ORIGINAL ARTICLE

Impact of Haemodialysis on Insulin Kinetics of Acute Kidney Injury Patients in Critical Care

Ummu K. Jamaludin · Paul D. Docherty · J. Geoffrey Chase · Geoffrey M. Shaw

Received: 18 February 2014/Accepted: 2 May 2014/Published online: 3 February 2015 © The Author(s) 2015. This article is published with open access at Springerlink.com

Abstract Critically ill patients are occasionally associated with an abrupt decline in renal function secondary to their primary diagnosis. The effect and impact of haemodialysis (HD) on insulin kinetics and endogenous insulin secretion in critically ill patients remains unclear. This study investigates the insulin kinetics of patients with severe acute kidney injury (AKI) who required HD treatment and glycaemic control (GC). Evidence shows that tight GC benefits the onset and progression of renal involvement in precocious phases of diabetic nephropathy for type 2 diabetes. The main objective of GC is to reduce hyperglycaemia while determining insulin sensitivity. Insulin sensitivity (S_I) is defined as the body response to the effects of insulin by lowering blood glucose levels. Particularly, this study used S_I to track changes in insulin levels during HD therapy. Model-based insulin sensitivity profiles were identified for 51 critically ill patients with severe AKI on specialized relative insulin nutrition titration GC during intervals on HD (OFF/ON) and after HD (ON/ OFF). The metabolic effects of HD were observed through changes in S_I over the ON/OFF and OFF/ON transitions. Changes in model-based S_I at the OFF/ON and ON/OFF

U. K. Jamaludin (🖂)

Faculty of Mechanical Engineering, Universiti Malaysia Pahang, 26600 Pekan, Pahang, Malaysia e-mail: ummu85@ump.edu.my

P. D. Docherty · J. Geoffrey Chase Department of Mechanical Engineering, Centre of Bioengineering, University of Canterbury, Private Bag 4800, Christchurch 8140, New Zealand

G. M. Shaw

transitions indicate changes in endogenous insulin secretion and/or changes in effective insulin clearance. Patients exhibited a median reduction of -29 % (interquartile range (IQR): [-58, 6 %], p = 0.02) in measured S_I after the OFF/ON dialysis transition, and a median increase of +9 % (IQR -15 to 28 %, p = 0.7) after the ON/OFF transition. Almost 90 % of patients exhibited decreased S_I at the OFF/ON transition, and 55 % exhibited increased S_I at the ON/OFF transition. Results indicate that HD commencement has a significant effect on insulin pharmacokinetics at a cohort and per-patient level. These changes in metabolic behaviour are most likely caused by changes in insulin clearance or/and endogenous insulin secretion.

Keywords Insulin sensitivity · Tight glycaemic control · Haemodialysis · Intensive care unit · Acute kidney injury

1 Introduction

Acute kidney injury (AKI) is a common complication among critically ill patients, especially for elderly patients with diabetes [1]. Approximately 36 % of critically ill patients are diagnosed with AKI [1–3] with a significant proportion progressing to severe AKI (Stage 3) [2], requiring weekly haemodialysis (HD) [4]. Several epidemiological studies have shown an increase in morbidity and mortality following the development of severe AKI [3, 5–7].

The increasing incidence of critically ill patients with severe AKI may be explained by several factors, including a rising incidence of sepsis [3, 4], major surgery (especially cardiothoracic), nephrotoxic medications, and chronic medical conditions [5]. With both uraemia and HD treatment, glycaemic control (GC) can be complicated [6] as

Department of Intensive Care Christchurch School of Medicine and Health Science, PO Box 4345, Christchurch 8140, New Zealand

GC affects insulin secretion, insulin clearance, gluconeogenesis [7], and peripheral tissue sensitivity of insulin [8]. Many studies have claimed that HD treatment is necessary to treat severe AKI patients by removing waste and toxins [9]. Other clinical studies have shown that HD treatment cleared plasma insulin through increased absorption [10-12] through the dialyzer membrane, lowering insulin concentration. Also, abnormalities in insulin secretion have important pathophysiological implications in the genesis of AKI, which is responsible for the progressive reduction in insulin requirement of diabetic patients who develop AKI [13]. However, the effect of renal failure on insulin kinetics in critically ill patients is still unclear due to the lack of pharmacokinetic studies on insulin secretion and clearance related to HD treatment. These unknown effects might have the potential to complicate metabolic management and the treatment itself.

In particular, insulin resistance is common in many severe AKI patients [4, 14, 15]. Hence, these patients are at risk of developing hyperglycaemia [6] with its associated negative outcomes [16, 17]. The mechanism of glucose intolerance in severe AKI patients is ambiguous [18]. DeFronzo et al. [21] and Mak [20] showed that insulin resistance among severe AKI patients improved during a 10-week course of HD treatment. However, the net effect of HD treatment on glycaemic regulation and insulin sensitivity (S_f) in a critically ill cohort is unknown.

This study uses dense clinical data and a model-based analysis to investigate changes in a clinically validated model-based S_I metric at HD transitions in a cohort of critically ill patients. It was hypothesized that the observed S_I would decrease during HD due to enhanced insulin clearance compared to the model and would be recaptured again when HD is stopped. These changes in model-based S_I would thus offer a unique observation of insulin kinetics and action in this population of critically ill patients with severe AKI that would better inform metabolic care.

