REVIEW



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The claudin-transcription factor signaling pathway

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ABSTRACT

Claudins (CLDNs) represent major transmembrane proteins of tight junctions and contribute to the barrier function. They also serve as anchors for several signaling proteins, but the underlying molecular basis has yet to be established. The present review covers the recent progress in our understanding of the CLDN signaling pathway in health and disease. We discuss the functional relevance of phosphotyrosine motifs in the C-terminal cytoplasmic domain of CLDNs and define mutual regulation between CLDNs and Src-family kinases (SFKs). In addition, we focus on the crosstalk between CLDN and transcription factor signaling. We also describe how aberrant CLDN-transcription factor signaling promotes or inhibits cancer progression. We propose that a link between various cell adhesion molecules and transcription factors coordinates a range of physiological and pathological events via activation or suppression of target genes.

Introduction

The claudin (CLDN) family is capable of forming tight-junction strands and thereby represents the backbone of tight junctions in vertebrate epithelial and endothelial cells, as well as in other types of cells.^{1,2} It is composed of 27 members in mammals, and a specific combination of CLDNs is expressed in a given cell/tissue type. CLDNs are tetraspan transmembrane proteins that include two extracellular loops (EC1 and EC2) and N- and C-terminal cytoplasmic domains. The CLDN-EC1 creates paracellular barriers or pores for selective ions and solutes,³⁻⁷ whereas both CLDN-EC1 and CLDN-EC2 contribute to cis- and transinteractions between CLDNs.⁷⁻¹¹ On the other hand, the C-terminal cytoplasmic domain of many CLDN subtypes contains specific sequence motifs such as PDZ domain-binding motifs and phosphorylation consensus sites^{5, 12–16} and receives or propagates a magnitude of intracellular signals as platforms; however, it remains poorly defined how CLDN signaling reaches the nucleus and regulates gene expression.¹⁷

CLDNs are absolutely required for human health, and their dysregulations are involved in the pathogenesis of diverse diseases.^{4-6,18-16} For instance, it is well known that mutations in several *CLDN* genes cause ARTICLE HISTORY

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various human hereditary diseases. In addition, CLDNs frequently show aberrant expression and/or localization in a wide variety of cancers, resulting in either promotion or repression of tumor progression, most probably by dysregulated CLDN signaling.^{22,26–}

³² Moreover, recent studies have established the region-selective CLDN5 breakdown in brain disorders, such as schizophrenia, depression, Alzheimer's disease, and multiple sclerosis.^{33–39} In this respect, we and others have previously reported how endothelial CLDN5 expression is regionally disrupted in these psychiatric disorders.^{40,41}

In the current review, we focus on the link between CLDN and transcription factor signaling because it is theoretically attributed to the organization of a broad range of cellular processes, including cell growth, survival, differentiation, polarity, migration, and metabolism, via regulation of the expression of corresponding target genes. We also discuss aberrant CLDN-transcription factor signaling in cancer. We do not describe recent progress in our understanding of numerous aspects of other tight-junction players, such as junctional adhesion molecules (JAMs), tight junction-associated MARVEL domain-containing proteins (TAMPs: occludin, tricellulin, and MarvelD3), and the ZO family of scaffolding proteins.42-55

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The CLDN/SFK/PI3K/AKT/transcription factor signaling cascade

We previously reported that retinoid X receptor a (RXR α)/retinoic acid receptor γ (RAR γ) and another member of the nuclear receptor superfamily, hepatocyte nuclear factor 4 α (HNF4 α), trigger the formation of mature cell-cell junctions and microvilli, expression of tight-junction markers (CLDN6, CLDN7, occludin, and ZO-1 α + variant) and a microvilli marker (ezrin/radixin/moesinbinding phosphoprotein 50 [EBP50]), as well as morphological differentiation into epithelial cells from stem cells.^{56–59} These effects are incredibly similar to the CLDN6-triggered ones,⁶⁰ implying a possible crosstalk between them. Along this line, we have recently identified the CLDN–transcription factor signaling pathway.⁶¹

Reciprocal regulation between CLDNs and SFKs

First, we showed, by using the corresponding deletion mutants, that the CLDN6-adhesion signal is transduced through the EC2 domain and the first half C-terminal cytoplasmic domains but not through the EC1 domain (Figure 1). Second, we paid attention to the four tyrosine residues in the C-terminal domain of CLDN6, which are completely conserved among vertebrates, and revealed that Y196/200, but not Y213/218, are definitely required for CLDN6signaling ability. Third, we disclosed that CLDN6 recruits and activates Src-family kinases (SFKs) in EC2-dependent and Y196/200dependent manners, and SFKs in turn phosphorylate CLDN6 at Y196/200 in an EC2-dependent fashion. The functional relevance of the EC2 in the CLDN6-adhesion signal was also supported by using the C-terminal half of Clostridium perfringens enterotoxin (C-CPE).^{60,61} Moreover, we have identified SFK members associated with CLDN6 and obtained evidence showing that recombinant proteins corresponding to the Src homology 2 (SH2) domains of certain SFKs directly bind to the C-terminal cytoplasmic of CLDN6 and the Y196/200domain containing peptide (our unpublished results).

Through a careful search for amino acid sequence of human and mouse CLDN1-20, both

Y196 and Y200 in the C-terminal cytoplasmic domain of CLDN6 are conserved in CLDN2/4/12 (Figures 2 and 3). It is also noteworthy that CLDN6Y196 and CLDN6Y200 are conserved in CLDN3/7/8/10/14/16 and CLDN1/5/9/17/18, respectively.

