tumor-infiltrating immune cells influences survival of lung cancer patients.68 Similarly, a recent study reported that the ablation of TGF-B downstream signaling in CD4 T cells restricts tumor progression by enhancing Th2 cell phenotype and causing the death of cancer cells in distant avascular regions.⁶⁹ Given that TGF- β is essential for sustained immune-suppressive Treg cells, enhanced immune suppression in the tumor niche can resist the immune checkpoint blockade treatment. An argument with this de Streel et al. revealed TGF-β secreted by GARP⁺ (glycoprotein-A repetitions predominant) Treg cells in human metastatic melanoma induces resistance to anti-PD-1 immunotherapy. The selective blockade of TGF-β isoform with anti-GARP:TGF-β1 monoclonal antibodies (mAbs) overcomes resistance to PD-1/Programmed death-ligand 1 (PD-L1) blockade in cancer patients.⁷⁰

In patients with esophageal squamous cell carcinoma, MDSC-secreted TGF-B augments PD-1 expression on CD8 T cells and develops resistance toward PD-1/PD-L1 blockade. Combinatorial therapy of PD-1/PD-L1 and TGF-B blockade restored tumor-antigen-specific CD8 T cells.⁷¹ Consistently, TGF-β blockade and immune checkpoint blockade boost Th1 polarization and increase the clonal expansion of CD8 T cells, improving the survival of patients with metastatic castration-resistant prostate cancer.72 Similar studies show the combinatorial blockade of PD-L1 and TGF- β enhances the intratumoral infiltration of CD8⁺ T cells and improves the survival of mice with colon and pancreatic adenocarcinoma.⁷³ An alternative study by Liu et al.74 shows dual blockade of TGF-B1 and GM-CSF (granulocyte-macrophage colony-stimulating factor) improves the effects of chemotherapy treatment by preventing polarization of TAMs toward immune-suppressive M2 phenotype and induces response of CD8 T cells in pancreatic cancer patients. The TAN polarization was known to regulate by TGF-β. TGF-β promotes the immunesuppressive N2 phenotype of TANs, and blockade of TGFβ results in enhanced infiltration and activation of the antitumor phenotype of TANs.75

Recent reports showed the relationship between TGF- β and the expression of negative receptors on immune cells. Li et al. observed osteopontin-induced TGF- β and upregulated PD-1 expression on immune cells. In patients with advanced triple-negative breast cancer (TNBC), the inhibition of PD-1 signaling impedes TGF- β expression and improves the treatment responses.⁷⁶ In addition, TGF- β downstream-activated SMAD2/3 induces PD-1 expression on intratumoral CD8 T cells that resulted in CD8 T cell

dysfunction, exhaustion, and increased tumor growth in gastric cancer patients.⁷⁷ In addition, Yang et al. demonstrated Notch1 signaling in B16 melanoma cells enhances tumor-induced immune suppression by promoting the expression of TGF- β . TGF- β hinders the proliferation and intratumoral infiltration of effector CD4 and CD8 T cells and reduces the level of IFN- γ by upregulating the expression of PD-1.⁷⁸ Thus, targeting the TGF- β -PD-1 axis may be beneficial specifically in combination with immune therapies for cancer treatment.

Several studies have shown the elevated levels of TGF-B associated with cancer patient's survival, and it regulates tumor-specific cytotoxic immune cells. Yu et al.79 showed tumor-cell-secreted TGF-β was inversely correlated with the frequency of CD8 T cells in the tumor niche, and it is associated with a worse prognosis of breast cancer patients. The higher expression of TGF- β was also shown to be inversely correlated with CD8 T cell infiltration in lung cancer patients, and altered expression of TGF- β can predict the therapeutic response to nivolumab.⁸⁰ In parallel, Wu et al. demonstrated TGF-β could induce the tumor-promoting suppressive phenotype of CD8 T cells in OC environment. Moreover, OC patients express an elevated level of TGF- β , and it is positively correlated with the frequency of CD8⁺ Treg cells.⁸¹ Besides, TGF- β was shown to alter the lineage plasticity of CD8 T cells in the tumor environment and make CD8 T cells more susceptible to TGF-β-induced immune suppression.⁸² In addition, in gastric cancer patients, CD8 T cell dysfunction was positively correlated with elevated levels of TGF- β , which can be used as a potential therapeutic marker.⁸³ Alternatively, TGF- β was shown to regulate effector functions of NK cells. SMAD2/3/4 activation downstream of TGF- β signaling suppresses T-bet, a positive regulator of IFN- γ , and plays a vital role in regulating NK-cell-mediated immune functions.⁸⁴ TGF- β is also known to regulate epithelial-mesenchymal transition (EMT) and regulates cancer metastasis.85,86 EMT plays an essential role in tissue repair, and remodeling and acquiring the metastasis phenotype of cancer cells^{87,88} indicates the importance of TGF-β in promoting tumor growth independent of immune cells.

The inhibitory effect of TGF- β on effector functions and cytotoxic activity of NK cells were well characterized. Fujii et al. showed TGF- β -induced suppression of NK-cellcytotoxic function could be inhibited via promoting the IL-15 signaling pathway. Mechanistically, the IL-15 superagonist/IL-15 receptor α fusion complex (IL-15SA/ IL-15RA) suppresses the Smad2/3-induced transcription and rescues cytotoxic gene expression of NK cells.⁸⁹ Recently, the efficacy of Chimeric antigen receptor (CAR) T cells was shown to be affected in the presence of TGF- β . Stüber et al. showed the negative impact of TGF- β on the efficiency of CAR T cells against TNBC. TGF- β reduces effector cytokine production, proliferation, viability, and cytotoxic activity of ROR1-CAR T cells. Furthermore, the kinase inhibitor SD-208 impedes the TGF- β -receptor signaling, protects CD8⁺ ROR1-CAR T cells, and sustains their antitumor function.⁹⁰ These findings highlight the importance of TGF- β in generating and maintaining a highly immune-suppressive tumor environment that restricts the activity of effector immune cells and promotes tumor growth. Therefore, targeting TGF- β alone or with other combinatorial therapies has notable clinical benefits for the treatment of cancer patients.

IL-4

IL-4 is mainly required for lineage-specific differentiation of Th2 cells and regulation of humoral immune responses. In addition to basophils and mast cells, IL-4 is predominantly secreted by Th2 cells via autocrine signaling.^{91,92} IL-4 plays a diverse role in the tumor microenvironment and exhibits immune-suppressive and antitumor activities.^{38,41} Here, we primarily focused on the immune-suppressive role of IL-4 in controlling antitumor immunity and supporting tumor growth. In a mouse model of primary lung carcinogenesis, IL-4 downstream STAT6-activated signaling in CD11b⁺ myeloid cells promotes M2 immune-suppressive phenotype, and triggers IL-4 expression in M2 cells by a positive feed-back loop and enhances lung cancer progression.⁹³

Further studies from Ito et al. revealed that blockade of IL-4 decreases the generation of immune-suppressive M2 phenotype and MDSCs in a specific tumor microenvironment and ameliorates tumor-specific CD8 T cell response in a mouse model of colon and breast cancer. In addition, blockade of IL-4 improves the responsiveness of anti-OX40 immunotherapy.⁹⁴ Similar studies in a mouse model of non-small-cell lung cancer showed blockade of IL-4 augments IL-12 production by tumor-antigen-specific dendritic cells that enhance infiltration and proliferation of effector T cells and reduce tumor burden.⁹⁵ In addition, Joshi et al.⁹⁶ demonstrated depletion of IL-4 augments the antitumor immunity and overall survival without any visible toxicity in mouse models of anaplastic thyroid cancer (ATC).

Several recent reports critically illustrated the significance of IL-4 signaling in the survival and proliferation of cancer cells. Nappo et al.⁹⁷ showed IL-4 downstreaminduced STAT6 signaling is required for increased survival and proliferation pathways, including Akt and NF- κ B (nuclear factor- κ B) pathways in prostate cancer cells. Likewise, IL-4 triggers the mitogen-activated protein kinases (MAPKs) and specifically induces proliferation of A253 (head and neck), MDA-MB-231 (breast), and SKOV-3 (ovarian) cells by c-Jun N-terminal kinase (JNK) activation.⁹⁸ Moreover, Li et al. observed enhanced growth of MCA205 fibrosarcoma in the presence of IL-4. Mechanistically, IL-4 upregulates the expression of IL-4R and anti-apoptotic genes in tumor cells, which is required for the survival of tumor cells. Furthermore, blockade of IL-4 by 11B11 mAb showed a reduction in tumor growth.⁴¹ Besides, Zhao et al.⁹⁹ showed IL-4-TBX21 signaling induces expression of cancer stemness markers and proliferation in lung cancer cells, indicating IL-4 signaling is an essential factor for cancer development in lung adenocarcinoma patients. The CD44^{hi} expressing tumor cells have been revealed to have tumor-initiating characteristics. Erb et al.¹⁰⁰ showed the expression of CD44 on prostate cancer cells was upregulated by IL-4 and increased their proliferation, migration, and clonogenic potential. These studies indicate the role of IL-4 signaling in the survival and proliferation of cancer cells.

