

Impact of copper toxicity on stone-head cabbage (*Brassica oleracea* var. *capitata*) in hydroponics

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ABSTRACT

Arable soils are frequently subjected to contamination with copper as the consequence of imbalanced fertilization with manure and organic fertilizers and/or extensive use of copper-containing fungicides. In the present study, the exposure of stone-head cabbage (*Brassica oleracea var. capitata*) to elevated Cu^{2+} levels resulted in leaf chlorosis and lesser biomass yield at $\geq 2~\mu M$. Root nitrate content was not statistically affected by Cu^{2+} levels, although it was substantially decreased at $\geq 5~\mu M~Cu^{2+}$ in the shoot. The decrease in nitrate contents can be related to lower nitrate uptake rates because of growth inhibition by Cu-toxicity. Shoot sulfate content increased strongly at $\geq 2~\mu M~Cu^{2+}$ indicating an increase in demand for sulfur under Cu stress. Furthermore, at $\geq 2~\mu M$ concentration, concentration of water-soluble non-protein thiol increased markedly in the roots and to a smaller level in the shoot. When exposed to elevated concentrations of Cu^{2+} the improved sulfate and water-soluble non-protein thiols need further studies for the evaluation of their direct relation with the synthesis of metal-chelating compounds (i.e., phytochelatins).

Subjects Agricultural Science, Biochemistry, Plant Science, Soil Science, Toxicology **Keywords** Leaf chlorosis, Nutrient uptake, Non-protein thiol, Cu contamination, Biomass, Sulfur, *Brassica*, Copper, Thiols, Hydroponics, Toxicity, Cabbage

INTRODUCTION

Transition metals such as copper (Cu), zinc (Zn) and molybdenum (Mo) are essential for the growth and development of plants, but they rapidly get toxic at higher levels (*Kopsell & Kopsell*, 2007). Cu contamination in agricultural soils as a consequence of mining metals, dispersal of sewage sludge, arbitrary and improper application of agrochemicals, addition of organic fertilizers and frequent use of irrigation with low quality water is a well-known problem (*Dach & Starmans*, 2005; *Yruela*, 2009). Cu is a redox active metal that can exist in both Cu²⁺ and Cu⁺ forms in living organisms. At the protein level, Cu serves as a co-factor for various enzymes such as Cu/Zn-superoxide dismutase (Cu/ZnSOD), cytochrome c oxidase, ascorbate oxidase, amino oxidase, laccase, plastocyanin (PC), and polyphenol

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oxidase (*Yruela, 2005*; *Yruela, 2009*; *Pilon et al., 2006*). However, redox cycling between Cu²⁺ and Cu⁺ could induce oxidative stress by producing highly toxic hydroxyl radicals (*Yruela, 2005*; *Yruela, 2009*).

Plants exposed to elevated levels of Cu show unspecific toxicity symptoms. Elevated Cu levels in soils primarily result in stunted root growth and leaf chlorosis (*Kopsell & Kopsell, 2007; Shahbaz et al., 2010a*). Copper toxicity-induced reduction in chlorophyll contents hinder the development of chloroplast, thalakoid membrane and photosystem II (PSII), which are considered as the most sensitive Cu toxicity sites (*Pätsikkä, Aroan & Tyystjärvi, 1998; Pätsikkä et al., 2002; Burkhead et al., 2009; Yruela, 2005; Yruela, 2009; Shahbaz et al., 2010b*). At cellular level, toxicity may lead to binding of sulfhydryl groups in proteins, insufficiency or excess of other essential ions, oxidative damage and reduced cell transport (*De Vos et al., 1993; Yruela, 2009*). Furthermore, Cu-toxicity can change the mineral composition of plants. For instance, Fe contents may decrease in the shoot (*Pätsikkä, Aroan & Tyystjärvi, 1998; Pätsikkä et al., 2002; Kopsell & Kopsell, 2007; Shahbaz et al., 2010b*), Ca and Mg may decrease in the root and Zn contents may increase in both root and shoot upon Cu exposure at elevated levels (*Shahbaz et al., 2010b*).

