



Astroglia in the Vulnerability to and Maintenance of Stress-Mediated Neuropathology and Depression

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Significant stress exposure and psychiatric depression are associated with morphological, biochemical, and physiological disturbances of astrocytes in specific brain regions relevant to the pathophysiology of those disorders, suggesting that astrocytes are involved in the mechanisms underlying the vulnerability to or maintenance of stress-related neuropathology and depression. To understand those mechanisms a variety of studies have probed the effect of various modalities of stress exposure on the metabolism, gene expression and plasticity of astrocytes. These studies have uncovered the participation of various cellular pathways, such as those for intracellular calcium regulation, neuroimmune responses, extracellular ionic regulation, gap junctions-based cellular communication, and regulation of neurotransmitter and gliotransmitter release and uptake. More recently epigenetic modifications resulting from exposure to chronic forms of stress or to early life adversity have been suggested to affect not only neuronal mechanisms but also gene expression and physiology of astrocytes and other glial cells. However, much remains to be learned to understand the specific role of those and other modifications in the astroglial contribution to the vulnerability to and maintenance of stress-related disorders and depression, and for leveraging that knowledge to achieve more effective psychiatric therapies.

Keywords: astrocytes, stress, depression, pathology, epigenetics, glutamate

INTRODUCTION

Stress and the systemic and neurophysiological responses it triggers are major risk factors for depression and other psychiatric disorders. In these disorders as well as in animal models for the effects of stress, astrocytes in several brain regions display morphological and molecular alterations (Bender et al., 2016; Murphy-Royal et al., 2019) that may result in grave consequences for the physiology and connectivity of brain circuits (Fields et al., 2015). A variety of chronic stress-inducing procedures in animals, such as chronic unpredictable stress, repeated fearful social interactions or recurrent restraint in rodents result in reductions in the numbers of astrocyte labeled for specific markers such as glial fibrillary acidic proteins (GFAP), in the levels the mRNA expression of the markers, and in the extent of the labeled processes that stem from their cell bodies in prefrontal cortical areas, the hippocampus or other brain regions (Banasr and Duman, 2008; Liu et al., 2009; Banasr et al., 2010; Araya-Callis et al., 2012; Imbe et al., 2012, 2013; Li L. F. et al., 2013; Li Y. S. et al., 2013; Tynan et al., 2013; Codeluppi et al., 2021). Interestingly, laboratory rats of the Wistar Kyoto strain, known to display spontaneously some depression-like behaviors, show comparable

lower levels of GFAP astrocyte immunostaining (Gosselin et al., 2009). Social defeat stress in a different species, the tree shrew, a marsupial mammal, leads to significant decrease in GFAP positive astrocyte and their cell body size in the hippocampus (Czeh et al., 2006).

Although most examples of chronic stress result in reduction of astrocytes' cell size or their GFAP expression as well as reduced levels of glutamate transporters, acute or short exposures to various stress procedures appear to augment rather than decrease markers of astrocytes (Bender et al., 2016; Yoshino et al., 2020). For example, two exposures in two consecutive days to electric shocks produced learned helplessness in rats, but they increased the expression of glutamate transporter 1 (GLT-1) and glutamine synthetase (Yoshino et al., 2020). Similar results in other studies of the effects of short exposure to stress or adversity, have led some researchers to suggest that moderate or short stress [with the exception perhaps of post-traumatic stress disorder (PTSD)] rather than contributing to eventual depression would increase resilience and that astrocytes alterations may also be involved in positive effects of short-term stresses (Franklin et al., 2012; Menard et al., 2017). Nonetheless, most of the examples of astrocyte involvement in stress effects and depression-related behaviors discussed below are mostly related to chronic stress or stress undergone in early life stages.

It must also be pointed out that repeated exposure to altered concentrations of glucocorticoids in chronic stress may be a major factor determining the pathological changes in astrocytes that contribute to depression-like behavior. For instance, chronic treatment with corticosterone in mice results in reduced numbers of GFAP-positive astrocytes, smaller volume of their cells bodies and shorter process length and induces hippocampal atrophy in mice (Zhang et al., 2015). More recently, exposure to chronic social defeat stress, lipopolysaccharide or corticosteroid analog dexamethasone (DEX) have been shown to result in marked downregulation of glucocorticoid receptor (GR) expression in astrocytes and much less so in neurons in mice (Lu et al., 2021). Absence of those receptors in astrocytes results in depression-like behaviors while astrocyte-specific restoration of the receptors in the medial prefrontal cortex greatly mitigated the depression-like behavior (Lu et al., 2021). Thus, depletion of GR expression in astrocytes of the prefrontal cortex and other brain regions may be a major factor in the various astrocyte-specific molecular changes and the mechanism by which they contribute to long lasting behavioral and emotional effects of stress.

ASTROCYTES AND GLUTAMATE NEUROTRANSMISSION AND REUPTAKE

Stress and the Role of NMDA-Type Glutamate Receptors

In animal models, stress that results in depression- and anxiety-like behaviors induces profound changes in various components of glutamate- and gamma-aminobutyric acid (GABA)-driven neurotransmission (Choudary et al., 2005; Kendell et al., 2005; Karolewicz et al., 2010; McKlveen et al., 2016; Réus et al., 2016).

At the neuronal level, chronic stress and excess glucocorticoids lead to decreased expression of glutamate receptors in the prefrontal cortex, an effect that is dependent on the activation of glucocorticoid receptors (Yuen et al., 2012). In human subjects, elevated levels for N-methyl-D-aspartate-type (NMDA) and metabotropic glutamate receptor subunits were measured in the locus coeruleus of depressed subjects but not in pyramidal cells of the prefrontal cortex (Chandley et al., 2014), although a study found increases of metabotropic glutamate receptor 2/3 in Brodmann's area 10 of the prefrontal cortex (Feyissa et al., 2010). However, the regulation of glutamatergic neurotransmission is also critically dependent on the ability to reuptake released glutamate and recycle it to glutamine by astrocyte processes surrounding synapses, glutamine then being returned to the presynaptic element for further generation and release of glutamate. Thus, astrocytes have a critical role not only in terminating synaptic actions of glutamate but also in ensuring the continuous supply of releasable glutamate (Had-Aissouni et al., 2002).

Astrocytic Glutamate Transporters and Glutamine Synthetase in the Vulnerability to Stress and Depression

Regulation of the extracellular content of glutamate at synaptic and extra-synaptic sites is highly dependent on the activity of glutamate transporters EAAT1 and EAAT2 in the membranes of astrocytes processes, while the recycling of glutamate to glutamine within astrocytes is also performed in astrocytes by the action of glutamine synthetase. In human subjects with depression there are lower levels of glutamate transporters in the prefrontal cortex (Miguel-Hidalgo et al., 2010; Nagy et al., 2015). Similarly, in rodents subjected to chronic stress and in other animal models exhibiting depressive behaviors there is also a decrease in the expression of astrocytic glutamate transporters (Zink et al., 2010; Gomez-Galan et al., 2013; Piao et al., 2019) or glutamate reuptake (Almeida et al., 2010; de Vasconcellos-Bittencourt et al., 2011) in the cortex and hippocampus as compared to non-stressed controls.

Reduction in astrocytic transport relevant to the manifestation of depressive behaviors is not limited to cortical regions of the brain but extends as well to subcortical structures that have been long known to substantially contribute to the pathophysiology of stress effects and depression. In mice with depression-like behaviors caused by early life stress, the immunostaining of GLT1 astrocyte transporters and the thickness of astrocyte processes are reduced in the corticotrophin-releasing factor (CRF)-expressing dorsal-medial (MPD) neurons of the hypothalamus, effects which are concomitant with increased excitability of the surrounding neurons (Gunn et al., 2013). Also in mice, decreased activity of GLT1 transporters in the habenula promotes depression-like behaviors and increases the behavioral vulnerability to depression-like behavioral effects of chronic stress (Cui et al., 2014). Loss of astrocyte glutamate transporter GLT-1 was observed also in the periaqueductal gray matter of rats subjected to chronic restraint stress (Imbe et al., 2012). Thus, the available evidence strongly suggests that

prolonged or repetitive stress results in a generalized or, at least, amply extensive decrease in the astrocytic cellular systems for glutamate reuptake, and that behavioral effects may be greatly influenced by the corresponding disturbances in neuronal excitability. This involvement of glutamate reuptake dysfunction in the depression-like behavioral effects of stress is supported also by the demonstration that administration of riluzole, a compound that stimulates the reuptake of glutamate by astrocytes, and also reduces its release, mitigates depression-like behaviors and prevents alterations of astrocyte metabolism and GFAP mRNA expression in rodent models of stress (Banar et al., 2010). Likewise, ceftriaxone, a beta-lactam antibiotic that favors glutamate uptake, results in antidepressant-like effects across models of stress in mice (Mineur et al., 2007).

