

ORIGINAL PAPER



Ozone and microstructural morphological changes of tooth enamel

ALIN DANIEL FLOARE^{1,2)}, DAVID FOCHT^{1,2)}, ADRIAN IOAN HAJDU^{1,2)}, IOANA CRISTINA NICULESCU TALPOȘ²⁾, OCTAVIA IULIA BĂLEAN^{1,2)}, CORNELIA VERONICA MUNTEAN^{3,4)}, DORIAN SEBEȘAN^{1,2)}, DANIELA ELISABETA JUMANCA^{1,2)}, ATENA GĂLUȘCAN^{1,2)}

¹⁾Department of Preventive, Community Dentistry and Oral Health, Translational and Experimental Clinical Research Centre in Oral Health (TEXC-OH), Timișoara, Romania

²⁾1st Department of Dentistry, Faculty of Dental Medicine, Victor Babeș University of Medicine and Pharmacy, Timișoara, Romania

³⁾Department of Applied Chemistry and Engineering of Inorganic Compounds and the Environment, Faculty of Industrial Chemistry and Environmental Engineering, Politehnica University Timișoara, Romania

⁴⁾Research Institute for Renewable Energy, Politehnica University Timișoara, Romania

Abstract

The paper aims to study the impact of ozone (O₃) treatment on the microstructural changes of the tooth enamel after the treatment at different time intervals. The ozonation was performed with gaseous O₃ produced by HealOzone X4, the demineralization level was measured with the DiagnoDent Pen 2190 device, and the microstructure changes of enamel surface were observed using scanning electron microscopy (SEM) analysis. The results showed the exposure to O₃ for 40–50 seconds enhanced enamel micro-hardness and ensures a rate of remineralization between 96.82–97.38%. In conclusion, in search of new minimally invasive solutions in the treatment of caries and to offer antimicrobial support of the oral cavity, the use of O₃ as an alternative therapy to classical solutions may be a viable solution in dentistry.

Keywords: enamel, ozone gas, scanning electron microscopy, remineralizing, caries.

Introduction

Ozone (O₃) is a highly reactive compound formed of three oxygen atoms that works as an oxidant and oxidizer [1]. As a non-invasive option for treating tooth decay, O₃ therapy has some distinctive features compared to other available modalities. The aim of treating carious lesions with O₃ is to decrease the causative microbiota and contributing risk factors, stop the caries process and stimulate remineralization. Enamel is the most complex and most mineralized hard tissue in the human body, with a content of 96% hydroxyapatite (HA) and 4% water and organic material [2]. Enamel can be damaged throughout life, as it is continuously exposed to the oral environment. The acidic environment created by acidic bacteria, acidic foods and drinks results in dental problems for four out of five temporary teeth in children and over half of permanent teeth in adults) [2, 3]. Because enamel is classified as a cellular tissue, it cannot regenerate or reshaped on its own; therefore, the lesions are irreversible and restored with artificial material.

The main issue of non-invasive approaches for preventing carious lesions and remineralization of enamel is the difficulty of controlling dental plaque. O₃ can remove proteins from carious lesions and allow calcium (Ca²⁺) and phosphate (PO₄²⁻) ions to diffuse through lesions, leading to remineralization [4]. In addition, O₃ has a significant environmental advantage, meaning a rapid degradation

of O₃ clinically causes its low cytotoxicity after contact with organic compounds. All these features suggest that O₃ could be widely used soon in restorative and preventive dentistry [4].

Because of its high oxidizing power, O₃ can oxidize the bacterial cell wall, inducing its lysis and the pyruvic acid produced by the bacteria, turning it into acetic acid and carbon dioxide (CO₂), stopping or reversing the progression of caries. While the antimicrobial qualities of O₃ are well known [5–9], its remineralization potential, by increasing the permeability of the dentin tubules, is less studied. As a demineralizing agent, O₃ seems to improve the diffusion of salivary ions to the surface of degraded dentin, thereby promoting remineralization [10–13]. In this context, O₃ can neutralize acid proteins produced by cariogenic bacteria, representing the osmotic stimulus responsible for the movement of fluids in the dentin tubules leading to hypersensitivity [12–14].

