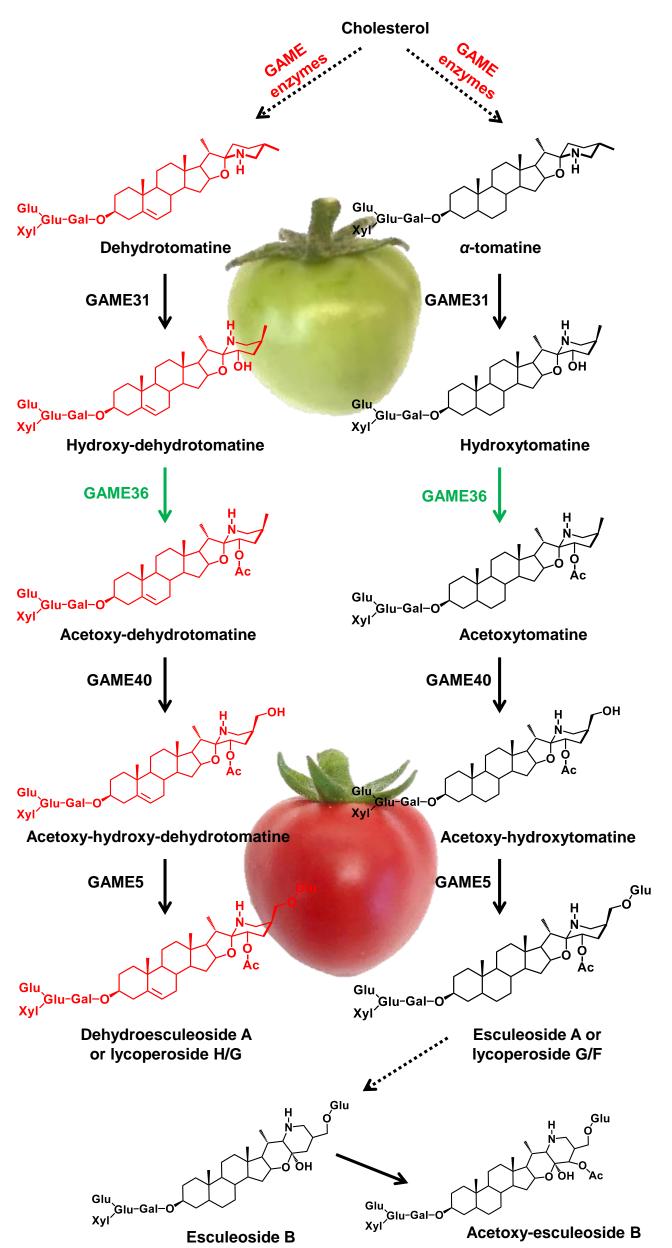
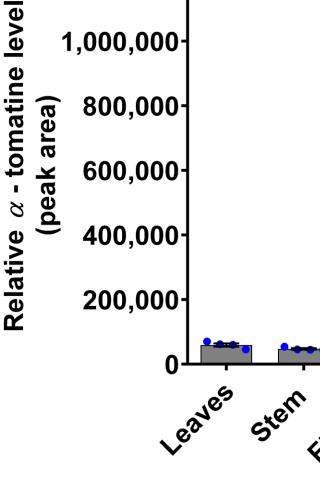
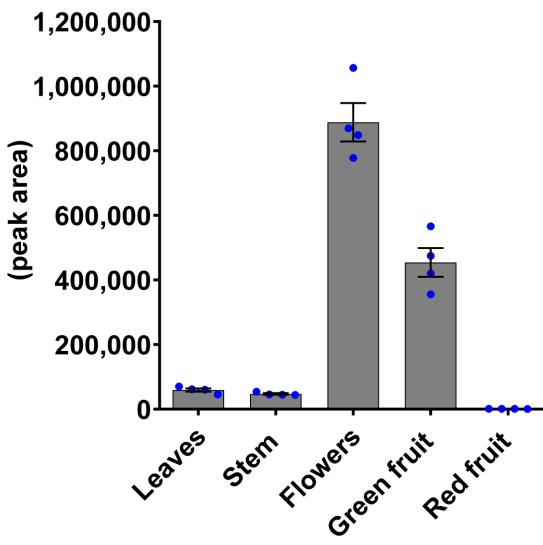
A BAHD-type Acyltransferase Concludes the Biosynthetic Pathway of Non-bitter Glycoalkaloids in Ripe Tomato Fruit

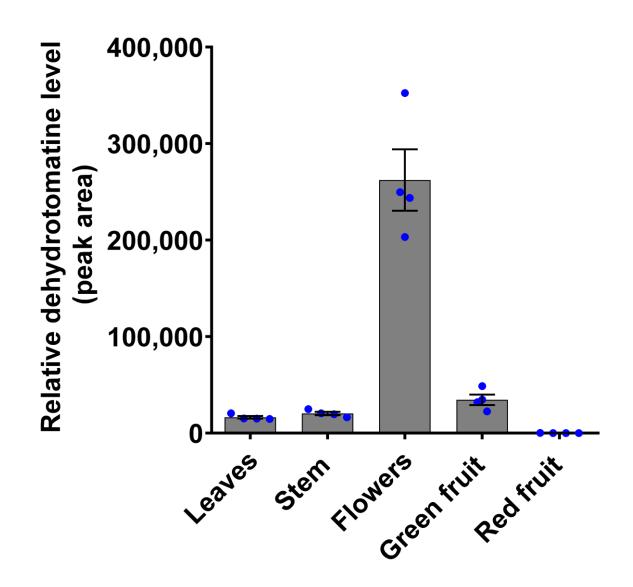
Sonawane et al.

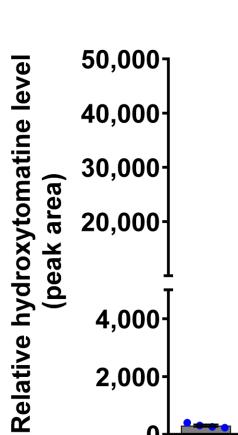


Supplementary Figure 1: Steroidal glycoalkaloids (SGAs) metabolism during tomato fruit development and ripening. In cultivated tomato, the ripening associated chemical shift in SGAs metabolism involves conversion of α -tomatine and dehydrotomatine (major SGAs in green fruit) to Esculeoside A and dehydroesculeoside A (major SGAs in red ripe fruit), respectively. The known SGA biosynthetic enzymes (i.e. GAME31, GAME40 and GAME5) in the Esculeoside A pathway are marked in black. GAME36 enzymatic step discovered in this study is shown in green. Throughout tomato fruit development and ripening, α -tomatine derived SGAs (shown in black) are highly abundant as compared to the dehydrotomatine-derived SGAs (marked in red). Dashed arrows represent multiple biosynthetic reactions whereas solid arrows represent a single step. GAME: GLYCOALKALOID METABOLISM; Glu: Glucose; Gal: Galactose; Xyl: Xylose; Rha: Rhamnose and Ac: Acetoxy.