Gap Junctions of Astrocytes in Stress and Depression

In animals under chronic stress and in humans with major depression) there are drastically low levels of astrocytic gap junction proteins connexin 43 (Cx43) and connexin 30 (Cx30) in the prefrontal cortex (PFC) as compared to controls (Ernst et al., 2011; Sun et al., 2012; Miguel-Hidalgo et al., 2014, 2018; Nagy et al., 2016). Likewise, some animal models of stress leading to depression-like behaviors have demonstrated a region-specific decrease in myelination markers after prolonged stress, partially reversible with antidepressant treatments or myelin-promoting agents (Liu et al., 2012, 2016). This is consistent with findings in the PFC of human subjects with depression, where there is reduced expression of myelin- and astrocyte-related mRNAs (Liu et al., 2012, 2016; Rajkowska et al., 2015; Miguel-Hidalgo et al., 2017, 2018). More recently, highly suggestive evidence of reduced gap junction coupling between astrocytes and oligodendrocytes has been also observed in the anterior cingulate cortex of subjects with depression (Tanti et al., 2019). Gap junctions formed by astrocyte connexons (made of Cx43 or Cx30 subunits) interacting with the corresponding connexons from oligodendrocytes (made of Cx47 or C32) (Giaume and McCarthy, 1996) have proven crucial for the maintenance of myelin, as mice with KO for different combinations of astrocyte and oligodendrocyte connexins demonstrate variable degrees of myelin vacuolization and degradation, and profound behavioral deficits (Lutz et al., 2009; Magnotti et al., 2011; Li et al., 2014; Basu and Sarma, 2018). Whether chronic stress or sustained high corticosteroids in the blood circulation or the brain results in deficient myelin formation or maintenance mediated by reduced astrocyte connexins is less well established, although recently published evidence from our laboratory, has provided the first proof-of-concept evidence that stress, or high corticosterone during myelination, results in concomitant myelin and Cx43/Cx30 alterations that involve lower immunoreactivity for myelin basic protein and those connexins *in vivo* (Miguel-Hidalgo et al., 2018) and *in vitro* (Miguel-Hidalgo et al., 2019), and decreased morphological indices of myelination *in vitro*. We also have observed that concurrent reductions of connexins and myelin proteins in cortical primary cell cultures *in vitro*

can both be prevented by the glucocorticoid receptor antagonist mifepristone (Miguel-Hidalgo et al., 2019).

Mechanisms by which glucocorticoids regulate the expression, turnover or function of Cx43-containing gap junctions have been partially characterized in cells from tissues outside of nervous system. In cells from these tissues corticosterone (CORT) causes Cx43 degradation through autophagy mechanisms (Gao et al., 2016) or the activation of the Akt/mTOR signaling pathway (Shen et al., 2016). Activation of GC receptors is involved in those mechanisms because antagonism with mifepristone blocks CORT effects. In addition, some *in vitro* studies with primary cortical astrocytes, and *in vivo* research in the cerebral cortex, have found that high CORT levels cause reduction in Cx43 protein (Xia et al., 2017, 2018) or mRNA (Samarasinghe et al., 2012; Carter et al., 2013) and that inactivation of p38 and c-fos/API pathways (Morioka et al., 2014) may also lead to reduced Cx43. Also *in vivo*, some antidepressants, or GC receptor antagonists such as mifepristone mitigate the reduction of Cx43 caused by chronic unpredictable stress in animals (Sun et al., 2012; Xia et al., 2017). However, the mechanisms by which glucocorticoid receptor activation leads to Cx43 reductions in the CNS, and particularly in the PFC in stress or in postmortem brains from subjects with depression, remain to be ascertained. Neurodevelopmental research in prenatal neural progenitors has also indicated that the inactivation and reduction Cx43 gap junctions can be mediated by non-genomic actions of caveolin-1-attached GC receptors through the activation of the Raf-RAS-ERK1/2 pathway (Pedram et al., 2006; Hammes and Levin, 2007; Samarasinghe et al., 2012), although other effects on astrocytes may depend on the genomic pathway of GR activation (Kumar et al., 1986). Furthermore caveolin-1 is strongly expressed in astrocytes (Yun et al., 2011). Excess CORT-related decreases of brain-derived neurotrophic factor (BDNF) may also contribute to the reduction of Cx43, as shown in hippocampal astrocytes although this mechanism might not operate in the PFC (Schaaf et al., 1998). Thus, increased autophagy, AKT/mTOR activation, ERK-activation/Cx43-reduction by non-genomic GR mechanisms, or excess GC-induced BDNF decrease may also lead to alterations in the structure and function of astrocyte gap junctions. However, it is unknown whether and which of those mechanisms operates in the effects of stress (or high CORT) on astrocyte connexins or their companion oligodendrocyte connexins in the PFC. Interestingly, in our recent *in vitro* studies of mixed rat cortical primary cell cultures, we observed that reduction of Cx43 with a high dose of CORT analog DEX, which has a very high effectivity for genomic GR actions, was noticeably smaller than with high CORT (Miguel-Hidalgo et al., 2019), suggesting that, besides the genome-acting GC receptors, reductions of Cx43 with stress may be substantially due to non-genomic effects of GR activation.

An important contribution of astrocyte Cx43 and Cx30 to the maintenance of myelin integrity has been established in rodent models with KO of Cx43 and Cx30 expression alone or in combination with their KO of Cx47 and Cx32 oligodendrocyte partners, where the KO leads in many instances to patent myelin disorganization (Lutz et al., 2009; Magnotti et al., 2011; Tress et al., 2012; Li et al., 2014). However, it is yet unknown whether

connexin decreases caused by high glucocorticoids or stress is part of mechanisms that would lead to reduced expression of myelin proteins or to anomalous myelination in the PFC and its connections. Clearly, the magnitude and extent of Cx43 and myelin changes detected after stress, though significant, are smaller than in connexin KO animals. However, during the last decade it has become clear that plasticity of myelin leading to significant, but not necessarily catastrophic, changes in the morphology or extent of myelin results in meaningful variations in function and connectivity (Nave, 2010; Nave and Ehrenreich, 2014; Fields, 2015; Fields et al., 2015).

Involvement of Astrocytes in Neuroimmune Activation Elicited by Stress-Responses

Depression and stress responses are closely accompanied by dysregulation of immune activation and by expression of neuroimmune mediators produced by microglia and astrocytes (Hodes et al., 2015; Miller and Raison, 2016). Astrocytes carry receptors for various cytokines and chemokines and can thus respond to some of those mediators released by surrounding microglial cells or provided in the blood circulation (Maeng and Hong, 2019). Some of these responses involve chemokine expression that would in turn increase the ability to attract peripheral immune cells to the brain (Hennessy et al., 2015). Importantly, the actions of immune mediators on astrocytes directly influence their ability to perform essential functions (Sofroniew, 2014) in the support of neuronal activity, such as neurotransmitter metabolism (Muller and Schwarz, 2007), glutamate transport (Haroon et al., 2017, 2018; Piao et al., 2019), expression of aquaporins, maintenance of gap junctions (John et al., 1999; Karpuk et al., 2011) or expression of connexin 43 in a nuclear factor- κ B (NF- κ B)-dependent manner (Zhao et al., 2006). Conversely, connexin 43, which is present in astrocyte gap junctions interacting with the blood brain barrier (BBB), works as a main factor in regulating immune cell recruitment and antigen presentation at the BBB (Boulay et al., 2015). Thus, the sensitivity of astrocytes to inflammatory and other immune system-derived factors and the feedback they may exert to further regulate immune responses may be major contributors to neuronal metabolism and connectivity deficits that are thought to underpin the emotional and behavioral pathology caused by stress and other depressogenic neural mechanisms.

The intracellular molecular pathways involved in stress-induced activation of immune responses by astrocytes may involve the regulation of NF- κ B by menin, because in non-stressed conditions this protein keeps in check the content of nervous system pro-inflammatory transcriptional activator NF- κ B and the consequent downstream stimulation of interleukin 1 β (IL-1 β), while chronic stress results in menin reduction and subsequent activation of NF- κ B and IL-1 β increase. Depression-like behaviors in the same animal model were prevented by the use of NF- κ B inhibitors or IL1 β antagonists (Leng et al., 2018; Zhuang et al., 2018). Likewise, a polymorphism of gene *MEN1* (which encodes menin) was found to be significantly linked to higher risk for the onset of depression (Leng et al., 2018). In

fact, the polymorphism reduces the binding of menin to NF- κ B component p65, effectively enhancing the activation of Nf- κ B and thus the production of IL1 β by astrocytes.

