



Ectopic Expression of the *ydaS* and *ydaT* Genes of the Cryptic Prophage Rac of *Escherichia coli* K-12 May Be Toxic but Do They Really Encode Toxins?: a Case for Using Genetic Context To Understand Function

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It was welcoming to read in *mSphere* the two recent publications (1, 2) from collaborative groups on their work on the essentiality and biology of the *Escherichia coli* Rac prophage putative repressor, RacR. My interest in the *rac* locus and its regulation stems from my recent demonstration that both the heat-labile and pertussis toxin-like enterotoxins of type II enterotoxigenic *E. coli* are encoded within full-sized Rac-like prophages (3). Over a decade ago, a computer algorithm (4) designed to identify potential toxin-antitoxin gene pairs flagged the *racR*-divergent *ydaST* genes of the Rac prophage as candidates, and the encoded polypeptides are still listed in PFAM (5) (PF15943, PF06254) as a toxin-antitoxin pair, despite a subsequent 2010 study finding no evidence for this (6). The two new RacR studies published in *mSphere* focus solely on the toxicity of these two genes at the expense of any insight into their toxicity, which could have been addressed by studying the corresponding loci in other temperate prophages of *E. coli*. Many prophages genes when ectopically expressed are detrimental to the bacterium but are likely to be beneficial (for the phage) during prophage induction. In the majority of lambdoid prophages, there are two functionally (but not always genetically) conserved transcriptional regulators divergently encoded from the phage repressor, the so-called phage immunity region (7). In the prototypical lambda phage, this locus encodes *ci* (repressor), *Cro* (lytic regulator), and *cII* (lysogenic regulator).

YdaS and *YdaT* were identified as *Cro* and *cII* homologs in the Rac prophage by Casjens (8) in 2003 and are also annotated as such in many databases, e.g., EcoGene (9). Coordinated expression of the lambda regulatory genes is essential for the lysis/lysogeny decision (10, 11), where the relative levels of *Cro* and *cII* proteins are controlled by both environmental inputs and other phage regulatory proteins (12–14). As recently shown for *ydaT* (15), overexpression of *cII* is highly toxic to *E. coli* (16, 17), yet *Cro* and *cII* are not considered toxins. It is highly likely that *racR*, *ydaS*, and *ydaT* form a similarly functioning regulatory locus. The *rac* regulatory circuit is fully functional—zygotic induction of prophage recombination genes occurs upon Hfr transfer of the *rac* locus to naive cells (18). That 1973 study identified the *rac* locus as the Rac prophage, named for recombination activation. Zygotic induction is lethal in up to 98% of recipient bacteria (19). Prophage repressors are essential only in the context of the lysogen (two other essential genes encode putative repressors of the defective prophages Qin and e14 [9]), and to focus on the phenotype of a particular locus without addressing its genetic context has led to what I consider the erroneous conclusion that the role of RacR is solely to repress the *ydaST* “toxin” genes—specifically, that the *ydaST-racR* module forms a “toxin-repressor” combination (1)—or that RacR-*YdaS*-*YdaT*

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form an atypical toxin-antitoxin system (2). Most likely, YdaS and YdaT proteins, while toxic to the host bacterium when overproduced or ectopically expressed, are actually essential regulatory components of the prophage genome.