We have found mutual regulation between certain CLDN subtypes (tyrosine residues corresponding to CLDN6Y200) and SFKs (our unpublished results). In addition, Li et al.⁶² have recently reported that CLDN11 is phosphorylated at two adjacent tyrosine residues, which are positioned at different sites from CLDN6Y196/200, as described above, and activates SRC. The C-terminal domain of CLDN1 is also associated with SRC although the involvement of the pY motifs is not determined.⁶³ It should also be noted that amino acids around pY, in particular 3-5 residues at the C-terminal side, influence the binding specificity of the SH2 domain.⁶⁴⁻⁶⁷ For instance, the C-terminal amino acids of the CLDN6Y200-corresponding tyrosine residue in human and mouse CLDN18 are very different from others; therefore, it does not seem to function as the pY motif. CLDN18 decrease PI3K and AKT known is to activities,68,69 suggesting this notion. Further studies are required to determine whether other CLDN subtypes, similar to CLDN6, couple with SFKs via the pY motifs in the C-terminal cytoplasmic domains.

SFKs are known to be activated by several cellcell and cell-matrix adhesion proteins lacking intrinsic kinase activity, such as E-cadherin, integrins, and cellular prion protein.⁶²,^{70–76}, In addition, engagement of JAMs, the JAML (junctional adhesion molecule-like) and CAR (coxsackie and adenovirus receptor), stimulates PI3K,⁷⁷which is the major downstream signal of SFKs. Besides, it is well known that certain signaling proteins, which contain the SH2, phosphotyrosine-binding (PTB), Hakaityrosine binding (HYB), C2, and pyruvate kinase M2 domains, bind to the pY motifs.^{66,67,78–81} Taken together, these findings strongly suggest that the pY motifs in the C-terminal cytoplasmic domains of diverse cell-cell and cell-matrix adhesion molecules generally serve as the signaling landscapes for SFKs and other pY motif-binding proteins.



Figure 1. Schematic model for regulation of the nuclear receptor activity by the CLDN–adhesion signaling. The schema is modified from that reported previously (Sugimoto et al., 2019). SH2/3: Src homology 2/3 domain; Kinase: kinase domain; AF1: activation function-1; DBD: DNA-binding domain; LBD: ligand-binding domain; RARE: retinoic acid response element; ERE: estrogen response element; RA: yellow circle; estrogen: pink circle.

196 200

CLDN6	CCTCPSGGSQGPSHYMARYSTSAPAISRGPSEYPTKNYV	
CLDN2	CFSCSSQRNRSN¥YDA¥QAQPLATRSSPNSYSLTGYV ^RPGQPPKVK	SEF
CLDN4	CCNCPPR-TDKPYSAKYSAARSAAASNYV	
CLDN12	YCTCKSLPSPSMHT <mark>Y</mark> SQPYSARPR-LSAIEIDIPVVSHT PFWQPLYSHP [^]	
CLDN1	CCSCPRKTTSYPTPRPYPKPAPSSGKDYV	
CLDN5	CCGAWVCTGRPDLSFPVK <mark>Y</mark> S-APRRPTATGDYDKK-NYV	
CLDN9	CCTCPPPQVERPRGPRLGYSIPSRS-GASGLDKRDYV	
CLDN17	CGFCCCNRKQGYRYPVPG <mark>Y</mark> RVPHTDKRRNTTMLSSTSYV K^ ^KT	
CLDN18	CIACRGLAPEETNYKAVS <mark>Y</mark> HASGHSVAYKPGYPSKHDYV ^GFKASTGF	GSNTKNKK
CLDN3	CCSCPPREKKYTVV <mark>Y</mark> SAPRSTGPGASLGTGYDRKDYV ATK^	IYDGGAR TEDEVQS
CLDN7	SCSCPGNESKAG <mark>Y</mark> RVPRSYPKSNSSKEYV	
CLDN8	CCVCCNEKSSSYR- Y SIPSHRTTQKSYHTPSVYSRSQYV F^ GKKS	
CLDN10	CFSISDNNKTPRYT <mark>Y</mark> NGATSVMSSRTKYHGGEDFKTTYV ^P	SKQFDKNAN
CLDN14	PYRPY CLSCQDEAYQPPAA <mark>Y</mark> KDNRAPSVTSATHPAYSGYRNDYV QAPPRATTTTANTAPA^ L	
CLDN16	CCLFKDVGPERNYP <mark>Y</mark> SLRKAYSAAGVSMAPRTETAKMYA YL^ ^KSYSA	
CLDN11	LCCAGDAQAFGENRFYYTAGSSSPTHAKSAHV-	
CLDN15	CSACCCGSDEDPAASARRVATSDQEGDSSFGKYGRNAYV	
CLDN19	CCTCPEPERPNSSPQPYRPGPSAAAREKLPASAKGPLGV ^PVV	
CLDN20	CTSCIKRNPEARLDPPTQQPISNTQLENN-STHNLKDYV	

Figure 2. Amino acid sequences of a part of the C-terminal cytoplasmic domain in human CLDN1-20. Tyrosine residues corresponding to CLDN6Y196/200 are highlighted and the conserved ones are indicated in red.

