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## **Design, Synthesis, and Antiviral and Fungicidal Activities of 4‑Oxo‑4***H***‑quinolin-1-yl Acylhydrazone Derivatives**

[Peipei](https://pubs.acs.org/action/doSearch?field1=Contrib&text1="Peipei+Cui"&field2=AllField&text2=&publication=&accessType=allContent&Earliest=&ref=pdf) Cui,[\\*](#page-8-0) [Kaisi](https://pubs.acs.org/action/doSearch?field1=Contrib&text1="Kaisi+Liu"&field2=AllField&text2=&publication=&accessType=allContent&Earliest=&ref=pdf) Liu, [Zhaokai](https://pubs.acs.org/action/doSearch?field1=Contrib&text1="Zhaokai+Yang"&field2=AllField&text2=&publication=&accessType=allContent&Earliest=&ref=pdf) Yang, [Ping](https://pubs.acs.org/action/doSearch?field1=Contrib&text1="Ping+Sun"&field2=AllField&text2=&publication=&accessType=allContent&Earliest=&ref=pdf) Sun, [Yanan](https://pubs.acs.org/action/doSearch?field1=Contrib&text1="Yanan+Meng"&field2=AllField&text2=&publication=&accessType=allContent&Earliest=&ref=pdf) Meng, [Qilong](https://pubs.acs.org/action/doSearch?field1=Contrib&text1="Qilong+Yang"&field2=AllField&text2=&publication=&accessType=allContent&Earliest=&ref=pdf) Yang, [Xinyang](https://pubs.acs.org/action/doSearch?field1=Contrib&text1="Xinyang+Wu"&field2=AllField&text2=&publication=&accessType=allContent&Earliest=&ref=pdf) Wu, [Yongkang](https://pubs.acs.org/action/doSearch?field1=Contrib&text1="Yongkang+Lv"&field2=AllField&text2=&publication=&accessType=allContent&Earliest=&ref=pdf) Lv, Yan [Yang,](https://pubs.acs.org/action/doSearch?field1=Contrib&text1="Yan+Yang"&field2=AllField&text2=&publication=&accessType=allContent&Earliest=&ref=pdf)[\\*](#page-8-0) and [Jian](https://pubs.acs.org/action/doSearch?field1=Contrib&text1="Jian+Wu"&field2=AllField&text2=&publication=&accessType=allContent&Earliest=&ref=pdf) Wu[\\*](#page-8-0)



antiviral and fungicidal activities. Most acylhydrazone derivatives exhibited moderate to good antiviral activities in vivo. The inactive, curative, and protective activities of compounds 4 (51.2, 47.6, and 46.3%), 11 (49.6, 43.0, and 45.2% at 500 mg/L), and 17 (47.1, 49.2, and 44.1%) were higher than those of ribavirin (39.2, 38.0, and 40.8%) at 500 mg/L. Molecular docking showed that compound 4 exhibited a stronger affinity to TMV coat protein (TMV-CP) than ribavirin, with a binding energy (−6.89 kcal/mol)



slightly lower than that of ribavirin (−6.08 kcal/mol). Microscale thermophoresis showed that compound 4 ( $K_d$  = 0.142 ± 0.060 μM) exhibited a strong binding ability to TMV-CP, superior to that of ribavirin ( $K_d$  = 0.512  $\pm$  0.257 μM). The results of transmission electron microscopy showed that compound 4 hindered the self-assembly and growth of TMV. The antifungal activities of most compounds were moderate at 50 mg/L, among which compounds 12 and 21 exhibited a 72.1 and 76.5% inhibitory rate against *Physalospora piricola*, respectively. Meanwhile, compound 16 exhibited a 60% inhibitory rate against *Cercospora arachidicola Hori* at 50 mg/L.

### ■ **INTRODUCTION**

Plant virus diseases cause approximately 5 billion euros in economic losses to world agriculture every year.<sup>[1](#page-9-0)</sup> As a widely distributed virus worldwide, tobacco mosaic virus (TMV) can infect various Solanaceae plants, and its infection is difficult to control in the field. The actual inhibitory effect of commercially available TMV inhibitor Ningnanmycin is lower than  $60\%$  in the field.<sup>[2](#page-9-0)</sup> Meanwhile, its use is limited by water stickiness and photosensitivity. $3$  Based on this, it is necessary to develop efficient TMV inhibitors.