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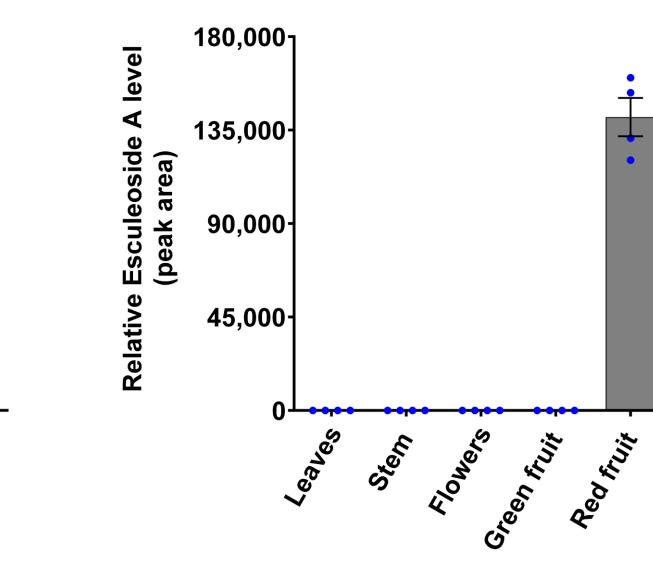
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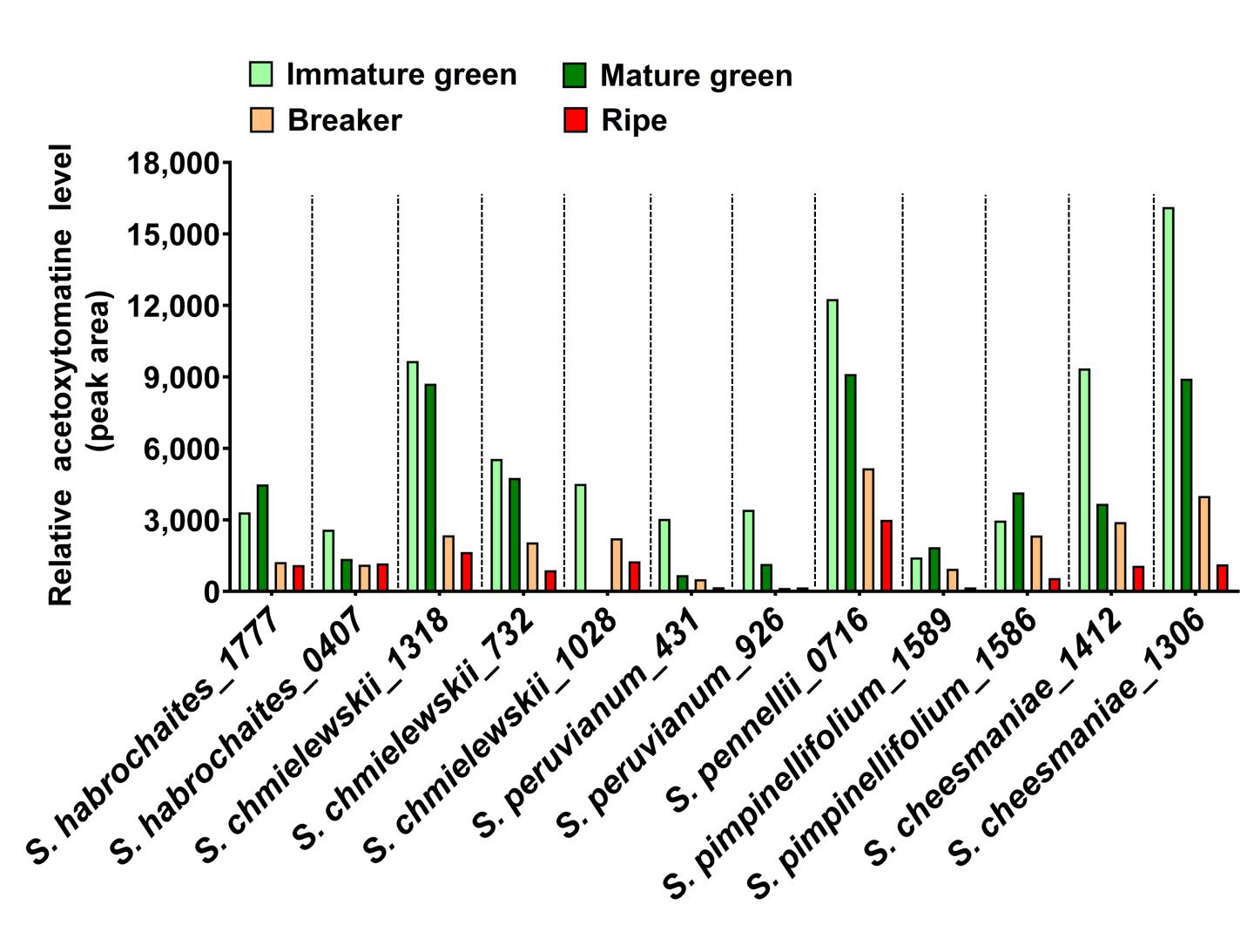
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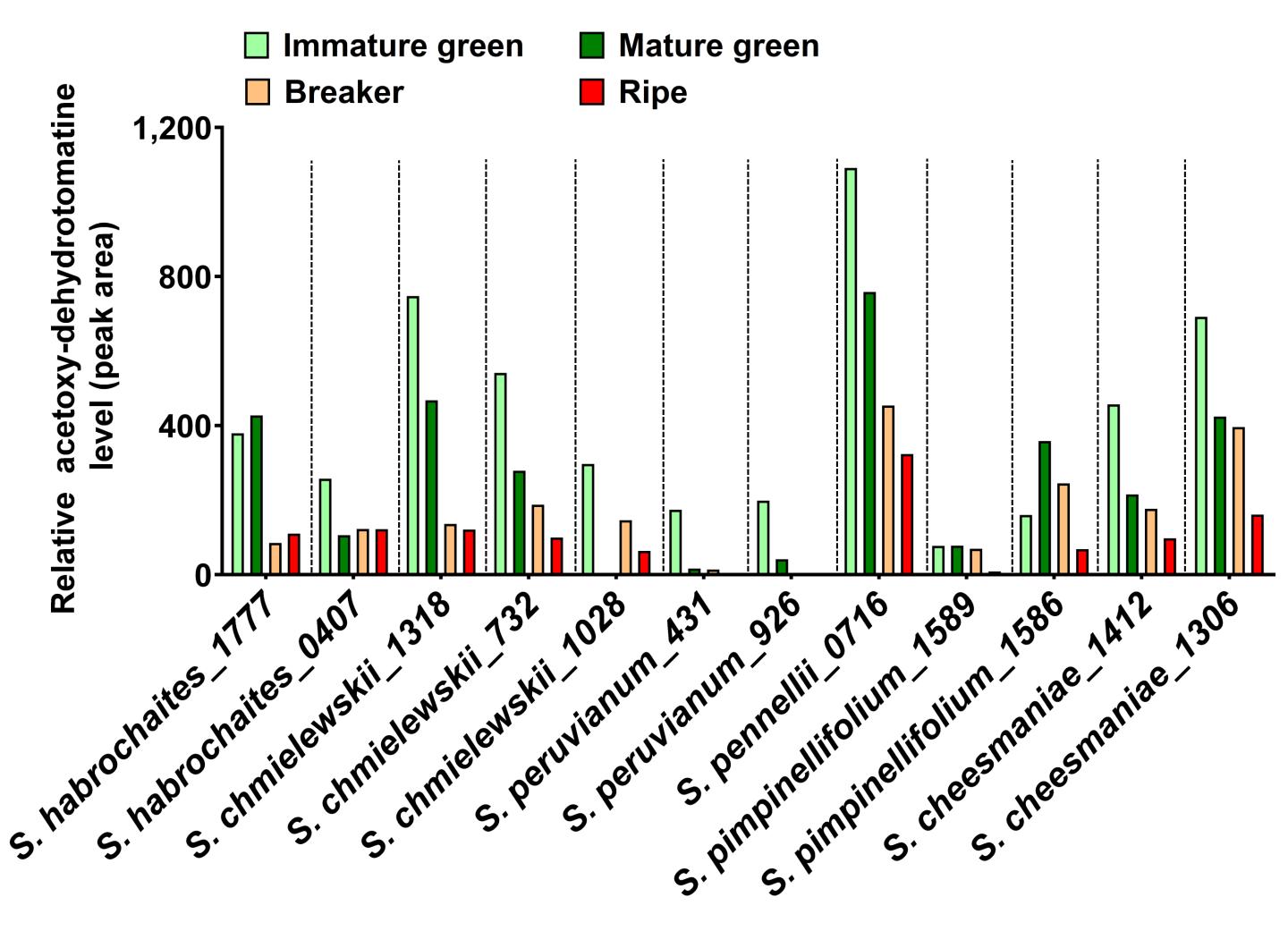
a

b

d

Supplementary Figure 2: Selected SGAs profiling in different tissues of cultivated tomato (*S. lycopersicum cv.* Micro Tom). (a-d) α -tomatine (a), dehydrotomatine (b), hydroxytomatine (c) and Esculeoside A (d) content was determined by LC-MS analysis. The peak areas for each SGA were determined using the TargetLynx software. The values indicate means of biological replicates ± standard error mean (n=4).