Astrocyte Thrombospondin in Depression and Stress-Related Synaptic Alterations

Release of extracellular matrix proteins thrombospondin-1 and -2 by astrocytes is a strong promoter of synapse formation *in vitro* and *in vivo* (Christopherson et al., 2005; Jayakumar et al., 2014). Since changes of synaptic plasticity are involved in the pathophysiology of stress responses, depression, and other psychiatric disorders (Duman and Aghajanian, 2012; Yuen et al., 2012; Treccani et al., 2014; Csabai et al., 2018), some researchers have targeted the putative involvement of astrocyte-expressed thrombospondins 1 (TSP1) and 2 in the consequences of stress, including the generation of depression-like behaviors, as well as in the synaptic effects of antidepressant treatments. For instance, TSP-1 synthesis and mRNA expression are increased in the hippocampus after repeated electroconvulsive therapy, one of the most effective antidepressant treatments, although pharmacological antidepressants are apparently ineffective (Okada-Tsuchioka et al., 2014, 2017). In human subjects, plasma levels of TSP-1 levels were also found to be decreased in depression, at least in females (Okada-Tsuchioka et al., 2020).

Astrocyte Processes and Aquaporin at the Blood Brain Barrier in Stress-Related Neuropathology

In the prefrontal cortex of human subjects with depression, the coverage of vessels by the aquaporin 4 (AQP-4)-immunoreactive endfeet of astrocytes is significantly reduced, even if the extent of those feet is not changed when labeled for the cytoskeletal protein GFAP (Rajkowska et al., 2013). Interestingly, in a genetic rat model for depression-like behaviors (Arauchi et al., 2018), AQP-4-positive astrocyte endfeet were also found to be reduced when compared with Wistar rats. Remarkably, electroconvulsive therapy was able to suppress depression-like behaviors and increased the extent of AQP-4 immunoreactive processes around blood vessels in the rat genetic model (Azis et al., 2019). In fact, the functions of astrocytes' processes abutting at the basal lamina of arterioles are not limited to the control of BBB exchanges, but also include the regulation of the neurovascular coupling that depends on arteriole smooth muscle responses to extracellular potassium ions (Longden et al., 2014). This regulation may be significantly affected by elevated glucocorticoids in stress because corticoids, besides affecting smooth muscle potassium channels, would also alter coverage of arterioles by astrocytes processes, thus adding another risk factor for astrocyte-induced blood flow dysregulation upon the stress response.

Chronic stress can also cause astrocyte mediated alterations of cerebrovascular regulation through increases in oxidative damage elicited by neuroinflammation and oxidative stress. These processes are increased in the brain of subjects with depression, involving decreased levels of antioxidants and

higher levels of products of lipid peroxidation or, in serum, accompanied by augmented amounts of malondialdehyde, decreased antioxidant enzymes such as glutathione peroxidase and catalase, and increased superoxide and nNOS (Burrage et al., 2018). Decreases in eNOS in astrocytes and endothelial cells after stress may displace the equilibrium between oxidative and antioxidant processes to favor excess generation of deleterious superoxide.

Astrocyte Epigenetic Markers in Stress and Depression

As presented above, there are major effects of stress and probably other depressogenic mechanisms on the expression of astrocytes' connexins 43 and 30 in brain cortical areas (Sun et al., 2012; Miguel-Hidalgo et al., 2014, 2018, 2019). These reductions also affect their corresponding mRNAs both in animal models of stress (Sun et al., 2012) and in humans subjects with depression (Nagy et al., 2016) and have been convincingly attributed to epigenetic marker alterations such as methylation of histone 3 at Cx43 and Cx30 genes in the prefrontal cortex using chromatin immunoprecipitation (Nagy et al., 2016). The epigenetic differences between subjects with depression and controls also extend to the pattern of DNA methylation for genes expressed at a high level in astrocytes as compared to other cell types (Nagy et al., 2015) including *GFAP*, *ALDH1L1*, *SLC1A3*, *GJA1*, *GJB6*, *GLUL* and *SOX9*, all of which are found to be reduced in the dorsolateral prefrontal cortex of subjects with depression (Nagy et al., 2015). Depression-linked histone methylation modifications in astrocytes also have been detected for genes involved in the signaling of trophic factors such as *BDNF*, which is altered in some subjects with depression and in stress models. The levels of a BDNF receptor variant expressed in astrocytes, TrkB.T1, were found to be decreased in the prefrontal cortex of depressed suicide completers and inversely correlated with the levels of histone 3 methylation at lysine 27 (Ernst et al., 2009a,b). It has been suggested that vulnerability to the effects of stress, at least of a social nature, could be related to the expression of the TrkB.T1 variant in astrocytes, because mice with higher expression levels of that variant express avoidance behaviors to a higher extent than control mice, behavior that would be related to the increased probability of epigenetic modifications at TrkB.T1 in astrocytes (Razzoli et al., 2011; Hodes et al., 2017).

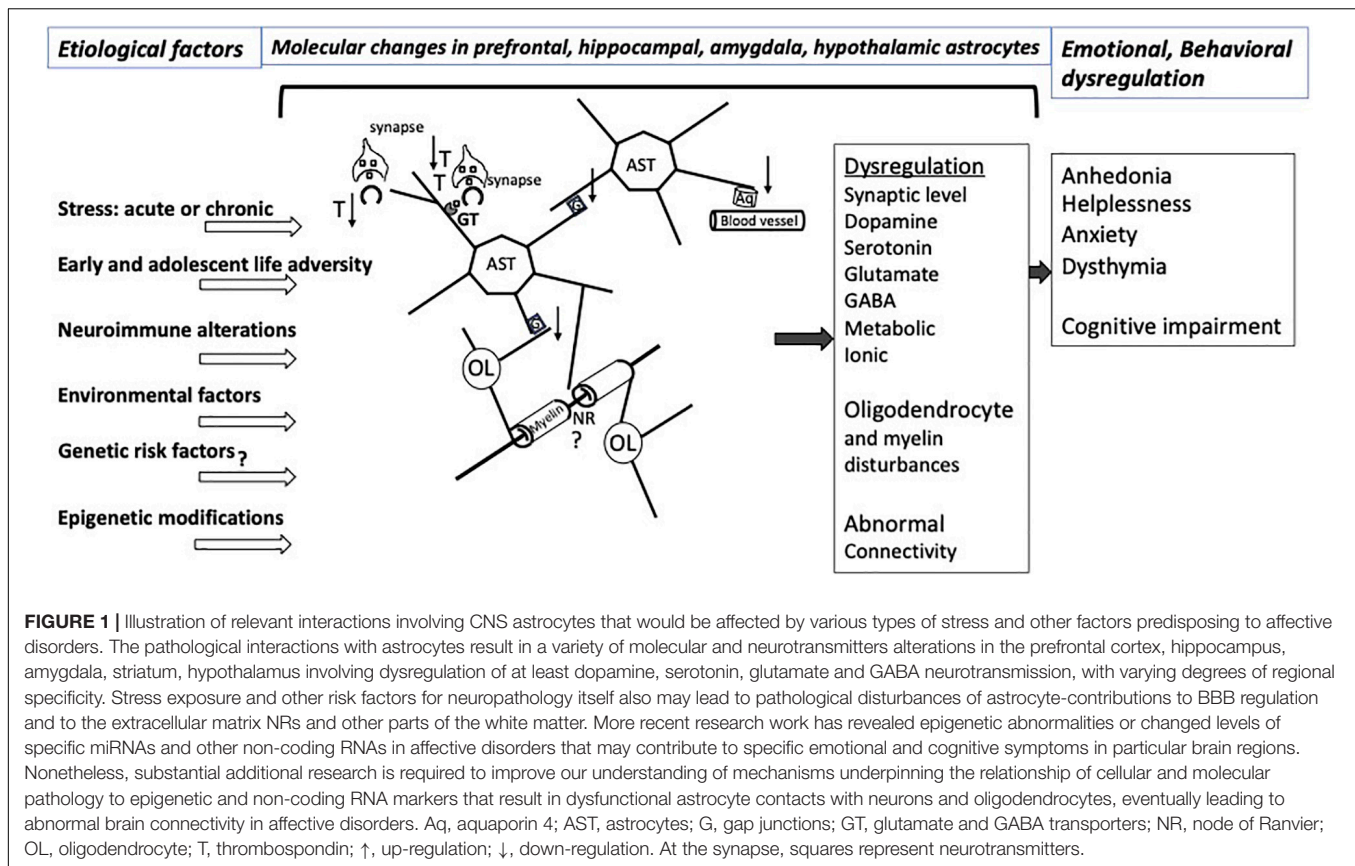
Further research into the regulation of astrocyte glutamate uptake has also indicated that epigenetic alterations of glutamate transporter genes *EAAT1* and *EAAT2* (*GLAST* and *GLT-1* respectively in rats) may confer vulnerability to the effects of stress or the manifestation of depression.