Given all the therapeutic effects of O₃ mentioned above, the mechanisms of action are also diversified, such as: (i) as an antimicrobial agent, O₃ breaks down viral capsid and interrupts the reproductive cycle by peroxidation; (ii) as an immunomodulatory and antioxidant agent, it stimulates immunoglobulin synthesis, preserves cellular redox state and raises cellular levels of glutathione peroxidase (GPx); (iii) the anti-hypoxic effect occurs by increasing hemoglobin oxygen saturation; (iv) the anti-inflammatory action is due to the impact of O₃ on the synthesis of interleukins,

prostaglandins and leukotrienes; (v) the bioenergetic function is exerted by activation of protein synthesis and an by enhancement of cellular metabolism; (vi) the detoxification activity occurs by activation of cellular aerobic processes (Krebs cycle, glycolysis and fatty acid oxidation); and (vii) the biosynthetic activity is achieved by enhancement of carbohydrate, protein and lipid metabolism [15].

O₃ can be applied to the tooth surface using either gas or water. It is a powerful bio-oxidation agent, is highly bactericidal and has been utilized in dentistry to treat dental caries (enamel and dentin), root caries and dental hypersensitivity [10].

Aim

The aim of this study was the analysis of the bio-oxidative effect and the microstructural changes of the tooth enamel following the use of O₃ at different time intervals.

Materials and Methods

Biological material

The study was performed on 15 permanent human teeth extracted for orthodontic reasons without harming them. Only fully erupted molars and whole molars without clinically detectable cavity lesions were included in the study, this being the main inclusion/exclusion criterion for molars studied. Teeth were collected and kept in saline until the start of the working method. All adherent soft tissues on the teeth were removed and cleaned by flushing. The samples were obtained using the same tooth on both sides to obtain the blank and working samples on the same enamel pattern by demineralization/ozonation (each tooth offered two samples). The experiment was done in duplicate. The 30 samples obtained were fixed in silicone material for better handling.

Demineralization and ozonation procedure

Teeth demineralization was performed with 35% phosphoric acid (H₃PO₄) (Gluma[®] Etch 35 Gel, Kulzer, Hanau, Germany), which was left on the enamel surface for 5 minutes for each half of the specimen separately for effective demineralization. The working specimens were successively subjected to ozonize with gaseous O₃ produced by HealOzone X4 (KaVo Dental & Co., Biberach, Germany), which releases O₃ at a fixed concentration of 2100 ppm, with a flow rate of 615 cm²/min for 10, 20, 40 and 50 seconds, respectively. The ozonize device comprises an air filter, vacuum pump, an O₃ generator, a hand piece fitted with a sealing silicone cup, and a flexible hose. The procedure is usually between 20 and 120 seconds per tooth. According to the specifications, the device produces O₃ at a concentration of 32 g/m³ at an exposure of 60 seconds. The initial mineralization degree of the enamel and the demineralization level was measured with the DiagnoDent Pen 2190 device (KaVo Dental & Co., Biberach, Germany). This equipment uses laser-induced fluorescence (LIF) technology to determine the quality of the structure of the tooth enamel, detecting the degree of its demineralization compared to healthy enamel. Fluorescent tooth enamel physiologically loses its properties with demineralization so that measurements with DiagnoDent Pen indicate on

a scale of 0–12 healthy enamel, 13–24 enamel lesions and 25–99 dentin lesion (depending on the degree of demineralization).

The rate of mineralization was calculated with the formula:

$$\text{Rate of mineralization [\%]} = [(SD - SR)/SD] \times 100$$

where, SD – the DiagnoDent score after demineralization; SR – the DiagnoDent score after remineralization at different time.

The working (ozonized) and control (demineralized) samples were then analyzed with the scanning electron microscopy (SEM) using Quanta 250 FEG (FEI Company, Hillsboro, Oregon, USA) scanning electron microscope operating at 10 kV and 10 mm working distance to highlight the structural changes produced in the enamel. The applied procedure is presented in the Figure 1.

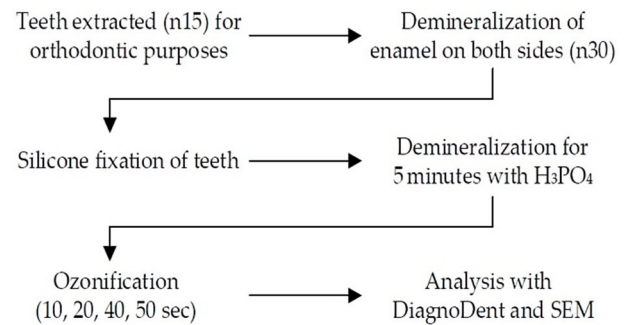


Figure 1 – The demineralization/ozonation procedure. H₃PO₄: Phosphoric acid; SEM: Scanning electron microscopy.

