



## Characterization the response of Korl:ICR mice to loperamide induced constipation

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Animal models of constipation induced with drugs and diet have been widely employed to investigate therapeutic effects and the action mechanism of drugs against this disease. ICR mice were selected to produce this disease model through oral administration of loperamide (Lop), even though SD rats are commonly utilized in studies of constipation. To compare the responses of ICR mice obtained from three different sources to constipation inducers, alterations in stool number, histopathological structure, mucin secretion and opioid-receptor downstream signaling pathway were measured in Korl:ICR (Korea FDA source), A:ICR (USA source) and B:ICR (Japan source) injected with low and high concentrations of Lop (LoLop and HiLop). The number, weight and moisture content of stools decreased significantly in the Lop treated group of all ICR relative to the Vehicle treated group. Additionally, decreased mucosa layer thickness, muscle thickness, and mucin secretion were observed in the transverse colon of Lop treated ICR mice, while a similar number of goblet cells and crypt of lieberkuhn were detected in the same group. Furthermore, a similar change in the level of G $\alpha$  expression and PKC phosphorylation was detected in the Lop treated group relative to the vehicle treated group, while some differences in the change pattern were observed in the B:ICR group. Therefore, these results of the present study provide strong additional evidence that Korl:ICR, A:ICR and B:ICR derived from different sources have a similar overall response to constipation induced by Lop injection, although there were a few differences in the magnitude of their responses.

**Keywords:** Korl:ICR, constipation, loperamide, excretion parameters, mucin secretion

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Constipation is a chronic gastrointestinal disorder characterized by infrequent bowel movements, difficulty during defecation, and sensation of incomplete bowel evacuation [1-3]. This disease is often caused by various factors such as insufficient dietary fiber intake, inadequate fluid intake, decreased physical activity, side effects of medication, hypothyroidism, and obstruction by colorectal cancer [4]. In addition, chronic constipation is classified into one of three groups by assessment of colonic transit

and anorectal function; slow transit constipation, pelvic floor dysfunction (functional defecatory disorders) and normal transit or irritable bowel syndrome [5]. However, more than half of patients with chronic constipation show normal transit (59%), while only 25% of patients show functional defecatory disorders, slow transit (13%) or a combination of these conditions (3%) [6].

To date, a wide variety of animal models for human diseases have been applied to investigate the therapeutic

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effects of chemical drugs or herbal medicines, as well as to study the mechanism of action of candidate drugs. The data obtained using these models have contributed greatly to our understanding of the complex motor patterns known to exist in the human colon [7]. Two methods are commonly used to produce constipation animal models, drug and diet induction. Oral administration and subcutaneous injection of Lop hydrochloride have been shown to successfully induce chronic constipation in many previous studies [8-9], while feeding a low-fiber diet containing 41.5% cornstarch, 24.5% milk casein, 10.0% sucrose, 10.0% dextrin, 7.0% mineral mixture, 6.0% corn oil, and 1.0% vitamin mixture for 5 weeks was induced the same condition [10].

Several strains of rodents have been utilized to conduct most experiments in studies of Lop-induced constipation, including SD rats, Wistar rats, Balb/c mice, ICR mice and ddY mice [8,11-14], although ICR mice have only been used in two studies [15-16]. However, most studies conducted to date have used only one strain of one animal species to produce animal models for constipation and to evaluate the anti-constipation activity of drugs and extracts from herbal medicines. Moreover, no studies have provided scientific evidence for comparative analysis of the response of ICR mice derived from different sources to chemicals that cause constipation.

Therefore, in the present study, we was compared the response of ICR mice (Korl:ICR, A:ICR and B:ICR) derived from three difference sources to Lop induced constipation and evaluated the characteristics Korl:ICR mice established by the Korea FDA. These results presented herein provide the first scientific evidence of a similar response to Lop induced constipation in the transverse colon of Korl:ICR, A:ICR and B:ICR, although there were slight differences in the magnitude of these effects.

## Materials and Methods

### Experimental design for animal study

The animal protocol used in this study was reviewed and approved based on the ethical procedures for scientific care set by the Pusan National University-Institutional Animal Care and Use Committee (PNU-IACUC; Approval Number PNU-2014-0572). Six-week-old male ICR mice were obtained from three difference sources. Specifically, Korl:ICR mice were kindly provided by the Department

of Laboratory Animal Resources of the National Institute of Food and Drug Safety Evaluation (NIFDS, Chungju, Korea). The other two group of ICR mice (A:ICR and B:ICR) were purchased from vendors located in the United States (Vendor A) and Japan (Vendor B). All mice were handled in the Pusan National University-Laboratory Animal Resources Center, which is accredited by the Korea Food and Drug Administration (FDA) (Accredited Unit Number-000231) and AAALAC International according to the National Institutes of Health guidelines (Accredited Unit Number; 001525). Animals were provided with *ad libitum* access to a standard irradiated chow diet (Samtako Inc.) consisting of moisture (12.5%), crude protein (25.43%), crude fat (6.06%), crude fiber (3.9%), crude ash (5.31%), calcium (1.14%), phosphorus (0.99%) and water. During the experiment, rats were maintained in a specific pathogen-free (SPF) state under a strict light cycle (lights on at 08:00 h and off at 20:00 h) at 23±2°C and 50±10% relative humidity.