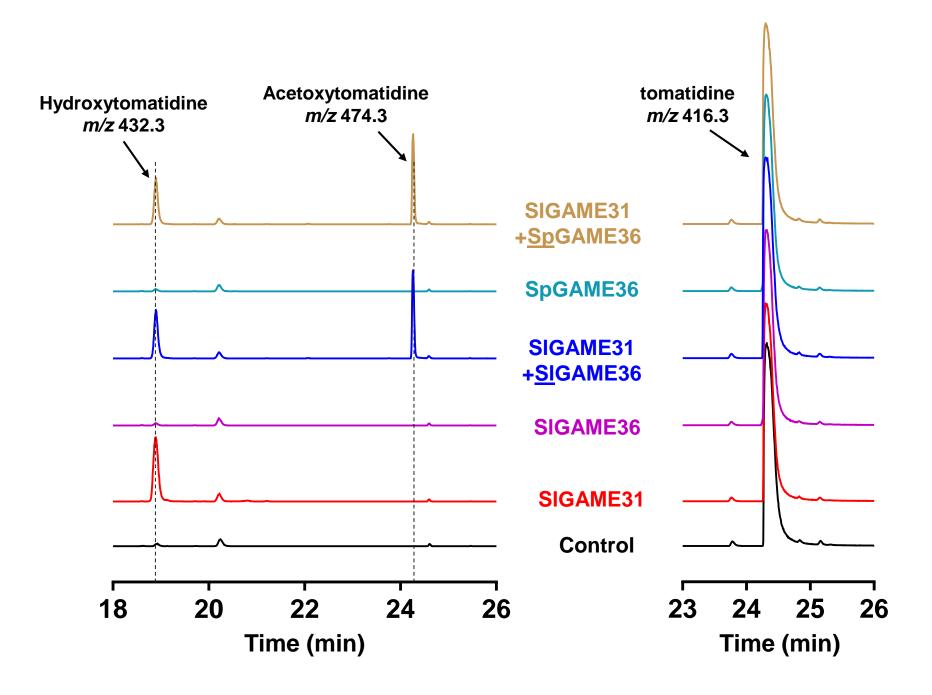




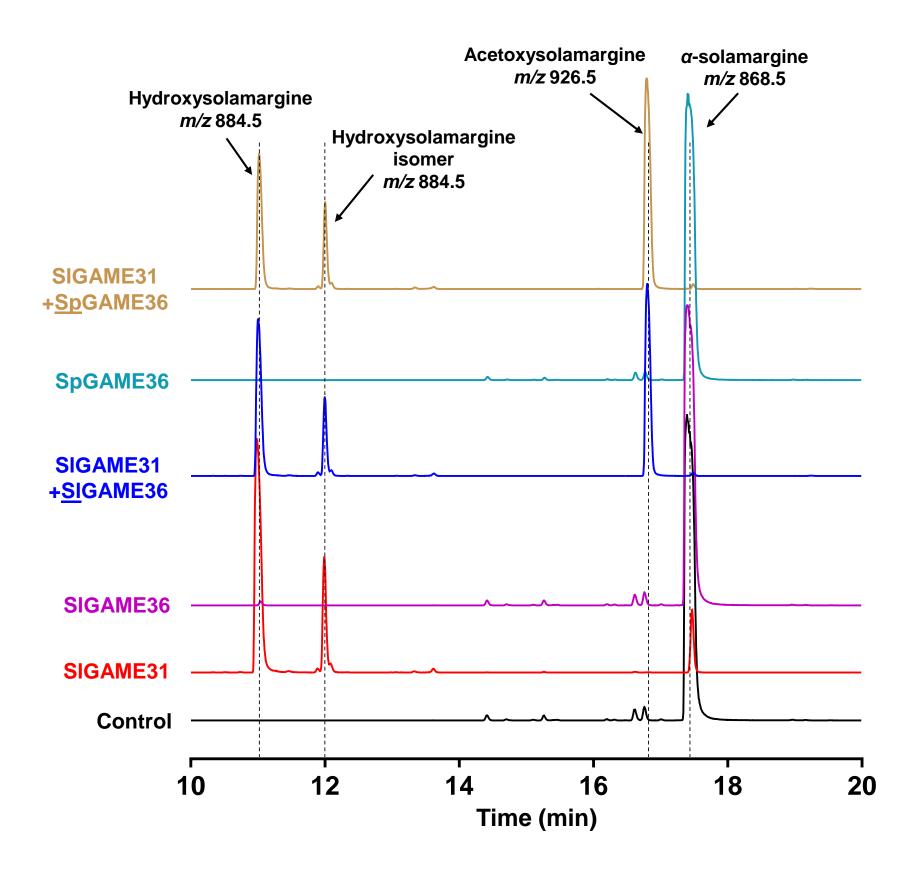
Supplementary Figure 3: Acetoxytomatine and acetoxydehydrotomatine levels in wild tomato accessions. SGA analysis was performed in four fruit developmental stages (i.e. immature green, mature green, breaker and ripe) of 12 wild tomato accessions by LC-MS (n=1, single replicate for each fruit ripening stage obtained by extracting several fruits of an individual wild tomato accession). The peak areas were determined using the TargetLynx software.

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Solyc09g014280	230	533	19		105	211	17	66	
Solyc07g049660	198	145	0		39	179	1	13	
Solyc11g008630	123	21	0		25	2	0	16	
Solyc09g092270	94	27	70		4	2	0	42	
Solyc12g005430	89	43	0		60	11	0	37	
Solyc12g096770	84	91	0		11	14	O	75	
Solyc12g087980	76	14	11		5	1	0	10	
Solyc08g075210	53	109	50	- <u>-</u>	 164	99	34	34	
Solyc04g079720	36	45	15		41	29	10	27	
Solyc12g010980	9	8	0		18	1	0	136	
Solyc07g006680	7	3	1		29	4	1	18	
Min.									Max.

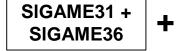
Supplementary Figure 4: Expression profile of BAHD candidate genes in cultivated tomato (*cv.* Micro Tom) tissue types and fruit developmental stages (RNA-seq expression data). Normalized FPKM (Fragments Per Kilobase of transcript per Million mapped reads) values were used to infer the expression profile. Fruit developmental stages- IG: immature green; MG: mature green; RR: red ripe. The BAHD candidate Solyc08g075210 (GAME36) characterized further in this study is marked in dashed black rectangle.

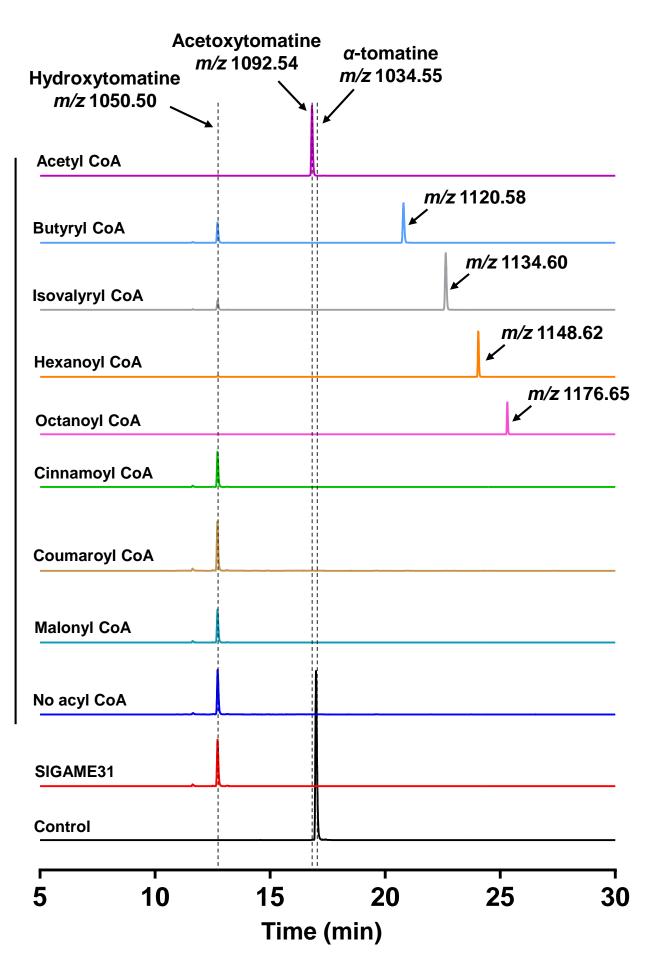


Supplementary Figure 5: LC-MS analysis of recombinant GAME31 and GAME36 enzyme assay with tomatidine substrate. Recombinant SIGAME36 [S. lycopersicum (cv. Micro Tom); shown in blue] and SpGAME36 (S. pennellii, in tan brown) enzymes convert tomatidine to acetoxytomatidine when assayed together with SIGAME31 (each enzyme produced separately in *E. coli* cells). Control reaction with tomatidine (shown in black) was performed using the protein extracts from *E. coli* cells transformed with empty vector. Individual assays of SIGAME31, SIGAME36 and SpGAME36 recombinant enzymes are shown in red, dark purple and turquoise colors, respectively. Mass to charge (m/z) is shown for substrate and assay products. MS/MS analysis and identification of assay products is provided in Supplementary Data 3.

