

Draft Genome Sequence of *Mycobacterium chelonae* Type Strain ATCC 35752

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***Mycobacterium chelonae* is a rapidly growing opportunistic nontuberculous mycobacterial (NTM) species that causes infections in humans and other hosts. Here, we report the draft genome sequence of *Mycobacterium chelonae* type strain ATCC 35752, consisting of 4.89 Mbp, 63.96% G+C content, 4,489 protein-coding genes, 48 tRNAs, and 3 rRNA genes.**

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Mycobacterium chelonae is an opportunistic environmental pathogen (1) found in diverse habitats, including water, sewage, and soil (2). *M. chelonae* can cause corneal, cutaneous, and pulmonary infections in humans and other hosts. In many health care settings, the occurrence and spread of atypical mycobacterial infections, including those caused by *M. abscessus* and *M. chelonae*, have increased in prevalence in recent years. *M. chelonae* outbreaks have been attributed to factors ranging from resistance to glutaraldehyde, a commonly used hospital disinfectant (3–6) to contaminated medical equipment (7) and contaminated reagents (8).

We have sequenced and report here the first draft genome of a representative of the *M. chelonae* species. The type strain ATCC 35752 was originally isolated from the lungs of sea turtles (*Chelona corticata*), and shares phylogenetic similarity to human pathogens of this species. The genome of the *M. chelonae* type strain ATCC 35752 was assembled using 400-bp sequence reads generated from the Life Technologies Ion Torrent PGM (Thermo, Fisher Sciences, Carlsbad, CA) and 250 × 250 bp paired-end reads from the Illumina MiSeq (Illumina, Inc., San Diego, CA), at 220× coverage. Sequence reads were quality filtered using the Fastx_Toolkit (http://hannonlab.cshl.edu/fastx_toolkit/) and assembled into scaffolded contigs using the Newbler v2.9 (454 Life Sciences, Branford, CT) software package. The contigs were ordered in relation to the *M. abscessus* subsp. *abscessus* ATCC 19977 genome as a reference (9), employing the progressive Mauve 2.3.1 whole-genome alignment algorithm (10). Genomic features were annotated using the NCBI Prokaryotic Genome Annotation Pipeline (11).

The *M. chelonae* ATCC 35752 draft genome is a pseudomolecule composed of 24 contigs, with linking sequences of 100 N's in between each contig, resulting in a total assembly size of 4,898,027 bp and a G+C content of 63.96%. The average contig length is 204,084 bp with an N_{50} of 462,745 bp. A total of 4,489 coding sequences (CDSs) were predicted, including 3,274 CDSs

(72.93%) with functional annotations and 1,215 CDSs (27.07%) annotated as hypothetical proteins. Our genome assembly contains 48 tRNAs, 1 noncoding RNA (ncRNA), and 1 rRNA cistron consisting of the 5S, 16S, and 23S rRNA genes. A comparative analysis between *M. chelonae* ATCC 35752 and *M. abscessus* ATCC 19977 revealed 3,803 orthologous CDSs that are shared between the two strains.

Whole-genome sequence alignments of *M. chelonae* ATCC 35752 and five representative *M. abscessus* genomes revealed an average of 620,054 single nucleotide polymorphisms (SNPs), including 628,647 SNPs compared to *M. abscessus* ATCC 19977, 629,586 SNPs compared to *M. bolletii* BD^T, 631,431 SNPs compared to *M. bolletii* M24, 621,641 SNPs compared to *M. massiliense* GO-06, and 588,967 SNPs compared to *M. massiliense* CCUG 48898 (= JCM15300). These SNP differences (12.02 to 12.85% of the *M. chelonae* ATCC 35752 genome) underscore the genomic divergence between *M. chelonae* and its sister taxa (*M. abscessus* complex).

Nucleotide sequence accession numbers. The draft genome sequence of *M. chelonae* ATCC 35752 has been deposited in NCBI GenBank under the accession no. [CP010946](https://www.ncbi.nlm.nih.gov/nuccore/CP010946), BioProject no. PRJNA251569, and BioSample no. SAMN02837161.