A link between CLDN/SFK and transcription factor signalings

Importantly, the CLDN6/SFK/PI3K/AKT axis targets the AKT-phosphorylation sites in the RAR γ and the estrogen receptor α (ER α) and stimulates their activities, thereby regulating the expression of respective target genes. This conclusion was drawn from the following results: (1) the CLDN6-induced cellular events were hindered in three distinct F9: $Rxra^{-/-}:Rarg^{-/-}:Cldn6$ cell lines, despite SFKs being activated; (2) AKT formed a complex with either RXRa/RARy or ERa; and (3) characterization of F9: $Rxra^{-/-}:Rarg^{-/-}:Cldn6:iRxra-Rarg2S379A$ (hereafter, "*i*" means doxycycline-inducible expression of a give gene) and F9: $Rxra^{-/-}:Rarg^{-/-}:Cldn6:iRxra-$ Rarg2S379E cells, as well as MCF-7:ESR1S518E cells, revealed that CLDN6 signaling directs S379 and S518 in mouse RARy and human ERa, respectively. In addition, CLDN6-provoked RARyS379 phosphorylation in mice resulted in releasing the nuclear receptor corepressor (NCoR) from several retinoic acid response elements (RAREs) of three

CLDN6	CCACSSGGTQGPRH <mark>Y</mark> MAC <mark>Y</mark> STSVPHSRGPSEYPTKNYV
CLDN2	CFSCSPQGNRTN <mark>Y</mark> YDG <mark>Y</mark> QAQPLATRSSPLFNSTGYV ^SAOOPKAKSE
CLDN4	CCSCPPRSNDKPYSAKYSAARSVPASNYV
CLDN12	YCACKSLSAPGMHT <mark>Y</mark> SQPYSSRSRLSAIEIDIPVVSHS SPFWQPLYSH [^]
CLDN1	SCSCPR-KTTS <mark>Y</mark> PTPRPYP-KPTPSSGKDYV
CLDN5	CCGAWVCTGRPEFSFPVK <mark>Y</mark> SAPRRPTANG-DYDKKNYV
CLDN9	CCTCPPSHFERPRGPRLGYSIPSRSGASGLDKRDYV
CLDN17	-CCCNRK-ERWHRYPVPAYRVPQKDNQRNVTVPRKSYV ^STS
CLDN18	LEASRPALALGPTPKTRRYTMEVPAQRTRYEIPSRFLL ^NLILPSTTMCNALRPLS
CLDN3	CCSCPPRDKYAKILYSAPRSTGPG-TGTGTAYDRKDYV PT^
CLDN7	SCSCPGSESKAA <mark>Y</mark> RAPRSYPKSNSSKEYV
CLDN8	CVFCCTERSNSYR- <mark>Y</mark> SVPSHRTTQEKRSPSIYSKSQYV
CLDN10	-CFSISDNNKTPRM <mark>Y</mark> TYNGPTSAMSSRPSKQFDKNAYV ^G ^TKYQGGEGDFKTTG
CLDN14	TTTATAPAYRPPAA <mark>Y</mark> KDNRAPSVT-SAAHSGYRLNDYV
CLDN16	CCLYLFKDVGPNYP <mark>Y</mark> AMRKPYSTAGVSMAYKAPRTETA ER [^] ^KS
CLDN19	CCTCPEPANSIPQPYRSGPSTAAREPVVKLPASVKGPL ER^ ^RSFHA
CLDN11	AVMCLVGGSGDAQSFGENRFYYSSGSSSPTHAKSAHV- CVIVCC [^]
CLDN13	-CVNIPVCRDFPRCIETPSAR-PSGANNDTLD
CLDN15	TCCCSSKEEPATRAGLPYKPSTVVIPRATSDYGKNAYV ^ESDISFGK
CLDN20	VIFCTSYIQKNQEPWIYPPKQKLSTTWQPKNRRAHNLK

196

200

Figure 3. Amino acid sequences of a part of the C-terminal cytoplasmic domain in mouse CLDN1-20. Tyrosine residues corresponding to CLDN6Y196/200 are highlighted and the conserved ones are indicated in red.

distinct RA target genes, including Cldn6. Since RXRa/RAR γ heterodimer appears to induce Cldn6 gene expression,^{56,61} the positive loop of the CLDN6–RAR γ cascade could contribute not only to triggering but also to the maintenance of CLDN6-initiated cellular events. Intriguingly, the AKT-consensus phosphorylation motifs are conserved in 14 of 48 members of human nuclear receptors, implying the biological relevance of this phosphorylation site.

Aberrant CLDN-transcription factor signaling in cancer

We have recently reported that aberrant CLDN6 expression in endometrial cancer tissues is significantly associated with several clinicopathological variables, such as surgical stage III/IV, histological type, histological grade 3, lymphovascular space involvement, lymph node metastasis, and distant metastasis.⁸² Additionally, we showed that the high CLDN6 expression in endometrial cancer represents an independent

prognosis marker, and the 5-y survival rate was approximately 30%, which was one-third of that in the low-expression group.

In an additional study, we found that aberrant CLDN6 expression promotes the malignant phenotypes of endometrial cancer in vitro and in vivo via hijacking the CLDN6-ERa axis.⁸³ For instance, we demonstrated that abnormal CLDN6-ERa signaling stimulates not only cell proliferation but also collective cell migration in the leading front of endometrial cancer cells. It is noteworthy that activated SFKs appear to be concentrated at the cell borders together with CLDN6 in Ishikawa:CLDN6 cells but not in parental Ishikawa cells (Figure 4). The EC2 domain and Y196/ 200 of CLDN6 were required to recruit and activate SFKs and to stimulate malignant phenotypes of endometrial cancer cells. In addition, the CLDN6/SFK/ PI3K pathway propagates both AKT and serum- and glucocorticoid-regulated kinase (SGK), which share a high degree of homology and the same consensus phosphorylation motif,⁸⁴ resulting in targeting S518 in human ERa and activating target genes in a ligandindependent manner. Furthermore, RNAseq and RT-

qPCR analyses indicated the presence of not only ERαdependent but also ERα-independent CLDN6 signaling (Figure 5). The identification of this machinery highlights the regulation of transcription factor activity by cell adhesion to advance tumor progression.