Due to their low toxicities, easy decompositions, and unique action mechanism, more and more natural-product-based antiviral agents have been developed and their antiviral mechanism has been investigated in recent years, $4$  including  $\alpha$ -amino phosphonates derivatives,<sup>5,[6](#page-9-0)</sup> chalcone derivatives,<sup>[7](#page-9-0),[8](#page-9-0)</sup> dithioacetal derivatives,<sup>[9](#page-9-0)-[11](#page-9-0)</sup> myricetin derivatives,<sup>[12](#page-9-0),[13](#page-9-0)</sup> ferulic acid derivatives,<sup>[14](#page-9-0)-[16](#page-9-0)</sup> indole derivatives,<sup>[17](#page-9-0),[18](#page-9-0)</sup> limonin deriva-tives,<sup>[19](#page-9-0)</sup> and so forth. However, few antiviral drugs are used in the field due to cost, synthetic complexity, and stability. $20,21$ Thus, it is still necessary to develop highly stable, easy to synthesize, and efficient plant-based virus inhibitors.

Quinoline alkaloids are widely distributed in nature and drugs, which exhibited broad insecticidal, antifungal, antiviral, antimalarial, anticancer, and anti-inflammatory activities.<sup>[22,23](#page-9-0)</sup>

Dictamnine (I) extracted from *D. dasycarpus* exhibited feeding deterrent activity against adults and larvae of *T. castaneum* as well as *S. zeamais* adults with EC<sub>50</sub> values of 57.6, 47.9, and 91.7 ppm, respectively. $24$  Liu et al. synthesized quinoline alkaloids II ( $EC_{50} = 0.41 \ \mu g/mL$ ) and III ( $EC_{50} = 0.55 \ \mu g/A$ mL) which displayed superior in vitro fungicidal activities against *Sclerotinia sclerotiorum*. [25](#page-9-0) Quinoline tricyclic derivative IV exhibited anti-HCV (Hepacivirus) ( $EC_{50} = 3.1 \mu M$ ) and anti-BVDV (Pestivirus) ( $EC_{50} = 1.2 \mu M$ ) activities.<sup>[26](#page-9-0)</sup> Wolf et al. synthesized quinoline derivative V which exhibited submicromolar antimalarial activity versus HB3 (chloroquine sensitive) (IC<sub>50</sub> = 17.5 nM) and Dd2 (chloroquine-resistant strains of *Plasmodium falciparum*)  $(IC_{50} = 22.7 \text{ nM})$  para-sites.<sup>[27](#page-9-0)</sup> Compound VI displayed promising cytotoxicity against PC-3 (IC<sub>50</sub> = 3.12  $\mu$ M), DU-145, NCIH460, and 4 T1 cell lines.<sup>[28](#page-9-0)</sup> Compound VII could inhibit TNF- $\alpha$  formation with an  $IC_{50}$  value of 2.3  $\mu$ M.<sup>[29](#page-9-0)</sup>

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Acylhydrazone structure is commonly used in drug design. In 2013, Kaplancikli et al. found that compound VIII exhibited larvicidal activity with  $LD_{50}$  values of 57.4 and 4.35 ppm and  $LD_{90}$  values of 297.8 and 19.1 ppm, respectively, at 24 and 48 h post treatment[.30](#page-10-0) In 2019, Yang et al. synthesized compound IX which exhibited higher curative, protective, and inactive activities against TMV and CMV than those of ningnanmycin.[31](#page-10-0) In 2024, Wu et al. found that arylhydrazone derivative X exhibited anti-TMV curative activity ( $EC_{50} = 139 \mu g/mL$ ), which was similar to that of ningnanmycin<sup>[32](#page-10-0)</sup> (Figure 1).

In our previous work, we found that the inactive, curative, and protective activities of echinopsine against TMV were 49.5, 46.1, and 42.6% at 500 mg/L, respectively.<sup>[33](#page-10-0)</sup> Further studies indicated that the inactive, curative, and protective activities of compound  $C$  (45.9, 31.5, and 47.8%) were also higher than those of commercial ribavirin (39.2, 38.0, and 40.8%) at 500 mg/L. Based on the broad spectrum bioactivities of quinoline and acylhydrazone structures, using compound C as the leading compound, a series of 4-oxo-4*H*quinolin-1-yl acylhydrazone derivatives were designed and synthesized (Figure 2). Their anti-TMV and fungicidal activities were evaluated for the first time.