The effects of IL-4 gene polymorphisms on cancer risk were well characterized. Jia et al.¹⁰¹ found that IL-4 rs2243250 and rs79071878 polymorphisms are associated with cancer sensitivity. Likewise, Shamoun et al.¹⁰² showed that the IL-4 SNP rs2243250 genetic polymorphism could be a useful prognostic marker for stage III colorectal cancer patients. The IL-4 rs2243250 and variable number tandem repeat (VNTR) polymorphisms were also associated with lung cancer and breast cancer susceptibility;^{103,104} this observation shows the critical relationship between IL-4 polymorphism and tumorigenesis. In contrast, IL-4 was induced upon radiation therapy and contributed to tumor growth and metastasis. Formentini et al.¹⁰⁵ revealed that radiationinduced IL-4 upregulates the expression of IL-4R α in human colorectal cancer cells and facilitates colorectal cancer metastasis. Similarly, Kim et al.¹⁰⁶ observed expression of IL-4 is highly elevated upon radiation in metastatic carcinoma tissues, and it is correlated with poor survival of cancer patients, indicating IL-4 can be a promising target to prevent radiation-induced immune suppression.

Interestingly, several reports demonstrated the altered expression of IL-4 is associated with the severity of cancer, and it regulates diverse immune infiltrates in the tumor milieu. In patients with papillary thyroid carcinoma (PTC), the increased serum level of IL-4 is associated with persistent disease. It is positively correlated with the production of antithyroglobulin antibodies.¹⁰⁷ In addition, tumor-cellsecreted IL-4 was shown to significantly impact effector tumor infiltrates, including TAMs and CD8 T cells. IL-4 was shown to enhance pancreatic-cancer-promoting functions of TAMs by inducing cathepsin protease activity in macrophages.¹⁰⁸ Furthermore, Apte et al.¹⁰⁹ observed tumor-cell-secreted IL-4 elevates expression of GATA3 and type 2 cytokines, which reduces the antitumor immune response mediated by CD8 T cells in a mouse model of thymic lymphoma. Likewise, Noble et al.¹¹⁰ showed IL-4 upregulates the expression of CD39 and IL-10 that induces peripheral tolerance and suppressive activity of tumorinfiltrating CD8 T cells. Moreover, treatment of CD8 T cells with IL-4 downregulates expression of CD127 and contributes to impaired CD8 T cell function and antitumor immune response.¹¹¹ In addition, Olver et al. showed the suppressive effects of tumor-cell-secreted IL-4 on intratumoral CD8 T cells. IL-4 alters the cytotoxic activity of tumor infiltrated CD8 T cells that resulted in delayed primary tumor clearance and increases possibilities for the development of secondary tumors indicating inhibition of IL-4 in the tumor milieu can improve the efficacy of CD8 T cells.¹¹² The role of IL-4 in modulating NK-cell functions is well characterized. Ohayon et al.¹¹³ showed a reduction of IFN- γ and TNF- α by IL-4-treated human NK cells and reduced cytotoxicity toward K562 cells. Likewise, human NK cells exposed to IL-4 differentially regulate and prime naïve T cells toward their regulatory phenotype via monocyte-derived dendritic cells and modulate the NK-cell interaction with Dendritic cells (DCs).¹¹⁴ In addition, IL-4 can reduce the functional ability of NK cells by secreting type-2 cytokine expression among innate cells and positively regulates tumor progression.^{34,115}

These findings indicate IL-4 has a dual role in the tumor microenvironment; it regulates the tumor cell survival, proliferation, and migration, and modulates antitumor immunity. It shows that IL-4 possesses an emerging therapeutic target to reduce tumor-induced immune suppression and boosts the efficacy of conventional cancer therapies. Thus, similar to other immune-suppressive cytokines, IL-4 is a critical regulator of tumor initiation and metastasis and may represent an attractive immune therapy target alone or in combination with currently available treatment regimens for the effective treatment of patients with cancer.

IL-35

IL-35 is a heterodimeric and relatively newly discovered cytokine compared to IL-10, TGF-β, and IL-4, belonging to the IL-12 family cytokine and made up of p35 and EBi3 subunits.^{116,117} IL-35 is well known to regulate adaptive immune responses and control effector T-cell-mediated immune responses in autoimmune diseases.^{118,119} In addition, IL-35 induces immune-suppressive iT35 cells from conventional T cells that control T cell proliferation promote exhaustion of CD8 T cells.^{22,23} Treg-cell-secreted IL-35 in tumor microenvironment enhances intratumoral T cell exhaustion by activating several inhibitory receptor expression and exhaustion-associated gene expression in tumorinfiltrating CD8 T cells and inhibits effective antitumor immunity.¹²⁰ Similar studies show the role of Treg-cellderived IL-35 promotes exhaustion and suppression in non-Treg cells.¹²¹ The expression of IL-35 is highly elevated in mouse and human pancreatic tumor samples and specifically inhibits the activity of CD8 T cells in STAT3-dependent mechanism.^{122,123}

Higher IL-35 levels were also detected in several tumor types, linked with tumor growth and overall survival. Zhu

et al.^{124,125} demonstrated enhanced expression of IL-35 in serum samples of prostate cancer patients was positively associated with metastasis and disease progression. Moreover, IL-35 contributes to prostate cancer progression via increasing cancer cell proliferation and tumor angiogenesis. Mechanistically, IL-35 blocks the infiltration of antitumor immune cells by favoring proliferation and intratumoral infiltration of Treg cells and MDSCs.^{124,125} The expression of IL-35 was also elevated in tumor tissues of patients with non-small cell lung cancer (NSCLC). It is correlated with overall and progression-free survival.¹²⁶ Likewise, the increased level of IL-35 was observed in children with acute lymphoblastic leukemia (ALL), and it is associated with the development of cancer.¹²⁷

The overexpression of IL-35 is associated with the progression of human gastric cancer. Fan et al.¹²⁸ revealed IL-35 is involved in the proliferation of gastric cancer cells by upregulating the expression of Ki67 and reducing apoptosis. Similarly, Gu et al.¹²⁹ observed that the enhanced serum level of IL-35 correlates with disease prognosis in gastric cancer patients. Furthermore, Chen et al. demonstrated the elevated level of IL-35 in serum and tumor tissues of breast cancer patients is associated with tumor progression. IL-35 facilitates the growth of breast cancer cells by augmenting the expression of epidermal growth factor receptor (EGFR), Ki67, and p53 suggests IL-35 is an essential factor for the progression of breast cancer.¹³⁰ Moreover, in colorectal cancer patients, high expression of IL-35 is directly correlated with the severity of malignancy and the clinical stage of the tumor. Mechanistically, the cancer-cell-secreted IL-35 recruits Treg cells in the tumor microenvironment and promotes immune suppression.¹³¹ In addition, the elevated level of IL-35 in diffuse large B-cell lymphoma (DLBCL) is correlated with overall survival in DLBCL patients.¹³² These observations showed that the elevated level of IL-35 is associated with tumor progression, and it can be served as a better prognostic marker. Overall, these findings uncover the potential role of IL-35 in the modulation of host innate and adaptive immune responses that promote tumor growth. Therefore, targeting or blockade of IL-35 has significant therapeutic benefits for the successful treatment of cancer patients.

Conclusion and future perspectives

Cytokines play a crucial role in shaping and dictating the tumor immune microenvironment that decides the fate of tumorigenesis. Several studies showed that the highly immune-suppressive tumor environment is mainly responsible for tumor growth and tumor resistance to multiple chemotherapeutic and immunotherapeutic drugs. For example, the acquired resistance to conventional immune-therapeutic treatment such as immune checkpoint blockade is predominantly due to elevated immune-suppressive cytokines in the tumor milieu.^{133,134} These immune-suppressive cytokines restrict the proliferation and effector functions of antitumor immune cells, alter their plasticity, and convert them into suppressive phenotypes. Due to restricted infiltration or absence of effector immune cells, the immune checkpoint blockades cannot exert optimum effects^{135–137} (Figure 1).