Constipation of ICR mice were induced based on traditional methods as described in previous studies, with some modification [8,9]. Briefly, 8-week-old ICR mice in each group (n=24) were assigned to either a non-constipation group (Vehicle treated group, n=8) or a constipation group (n=16). Constipation was induced by Lop injection at regular intervals. First, ICR mice were subcutaneously injected with Lop (Sigma-Aldrich, MO, USA) (4 mg/kg weight) in 0.9% sodium chloride twice a day for 4 days. After a 3 day stationary phase, two different concentrations (4 mg/kg or 8 mg/kg) of Lop were administered to two subgroups (LoLop treated subgroup and HiLop treated subgroup) in the same way for 4 days. Additionally, a vehicle treated group was injected with 0.9% sodium chloride alone (Figure 1). At 24 h after final treatment, all animals were euthanized using CO<sub>2</sub> gas, after which tissue samples were acquired and stored in Eppendorf tubes at -70°C until assay.

### Analysis of food intake, water intake and body weight

Alterations in food intake, water consumption and body weight of ICR mice derived from three different sources treated with Vehicle or Lop were measured daily at 10:00 am throughout the experimental period using an electrical balance and a measuring cylinder. All measurements were performed three times to ensure accuracy.

### Measurement of stool parameters

To collect pure stool and urine without any contamination, all ICR mice were bred in metabolic cages (Daejong Instrument Industry Co., LTD, Seoul, Korea) during the experimental period. The stool number and weight were measured as described in previous studies [9,17]. The stools excreted from each ICR mouse was collected at 10:00 am, after which stool samples were weighed three times per sample using an electric balance and the number of stools was counted three times. Also, the reduction rate of stool number was calculated using the following formula;

$$\text{Reduction rat of stool number}=(SV-SL)/SV\times 100$$

SV was represented stool number of Vehicle-treated group, while SL was indicated the stool number of Lop-treated group.

Furthermore, the stool moisture content was analyzed using the following formula;

$$\text{Stool moisture content}=(A-B)/A\times 100$$

A is indicated the weight of stools firstly collected after Lop administration, while B is represented the weight of stools after the dry at 60°C for 12 hr.

### Western blotting

Total proteins (30 mg) collected from the transverse colons of subset groups (Vehicle, LoLop and HiLop treated ICR mice) were separated by 4-20% sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) for 3 h, after which the resolved proteins were transferred to nitrocellulose membranes for 2 h at 40 V. Each membrane was then incubated separately with primary antibody, anti-G $\alpha$  (Abcame, Cambridge, UK), anti-PKC (Cell Signaling Technology Inc., Cambridge, MA, USA), anti-p-PKC (Cell Signaling Technology Inc.), or anti-actin (Sigma-Aldrich, St. Louis, MO, USA) overnight at 4°C. Next, the membranes were washed with washing buffer (137 mM NaCl, 2.7 mM KCl, 10 mM Na<sub>2</sub>HPO<sub>4</sub>, 2 mM KH<sub>2</sub>PO<sub>4</sub>, and 0.05% Tween 20) and incubated with horseradish peroxidase-conjugated goat anti-rabbit IgG (Zymed Laboratories, South San Francisco, CA, USA) at a dilution of 1:1,000 and room temperature for 2 h. Finally, the membrane blots were developed using Chemiluminescence Reagent Plus kits (Pfizer, New York, NY, USA and Pharmacia, New York, NY, USA).

### Histopathological analysis

Transverse colons collected from ICR mice treated with vehicle and Lop were fixed with 10% formalin for 12 h, embedded in paraffin wax, and then sectioned into 5  $\mu$ m thick slices that were stained with hematoxylin & eosin (H&E, Sigma-Aldrich). Morphological features of these sections were observed by light microscopy, after which the mucosa thickness, muscle thickness, flat luminal surface thickness, number of goblet cells and number of crypt of lieberkuhn were measured using Leica Application Suite (Leica Microsystems, Switzerland). The three different sites in the transverse colon of five rats per group were evaluated in triplicate. Data represent the mean $\pm$ SD from three replicates. A *P* value of <0.05 was considered significant after One-way ANOVA.