### ■ **MATERIALS AND METHODS**

**Instruments.** <sup>1</sup> H NMR spectra were recorded at 400 MHz using a Bruker AV400 spectrometer in a CDCl<sub>3</sub> or DMSO- $d_6$ solution with tetramethylsilane as the internal standard. Highresolution mass spectrometry (HRMS) data were obtained on a Thermo Q Exactive Focus-MS instrument. The melting



Figure 2. Design of target compounds.

points were determined on an SWG X-4 binocular microscope melting point apparatus without correction.

*General Synthesis.* Ribavirin (Topscience Co., Ltd.), chlorothalonil (Bailing Agrochemical Co., Ltd.), carbendazim (Bailing Agrochemical Co., Ltd.), and other reagents were purchased from commercial sources and used as received. Echinopsine was synthesized according to literature.<sup>[34](#page-10-0)</sup> All anhydrous solvents were dried and purified according to the standard techniques.

*Synthesis of (4-Oxo-4H-quinolin-1-yl)-acetic Acid Ethyl Ester (B).* To a round bottomed flask was added *N,N*dimethylformamide (100 mL), compound A (4.35 g, 30 mmol), potassium carbonate (6.22 g, 45 mmol), and ethyl bromoacetate (8.02 g, 48 mmol). The reaction mixture was stirred for 12 h at room temperature. Water was added and the reaction mixture was extracted with dichloromethane three times. The organic phase was combined, washed with brine, dried over anhydrous  $Na<sub>2</sub>SO<sub>4</sub>$ , and evaporated under reduced pressure. The residue was subjected to column chromatography eluted with dichloromethane/methanol  $(v/v, 20/1)$  to give compound B as a white solid (5.36 g, 77.4%); mp 157−

<span id="page-2-0"></span>

Figure 3. Synthesis of 4-oxo-4*H*-quinolin-1-yl acylhydrazone.

158 °C. <sup>1</sup>H NMR (400 Hz, CDCl<sub>3</sub>): *δ* 8.45 (d, *J* = 8.0 Hz, 1H), 7.66−7.62 (m, 1H), 7.47 (d, *J* = 7.6 Hz, 1H), 7.40−7.37 (m, 1H), 7.19 (d, *J* = 8.4 Hz, 1H), 6.29 (d, *J* = 7.6 Hz, 1H), 4.77 (s, 2H), 4.24 (q, *<sup>J</sup>* <sup>=</sup> 7.2 Hz, 2H), 1.25 (t, *<sup>J</sup>* <sup>=</sup> 7.2 Hz, 3H). 13C NMR (100 Hz, CDCl3): *<sup>δ</sup>*: 178.4, 167.2, 143.7, 140.2, 132.5, 127.3, 127.1, 124.0, 114.6, 110.8, 62.4, 54.0, 14.1. ESI-HRMS  $(m/z)$ : calcd for C<sub>13</sub>H<sub>14</sub>NO<sub>3</sub> [M + H]<sup>+</sup>, 232.0974; found, 232.0965.

*Synthesis of (4-Oxo-4H-quinolin-1-yl)-acetic Acid Hydrazide (C).* Compound B (2.31 g, 10 mmol) and hydrazine hydrate (5 g, 80%, 100 mmol) were dissolved in methanol (300 mL). The reaction mixture was refluxed until compound B was completely consumed. The reaction mixture was cooled to room temperature, and methanol was evaporated under reduced pressure until a large amount of white solid precipitated. The residue was filtered under reduced pressure to afford compound C as a white solid (1.78 g, 81.8%). mp 240−241 °C. <sup>1</sup> H NMR (400 Hz, DMSO-*d*6): *δ* 9.53 (s, 1H), 8.17 (d, *J* = 7.2 Hz, 1H), 7.92 (d, *J* = 7.6 Hz, 1H), 7.72−7.68 (m, 1H), 7.44 (d, *J* = 8.8 Hz, 1H), 7.39−7.35 (m, 1H), 6.07  $(d, J = 7.6 \text{ Hz}, 1H)$ , 4.86 (s, 2H), 4.36 (s, 2H). <sup>13</sup>C NMR (100 Hz, DMSO-d<sub>6</sub>): δ 176.5, 166.0, 145.7, 140.3, 132.0, 126.5, 125.7, 123.3, 116.0, 108.7, 52.8. ESI-HRMS (*m*/*z*): calcd for  $C_{11}H_{12}N_3O_2$  [M + H]<sup>+</sup>, 218.0930; found, 218.0924.