2 Materials and Methods

2.1 Patient Cohort

Retrospective blood glucose (*G*) measurements, nutrition administration rates (*P*), and insulin delivery (U_X) data used in this study were obtained from the Specialized Relative Insulin Nutrition Titration (SPRINT) pilot study of 371 critically ill patients who required GC [21]. 51 of the 371 patients had severe AKI treated with HD. The exogenous insulin and nutrition given to these patients were optimized to maximise blood glucose time in the range of 4.0–7.0 mmol L⁻¹, minimising hyperglycaemia, via patient-specific nutrition and insulin administration [12].

The 51 severe AKI patients were treated with HD with a polysulfone (PS) dialyzer membrane (APS-15SA: Asahi Medical Co., Ltd, Tokyo). This PS dialyzer membrane has been reported to affect plasma insulin clearance during HD treatment [22, 23]. Patients were subjected to HD three times weekly (in a fasting state) for a minimum of 4 h in the Christchurch Hospital Intensive Care Unit (ICU).

Study inclusion from 51 severe AKI patients required a minimum of 5 h of patient data before dialysis, followed by at least 6 h of dialysis, and then at least 5 h after dialysis. The clinical details of this cohort are summarized in Table 1. The APACHE III diagnosis for these patients can be divided into 5 main groups: Sepsis, Cardiovascular, Trauma, Respiratory, and Diabetes. Full details on SPRINT can be obtained elsewhere [21].

2.2 Limitation of Study

The study is mainly focused on the (intermittent) HD treatment as one of many treatments in managing severe AKI. Continuous renal replacement therapy (life-supporting treatments) is commonly instituted in critical care when severe AKI is diagnosed. Continuous renal replacement therapy [9] includes:

- (a) HD
- (b) peritoneal dialysis
- (c) hemofiltration
- (d) renal transplantation

All of these treatments are regarded as life-extending treatments to support renal function. With dense clinical data obtained from the SPRINT glycaemic protocol [21], it is suggested for most critically ill patients to be treated with HD in order to avoid other metabolic complications

Table 1 SPRINT cohort baseline variables (N = 51)

	Median	[IQR]
Age (years)	65	46–73
% male	76 %	
APACHE II score	24	19–30
APACHE III Diagnosis	Number of patients	%
Trauma	9	18
Cardiovascular	13	25
Sepsis	20	39
Respiratory	7	14
^a Diabetes	2	4

^a Patients diagnosed with type 1 diabetes mellitus (T1DM)

Data are expressed as median values [IQR] (APACHE = Acute Physiology And Chronic Health Evaluation)

such as sepsis and trauma [15]. Furthermore, the effects of continuous renal replacement inter-therapies (as mentioned above) can be discovered with a larger cohort with different AKI by observing the changes in insulin sensitivity that affects insulin kinetics.

2.3 Identification of Model-Based S_I

Model-based S_I was identified hourly by fitting *G* measurements with estimated endogenous insulin secretion using the ICING (Intensive Control Insulin-Nutrition-Glucose) model [24] with modification. An integral-based method [25] and clinical data were used to identify a patient-specific stepwise S_I profile with a 1 h resolution. The model nomenclatures are given in Table 2. It is mathematically defined as:

•
$$Q(t) = n_I (I(t) - Q(t)) - n_C \frac{Q(t)}{1 + \alpha_G Q(t)}$$
 (2)

$$\begin{split} \mathbf{\hat{I}}(t) &= -n_K I(t) - n_L \frac{I(t)}{1 + \alpha_I I(t)} - n_I (I(t) - Q(t)) + \frac{u_{ex}}{V_I} \\ &+ (1 - x_L) \frac{u_{en}(G)}{V_I} \end{split}$$

$$P_1(t) = -d_1 P_1 + D(t) \tag{4}$$

(3)

$$P_2(t) = -\min(d_2 P_2, P_{\max}) + d_1 P_1$$
(5)

$$P(t) = \min(d_2 P_2, P_{\max}) + PN(t)$$
(6)

$$u_{en}(G) = \begin{cases} u_{\min}, & u_{\min} > k_1 G(t) + k_2 \\ k_1 G(t) + k_2, & u_{\min} \le k_1 G(t) + k_2 \le u_{\max} \\ u_{\max}, & u_{\max} < k_1 G(t) + k_2 \end{cases}$$
(7)

Model estimation of endogenous insulin secretion $(u_{en}(G))$ is in the range of 16.7–266.7 mU min⁻¹ as a function of glycaemic level (G) [26]. This overall metabolic model has been clinically validated with a median prediction error of less than 5 % [27]. The model has been used in several clinical GC trials and insulin sensitivity tests [28, 29].

2.4 Hypothesis

Specifically, constant n_K is used in model ICING describes in Eqs. (1)–(6). While, u_{en} model in Eq. (7) is dependent on *G* values. Therefore, the unmodeled changes due to HD or any other effect [30] are reflected in the model-based S_I . Two dialysis transitions, OFF/ON and ON/OFF, are examined in this study.