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REFERENCES

1. Brown-Elliott BA, Wallace RJ. 2002. Clinical and taxonomic status of pathogenic nonpigmented or late-pigmenting rapidly growing mycobacteria. *Clin Microbiol Rev* 15:716–746. <http://dx.doi.org/10.1128/CMR.15.4.716-746.2002>.
2. Carson LA, Bland LA, Cusick LB, Favero MS, Bolan GA, Reingold AL, Good RC. 1988. Prevalence of nontuberculous mycobacteria in water supplies of hemodialysis centers. *Appl Environ Microbiol* 54:3122–3125.
3. Leao SC, Tortoli E, Viana-Niero C, Ueki SY, Lima KV, Lopes ML, Yubero J, Menendez MC, Garcia MJ. 2009. Characterization of mycobacteria from a major Brazilian outbreak suggests that revision of the taxonomic status of members of the *Mycobacterium chelonae*-*M. Abscessus* group is needed. *J Clin Microbiol* 47:2691–2698. <http://dx.doi.org/10.1128/JCM.00808-09>.
4. Chroneou A, Zimmerman SK, Cook S, Willey S, Eyre-Kelly J, Zias N, Shapiro DS, Beamis Jr JF, Craven DE. 2008. Molecular typing of *Mycobacterium chelonae* isolates from a pseudo-outbreak involving an automated bronchoscope washer. *Infect Control Hosp Epidemiol* 29:1088–1090. <http://dx.doi.org/10.1086/591451>.
5. Meyers H, Brown-Elliott BA, Moore D, Curry J, Truong C, Zhang Y, Wallace Jr RJ. 2002. An outbreak of *Mycobacterium chelonae* infection following liposuction. *Clin Infect Dis* 34:1500–1507. <http://dx.doi.org/10.1086/340399>.
6. Kressel AB, Kidd F. 2001. Pseudo-outbreak of *Mycobacterium chelonae* and *Methylobacterium mesophilicum* caused by contamination of an automated endoscopy washer. *Infect Control Hosp Epidemiol* 22:414–418. <http://dx.doi.org/10.1086/501926>.
7. Fraser VJ, Jones M, Murray PR, Medoff G, Zhang Y, Wallace RJ. 1992. Contamination of flexible fiberoptic bronchoscopes with *Mycobacterium chelonae* linked to an automated bronchoscope disinfection machine. *Am Rev Respir Dis* 145:853–855. http://dx.doi.org/10.1164/ajrccm/145.4_Pt_1.853.
8. Drage LA, Ecker PM, Orenstein R, Phillips PK, Edson RS. 2010. An outbreak of *Mycobacterium chelonae* infections in tattoos. *J Am Acad Dermatol* 62:501–506. <http://dx.doi.org/10.1016/j.jaad.2009.03.034>.
9. Ripoll F, Pasek S, Schenowitz C, Dossat C, Barbe V, Rottman M, Macheras E, Heym B, Herrmann J-L, Daffé M, Brosch R, Risler J-L, Gaillard J-L. 2009. Non mycobacterial virulence genes in the genome of the emerging pathogen *Mycobacterium abscessus*. *PLoS One* 4:e5660. <http://dx.doi.org/10.1371/journal.pone.0005660>.
10. Darling AE, Mau B, Perna NT. 2010. progressiveMauve: multiple genome alignment with gene gain, loss and rearrangement. *PLoS One* 5:e11147. <http://dx.doi.org/10.1371/journal.pone.0011147>.
11. Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Ciufu S, Li W. 2013. Prokaryotic Genome Annotation Pipeline. In Beck J, Benson D, Coleman J, Hoepfner M, Johnson M, Maglott M, Mizrahi I, Morris R, Ostell J, Pruitt K, Rubinstein W, Sayers E, Sirotkin K, Tatusova T (ed), *The NCBI handbook*, 2nd ed., NCBI, Bethesda, MD. <http://www.ncbi.nlm.nih.gov/books/NBK174280>.