Root growth is more severely affected by elevated Cu than shoot growth and the major proportion of Cu uptake retains in the root. Increased Cu contents in the plant tissues induce the synthesis of metal-binding compounds (*viz.* phytochelatins), which are most likely glutathione-derived compounds. (*Inouhe, 2005*; *Ernst et al., 2008*). Inductions of phytochelatins presume that more sulfur is needed for synthesis of these compounds, which results into higher absorption, and incorporation of sulfate. Nonetheless, the role of phytochelatins in detoxification of Cu is not very clear yet (*Ernst et al., 2008*; *Yruela, 2009*; *Shahbaz et al., 2010a*).

Brassica and other vegetable crops are often grown in the surrounding areas of big cities and industrial areas in developing countries like Pakistan, where they may be subjected to air and heavy metals pollution (Yang, Stulen & De Kok, 2006). The direct application of sewage water to vegetables is not only the source of many nutrients, but it is often contaminated with high levels of Cu and other heavy metals. As a result of continues untreated sewage application, heavy metals not only accumulate in the soil but also in vegetables (Younas et al., 1998; Butt et al., 2005). High Cu content in crop plants might not only negatively affect plant growth and functioning, but will also enter the food chain (Brun et al., 2001).

The present study used the hydroponics system which allows very close control over water soluble Cu and other mineral concentrations as compared to soil-grown system. Cabbage is a very important vegetable in all over the world. In Pakistan, cabbage is cultivated on almost 4.9 thousand hectares with 76.7 thousand tonnes annual production (FAO, 2013). The present study was aimed to investigate the response of growth, pigment contents and sulfur metabolism of stone-head cabbage grown in hydroponics to copper exposure.

MATERIAL AND METHODS

Stone-head cabbage (*Brassica oleracea var. capitata* F_1) seeds were germinated to sand in a green house. The seedlings collected at ten days after germination were transferred on an aerated 25% modified Hoagland nutrient solution in a 11 liter container (15.8" $L \times 10.3$ " $W \times 7$ "H; 3 plants set $^{-1}$ and 12 sets container $^{-1}$) in a greenhouse for 10 days. The nutrient solution consists of 1.25 mM Ca(NO₃)₂.4H₂O, 1.25 mM KNO₃, 0.25 mM KH₂PO₄, 0/0.5 mM MgSO₄.7H₂O, 11.6 μ M H₃BO₃, 2.4 μ M MnCl₂.4H₂O, 0.24 μ M ZnSO₄.7H₂O, 0.08 μ M CuSO₄.5H₂O, 0.13 μ M Na₂MoO₄.2H₂O and 22.5 μ M Fe³⁺-EDTA with supplemental concentrations of 0, 2, 5 and 10 μ M CuCl₂ and pH 5.9–6.0. The nutrient solution was continuously aerated with Aqua-Supreme—Air Pump—Model AP-4.The photoperiod was 14 h. 30 and 25 °C (\pm 5 °C) temperatures were set for day and night respectively, whereas the relative humidity was maintained at 60–70%.

Pigment contents

Whole shoot was homogenized (in 100% acetone 10 mL per g FW) followed by centrifugation at 800 g for 20 min. *Lichtenthaler* (1987) was followed for the determination of chlorophyll a, b and total carotenoid contents.

Nitrate and sulfate contents

Frozen root and shoot material was homogenized in de-mineralized water (10 mL per g fresh weight) and one layer of Miracloth filter was used to filter the homogenate. The supernatant was incubated in a water bath at 100 °C for 10 min. The remainder was centrifuged for 15 min (0 °C) at 30,000 g. The anions were separated by HPLC and *Maas et al.* (1986) was followed for their refractometric determination using a Knauer differential refractometer (model 98.00, Bad Homburg, Germany).

Water-soluble non-protein thiols

Extraction medium containing 80 mM sulfosalicylic acid, 1 mM EDTA, and 0.15% (w/v) ascorbic acid with an Ultra Turrax at 0 °C (10 mL per g fresh weight) was used for the homogenization of fresh plant matter. The resultant homogenous material was passed through one layer of Miracloth which was then centrifuged at 30,000 g for 15 min (0 °C). *De Kok, Buwalda & Bosma* (1988) was followed for the determination of total water-soluble non-protein thiol content colorimetrically at 413 nm after reaction with 5, 5′-dithiobis [2-nitrobenzoic acid].