Results

The remineralizing potential of ozone

In Figure 2 is presented the demineralized tooth (Figure 2A) compared with a tooth treated with O₃ after demineralization (Figure 2B). In Figure 3 is presented the score of DiagnoDent Pen 2190 measurement and in Figures 4 and 5, the microscopic aspect (SEM) of tooth enamel before and after exposure to O₃ for 10 seconds and 50 seconds, respectively.

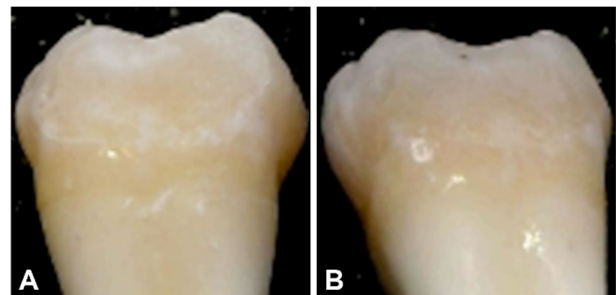


Figure 2 – The tooth aspect before and after exposure to the ozone treatment: (A) Demineralized tooth; (B) Tooth treated with ozone after demineralization.

Values are reported as mean values ± standard deviations of total measurements. A *t*-test was applied to compare mean differences between samples; data within the same row with different superscripts are significantly different ($p < 0.05$); data within the same row with the same superscripts are not significantly different ($p > 0.05$).

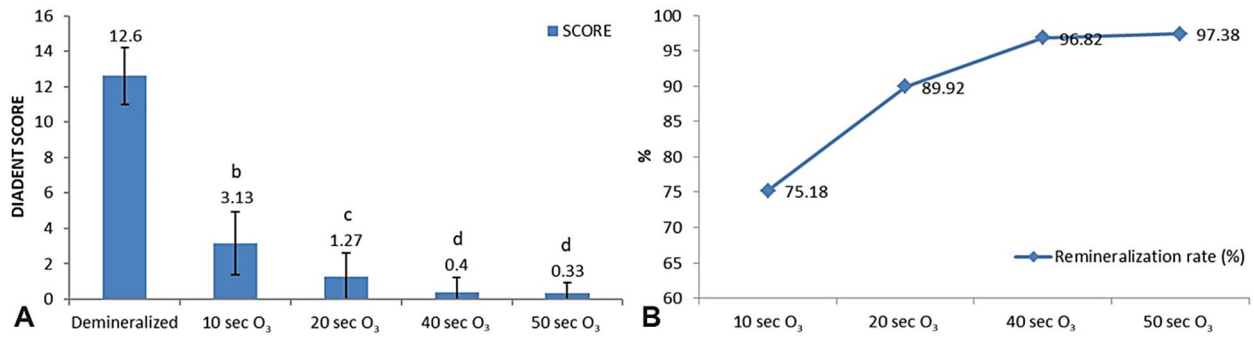


Figure 3 – DiagnoDent Pen 2190 measurement of the demineralized/ozonized enamel: (A) DiagnoDent score; (B) The remineralization rate. O₃: Ozone.