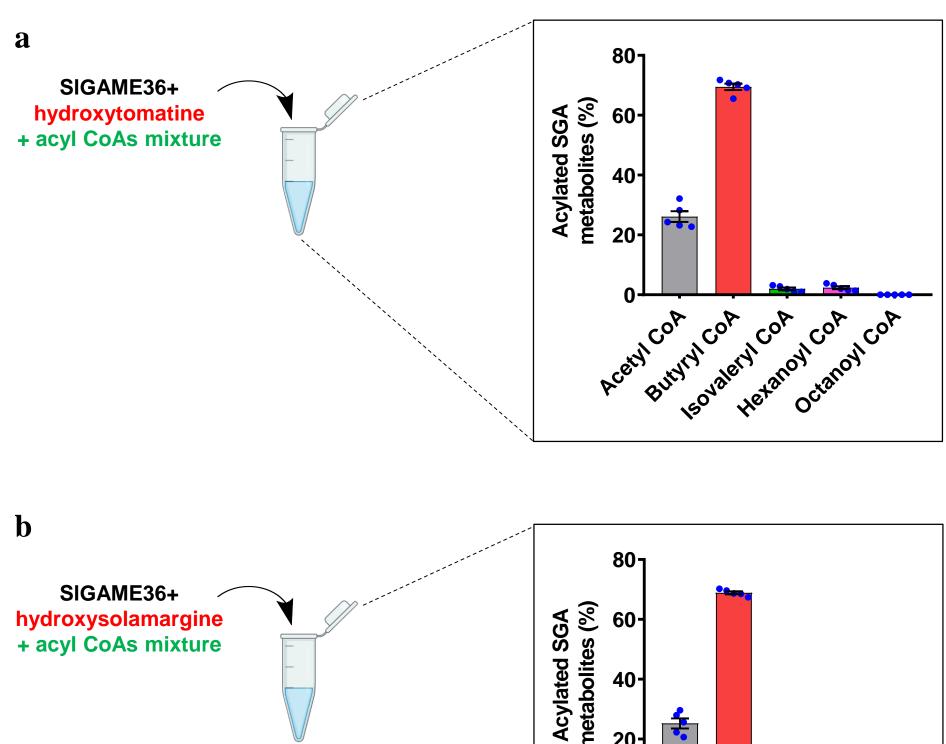


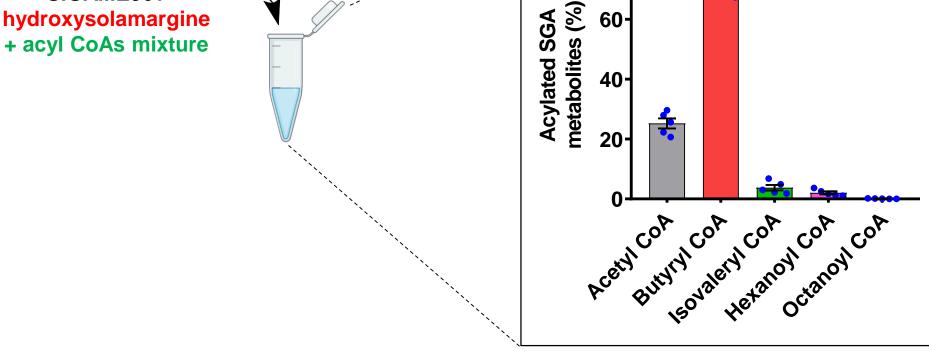
Supplementary Figure 6: In vitro coupled enzyme assays using recombinant GAME31 and GAME36 proteins. Aligned chromatograms (extracted ion) of assay products (i.e. hydroxysolamargine and acetoxysolamargine) and substrate (α -solamargine) are presented here. Blue: Recombinant SIGAME36 and SIGAME31 enzyme assay; Tan brown: Recombinant SpGAME36 and SIGAME31 enzyme assay; Black: control reaction using protein extracts from empty vector transformed *E. coli* cells; other (red, dark purple and turquoise): individual assays of SIGAME31, SIGAME36 and SpGAME36 recombinant enzymes with α -solamargine substrate. The recombinant SIGAME31, SIGAME36, SpGAME36 enzymes were produced separately in *E. coli* cells. Enzyme assays analysis was carried out by LC-MS (40 min. run). Mass to charge (m/z) is shown for substrate and assay products. MS/MS analysis and identification of assay products is provided in Supplementary Data 3.





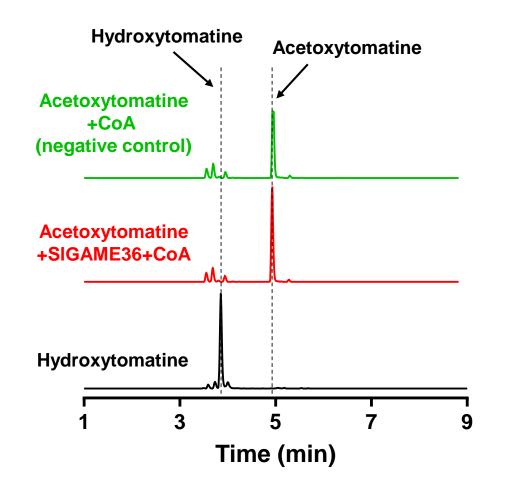
Supplementary Figure 7: *In vitro* coupled enzyme assays of recombinant GAME31 and GAME36 proteins using different CoA thioesters (acyl donors) as substrates. Aligned chromatograms (extracted ion) of assay products (i.e. acylated SGAs) and substrates (α -tomatine, hydroxytomatine) are presented here. Control reaction (in black) was performed using protein extracts from empty vector transformed *E. coli* cells. Enzyme assays analysis was carried out by LC-MS (40 min. run). Mass to charge (*m/z*) is shown for substrate and assay products.

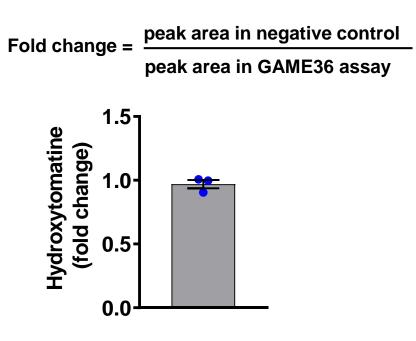




Supplementary Figure 8: In vitro production of acylated SGAs by GAME36 using different acyl-CoA donors with hydroxytomatine (a) and hydroxysolamargine (b) substrates. Equimolar mixture of acyl-CoAs (acetyl, butyryl, isovalyryl, hexanoyl, and octanoyl) each at 0.2 mM were included in the enzymatic reaction with either substrate (hydroxytomatine and hydroxysolamargine). Each assay reaction was monitored at five different time points (5, 10, 20, 40 and 60 min.), then terminated and extracted after completion of given time point. Peak areas of each acylated product were normalized to the sum of all detected acylated SGA metabolites at given time point of assay reaction, and plotted as a percentage. Values indicates means ± standard error mean (n=5, obtained for each assay reaction from five different time points). Enzyme assays analysis was carried out by LC-MS.

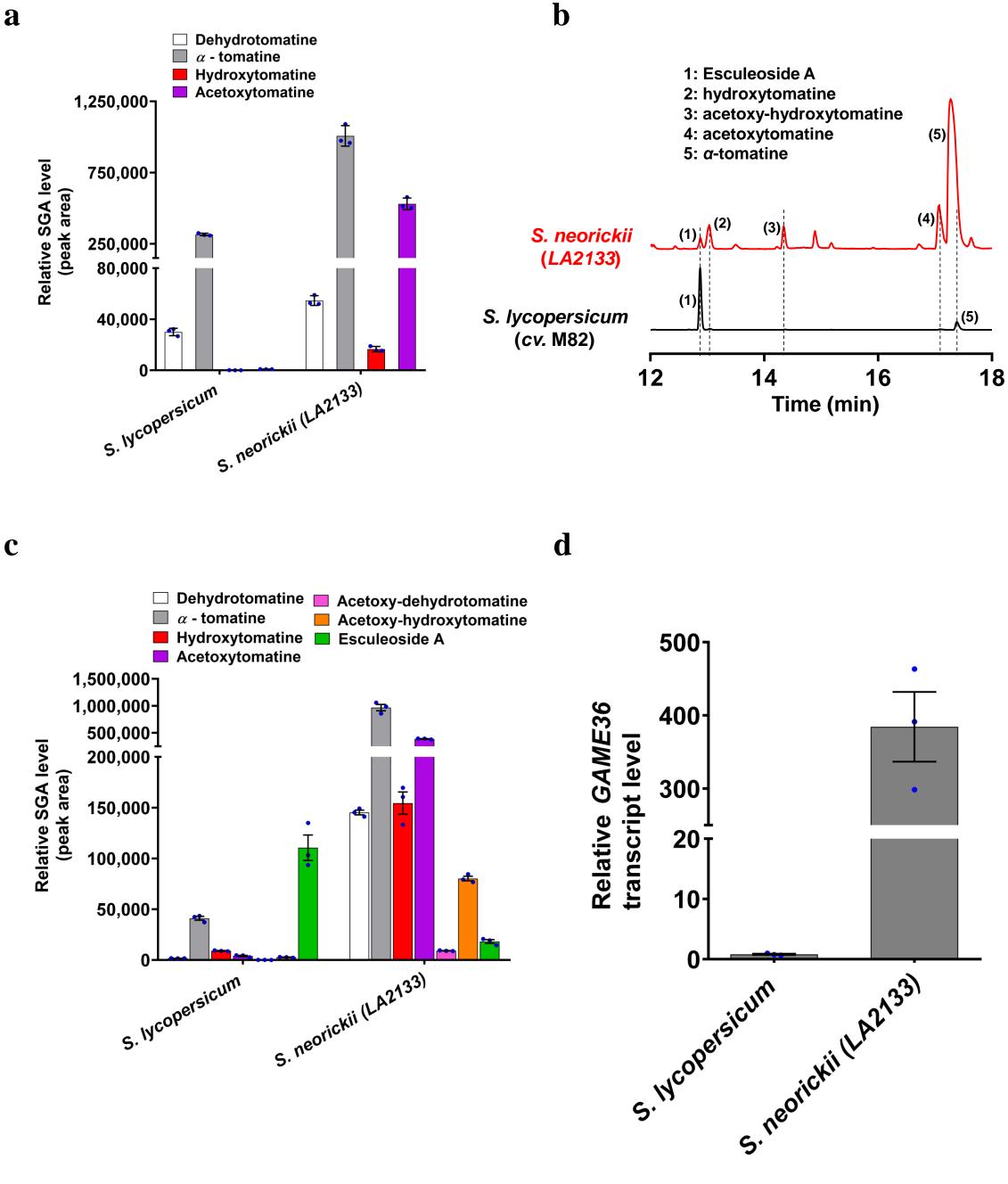
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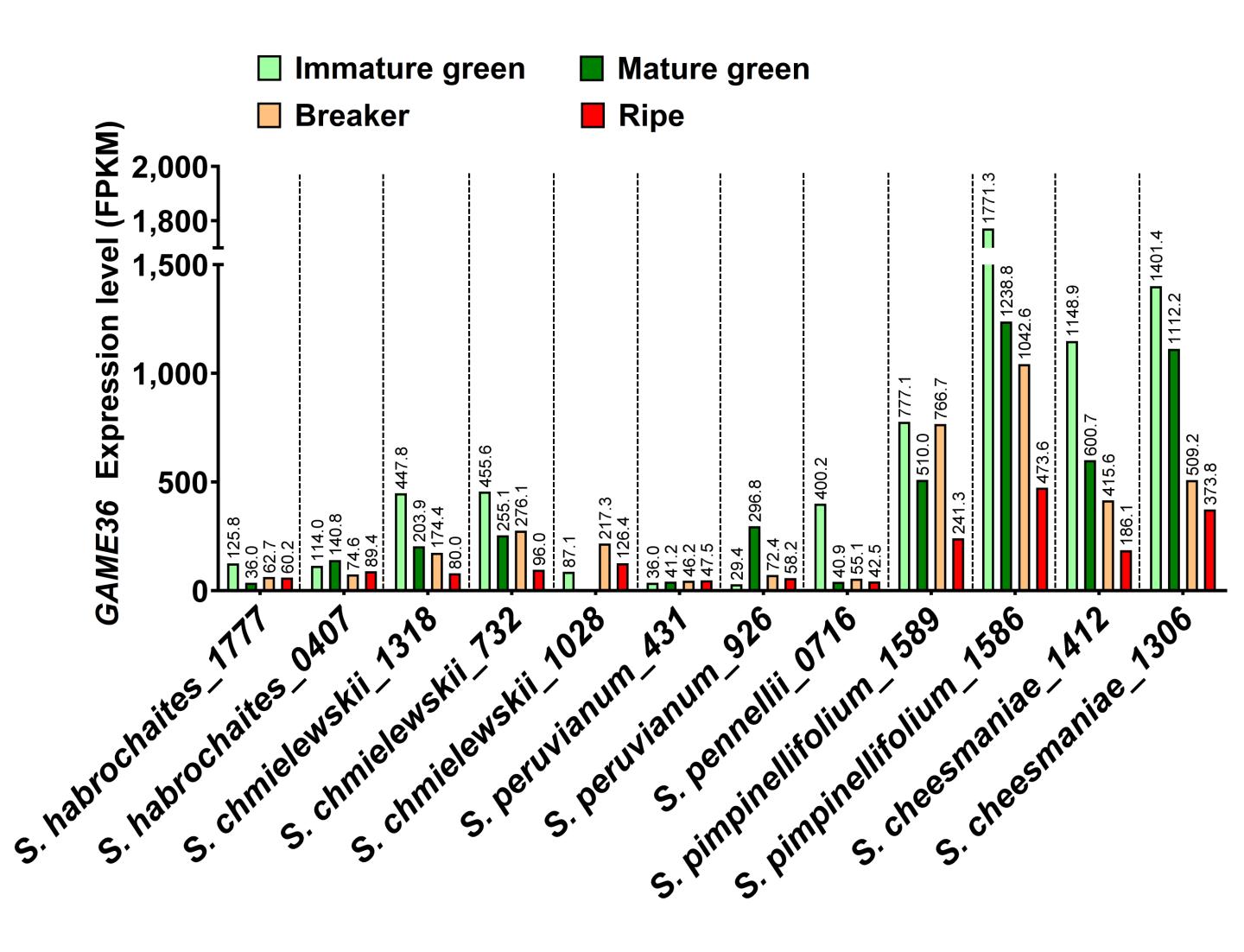