Another issue that should be mentioned is other abnormal CLDN-transcription factor signaling in health and disease. The CLDN18/Yes-associated protein (YAP) pathway regulates the homeostasis of normal lung stem and progenitor cells, and its deficiency promotes tumorigenesis and progression of lung and gastric adenocarcinoma.^{69,85–87} By contrast, CLDN2 activates YAP, leading to self-renewal of human colorectal cancer stem-like cells.⁸⁸ Since CLDN2 has two conserved pY in the C-terminal cytoplasmic domain as described above, it should also be verified whether CLDN2/SFK signaling is involved in the self-renewal of various cancer stem-like cells.

Conclusions and future directions

The work discussed in the present review highlights the CLDN-transcription factor signaling pathway,



Figure 4. Aberrant SFK activation in human endometrial cancer cells by CLDN6. Scale bar, 20 µm.



Figure 5. The ERα-dependent and independent CLDN6 signaling in endometrial cancer. A–F indicate signaling proteins. TF, transcription factor.

notably the CLDN/SFK/PI3K/AKT/nuclear receptor cascade. Cell-cell and cell-matrix adhesion molecules are indispensable not only for proper tissue integrity but also for signaling properties that coordinate a wide range of cell behaviors. In other words, appropriate tissue formation connected by various cell adhesion proteins should be pre-requested for normal cell-adhesion signal. Since cell-cell and cell-matrix adhesion proteins are broadly expressed in distinct cell types, we propose that various combinations of cell adhesion molecules and transcription factors coordinate diverse physiological and pathological processes, including cancer. The cell-adhesion signals most probably lead to posttranslational modification of transcription factors, thereby regulating their activities. In future, it would be interesting to generalize the cell adhesion-transcription factor signaling pathway in health and disease.

Author Contributions

H.C. wrote the paper, and H.C. and K.S. prepared figures and agreed to the final version of the manuscript.

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Disclosure of Potential Conflicts of Interest

The authors declare no conflict of interest.

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References

- Furuse M, Fujita K, Hiiragi T, Fujimoto K, Tsukita S. Claudin-1 and -2: novel integral membrane proteins localizing at tight junctions with no sequence similarity to occludin. J Cell Biol. 1998;141(7):1539–1550. doi:10.1083/jcb.141.7.1539.
- Furuse M, Sasaki H, Fujimoto K, Tsukita S. A single gene product, claudin-1 or -2, reconstitutes tight junction strands and recruits occludin in fibroblasts. J Cell Biol. 1998;143(2):391-401. doi:10.1083/jcb.143.2.391.
- Furuse M, Tsukita S. Claudins in occluding junctions of humans and flies. Trends Cell Biol. 2006;16(4):181–188. doi:10.1016/j.tcb.2006.02.006.

- 4. Van Itallie CM, Anderson JM. Claudins and epithelial paracellular transport. Annu Rev Physiol. 2006;68 (1):403–429. doi:10.1146/annurev. physiol.68.040104.131404.
- Chiba H, Osanai M, Murata M, Kojima T, Sawada N. Transmembrane proteins of tight junctions. Biochim Biophys Acta. 2008;1778(3):588–600. doi:10.1016/j. bbamem.2007.08.017.
- Zihni C, Mills C, Matter K, Balda MS. Tight junctions: from simple barriers to multifunctional molecular gates. Nat Rev Mol Cell Biol. 2016;17(9):564–580. doi:10.1038/nrm.2016.80.
- Tsukita S, Tanaka H, Tamura TA. The claudins: from tight junctions to biological systems. Trends Biochem Sci. 2019;44(2):141–152. doi:10.1016/j.tibs.2018.09.008.
- Krause G, Winkler L, Mueller SL, Haseloff RF, Piontek J, Blasig IE. Structure and function of claudins. Biochim Biophys Acta. 2008;1778 (3):631–645. doi:10.1016/j.bbamem.2007.10.018.
- Piontek J, Winkler L, Wolburg H, Müller SL, Zuleger N, Piehl C, Wiesner B, Krause G, Blasig IE. Formation of tight junction: determinants of homophilic interaction between classic claudins. FASEB J. 2008;22(1):146–158. doi:10.1096/fj.07-8319com.
- Suzuki H, Nishizawa T, Tani K, Yamazaki Y, Tamura A, Ishitani R, Dohmae N, Tsukita S, Nureki O, Fujiyoshi Y. Crystal structure of a claudin provides insight into the architecture of tight junctions. Science. 2014;344(6181):304–307. doi:10.1126/science.1248571.
- Haseloff RF, Piontek J, Blasig IE. Chapter 5 the investigation of cis- and trans-interactions between claudins. In: Yu AS L editor. Current topics in membranes. Academic Press, USA; 2010. p. 97–112.
- Itoh M, Furuse M, Morita K, Kubota K, Saitou M, Tsukita S. Direct binding of three tight junction-associated MAGUKs, ZO-1, ZO-2, and ZO-3, with the COOH termini of claudins. J Cell Biol. 1999;147(6):1351–1363. doi:10.1083/jcb.147.6.1351.
- Tsukita S, Furuse M, Itoh M. Multifunctional strands in tight junctions. Nat Rev Mol Cell Biol. 2001;2 (4):285–293. doi:10.1038/35067088.
- González-Mariscal L, Garay E, Quirós M. Chapter 6 regulation of claudins by posttranslational modifications and cell-signaling cascades. In: Yu AS L editor. Current topics in membranes. Academic Press, USA; 2010. p. 113–150.
- Van Itallie CM, Anderson JM. Phosphorylation of tight junction transmembrane proteins: many sites, much to do. Tissue Barriers. 2018;6(1):e1382671. doi:10.1080/ 21688370.2017.1382671.
- 16. Tabariès S, McNulty A, Ouellet V, Annis MG, Dessureault M, Vinette M, Hachem Y, Lavoie B, Omeroglu A, Simon H-G, et al. Afadin cooperates with claudin-2 to promote breast cancer metastasis. Genes Dev. 2019;33(3–4):180–193. doi:10.1101/ gad.319194.118.