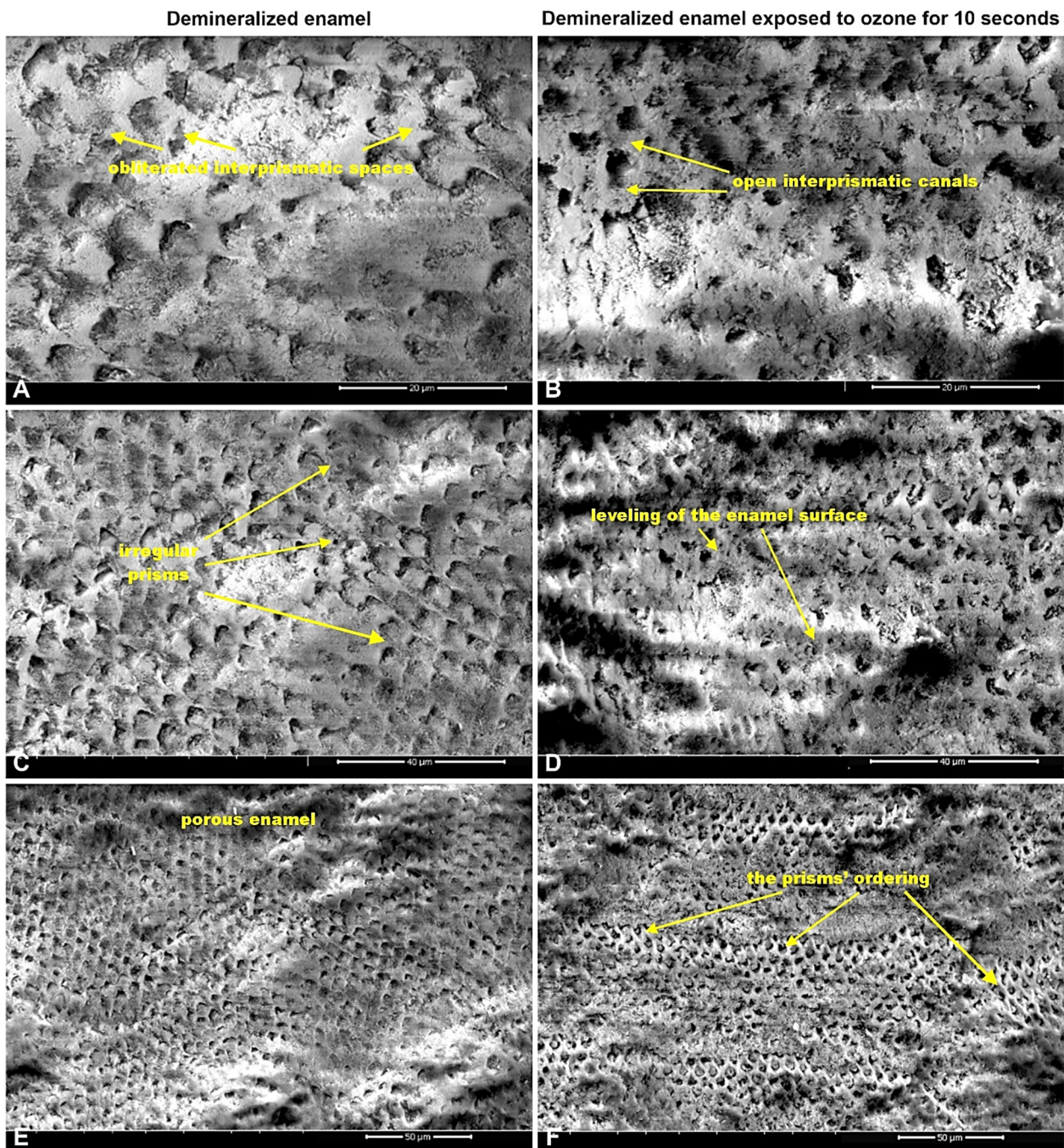


Figure 4 – Microscopic aspect of tooth enamel before (A, C and E) and after exposure to 10 seconds ozone (B, D, and F). Microscopic aspect of demineralized enamel: (A) ×2000; (C) ×1000; (E) ×500. Microscopic aspect of demineralized enamel exposed to ozone: (B) ×2000; (D) ×1000; (F) ×500. Scale bar: (A and B) 20 μm; (C and D) 40 μm; (E and F) 50 μm.