Similarly, multiple radiation therapies are well known to induce immune suppression, primarily due to the secretion of regulatory cytokines either by tumor cells or by immune cells in the tumor niche.¹³⁸⁻¹⁴⁰ Therefore, targeting or depleting these cytokines has significant clinical benefits. Several cytokines play an immune-suppressive role in tumorigenesis depending on the type and tumor immune contexture. However, in this review, we mainly focused on IL-10, TGF-β, IL-4, and IL-35, since several recent studies described above showed their role in limiting antitumor immunity and progression of tumor growth. In addition, the cytokines secreted by tumor cells as well as immune cells such as IL-23, IL-27, IL-39, IL-1β, IL-13, IL-21, and IL-17 are well known to regulate tumor immune contexture and can participate in immune suppression; however, their precise role and importance in contributing tumor growth are out of the scope of this review.

The critical challenge in developing better immunetherapeutic drugs is to combat immune suppression within the tumor milieu. The tumors typically resist the infiltration of immune cells, considered immunologically "cold tumors."¹⁴¹ The immune-suppressive cytokines play a vital role in developing the "cold" nature of the tumor.¹⁴² In addition, tumor-cell-expressed markers and secreted factors such as soluble inhibitory factors, inhibitory receptors/ ligands, and immune-suppressive cytokines/chemokines further facilitate the "cold" nature of the tumor.^{143–145} The overreaching goal is to target these factors alone or combine them with available conventional therapies to achieve maximum tumor clearance. A blockade of immune-suppressive cytokines such as IL-10, TGF-B, IL-4, and IL-35 might play an essential role in making tumors sensitive to conventional therapies. Blockade or neutralizing the activities of these cytokines might convert the immunological "cold" nature of the tumor to a "hot" tumor. The immunological "hot" tumor further facilitates the infiltration and proliferation of antitumor immune cells and makes the tumor more responsive toward conventional immune therapies such as immune checkpoint blockades (Figure 2). Future studies should delineate these cytokines' detailed mechanism of action in regulating immune-suppressive cell phenotype and tumor growth. Additional studies are required to monitor their systemic versus localized effect and check whether these cytokines can induce any adverse effects like toxic or inflammatory reactions. Thus,



Figure 2. Targeting the immune-suppressive IL-10, TGF- β , IL-4, and IL-35 reduces tumor growth.

IL: interleukin; TGF- β : transforming growth factor- β ; PFN: perforins, GzmB: granzyme B.

Neutralizing or specific blockade of IL-10, TGF- β , IL-4, and IL-35 augments the infiltration and effector functions of antitumor immune cells while limiting the activity of immune-suppressive cells in the tumor microenvironment. Therefore, targeting immune-suppressive cytokines promotes tumor cell apoptosis and increases the efficacy of conventional immune or chemotherapeutic agents.

targeting highly abundant immune-suppressive cytokines such as IL-10, TGF- β , IL-4, and IL-35 in the tumor microenvironment considerably helps to revert immune suppression, enhancing the efficacy of conventional cancer therapies and activating antitumor immunity, and controlling tumorigenesis.

Limitations of the study

The initiation and development of cancer is a multistep process. Although immune cells and their secreted immunesuppressive cytokines play a vital role in dictating the fate of tumorigenesis, several other important factors can modulate tumor growth. Several growth factors, chemokines, and tumor-cell-secreted extracellular factors can significantly influence immune-suppressive cytokines. In addition, this review mainly focused on immune-suppressive IL-10, TGF- β , IL-4, and IL-35 cytokines; however, several other cytokines exist, such as IL-13, IL-5, IL-1 β , IL-2, and IL-6, and they can play immune-suppressive roles, which controls antitumor immunity and promotes tumor growth. Apart from the immune-suppressive functions of IL-10, TGF- β , IL-4, and IL-35, these cytokines might act directly on tumor cells and control their growth and proliferation. Nevertheless, this review predominantly revealed their immune-suppressive functions of pro-tumorigenic effects. Therefore, further detailed studies are necessary to elucidate distinct functions of IL-10, TGF- β , IL-4, and IL-35 in tumor milieu and their role in maintaining equilibrium between protumor and antitumor activities (see Table 1).

Cytokine	Observations and mechanism of action	Reference
IL-10	 Pegilodecakin (pegylated IL-10) with anti-PD-1 monoclonal antibodies showed antitumor activity in patients with renal cell carcinoma and non- small-cell carcinoma 	Naing et al. ¹⁴⁶
	 Pegilodecakin in combination with anti-PD-1 enhances effector CD8 T cell response in renal cell carcinoma patients (phase 1 clinical study) 	Tannir et al. ⁵⁹
	 Combinatorial treatment of PD-1 blockade and IL-10 neutralization enhances antitumor immunity against ovarian cancer 	Lamichhane et al. ¹⁴⁷
	 IL-10 blockade leads to complete melanoma rejection 	Llodiz et al. ¹⁴⁸
	 Pegylated IL-10 (pegilodecakin) induces the proliferation of antigen- activated intratumoral CD8+ T cells 	Autio and Oft ¹⁴⁹
	 IL-10 immunomodulator MK-1966 activates cell-mediated immunity against cancer cells 	Oft, ¹⁵⁰ Heumann and Azad ¹⁵¹
	 Phase 2 clinical trial (NCT03382912) studying AM0010 (peg-IL-10) in combination with Nivolumab enhances tumor clearance in advanced NSCLC patients 	Choucair et al. ¹⁵²
	 Phase I clinical trial AM0010: PEGylated human IL-10 shows effectiveness against pancreatic cancer, colorectal cancer, and advanced solid tumors; in addition, anti-PD-1 enhances intratumoral CD8 T cell response 	Naing et al., ¹⁵³ Papadopoulos et al., ¹⁵⁴ Infante et al. ¹⁵⁵
	 Cetuximab-based IL-10 fusion protein (CmAb-(IL10)2) improves antitumor effects in advanced tumors 	Qiao et al. ⁴⁵
	 L-10 neutralization induces antigen-specific IFN-gamma response 	Liu et al. ¹⁵⁶
	 Administration of CpG and blockade of IL-10 signaling showed robust antitumor therapeutic activity 	Vicari et al. ¹⁵⁷
TGF-β	 Blockade of TGF-β with radiation therapy induces an abscopal effect and controls tumor growth; a phase I/II clinical trial in metastatic breast cancer patients (NCT01401062) 	Diamond et al. ¹⁵⁸
	 TGF-β blockade increases responsiveness to anti-PD-1 and radiation therapy in metastatic breast cancer patients 	Formenti et al. ¹⁵⁹
	- Galunisertib (LY2157299 monohydrate) inhibits TGF- β signaling and controls tumor progression	Herbertz et al. ¹⁶⁰
	 PF-03446962, a human monoclonal antibody, blocks TGF-β receptors and shows effectiveness in patients with urothelial cancer (phase 2 clinical trials NCT01911273 and NCT01486368) 	Necchi et al. ¹⁶¹
	 Small molecule TGF-β inhibitors such as Trabedersen, Galunisertib, Gradalis, PF-03446962, and NIS793 show effectiveness against colorectal cancer 	Gachpazan et al. ¹⁶²
	 Anti-TGF-β receptor monoclonal antibody LY3022859 controls tumor growth in patients with advanced solid tumors (phase 1 clinical trial NCT01646203) 	Tolcher et al. ¹⁶³
	 Phase I study of PF-03446962 targeting TGF-β receptors has clinical activity in patients with HCC (NCT00557856) 	Simonelli et al. ¹⁶⁴
	 A bifunctional fusion protein M7824 targets TGF-β and PD-L1 and has had promising efficacy in NSCLC patients (phase 1 clinical trial NCT02517398) 	Paz-Ares et al. ¹⁶⁵
	 M7824 in patients with heavily pretreated colorectal cancer (phase I clinical trial NCT02517398) 	Kopetz et al. ¹⁶⁶
	 The blockade of TGF-β has illustrated promising antitumor properties in several preclinical models 	Ciardiello et al. ¹⁶⁷
	 M7824, phase 1 evaluation in cervical and other human papillomavirus– associated cancers (NCT03427411) 	Allan et al. ¹⁶⁸
	 NCT02699515, a phase 1 clinical trial targeting TGF-β in patients with advanced solid tumors, including HCC, showed the best overall response 	Doi et al. ¹⁶⁹
	 A randomized phase 2b clinical trial (NCT00431561) targets TGF-β2 in adult patients with grade III/IV high-grade glioma 	Hwang et al. ¹⁷⁰

Table 1. Clinical implications of IL-10, TGF- β , IL-4, and IL-35 in cancer immunotherapy.