Specificity of epigenetic changes in astrocytes as compared to changes in neurons is underscored by astrocyte-specific higher or lower expression of some of the enzymes that are in charge of executing epigenetic modifications, astrocytes featuring higher mRNA expression of histone deacetylase Hdac8, sirtuin Sirt8, histone methyl-transferase Mll3 or histone demethylase Kdm5d, while underexpressing histone deacetylase Hdac5, Hdac11 or histone demethylase Kdm2b (Zhang et al., 2014; Jakovcevski et al., 2016). This capability of astrocytes to differentially

contribute to epigenetic changes may be of great relevance to the vulnerability to stress-related disorders and depression since disturbances in the expression of epigenetic enzymes and in the epigenetic changes themselves have been found to distinguish specific brain regions of subjects with depression as compared to non-psychiatric human subjects (Lopizzo et al., 2015; Jakovcevski et al., 2016). Other studies have pointed to factors involved in regulating the transcription of glutamate transporters such as the astrocyte-elevated gene which acts as corepressor of factor YY1 to repress the transcription of *GLT-1* and diminish glutamate uptake by astrocytes. YY1 also mediates repression of *GLAST* and *GLT-1* expression after exposure to inflammatory cytokine tumor necrosis factor α (TNF- α) by recruiting histone deacetylases to the promoter of glutamate transporter genes (Pajarillo et al., 2019). Some of the changes in epigenetic regulation of glutamate transporter expression may also be involved in depression-like behaviors and cognitive alterations secondary to neurological conditions, such as subarachnoid hemorrhage, in which astrocyte histone deacetylase 2 negatively regulates the expression of *GLT-1* (Tao et al., 2020).

Effects of Early Stress Exposure on Astrocytes and Their Role in the Development of Depression

Paramount among the studies addressing the vulnerability to depressive and anxiety symptoms are those focusing on the effects of early life adversity and stress on the probability of developing depressive and other psychiatric illnesses during and adolescence and adulthood (Chen and Baram, 2016). Various of these studies have been based on different forms of early adversity, including maternal separation/deprivation, malnutrition, overnutrition or immune challenges. A recent review has stressed the importance of early adversity in the responses of astrocytes to immune challenges later in life, as well the effects of disturbed astrocyte metabolism due to malnutrition, which could set the stage for astrocyte priming as a factor for the level of vulnerability to affective and other psychiatric disorders (Abbink et al., 2019). In the hippocampus, the multifaceted physiological roles of astrocytes and their involvement in functional plasticity such as long-term potentiation have been recently reviewed by Çalişkan et al. (2020), also indicating that early or juvenile stress results in reduced GFAP expression and diminished complexity of morphological features in astrocytes as well as reduced uptake of glutamate and GABA, and impaired conversion of glutamate to glutamine (Çalişkan et al., 2020). Repeated or chronic maternal separation in infant rats during the lactating period was also found to cause drastic reductions in the number of GFAP-immunoreactive astrocytes in the prefrontal cortex, hippocampus and striatum (Banqueri et al., 2019). For instance, maternal separation stress results in depletion of GFAP and S100b immunoreactive astrocytes in the medial precentral cortex of the Wistar rat strain (Musholt et al., 2009). Similarly, in Fischer rats, which are constitutively highly responsive to stress, 14 days of early maternal deprivation leads to reduced density of GFAP-immunoreactive astrocytes in the prefrontal cortex, hippocampal regions and the amygdala (Leventopoulos et al., 2007) in adult



animals. In guinea pigs, antenatal stress of dams results as well in reduced GFAP immunostaining of CA1 hippocampal astrocytes when examined in 21-days old offspring (Bennett et al., 2015). In view of the lower immunoreactivity of GFAP immunoreactive astrocytes and the lower levels of GFAP observed in the prefrontal cortex in postmortem human brains from younger subjects with major depression (as compared to older subjects or to non-psychiatric controls) (Miguel-Hidalgo et al., 2000; Si et al., 2004), early stress, adversity or excessive immune activation may prime astrocytes to facilitate, at least in some human subjects, manifestations of depression in late-adolescence and adulthood.

Since early life stress (ELS) also involves the activation of hormonal responses regulated by the hypothalamic-pituitary-adrenal axis, some research has addressed cellular and neurotransmitter dysfunction in the hypothalamus and a putative involvement of astrocytes. In mature hypothalamic neurons, the ability of neurosteroids to reduce overactivation was found to be curtailed by ELS. This effect is dependent on an excessive glutamatergic activation that derives in part from impaired astrocytic regulation of extracellular glutamate levels (Gunn et al., 2013). This alteration was concomitant with morphological changes of GFAP-immunoreactive astrocyte processes and a reduction of the immunoreactivity of astrocytic glutamate transporters (Gunn et al., 2013).

Long-lasting astrocyte glutamate transport deficits may lead to dysfunction of specific circuits known to be involved in

the pathophysiology of affective and psychiatric disorders. For instance, in the medial prefrontal cortex of rodents, astrocytic glutamate transport activity plays a major role in modulating the connections between the prefrontal cortex and the serotonergic neurons of the raphe nucleus (Fullana et al., 2020). Accordingly, prolonged suppression of the astrocyte-dependent glutamate transport capacity in the prefrontal cortex of mice results in depression-like behaviors (Fullana et al., 2019a,b), although the effects of acute vs chronic stress on the connections between the various subdivisions of the PFC and the serotonergic raphe neurons remain to be fully elucidated (Fullana et al., 2020).

CONCLUSION

Astrocytes' multifaceted involvement in various aspects of neuronal metabolism, support of neurotransmission, supply of neurotrophic factors, neuroimmune reactivity, myelin maintenance, and regulation of signal propagation as well as their capacity to directly respond to hormonal and other molecular adaptations caused by stress sets them to be major players in determining the behavioral and emotional vulnerability to the effects of stress (Figure 1). These effects may be long lasting because at different stages of development and in adulthood there is substantial epigenetic regulation of gene expression in astrocytes, and the astrocytes themselves are crucial

in supporting the development of other neural cells, synaptic connections, and axonal pathways in the central nervous system. Some of the epigenetic and molecular mechanisms regarding glutamate transport, gap junction communication and neuroimmune regulation that are related to stress effects are starting to be ascertained. However, much remains to be learned to understand the specific role of those and other modifications in the astroglial contribution to the vulnerability to and maintenance of stress-related disorders and depression, and for leveraging that knowledge to achieve more effective psychiatric therapies.

REFERENCES

Abbink, M. R., Van Deijk, A. F., Heine, V. M., Verheijen, M. H., and Korosi, A. (2019). The involvement of astrocytes in early-life adversity induced programming of the brain. *Glia* 67, 1637–1653. doi: 10.1002/glia.23625

Almeida, R. F., Thomazi, A. P., Godinho, G. F., Saute, J. A., Wofchuk, S. T., Souza, D. O., et al. (2010). Effects of depressive-like behavior of rats on brain glutamate uptake. *Neurochem. Res.* 35, 1164–1171. doi: 10.1007/s11064-010-0169-4

Arauchi, R., Hashioka, S., Tsuchie, K., Miyaoka, T., Tsumori, T., Limoa, E., et al. (2018). Gunn rats with glial activation in the hippocampus show prolonged immobility time in the forced swimming test and tail suspension test. *Brain Behav.* 8:e01028. doi: 10.1002/brb3.1028

Araya-Callis, C., Hiemke, C., Abumaria, N., and Flugge, G. (2012). Chronic psychosocial stress and citalopram modulate the expression of the glial proteins GFAP and NDRG2 in the hippocampus. *Psychopharmacology (Berl.)* 224, 209–222. doi: 10.1007/s00213-012-2741-x

Azis, I. A., Hashioka, S., Tsuchie, K., Miyaoka, T., Abdullah, R. A., Limoa, E., et al. (2019). Electroconvulsive shock restores the decreased coverage of brain blood vessels by astrocytic endfeet and ameliorates depressive-like behavior. *J. Affect. Disord.* 257, 331–339. doi: 10.1016/j.jad.2019.07.008

Banasr, M., Chowdhury, G. M., Terwilliger, R., Newton, S. S., Duman, R. S., Behar, K. L., et al. (2010). Glial pathology in an animal model of depression: reversal of stress-induced cellular, metabolic and behavioral deficits by the glutamate-modulating drug riluzole. *Mol. Psychiatry* 15, 501–511. doi: 10.1038/mp.2008.106

Banasr, M., and Duman, R. S. (2008). Glial loss in the prefrontal cortex is sufficient to induce depressive-like behaviors. *Biol. Psychiatry* 64, 863–870. doi: 10.1016/j.biopsych.2008.06.008

Banqueri, M., Mendez, M., Gomez-Lazaro, E., and Arias, J. L. (2019). Early life stress by repeated maternal separation induces long-term neuroinflammatory response in glial cells of male rats. *Stress* 22, 563–570. doi: 10.1080/10253890.2019.1604666