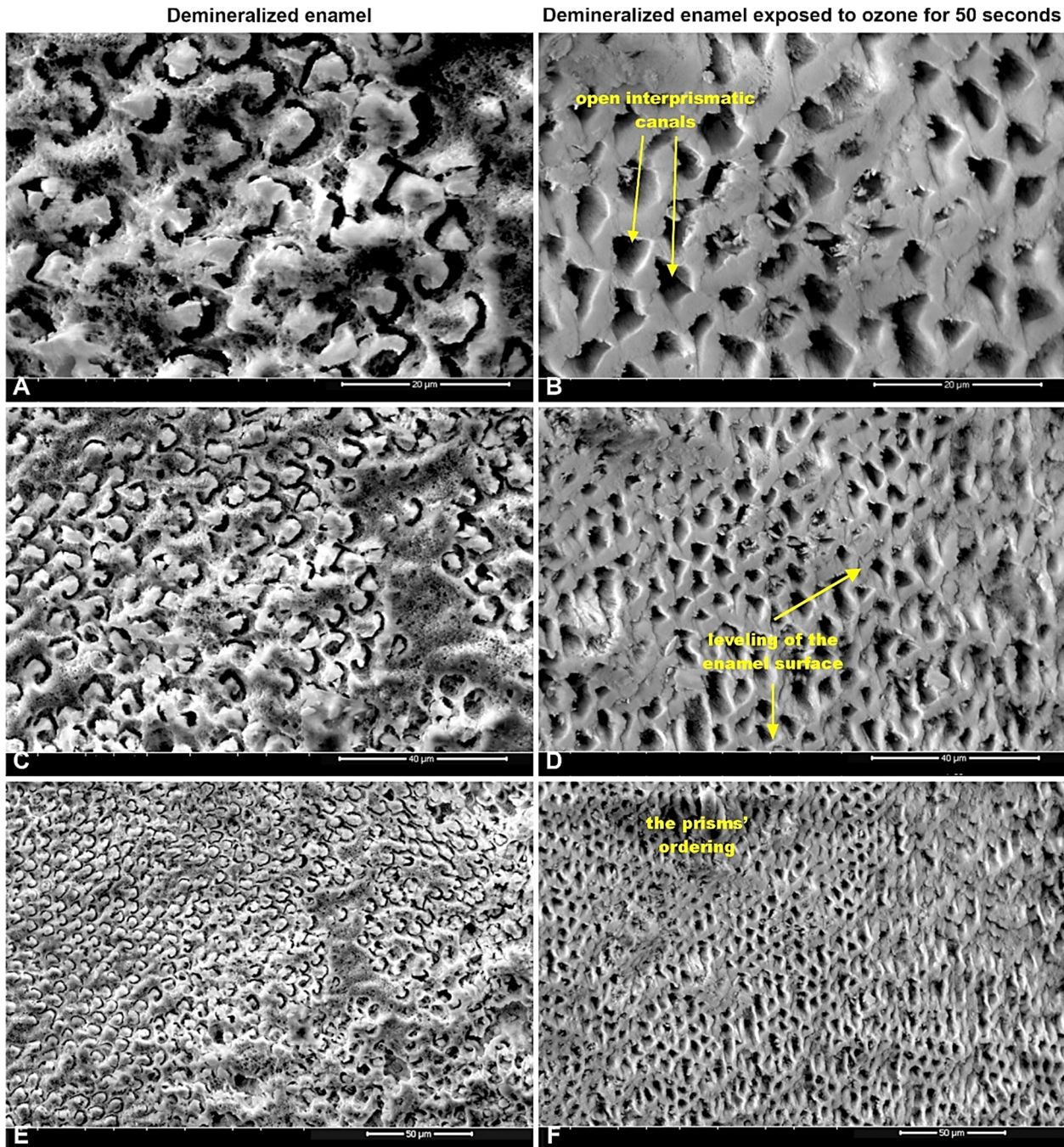


Figure 5 – Microscopic aspect of tooth enamel before (A, C and E) and after exposure to 50 seconds ozone (B, D, and F). Microscopic aspect of demineralized enamel: (A) $\times 2000$; (C) $\times 1000$; (E) $\times 500$. Microscopic aspect of demineralized enamel exposed to ozone: (B) $\times 2000$; (D) $\times 1000$; (F) $\times 500$. Scale bar: (A and B) $20\ \mu\text{m}$; (C and D) $40\ \mu\text{m}$; (E and F) $50\ \mu\text{m}$.

The results of the test performed using DiagnoDent Pen 2190 equipment before and after O_3 exposure (Figure 2, A and B) shown that initially, after demineralization, the scores obtained are between 11–14, recommended intensive prophylaxis measures, including ozonized. After a reduced exposure of only 10 seconds to the O_3 treatment, a significant decrease of DiagnoDent Pen 2190 scores is observed, with values between 1–5, respectively after 20 seconds the values obtained decrease between 0–3, and after 40–50 seconds the scores obtained be less than 1. The exposure to O_3 gas for 40 seconds dehydrated the enamel and increased its micro-hardness.

The statistical analysis using *t*-test shows statistical relevance ($p < 0.05$) between demineralization sample (after

acidic treatment and before O_3 exposure) and experimental trials (with O_3 treatment at different exposure time). Thus, it can be said that the dental structures were remineralized in proportions compared to the control samples depending on the O_3 exposure. After 10 seconds of exposure, the remineralization rate was 75.18% and increased at 89.92% after 20 seconds. Higher rates (96.82% and 97.38%) were recorded after 40 seconds, respectively 50 seconds of O_3 exposure. According to *t*-test, there is a dominant difference between the values recorded at the exposed sample at 10 seconds from the initial sample. No significant differences are recorded between the score obtained after 40 and 50 seconds to O_3 exposure.

Tooth enamel, like bones, is the hardest tissue in the

body, being even more mineralized than the skeleton and having a complex three-dimensional structure due to the presence of HA crystals. Under normal conditions, mammals have a uniform and orderly spatial arrangement of these tightly packed crystals. Thus, the HA crystals of the enamel are organized in the form of prisms (aligned crystals) and in the form of an interprismatic substance [16, 17].

