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TRYPTOPHAN RECOVERY INDEX AS A NEW BIOMARKER FOR FITNESS

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ABSTRACT

The maximal oxygen uptake (VO₂max) and maximal power output (P_{max}) are commonly used parameters to evaluate the endurance fitness status. A connection between exercise and the kynurenine pathway (KP), which describes the metabolism of unused tryptophan, has already been reported. However, a potential association of the KP with endurance fitness levels remains unknown. In this study, adolescent competitive athletes performed an exhaustive incremental exercise test. Blood samples were taken before, directly after, and 30 minutes after the end of exercise. Tryptophan (Trp), kynurenine (Kyn) and kynurenic acid (KA) serum levels were determined by high-performance liquid chromatography (HPLC). Forty-four male and 27 female athletes (median age: 16 years) were recruited. During exhaustive exercise tests, Trp initially declined and then increased 30 minutes after discontinuing exercise. Similar findings were observed for Kyn, whereas KA levels behaved inversely. After incremental exhaustive exercise the relative increase of Trp concentrations, termed the tryptophan-recovery-index (TRI), showed a highly significant positive correlation with VO₂max and P_{max} (r=0.468 and 0.491, p-values <0.001). There was a significant gender-difference with higher levels of all metabolites at all measured time points in male participants. In the present study, a highly significant correlation was found between the TRI and the maximal oxygen uptake in well-trained athletes. The implementation of TRI can therefore be suggested as a biomarker for physical fitness.

Keywords: Tryptophan, kynurenine, kynurenic acid, kynurenine pathway, exercise

INTRODUCTION

The evaluation of fitness is a much-debated topic not only amongst sports medicine specialists, but also in popular science. While the ideal fitness parameter depends on the kind of sport, the maximal oxygen uptake (VO_2max) and the maximal power output (P_{max}) during an exhaustive incremental exercise test are commonly used markers. Beside these the aerobic and anaerobic threshold are also frequently applied.

Recently, different authors investigated a possible connection between exercise and the kynurenine pathway (KP) (Martin et al., 2020). This pathway describes the metabolism of approximately 90-99 % of the unused essential amino acid tryptophan (Trp) (Palego et al., 2016; Richard et al., 2009; Stone and Darlington, 2002). The first enzymes of the are the tryptophan-2,3-dioxygenase KP (TDO) and different subtypes of the indoleamine-2,3-dioxygenase (IDO) (Chen and Guillemin, 2009; Ball et al., 2007), which metabolize Trp to kynurenine (Kyn). Since the main stimulator of IDO is interferon-y (IFN- γ), the expression and activity of IDO are strongly related to inflammatory processes (Chen and Guillemin, 2009; Grant et al., 2000). Consequently, changes in the KP have been described in a variety of diseases (Nagy et al., 2017), also including psychiatric disorders, such as depression (Chen and Guillemin, 2009; Song et al. 2017). While exercise is thought to be beneficial in individuals with mental disorders, it was supposed that the mood enhancing effects of exercise could be caused by changes of the KP (Allison et al., 2019; Agudelo et al., 2019).

Results of previously published studies examining exercise related alterations of the KP are inconsistent. Data have shown a decrease of Trp while Kyn mostly remained unchanged. Currently, the exercise induced kinetics of KA are poorly investigated (Metcalfe et al., 2018). It is not fully understood yet, why the KP changes after exercise and how exercise can influence the KP. While many authors described a higher activity of KP-enzymes (Strasser et al., 2016a; b; Ito et al., 2003; Koliamitra et al., 2019) a shift of KP-metabolites into other body compartments could be another explanation for the metabolic kinetics (Areces et al., 2015). In trained athletes, a permanent adaptation of the KP can be hypothesized. As a consequence, it may be possible to deduce the fitness level by investigating alterations of the KP. This is of great interest, considering that there is no biochemical parameter available, which can estimate somebody's fitness level.

Therefore, the aim of the present study was to investigate possible associations between the kinetics of Trp metabolites and fitness parameters. We studied adolescent trained athletes during incremental exhaustive exercise testing for changes in the KP. The main focus of this study was on the recovery efficiency.