**General Procedure for the Preparation of Compounds 1**−**25.** The mixture of methanol (50.0 mL), compound C (1 mmol), and aldehydes  $D_1-D_{25}$  (1 mmol) was refluxed for 12 h. Then, the reaction mixture was cooled to room temperature, and methanol was evaporated under reduced pressure until a large amount of solid precipitated. The residue was filtered under reduced pressure and the precipitate was washed with methanol to afford compounds 1− 25. The data of compounds 1−25 are presented in the Supporting [Information.](https://pubs.acs.org/doi/suppl/10.1021/acsomega.4c05046/suppl_file/ao4c05046_si_001.pdf)

**Biological Assay.** The anti-TMV and fungicidal activities of compounds were tested according to the methods reported in the literature,  $35,36$  $35,36$  which can also be found in the [Supporting](https://pubs.acs.org/doi/suppl/10.1021/acsomega.4c05046/suppl_file/ao4c05046_si_001.pdf) [Information](https://pubs.acs.org/doi/suppl/10.1021/acsomega.4c05046/suppl_file/ao4c05046_si_001.pdf).

**Morphological Observation via Transmission Electron Microscopy.** The samples were treated according to the method reported in the literature.<sup>37</sup> The support membrane was dipped in the compound solution for 10 s, then redyed using phosphotungstic acid, dried, and then observed under a transmission electron microscope. The details can be found in the Supporting [Information](https://pubs.acs.org/doi/suppl/10.1021/acsomega.4c05046/suppl_file/ao4c05046_si_001.pdf).

**Molecular Docking.** Molecular docking was performed on the MOE software. The structure preparation function in the MOE was used to prepare the protein structure. The preparation process included assessing the quality of the protein-structure data using defined temperature factors, protein geometry checks, and electron-density checks; adding hydrogen atoms and optimizing their positions; and performing the final energy minimization of the structure. Next, these ligands were minimized by the Amber 10 force field.

Following this, molecular docking was performed with the dock application of the MOE, with two rounds of calculation. A collection of poses were generated from the pool of ligand conformations using the Triangle Matcher method and were further refined using the rigid receptor method in MOE. Finally, the generalized-Born/volume-integral implicit-solvent model developed by Labute, GBVI/WSA dG, was used for scoring each of the generated poses. For each compound, the pose with the lowest score was retained. The compounds were ranked lowest to highest on the basis of their scores.

**Microscale Thermophoresis.** TMV coat protein (TMV-CP) was purified according to the procedure reported in a previous study.<sup>[38](#page-10-0)</sup> The required concentration of compound 4 was mixed with 10<sup>−</sup><sup>2</sup> mL of the same volume of labeled protein (TMV-CP) and incubated for 5 min. Samples were loaded into special glass capillaries, and the dissociation constant  $(K_d)$ values of compounds were measured at 25 °C via microscale thermophoresis using the Monolith NT.115 software (Nano Temper Technologies)[.39](#page-10-0),[40](#page-10-0) Additional details can be found in the Supporting [Information](https://pubs.acs.org/doi/suppl/10.1021/acsomega.4c05046/suppl_file/ao4c05046_si_001.pdf).

### ■ **RESULTS AND DISCUSSION**

**Synthesis of Target Compounds.** The synthetic route of 4-oxo-4*H*-quinolin-1-yl acylhydrazone is given in [Figure](#page-2-0) 3. The nucleophilic substitution of compound A and ethyl bromoacetate afforded compound B in 77.4% yield, which reacted with hydrazine hydrate to afford compound C in 81.8% yield. The condensation of compound C and aldehydes  $D_1-D_{25}$ afford compounds 1−25 in 42.8−95.5% yields, respectively. Compounds 1−25 precipitated from methanol, which made the purification of these compounds easy and suitable for largescale production.