Basu, R., and Sarma, J. D. (2018). Connexin 43/47 channels are important for astrocyte/ oligodendrocyte cross-talk in myelination and demyelination. *J. Biosci.* 43, 1055–1068. doi: 10.1007/s12038-018-9811-0

Bender, C. L., Calfa, G. D., and Molina, V. A. (2016). Astrocyte plasticity induced by emotional stress: a new partner in psychiatric physiopathology? *Prog. Neuropsychopharmacol. Biol. Psychiatry* 65, 68–77. doi: 10.1016/j.pnpbp.2015.08.005

Bennett, G. A., Palliser, H. K., Shaw, J. C., Walker, D., and Hirst, J. J. (2015). Prenatal stress alters hippocampal neuroglia and increases anxiety in childhood. *Dev. Neurosci.* 37, 533–545. doi: 10.1159/000437302

Boulay, A. C., Mazeraud, A., Cisternino, S., Saubamea, B., Maily, P., Jourden, L., et al. (2015). Immune quiescence of the brain is set by astroglial connexin 43. *J. Neurosci.* 35, 4427–4439. doi: 10.1523/JNEUROSCI.2575-14.2015

Burrage, E., Marshall, K. L., Santanam, N., and Chantler, P. D. (2018). Cerebrovascular dysfunction with stress and depression. *Brain Circ.* 4, 43–53. doi: 10.4103/bc.bc_6_18

Çalışkan, G., Muller, A., and Albrecht, A. (2020). Long-term impact of early-life stress on hippocampal plasticity: spotlight on astrocytes. *Int. J. Mol. Sci.* 21:4999. doi: 10.3390/ijms21144999

AUTHOR CONTRIBUTIONS

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Carter, B. S., Hamilton, D. E., and Thompson, R. C. (2013). Acute and chronic glucocorticoid treatments regulate astrocyte-enriched mRNAs in multiple brain regions in vivo. *Front. Neurosci.* 7:139. doi: 10.3389/fnins.2013.00139

Chandleley, M. J., Szebeni, A., Szebeni, K., Crawford, J. D., Stockmeier, C. A., Turecki, G., et al. (2014). Elevated gene expression of glutamate receptors in noradrenergic neurons from the locus coeruleus in major depression. *Int. J. Neuropsychopharmacol.* 17, 1569–1578. doi: 10.1017/S1461145714000662

Chen, Y., and Baram, T. Z. (2016). Toward understanding how early-life stress reprograms cognitive and emotional brain networks. *Neuropsychopharmacology* 41, 197–206. doi: 10.1038/npp.2015.181

Choudary, P. V., Molnar, M., Evans, S. J., Tomita, H., Li, J. Z., Vawter, M. P., et al. (2005). Altered cortical glutamatergic and GABAergic signal transmission with glial involvement in depression. *Proc. Natl. Acad. Sci. U.S.A.* 102, 15653–15658. doi: 10.1073/pnas.0507901102

Christopherson, K. S., Ullian, E. M., Stokes, C. C., Mullowney, C. E., Hell, J. W., Agah, A., et al. (2005). Thrombospondins are astrocyte-secreted proteins that promote CNS synaptogenesis. *Cell* 120, 421–433. doi: 10.1016/j.cell.2004.12.020

Codeluppi, S. A., Chatterjee, D., Prevot, T. D., Bansal, Y., Misquitta, K. A., Sibille, E., et al. (2021). Chronic stress alters astrocyte morphology in mouse prefrontal cortex. *Int. J. Neuropsychopharmacol.* 24, 842–853. doi: 10.1093/ijnp/pyab052

Csabai, D., Wiborg, O., and Czeh, B. (2018). Reduced synapse and axon numbers in the prefrontal cortex of rats subjected to a chronic stress model for depression. *Front. Cell. Neurosci.* 12:24. doi: 10.3389/fncel.2018.00024

Cui, W., Mizukami, H., Yanagisawa, M., Aida, T., Nomura, M., Isomura, Y., et al. (2014). Glial dysfunction in the mouse habenula causes depressive-like behaviors and sleep disturbance. *J. Neurosci.* 34, 16273–16285. doi: 10.1523/JNEUROSCI.1465-14.2014

Czeh, B., Simon, M., Schmeling, B., Hiemke, C., and Fuchs, E. (2006). Astroglial plasticity in the hippocampus is affected by chronic psychosocial stress and concomitant fluoxetine treatment. *Neuropsychopharmacology* 31, 1616–1626. doi: 10.1038/sj.npp.1300982

de Vasconcellos-Bittencourt, A. P., Vendite, D. A., Nassif, M., Crema, L. M., Frozza, R., Thomazi, A. P., et al. (2011). Chronic stress and lithium treatments alter hippocampal glutamate uptake and release in the rat and potentiate necrotic cellular death after oxygen and glucose deprivation. *Neurochem. Res.* 36, 793–800. doi: 10.1007/s11064-011-0404-7

Duman, R. S., and Aghajanian, G. K. (2012). Synaptic dysfunction in depression: potential therapeutic targets. *Science* 338, 68–72. doi: 10.1126/science.1222939

Ernst, C., Chen, E. S., and Turecki, G. (2009a). Histone methylation and decreased expression of TrkB.T1 in orbital frontal cortex of suicide completers. *Mol. Psychiatry* 14, 830–832. doi: 10.1038/mp.2009.35

Ernst, C., Deleva, V., Deng, X., Sequeira, A., Pomarenski, A., Klempan, T., et al. (2009b). Alternative splicing, methylation state, and expression profile of tropomyosin-related kinase B in the frontal cortex of suicide completers. *Arch. Gen. Psychiatry* 66, 22–32. doi: 10.1001/archpsyc.66.1.22

Ernst, C., Nagy, C., Kim, S., Yang, J. P., Deng, X., Hellstrom, I. C., et al. (2011). Dysfunction of astrocyte connexins 30 and 43 in dorsal lateral prefrontal cortex of suicide completers. *Biol. Psychiatry* 70, 312–319. doi: 10.1016/j.biopsych.2011.03.038

Feyissa, A. M., Woolverton, W. L., Miguel-Hidalgo, J. J., Wang, Z., Kyle, P. B., Hasler, G., et al. (2010). Elevated level of metabotropic glutamate receptor 2/3