MATERIALS AND METHODS

Study population and exercise testing

A total of 71 athletes (44 males, 27 females) were included in this study after written informed consent of the patient or the legal guardian. The inclusion criteria were athletes aged between 14 and 18 years, visiting the outpatient clinic for a voluntary performance review. Each study participant underwent a clinical examination for the suitability of a physical load (12-lead ECG), determination of the resting heart rate (Assy Cam 14®, GE Healthcare, Chicago, Illinois, USA), resting blood pressure (oscillatory, Boso Medicus®, BOSCH+SOHN, Jungingen, Germany), physical examination, an estimation of the total body fat (Caliper, John Bull, British Indicators Ltd., St Albans, UK) (Jackson and Pollock, 1978), and a history of the accomplished training. Athletes were excluded from the study in case of incomplete study parameters.

An exhaustive incremental exercise protocol was performed on a bicycle ergometer (Excalibur Sport®, Lode, Groningen, Netherlands). During the test, we recorded a 12-lead ECG and a continuous spirometry (both Jaeger Oxycon ProTM, Hoechberg, Germany), and a pulse oximetry (BCI® Autocorr® Pulse Oximeter, Minneapolis, USA). Blood was drawn from an indwelling venous cannula before (t₁), immediately after the exhaustive incremental exercise test (t₂), and after a 30 minutes recovery period (t₃). Ethical approval was provided by the local Ethical Committee 27-406 ex 14/15. The study was performed according to the declaration of Helsinki. Informed written consent was provided by all participants.

Analysis of KP key metabolites

Blood samples were drawn with 2.8 mL standard serum tubes (*Vacuette*®, *Greiner Bio-One*, Kremsmünster, Austria) and were allowed to clot for 30 minutes. After centrifugation at 4000 x g for 10 minutes, 1 mL serum portions of the supernatant were harvested and stored at -80 °C until further evaluation.

Trp, Kyn and KA were measured by highperformance liquid chromatography (HPLC) coupled with a simultaneous ultraviolet and fluorometric detection system (Hervé et al., 1996; Xiao et al., 2008). All evaluations were performed according to the published guidelines (FDA, 2018). Within-day coefficients of variation (CVs) at different concentrations were in the range between 1.7 % to 4.3 % for Kyn, 0.7 % to 2.9 % for Trp and 2.6 % to 4.5 % for KA. The between-day CVs were 2.0 % to 5.4 %, 6.3 % to 9.3 %, and 8.4 % to 11.6 %, respectively.

Statistical analysis

Non-normally distributed data were displayed as medians and interquartile ranges (IQR). Correlation analyses between parameters were performed using a Spearman's rho (ρ) test. For comparison of independent groups (gender differences) a Mann-Whitney-U-test was applied. Differences between dependent variables (KP kinetics) were examined with a Friedmann-test. In case of significant results a pairwise comparison with a Wilcoxon-test followed by the Bonferroni-Holm correction for multiple testing was performed. Associations between parameters were calculated with simple linear regression model. Pvalues <0.05 were regarded as statistically significant. Statistical analysis was performed with SPSS® Statistics 22 (IBM®, Armonk, USA).

Tryptophan recovery index (TRI), kynurenine recovery index (KRI) and

kynurenic acid recovery index (KARI) were calculated by the following formulae:

$$TRI = \frac{Trp(t_3) - Trp(t_2)}{Trp(t_2)} x \ 100,$$
$$KRI = \frac{Kyn(t_3) - Kyn(t_2)}{Kyn(t_2)} x \ 100,$$
$$KARI = \frac{KA(t_3) - KA(t_2)}{Kyn(t_2)} x \ 100.$$

$$KA(t_2)$$

RESULTS

Study population characteristics

A total of 71 athletes (44 males, 27 females) with a median age of 16 (range: 14-18) years were examined. Detailed data for the participants are given in Table 1. Median concentrations of Trp, Kyn and KA at t₁ were significantly higher in men compared to women (Trp: 70.3 vs. 60.3 μ mol/L, p=0.004; Kyn: 3.06 vs. 2.66 μ mol/L, p<0.001; KA: 47.1 vs. 39.9 nmol/L, p=0.005).

Exercise induced KP kinetics

Independently from sex, Trp and Kyn concentrations significantly decreased after exhaustive effort (t_2 vs. t_1), while KA significantly increased (all p-values ≤ 0.001). At t_3 Trp and Kyn concentrations increased (all p-values < 0.001) while KA (p<0.001) decreased significantly compared to t_2 . At t_3 , Trp was still significantly lower (p<0.001 in men and p=0.003 in women) than at t_1 . However, there was no significant change between Kyn concentrations between these points of time. Only in men, KA concentrations were significantly increased at t_3 compared to t_1 (p < 0.001). The kinetic of Trp blood concentrations is demonstrated in Figure 1.