(Continued)

Table I. (Continued)

Cytokine	Observations and mechanism of action	Reference
IL-4	 Phase II trial of recombinant human IL-4 in patients with advanced renal cell carcinoma had promising growth inhibitory and immunologic effects 	Whitehead et al. ¹⁷¹
	• A phase I trial of continuous infusion IL-4 in cancer patients enhances responses of IL-2 sensitive malignancies	Sosman et al. ¹⁷²
	• IL-4 may favor the clonogenic growth of prostate cancer stem cells	Nappo et al. ⁹⁷
	 Targeting IL-4 enhances the clinical efficacy of CAR T cells in solid tumors by promoting tumor-targeted cytotoxicity in CAR T cells 	Wang et al. ¹⁷³
	 Neutralizing the effects of IL-4 reduces Survival of Colon Cancer Stem Cells 	Francipane et al. ¹⁷⁴
	 Phase 2b clinical trial MDNA55-05 focused on IL-4R for superior treatment outcomes in glioblastoma patients 	Lézard ¹⁷⁵
	 Blockade of IL-4 reduces cancer-promoting functions of TAMs and controls pancreatic islet and mammary cancer development 	Gocheva et al. ¹⁰⁸
	 Targeting IL-4R by MDNA55 may improve glioblastoma patient outcomes and help to guide patient selection strategies 	Randazzo et al. ¹⁷⁶
	 IL-4R-targeted liposomal doxorubicin boosts cellular uptake and antitumor efficacy in colorectal cancer 	Yang et al. ¹⁷⁷
	 Targeting IL-4R revealed possible tumor-directed cancer therapy in medulloblastoma 	Joshi et al. ¹⁷⁸
IL-35	 Blockade of IL-35 limits T cell exhaustion and controls melanoma and colorectal tumor growth 	Turnis et al. ¹⁷⁹
	 IL-35 blockade enhances antitumor T cell functions in preclinical models of pancreatic cancer 	Mirlekar et al. ^{122,123}
	 Depletion of IL-35 reduces the severity of pancreatic cancer by enhancing CD8 T cell response 	Takahashi et al. ¹⁸⁰
	 Targeting IL-35 boosts the antitumor activity of T cells in NSCLC 	Wang et al. ¹⁸¹
	 IL-35 signaling is shown to promote angiogenesis and growth of xenograft pancreatic tumors 	Huang et al. ¹⁸²

IL: interleukin; TGF- β : transforming growth factor- β ; PD-1: programmed cell death protein-1; NSCLC: non-small cell lung cancer; IFN: interferon; HCC: hepatocellular carcinoma; TAMs: tumor-associated macrophages; CAR: chimeric antigen receptor.

Author's note

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References

- 1. Lee S and Margolin K. Cytokines in cancer immunotherapy. *Cancers* 2011; 3(4): 3856–3893.
- Berraondo P, Sanmamed MF, Ochoa MC, et al. Cytokines in clinical cancer immunotherapy. *Br J Cancer* 2019; 120(1): 6–15.

- 3. Valencia JC, Egbukichi N and Erwin-Cohen RA. Autoimmunity and cancer, the paradox comorbidities challenging therapy in the context of preexisting autoimmunity. *J Interferon Cytokine Res* 2019; 39(1): 72–84.
- 4. Mortier E, Ma A, Malynn BA, et al. Modulating cytokines as treatment for autoimmune diseases and cancer. *Front Immunol* 2020; 11: 8636.
- 5. Baylin SB and Jones PA. Epigenetic determinants of cancer. *Cold Spr Harbor Persp Biol* 2016; 8(9): a019505.
- 6. Miranda-Gonçalves V, Lameirinhas A, Henrique R, et al. Metabolism and epigenetic interplay in cancer: regulation and putative therapeutic targets. *Front Genet* 2018; 9: 427.
- 7. Disis ML. Immune regulation of cancer. *J Clin Oncol* 2010; 28(29): 4531.
- 8. Zamarron BF and Chen W. Dual roles of immune cells and their factors in cancer development and progression. *Int J Biol Sci* 2011; 7(5): 651–658.
- 9. Gonzalez H, Hagerling C and Werb Z. Roles of the immune system in cancer: from tumor initiation to metastatic progression. *Genes Dev* 2018; 32(19–20): 1267–1284.
- Zhang Y, Guan X-Y and Jiang P. Cytokine and chemokine signals of T-cell exclusion in tumors. *Front Immunol* 2020; 11: 594609.
- Burkholder B, Huang R-Y, Burgess R, et al. Tumor-induced perturbations of cytokines and immune cell networks. *Biochim Biophys Acta* 2014; 1845(2): 182–201.

- Lindau D, Gielen P, Kroesen M, et al. The immunosuppressive tumour network: myeloid-derived suppressor cells, regulatory T cells and natural killer T cells. *Immunology* 2013; 138(2): 105–115.
- 13. Liu Y and Cao X. Immunosuppressive cells in tumor immune escape and metastasis. *J Mol Med* 2016; 94(5): 509–522.
- Ostrand-Rosenberg S and Fenselau C. Myeloid-derived suppressor cells: immune-suppressive cells that impair antitumor immunity and are sculpted by their environment. *J Immunol* 2018; 200(2): 422–431.
- Singh S, Mehta N, Lilan J, et al. Initiative action of tumorassociated macrophage during tumor metastasis. *Biochim Open* 2017; 4: 8–18.
- Pati S, Chowdhury A, Mukherjee S, et al. Regulatory lymphocytes: the dice that resolve the tumor endgame. *Appl Cancer Res* 2020; 40(1): 1–9.
- Horii M and Matsushita T. Regulatory B cells and T cell regulation in cancer. J Mol Biol 2020; 433: 166685.
- Smith HA and Kang Y. The metastasis-promoting roles of tumorassociated immune cells. J Mol Med 2013; 91(4): 411–429.
- Taylor A, Verhagen J, Blaser K, et al. Mechanisms of immune suppression by interleukin-10 and transforming growth factorβ: the role of T regulatory cells. *Immunology* 2006; 117(4): 433–442.
- Jarnicki AG, Lysaght J, Todryk S, et al. Suppression of antitumor immunity by IL-10 and TGF-β-producing T cells infiltrating the growing tumor: influence of tumor environment on the induction of CD4+ and CD8+ regulatory T cells. *J Immunol* 2006; 177(2): 896–904.
- Cote-Sierra J, Foucras G, Guo L, et al. Interleukin 2 plays a central role in Th2 differentiation. *Proc Natl Acad Sci USA* 2004; 101(11): 3880–3885.
- Collison LW, Chaturvedi V, Henderson AL, et al. IL-35mediated induction of a potent regulatory T cell population. *Nat Immunol* 2010; 11(12): 1093–1101.
- Collison LW, Delgoffe GM, Guy CS, et al. The composition and signaling of the IL-35 receptor are unconventional. *Nat Immunol* 2012; 13(3): 290–299.
- Kim R, Emi M, Tanabe K, et al. Tumor-driven evolution of immunosuppressive networks during malignant progression. *Cancer Res* 2006; 66(11): 5527–5536.
- Labani-Motlagh A, Ashja-Mahdavi M and Loskog A. The tumor microenvironment: a milieu hindering and obstructing antitumor immune responses. *Front Immunol* 2020; 11: 940.
- Li L, Yu R, Cai T, et al. Effects of immune cells and cytokines on inflammation and immunosuppression in the tumor microenvironment. *Int Immunopharmacol* 2020; 88: 106939.
- Junttila IS. Tuning the cytokine responses: an update on interleukin (IL)-4 and IL-13 receptor complexes. *Front Immunol* 2018; 9: 888.
- Batlle E and Massagué J. Transforming growth factor-β signaling in immunity and cancer. *Immunity* 2019; 50(4): 924–940.
- O'shea J. Targeting the Jak/STAT pathway for immunosuppression. Ann Rheumat Dis 2004; 63(Suppl. 2): ii67–ii71.
- Yang L, Li A, Lei Q, et al. Tumor-intrinsic signaling pathways: key roles in the regulation of the immunosuppressive tumor microenvironment. *J Hematol Oncol* 2019; 12(1): 1–14.
- Tedder TF and Leonard WJ. Regulatory B cells: IL-35 and IL-21 regulate the regulators. *Nat Rev Rheumatol* 2014; 10(8): 452–453.