- Cavallaro U, Dejana E. Adhesion molecule signalling: not always a sticky business. Nat Rev Mol Cell Biol. 2011;12(3):189–197. doi:10.1038/nrm3068.
- Günzel D, Yu ASL. Claudins and the modulation of tight junction permeability. Physiol Rev. 2013;93 (2):525-569. doi:10.1152/physrev.00019.2012.
- 19. Sawada N. Tight junction-related human diseases. Pathol Int. 2013;63(1):1-12. doi:10.1111/pin.12021.
- Barmeyer C, Schulzke JD, Fromm M. Claudin-related intestinal diseases. Semin Cell Dev Biol. 2015;42:30–38. doi:10.1016/j.semcdb.2015.05.006.
- Tawar RG, Colpitts CC, Lupberger J, El-Saghire H, Zeisel MB, Baumert TF. Claudins and pathogenesis of viral infection. Semin Cell Dev Biol. 2015;42:39–46. doi:10.1016/j.semcdb.2015.04.011.
- Zeisel MB, Dhawan P, Baumert TF. Tight junction proteins in gastrointestinal and liver disease. Gut. 2019;68(3):547–561. doi:10.1136/gutjnl-2018-316906.
- 23. Culemann S, Grüneboom A, Nicolás-Ávila JÁ, Weidner D, Lämmle KF, Rothe T, Quintana JA, Kirchner P, Krljanac B, Eberhardt M, et al. Locally renewing resident synovial macrophages provide a protective barrier for the joint. Nature. 2019;572 (7771):670–675. doi:10.1038/s41586-019-1471-1.
- Zhu Z, Yu J, Lin W, Tang H, Zhang W, Lu B. Molecular hydrogen accelerates the reversal of acute obstructive cholangitis-induced liver dysfunction by restoring gap and tight junctions. Mol Med Rep. 2019;19 (6):5177–5184. doi:10.3892/mmr.2019.10179.
- Zuo L, Kuo W-T, Turner JR. Tight junctions as targets and effectors of mucosal immune homeostasis. Cell Mol Gastroenterol Hepatol. 2020;10(2):327–340. doi:10.1016/j.jcmgh.2020.04.001.
- Oliveira SS, Morgado-Díaz JA. Claudins: multifunctional players in epithelial tight junctions and their role in cancer. Cell Mol Life Sci. 2007;64(1):17–28. doi:10.1007/s00018-006-6314-1.
- Valle BL, Morin PJ. Chapter 13 claudins in cancer biology. In: Yu AS L editor. Current topics in membranes. Academic Press, United States; 2010. p. 293–333.
- Turksen K, Troy T-C. Junctions gone bad: claudins and loss of the barrier in cancer. Biochim Biophys Acta. 2011;1816(1):73–79. doi:10.1016/j.bbcan.2011.04.001.
- Osanai M, Takasawa A, Murata M, Sawada N. Claudins in cancer: bench to bedside. Pflugers Arch. 2017;469 (1):55–67. doi:10.1007/s00424-016-1877-7.
- Tabariès S, Siegel PM. The role of claudins in cancer metastasis. Oncogene. 2017;36(9):1176–1190. doi:10.1038/onc.2016.289.
- Gowrikumar S, Singh AB, Dhawan P. Role of claudin proteins in regulating cancer stem cells and chemoresistance-potential implication in disease prognosis and therapy. Int J Mol Sci. 2019;21(1):53. doi:10.3390/ijms21010053.
- 32. Bhat AA, Syed N, Therachiyil L, Nisar S, Hashem S, Macha MA, Yadav SK, Krishnankutty R,

Muralitharan S, Al-Naemi H, et al. Claudin-1, a double-edged sword in cancer. Int J Mol Sci. 2020;21 (2):569. Available from: http://dx.doi.org/10.3390/ ijms21020569.