- in the prefrontal cortex in major depression. *Prog. Neuropsychopharmacol. Biol. Psychiatry* 34, 279–283. doi: 10.1016/j.pnpbp.2009.11.018
- Fields, R. D. (2015). A new mechanism of nervous system plasticity: activity-dependent myelination. *Nat. Rev. Neurosci.* 16, 756–767. doi: 10.1038/nrn4023
- Fields, R. D., Woo, D. H., and Basser, P. J. (2015). Glial regulation of the neuronal connectome through local and long-distant communication. *Neuron* 86, 374–386. doi: 10.1016/j.neuron.2015.01.014
- Franklin, T. B., Saab, B. J., and Mansuy, I. M. (2012). Neural mechanisms of stress resilience and vulnerability. *Neuron* 75, 747–761. doi: 10.1016/j.neuron.2012.08.016
- Fullana, M. N., Covelo, A., Bortolozzi, A., Araque, A., and Artigas, F. (2019a). In vivo knockdown of astroglial glutamate transporters GLT-1 and GLAST increases excitatory neurotransmission in mouse infralimbic cortex: relevance for depressive-like phenotypes. *Eur. Neuropsychopharmacol.* 29, 1288–1294. doi: 10.1016/j.euroneuro.2019.09.004
- Fullana, M. N., Ruiz-Bronchal, E., Ferrer-Coy, A., Juárez-Escoto, E., Artigas, F., and Bortolozzi, A. (2019b). Regionally selective knockdown of astroglial glutamate transporters in infralimbic cortex induces a depressive phenotype in mice. *Glia* 67, 1122–1137. doi: 10.1002/glia.23593
- Fullana, N., Gasull-Camos, J., Tarres-Gatius, M., Castane, A., Bortolozzi, A., and Artigas, F. (2020). Astrocyte control of glutamatergic activity: downstream effects on serotonergic function and emotional behavior. *Neuropharmacology* 166:107914. doi: 10.1016/j.neuropharm.2019.107914
- Gao, J., Cheng, T. S., Qin, A., Pavlos, N. J., Wang, T., Song, K., et al. (2016). Glucocorticoid impairs cell-cell communication by autophagy-mediated degradation of connexin 43 in osteocytes. *Oncotarget* 7, 26966–26978. doi: 10.18632/oncotarget.9034
- Giaume, C., and McCarthy, K. D. (1996). Control of gap-junctional communication in astrocytic networks. *Trends Neurosci.* 19, 319–325. doi: 10.1016/0166-2236(96)10046-1
- Gomez-Galan, M., De Bundel, D., Van Eeckhaut, A., Smolders, I., and Lindskog, M. (2013). Dysfunctional astrocytic regulation of glutamate transmission in a rat model of depression. *Mol. Psychiatry* 18, 582–594. doi: 10.1038/mp.2012.10
- Gosselin, R. D., Gibney, S., O'malley, D., Dinan, T. G., and Cryan, J. F. (2009). Region specific decrease in glial fibrillary acidic protein immunoreactivity in the brain of a rat model of depression. *Neuroscience* 159, 915–925. doi: 10.1016/j.neuroscience.2008.10.018
- Gunn, B. G., Cunningham, L., Cooper, M. A., Corteen, N. L., Seifi, M., Swinny, J. D., et al. (2013). Dysfunctional astrocytic and synaptic regulation of hypothalamic glutamatergic transmission in a mouse model of early-life adversity: relevance to neurosteroids and programming of the stress response. *J. Neurosci.* 33, 19534–19554. doi: 10.1523/JNEUROSCI.1337-13.2013
- Had-Aissouni, L., Re, D. B., Nieoullon, A., and Kerkerian-Le Goff, L. (2002). Importance of astrocytic inactivation of synaptically released glutamate for cell survival in the central nervous system—are astrocytes vulnerable to low intracellular glutamate concentrations? *J. Physiol. Paris* 96, 317–322. doi: 10.1016/s0928-4257(02)00022-0
- Hammes, S. R., and Levin, E. R. (2007). Extracellular steroid receptors: nature and actions. *Endocr. Rev.* 28, 726–741. doi: 10.1210/er.2007-0022
- Haroon, E., Chen, X., Li, Z., Patel, T., Woolwine, B. J., Hu, X. P., et al. (2018). Increased inflammation and brain glutamate define a subtype of depression with decreased regional homogeneity, impaired network integrity, and anhedonia. *Transl. Psychiatry* 8:189. doi: 10.1038/s41398-018-0241-4
- Haroon, E., Miller, A. H., and Sanacora, G. (2017). Inflammation, glutamate, and glia: a trio of trouble in mood disorders. *Neuropsychopharmacology* 42, 193–215. doi: 10.1038/npp.2016.199
- Hennessy, E., Griffin, E. W., and Cunningham, C. (2015). Astrocytes are primed by chronic neurodegeneration to produce exaggerated chemokine and cell infiltration responses to acute stimulation with the cytokines IL-1beta and TNF-alpha. *J. Neurosci.* 35, 8411–8422. doi: 10.1523/JNEUROSCI.2745-14.2015
- Hodes, G. E., Kana, V., Menard, C., Merad, M., and Russo, S. J. (2015). Neuroimmune mechanisms of depression. *Nat. Neurosci.* 18, 1386–1393. doi: 10.1038/nn.4113
- Hodes, G. E., Walker, D. M., Labonte, B., Nestler, E. J., and Russo, S. J. (2017). Understanding the epigenetic basis of sex differences in depression. *J. Neurosci. Res.* 95, 692–702. doi: 10.1002/jnr.23876
- Imbe, H., Kimura, A., Donishi, T., and Kaneoke, Y. (2012). Chronic restraint stress decreases glial fibrillary acidic protein and glutamate transporter in the periaqueductal gray matter. *Neuroscience* 223, 209–218. doi: 10.1016/j.neuroscience.2012.08.007
- Imbe, H., Kimura, A., Donishi, T., and Kaneoke, Y. (2013). Effects of restraint stress on glial activity in the rostral ventromedial medulla. *Neuroscience* 241, 10–21. doi: 10.1016/j.neuroscience.2013.03.008
- Jakovcevski, M., Akbarian, S., and Di Benedetto, B. (2016). Pharmacological modulation of astrocytes and the role of cell type-specific histone modifications for the treatment of mood disorders. *Curr. Opin. Pharmacol.* 26, 61–66. doi: 10.1016/j.coph.2015.10.002
- Jayakumar, A. R., Tong, X. Y., Curtis, K. M., Ruiz-Cordero, R., Shamaladevi, N., Abuzamel, M., et al. (2014). Decreased astrocytic thrombospondin-1 secretion after chronic ammonia treatment reduces the level of synaptic proteins: in vitro and in vivo studies. *J. Neurochem.* 131, 333–347. doi: 10.1111/jnc.12810
- John, G. R., Scemes, E., Suadicani, S. O., Liu, J. S., Charles, P. C., Lee, S. C., et al. (1999). IL-1beta differentially regulates calcium wave propagation between primary human fetal astrocytes via pathways involving P2 receptors and gap junction channels. *Proc. Natl. Acad. Sci. U.S.A.* 96, 11613–11618. doi: 10.1073/pnas.96.20.11613
- Karolewicz, B., Maciag, D., O'dwyer, G., Stockmeier, C. A., Feyissa, A. M., and Rajkowska, G. (2010). Reduced level of glutamic acid decarboxylase-67 kDa in the prefrontal cortex in major depression. *Int. J. Neuropsychopharmacol.* 13, 411–420. doi: 10.1017/S1461145709990587
- Karpuk, N., Burkovetskaya, M., Fritz, T., Angle, A., and Kielian, T. (2011). Neuroinflammation leads to region-dependent alterations in astrocyte gap junction communication and hemichannel activity. *J. Neurosci.* 31, 414–425. doi: 10.1523/JNEUROSCI.5247-10.2011
- Kendell, S. F., Krystal, J. H., and Sanacora, G. (2005). GABA and glutamate systems as therapeutic targets in depression and mood disorders. *Expert Opin. Ther. Targets* 9, 153–168. doi: 10.1517/14728222.9.1.153
- Kumar, S., Holmes, E., Scully, S., Birren, B. W., Wilson, R. H., and De Vellis, J. (1986). The hormonal regulation of gene expression of glial markers: glutamine synthetase and glycerol phosphate dehydrogenase in primary cultures of rat brain and in C6 cell line. *J. Neurosci. Res.* 16, 251–264. doi: 10.1002/jnr.490160122
- Leng, L., Zhuang, K., Liu, Z., Huang, C., Gao, Y., Chen, G., et al. (2018). Menin deficiency leads to depressive-like behaviors in mice by modulating astrocyte-mediated neuroinflammation. *Neuron* 100, 551–563.e7. doi: 10.1016/j.neuron.2018.08.031
- Leventopoulos, M., Ruedi-Bettschen, D., Knuesel, I., Feldon, J., Pryce, C. R., and Opacka-Juffry, J. (2007). Long-term effects of early life deprivation on brain glia in Fischer rats. *Brain Res.* 1142, 119–126. doi: 10.1016/j.brainres.2007.01.039
- Li, L. F., Yang, J., Ma, S. P., and Qu, R. (2013). Magnolol treatment reversed the glial pathology in an unpredictable chronic mild stress-induced rat model of depression. *Eur. J. Pharmacol.* 711, 42–49. doi: 10.1016/j.ejphar.2013.04.008
- Li, T., Giaume, C., and Xiao, L. (2014). Connexin-mediated glia networking impacts myelination and remyelination in the central nervous system. *Mol. Neurobiol.* 49, 1460–1471. doi: 10.1007/s12035-013-8625-1
- Li, Y. S., Hong, Y. F., He, J., Lin, J. X., Shan, Y. L., Fu, D. Y., et al. (2013). Effects of magnolol on impairment of learning and memory abilities induced by scopolamine in mice. *Biol. Pharm. Bull.* 36, 764–771. doi: 10.1248/bpb.12-00880
- Liu, J., Dietz, K., Deloyht, J. M., Pedre, X., Kelkar, D., Kaur, J., et al. (2012). Impaired adult myelination in the prefrontal cortex of socially isolated mice. *Nat. Neurosci.* 15, 1621–1623. doi: 10.1038/nn.3263
- Liu, J., Dupree, J. L., Gacias, M., Frawley, R., Sikder, T., Naik, P., et al. (2016). Clemastine enhances myelination in the prefrontal cortex and rescues behavioral changes in socially isolated mice. *J. Neurosci.* 36, 957–962. doi: 10.1523/JNEUROSCI.3608-15.2016
- Liu, Q., Li, B., Zhu, H. Y., Wang, Y. Q., Yu, J., and Wu, G. C. (2009). Clomipramine treatment reversed the glial pathology in a chronic unpredictable stress-induced rat model of depression. *Eur. Neuropsychopharmacol.* 19, 796–805. doi: 10.1016/j.euroneuro.2009.06.010
- Longden, T. A., Dabertrand, F., Hill-Eubanks, D. C., Hammack, S. E., and Nelson, M. T. (2014). Stress-induced glucocorticoid signaling remodels neurovascular coupling through impairment of cerebrovascular inwardly rectifying K+