The SEM analysis of the O₃-exposed enamel reveals morphological changes in its roughness depending on the exposure times. Microscopically aspect of demineralized enamel recorded at ×500 magnification shown porous aspect of enamel (Figure 4A), respectively irregular prisms when the magnification was increased at ×1000 (Figure 4C) and obliterated interprismatic spaces at ×2000 (Figure 4E). The exposure at O₃ treatment for 10 seconds leads prisms ordering, leveling of the enamel surface and to the opening of interprismatic canals (Figure 4, B, D and F). The phenomena of enamel quality improvement are even more pronounced after exposure of 50 seconds to O₃ treatment (Figure 5, A–F).

These changes depend on the exposure time and the opportunity to disrupt the proteins embedded in the enamel matrix by bio-oxidation. Oxygen penetrates more easily into the structure of the enamel in the organic phase. On the surface of the demineralized enamel, the appearance of porous enamel was noticed because of the dissolution of the HA crystals around the enamel prisms. Also, irregular prisms and obliterated interprismatic spaces with the appearance of rough enamel was highlighted (Figure 4, A–C). After ozonized, the SEM images show a leveling of the enamel surface, open interprismatic canals, and the prisms' ordering.

At the higher the exposure, the better the appearance of the honeycomb with an orderly structure of the HA prisms and the cleaning of the interprismatic spaces was noticed (Figures 4 and 5).

☞ Discussions

In this study, the remineralizing potential of O₃ was evaluated using two methods: (i) the measurement of fissure carries using DiagnoDent Pen 2190; and (ii) analysis of the microscopic aspect (SEM) of tooth enamel before and after exposure to O₃ treatment. The use of DiagnoDent Pen 2190 offers advantages in minimally invasive therapy and completes the clinical diagnosis and dental radiography. The smallest changes can be established and treated accordingly, which cannot be seen with the naked eye up to a depth of up to 2 mm from the dentin. The scores obtained in case of demineralization diagnosis using DiagnoDent Pen 2190 recommend the prophylactic treatment, as follows: (i) for values between 0–12, normal prophylaxis measures are recommended (e.g., fluoride toothpaste); (ii) for values between 13 and 24, intensive prophylaxis measures are recommended (egg fluoridation or ozonizing); (iii) for values above 25, minimally invasive restorative procedures are recommended using filling materials and intensive prophylaxis, respectively for large lesions, classical restoration, depending on the risk assessment and on the diagnosis. The results analysis using DiagnoDent Pen 2190 measurement of the demineralized/ozonizing enamel shown that the scores decreased after the application of O₃ (Figure 2, A and B). The level of reduction, compared

to the score obtained after demineralization was 75.18% after 10 seconds, 89.92% after 20 seconds, respectively 96.82% after 40 seconds and 97.38% after 50 seconds of remineralization. The results confirm the research obtained by other authors who showed that O₃ was effective on remineralization [15, 18].

SEM was used by several studies to assess the demineralization/remineralization effect, SEM analysis is often used to evaluate enamel processes, both in cariology, the adhesion of enamel materials, orthodontics [19, 20].

Baysan *et al.* (2007), evaluated the clinical inversion of root caries by exposure for 10 seconds to gaseous O₃ at 2100 ppm, succeeded by application for 5 seconds of xylitol and fluoride. The study found that after six months, O₃-treated lesions were significantly reduced compared to controls. Similar results were reported after 18 months, when primary root caries lesions were treated with gaseous O₃ (2100 ppm for 10 seconds), followed by daily use of the remineralization kit with patent [21]. The experimental studies recommend the exposure to O₃ gas for 40 seconds to enhance the enamel micro-hardness [15, 22]. Our study confirms the maximum efficiency of exposure for 40 seconds to O₃ on the remineralization of the enamel, but significant effects can be registered at an exposure of only 10 seconds.

The role played by O₃ in this process as a powerful oxidant and proteolytic agent that can disrupt the organic content of demineralized enamel and enhance the diffusion of remineralizing agents. O₃ oxidizes pyruvic acid (responsible for lowering pH and the carry process) into acetate and CO₂ and promote remineralization because acetate has a greater pKa and promotes pH buffering in the resting plate [23].