- 33. Menard C, Pfau ML, Hodes GE, Kana V, Wang VX, Bouchard S, Takahashi A, Flanigan ME, Aleyasin H, LeClair KB, et al. Social stress induces neurovascular pathology promoting depression. Nat Neurosci. 2017;20 (12):1752–1760. doi:10.1038/s41593-017-0010-3.
- 34. Nishiura K, Ichikawa-Tomikawa N, Sugimoto K, Kunii Y, Kashiwagi K, Tanaka M, Yokoyama Y, Hino M, Sugino T, Yabe H, et al. PKA activation and endothelial claudin-5 breakdown in the schizophrenic prefrontal cortex. Oncotarget. 2017;8(55):93382–93391. doi:10.18632/oncotarget.21850.
- Greene C, Campbell M. Tight junction modulation of the blood brain barrier: CNS delivery of small molecules. Tissue Barriers. 2016;4(1):e1138017. doi:10.1080/21688370.2015.1138017.
- 36. Yamazaki Y, Shinohara M, Shinohara M, Yamazaki A, Murray ME, Liesinger AM, Heckman MG, Lesser ER, Parisi JE, Petersen RC, et al. Selective loss of cortical endothelial tight junction proteins during Alzheimer's disease progression. Brain. 2019;142(4):1077–1092. doi:10.1093/brain/awz011.
- 37. Niu J, Tsai H-H, Hoi KK, Huang N, Yu G, Kim K, Baranzini SE, Xiao L, Chan JR, Fancy SPJ. Aberrant oligodendroglial-vascular interactions disrupt the blood-brain barrier, triggering CNS inflammation. Nat Neurosci. 2019;22(5):709–718. doi:10.1038/s41593-019-0369-4.
- 38. Dudek KA, Dion-Albert L, Lebel M, LeClair K, Labrecque S, Tuck E, Ferrer Perez C, Golden SA, Tamminga C, Turecki G, et al. Molecular adaptations of the blood-brain barrier promote stress resilience vs. Depression Proc Natl Acad Sci U S A. 2020;117 (6):3326–3336. doi:10.1073/pnas.1914655117.
- Greene C, Hanley N, Campbell M. Blood-brain barrier associated tight junction disruption is a hallmark feature of major psychiatric disorders. Transl Psychiatry. 2020;10(1):373. doi:10.1038/s41398-020-01054-3.
- Greene C, Hanley N, Campbell M. Claudin-5: gatekeeper of neurological function. Fluids Barriers CNS. 2019;16(1):3. doi:10.1186/s12987-019-0123-z.
- Chiba H, Ichikawa-Tomikawa N, Imura T, Sugimoto K. The region-selective regulation of endothelial claudin-5 expression and signaling in brain health and disorders. J Cell Physiol. 2021. Available from 10.1002/jcp.30357
- Spadaro D, Le S, Laroche T, Mean I, Jond L, Yan J, Citi S. Tension-dependent stretching activates ZO-1 to control the junctional localization of its interactors. Curr Biol. 2017;27(3783–3795.e8):3783–3795.e8. doi:10.1016/j.cub.2017.11.014.
- 43. Zenker J, White MD, Gasnier M, Alvarez YD, Lim HYG, Bissiere S, Biro M, Plachta N. Expanding actin rings zipper the mouse embryo for blastocyst

formation. Cell. 2018;173(3):776-791.e17. doi:10.1016/ j.cell.2018.02.035.

- 44. Bendriem RM, Singh S, Aleem AA, Antonetti DA, Ross ME. Tight junction protein occludin regulates progenitor self-renewal and survival in developing cortex. Elife. 2019;8:664. doi:10.7554/eLife.49376.
- Beutel O, Maraspini R, Pombo-García K, Martin-Lemaitre C, Honigmann A. Phase separation of zonula occludens proteins drives formation of tight junctions. Cell. 2019;179(4):923–936.e11. doi:10.1016/j. cell.2019.10.011.
- 46. Kuo W-T, Shen L, Zuo L, Shashikanth N, Ong MLDM, Wu L, Zha J, Edelblum KL, Wang Y, Wang Y, et al. Inflammation-induced occludin downregulation limits epithelial apoptosis by suppressing caspase-3 expression. Gastroenterology. 2019;157(5):1323–1337. doi:10.1053/j.gastro.2019.07.058.
- 47. Otani T, Nguyen TP, Tokuda S, Sugihara K, Sugawara T, Furuse K, Miura T, Ebnet K, Furuse FM. Claudins and JAM-A coordinately regulate tight junction formation and epithelial polarity. J Cell Biol. 2019;218(10):3372–3396. doi:10.1083/jcb.201812157.
- Schwayer C, Shamipour S, Pranjic-Ferscha K, Schauer A, Balda M, Tada M, Matter K, Heisenberg C-P. Mechanosensation of tight junctions depends on ZO-1 phase separation and flow. Cell. 2019;179(4):937–952.e18. doi:10.1016/j. cell.2019.10.006.
- 49. Barnat M, Capizzi M, Aparicio E, Boluda S, Wennagel D, Kacher R, Kassem R, Lenoir S, Agasse F, Braz BY, et al. Huntington's disease alters human neurodevelopment. Science. 2020;369(6505):787–793. doi:10.1126/science.aax3338.
- Belardi B, Hamkins-Indik T, Harris AR, Kim J, Xu K, Fletcher DA, Weak A. Link with actin organizes tight junctions to control epithelial permeability. Dev Cell. 2020;54(792-804.e7):792-804.e7. doi:10.1016/j. devcel.2020.07.022.
- Citi S. Cell Biology: Tight junctions as biomolecular condensates. Curr Biol. 2020;30(2):R83–6. doi:10.1016/ j.cub.2019.11.060.
- Hartmann C, Schwietzer YA, Otani T, Furuse M, Ebnet K. Physiological functions of junctional adhesion molecules (JAMs) in tight junctions. Biochim Biophys Acta. 2020;1862(9):183299. doi:10.1016/j. bbamem.2020.183299.
- 53. Lauko A, Mu Z, Gutmann DH, Naik UP, Lathia JD. Junctional adhesion molecules in cancer: a paradigm for the diverse functions of cell-cell interactions in tumor progression. Cancer Res. 2020;80(22):4878–4885. doi:10.1158/0008-5472. CAN-20-1829.
- 54. Otani T, Furuse M. Tight junction structure and function revisited. Trends Cell Biol. 2020;30(10):805-817. doi:10.1016/j.tcb.2020.08.004.
- 55. Zhou T, Lu Y, Xu C, Wang R, Zhang L, Lu P. Occludin protects secretory cells from ER stress by facilitating

SNARE-dependent apical protein exocytosis. Proc Natl Acad Sci U S A. 2020;117(9):4758–4769. doi:10.1073/ pnas.1909731117.