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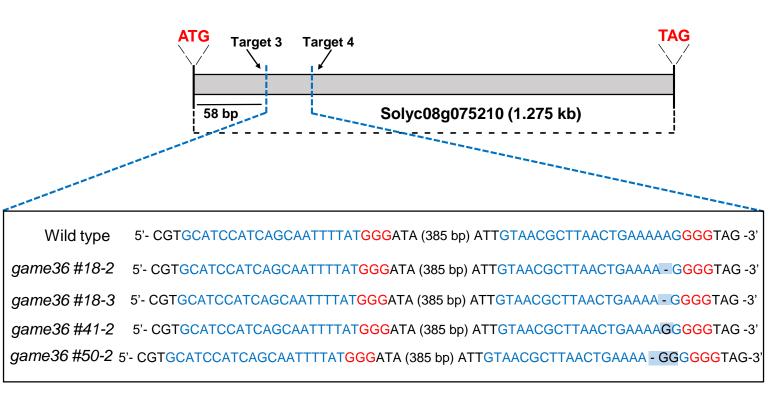
Supplementary Figure 9: Reversible reactions test catalyzed by SIGAME36. Recombinant SIGAME36 enzyme did not catalyze reversible reaction when assayed together with acetoxytomatine and CoA. (a) Aligned extracted ion chromatograms of expected assay product (hydroxytomatine) and substrate (acetoxytomatine) are presented here. Black: hydroxytomatine; Red: GAME36 assay reaction; Green: negative control reaction using acetoxytomatine and CoA, but without GAME36 enzyme. Enzyme assays analysis was carried out by LC-MS (15 min. run). Mass to charge (m/z) is shown for metabolites. (b) Hydroxytomatine fold change between negative control and GAME36 enzyme assay. As acetoxytomatine used in the 'reversible' reaction assay contained traces of hydroxytomatine, fold change of hydroxytomatine peak area between negative control and assay with GAME36 is presented. Values in panel b indicates means of fold change ± standard error mean (n=3) obtained from each three independent negative control and GAME36 reactions.



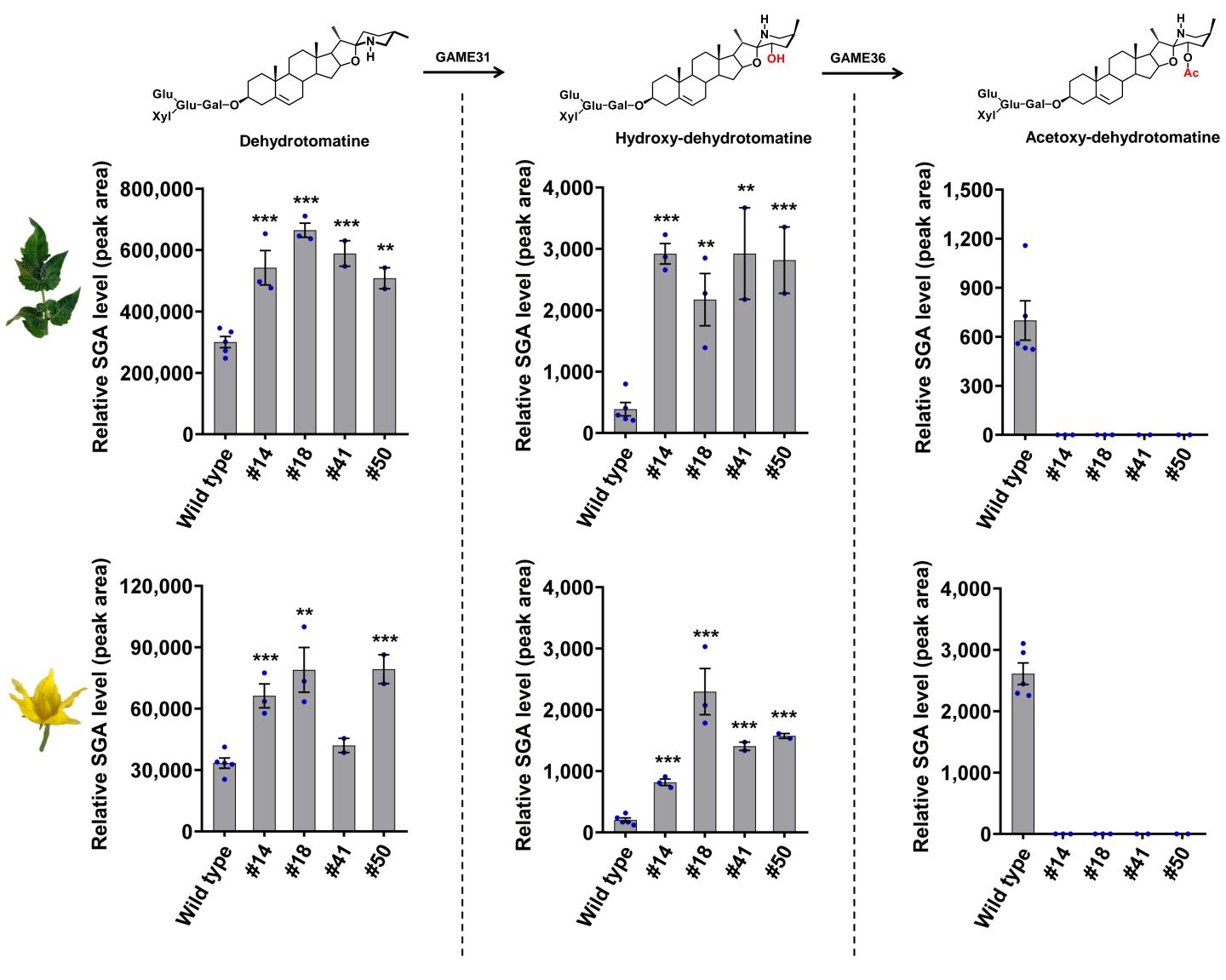
Supplementary Figure 10: LC-MS based profiling of SGAs and GAME36 gene expression in wild tomato S. neorickii LA2133. (a) α-tomatine and downstream SGAs (e.g. hydroxytomatine, acetoxytomatine etc.) content in leaves of S. neorickii LA2133 accession and cultivated tomato (S. lycopersicum cv. M82). (b) Comparison of ripening associated SGA profiles between cultivated tomato and S. neorickii LA2133 accession. Representative aligned extracted ion chromatograms for ripe fruit SGAs are shown. (c) Ripe fruit SGAs repertoire in cultivated tomato and S. neorickii LA2133 accession. Not as previously reported, S. neorickii LA2133 accumulates acetoxytomatine and downstream intermediates in the Esculeoside A pathway. LC-MS was used for targeted SGA profiling. (d) GAME36 gene expression in leaves of cultivated tomato (cv. M82) and wild tomato S. neorickii LA2133 accession as determined by qRT-PCR. Values in panels (a, c and d) represent the means of three biological replicates ± standard error mean (n=3).