2.4.1 OFF/ON Transition

The HD PS membrane is known to absorb plasma insulin during dialysis treatment [23]. Therefore, it was hypothesized that after the OFF/ON transition, S_I will decrease given a model assumption of a fixed u_{en} which dependent on *G* and a constant n_K .

2.4.2 ON/OFF Transition

By using the same dialyzer membrane with the assumptions of a fixed u_{en} (dependable on *G*) and a constant n_K , it was hypothesized that S_I will increase as the plasma insulin level recovers to higher levels after HD treatment ends.

Thus, changes in S_I due to HD might be caused either by:

- (a) changes in u_{en} due to HD treatment [14, 23]
- (b) changes in the effective insulin clearance $(n_K \text{ in the model})$ [31, 32]

However, only changes and the net effect of S_I after both transitions were tracked so the separate effects could not be delineated. In particular, a rising of u_{en} that is based on the assumption of a fixed u_{en} (dependable on *G*) leads to an increase in the observed S_I , while rising n_K leads to an apparent reduction in S_I . If S_I is decreasing, it means that the effect of insulin clearance increases outweighs the effect of u_{en} increases.

2.5 Calculations and Statistical Analysis

Numerical calculations and parameter identification were performed using MATLAB (The MathWorks Inc., Natick, MA). The proportional difference in $S_I(\Delta S_I)$ was calculated as:

$$\Delta S_I = 2 \frac{S_{I(after)} - S_{I(before)}}{\left(S_{I(before)} + S_{I(after)}\right)}$$
(8)

Blood glucose changes, ΔG , were calculated in a manner similar to that for ΔS_I to assess any changes in glycaemia that could affect results.

This analysis uses a 2 h moving average to reduce the effect of measurement error and the influence of transient effects. S_I profiles are identified over periods starting at 3 h before dialysis commencement until 4 h after dialysis ends. This range ensures full settling of patient responses after transitions. The patients' blood glucose and insulin sensitivity at both OFF/ON and ON/OFF dialysis transitions are illustrated on distribution and Bland–Altman plots.

Table 2 Nomenclature of ICING-2 Model

Parameters	Description			Unit
G	Blood glucose level			mmol L^{-1}
Q	Interstitial insulin level			mU L^{-1}
Ι	Plasma insulin level			mU L^{-1}
P_1	Stomach glucose content			mmol
P_2	Gut glucose content			mmol
Р	Rate of glucose appearance in plasma			$mmol min^{-1}$
u _{en}	Endogenous insulin secretion rate			$mU min^{-1}$
Parameters an	nd kinetic values of ICING-2 model based on diabetic status			
EGP	Endogenous glucose production rate	1.16		$mmol min^{-1}$
CNS	Central nervous system glucose uptake	0.3		$mmol min^{-1}$
p_G	Patient endogenous glucose removal	0.006		\min^{-1}
S_I	Insulin sensitivity			$L m U^{-1} m in^{-1}$
α_G	Saturation parameter of insulin-mediated glucose removal	0.0154		$L mU^{-1}$
V_G	Plasma glucose distribution volume	13.3		L
n _I	Plasma-interstitium insulin diffusion rate	0.006		\min^{-1}
n _C	Receptor-bound insulin degradation	0.006		\min^{-1}
n _K	Renal insulin clearance	0.0542		\min^{-1}
n_L	Hepatic insulin clearance	0.1578		\min^{-1}
α_I	Saturation parameter for hepatic insulin clearance	0.0017		$L mU^{-1}$
V_I	Insulin distribution volume	4.0		L
x_L	First pass hepatic clearance	0.67		
d_1	Rate of glucose transport through the enteral route into the bloodstream	0.0347		\min^{-1}
d_2		0.0069		\min^{-1}
P _{max}	Maximal gut glucose flux	6.11		$mmol min^{-1}$
u _{max}	Maximum pancreatic secretion rate	266.7		$mU min^{-1}$
u _{min}	Minimum pancreatic secretion rate	16.7		$mU min^{-1}$
k_I	Pancreatic insulin secretion glucose-sensitivity	*NGT	14.9	mU L $mmol^{-1} min^{-1}$
		*T2DM	4.9	
		*T1DM	0.0	
<i>k</i> ₂	Pancreatic insulin secretion offset	*NGT	-49.9	$mU \cdot min^{-1}$
		*T1DM	16.7	
Exogenous in	put variables of ICING-2 model			
u _{ex}	Intravenous insulin input rate			$mU min^{-1}$
D	Oral glucose input rate from enteral nutrition			$mmol min^{-1}$
PN	Intravenous glucose input rate from parenteral nutrition			$mmol min^{-1}$

* NGT = normal glucose tolerance, T1DM = type 1 diabetes mellitus, T2DM = type 2 diabetes mellitus

Non-parametric Wilcoxon rank sum tests were used to assess ΔS_I and ΔG over the cohort at each transition.

3 Results

Figures 1 and 2 show Bland–Altman plots of ΔG and ΔS_I over the OFF/ON and ON/OFF dialysis transitions. ΔG and ΔS_I at the ON/OFF transition are unbiased overall, but S_I is

biased (median $\Delta S_I = -29$ %, p = 0.02) over the OFF/ ON dialysis transition (Fig. 1b).