**Configuration of Compounds 1**−**25.** It is interesting that compounds 1–25 exhibited two sets of signals in the  $^1\rm H$ NMR spectra at room temperature. Initially, this phenomenon was considered to be caused by the impurities in the compounds. Further purification of these compounds ruled out this possibility. According to previous literature,  $41,42$  $41,42$  the condensation of hydrazide and aldehyde afforded both cis and trans isomers (Figure 4), the mutual transformation of which



may be hindered at room temperature, resulting in two sets of peaks on <sup>1</sup>H NMR. The ratio of cis and trans isomers can be determined by integral area of peaks in  $^1\mathrm{H}$  NMR, which can be attributed to the fact that the relative thermodynamical stabilities of cis isomers and trans isomers were different. The hypothesis was confirmed by variable temperature  ${}^{1}\mathrm{H}$ NMR studies of compound 2 in DMSO- $d_6$  [\(Figure](#page-4-0) 5). The

coalescence of signals was observed in the  ${}^{1}H$  NMR spectrum with the increase of temperature  $(20-40-60-80 \degree \text{C})$ .

**Preliminary Structure**−**Activity Relationship Analysis.** *In Vivo Anti-TMV Activity.* The anti-TMV activities in vivo of compounds B, C, and 1−25 are listed in [Table](#page-5-0) 1. In order to make the antiviral activity results more reliable, commercial plant virus inhibitor ribavirin was taken as a control. All compounds exhibited moderate to good anti-TMV activities, especially, compound 4 (51.2, 47.6, and 46.3% at 500 mg/L; 16.8, 14.0, and 15.8% at 100 mg/L) and compound 11 (49.6, 43.0, and 45.2% at 500 mg/L; 19.8, 10.1, and 20.9% at 100 mg/L) exhibited higher inactive, curative, and protective activities against TMV than ribavirin (39.2, 38.0, and 40.8% at 500 mg/L; 12.1, 10.1, and 13.4% at 100 mg/L) both at 500 and 100 mg/L. Compound 17 (47.1, 49.2, and 44.1%) exhibited higher inactive, curative, and protective activities against TMV than ribavirin at 500 mg/L, while its inactive (7.5%) activity was lower than that of ribavirin (12.1%) at 100 mg/L. The inactive, curative, and protective activities of compound 23 (40.1, 32.5, and 43.9%) were equivalent to those of ribavirin at 500 mg/L, while the compound did not exhibit any inactive and protective activities at 100 mg/L. Biphenylsubstituted acylhydrazone derivative 11 (49.6, 43.0, and 45.2% at 500 mg/L) exhibited equivalent antiviral activity to corresponding 3-positon hydrazone derivative of echinopsine  $(46.2, 45.0, \text{ and } 41.7\% \text{ at } 500 \text{ mg/L}).^{33}$  $(46.2, 45.0, \text{ and } 41.7\% \text{ at } 500 \text{ mg/L}).^{33}$  $(46.2, 45.0, \text{ and } 41.7\% \text{ at } 500 \text{ mg/L}).^{33}$ 

The inactive, curative, and protective activities of hydrazide  $C$  (45.9, 31.5, and 47.8% at 500 mg/L) were higher than that of compound B, which indicated that the introduction of a hydrazide functional group increases the antiviral activity. Substituted benzaldehydes and heterocyclic aldehydes were selected to react with hydrazide C to investigate the effect of phenyl rings and heterocycles. Overall, introducing the substituted phenyl rings may increase or decrease antiviral activities, while introducing heterocycles generally makes the antiviral activity lower than compound C. For derivatives containing electron-donating groups substituted phenyl (2− 11), the structure−activity relationship shows the following: *m*-methoxy (4) > *p*-phenyl (11) > *p*-1-pyrazolyl (9) > nonsubstituent (1) > *p*-dimethylamino (8) > *p*-methyl (2) > *p-tert*-butyl (3) > methylthio (10). For derivatives containing electron-withdrawing groups substituted phenyl (12−18), the structure−activity relationship shows the following: *p*-trifluoromethyl (17) > *p*-methylsulfonyl (18) > *p*-bromo (14) > nonsubstituent (1) > *p*-fluoro (12) > *p*-chloro (13) > *p*-nitro (15). Thus, there was no obvious linear relationship between anti-TMV activity and electron-donating and electron-withdrawing abilities. The antiviral activities of derivatives containing monosubstituted benzene were higher than those containing polysubstituted benzene. For instance, compared with the disubstituted compound 5 (inactive, 11.3%, 500 mg/ L), the monosubstituted compound 4 (inactive, 51.2%, 500 mg/L) exhibited higher activity. For derivatives containing heterocycles, derivatives containing benzoheterocyclic rings exhibited higher antiviral activities than that containing furan, pyridine, thiazole, and thiophene.