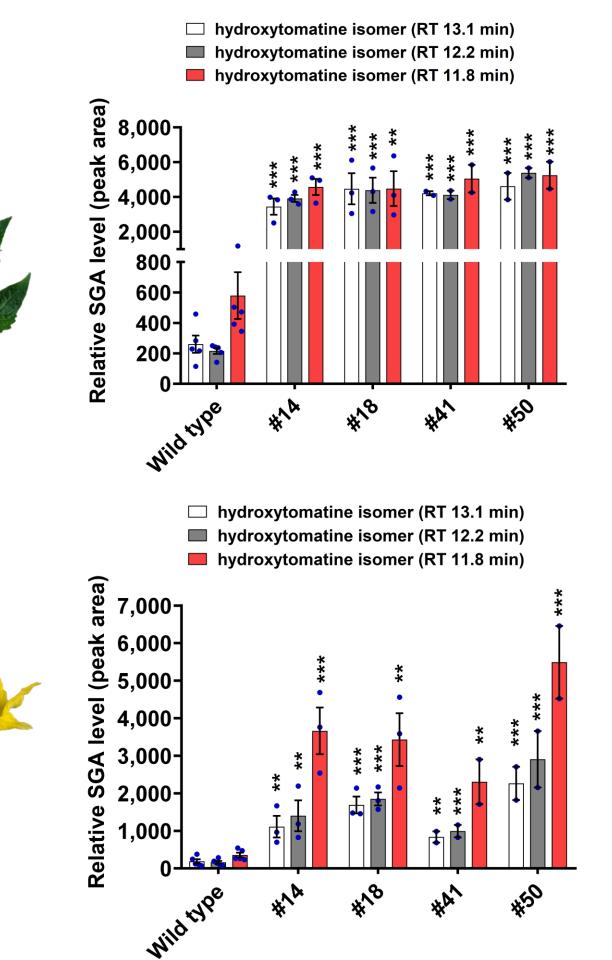
Supplementary Figure 11: GAME36 expression levels in four fruit developmental stages of wild tomato accessions (normalized RNA-seq data). FPKM: Fragments Per Kilobase of transcript per Million mapped reads.



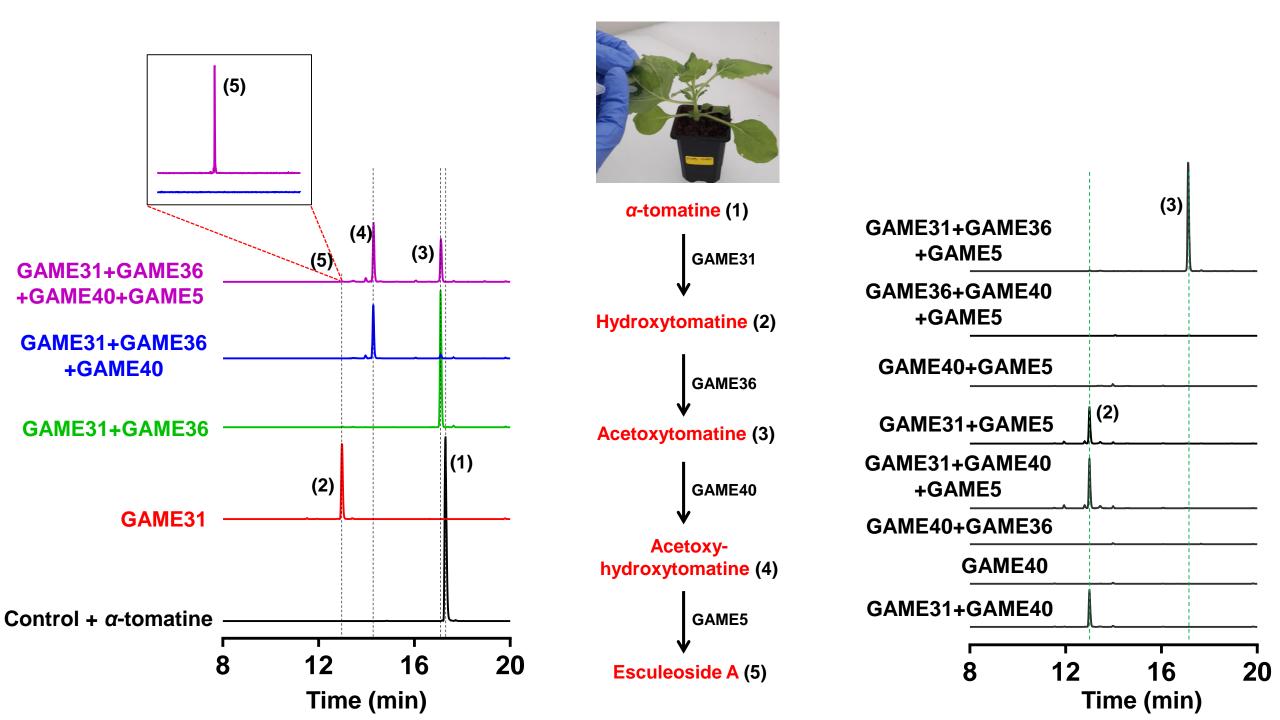
Supplementary Figure 12: Schematic representation of *GAME36* with location of guide RNA sequences, CRISPR-Cas9 cleavage sites and *game36* mutant sequences. PAMs and inserted bases are shown in red and blue respectively and deleted bases are replaced by a dash. Four independent (#14, #18, #41 and #50) homozygous *game36* mutant lines were generated in tomato (*cv.* Micro Tom) background.



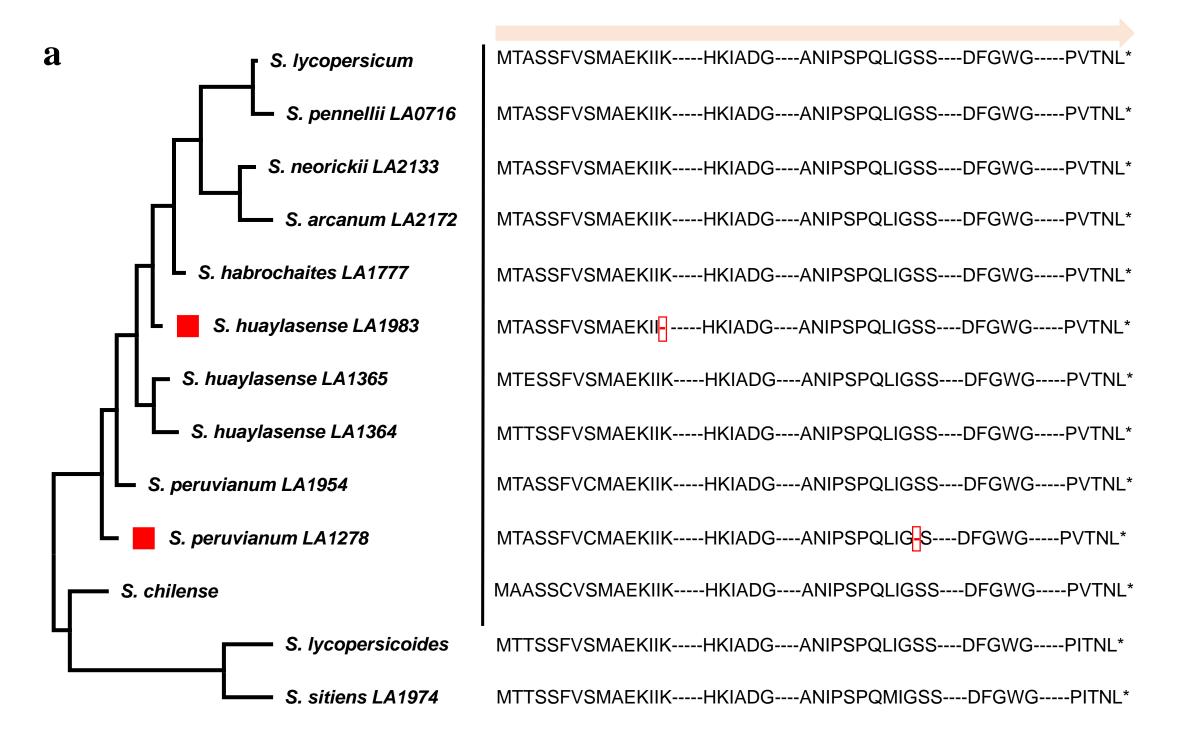
Supplementary Figure 13: Levels of dehydrotomatine-derived SGAs detected in leaves and flowers of *game36* mutant lines as compared to wild type ones. Acetoxy-dehydrotomatine was not detected in the *game36* mutant lines. Four independent (#14, #18, #41 and #50) homozygous *game36* mutant lines were generated in tomato (cv. Micro Tom) background. The values indicate means of biological replicates ± standard error mean (n=5 for wild type, n=3 for #14 and #18 lines, and n=2 for #41 and #50 lines genotype). Asterisks indicate significant changes compared to wild type samples as calculated by two tailed Student's t-test (*P-value < 0.05; **P-value < 0.01; ***P-value < 0.001). LC-MS was used for targeted profiling of SGAs.