At the OFF/ON dialysis transition, the G distribution is effectively maintained within 4–7 mmol L^{-1} except for two patients with sepsis and cardiovascular diagnoses.

Figure 3 shows ΔS_I over 6 h at the OFF/ON and ON/ OFF dialysis transitions. Patients diagnosed with pancreatitis, diabetes, and other metabolic dysfunctions showed larger variance in ΔS_I (>-150 %). However, the trend at

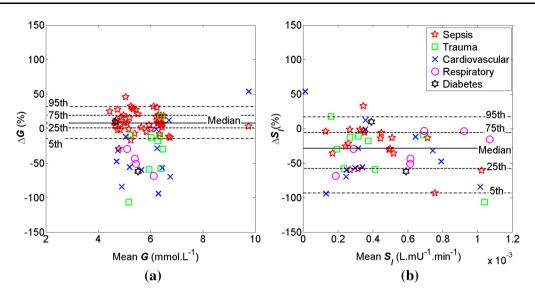
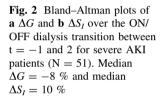
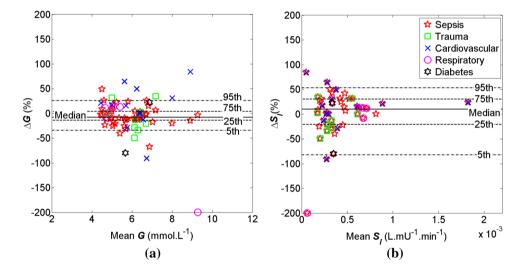


Fig. 1 Bland–Altman plots of **a** ΔG and **b** ΔS_I over the OFF/ON dialysis transition between t = -1 and 2 for severe AKI patients (N = 51). Median $\Delta G = 8$ % and median $\Delta S_I = -29$ %





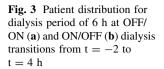
the OFF/ON transition in Fig. 3a (and Fig. 1) is much clearer.

Table 3 summarises ΔS_I over the OFF/ON and ON/OFF transitions. S_I decreased after the OFF/ON dialysis transition until t = 2 h, where it settled with median $\Delta S_I = -29 \%$ (interquartile range (IQR): [-58,6] %, p = 0.02). There were a comparatively low number of confounders, indicating a relatively strong effect. Median ΔS_I increased by 9 % for the ON/OFF transition (Table 3), (IQR: [-15,28] %; p = 0.7) at t = 3 h after the ON/OFF transition. The number of confounders is significantly higher for the ON/OFF transition and the *p* values indicate that the hypothesized effect cannot be confirmed at this transition. *G* remains effectively constant at both transitions (Figs. 1, 2). However, changes in S_I outcomes were not significant (p > 0.05) even 4 h after the ON/OFF transition.

An extended dialysis interval (>10 h) of ΔS_I for both OFF/ON and ON/OFF dialysis transitions across the N = 26 subjects with sufficient data is shown in Table 4. S_I decreased during the OFF/ON dialysis interval until t = 8 h, where it settled to a median reduction of -25 % (IQR: [-10, -51] %; p = 0.04). There were only 2 confounders ($\Delta S_I > 0$) from the 26 patients at t = 8 h. However, while the ON/OFF transition results improved relative to the hypothesized effect, the results were still insignificant (p > 0.07).

4 Discussion

This study investigated the effect of dialysis on renal insulin clearance, endogenous insulin secretion, and



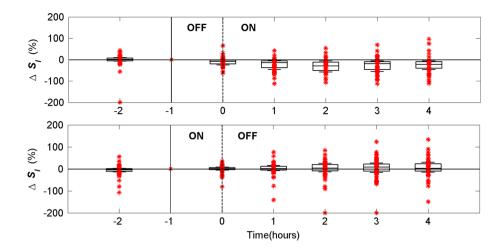


Table 3 Results for OFF/ON and ON/OFF dialysis transitions of 6 h with inverted S_I confounders (t = -2 to t = 4, N = 51)

Time t (hr)	OFF/ON	(N = 51),	expect ΔS_I	< 0		ON/OFF (N = 51), expect $\Delta S_I > 0$						
	Q _I (%)	Q ₂ (%)	Q ₃ (%)	p value	$\Delta S_I > 0 \%$ (confounders)		Q1 (%)	Q ₂ (%)	Q ₃ (%)	p value	$\Delta S_I < 0 \%$ (confounders)	
-2	-7	1	10	0.9	31	61	-13	-4	3	0.7	32	63
-1	0	0	0	1	0	0	0	0	0	1	0	0
0	-24	-7	-1	0.3	10	20	-5	1	8	0.9	24	47
1	-45	-14	-2	0.05	7	14	-7	2	15	0.9	23	45
2	-58	-29	-6	0.02	5	10	-13	4	26	0.8	21	41
3	-55	-19	-5	0.03	9	18	-15	9	28	0.7	23	45
4	-46	-22	-5	0.03	6	12	-15	3	31	0.5	24	47