- channel function. *Proc. Natl. Acad. Sci. U.S.A.* 111, 7462–7467. doi: 10.1073/pnas.1401811111
- Lopizzo, N., Bocchio Chiavetto, L., Cattane, N., Plazzotta, G., Tarazi, F. I., Pariante, C. M., et al. (2015). Gene-environment interaction in major depression: focus on experience-dependent biological systems. *Front. Psychiatry* 6:68. doi: 10.3389/fpsy.2015.00068
- Lu, C.-L., Ren, J., Mo, J.-W., Fan, J., Guo, F., Chen, L.-Y., et al. (2021). Glucocorticoid receptor-dependent astrocytes mediate stress vulnerability. *Biol. Psychiatry*. doi: 10.1016/j.biopsych.2021.11.022
- Lutz, S. E., Zhao, Y., Gulinello, M., Lee, S. C., Raine, C. S., and Brosnan, C. F. (2009). Deletion of astrocyte connexins 43 and 30 leads to a dysmyelinating phenotype and hippocampal CA1 vacuolation. *J. Neurosci.* 29, 7743–7752. doi: 10.1523/JNEUROSCI.0341-09.2009
- Maeng, S. H., and Hong, H. (2019). Inflammation as the potential basis in depression. *Int. Neurol.* J. 23, S63–S71. doi: 10.5213/inj.1938226.113
- Magnotti, L. M., Goodenough, D. A., and Paul, D. L. (2011). Deletion of oligodendrocyte Cx32 and astrocyte Cx43 causes white matter vacuolation, astrocyte loss and early mortality. *Glia* 59, 1064–1074. doi: 10.1002/glia.21179
- McKlveen, J. M., Morano, R. L., Fitzgerald, M., Zoubovsky, S., Cassella, S. N., Scheimann, J. R., et al. (2016). Chronic stress increases prefrontal inhibition: a mechanism for stress-induced prefrontal dysfunction. *Biol. Psychiatry* 80, 754–764. doi: 10.1016/j.biopsych.2016.03.2101
- Menard, C., Pfau, M. L., Hodes, G. E., and Russo, S. J. (2017). Immune and neuroendocrine mechanisms of stress vulnerability and resilience. *Neuropsychopharmacology* 42, 62–80. doi: 10.1038/npp.2016.90
- Miguel-Hidalgo, J. J., Baucom, C., Dilley, G., Overholser, J. C., Meltzer, H. Y., Stockmeier, C. A., et al. (2000). Glial fibrillary acidic protein immunoreactivity in the prefrontal cortex distinguishes younger from older adults in major depressive disorder. *Biol. Psychiatry* 48, 861–873. doi: 10.1016/s0006-3223(00)00999-9
- Miguel-Hidalgo, J. J., Carter, K., Deloach, P. H., Sanders, L., and Pang, Y. (2019). Glucocorticoid-induced reductions of myelination and connexin 43 in mixed central nervous system cell cultures are prevented by mifepristone. *Neuroscience* 411, 255–269. doi: 10.1016/j.neuroscience.2019.05.050
- Miguel-Hidalgo, J. J., Hall, K. O., Bonner, H., Roller, A. M., Syed, M., Park, C. J., et al. (2017). MicroRNA-21: expression in oligodendrocytes and correlation with low myelin mRNAs in depression and alcoholism. *Prog. Neuropsychopharmacol. Biol. Psychiatry* 79, 503–514. doi: 10.1016/j.pnpbp.2017.08.009
- Miguel-Hidalgo, J. J., Moulana, M., Deloach, P. H., and Rajkowska, G. (2018). Chronic unpredictable stress reduces immunostaining for connexins 43 and 30 and myelin basic protein in the rat prelimbic and orbitofrontal cortices. *Chronic Stress* 2, 1–12. doi: 10.1177/2470547018814186
- Miguel-Hidalgo, J. J., Waltzer, R., Whitton, A. A., Austin, M. C., Rajkowska, G., and Stockmeier, C. A. (2010). Glial and glutamatergic markers in depression, alcoholism, and their comorbidity. *J. Affect. Disord.* 127, 230–240. doi: 10.1016/j.jad.2010.06.003
- Miguel-Hidalgo, J. J., Wilson, B. A., Hussain, S., Meshram, A., Rajkowska, G., and Stockmeier, C. A. (2014). Reduced connexin 43 immunolabeling in the orbitofrontal cortex in alcohol dependence and depression. *J. Psychiatr. Res.* 55, 101–109. doi: 10.1016/j.jpsychires.2014.04.007
- Miller, A. H., and Raison, C. L. (2016). The role of inflammation in depression: from evolutionary imperative to modern treatment target. *Nat. Rev. Immunol.* 16, 22–34. doi: 10.1038/nri.2015.5
- Mineur, Y. S., Picciotto, M. R., and Sanacora, G. (2007). Antidepressant-like effects of ceftriaxone in male C57BL/6J mice. *Biol. Psychiatry* 61, 250–252. doi: 10.1016/j.biopsych.2006.04.037
- Morioka, N., Suekama, K., Zhang, F. F., Kajitani, N., Hisaoka-Nakashima, K., Takebayashi, M., et al. (2014). Amitriptyline up-regulates connexin43-gap junction in rat cultured cortical astrocytes via activation of the p38 and c-Fos/AP-1 signalling pathway. *Br. J. Pharmacol.* 171, 2854–2867. doi: 10.1111/bph.12614
- Muller, N., and Schwarz, M. J. (2007). The immune-mediated alteration of serotonin and glutamate: towards an integrated view of depression. *Mol. Psychiatry* 12, 988–1000. doi: 10.1038/sj.mp.4002006
- Murphy-Royal, C., Gordon, G. R., and Bains, J. S. (2019). Stress-induced structural and functional modifications of astrocytes-further implicating glia in the central response to stress. *Glia* 67, 1806–1820. doi: 10.1002/glia.23610
- Musholt, K., Cirillo, G., Cavaliere, C., Rosaria Bianco, M., Bock, J., Helmeke, C., et al. (2009). Neonatal separation stress reduces glial fibrillary acidic protein- and S100beta-immunoreactive astrocytes in the rat medial precentral cortex. *Dev. Neurobiol.* 69, 203–211. doi: 10.1002/dneu.20694
- Nagy, C., Suderman, M., Yang, J., Szyf, M., Mechawar, N., Ernst, C., et al. (2015). Astrocytic abnormalities and global DNA methylation patterns in depression and suicide. *Mol. Psychiatry* 20, 320–328. doi: 10.1038/mp.2014.21
- Nagy, C., Torres-Platas, S. G., Mechawar, N., and Turecki, G. (2016). Repression of astrocytic connexins in cortical and subcortical brain regions and prefrontal enrichment of H3K9me3 in depression and suicide. *Int. J. Neuropsychopharmacol.* 20, 50–57. doi: 10.1093/ijnp/pyw071
- Nave, K. A. (2010). Myelination and the trophic support of long axons. *Nat. Rev. Neurosci.* 11, 275–283. doi: 10.1038/nrn2797
- Nave, K. A., and Ehrenreich, H. (2014). Myelination and oligodendrocyte functions in psychiatric diseases. *JAMA Psychiatry* 71, 582–584. doi: 10.1001/jamapsychiatry.2014.189
- Okada-Tsuchioka, M., Omori, W., Kajitani, N., Shibasaki, C., Itagaki, K., and Takebayashi, M. (2020). Decreased serum levels of thrombospondin-1 in female depressed patients. *Neuropsychopharmacol. Rep.* 40, 39–45. doi: 10.1002/npr2.12088
- Okada-Tsuchioka, M., Segawa, M., Kajitani, N., Hisaoka-Nakashima, K., Shibasaki, C., Morinobu, S., et al. (2014). Electroconvulsive seizure induces thrombospondin-1 in the adult rat hippocampus. *Prog. Neuropsychopharmacol. Biol. Psychiatry* 48, 236–244. doi: 10.1016/j.pnpbp.2013.10.001
- Okada-Tsuchioka, M., Shibasaki, C., and Takebayashi, M. (2017). “How does electroconvulsive therapy work in the brain?–Involvement of the astrocyte-derived synaptogenic factor, Thrombospondin-1,” in *Advances in Modern Medicine*, Vol. 242, eds K. Taniyama and W. Kamiike (Sharjah: Bentham Science Publishers), 242–250. doi: 10.2174/9781681080239117010028
- Pajarillo, E., Rizor, A., Lee, J., Aschner, M., and Lee, E. (2019). The role of astrocytic glutamate transporters GLT-1 and GLAST in neurological disorders: potential targets for neurotherapeutics. *Neuropharmacology* 161:107559. doi: 10.1016/j.neuropharm.2019.03.002
- Pedram, A., Razandi, M., and Levin, E. R. (2006). Nature of functional estrogen receptors at the plasma membrane. *Mol. Endocrinol.* 20, 1996–2009. doi: 10.1210/me.2005-0525
- Piao, C. S., Holloway, A. L., Hong-Routson, S., and Wainwright, M. S. (2019). Depression following traumatic brain injury in mice is associated with down-regulation of hippocampal astrocyte glutamate transporters by thrombin. *J. Cereb. Blood Flow Metab.* 39, 58–73. doi: 10.1177/0271678X17742792
- Rajkowska, G., Hughes, J., Stockmeier, C. A., Javier Miguel-Hidalgo, J., and Maciag, D. (2013). Coverage of blood vessels by astrocytic endfeet is reduced in major depressive disorder. *Biol. Psychiatry* 73, 613–621. doi: 10.1016/j.biopsych.2012.09.024
- Rajkowska, G., Mahajan, G., Maciag, D., Sathyanesan, M., Iyo, A. H., Moulana, M., et al. (2015). Oligodendrocyte morphometry and expression of myelin-related mRNA in ventral prefrontal white matter in major depressive disorder. *J. Psychiatr. Res.* 65, 53–62. doi: 10.1016/j.jpsychires.2015.04.010
- Razzoli, M., Domenici, E., Carboni, L., Rantamaki, T., Lindholm, J., Castren, E., et al. (2011). A role for BDNF/TrkB signaling in behavioral and physiological consequences of social defeat stress. *Genes Brain Behav.* 10, 424–433. doi: 10.1111/j.1601-183X.2011.00681.x
- Réus, G. Z., Abelaira, H. M., Tuon, T., Titus, S. E., Ignácio, Z. M., Rodrigues, A. L. S., et al. (2016). “Chapter five – glutamatergic NMDA receptor as therapeutic target for depression,” in *Advances in Protein Chemistry and Structural Biology*, ed. R. Donev (Cambridge, MA: Academic Press), 169–202. doi: 10.1016/bs.apcsb.2015.10.003
- Samarasinghe, R. A., Witchell, S. F., and Defranco, D. B. (2012). Cooperativity and complementarity: synergies in non-classical and classical glucocorticoid signaling. *Cell Cycle* 11, 2819–2827. doi: 10.4161/cc.21018
- Schaaf, M. J., De Jong, J., De Kloet, E. R., and Vreugdenhil, E. (1998). Downregulation of BDNF mRNA and protein in the rat hippocampus by corticosterone. *Brain Res.* 813, 112–120. doi: 10.1016/s0006-8993(98)01010-5
- Shen, C., Kim, M. R., Noh, J. M., Kim, S. J., Ka, S. O., Kim, J. H., et al. (2016). Glucocorticoid suppresses Connexin 43 expression by inhibiting the Akt/mTOR signaling pathway in osteoblasts. *Calcif. Tissue Int.* 99, 88–97. doi: 10.1007/s00223-016-0121-y