☞ Conclusions

In search of new minimally invasive solutions in the treatment of caries and to offer antimicrobial support of the oral cavity, the use of O₃ as an alternative therapy to classical solutions may be a viable solution in dentistry. Even though the use of O₃ in routine dental medicine is not yet universally accepted due to lack knowledge, limited training and evidence in the literature, the results obtained in this study are encouraging and offer new approaches in dental technology.

☞ Conflict of interests

The authors declare no conflict of interests.

☞ Funding

This research was funded by Victor Babeş University of Medicine and Pharmacy, Timișoara, Romania.

☞ References

- [1] Suh Y, Patel S, Kaitlyn R, Gandhi J, Joshi G, Smith NL, Khan SA. Clinical utility of ozone therapy in dental and oral medicine. *Med Gas Res*, 2019, 9(3):163–167. <https://doi.org/10.4103/2045-9912.266997> PMID: 31552882 PMCID: PMC6779001
- [2] Lacruz RS, Habelitz S, Wright JT, Paine ML. Dental enamel formation and implications for oral health and disease. *Physiol Rev*, 2017, 97(3):939–993. <https://doi.org/10.1152/physrev.0030.2016> PMID: 28468833 PMCID: PMC6151498
- [3] Kreulen CM, Van 't Spijker A, Rodriguez JM, Bronkhorst EM, Creugers NHJ, Bartlett DW. Systematic review of the prevalence of tooth wear in children and adolescents. *Caries Res*, 2010, 44(2):151–159. <https://doi.org/10.1159/000308567> PMID: 20389070