**TEM Analysis.** The inhibitory effect of compound 4 and ribavirin (500 mg/L) on the morphology of TMV particles was observed by transmission electron microscopy (TEM) [\(Figure](#page-6-0) [6](#page-6-0)). TMV particles in the blank control exhibited a relatively complete rod shape, while TMV particles treated with a compound 4 solution with a concentration of 500 mg/ $L$ fragmented into shorter rods, similar to the effect of the 500

<span id="page-4-0"></span>



mg/L ribavirin control. It was speculated that compound 4 inhibited the assembly of TMV particles.

**Molecular Docking.** The molecular docking diagrams of ribavirin and compound 4 with TMV-CP are shown in [Figure](#page-6-0) [7](#page-6-0). Both compound 4 and ribavirin can be docked into the

pocket of TMV-CP. Compound 4 was larger than ribavirin so that it bonded more tightly to the protein pocket than ribavirin. Thus, the docking pocket between compound 4 and TMV-CP was more in line with the lock-key principle than that between ribavirin.

### <span id="page-5-0"></span>Table 1. In Vivo Antiviral Activities of Compounds B, C, and <sup>1</sup>−<sup>25</sup> against TMV*<sup>a</sup>*



<sup>a</sup>A one-way analysis of variance followed by Duncan's test was used for significant differences at  $p < 0.05$ , marking with the letters. Different letters indicate significant differences. <sup>b</sup>Ribavirin was used as the con

<span id="page-6-0"></span>

Figure 6. TEM images of TMV particles at 200 nm: (A) blank control, (B) compound 4, and (C) ribavirin.



Figure 7. Molecular docking diagrams of ribavirin with TMV-CP (A,C) and compound 4 with TMV-CP (B,D).

As shown in Figure 7C,D, the carbonyl group of the quinolone structure of compound 4 can form hydrogen bonding with amino acids Asn73, Tyr139, and Gln257. The benzene ring of compound 4 formed a Pi−cation interaction with Lys268. Compound 4 also had hydrophobic interactions with amino acids Glu131, Arg134, Glu222, and Lys253. Hydrogen bonding interactions, charge interactions, and hydrophobic interactions all played important roles in the binding of compound 4 and TMV-CP. Ribavirin formed hydrogen bonds with amino acids Arg134, Glu222, Ser255,

Asp266, and Lys268 and also formed Pi−anion interactions with amino acid Glu131.

Although compound 4 exhibited slightly fewer hydrogen bonding interactions with TMV-CP than ribavirin, the nonpolar interactions (van der Waals forces and hydrophobic interactions) between compound 4 and TMV-CP were superior to ribavirin. Compound 4 was more compatible with the binding pocket of TMV-CP and exhibited antiviral activities that were better than those of ribavirin. According to molecular simulation data in our previous work, $43$  the TMV-CP and compound 4 complex was stable.

**Microscale Thermophoresis Analysis.** Microscale thermophoresis results of compound 4 and ribavirin with TMV-CP are shown in Figure 8, which indicated that compound  $4(K_d =$  $0.142 \pm 0.060 \mu M$ ) showed stronger binding affinities toward TMV-CP than ribavirin ( $K_d$  = 0.512  $\pm$  0.257  $\mu$ M). These results were consistent with the molecular docking and anti-TMV inactivation activity results.

**Fungicidal Activity.** The fungicidal activities of compounds 1−25 were also evaluated and commercial fungicides carbendazim and chlorothalonil were used as the positive control [\(Table](#page-7-0) 2). All compounds exhibited moderate bactericidal activities against 14 kinds of phytopathogenic fungi, which were attributed to their poor solubilities. Compound 4 exhibited more than 40% inhibitory rate against three fungi at 50 mg/L. Compounds 15 and 16 exhibited 72.1 and 76.5% inhibitory rate against *Physalospora piricola* at 50 mg/L, respectively. Compound 19 exhibited a 60% inhibitory rate against *Cercospora arachidicola Hori* at 50 mg/L.

### ■ **CONCLUSIONS**

In summary, a series of 4-*oxo*-4H-quinolin-1-yl acylhydrazone derivatives were synthesized. Their anti-TMV activities and fungicidal activities were investigated. The bioassays results



Figure 8. Microscale thermophoresis results for compounds 4 (A) and ribavirin (B) with TMV-CP.