The results show ΔS_I quartiles and number of confounders with ΔS_I in direction opposite to median trend hypothesized. $Q_2 = \text{median} = 50 \%$ percentiles result. * *p* values measured using Wilcoxon rank sum tests, $Q_1 = 25 \%$ percentile, $Q_3 = 75 \%$ percentile

effective plasma insulin through a clinically validated model-based ΔS_I metric at both OFF/ON and ON/OFF dialysis transitions. Significant insulin sensitivity changes were observed at the OFF/ON dialysis transition (p = 0.02). This analysis indicates that model-based S_I decreased over the initial 4 h after HD started and that the changes occurred as rapidly as 2 h. This result implies that dialysis significantly affects plasma insulin levels via changes in renal insulin clearance and/or endogenous insulin secretion, compared to baseline model assumptions.

Glucose intolerance among critically ill patients with severe AKI occurs with significant inhibition of insulin secretion and a state of peripheral insulin resistance [14, 33] on top of insulin resistance from the critical illness [34]. It has also been reported that in patients with severe AKI, insulin resistance occurred even though glomerular filtration rate (GFR) values were still within the normal range [35]. The effect of insulin resistance can be exacerbated by impairment of the role of insulin in maintaining the hepatic glucose balance [19]. Specifically, an inability of insulin to stimulate hepatic glucose uptake with decreasing S_I has been observed in severe AKI patients [36]. Thus, understanding the pharmacokinetics of insulin during dialysis is clinically important.

Plasma insulin is reduced by enhanced insulin clearance due to the PS dialyzer membrane [22, 23] used during HD treatment in this study. It is suggested that the most significant reduction in plasma insulin during HD treatment is through absorption of insulin across the PS membrane [23], where the equilibrium amount of insulin absorbed was greatest in positively charged membranes [31]. A significant uptake and degradation of insulin may occur when renal insulin clearance significantly exceeds the glomerular filtration rate [6], as would occur in HD treatment. This enhanced insulin clearance rate and accumulation of dialyzable uraemic toxins can cause inhibition of insulin degradation and can be sufficiently normalized by HD treatment [20].

Plasma insulin levels also depend on endogenous insulin secretion. Physiologically, u_{en} is determined by the

Table 4 Extended results for OFF/ON and ON/OFF dialysis transitions of >10 h with inverted S_I confounders (t = -2 to t = 10, N = 26)

Time t (hr)	OFF/ON	N(N=26)	6), expect	$\Delta S_I < 0$			ON/OFF (N = 26), expect $\Delta S_I > 0$					
	Q _I (%)	Q ₂ (%)	Q ₃ (%)	p value	$\Delta S_I > 0 \%$ (confounders)		Q ₁ (%)	Q_2 (%) Q_3 (%)		p value	$\Delta S_I < 0 \%$ (confounders)	
-2	-4	3	13	0.9	16	62	-15	-6	1	0.7	19	73
-1	0	0	0	1	0	0	0	0	0	1	0	0
0	-25	-10	-2	0.4	4	15	-5	3	9	0.8	10	28
1	-44	-23	-4	0.1	3	12	-7	8	17	0.8	11	42
2	-56	-30	-6	0.09	2	8	-8	13	32	0.6	9	35
3	-55	-19	-2	0.09	6	23	-4	18	42	0.3	9	35
4	-53	-24	-4	0.1	4	15	-13	14	36	0.3	10	28
5	-53	-29	-8	0.08	4	15	-12	22	44	0.2	10	28
6	-44	-25	-8	0.1	4	15	-6	23	47	0.1	8	31
7	-40	-21	-9	0.07	3	12	-5	18	50	0.07	7	27
8	-51	-25	-10	0.04	2	8	-5	19	39	0.07	7	27
9	-47	-18	-7	0.04	1	4	-6	22	38	0.07	8	31
10	-45	-12	-3	0.09	5	19	-4	22	41	0.08	8	31

The results show ΔS_I quartiles and number of confounders with ΔS_I in direction opposite to median trend hypothesized. $Q_2 = \text{median} = 50 \%$ percentile result. * *p* values measured using Wilcoxon rank sum tests, $Q_1 = 25 \%$ percentile, $Q_3 = 75 \%$ percentile

glycaemic level and the ability of β cells to respond to blood glucose level and its changes. However, it was suggested that an increase in endogenous insulin secretion may occur in response to HD treatment with a PS membrane dialyzer due to reductions in plasma insulin [10, 22]. In particular, a PS membrane can reduce plasma insulin significantly in HD [10, 23]. Thus, S_I is also expected to decrease with an unmodeled increase in u_{en} during the initial period of HD treatment to maintain the *G* level.