Correlations between kinetics of KP metabolites with fitness parameters

After incremental exhaustive exercise the relative increase of Trp concentrations (TRI) showed a highly significant positive correlation with VO₂max and P_{max} (r=0.468 and 0.491, p-values <0.001). The correlations of VO₂max and P_{max} were stronger in female (r=0.713 and 0.794, p-values <0.001) than in

male athletes (r=0.429 and r=0.452, p=0.004 and p=0.002). A significant positive correlation between the relative increase of Kyn concentrations (KRI) and fitness parameters (VO₂max and P_{max}) (r=0.433 and 0.581, p= 0.024 and p=0.001) was observed in females, only. No significant correlation was found between the relative decrease of KA (KARI) and fitness parameters. Detailed results of the TRI are shown in Table 2 and Figure 2. The anonymized raw data of the present study are provided in Supplementary data (Table 1).

DISCUSSION

In this study, we found a strong correlation between the increase of Trp concentrations after exercise and fitness parameters in adolescent athletes. To the best of our knowledge, this is the first study describing the ratio of Trp concentrations immediately and 30 minutes after exercise in correlation to the fitness parameters VO_2max and P_{max} . This ratio, called TRI, describes the recovery efficiency of Trp.

Anthropometric data									
	Male			Female					
Ν	44			27					
Age (years)	15.8 (15.0-16.8)			16.2 (15.0-16.7)					
Height (cm)	177 (171-180)			164 (1.63-1.69)					
Weight (kg)	64.0 (58.3-71.0)			55.0 (51.0-59.0)					
BMI (m²/kg)	20.7 (19.0-21.7)			20.3 (19.1-21.9)					
Body fat (%)	10.5 (9.2-12.1)			20.0 (18.6-20.9)					
Fitness data									
HR _{maximum} (bpm)	199 (194-205)		202 (194-206)						
Weekly training (h)	12.5 (10.6-16.0)		13.0 (8.0-17.0)						
VO2max (mL/kg/min)	57.7 (53.1-62.2)		45.6 (43.6-54.8)						
P _{max} (W/kg)	4.62 (4.32-5.09)		3.81 (3.40-4.26)						
Measured biochemical data									
	t1	t ₂	t ₃	t 1	t2	t ₃			
Tryptophan (µmol/L)	70.3	53.7	64.4	60.3	50.1	58.4			
	(66.1-	(48.2-	(57.4-	(57.5-	(45.0-	(55.2-			
	77.1)	56.9)	70.0)	71.1)	52.6)	63.6)			
Kynurenine (µmol/L)	3.06	2.86	2.91	2.66	2.31	2.57			
	(2.71-	(2.47-	(2.61-	(2.11-	(2.15-	(2.25-			
	3.57)	3.27)	3.59)	2.87)	2.62)	2.83)			
Kynurenic acid	47.1	65.4	50.6	39.9	58.2	40.9			
(nmol/L)	(38.9- 55.6)	(55.1-	(42.0-	(30.7-	(44.0-	(32.8-			
	· · ·	83.6)	64.0)	46.0)	63.1)	50.1)			
Calculated biochemical data									
TRI (%)		22.3 (17.1-26.1)			21.1 (13.8-25.7)				
KRI (%)		6.03 (2.7-10.3)			7.14 (0.59-9.7)				
KARI (%)		-23.6 (29.2-15.4)		-24.0 (30.6-16.8)					

Data are presented as medians (Q1-Q3). HR, heart rate; bpm, beats per minute; BMI, body mass index; VO₂max, measured maximal oxygen capacity, P_{max}, maximal power performed during incremental exercise testing; TRI, tryptophan recovery index; KRI, kynurenine recovery index; KARI, kynurenic acid recovery index



Figure 1: Exercise induced kinetic of Trp blood concentrations before (t1), immediately after an exhaustive incremental exercise test (t2), and after 30 minutes of recovery (t3). **p=0.003; ***p<0.001