- 56. Kubota H, Chiba H, Takakuwa Y, Osanai M, Tobioka H, Kohama G, Mori M, Retinoid SN. X receptor alpha and retinoic acid receptor gamma mediate expression of genes encoding tight-junction proteins and barrier function in F9 cells during visceral endodermal differentiation. Exp Cell Res. 2001;263 (1):163–172. doi:10.1006/excr.2000.5113.
- 57. Chiba H, Gotoh T, Kojima T, Satohisa S, Kikuchi K, Osanai M, Sawada N. Hepatocyte nuclear factor (HNF)-4alpha triggers formation of functional tight junctions and establishment of polarized epithelial morphology in F9 embryonal carcinoma cells. Exp Cell Res. 2003;286(2):288–297. doi:10.1016/S0014-4827(03) 00116-2.
- Satohisa S, Chiba H, Osanai M, Ohno S, Kojima T, Saito T, Sawada N. Behavior of tight-junction, adherens-junction and cell polarity proteins during HNF-4alpha-induced epithelial polarization. Exp Cell Res. 2005;310(1):66–78. doi:10.1016/j.yexcr.2005.06.025.
- Chiba H, Sakai N, Murata M, Osanai M, Ninomiya T, Kojima T, Sawada N. The nuclear receptor hepatocyte nuclear factor 4alpha acts as a morphogen to induce the formation of microvilli. J Cell Biol. 2006;175 (6):971–980. doi:10.1083/jcb.200608012.
- 60. Sugimoto K, Ichikawa-Tomikawa N, Satohisa S, Akashi Y, Kanai R, Saito T, Sawada N, Chiba H. The tight-junction protein claudin-6 induces epithelial differentiation from mouse F9 and embryonic stem cells. PLoS One. 2013;8(10):e75106. doi:10.1371/journal. pone.0075106.
- 61. Sugimoto K, Ichikawa-Tomikawa N, Kashiwagi K, Endo C, Tanaka S, Sawada N, Watabe T, Higashi T, Chiba H. Cell adhesion signals regulate the nuclear receptor activity. Proc Natl Acad Sci U S A. 2019;116(49):24600-24609. doi:10.1073/ pnas.1913346116.
- 62. Li C-F, Chen J-Y, Ho Y-H, Hsu W-H, Wu L-C, Lan H-Y, Hsu D-S-S, Tai S-K, Chang Y-C, Yang M-H. Snail-induced claudin-11 prompts collective migration for tumour progression. Nat Cell Biol. 2019;21 (2):251–262. doi:10.1038/s41556-018-0268-z.
- Singh AB, Sharma A, Dhawan P. Claudin-1 expression confers resistance to anoikis in colon cancer cells in a Src-dependent manner. Carcinogenesis. 2012;33 (12):2538–2547. doi:10.1093/carcin/bgs275.
- Huang H, Li L, Wu C, Schibli D, Colwill K, Ma S, Li C, Roy P, Ho K, Songyang Z, et al. Defining the specificity space of the human SRC homology 2 domain. Mol Cell Proteomics. 2008;7(4):768–784. doi:10.1074/mcp. M700312-MCP200.
- Filippakopoulos P, Müller S, Knapp S. SH2 domains: modulators of nonreceptor tyrosine kinase activity. Curr Opin Struct Biol. 2009;19(6):643–649. doi:10.1016/j.sbi.2009.10.001.