The model-based ΔS_I at the ON/OFF dialysis transition in this study was insignificant (p > 0.05). It is assumed that acute intravenous (i.v.) administration of 1,25-dihydroxyvitamin D_3 (1,25(OH)₂ D_3) given to severe AKI patients during HD may increase insulin secretion and reverse glucose intolerance [20]. An improvement in glucose metabolism has been observed in some studies via lower mean glucose during dialysis and a more rapid disappearance rate of glucose in the immediate post-dialysis period [37]. In general, glucose metabolism and renal function are expected to increase gradually after postdialysis when toxic substances that are suspected of hindering renal function have been extracted. Long-term (4.9 weeks) HD treatment has been shown to normalize insulin sensitivity and result in a marked improvement in glucose metabolism [11], but this might not completely normalize glucose utilization [19]. Overall, it is impossible to delineate the effects that contribute to changes in S_I in this study, due to model identifiability issues [16] and the side effects of other diagnosed critical illnesses apart from severe AKI [18]. Over longer intervals, as in Table 4, interpatient or intra-patient variation may further obscure the observation of the effect itself [28].

Thus, a substantial change in S_I at the OFF/ON dialysis transition indicates a strong and fast process of the cleaning and clearing of toxic substances from blood, improving effective S_I due to either decreased u_{en} or increased n_K clearance. However, at the ON/OFF dialysis transition, the recovery process to regulate and normalize blood is a lot slower physiologically. Hence, the modelbased S_I after dialysis may be expected to remain unchanged, as observed here, even for extended periods after HD treatment.

The model-based S_I is an indication of overall glucose metabolism of critically ill patients and does not necessarily reflect the precise cellular physiology of peripheral insulin sensitivity. The model-based ΔS_I at a cohort level used in this study are unlikely to be caused by actual variance in true peripheral S_I at a cellular level. In particular, there is no apparent stimulus induced by HD to directly affect S_I . Thus, ΔS_I reflects changes in renal clearance or/and endogenous insulin secretion, which in turn result in changes in the model-based S_I calculated based on fixed assumptions for these values.

In particular, the ICING model prediction of u_{en} is made in terms of blood glucose level in the absence of direct measurement of C-peptide. Hence, the effect of dialysis on u_{en} cannot be defined patient-specifically by the model without added data that was not available in this study. Alternatively, it has been reported that endogenous insulin secretion is also affected by exogenous insulin [38]. As plasma insulin levels are suspected to decrease during dialysis, it is suspected that endogenous insulin secretion would increase at a cohort level. Thus, an increase in the model-based S_I over time should be observed at the OFF/ ON transition, contrary to the observations here. Hence, u_{en} dependence on blood glucose level would confound the observed effect and can be discounted as a contributor to the shifts in the model-based S_I in this study. Thus, it is most likely that effective renal clearance increases during HD and decreases after HD treatment.

Effective u_{en} identification cannot be undertaken with only glucose data [16, 39]. Thus, a direct measurement of C-peptide should be included for direct quantification of the effects contributing to the glycaemia of severe AKI patients. The results of this study could be used to confirm these results in order to power a further study that segregates these potential contributing effects.

All critically ill patients with severe AKI in this study were undergoing SPRINT tight glycaemic control (TGC), where the minimal changes in G illustrated that SPRINT was successful in controlling glycaemia during these transitions. A consistent trend of ΔG below the 5th percentile was observed for patients with cardiovascular, respiratory, and trauma compared to the patient diagnosed with sepsis, as shown in Fig. 1a. It portrays sepsis complications due to severe infections and multiple organ dysfunctions [40], which were not the main focus in this study. Thus, glycaemic levels and the tightness of this protocol are used to ensure that the analysis results are not biased by variations in glucose levels which can affect stress response and thus S_I levels. SPRINT clinical results were also compared with the simulation results to validate and minimize the prediction errors of the protocol. Although there were two diagnosed diabetics, almost no bias in ΔG was observed. Thus, the confounding factor plays no role.

Overall, this investigation suggests that the most likely contributor to the observed changes in S_I was the HD insulin clearance, which was modeled by the renal insulin clearance term. The effect of HD on plasma insulin and the mechanism of insulin clearance among critically ill patients with severe AKI were shown in this study to be a contributor to overall effective S_I , which determines the glycaemic level, all else being equal. However, further indepth studies must be undertaken to measure the specific effects of HD in different AKI stages. A prospective cohort and clinical studies with direct insulin and C-peptide assays on this cohort may lead to a better understanding of insulin kinetics during HD treatment. A broad comparison from a different cohort of varied HD duration with mixed levels of insulin will also clarify the effects of ΔS_I , revealing further details in the underlying contributors of specific insulin resistance.

5 Conclusion

The distinct change in model-based insulin sensitivity during HD treatment was a significant and observable aspect of critically ill patient physiology. The findings were consistent with the presence of effects of HD treatment in a majority of severe AKI patients from other studies. Clinically, the effect of the main contributors (n_K and u_{en}) to effective insulin sensitivity changes during HD from a baseline model or clinical assumptions suitable for other patients should also be considered in GC. However, the precise pharmaco-kinetics/dynamics driving this change remain ambiguous. These results justify a larger cohort investigation with specific measurement of insulin secretion and renal clearance to differentiate these impacts.

Acknowledgments This work was performed with collaboration from the Department of Intensive Care Christchurch School of Medicine and Health Sciences, University of Otago, and the Department of Mechanical Engineering, Centre of Bioengineering, University of Canterbury.

Open Access This article is distributed under the terms of the Creative Commons Attribution License which permits any use, distribution, and reproduction in any medium, provided the original author(s) and the source are credited.