- Uzhachenko RV and Shanker A. CD8+ T lymphocyte and NK cell network: circuitry in the cytotoxic domain of immunity. *Front Immunol* 2019; 10: 1906.
- 33. Krijgsman D, Hokland M and Kuppen PJ. The role of natural killer T cells in cancer: a phenotypical and functional approach. *Front Immunol* 2018; 9: 367.
- Setrerrahmane S and Xu H. Tumor-related interleukins: old validated targets for new anti-cancer drug development. *Mol Cancer* 2017; 16(1): 1–17.
- Odenthal J, Takes R and Friedl P. Plasticity of tumor cell invasion: governance by growth factors and cytokines. *Carcinogenesis* 2016; 37(12): 1117–1128.
- Mannino MH, Zhu Z, Xiao H, et al. The paradoxical role of IL-10 in immunity and cancer. *Cancer Lett* 2015; 367(2): 103–107.
- 37. Massagué J. TGFβ in cancer. Cell 2008; 134(2): 215–230.
- Li Z, Chen L and Qin Z. Paradoxical roles of IL-4 in tumor immunity. *Cell Mol Immunol* 2009; 6(6): 415–422.
- 39. Suzuki A, Leland P, Joshi BH, et al. Targeting of IL-4 and IL-13 receptors for cancer therapy. *Cytokine* 2015; 75(1): 79–88.
- Yazdani Z, Rafiei A, Golpour M, et al. IL-35, a double-edged sword in cancer. J Cell Biochem 2020; 121(3): 2064–2076.
- Li Z, Jiang J, Wang Z, et al. Endogenous interleukin-4 promotes tumor development by increasing tumor cell resistance to apoptosis. *Cancer Res* 2008; 68(21): 8687–8694.
- Ouyang W and O'Garra A. IL-10 family cytokines IL-10 and IL-22: from basic science to clinical translation. *Immunity* 2019; 50(4): 871–891.
- Hassuneh MR, Nagarkatti M and Nagarkatti PS. Role of interleukin-10 in the regulation of tumorigenicity of a T cell lymphoma. *Leuk Lymphoma* 2013; 54(4): 827–834.
- Wang X, Wong K, Ouyang W, et al. Targeting IL-10 family cytokines for the treatment of human diseases. *Cold Spr Harbor Persp Biol* 2019; 11(2): a028548.
- Qiao J, Liu Z, Dong C, et al. Targeting tumors with IL-10 prevents dendritic cell-mediated CD8+ T cell apoptosis. *Cancer Cell* 2019; 35(6): 901–915.e4.
- 46. Naing A, Infante JR, Papadopoulos KP, et al. PEGylated IL-10 (Pegilodecakin) induces systemic immune activation, CD8+ T cell invigoration and polyclonal T cell expansion in cancer patients. *Cancer Cell* 2018; 34(5): 775–791.e3.
- Mumm JB, Emmerich J, Zhang X, et al. IL-10 elicits IFNγdependent tumor immune surveillance. *Cancer Cell* 2011; 20(6): 781–796.
- Ruffell B, Chang-Strachan D, Chan V, et al. Macrophage IL-10 blocks CD8+ T cell-dependent responses to chemotherapy by suppressing IL-12 expression in intratumoral dendritic cells. *Cancer Cell* 2014; 26(5): 623–637.
- 49. Li C and Zuo W. IL-10 in vitro could enhance IFNγ expression and suppress PD-1 expression in CD8 T cells from esophageal cancer patients. *Exp Cell Res* 2019; 379(2): 159–165.
- Wang X, Li J, Lu C, et al. IL-10-producing B cells in differentiated thyroid cancer suppress the effector function of T cells but improve their survival upon activation. *Exp Cell Res* 2019; 376(2): 192–197.
- Hu H-T, Ai X, Lu M, et al. Characterization of intratumoral and circulating IL-10-producing B cells in gastric cancer. *Exp Cell Res* 2019; 384(2): 111652.

- 52. Zhang H, Li R, Cao Y, et al. Poor clinical outcomes and immunoevasive contexture in intratumoral IL-10producing macrophages enriched gastric cancer patients. *Ann Surg.* Epub ahead of print 11 June 2021. DOI: 10.1097/ SLA.0000000000004037.
- 53. Smith LK, Boukhaled GM, Condotta SA, et al. Interleukin-10 directly inhibits CD8+ T cell function by enhancing N-glycan branching to decrease antigen sensitivity. *Immunity* 2018; 48(2): 299–312.e5.
- 54. Li L, Ma Y and Xu Y. Follicular regulatory T cells infiltrated the ovarian carcinoma and resulted in CD8 T cell dysfunction dependent on IL-10 pathway. *Int Immunopharmacol* 2019; 68: 81–87.
- 55. Plaumann J, Engelhardt M, Awwad MH, et al. IL-10 inducible CD8+ regulatory T-cells are enriched in patients with multiple myeloma and impact the generation of antigen-specific T-cells. *Cancer Immunol Immunother* 2018; 67(11): 1695–1707.
- 56. Liu Z, Zhou Q, Wang Z, et al. Intratumoral TIGIT+ CD8+ T-cell infiltration determines poor prognosis and immune evasion in patients with muscle-invasive bladder cancer. J Immunother Cancer 2020; 8(2): e000978.
- Matsuzaki J, Gnjatic S, Mhawech-Fauceglia P, et al. Tumorinfiltrating NY-ESO-1–specific CD8+ T cells are negatively regulated by LAG-3 and PD-1 in human ovarian cancer. *Proc Natl Acad Sci* 2010; 107(17): 7875–7880.
- Tannir NM, Naing A, Infante JR, et al. Pegilodecakin with nivolumab (nivo) or pembrolizumab (pembro) in patients (pts) with metastatic renal cell carcinoma (RCC). *Am Soc Clin Oncol* 2018; 36: 4509.
- 59. Tannir NM, Papadopoulos KP, Wong DJ, et al. Pegilodecakin as monotherapy or in combination with anti-PD-1 or tyrosine kinase inhibitor in heavily pretreated patients with advanced renal cell carcinoma: final results of cohorts A, G, H and I of IVY Phase I study. *Int J Cancer* 2021; 149(2): 403–408.
- Marcon F, Zuo J, Pearce H, et al. NK cells in pancreatic cancer demonstrate impaired cytotoxicity and a regulatory IL-10 phenotype. *Oncoimmunology* 2020; 9(1): 1845424.
- Sun C, Xu J, Huang Q, et al. High NKG2A expression contributes to NK cell exhaustion and predicts a poor prognosis of patients with liver cancer. *Oncoimmunology* 2017; 6(1): e1264562.
- Sun H, Huang Q, Huang M, et al. Human CD96 correlates to natural killer cell exhaustion and predicts the prognosis of human hepatocellular carcinoma. *Hepatology* 2019; 70(1): 168–183.
- Ostapchuk YO, Cetin EA, Perfilyeva YV, et al. Peripheral blood NK cells expressing HLA-G, IL-10 and TGF-β in healthy donors and breast cancer patients. *Cell Immunol* 2015; 298(1–2): 37–46.
- 64. van den Bulk J, de Miranda NF and Ten Dijke P. Therapeutic targeting of TGF-β in cancer: hacking a master switch of immune suppression. *Clin Sci* 2021; 135(1): 35–52.
- 65. Xu L, Kitani A and Strober W. Molecular mechanisms regulating TGF-β-induced Foxp3 expression. *Mucosal Immunol* 2010; 3(3): 230–238.
- 66. Moo-Young TA, Larson JW, Belt BA, et al. Tumor derived TGF-Beta mediates conversion of CD4+ Foxp3+ regulatory T cells in a murine model of pancreas cancer. *J Immunother* 2009; 32(1): 12–21.
- Sheng W, Liu Y, Chakraborty D, et al. Simultaneous Inhibition of LSD1 and TGF-B enables eradication of poorly immunogenic

tumors with anti-PD-1 treatment. *Cancer Disc* 2021; 11: 1970–1981.