For mucin staining, transverse colons collected from ICR mice were fixed with 10% formalin for 48 h, embedded in paraffin wax, and then sectioned into 3  $\mu$ m thick slices that were subsequently deparaffinized with xylene and rehydrated. Next, the tissue sections on the slides were rinsed with distilled water and stained with an Alcian Blue Stain kit (IHC WORLD, Woodstock, MD, USA). Finally, the morphological features in the stained colon sections were observed by light microscopy.

### Statistical analysis

One-way ANOVA was used to determine whether or not significant differences existed between the Korl:ICR group and the A:ICR or B:ICR group (SPSS for Windows, Release 10.10, Standard Version, Chicago, IL, USA). All values were expressed as the means $\pm$ SD. A *P* value of <0.05 was considered significant.

## Results

### Establishment of induction condition for constipation

To establish the induction conditions for constipation, Lop (5 mg/kg) was administered to Korl:ICR mice using two different methods, oral administration and subcutaneous injection, for 3 continuous days. However, a significant decrease of stool number and weight was not detected in any ICR groups (Table 1). We next tried a novel strategy consisting of two injection phases and one stationary phase to induce constipation. During the first 4 day, 4 mg/kg of Lop were subcutaneously injected into Korl:ICR mice, after which they were allowed to rest for 3 days. Following the resting period, Korl:ICR mice

**Table 1.** Alteration on the excretion parameters after Lop administration using two different methods

Category	Continues injection			Interval injection	
	Vehicle	LoLop	HiLop	LoLop	HiLop
Food intake (g/day)	8.83±1.14	7.50±0.76	7.95±0.65	6.50±0.54*	6.87±0.87*
Water consumption (mL)	15.21±2.84	14.00±2.74	14.5±2.33	14.98±2.51	13.52±1.85*
Stool number (ea)	87.00±12.03	82.13±16.19	78.86±13.81	69.11±10.9*	44.83±12.21*
Stool weight (g)	1.40±0.12	1.28±0.19	1.25±0.11	1.02±0.18*	0.74±0.16*
Water contents (%)	45.45±1.7	45.80±0.64	40.73±0.71	39.00±0.50*	29.00±1.60*

Data represent the means±SD from three replicates. \*,  $P<0.05$  compared to Vehicle treated group.

**Table 2.** Measurement of body weight, feeding behavior, stool and urine secretion in Lop-induced constipated ICR mice

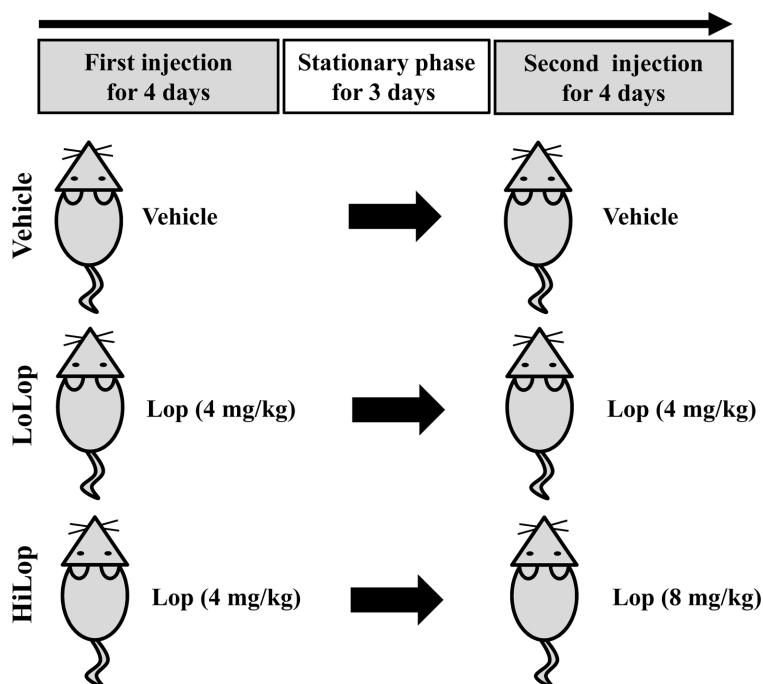
Category	Subgroup	Group		
		Kort:ICR	A:ICR	B:ICR
Body weight (g)	Vehicle	30.33±4.64	32.5±0.71	35.33±1.65
	LoLop	30.83±1.25	31.33±0.71	35.83±1.43
	HiLop	31.63±2.65	34.13±1.43	34.75±1.48
Food intake (g/day)	Vehicle	8.50±3.63	18.83±2.46*	9.17±3.47
	LoLop	7.50±1.78	18.33±1.54*	10.33±0.47*
	HiLop	6.38±1.14	17.25±2.56*	8.63±1.95
Water consumption (mL)	Vehicle	14.21±2.84	28.33±6.24*	13.33±4.71
	LoLop	14.98±3.51	28.33±8.5*	15.67±3.3
	HiLop	13.52±1.85	25.00±6.12*	11.25±2.17
Stool number (ea)	Vehicle	85.00±14.20	115.00±4.64*	72.00±11.89
	LoLop	69.00±10.90 (18.82±1.52)	98.00±3.42* (14.78±0.93)*	55.55±13.49 (22.85±5.33)*
	HiLop	45.00±12.21 (47.06±5.27)	74.00±6.71* (35.65±3.11)*	56.91±7.88 (20.96±4.34)*
Stool weight (g)	Vehicle	1.16±0.12	1.68±0.19*	1.19±0.31
	LoLop	1.02±0.23 (13.19±3.17)	1.57±0.24* (6.97±0.22)*	0.92±0.07 (18.70±4.15)
	HiLop	0.64±0.18 (45.85±5.15)	1.08±0.22* (36.38±3.20)	0.73±0.12 (37.01±4.26)
Stool moisture content (%)	Vehicle	41.61±0.06	64.17±0.08*	57.46±0.04*
	LoLop	39.57±0.05	54.78±0.07*	28.48±0.01*
	HiLop	29.43±0.16	41.98±0.09*	12.82±0.04*
Urine volume (mL)	Vehicle	1.87±0.16	2.91±0.41*	2.09±0.26
	LoLop	2.16±0.22	1.94±0.59	2.07±0.43
	HiLop	2.75±0.33	2.23±0.46	1.75±0.25*