Table O Circula line and an action of TDL KDL and KADL with fits.	
Table 2: Simple linear regression of TRI, KRI and KARI with fitne	ess parameters

Correlation	on	All patients r	Male r	Female r
TRI	P _{max}	0.491**	0.452**	0.794**
TRI	VO₂max	0.468**	0.429**	0.713**
KRI	P _{max}	0.218	0.129	0.581**
KRI	VO₂max	0.150	-0.003	0.433*
KARI	P _{max}	-0.066	-0.156	-0.218
KARI	VO₂max	-0.003	-0.077	-0.131

TRI, tryptophan recovery index; KRI, kynurenine recovery index; KARI, kynurenic acid recovery index. *p<0.05; **p<0.01



We observed a decrease of Trp after exercise, which is in line with previous studies in animals and humans (Metcalfe et al., 2018; Strasser et al., 2016a, b; Areces et al., 2015; Mudry et al., 2016; Lewis et al., 2010; Bansi et al., 2018; Melancon et al., 2014). In rats, Ito et al. observed a mean decrease of 43.5 % of blood Trp concentrations after treadmill exercise with exhaustion (Ito et al., 2003). Mudry et al. found slightly decreased Trp levels in diabetic patients and healthy controls after 30 min training compared to baseline concentrations (Mudry et al., 2016). Also in athletes Trp declined after exhaustive exercise (Strasser et al., 2016a, b), even during ultramarathon running (Yamada et al., 2016). In our study, we observed a decline of Kyn after exercise. In contrast to Trp, contradicting results have been published regarding Kyn alterations after exercise. Data from Yamada et al. (2016) are consistent with our findings. They found a decrease of Kyn after 35 km ultramarathon running. However, other authors describe an increase of Kyn after exercise (Strasser et al., 2016a, b; Mudry et al., 2016; Ito et al., 1999). Contrary to Trp and Kyn, we found a distinct increase (approximately 40 %) of KA after exercise. Little data are available about alterations of KA after exercise. Our findings confirm one previously published study (Mudry et al., 2016).

Nevertheless, the explanation for alterations of Trp and KP metabolites during and after exercise is an emerging topic of current discussions. Previous studies reported a shift of amino acids due to exercise from the vascular compartment into skeletal muscle (Strasser et al., 2016b; Areces et al., 2015). This shift may be facilitated by an enhanced expression of the large amino acid transporter 1 (LAT1), which carries Trp into skeletal muscle (Martin et al., 2020; Pillon et al., 2020). A higher physiological energy consumption in skeletal muscle could be one possible explanation for the observed Trp kinetics for energy production (Agudelo et al., 2019). Moreover, during exercise, the PPAR- γ coactivator-1 α 1 (PGC-1 α 1) enhances the expression of kynurenine-aminotransferases (KATs) in skeletal muscle elevating the production of KA (Agudelo et al., 2019; Agudelo et al., 2014). Subsequently, more KA is shifted from skeletal muscle into the circulation (Martin et al., 2020). This theory is in line with our data showing a Kyn decrease and KA elevation after exercise. Allison et al. showed that a 12-week exercise program increases the expression of skeletal muscle transcription factors PGC-1 α , PPAR α and PPAR δ , but they did not see an effect on the kynurenine metabolism (Allison et al., 2019).

We found a strong association between the TRI and the conventional fitness parameters. In comparison Strasser et al. described a positive correlation between the maximal oxygen consumption (VO₂max) and values of Trp (Strasser et al., 2016b). However, they did neither assess the association between the rebound of Trp after a recovery period nor the fitness status.

It is well known that physical exercise activates the coactivator PGC-1a1, a master coactivator of cellular adaptive processes, which improves fuel supply, uptake and utilization (Correia et al., 2015), but also the expression of KATs (Agudelo et al., 2014). The increased activity of the KATs might lead to higher blood glutamate concentrations, which are important for energy utilization. Moreover, the PGC-1 α 1 regulated pathway in the trained muscle is considered to use Kyn as a supporter for aspartate biosynthesis and mitochondrial function (Agudelo et al., 2019). It might be possible that in trained athletes this pathway is gained. After exercise the no longer needed Trp and Kyn is shifted back faster into the vascular compartment in better trained athletes. Therefore, the TRI is higher in these individuals.