REFERENCES

- Krishnamurthi R, Ghosh S, Khedkar S, Seshasayee ASN. 2017. Repression of YdaS toxin is mediated by transcriptional repressor RacR in the cryptic *rac* prophage of *Escherichia coli* K-12. *mSphere* 2:e00392-17. <https://doi.org/10.1128/mSphere.00392-17>.
- Bindal G, Krishnamurthi R, Seshasayee ASN, Rath D. 2017. CRISPR-Cas-mediated gene silencing reveals RacR to be a negative regulator of YdaS and YdaT toxins in *Escherichia coli* K-12. *mSphere* 2:e00483-17. <https://doi.org/10.1128/mSphere.00483-17>.
- Jobling MG. 2016. The chromosomal nature of LT-II enterotoxins solved: a lambdoid prophage encodes both LT-II and one of two novel pertussis-toxin-like toxin family members in type II enterotoxigenic *Escherichia coli*. *Pathog Dis* 74:ftw001. <https://doi.org/10.1093/femspd/ftw001>.
- Sevin EW, Barloy-Hubler F. 2007. RASTA-Bacteria: a web-based tool for identifying toxin-antitoxin loci in prokaryotes. *Genome Biol* 8:R155. <https://doi.org/10.1186/gb-2007-8-8-r155>.
- Finn RD, Bateman A, Clements J, Coghill P, Eberhardt RY, Eddy SR, Heger A, Hetherington K, Holm L, Mistry J, Sonnhammer ELL, Tate J, Punta M. 2014. Pfam: the protein families database. *Nucleic Acids Res* 42:D222–D230. <https://doi.org/10.1093/nar/gkt1223>.
- Christensen-Dalsgaard M, Jørgensen MG, Gerdes K. 2010. Three new RelE-homologous mRNA interferases of *Escherichia coli* differentially induced by environmental stresses. *Mol Microbiol* 75:333–348. <https://doi.org/10.1111/j.1365-2958.2009.06969.x>.
- Degnan PH, Michalowski CB, Babić AC, Cordes MHJ, Little JW. 2007. Conservation and diversity in the immunity regions of wild phages with the immunity specificity of phage λ . *Mol Microbiol* 64:232–244. <https://doi.org/10.1111/j.1365-2958.2007.05650.x>.
- Casjens S. 2003. Prophages and bacterial genomics: what have we learned so far? *Mol Microbiol* 49:277–300. <https://doi.org/10.1046/j.1365-2958.2003.03580.x>.
- Zhou J, Rudd KE. 2013. EcoGene 3.0. *Nucleic Acids Res* 41:D613–D624. <https://doi.org/10.1093/nar/gks1235>.
- Echols H. 1986. Bacteriophage λ development: temporal switches and the choice of lysis or lysogeny. *Trends Genet* 2:26–30. [https://doi.org/10.1016/0168-9525\(86\)90165-4](https://doi.org/10.1016/0168-9525(86)90165-4).
- Vohradsky J. 2017. Lambda phage genetic switch as a system with critical behaviour. *J Theor Biol* 431:32–38. <https://doi.org/10.1016/j.jtbi.2017.07.024>.
- Lee S, Lewis DEA, Adhya S. 2018. The developmental switch in bacteriophage λ : a critical role of the Cro protein. *J Mol Biol* 430:58–68. <https://doi.org/10.1016/j.jmb.2017.11.005>.
- Golding I. 2011. Decision making in living cells: lessons from a simple system. *Annu Rev Biophys* 40:63–80. <https://doi.org/10.1146/annurev-biophys-042910-155227>.
- Casjens SR, Hendrix RW. 2015. Bacteriophage lambda: early pioneer and still relevant. *Virology* 479–480:310–330. <https://doi.org/10.1016/j.virol.2015.02.010>.
- Campos M, Dobihal GS, Jacobs-Wagner C. 2017. Genome-wide phenotypic analysis of growth, cell morphogenesis and cell cycle events in *Escherichia coli*. *bioRxiv* <https://doi.org/10.1101/101832>.
- Rajamanickam K, Hayes S. 2018. The bacteriophage lambda CII phenotypes for complementation, cellular toxicity and replication inhibition are suppressed in cII-*oop* constructs expressing the small RNA OOP. *Viruses* 10:115. <https://doi.org/10.3390/v10030115>.
- Kedzierska B, Glinkowska M, Iwanicki A, Obuchowski M, Sojka P, Thomas MS, Wegrzyn G. 2003. Toxicity of the bacteriophage λ cII gene product to *Escherichia coli* arises from inhibition of host cell DNA replication. *Virology* 313:622–628. [https://doi.org/10.1016/S0042-6822\(03\)00376-3](https://doi.org/10.1016/S0042-6822(03)00376-3).
- Low B. 1973. Restoration by the *rac* locus of recombinant forming ability in *recB*—and *recC*—merozygotes of *Escherichia coli* K-12. *Mol Gen Genet* 122:119–130. <https://doi.org/10.1007/BF00435185>.
- Feinstein SI, Low KB. 1982. Zygotic induction of the *rac* locus can cause cell death in *E. coli*. *Mol Gen Genet* 187:231–235. <https://doi.org/10.1007/BF00331122>.