- 66. Liu BA, Engelmann BW, Jablonowski K, Higginbotham K, Stergachis AB, Nash PD. SRC homology 2 domain binding sites in insulin, IGF-1 and FGF receptor mediated signaling networks reveal an extensive potential interactome. Cell Commun Signal. 2012;10(1):27. doi:10.1186/1478-811X-10-27.
- Mayer BJ. The discovery of modular binding domains: building blocks of cell signalling. Nat Rev Mol Cell Biol. 2015;16(11):691–698. doi:10.1038/nrm4068.
- 68. Shimobaba S, Taga S, Akizuki R, Hichino A, Endo S, Matsunaga T, Watanabe R, Yamaguchi M, Yamazaki Y, Sugatani J, et al. Claudin-18 inhibits cell proliferation and motility mediated by inhibition of phosphorylation of PDK1 and Akt in human lung adenocarcinoma A549 cells. Biochim Biophys Acta. 2016;1863(6):1170–1178. doi:10.1016/j.bbamcr.2016.02.015.
- 69. Luo J, Chimge N-O, Zhou B, Flodby P, Castaldi A, Firth AL, Liu Y, Wang H, Yang C, Marconett CN, et al. CLDN18.1 attenuates malignancy and related signaling pathways of lung adenocarcinoma in vivo and in vitro. Int J Cancer. 2018;143(12):3169–3180. doi:10.1002/ijc.31734.
- Brunton VG, Irj M, Frame MC. Cell adhesion receptors, tyrosine kinases and actin modulators: a complex three-way circuitry. Biochim Biophys Acta. 2004;1692 (2-3):121-144. doi:10.1016/j.bbamcr.2004.04.010.
- McLachlan RW, Kraemer A, Helwani FM, Kovacs EM, Yap AS, Mostov K. E-cadherin adhesion activates c-Src signaling at cell-cell contacts. Mol Biol Cell. 2007;18 (8):3214–3223. doi:10.1091/mbc.e06-12-1154.
- 72. Cabodi S, Di Stefano P, Leal M, Del Pc TA, Bisaro B, Morello V, Damiano L, Aramu S, Repetto D, Tornillo G, et al. Integrins and signal transduction. Adv Exp Med Biol. 2010;674:43–54.
- Canel M, Serrels A, Frame MC, Brunton VG. E-cadherin-integrin crosstalk in cancer invasion and metastasis. J Cell Sci. 2013;126(2):393–401. doi:10.1242/jcs.100115.
- 74. Xiao R, Xi X-D, Chen Z, Chen S-J S-J, Meng G. Structural framework of c-Src activation by integrin β 3. Blood. 2013;121(4):700–706. doi:10.1182/blood-2012-07-440644.
- Ochs K, Málaga-Trillo E. Common themes in PrP signaling: the Src remains the same. Front Cell Dev Biol. 2014;2:63. doi:10.3389/fcell.2014.00063.
- Hamidi H, Ivaska J. Every step of the way: integrins in cancer progression and metastasis. Nat Rev Cancer. 2018;18(9):533–548. doi:10.1038/s41568-018-0038-z.
- 77. Witherden DA, Verdino P, Rieder SE, Garijo O, Mills RE, Teyton L, Fischer WH, Wilson IA, Havran WL. The junctional adhesion molecule JAML is a costimulatory receptor for epithelial gammadelta T cell activation. Science. 2010;329(5996):1205–1210. doi:10.1126/science.1192698.
- Yaffe MB. MAGUK SH3 domains-swapped and stranded by their kinases? Structure. 2002;10(1):3-5. doi:10.1016/S0969-2126(01)00705-5.

- Lim WA, Pawson T. Phosphotyrosine signaling: evolving a new cellular communication system. Cell. 2010;142(5):661–667. doi:10.1016/j.cell.2010.08.023.
- Liu BA, Nash PD. Evolution of SH2 domains and phosphotyrosine signalling networks. Philos Trans R Soc Lond B Biol Sci. 2012;367(1602):2556–2573. doi:10.1098/rstb.2012.0107.
- Wagner MJ, Stacey MM, Liu BA, Pawson T. Molecular mechanisms of SH2- and PTB-domain-containing proteins in receptor tyrosine kinase signaling. Cold Spring Harb Perspect Biol. 2013;5(12):a008987. doi:10.1101/ cshperspect.a008987.
- 82. Kojima M, Sugimoto K, Tanaka M, Endo Y, Kato H, Honda T, Furukawa S, Nishiyama H, Watanabe T, Soeda S, et al. Prognostic significance of aberrant claudin-6 expression in endometrial cancer. Cancers. 2020;12(10):2748. Available from: http://dx.doi.org/10. 3390/cancers12102748.
- 83. Kojima M, Sugimoto K, Kobayashi M, Ichikawa-Tomikawa N, Kashiwagi K, Watanabe T, Soeda S, Fujimori K, Chiba H. Aberrant claudin-6-adhesion signaling promotes endometrial cancer progression via estrogen receptor α. Mol Cancer Res [*in press*; Available from]. 2021. https://mcr.aacrjournals.org/con tent/early/2021/03/16/1541-7786.MCR-20-0835
- 84. Lang F, Böhmer C, Palmada M, Seebohm G, Strutz-Seebohm N, Vallon V. (Patho)physiological significance of the serum- and glucocorticoid-inducible kinase

isoforms. Physiol Rev. 2006;86(4):1151–1178. doi:10.1152/physrev.00050.2005.

- 85. Hagen SJ, Ang L-H, Zheng Y, Karahan SN, Wu J, Wang YE, Caron TJ, Gad AP, Muthupalani S, Fox JG. Loss of tight junction protein claudin 18 promotes progressive neoplasia development in mouse stomach. Gastroenterology. 2018;155(6):1852–1867. doi:10.1053/ j.gastro.2018.08.041.
- Zhou B, Flodby P, Luo J, Castillo DR, Liu Y, Yu F-X, McConnell A, Varghese B, Li G, Chimge N-O, et al. Claudin-18-mediated YAP activity regulates lung stem and progenitor cell homeostasis and tumorigenesis. J Clin Invest. 2018;128(3):970–984. doi:10.1172/ JCI90429.
- 87. Suzuki K, Sentani K, Tanaka H, Yano T, Suzuki K, Oshima M, Yasui W, Tamura A, Tsukita S. Deficiency of stomach-type claudin-18 in mice induces gastric tumor formation independent of H pylori infection. Cell Mol Gastroenterol Hepatol. 2019;8(1):119–142. doi:10.1016/j. jcmgh.2019.03.003.
- Paquet-Fifield S, Koh SL, Cheng L, Beyit LM, Shembrey C, Mølck C, Behrenbruch C, Papin M, Gironella M, Guelfi S, et al. Tight junction protein claudin-2 promotes self-renewal of human colorectal cancer stem-like cells. Cancer Res. 2018;78 (11):2925-2938. doi:10.1158/0008-5472.CAN-17-1869.