Data represent the means±SD from three replicates. \*,  $P<0.05$  compared to Kort:ICR. Reduce rate (%) of stool number and weight were represented in parenthesis.

received two different concentrations of Lop, 4 mg/kg (LoLop) and 8 mg/kg (HiLop), for 4 days. The number, weight and moisture content of stool decreased by 15-47%, 12-44% and 15-79% in the LoLop and HiLop treated subgroup, while feeding behavior related factors were maintained at a constant level (Table 1). Taken together, these results indicate that the interval injection of Lop can be used to induce constipation in ICR mice derived from three different sources.

#### Alteration of feeding behavior and excretion parameters

We investigated whether Lop treatment could affect the feeding behavior and excretion parameters of constipated ICR mice derived from three different sources. To accomplish this, food intake, water consumption, and stool number and weight were measured in the three constipated ICR groups after Lop treatment. As shown in Table 2, three related factors, feeding behavior, body weight, and food and water consumption, did not differ



**Figure 1.** Strategy for induction of chronic constipation using Lop. After the first injection of Lop (4 mg/kg) for 4 days, ICR mice were allowed to rest for 3 days. Two different concentrations (4 and 8 mg/kg) of Lop were then subcutaneously injected into three groups of ICR mice to produce constipated mice.

significantly between the Vehicle treated subgroup and the Lop treated subgroups within each group, although water consumption was slightly lower in the HiLop treated subgroup. Among these parameters, food intake and water consumption were higher in the A:ICR group than the Korl:ICR. However, the stool number, weight and moisture content were dramatically decreased in the Lop treated subgroup relative to the Vehicle treated subgroup. Furthermore, these dose dependent decreases were commonly observed in the Korl:ICR, A:ICR and B:ICR group, although slight differences in the decrease rate of stool number were observed (Table 2, Figure 2). Specifically, the stool number of the B:ICR showed a lower decrease rate in the Lop treated subgroup and the decrease rate did not depend on the increase in Lop concentration (Table 2, Figure 2). Therefore, the above results suggest that Lop injection at regular intervals can successfully decrease some excretion parameters, but does not lead to any significant alterations in feeding behavior. Furthermore, these changes were commonly observed in Korl:ICR, A:ICR and B:ICR mice.