<span id="page-7-0"></span>

showed that compounds 4 and 11 exhibited higher inactive, curative, and protective activities than ribavirin both at 500 and 100 mg/L. The inactive, curative, and protective activities of compound 17 against TMV were higher than those of ribavirin at 500 mg/L, while its inactive activities were lower than those of ribavirin at 100 mg/L. Molecular docking, TEM, and microscale thermophoresis experiments showed that compound 4 hindered the self-assembly of TMV-CP and thus prevented TMV from infecting tobacco plants. Compounds 15 and 16 exhibited 72.1 and 76.5% inhibitory rate against *P. piricola* at 50 mg/L, respectively. Compound 19 exhibited a 60% inhibitory rate against *C. arachidicola Hori* at 50 mg/L. Further studies on structural optimization are in progress in our laboratory.

# ■ **ASSOCIATED CONTENT**<br>**<b>③** Supporting Information

The Supporting Information is available free of charge at [https://pubs.acs.org/doi/10.1021/acsomega.4c05046](https://pubs.acs.org/doi/10.1021/acsomega.4c05046?goto=supporting-info) .

> <sup>1</sup>H NMR, <sup>13</sup>C NMR, and ESI-HRMS data for compounds  $1-25$ ; <sup>1</sup>H NMR and <sup>13</sup>C NMR spectrum for compounds B, C, and 1-25; detailed bioassay procedures for the anti-TMV activities; and detailed bioassay procedures for the fungicidal activities ([PDF](https://pubs.acs.org/doi/suppl/10.1021/acsomega.4c05046/suppl_file/ao4c05046_si_001.pdf) )

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<span id="page-8-0"></span>Bc, Botrytis cinerea; Rs, Rhizoctonia solani; Fc, Fusarium oxysporium t. sp. cucumens; Ch, Cercospora arachidicola Hori; Pp, Physalospora piricola; Rc, Rhizoctonia cerealis; Bun, Bipolaris maydis; Wa, watermelon  $100\,\pm\,0.0$ "Average of three replicates; all results are expressed as mean ± SD. <sup>b</sup>As, Alternaria solarii Fg, Fusarium graminearum; Mg, magnaporthe grisea; Pc, Phytophthora capsici; Ss, Sclevotinia sclerotiorum; Bc, Bm, Bipolaris maydis; Wa, watermelor chlorothalonil 100 ± 0. Sclerotinia sclerotiorum;  $\text{Comp} \textbf{d}$  As<sup>*b*</sup>  $\text{Fg}^b$   $\text{Mg}^b$   $\text{Pc}^b$   $\text{Be}^b$   $\text{Be}^b$   $\text{Fe}^b$   $\text{Pc}^b$   $\text{Pc}^b$   $\text{Rc}^b$   $\text{Bm}^b$   $\text{Bm}^b$  $\text{F}_{\text{m}}$  $100\,\pm\,0.0$  $w_a^b$ Ss,  $1.2$  $86 \pm 1$ graminearum; Mg, magnaporthe grisea; Pc, Phytophthora capsici;  $Bm<sup>2</sup>$ Rc, Rhizoctonia cerealis;  $0.7$  $91 \pm$  $Rc^b$  $91\pm1.3$  $Pp^b$ Botrytis cinerea; Rs, Rhizoctonia solani; Fc, Fusarium oxysporium f. sp. cucumeris; Ch, Cercospora arachidicola Hori; Pp, Physalospora piricola;  $100 \pm 0.0$  $\operatorname{Ch}^b$  $100\,\pm\,0.0$  $F_c^b$ <sup>b</sup>As, Alternaria solani; Fg, Fusarium  $Rs$ <sup> $t$ </sup>  $50$  $0.0 \pm 0.0$  $Bc^b$  $\mathrm{^{15}S^2}$  $50$ results are expressed as mean  $\pm$  SD.  $73\pm1.3$  $\mathrm{Pc}^b$  $0.0\,\pm\,0.0$  $Mg^{b}$ anthracnose; and Fm, Fusarium moniliforme. *anthracnose*; and Fm, *Fusarium moniliforme*.  $73 \pm 0.9$  $\mathbb{A}^{\mathcal{A}}$  $\exists$ <sup>a</sup>Average of three replicates;  $0.0$  $100 \pm 0$  $As^b$ chlorothalonil  $Compd$ 

Table 2. continued

2. continued

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### **Notes**

The authors declare no competing financial interest.

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