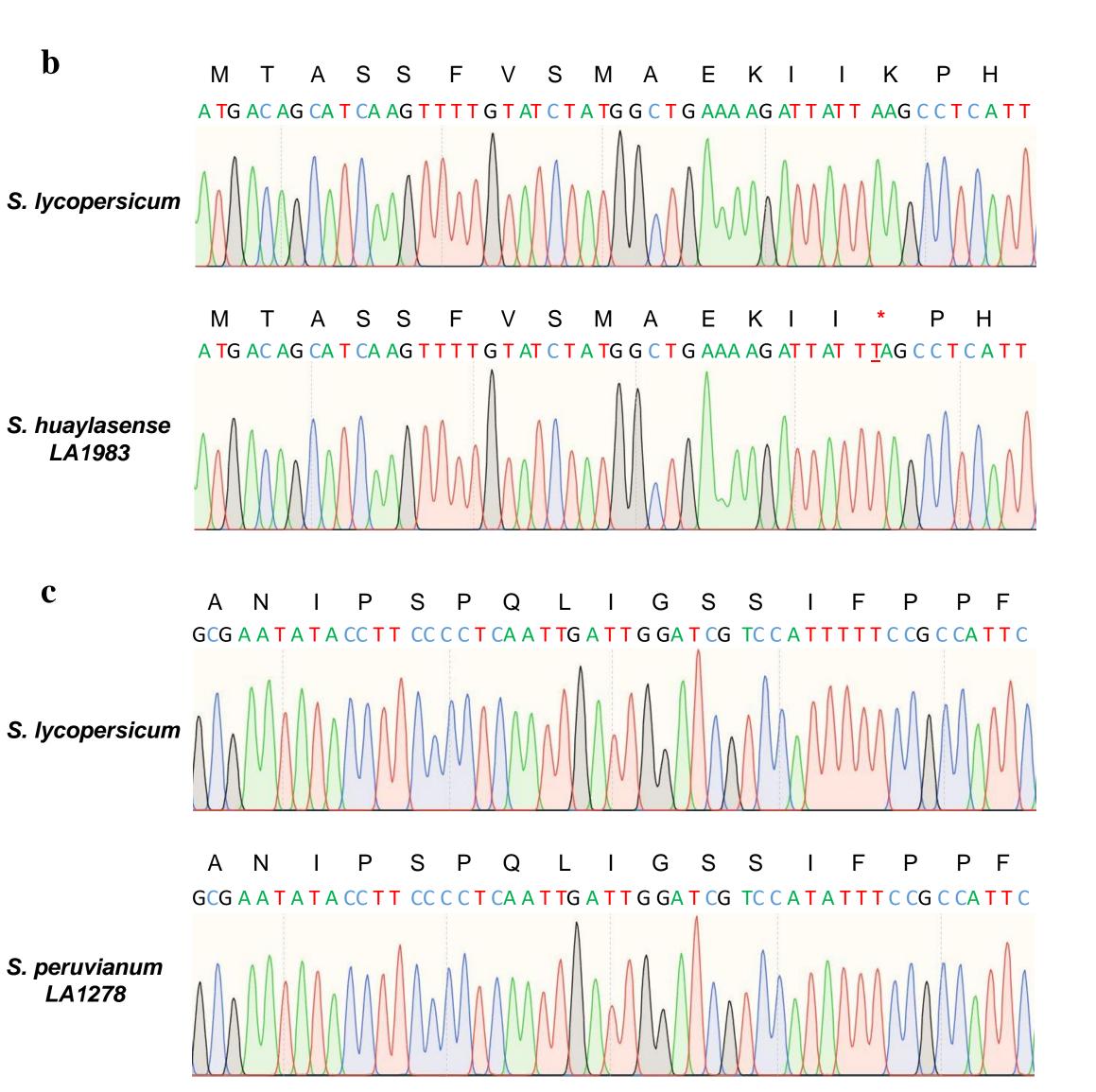


Supplementary Figure 14: Levels of hydroxytomatine isomers in game36 mutant tomato leaves (upper panel) and flowers (lower panel). Knockout of game36 in cultivated tomato resulted in significant accumulation of hydroxytomatine and its isomers. These isomers are typically present in minor quantities in wild type tissues. Four independent (#14, #18, #41 and #50) homozygous game36 mutant lines were generated in tomato (cv. Micro Tom) background. The values indicate means of biological replicates ± standard error mean (n=5 for wild type, n=3 for #14 and #18 lines, and n=2 for #41 and #50 lines genotype). Asterisks indicate significant changes compared to wild type samples as calculated by two tailed Student's t-test (*P-value < 0.05; **P-value < 0.01; ***P-value < 0.001). LC-MS was used for SGAs analysis.

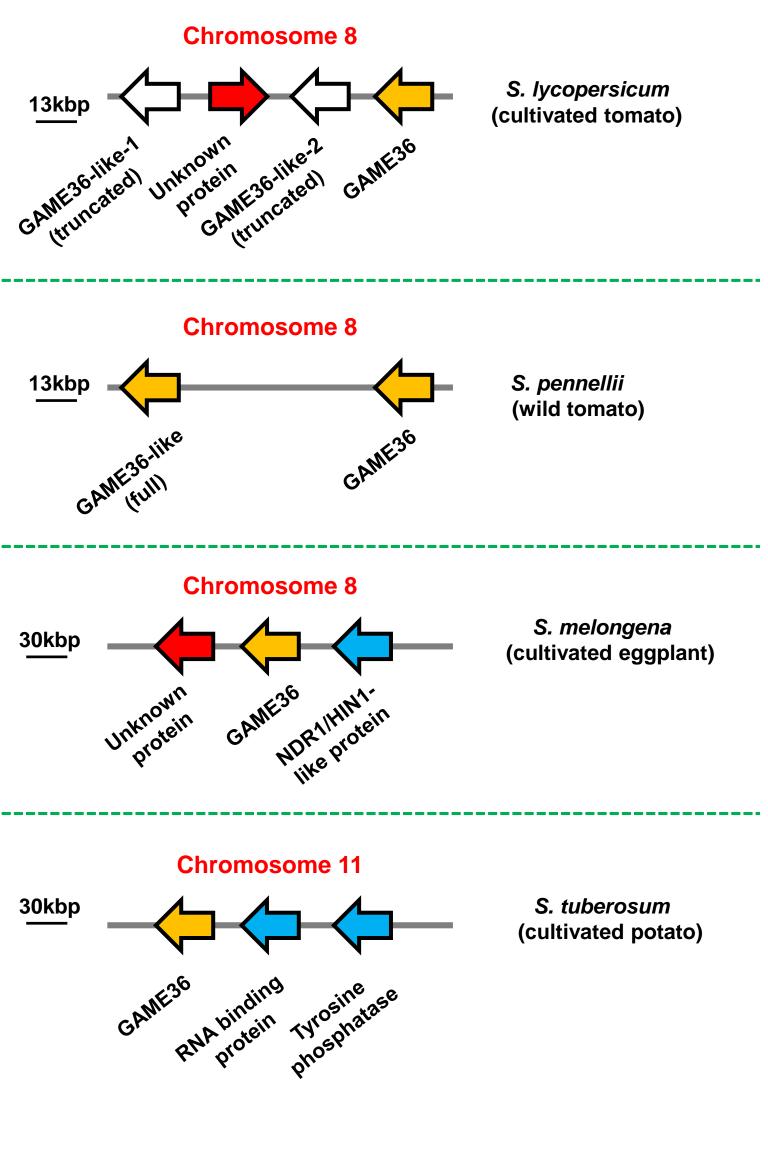


Supplementary Figure 15: Transient co-expression of four tomato GAME genes (SIGAME31, SIGAME36, SIGAME40 and SIGAME5) in *N. benthamiana* enables step-by-step conversion of co**infiltrated** *α*-tomatine substrate into Esculeoside A. Esculeoside A pathway intermediates were observed upon the transient expression of corresponding GAME genes (left panel). Extracted ion chromatograms from LC-MS are shown. Various GAME genes combinations (right panel, shown in black) were included in the transient expression experiments to test the promiscuity of GAME enzymes on Esculeoside A pathway intermediates. Leaves co-infiltrated with empty vector and α -tomatine were used as control (shown in black, left panel). m/z: mass to charge.