- Marwitz S, Ballesteros-Merino C, Jensen SM, et al. Phosphorylation of SMAD3 in immune cells predicts survival of patients with early stage non-small cell lung cancer. *J Immunother Cancer* 2021; 9(2): e001469.
- Liu M, Kuo F, Capistrano KJ, et al. TGF-β suppresses type 2 immunity to cancer. *Nature* 2020; 587(7832): 115–120.
- 70. de Streel G, Bertrand C, Chalon N, et al. Selective inhibition of TGF-β produced by GARP-expressing Tregs overcomes resistance to PD-1/PD-L1 blockade in cancer. *Nat Commun* 2020; 11(1): 1–15.
- Chen X, Wang L, Li P, et al. Dual TGF-β and PD-1 blockade synergistically enhances MAGE-A3-specific CD8+ T cell response in esophageal squamous cell carcinoma. *Int J Cancer* 2018; 143(10): 2561–2574.
- Jiao S, Subudhi SK, Aparicio A, et al. Differences in tumor microenvironment dictate T helper lineage polarization and response to immune checkpoint therapy. *Cell* 2019; 179(5): 1177–1190.e13.
- 73. Sow HS, Ren J, Camps M, et al. Combined inhibition of TGFβ signaling and the PD-L1 immune checkpoint is differentially effective in tumor models. *Cells* 2019; 8(4): 320.
- 74. Liu Q, Wu H, Li Y, et al. Combined blockade of TGf-β and GM-CSF improves chemotherapeutic effects for pancreatic cancer by modulating tumor microenvironment. *Cancer Immunol Immunother* 2020; 69(8): 1477–1492.
- Fridlender ZG, Sun J, Kim S, et al. Polarization of tumorassociated neutrophil phenotype by TGF-β: "N1" versus "N2" TAN. *Cancer Cell* 2009; 16(3): 183–194.
- Li Q, Wang Y, Jia W, et al. Low-dose anti-angiogenic therapy sensitizes breast cancer to PD-1 blockade. *Clin Cancer Res* 2020; 26(7): 1712–1724.
- 77. Shen Y, Teng Y, Lv Y, et al. PD-1 does not mark tumor-infiltrating CD8+ T cell dysfunction in human gastric cancer. J Immunother Cancer 2020; 8(2): e000422.
- 78. Yang Z, Qi Y, Lai N, et al. Notch1 signaling in melanoma cells promoted tumor-induced immunosuppression via upregulation of TGF-β. *J Exp Clin Cancer Res* 2018; 37(1): 1–13.
- 79. Yu Yang JL, Jiao SC, Wang JD, et al. TGF-β precursor and CD8 are potential prognostic and predictive markers in operated breast cancer. *J Huazhong Univ Sci Technolog Med Sci* 2014; 34(1): 51–58.
- Nakazawa N, Yokobori T, Kaira K, et al. High stromal TGFBI in lung cancer and intratumoral CD8-positive T cells were associated with poor prognosis and therapeutic resistance to immune checkpoint inhibitors. *Ann Surg Oncol* 2020; 27(3): 933–942.
- Wu M, Chen X, Lou J, et al. TGF-β contributes to CD8+ Treg induction through p38 MAPK signaling in ovarian cancer microenvironment. *Oncotarget* 2016; 7(28): 44534.
- Zloza A, Jagoda MC, Lyons GE, et al. CD8 co-receptor promotes susceptibility of CD8+ T cells to transforming growth factor-β (TGF-β)-mediated suppression. *Cancer Immunol Immunother* 2011; 60(2): 291–297.
- Li R, Zhang H, Cao Y, et al. Lauren classification identifies distinct prognostic value and functional status of intratumoral CD8+ T cells in gastric cancer. *Cancer Immunol Immunother* 2020; 69: 1327–1336.

- Yu J, Wei M, Becknell B, et al. Pro- and antiinflammatory cytokine signaling: reciprocal antagonism regulates interferongamma production by human natural killer cells. *Immunity* 2006; 24(5): 575–590.
- 85. Hao Y, Baker D and Ten Dijke P. TGF-β-mediated epithelialmesenchymal transition and cancer metastasis. *Int J Mol Sci* 2019; 20(11): 2767.
- 86. Xu J, Lamouille S and Derynck R. TGF-β-induced epithelial to mesenchymal transition. *Cell Res* 2009; 19(2): 156–172.
- Wendt MK, Allington TM and Schiemann WP. Mechanisms of the epithelial–mesenchymal transition by TGF-β. *Future Oncol* 2009; 5(8): 1145–1168.
- Derynck R, Muthusamy BP and Saeteurn KY. Signaling pathway cooperation in TGF-β-induced epithelial–mesenchymal transition. *Curr Opin Cell Biol* 2014; 31: 56–66.
- Fujii R, Jochems C, Tritsch SR, et al. An IL-15 superagonist/ IL-15Rα fusion complex protects and rescues NK cell-cytotoxic function from TGF-β1-mediated immunosuppression. *Cancer Immunol Immunother* 2018; 67(4): 675–689.
- 90. Stüber T, Monjezi R, Wallstabe L, et al. Inhibition of TGF-βreceptor signaling augments the antitumor function of ROR1specific CAR T-cells against triple-negative breast cancer. J Immunother Cancer 2020; 8(1): e000676.
- Zhou L, Chong MM and Littman DR. Plasticity of CD4+ T cell lineage differentiation. *Immunity* 2009; 30(5): 646–655.
- Rengarajan J, Szabo SJ and Glimcher LH. Transcriptional regulation of Th1/Th2 polarization. *Immunol Today* 2000; 21(10): 479–483.
- Fu C, Jiang L, Hao S, et al. Activation of the IL-4/STAT6 signaling pathway promotes lung cancer progression by increasing M2 myeloid cells. *Front Immunol* 2019; 10: 2638.
- 94. Ito Shirota H, Kasahara Y, Saijo K, et al. IL-4 blockade alters the tumor microenvironment and augments the response to cancer immunotherapy in a mouse model. *Cancer Immunol Immunother* 2017; 66(11): 1485–1496.
- Maier B, Leader AM, Chen ST, et al. A conserved dendriticcell regulatory program limits antitumour immunity. *Nature* 2020; 580(7802): 257–262.
- Joshi BH, Suzuki A, Fujisawa T, et al. Identification, characterization, and targeting of IL-4 receptor by IL-4-Pseudomonas exotoxin in mouse models of anaplastic thyroid cancer. *Discov Med* 2015; 20(111): 273–284.
- Nappo G, Handle F, Santer F, et al. The immunosuppressive cytokine interleukin-4 increases the clonogenic potential of prostate stem-like cells by activation of STAT6 signalling. *Oncogenesis* 2017; 6(5): e342.
- Roca H, Craig MJ, Ying C, et al. IL-4 induces proliferation in prostate cancer PC3 cells under nutrient-depletion stress through the activation of the JNK-pathway and survivin upregulation. J Cell Biochem 2012; 113(5): 1569–1580.
- 99. Zhao S, Shen W, Yu J, et al. TBX21 predicts prognosis of patients and drives cancer stem cell maintenance via the TBX21–IL-4 pathway in lung adenocarcinoma. *Stem Cell Res Ther* 2018; 9(1): 1–15.
- 100. Erb HH, Guggenberger F, Santer FR, et al. Interleukin-4 induces a CD44high/CD49bhigh PC3 subpopulation with tumor-initiating characteristics. *J Cell Biochem* 2018; 119(5): 4103–4112.
- 101. Jia Y, Xie X, Shi X, et al. Associations of common IL-4 gene polymorphisms with cancer risk: a meta-analysis. *Mol Med Rep* 2017; 16(2): 1927–1945.