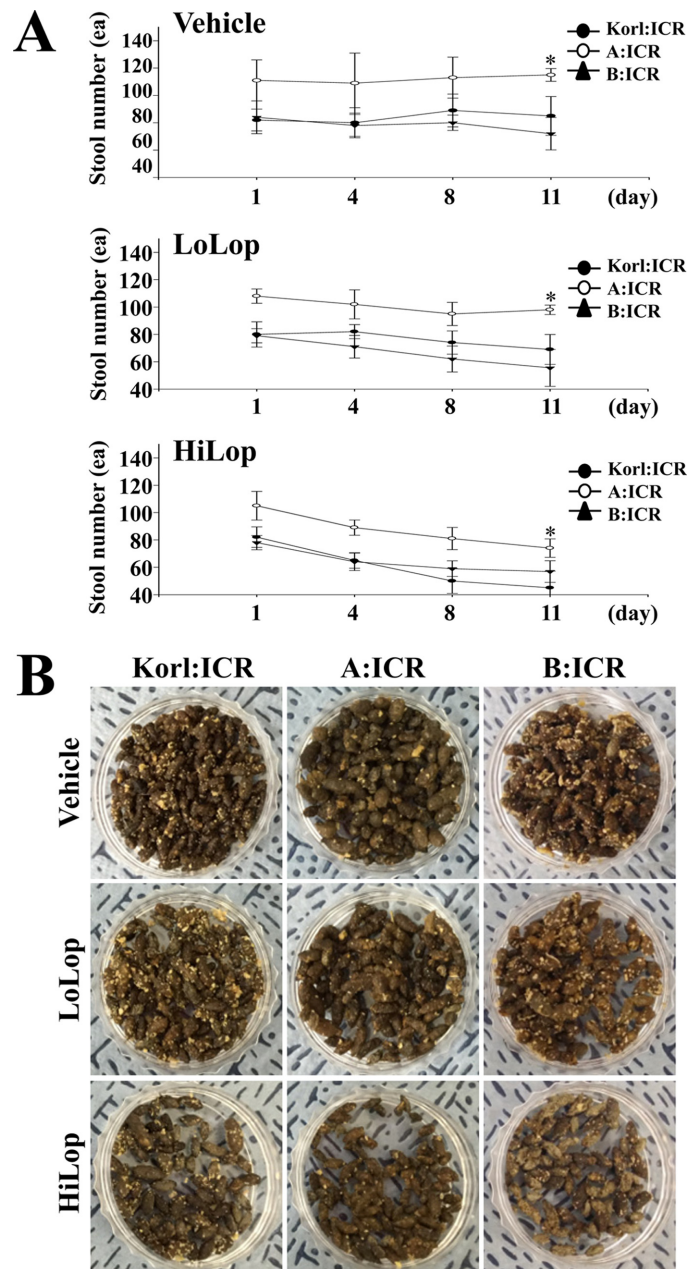
#### Histological alterations in the transverse colon

Several histological parameters were measured in H&E stained transverse colons of Korl:ICR, A:ICR and

B:ICR following treatment with Lop. Although there were a few variations in the statistical significance of each group, histopathological changes were observed in all Lop treated subgroups. The muscle thickness and mucosa layer length was significantly shorter in the Lop treated subgroup than the Vehicle treated subgroup, and these decreases depended on the increase of Lop concentration. Furthermore, the highest levels of muscle thickness and mucosa layer length were detected in the transverse colon of A:ICR (Figure 3). Therefore, the above results suggest that the histopathological structure of constipation was successfully induced by Lop treatment of Korl:ICR, A:ICR and B:ICR mice from three different sources.

#### Alteration of ability to secrete mucin

To characterize the ability to secrete mucin in the transverse colon of ICR mice derived from three different sources following Lop injection, the levels of mucin in tissue sections stained with Alcian blue were observed. The tissue region stained with dark blue color was concentrated in the crypt in the mucosa layer of the transvers colon of all subset groups. However, the total level of mucin decreased dramatically in the Lop treated subgroup compared to the Vehicle treated subgroup.