Male athletes had a higher VO₂max compared to female athletes, which correspond with the higher baseline values of the KP. This gender difference was already discussed in prior studies (Strasser et al., 2016b). It was suggested that estrogen and progesterone activate the KP downstream (de Bie et al., 2016). However, we found significantly higher Trp, Kyn and KA in men, which partly is in line with a previous report describing higher Kyn blood concentrations in men, but no differences in other KP metabolites (Strasser et al., 2016b). Another study of the same study group reported lower Kyn levels in women (Strasser et al., 2016a). In contrast to conventional fitness parameters, the TRI might give information about the individual fitness status independently from sex.

Further studies are required to optimize the predictive value of the TRI including duration and degree of the necessary exercise, duration of observation and recovery. If it is possible to determine the TRI in a simple way, it could be a valuable parameter to evaluate the fitness status.

Some limitations of this study should be mentioned. The last meal before blood taking was not standardized due to comfort for our athletes. This circumstance could have an influence on the baseline values of Trp metabolism. Moreover, inflammatory markers, such as interleukin-6 (IL-6) or cortisol were not assessed.

CONCLUSIONS

In the present study, a highly significant correlation could be found between the TRI and the maximal oxygen uptake in welltrained athletes. The authors of this study suggest to use the TRI as a biomarker for physical fitness.

Conflict of interest

The authors declare that there is no conflict of interest.

REFERENCES

Agudelo LZ, Femenia T, Orhan F, Porsmyr-Palmertz M, Goiny M, Martinez-Redondo V, et al. Skeletal muscle PGC-1alpha1 modulates kynurenine metabolism and mediates resilience to stress-induced depression. Cell. 2014;159:33-45.

Agudelo LZ, Ferreira DMS, Dadvar S, Cervenka I, Ketscher L, Izadi M, et al. Skeletal muscle PGC-1alpha1 reroutes kynurenine metabolism to increase energy efficiency and fatigue-resistance. Nat Commun. 2019;10(1):2767.

Allison DJ, Nederveen JP, Snijders T, Bell KE, Kumbhare D, Phillips SM, et al. Exercise training impacts skeletal muscle gene expression related to the kynurenine pathway. Am J Physiol Cell Physiol. 2019; 316:C444-C8.

Areces F, Gonzalez-Millan C, Salinero JJ, Abian-Vicen J, Lara B, Gallo-Salazar C, et al. Changes in serum free amino acids and muscle fatigue experienced during a half-ironman triathlon. PLoS One. 2015;10 (9):e0138376.

Ball HJ, Sanchez-Perez A, Weiser S, Austin CJ, Astelbauer F, Miu J, et al. Characterization of an indoleamine 2,3-dioxygenase-like protein found in humans and mice. Gene. 2007;396:203-13.

Bansi J, Koliamitra C, Bloch W, Joisten N, Schenk A, Watson M, et al. Persons with secondary progressive and relapsing remitting multiple sclerosis reveal different responses of tryptophan metabolism to acute endurance exercise and training. J Neuroimmunol. 2018; 314:101-5.

Chen Y, Guillemin GJ. Kynurenine pathway metabolites in humans: disease and healthy States. Int J Tryptophan Res. 2009;2:1-19.

Correia JC, Ferreira DM, Ruas JL. Intercellular: local and systemic actions of skeletal muscle PGC-1s. Trends Endocrinol Metab. 2015;26:305-14.

de Bie J, Lim CK, Guillemin GJ. Kynurenines, gender and neuroinflammation; showcase schizophrenia. Neurotox Res. 2016;30:285-94.

FDA, Food and Drug Administration, U.S. Department of Health and Human Services, FDA, Center for Drug Evaluation and Research (CDER), Center for Veterinary Medicine (CVM). Bioanalytical method validation - guidance for industry. Rockeville, MA: FDA, 2018.

Grant RS, Naif H, Espinosa M, Kapoor V. IDO induction in IFN-gamma activated astroglia: a role in improving cell viability during oxidative stress. Redox Rep. 2000;5:101-4.

Hervé C, Beyne P, Jamault H, Delacoux E. Determination of tryptophan and its kynurenine pathway metabolites in human serum by high-performance liquid chromatography with simultaneous ultraviolet and fluorimetric detection. J Chromatogr B Biomed Appl. 1996;675:157-61. Ito Y, Saito K, Maruta K, Nakagami Y, Koike T, Oguri Y, et al. Kynurenine concentration of serum was increased by exercise. Adv Exp Med Biol. 1999;467: 717-22.