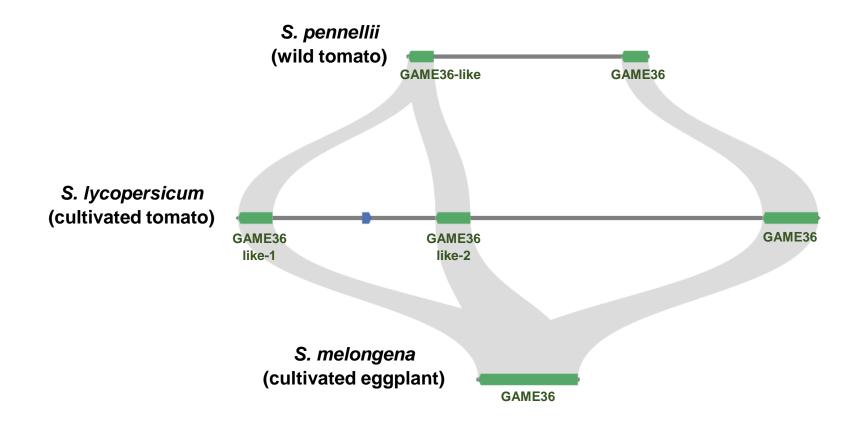




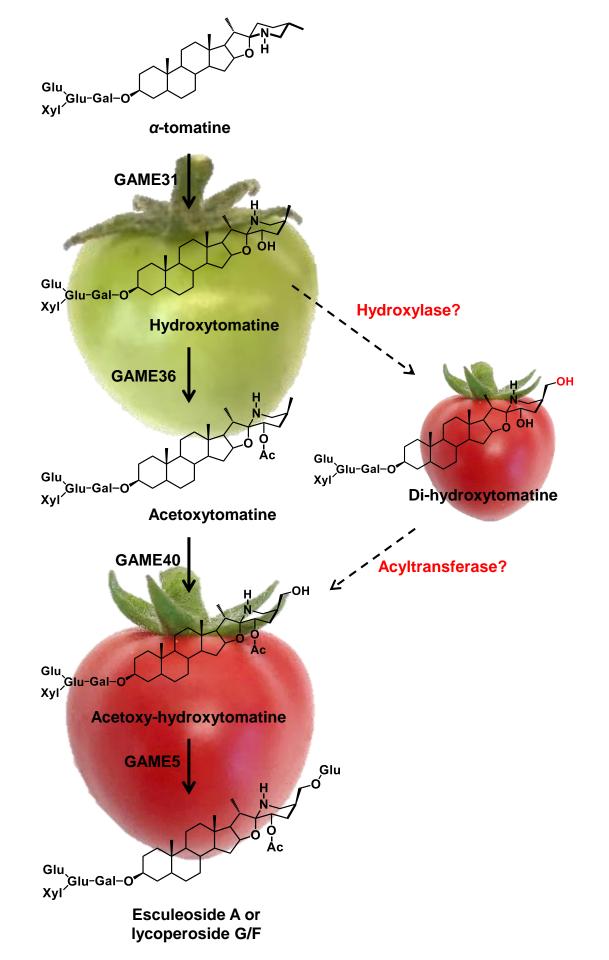
Supplementary Figure 16: GAME36 variant proteins in tomato clade of the genus Solanum. (a) GAME36 genotype variants from cultivated and wild tomato species were obtained from NCBI and other tomato public databases. Two wild tomato accessions, *S. huaylasense* LA1983 and *S. peruvianum* LA1278 were predicted to encode truncated GAME36 proteins due to introduction of premature stop codons in sequence (shown in red dash). As typically observed for BAHD acyltransferase family proteins, GAME36 protein sequences analyzed here also contains two conserved amino acid motifs HKIAD and DFGWG. (b, c) Sanger sequencing results of cloned *GAME36* sequences from *S. huaylasense* LA1983 (b) and *S. peruvianum* LA1278 (c) accessions. Among these two species, nonsense mutation was confirmed only in *S. huaylasense* LA1983, and not in *S. peruvianum* LA1278 as predicted (see panel a) Sequencing chromatograms from wild tomato accessions were compared to cultivated tomato (*S. lycopersicum cv.* Moneymaker) *GAME36* sequence.



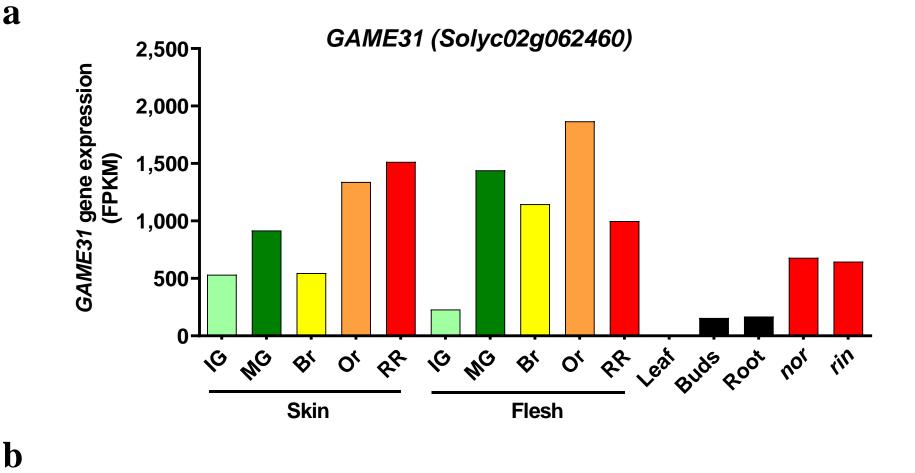
Supplementary Figure 17: Genomic organization of GAME36 and GAME36-like genes associated with SGA metabolism in selected cultivated and wild Solanum species. GAME36 and GAME36-like (full length) are shown in yellow block arrow. Truncated GAME36-like genes (white block arrows), unknown proteins (red block arrow) and other genes (blue block arrow) located next to GAME36 on chromosomal region of respective Solanum species are presented. The arrowheads represent direction of transcription.

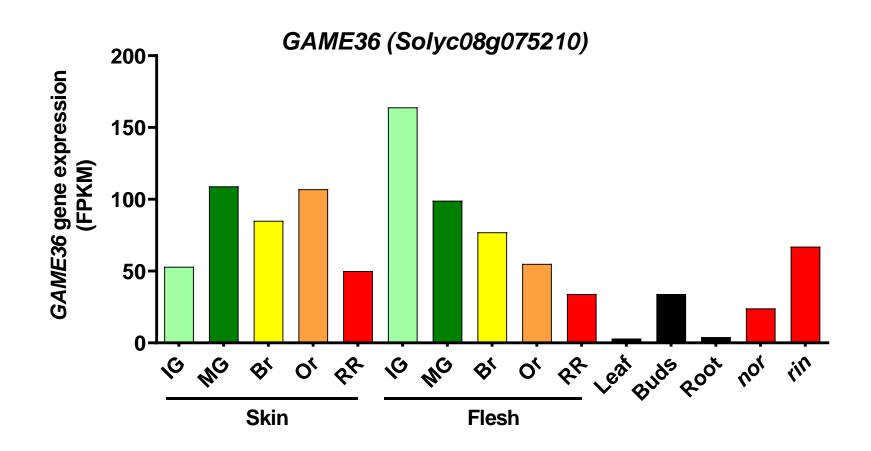


Supplementary Figure 18: Syntenic alignment of GAME36 and GAME36like genes between cultivated tomato, cultivated eggplant and wild *S. pennellii*.

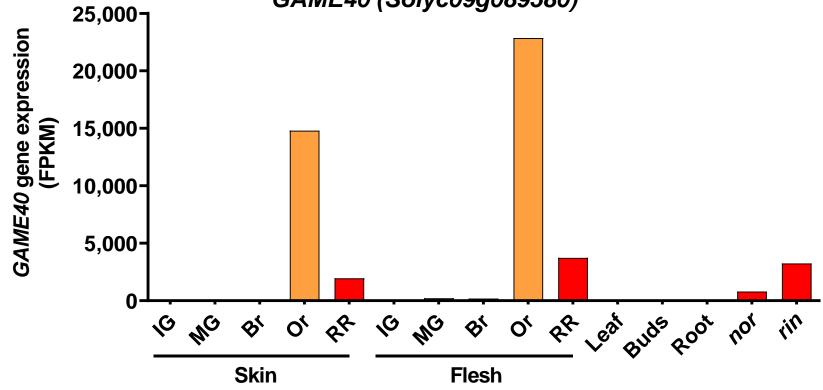


Supplementary Figure 19: Alternative minor biosynthetic route to Esculeoside A formation in tomato red ripe fruit. The major biosynthetic route for Esculeoside A production from α -tomatine involving four GAME enzymes is shown in black arrows. The alternative route involving dibold hydroxytomatine as pathway intermediate is shown in dashed arrows, while proposed enzymes in this pathway are marked in red. GAME: GLYCOALKALOID METABOLISM; Glu: Glucose; Gal: Galactose; Xyl: Xylose; Rha: Rhamnose and Ac: Acetoxy.



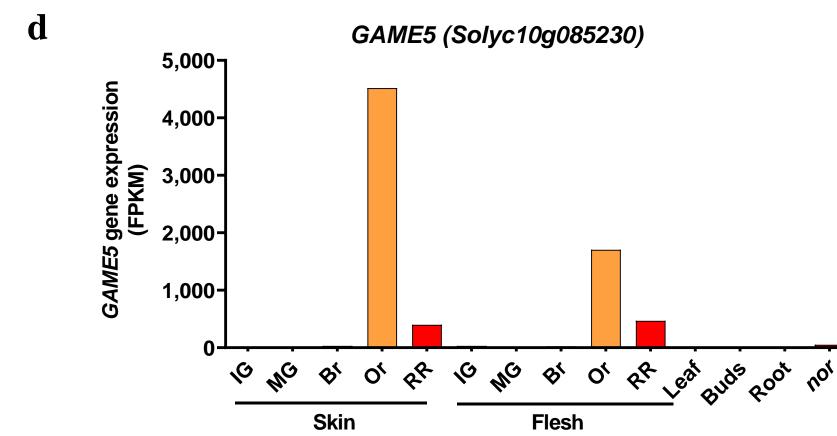






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C



Supplementary Figure 20: Expression of *GAME genes* involved in Esculeoside A pathway. (a-d) Normalized *GAME31* (a), *GAME36* (b), *GAME40* (c), *GAME5* (d) expression level in tomato (*cv.* Micro Tom) tissue types, fruit developmental stages and ripening impaired mutants (RNA-seq expression data). Fruit developmental stages- IG: immature green; MG: mature green; Br: breaker; Or: orange; RR: red ripe. FPKM: Fragments Per Kilobase of transcript per Million mapped reads.