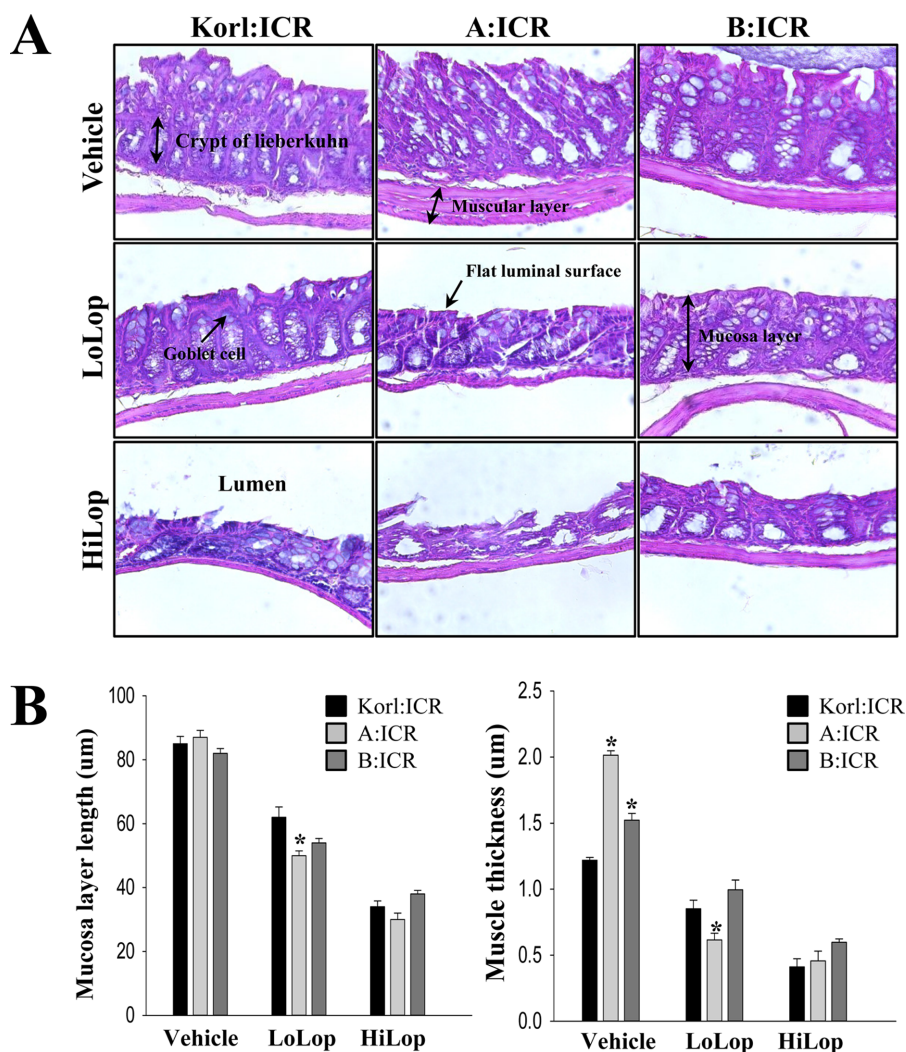


**Figure 2.** Alteration of the number and morphology of stools in three ICR groups after Lop injection. (A) The number of stools were observed during two Lop injection phases and one stationary phase in Vehicle, LoLop and HiLop treated mice. Numbers were counted three times and data represent the means $\pm$ SD of three replicates. \*,  $P < 0.05$  compared to the Korl:ICR group. (B) Following the final treatment, total stools were collected from metabolic cages and their morphology was observed.

Furthermore, three ICR groups showed similar patterns in the ability to secrete mucin following Lop injection (Figure 4). Therefore, the results of the present study suggest that suppression of the ability to secrete mucin may be successfully induced by Lop injection in the transverse colon of Korl:ICR, A:ICR and B:ICR from three different sources.

#### Alteration of G protein signaling pathway

Finally, we examined whether constipation induced by Lop injection was accompanied by altered G protein signaling by measuring the expression of  $G\alpha$  and PKC protein in the transverse colon of subset groups.  $G\alpha$  expression was lower in the LoLop treated subgroup than the Vehicle treated subgroup. Following HiLop



**Figure 3.** Alteration of histopathological structure in Lop-induced constipated ICR mice. H&E stained sections of transverse colons rats from the Vehicle treated group, LoLop treated group or HiLop treated group were observed at 200× magnification using a light microscope. The length of the mucosa layer and muscle thickness in each tissue were measured using Leica Application Suite. Data represent the means±SD of three replicates. \*,  $P < 0.05$  compared to the Korl:ICR group.

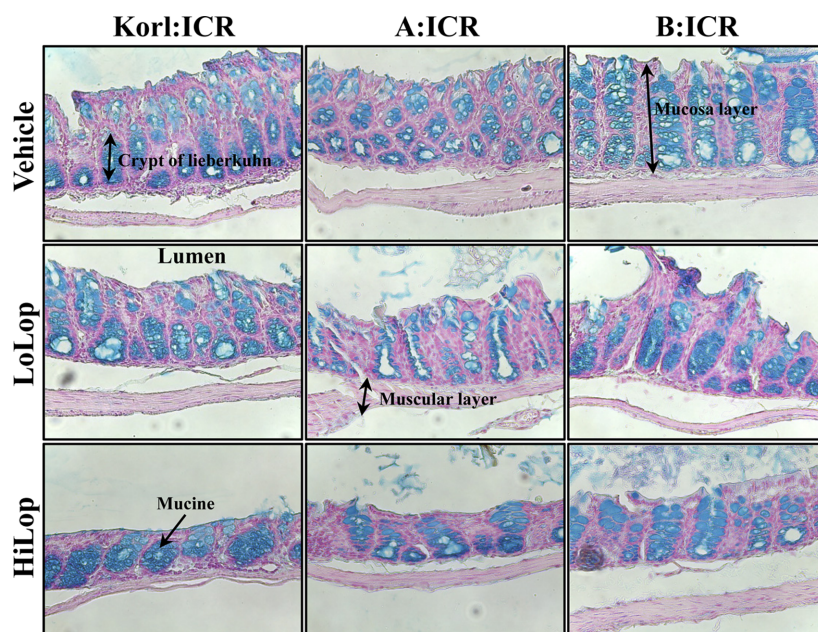
injection, this level was slightly increased relative to that of the LoLop treated group, regardless of concentration decrease. In addition, these patterns of  $G\alpha$  expression were commonly observed in Korl:ICR, A:ICR and B:ICR mice (Figure 5).