References

- Thomas, G., Rojas, M. C., Epstein, S. K., Balk, E. M., Liangos, O., & Jaber, B. L. (2007). Insulin therapy and acute kidney injury in critically ill patients—a systematic review. *Nephrology*, *Dialysis, Transplantation*, 22, 2849–2855.
- Mactier, R., Hoenich, N., & Breen, C. (2011). Renal association clinical practice guideline on haemodialysis. *Nephron Clinical Practice*, 118, c241–c286.
- Bagshaw, S. M., Uchino, S., Bellomo, R., Morimatsu, H., Morgera, S., Schetz, M., et al. (2007). Septic acute kidney injury in critically ill patients: Clinical characteristics and outcomes. *Clinical Journal of the American Society of Nephrology*, 2, 431–439.
- Midtvedt, K., Hartmann, A., Hjelmesaeth, J., Lund, K., & Bjerkely, B. L. (1998). Insulin resistance is a common denominator of post-transplant diabetes mellitus and impaired glucose tolerance in renal transplant recipients. *Nephrology, Dialysis, Transplantation, 13*, 427–431.
- Hoste, E. A. J., Lameire, N. H., Vanholder, R. C., Benoit, D. D., Decruyenaere, J. M. A., & Colardyn, F. A. (2003). Acute renal failure in patients with sepsis in a surgical ICU: Predictive factors, incidence, comorbidity, and outcome. *Journal of the American Society of Nephrology*, 14, 1022–1030.
- Mak, R. H. K. (2000). Impact of end-stage renal disease and dialysis on glycemic control. *Seminars in Dialysis*, 13, 4–8.
- Van den Berghe, G., Wouters, P., Weekers, F., Verwaest, C., Bruyninckx, F., Schetz, M., et al. (2001). Intensive insulin therapy in the critically ill patients. *New England Journal of Medicine*, 345, 1359–1367.
- Shrishrimal, K., Hart, P., & Michota, F. (2009). Managing diabetes in hemodialysis patients: Observations and recommendations. *Cleveland Clinic Journal of Medicine*, 76, 649–655.

- Abe, M., Kaizu, K., & Matsumoto, K. (2007). Plasma insulin is removed by hemodialysis: Evaluation of the relation between plasma insulin and glucose by using a dialysate with or without glucose. *Therapeutic Apheresis*, 11, 280–287.
- Kobayashi, S., Maejima, S., Ikeda, T., & Nagase, M. (2000). Impact of dialysis therapy on insulin resistance in end-stage renal disease: Comparison of haemodialysis and continuous ambulatory peritoneal dialysis. *Nephrology, Dialysis, Transplantation*, 15, 65–70.
- Stenvinkel, P., Bolinder, J., & Alvestrand, A. (1992). Effects of insulin on renal haemodynamics and the proximal and distal tubular sodium handling in healthy subjects. *Diabetologia*, 35, 1042–1048.
- Mak, R. H. K., & De Fronzo, R. A. (1992). Glucose and insulin metabolism in uremia. *Nephron*, 61, 377–382.
- Mak, R. H. K. (1995). Insulin secretion and growth failure in uremia. *Pediatric Research*, 38, 379–383.
- 15. Cruz, D. N., Ferrer-Nadal, A., Piccinni, P., Goldstein, S. L., Chawla, L. S., Alessandri, E., Belluomo Anello, C., Bohannon, W., Bove, T., Brienza, N., Carlini, M., Forfori, F., Garzotto, F., Gramaticopolo, S., Iannuzzi, M., Montini, L., Pelaia, P., Ronco, C., Investigators, F. T. N. (2014). Utilization of small changes in serum creatinine with clinical risk factors to assess the risk of AKI in critically Ill adults. *Clinical Journal of the American Society of Nephrology*, 9, 663–672.
- Docherty, P. (2011). Evaluation and development of the dynamic insulin sensitivity and secretion test for numerous clinical applications. PhD Thesis, Mechanical Engineering, University of Canterbury, Christchurch, New Zealand.
- Krinsley, J. S. (2003). Association between hyperglycemia and increased hospital mortality in a heterogeneous population of critically ill patients. *Mayo Clinic Proceedings*, 78, 1471–1478.
- Hampers, C. L., Soeldner, J. S., Doak, P. B., & Merrill, J. P. (1966). Effect of chronic renal failure and hemodialysis on carbohydrate metabolism. *Journal of Clinical Investigation*, 45, 1719–1731.
- DeFronzo, R. A., Alvestrand, A., Smith, D., Hendler, R., Hendler, E., & Wahren, J. (1981). Insulin resistance in uremia. *Journal of Clinical Investigation*, 67, 563–568.
- Mak, R. H. K. (1994). Renal disease, insulin resistance and glucose intolerance. *Diabetes*, 2, 19–28.
- Chase, J. G., Shaw, G. M., Le Compte, A., Lonergan, T., Willacy, M., Wong, X. W., et al. (2008). Implementation and evaluation of the SPRINT protocol for tight glycaemic control in critically ill patients: A clinical practice change. *Critical Care*, 12, R49.
- Jaber, B. L., Cendoroglo, M., Balakrishnan, V. S., Perianayagam, M. C., Karsou, S. A., Ruthazer, R., et al. (2000). Impact of dialyzer membrane selection on cellular responses in acute renal failure: A crossover study. *Kidney International*, 57, 2107–2116.
- Abe, M., Okada, K., Ikeda, K., Matsumoto, S., Soma, M., & Matsumoto, K. (2011). Characterization of insulin adsorption behavior of dialyzer membranes used in hemodialysis. *Artificial Organs*, 35, 398–403.
- 24. Lin, J., Parente, J. D., Chase, J. G., Shaw, G. M., Blakemore, A. J., LeCompte, A. J., et al. (2011). Development of a model-based clinical sepsis biomarker for critically ill patients. *Computer Methods and Programs in Biomedicine*, 102, 149–155.
- Hann, C. E., Chase, J. G., Lin, J., Lotz, T., Doran, C. V., & Shaw, G. M. (2005). Integral-based parameter identification for longterm dynamic verification of a glucose-insulin system model. *Computer Methods and Programs in Biomedicine*, 77, 259–270.