- Si, X. H., Miguel-Hidalgo, J. J., O'dwyer, G., Stockmeier, C. A., and Rajkowska, G. (2004). Age-dependent reductions in the level of glial fibrillary acidic protein in the prefrontal cortex in major depression. *Neuropsychopharmacology* 29, 2088–2096. doi: 10.1038/Sj.Npp.1300525
- Sofroniew, M. V. (2014). Multiple roles for astrocytes as effectors of cytokines and inflammatory mediators. *Neuroscientist* 20, 160–172. doi: 10.1177/1073858413504466
- Sun, J. D., Liu, Y., Yuan, Y. H., Li, J., and Chen, N. H. (2012). Gap junction dysfunction in the prefrontal cortex induces depressive-like behaviors in rats. *Neuropsychopharmacology* 37, 1305–1320. doi: 10.1038/npp.2011.319
- Tanti, A., Lutz, P. E., Kim, J., O'leary, L., Theroux, J. F., Turecki, G., et al. (2019). Evidence of decreased gap junction coupling between astrocytes and oligodendrocytes in the anterior cingulate cortex of depressed suicides. *Neuropsychopharmacology* 44, 2099–2111. doi: 10.1038/s41386-019-0471-z
- Tao, K., Cai, Q., Zhang, X., Zhu, L., Liu, Z., Li, F., et al. (2020). Astrocytic histone deacetylase 2 facilitates delayed depression and memory impairment after subarachnoid hemorrhage by negatively regulating glutamate transporter-1. *Ann. Transl. Med.* 8:691. doi: 10.21037/atm-20-4330
- Treccani, G., Musazzi, L., Perego, C., Milanese, M., Nava, N., Bonifacino, T., et al. (2014). Stress and corticosterone increase the readily releasable pool of glutamate vesicles in synaptic terminals of prefrontal and frontal cortex. *Mol. Psychiatry* 19, 433–443. doi: 10.1038/mp.2014.5
- Tress, O., Maglione, M., May, D., Pivneva, T., Richter, N., Seyfarth, J., et al. (2012). Panglial gap junctional communication is essential for maintenance of myelin in the CNS. *J. Neurosci.* 32, 7499–7518. doi: 10.1523/JNEUROSCI.0392-12.2012
- Tynan, R. J., Beynon, S. B., Hinwood, M., Johnson, S. J., Nilsson, M., Woods, J. J., et al. (2013). Chronic stress-induced disruption of the astrocyte network is driven by structural atrophy and not loss of astrocytes. *Acta Neuropathol.* 126, 75–91. doi: 10.1007/s00401-013-1102-0
- Xia, C. Y., Chu, S. F., Zhang, S., Gao, Y., Ren, Q., Lou, Y. X., et al. (2017). Ginsenoside Rg1 alleviates corticosterone-induced dysfunction of gap junctions in astrocytes. *J. Ethnopharmacol.* 208, 207–213. doi: 10.1016/j.jep.2017.06.031
- Xia, C. Y., Wang, Z. Z., Zhang, Z., Chen, J., Wang, Y. Y., Lou, Y. X., et al. (2018). Corticosterone impairs gap junctions in the prefrontal cortical and hippocampal astrocytes via different mechanisms. *Neuropharmacology* 131, 20–30. doi: 10.1016/j.neuropharm.2017.12.003
- Yoshino, K., Oda, Y., Kimura, M., Kimura, H., Nangaku, M., Shirayama, Y., et al. (2020). The alterations of glutamate transporter 1 and glutamine synthetase in the rat brain of a learned helplessness model of depression. *Psychopharmacology (Berl.)* 237, 2547–2553. doi: 10.1007/s00213-020-05555-3
- Yuen, E. Y., Wei, J., Liu, W., Zhong, P., Li, X., and Yan, Z. (2012). Repeated stress causes cognitive impairment by suppressing glutamate receptor expression and function in prefrontal cortex. *Neuron* 73, 962–977. doi: 10.1016/j.neuron.2011.12.033
- Yun, J. H., Park, S. J., Jo, A., Kang, J. L., Jou, I., Park, J. S., et al. (2011). Caveolin-1 is involved in reactive oxygen species-induced SHP-2 activation in astrocytes. *Exp. Mol. Med.* 43, 660–668. doi: 10.3858/emmm.2011.43.12.075
- Zhang, H., Zhao, Y., and Wang, Z. (2015). Chronic corticosterone exposure reduces hippocampal astrocyte structural plasticity and induces hippocampal atrophy in mice. *Neurosci. Lett.* 592, 76–81. doi: 10.1016/j.neulet.2015.03.006
- Zhang, Y., Chen, K., Sloan, S. A., Bennett, M. L., Scholze, A. R., O'Keeffe, S., et al. (2014). An RNA-sequencing transcriptome and splicing database of glia, neurons, and vascular cells of the cerebral cortex. *J. Neurosci.* 34, 11929–11947. doi: 10.1523/JNEUROSCI.1860-14.2014
- Zhao, Y., Rivieccio, M. A., Lutz, S., Scemes, E., and Brosnan, C. F. (2006). The TLR3 ligand polyI:C downregulates connexin 43 expression and function in astrocytes by a mechanism involving the NF-kappaB and PI3 kinase pathways. *Glia* 54, 775–785. doi: 10.1002/glia.20418
- Zhuang, K., Huang, C., Leng, L., Zheng, H., Gao, Y., Chen, G., et al. (2018). Neuron-specific menin deletion leads to synaptic dysfunction and cognitive impairment by modulating p35 expression. *Cell Rep.* 24, 701–712. doi: 10.1016/j.celrep.2018.06.055
- Zink, M., Vollmayr, B., Gebicke-Haerter, P. J., and Henn, F. A. (2010). Reduced expression of glutamate transporters vGluT1, EAAT2 and EAAT4 in learned helpless rats, an animal model of depression. *Neuropharmacology* 58, 465–473. doi: 10.1016/j.neuropharm.2009.09.005

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