However, PKC phosphorylation showed the different patterns than  $G\alpha$  expression. Specifically, the level of PKC phosphorylation was dramatically decreased in the Lop treated subgroup relative to the Vehicle treated subgroup in a dose dependent manner, although the decrease rate varied. The lowest decrease in PKC phosphorylation was detected in the LoLop treated subgroup of the A:ICR mice (Figure 5B). Nevertheless, the results indicate that the G protein signaling pathway

of Korl:ICR, A:ICR and B:ICR mice showed a similar response to Lop injection.

## Discussion

Animal models for constipation have been induced by various agents including Lop [8], clonidine [15], morphine [18], opioid receptor antagonist [19], clozapine [20] and carbon [21-22]. Among these, Lop and carbon have been applied to produce constipation models in many studies. In addition, appropriate rodent species with high susceptibility have been selected from the many available mouse and rat models. However, ICR mice have rarely been used to produce chronic constipation models, even



**Figure 4.** Mucin secretion in the transverse colon of Lop-induced constipated ICR mice. Mucin secreted from crypt layer cells was stained with alcian blue at pH 2.5 and their morphology were observed at 200× magnification. Five to six rats per group were assayed in triplicate by alcian blue staining.

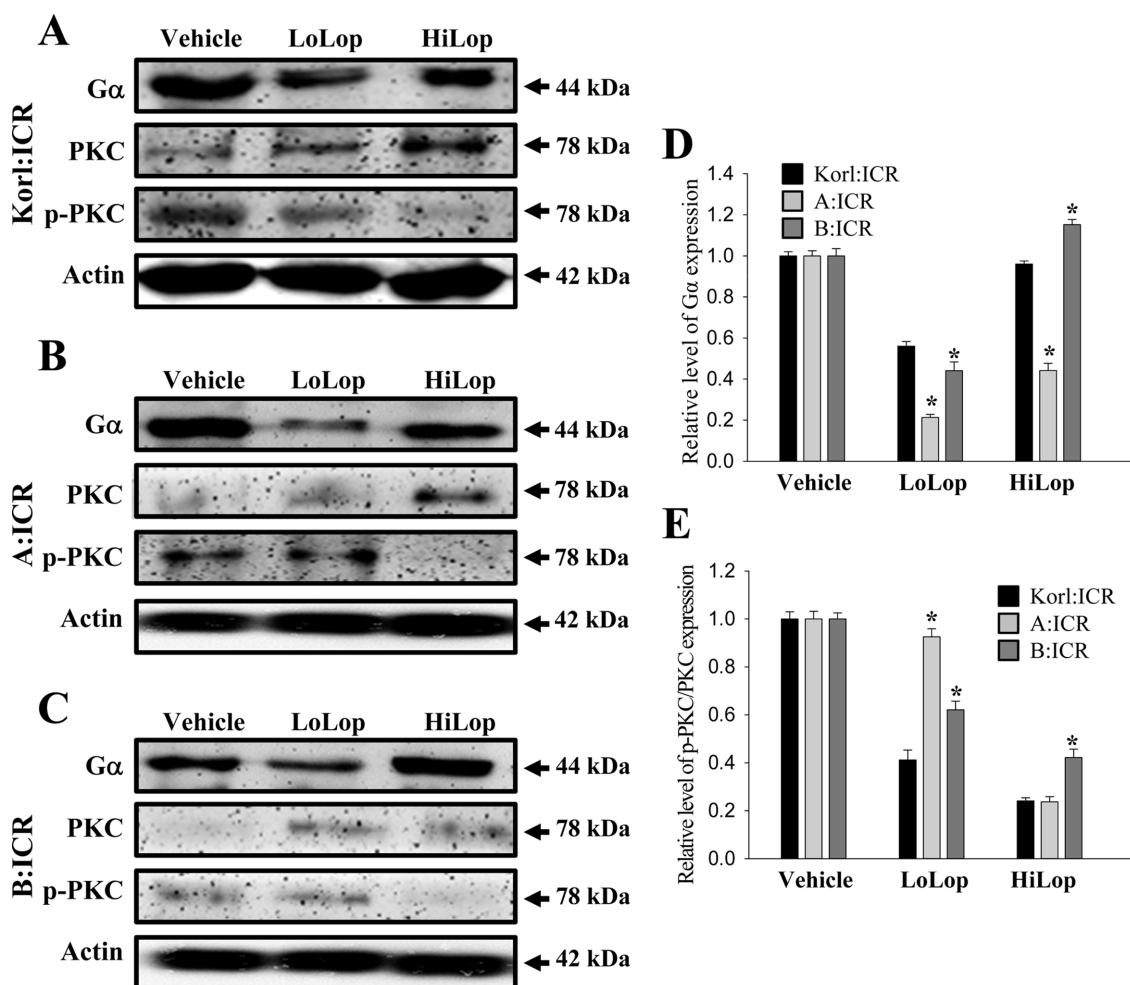
though they also widely used in toxicology (safety and efficacy testing), cancer research, aging, transgenesis experiments and gene mapping strategies [23-25]. In this study, we investigated whether Korl:ICR, A:ICR and B:ICR mice could be utilized to investigate chronic constipation. We also applied a novel strategy in which two different concentrations (4 and 8  $\mu\text{g}/\text{mL}$ ) of Lop were utilized to produce an animal model for constipation and investigated differences in the responses to these chemicals among ICR mice derived from three different sources. The results presented here provide the first evidence that Korl:ICR, A:ICR and B:ICR show an overall similar response to Lop induced constipation, although some differences were detected in  $G\alpha$  expression and PKC phosphorylation.