- [4] Elsayad II. Chemical analysis and surface morphology of enamel following ozone application with different concentrations and exposure times. *J Adv Res*, 2011, 2(2):131–136. <https://doi.org/10.1016/j.jare.2010.10.001> <https://www.sciencedirect.com/science/article/pii/S209012321000113X>
- [5] Ximenes M, Cardoso M, Astorga F, Arnold R, Pimenta LA, Viera RS. Antimicrobial activity of ozone and NaF–Chlorhexidine on early childhood caries. *Braz Oral Res*, 2017, 31:e2. <https://doi.org/10.1590/1807-3107BOR-2017.vol31.0002> PMID: 28076495
- [6] Sen S, Sen S. Ozone therapy a new vista in dentistry: integrated review. *Med Gas Res*, 2020, 10(4):189–192. <https://doi.org/10.4103/2045-9912.304226> PMID: 33380587 PMCID: PMC8092153
- [7] Elvis AM, Ekta JS. Ozone therapy: a clinical review. *J Nat Sci Biol Med*, 2011, 2(1):66–70. <https://doi.org/10.4103/0976-9668.82319> PMID: 22470237 PMCID: PMC3312702
- [8] Viebahn-Haensler R, León Fernández OS. Ozone in medicine. The low-dose ozone concept and its basic biochemical mechanisms of action in chronic inflammatory diseases. *Int J Mol Sci*, 2021, 22(15):7890. <https://doi.org/10.3390/ijms22157890> PMID: 34360655 PMCID: PMC8346137
- [9] Santos GM, Pacheco RL, Bussadori SK, Santos EM, Riera R, de Oliveira Cruz Latorraca C, Mota P, Benavent Caldas Bellotto EF, Martimbianco ALC. Effectiveness and safety of ozone therapy in dental caries treatment: systematic review and meta-analysis. *J Evid Based Dent Pract*, 2020, 20(4):101472. <https://doi.org/10.1016/j.jebdp.2020.101472> PMID: 33303100
- [10] Almaz ME, Sönmez İŞ. Ozone therapy in the management and prevention of caries. *J Formos Med Assoc*, 2015, 114(1):3–11. <https://doi.org/10.1016/j.jfma.2013.06.020> PMID: 23969041
- [11] Grocholewicz K, Matkowska-Cichocka G, Makowiecki P, Drożdżik A, Ey-Chmielewska H, Dzielulska A, Tomasiak M, Trybek G, Janiszewska-Olszowska J. Effect of nano-hydroxyapatite and ozone on approximal initial caries: a randomized clinical trial. *Sci Rep*, 2020, 10(1):11192. <https://doi.org/10.1038/s41598-020-67885-8> PMID: 32636425 PMCID: PMC7341873
- [12] Karlsson L, Kjaeldgaard M. Ozone treatment on dentin hypersensitivity surfaces – a pilot study. *Open Dent J*, 2017, 11:65–70. <https://doi.org/10.2174/1874210601711010065> PMID: 28356999 PMCID: PMC5345329
- [13] Raafat Abdelaziz R, Mosallam RS, Yousry MM. Tubular occlusion of simulated hypersensitive dentin by the combined use of ozone and desensitizing agents. *Acta Odontol Scand*, 2011, 69(6):395–400. <https://doi.org/10.3109/00016357.2011.572290> PMID: 21453220
- [14] Luppieri V, Manfra A, Ronfani L, Chermetz M, Cadenaro M. Ozone therapy for early childhood caries (ECC) treatment: an *in vivo* prospective study. *Appl Sci*, 2022, 12(4):1964. <https://doi.org/10.3390/app12041964> <https://www.mdpi.com/2076-3417/12/4/1964>
- [15] Floare AD, Scurtu AD, Balean OI, Chioran D, Buzatu R, Sava Rosianu R, Alexa VT, Jumanca D, Rusu LC, Racea RC, Coricovac D, Pinzaru I, Dehelean CA, Galuscan A. The biological effects of ozone gas on soft and hard dental tissues and the impact on human gingival fibroblasts and gingival keratinocytes. *Processes*, 2021, 9(11):1978. <https://doi.org/10.3390/pr9111978> <https://www.mdpi.com/2227-9717/9/11/1978>
- [16] Beniash E, Stifler CA, Sun CY, Jung GS, Qin Z, Buehler MJ, Gilbert PUPA. The hidden structure of human enamel. *Nat Commun*, 2019, 10(1):4383. <https://doi.org/10.1038/s41467-019-12185-7> PMID: 31558712 PMCID: PMC6763454
- [17] Li X, Wang J, Joiner A, Chang J. The remineralisation of enamel: a review of the literature. *J Dent*, 2014, 42(Suppl 1):S12–S20. [https://doi.org/10.1016/S0300-5712\(14\)50003-6](https://doi.org/10.1016/S0300-5712(14)50003-6) PMID: 24993850
- [18] Unal M, Oztas N. Remineralization capacity of three fissure sealants with and without gaseous ozone on non-cavitated incipient pit and fissure caries. *J Clin Pediatr Dent*, 2015, 39(4):364–370. <https://doi.org/10.17796/1053-4628-39.4.364> PMID: 26161609
- [19] Dudea D, Florea A, Miha C, Câmpeanu R, Nicola C, Benga Gh. The use of scanning electron microscopy in evaluating the effect of a bleaching agent on the enamel surface. *Rom J Morphol Embryol*, 2009, 50(3):435–440. PMID: 19690771
- [20] Ciocan DI, Stanciu D, Popescu MA, Miculescu F, Plotog I, Vârzaru G, Ciocan LT. Electron microscopy analysis of different orthodontic brackets and their adhesion to the tooth enamel. *Rom J Morphol Embryol*, 2014, 55(2 Suppl):591–596. PMID: 25178330
- [21] Holmes J. Clinical reversal of root caries using ozone, double-blind, randomised, controlled 18-month trial. *Gerodontology*, 2003, 20(2):106–114. <https://doi.org/10.1111/j.1741-2358.2003.00106.x> PMID: 14697022
- [22] Libonati A, Di Taranto V, Mea A, Montemurro E, Gallusi G, Angotti V, Nardi R, Paglia L, Marzo G, Campanella V. Clinical antibacterial effectiveness of Healozone Technology after incomplete caries removal. *Eur J Paediatr Dent*, 2019, 20(1):73–78. <https://doi.org/10.23804/ejpd.2019.20.01.14> PMID: 30919649
- [23] Srinivasan SR, Amaechi BT. Ozone: a paradigm shift in dental therapy. *J Global Oral Health*, 2019, 2(1):68–77. https://doi.org/10.25259/JGOH_56_2019 <https://jglobaloralhealth.org/ozone-a-paradigm-shift-in-dental-therapy/>

Corresponding authors

Ioana Cristina Niculescu Talpoș, Assistant Professor, DMD, PhD, 1st Department of Dentistry, Faculty of Dental Medicine, Victor Babeș University of Medicine and Pharmacy, 2 Eftimie Murgu Square, 300041 Timișoara, Romania; Phone +40722–811 688, e-mail: cristinaltalpos@yahoo.com

Octavia Iulia Bălean, Assistant Professor, DMD, PhD, 1st Department of Dentistry, Faculty of Dental Medicine, Victor Babeș University of Medicine and Pharmacy, 2 Eftimie Murgu Square, 300041 Timișoara, Romania; Phone +40748–677 191, e-mail: balean.octavia@umft.ro

Received: July 9, 2022

Accepted: December 26, 2022