- Pretty, C. (2012). "Analysis, classification and management of insulin sensitivity variability in a glucose-insulin system model for critical illness," PhD, Mechanical Engineering Department, University of Canterbury, Christchurch.
- Lin, J., Razak, N. N., Pretty, C. G., Le Compte, A., Docherty, P., Parente, J. D., et al. (2011). A physiological intensive control insulin-nutrition-glucose (ICING) model validated in critically ill patients". *Computer Methods and Programs in Biomedicine*, 102, 192–205.
- Pretty, C., Le Compte, A., Chase, J. G., Shaw, G. M., Preiser, J., Penning, S., & Desaive, T. (2012). Variability of insulin sensitivity during the first 4 days of critical illness: Implications for tight glycaemic control. *Annals Intensive Care*, 2, 17.
- Penning, S., Le Compte, A., Moorhead, K., Desaive, T., Massion, P., Preiser, J., Shaw, G. M., Chase, J. G. First pilot trial of the STAR-liege protocol for tight glycemic control in critically ill patients. *Computer Methods Programs Biomed.*, "to be published".
- Jamaludin, U. K., Docherty, P. D., Chase, J. G., Le Compte, A., Shaw, G. M., Desaive, T., & Preiser, J. (2012). Observation of incretin effects during enteral feed transitions of critically ill patients. *e-SPEN Journal*, 7, e154–e159.
- Zhang, S., Saito, K., Matsumoto, H., Minagawa, M., & Tanioka, A. (2008). Characterization of insulin adsorption behavior on amphoteric charged membranes. *Polymer Journal*, 40, 837–841.
- 32. Abe, M., Okada, K., Maruyama, T., Ikeda, K., Kaizu, K., & Matsumoto, K. (2009). Comparison of the effects of polysulfone and polyester-polymer alloy dialyzers on glycemic control in diabetic patients undergoing hemodialysis. *Clinical Nephrology*, 71, 514–520.
- DeFronzo, R. A., Tobin, J. D., Rowe, J. W., & Andres, R. (1978). Glucose intolerance in uremia: Quantification of pancreatic beta cell sensitivity to glucose and tissue sensitivity to insulin. *Journal* of Clinical Investigation, 62, 425–435.
- Basi, S., Pupim, L. B., Simmons, E. M., Sezer, M. T., Shyr, Y., Freedman, S., et al. (2005). Insulin resistance in critically ill patients with acute renal failure. *American Journal of Physiology Renal Physiology*, 289, F259–F264.
- Fliser, D., Pacini, G., Engelleiter, R., Kautzky-Willer, A., Prager, R., Franek, E., & Ritz, E. (1998). Insulin resistance and hyperinsulinemia are already present in patients with incipient renal disease. *Kidney International*, 53, 1343–1347.
- Valera Mora, M. E., Scarfone, A., Calvani, M., Greco, A. V., & Mingrone, G. (2003). Insulin clearance in obesity. *Journal of the American College of Nutrition*, 22, 487–493.
- Alfrey, A. C., Sussman, K. E., & Holmes, J. H. (1967). Changes in glucose and insulin metabolism induced by dialysis in patients with chronic uremia. *Metabolism, Clinical and Experimental*, 16, 733–740.
- Argoud, G. M., Schade, D. S., & Eaton, R. P. (1987). Insulin suppresses its own secretion in vivo. *Diabetes*, 36, 959–962.
- 39. Docherty, P., Chase, J. G., Lotz, T., Hann, C. E., Shaw, G. M., Berkeley, J. E., et al. (2009). DISTq: An iterative analysis of glucose data for low-cost, real-time and accurate estimation of insulin sensitivity. *Open Medicine Information Journal*, *3*, 65–67.
- 40. Bone, R. C., Balk, R. A., Cerra, F. B., Dellinger, R. P., Fein, A. M., Knaus, W. A., et al. (1992). Definitions for sepsis and organ failure and guidelines for the use of innovative therapies in sepsis. The ACCP/SCCM consensus conference committee. American college of chest physicians/society of critical care medicine. *Chest*, 101, 1644–1655.