Lop is well documented as a constipation inducer and commonly used to induce models of spastic constipation. Lop inhibits intestinal motility and fluid secretion, leading to reduced fecal evacuation time and intestinal luminal transit [26]. In the case of SD rats, constipation was successfully induced by subcutaneous injection of 3-5 mg/kg Lop for 3-6 continuous days in most experiments [8]. However, constipation phenotypes such as decreased stool number and weight in ICR mice were observed after oral administration of 5 mg/kg Lop [15-16]. In the present study, we attempted to induce constipation in ICR mice derived from three different sources using Lop

injection. Following subcutaneous injection of Lop (4 mg/kg) for 4 days, mice were allowed to rest for 3 days, after which they were injected with 4 or 8 mg/kg of Lop for 4 days. This strategy successfully induced the constipation in ICR mice, although their pathological symptoms varied. Actually, the interval injection strategy has been widely applied to induce immunological responses such as vaccination and allergen immunotherapy [27].

Since being developed by the Dr. T. S. Hauschka of Fox Chase Cancer Center in 1948, the ICR mice used in this study have been extensively applied to toxicology and pharmacology studies and product safety testing because they have good reproductive performance and rapid growth [23-25]. In addition, ICR mice have been used in several comparison studies to characterize the response of physiological properties to chemicals or hormones. Studies using his model have focused on the incidence of 5-azacytidine-induced exencephaly [28], lung injury by butylated hydroxytoluene [29], styrene-induced hepato/pneumotoxicity [30] and the response to growth hormone by chondrocytes [31]. However, the response to Lop has not been examined in ICR mice from different sources to date. The results presented herein provide the first evidence that a similar response can be detected in Korl:ICR, A:ICR and B:ICR provided from the Korea FDA or commercial vendors, although there were slight differences in the response rate within





**Figure 5.** Alteration of the level of  $G\alpha$  expression and PKC phosphorylation. The  $G\alpha$  expression and PKC phosphorylation within opioid-receptor downstream signaling pathway was measured by Western blotting using HRP-labeled anti-rabbit IgG antibody. After the intensity of each band was determined using an imaging densitometer, the relative levels of protein were calculated based on the intensity of actin protein. Data represent the means  $\pm$  SD of three replicates. \*,  $P < 0.05$  compared to the Korl:ICR group.

each analysis factor. Moreover, further studies are needed to investigate the response to other constipation inducers, including carbon and opioids.

Overall, we established the induction conditions for constipation using Lop in ICR mice as well as investigated the responses of ICR mice provided from different sources to two concentrations of Lop with constipation inducing activity. Three ICR groups, Korl:ICR, A:ICR and B:ICR, showed a similar response of stool parameters, histopathological structure and signaling pathways of opioid receptors to constipation inducers. Therefore, our results suggest that Korl:ICR mice, as well as ICR mice from other commercial vendors can be widely applied to produce animal models for chronic constipation.

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**Conflict of interests** The authors declare that there is no financial conflict of interests to publish these results.

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