Ito Y, Yonekura R, Maruta K, Koike T, Nakagami Y, Shibata K, et al. Tryptophan metabolism was accelerated by exercise in rat. Adv Exp Med Biol. 2003;527: 531-5.

Jackson AS, Pollock ML. Generalized equations for predicting body density of men. Br J Nutr. 1978;40: 497-504.

Koliamitra C, Javelle F, Joisten N, Shimabukuro-Vornhagen A, Bloch W, Schenk A, et al. Do acute exerciseinduced activations of the kynurenine pathway induce regulatory T-cells on the long-term? - A theoretical frame work supported by pilot data. J Sports Sci Med. 2019;18:669-73.

Lewis GD, Farrell L, Wood MJ, Martinovic M, Arany Z, Rowe GC, et al. Metabolic signatures of exercise in human plasma. Sci Transl Med. 2010;2(33):33ra7.

Martin KS, Azzolini M, Lira Ruas J. The kynurenine connection: how exercise shifts muscle tryptophan metabolism and affects energy homeostasis, the im-mune system, and the brain. Am J Physiol Cell Physiol. 2020;318:C818-C30.

Melancon MO, Lorrain D, Dionne IJ. Changes in markers of brain serotonin activity in response to chronic exercise in senior men. Appl Physiol Nutr Metab. 2014;39:1250-6.

Metcalfe AJ, Koliamitra C, Javelle F, Bloch W, Zimmer P. Acute and chronic effects of exercise on the kynurenine pathway in humans - A brief review and future perspectives. Physiol Behav. 2018;194:583-7.

Mudry JM, Alm PS, Erhardt S, Goiny M, Fritz T, Caidahl K, et al. Direct effects of exercise on kynurenine metabolism in people with normal glucose tolerance or type 2 diabetes. Diabetes Metab Res Rev. 2016;32:754-61.

Nagy BM, Nagaraj C, Meinitzer A, Sharma N, Papp R, Foris V, et al. Importance of kynurenine in pulmo-nary hypertension. Am J Physiol Lung Cell Mol Physiol. 2017;313:L741-L51. Palego L, Betti L, Rossi A, Giannaccini G. Tryptophan biochemistry: structural, nutritional, metabolic, and medical aspects in humans. J Amino Acids. 2016;2016: 8952520.

Pillon NJ, Gabriel BM, Dollet L, Smith JAB, Sardon Puig L, Botella J, et al. Transcriptomic profiling of skeletal muscle adaptations to exercise and inactivity. Nat Commun. 2020;11(1):470.

Richard DM, Dawes MA, Mathias CW, Acheson A, Hill-Kapturczak N, Dougherty DM. L-Tryptophan: basic metabolic functions, behavioral research and therapeutic indications. Int J Tryptophan Res. 2009;2: 45-60.

Song P, Ramprasath T, Wang H, Zou MH. Abnormal kynurenine pathway of tryptophan catabolism in cardiovascular diseases. Cell Mol Life Sci. 2017;74:2899-916.

Stone TW, Darlington LG. Endogenous kynurenines as targets for drug discovery and development. Nat Rev Drug Discov. 2002;1:609-20.

Strasser B, Geiger D, Schauer M, Gatterer H, Burtscher M, Fuchs D. Effects of exhaustive aerobic exercise on tryptophan-kynurenine metabolism in trained athletes. PLoS One. 2016a;11(4):e0153617.

Strasser B, Geiger D, Schauer M, Gostner JM, Gatterer H, Burtscher M, et al. Probiotic supplements beneficially affect tryptophan-kynurenine metabolism and reduce the incidence of upper respiratory tract infections in trained athletes: a randomized, double-blinded, placebo-controlled trial. Nutrients. 2016b;8(11):752.

Xiao LD, Luo XB, Pi LG, Tang AG. Simultaneous determination of kynurenine and kynurenic acid concentrations in human serum by HPLC with dual wavelengths fluorescence detection. Clin Chim Acta. 2008; 395:178-80.

Yamada N, Shibata K, Fuku M, Kuriki K, Goto C, Tokudome Y, et al. Changes of tryptophan metabolism in Japanese runners during an ultra-marathon race. Sport Sci Health. 2016;12(1):77-83.