- 102. Shamoun L, Skarstedt M, Andersson RE, et al. Association study on IL-4, IL-4R α and IL-13 genetic polymorphisms in Swedish patients with colorectal cancer. *Clin Chim Acta* 2018; 487: 101–106.
- 103. Tan N, Song J, Yan M, et al. Association between IL-4 tagging single nucleotide polymorphisms and the risk of lung cancer in China. *Mol Genet Genomic Med* 2019; 7(4): e00585.
- 104. Ibrahimi M, Jamalzei B, Akbari M, et al. Association between interleukin 4 (IL-4) VNTR, gene polymorphism, and breast cancer susceptibility in Iranian population: experimental and web base analysis. *Bratisl Lek Listy* 2018; 119(10): 651–654.
- 105. Formentini A, Braun P, Fricke H, et al. Expression of interleukin-4 and interleukin-13 and their receptors in colorectal cancer. *Int J Colorectal Dis* 2012; 27(10): 1369–1376.
- 106. Kim ES, Choi YE, Hwang SJ, et al. IL-4, a direct target of miR-340/429, is involved in radiation-induced aggressive tumor behavior in human carcinoma cells. *Oncotarget* 2016; 7(52): 86836.
- 107. Stanciu AE, Serdarevic N, Hurduc AE, et al. IL-4, IL-10 and high sensitivity-CRP as potential serum biomarkers of persistent/recurrent disease in papillary thyroid carcinoma with/ without Hashimoto's thyroiditis. *Scand J Clin Lab Invest* 2015; 75(7): 539–548.
- 108. Gocheva V, Wang H-W, Gadea BB, et al. IL-4 induces cathepsin protease activity in tumor-associated macrophages to promote cancer growth and invasion. *Genes Dev* 2010; 24(3): 241–255.
- 109. Apte SH, Groves P, Olver S, et al. IFN-γ inhibits IL-4–induced type 2 cytokine expression by CD8 T cells in vivo and modulates the anti-tumor response. *J Immunol* 2010; 185(2): 998–1004.
- 110. Noble A, Mehta H, Lovell A, et al. IL-12 and IL-4 activate a CD39-dependent intrinsic peripheral tolerance mechanism in CD8+ T cells. *Eur J Immunol* 2016; 46(6): 1438–1448.
- 111. Crawley AM, Vranjkovic A, Young C, et al. Interleukin-4 downregulates CD127 expression and activity on human thymocytes and mature CD8+ T cells. *Eur J Immunol* 2010; 40(5): 1396–1407.
- 112. Olver S, Groves P, Buttigieg K, et al. Tumor-derived interleukin-4 reduces tumor clearance and deviates the cytokine and granzyme profile of tumor-induced CD8+ T cells. *Cancer Res* 2006; 66(1): 571–580.
- 113. Ohayon DE, Krishnamurthy D, Brusilovsky M, et al. IL-4 and IL-13 modulate natural killer cell responses under inflammatory conditions. *Am Assoc Immnol* 2017; 198: 194.11.
- 114. Agaugué S, Marcenaro E, Ferranti B, et al. Human natural killer cells exposed to IL-2, IL-12, IL-18, or IL-4 differently modulate priming of naive T cells by monocyte-derived dendritic cells. *Blood: J Am Soc Hematol* 2008; 112(5): 1776–1783.
- 115. O'Brien TF, Bao K, Dell'Aringa M, et al. Cytokine expression by invariant natural killer T cells is tightly regulated throughout development and settings of type-2 inflammation. *Mucosal Immunol* 2016; 9(3): 597–609.
- Vignali DA and Kuchroo VK. IL-12 family cytokines: immunological playmakers. *Nat Immunol* 2012; 13(8): 722–728.
- Wojno EDT, Hunter CA and Stumhofer JS. The immunobiology of the interleukin-12 family: room for discovery. *Immunity* 2019; 50(4): 851–870.
- 118. Su L-C, Liu X-Y, Huang A-F, et al. Emerging role of IL-35 in inflammatory autoimmune diseases. *Autoimmun Rev* 2018; 17(7): 665–673.

- Choi J, Leung PS, Bowlus C, et al. IL-35 and autoimmunity: a comprehensive perspective. *Clin Rev Allergy Immunol* 2015; 49(3): 327–332.
- 120. Sawant DV, Yano H, Chikina M, et al. Adaptive plasticity of IL-10+ and IL-35+ T reg cells cooperatively promotes tumor T cell exhaustion. *Nat Immunol* 2019; 20(6): 724–735.
- 121. Sullivan JA, Tomita Y, Jankowska-Gan E, et al. Treg-cellderived IL-35-coated extracellular vesicles promote infectious tolerance. *Cell Rep* 2020; 30(4): 1039–1051.e5.
- 122. Mirlekar B, Michaud D, Searcy R, et al. IL35 hinders endogenous antitumor T-cell immunity and responsiveness to immunotherapy in pancreatic cancer. *Cancer Immunol Res* 2018; 6(9): 1014–1024.
- 123. Mirlekar B, Michaud D, Lee SJ, et al. B cell-derived IL35 drives STAT3-dependent CD8+ T-cell exclusion in pancreatic cancer. *Cancer Immunol Res* 2020; 8(3): 292–308.
- 124. Zhu J, Wang Y, Li D, et al. Interleukin-35 promotes progression of prostate cancer and inhibits anti-tumour immunity. *Cancer Cell Int* 2020; 20: 487–413.
- 125. Zhu J, Yang X, Wang Y, et al. Interleukin-35 is associated with the tumorigenesis and progression of prostate cancer. *Oncol Lett* 2019; 17(6): 5094–5102.
- 126. Zhang T, Nie J, Liu X, et al. Correlation analysis among the level of IL-35, microvessel density, lymphatic vessel density, and prognosis in non-small cell lung cancer. *Clin Transl Sci* 2021; 14(1): 389–394.
- 127. Solati H, Zareinejad M, Ghavami A, et al. IL-35 and IL-18 serum levels in children with acute lymphoblastic leukemia: the relationship with prognostic factors. *J Pediatr Hematol Oncol* 2020; 42(4): 281–286.
- 128. Fan Y-G, Zhai J-M, Wang W, et al. IL-35 over-expression is associated with genesis of gastric cancer. *Asian Pac J Cancer Prev* 2015; 16(7): 2845–2849.
- 129. Gu JH, Wang XG, Wang LQ, et al. Serum level of interleukin-35 as a potential prognostic factor for gastric cancer. *Asia Pac J Clin Oncol* 2021; 17(1): 52–59.
- 130. Chen G, Liang Y, Guan X, et al. Circulating low IL-23: IL-35 cytokine ratio promotes progression associated with poor prognosis in breast cancer. *Am J Transl Res* 2016; 8(5): 2255– 2264.
- 131. Zeng J-C, Zhang Z, Li T-Y, et al. Assessing the role of IL-35 in colorectal cancer progression and prognosis. *Int J Clin Exp Pathol* 2013; 6(9): 1806–1816.
- 132. Larousserie F, Kebe D, Huynh T, et al. Evidence for IL-35 expression in diffuse large B-cell lymphoma and impact on the patient's prognosis. *Front Oncol* 2019; 9: 563.
- 133. Kalbasi A and Ribas A. Tumour-intrinsic resistance to immune checkpoint blockade. *Nat Rev Immunol* 2020; 20(1): 25–39.
- 134. Huang Q, Lei Y, Li X, et al. A highlight of the mechanisms of immune checkpoint blocker resistance. *Front Cell Dev Biol* 2020; 8: 580140.
- 135. Barrueto L, Caminero F, Cash L, et al. Resistance to checkpoint inhibition in cancer immunotherapy. *Transl Oncol* 2020; 13(3): 100738.
- 136. Fares CM, Van Allen EM, Drake CG, et al. Mechanisms of resistance to immune checkpoint blockade: why does checkpoint inhibitor immunotherapy not work for all patients? *Am Soc Clin Oncol Educ Book* 2019; 39: 147–164.
- 137. Fujiwara Y, Mittra A, Naqash AR, et al. A review of mechanisms of resistance to immune checkpoint inhibitors and

potential strategies for therapy. *Cancer Drug Resist* 2020; 3(3): 252–275.

- 138. Walle T, Martinez Monge R, Cerwenka A, et al. Radiation effects on antitumor immune responses: current perspectives and challenges. *Ther Adv Med Oncol* 2018; 10: 742575.
- 139. Jeong H, Bok S, Hong B-J, et al. Radiation-induced immune responses: mechanisms and therapeutic perspectives. *Blood Res* 2016; 51(3): 157–163.
- 140. Kaur P and Asea A. Radiation-induced effects and the immune system in cancer. *Front Oncol* 2012; 2: 191.
- 141. Bonaventura P, Shekarian T, Alcazer V, et al. Cold tumors: a therapeutic challenge for immunotherapy. *Front Immunol* 2019; 10: 168.
- 142. Duan Q, Zhang H, Zheng J, et al. Turning cold into hot: firing up the tumor microenvironment. *Trends Cancer* 2020; 6(7): 605–618.
- 143. Vareki SM. High and low mutational burden tumors versus immunologically hot and cold tumors and response to immune checkpoint inhibitors. *J Immunother Cancer* 2018; 6(1): 1–5.
- 144. Liu Y-T and Sun Z-J. Turning cold tumors into hot tumors by improving T-cell infiltration. *Theranostics* 2021; 11(11): 5365–5386.
- 145. Van der Woude LL, Gorris MAJ, Halilovic A, et al. Migrating into the tumor: a roadmap for T cells. *Trends Cancer* 2017; 3(11): 797–808.
- 146. Naing A, Wong DJ, Infante JR, et al. Pegilodecakin combined with pembrolizumab or nivolumab for patients with advanced solid tumours (IVY): a multicentre, multicohort, open-label, phase 1b trial. *Lancet Oncol* 2019; 20(11): 1544–1555.
- 147. Lamichhane P, Karyampudi L, Shreeder B, et al. IL10 release upon PD-1 blockade sustains immunosuppression in ovarian cancer. *Cancer Res* 2017; 77(23): 6667–6678.
- 148. Llopiz D, Aranda F, Díaz-Valdés N, et al. Vaccine-induced but not tumor-derived Interleukin-10 dictates the efficacy of Interleukin-10 blockade in therapeutic vaccination. *Oncoimmunology* 2016; 5(2): e1075113.
- 149. Autio K and Oft M. Pegylated interleukin-10: clinical development of an immunoregulatory cytokine for use in cancer therapeutics. *Curr Oncol Rep* 2019; 21(2): 1–7.
- 150. Oft M. Immune regulation and cytotoxic T cell activation of IL-10 agonists: preclinical and clinical experience. *Semin Immunol* 2019; 44: 101325.
- 151. Heumann T and Azad N. Next-generation immunotherapy for pancreatic ductal adenocarcinoma: navigating pathways of immune resistance. *Cancer Meta Rev* 2021: 1–26.
- 152. Choucair K, Kelso JD, Duff JR, et al. Interleukin 10-mediated response and correlated anemia in a patient with advanced non-small cell lung carcinoma. *Case Rep Oncol* 2019; 12(1): 297–303.
- 153. Naing A, Papadopoulos KP, Infante JR, et al. Clinical activity and safety of pegylated human IL-10 (AM0010) in combination with anti-PD1. *Am Soc Clin Oncol* 2016; 34: 3018.
- 154. Papadopoulos KP, Naing A, Infante JR, et al. Anti-tumor activity of PEGylated human IL-10 (AM0010) in patients with pancreatic or colorectal cancer. *Am Soc Clin Oncol* 2016; 34: 3082.
- 155. Infante JR, Naing A, Papadopoulos KP, et al. A first-in-human dose escalation study of PEGylated recombinant human IL-10 (AM0010) in advanced solid tumors. *Am Soc Clin Oncol* 2015; 33: 3017.

- 156. Liu XS, Dyer J, Leggatt GR, et al. Overcoming original antigenic sin to generate new CD8 T cell IFN-γ responses in an antigen-experienced host. *J Immunol* 2006; 177(5): 2873– 2879.
- 157. Vicari AP, Chiodoni C, Vaure C, et al. Reversal of tumorinduced dendritic cell paralysis by CpG immunostimulatory oligonucleotide and anti–interleukin 10 receptor antibody. J Exp Med 2002; 196(4): 541–549.
- 158. Diamond J, Vanpouille-Box C, Barcellos-Hoff MH, et al. The abscopal effect of local radiotherapy is induced by TGFβ blockade. *J Immunother Cancer* 2013; 1(1): 115.
- 159. Formenti SC, Hawtin RE, Dixit N, et al. Baseline T cell dysfunction by single cell network profiling in metastatic breast cancer patients. *J Immunother Cancer* 2019; 7(1): 1–8.
- 160. Herbertz S, Sawyer JS, Stauber AJ, et al. Clinical development of galunisertib (LY2157299 monohydrate), a small molecule inhibitor of transforming growth factor-beta signaling pathway. *Drug Des Devel Ther* 2015; 9: 4479–4499.
- 161. Necchi A, Giannatempo P, Mariani L, et al. PF-03446962, a fully-human monoclonal antibody against transforming growth-factor β (TGFβ) receptor ALK1, in pre-treated patients with urothelial cancer: an open label, single-group, phase 2 trial. *Invest New Drugs* 2014; 32(3): 555–560.
- 162. Gachpazan M, Kashani H, Hassanian SM, et al. Therapeutic potential of targeting transforming growth factor-beta in colorectal cancer: rational and progress. *Curr Pharm Des* 2019; 25(38): 4085–4089.
- 163. Tolcher AW, Berlin JD, Cosaert J, et al. A phase 1 study of anti-TGFβ receptor type-II monoclonal antibody LY3022859 in patients with advanced solid tumors. *Cancer Chemother Pharmacol* 2017; 79(4): 673–680.
- 164. Simonelli M, Zucali P, Santoro A, et al. Phase I study of PF-03446962, a fully human monoclonal antibody against activin receptor-like kinase-1, in patients with hepatocellular carcinoma. *Ann Oncol* 2016; 27(9): 1782–1787.
- 165. Paz-Ares LG, Kim TM, Vicente Baz D, et al. Results from a second-line (2L) NSCLC cohort treated with M7824 (MSB0011359C), a bifunctional fusion protein targeting TGF-β and PD-L1. *Am Soc Clin Oncol* 2018; 36: 9017.
- 166. Kopetz S, Spira AI, Wertheim M, et al. M7824 (MSB0011359C), a bifunctional fusion protein targeting PD-L1 and TGF-β, in patients with heavily pretreated CRC: preliminary results from a phase I trial. *Am Soc Clin Oncol* 2018; 36: 764.
- 167. Ciardiello D, Elez E, Tabernero J, et al. Clinical development of therapies targeting TGFβ: current knowledge and future perspectives. *Ann Oncol* 2020; 31(10): 1336–1349.
- 168. Allan S, Braiteh F, Aller EC, et al. P37 phase 1 evaluation of bintrafusp alfa (M7824), a bifunctional fusion protein targeting TGF-β and PD-L1, in cervical cancer. *BMJ Spec J* 2019; 29: A72–A73.
- 169. Doi T, Fujiwara Y, Koyama T, et al. Phase I study of the bifunctional fusion protein bintrafusp alfa in Asian patients

with advanced solid tumors, including a hepatocellular carcinoma safety-assessment cohort. *Oncologist* 2020; 25(9): e1292.

- 170. Hwang L, Nam D, Uckun F, et al. OT-101/Chemotherapy—a novel mechanism of action (MOA) in glioblastoma immunization therapy. *AACR* 2019; 79: 5029.
- 171. Whitehead RP, Lew D, Flanigan RC, et al. Phase II trial of recombinant human interleukin-4 in patients with advanced renal cell carcinoma: a southwest oncology group study. J Immunother 2002; 25(4): 352–358.
- 172. Sosman J, Fisher SG, Kefer C, et al. A phase I trial of continuous infusion interleukin-4 (IL-4) alone and following interleukin-2 (IL-2) in cancer patients. *Ann Oncol* 1994; 5(5): 447–452.
- 173. Wang Y, Jiang H, Luo H, et al. An IL-4/21 inverted cytokine receptor improving CAR-T cell potency in immunosuppressive solid-tumor microenvironment. *Front Immunol* 2019; 10: 1691.
- 174. Francipane MG, Alea MP, Lombardo Y, et al. Crucial role of interleukin-4 in the survival of colon cancer stem cells. *Cancer Res* 2008; 68(11): 4022–4025.
- 175. Lézard L. Medicenna presents clinical validation of the IL4 receptor as a biomarker for MDNA55 in recurrent glioblastoma, https://www.newswire.ca/news-releases/medicenna-presents-clinical-validation-of-the-il4-receptor-as-a-biomarkerfor-mdna55-in-recurrent-glioblastoma-857957449.html
- 176. Randazzo D, Achrol A, Aghi MK, et al. MDNA55: a locally administered IL4 guided toxin as a targeted treatment for recurrent glioblastoma. *Am Soc Clin Oncol* 2019; 37: 2039.
- 177. Yang C-Y, Liu H-W, Tsai Y-C, et al. Interleukin-4 receptortargeted liposomal doxorubicin as a model for enhancing cellular uptake and antitumor efficacy in murine colorectal cancer. *Cancer Biol Ther* 2015; 16(11): 1641–1650.
- 178. Joshi B, Leland P, Silber J, et al. IL-4 receptors on human medulloblastoma tumours serve as a sensitive target for a circular permuted IL-4-Pseudomonas exotoxin fusion protein. *Br J Cancer* 2002; 86(2): 285–291.
- 179. Turnis ME, Sawant DV, Szymczak-Workman AL, et al. Interleukin-35 limits anti-tumor immunity. *Immunity* 2016; 44(2): 316–329.
- 180. Takahashi R, Macchini M, Sunagawa M, et al. Interleukin-1β-induced pancreatitis promotes pancreatic ductal adenocarcinoma via B lymphocyte–mediated immune suppression. *Gut* 2021; 70(2): 330–341.
- 181. Wang H-M, Zhang X-H, Feng M-M, et al. Interleukin-35 suppresses the antitumor activity of T cells in patients with non-small cell lung cancer. *Cell Physiol Biochem* 2018; 47(6): 2407–2419.
- 182. Huang C, Li Z, Li N, et al. Interleukin 35 expression correlates with microvessel density in pancreatic ductal adenocarcinoma, recruits monocytes, and promotes growth and angiogenesis of xenograft tumors in mice. *Gastroenterology* 2018; 154(3): 675–688.