RESULTS

Plant biomass in response to Cu exposure

Exposure of stone-head cabbage to higher concentrations of Cu^{2+} ($\geq 2~\mu M$) in nutrient solution caused chlorosis of both the shoot and young emerging leaves, that ultimately reduced both root and shoot biomass production (Fig. 1). A 10 day exposure to increasing Cu^{2+} concentrations in nutrient media led to a significant reduction of both root and shoot biomass production at $\geq 2~\mu M~Cu^{2+}$. Shoot to root ratio improved at $\geq 5~\mu M~Cu^{2+}$, demonstrating that when exposed to copper, root growth was more affected than shoot

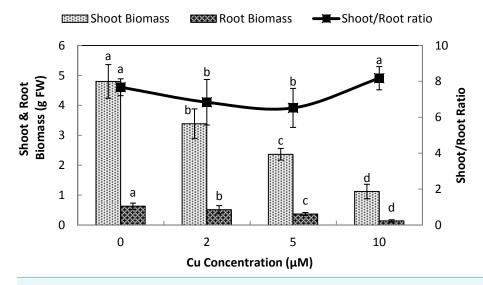


Figure 1 Impact of elevated levels of Cu^{2+} on biomass production of stone-head cabbage (*Brassica oleracea* var. *capitata*). 10-day-old seedlings of stone-head cabbage (*Brassica oleracea* var. *capitata*) were grown on a 25% Hoagland solution containing 0, 2, 5 and 10 μ M CuCl₂ in the root environment. Data on biomass production (g FW) and shoot/root ratio represent the mean of 2 independent experiments with 9 measurements having 3 plants in each treatment (\pm SD). Means with different letters differ significantly at $p \leq 0.01$ (Student's t-test).

growth (Fig. 1). Root dry matter content increased at 10 μ M Cu²⁺, whereas shoot dry matter content increased at >5 μ M Cu²⁺ (Fig. 2).

Pigment content in response to Cu exposure

The total chlorophyll (Chl. a+b) and carotenoid contents of stone-head cabbage were significantly decreased upon exposure at $\geq 2~\mu M~Cu^{2+}$ (Fig. 3). There were significant decreases in chlorophyll a/b and chlorophyll/carotenoid ratios when exposed to increased Cu^{2+} concentrations (10 $\mu M~Cu^{2+}$). Ten $\mu M~Cu^{2+}$ exposure resulted in the start of rapid development of shoot chlorosis and significantly faster reduction in chlorophyll a contents of chlorophyll b and carotenoids, ultimately leading to a significant reduction in chlorophyll a/b and chlorophyll/carotenoid ratios (Fig. 3).

Sulfate and water-soluble non-protein thiol contents in response to Cu exposure

Elevated Cu^{2+} levels showed a significant effect on concentration of the nitrate, sulfate and water-soluble non-protein thiol in stone-head cabbage. The nitrate contents of the roots showed a non significant response to the Cu exposure at different levels, however in shoots it were significantly decreased at \geq 5 μ M Cu^{2+} (Fig. 4). Sulfate contents in the roots were not affected; however, Cu^{2+} treatments of \geq 2 μ M substantially increased the sulfate contents of the shoot (Fig. 4). There was slight decrease in nitrate to sulfate ratio in the root and a strong decrease in the shoot when exposed to elevated levels of Cu (Fig. 4). Further-

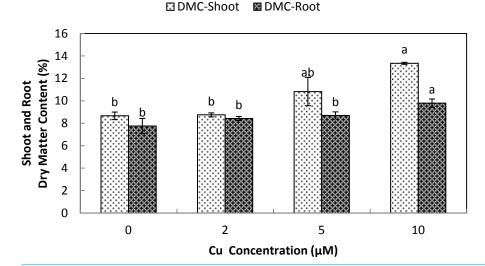


Figure 2 Impact of elevated levels of Cu^{2+} on dry matter content of stone-head cabbage (*Brassica oleracea* var. *capitata*). 10-day-old seedlings of stone-head cabbage (*Brassica oleracea* var. *capitata*) were grown on a 25% Hoagland solution containing 0, 2, 5 and 10 μ M CuCl₂ in the root environment. Data on dry matter content (%) represent the mean of 2 independent experiments with 9 measurements having 3 plants in each treatment (\pm SD). Means with different letters differ significantly at $p \le 0.01$ (Student's t-test).

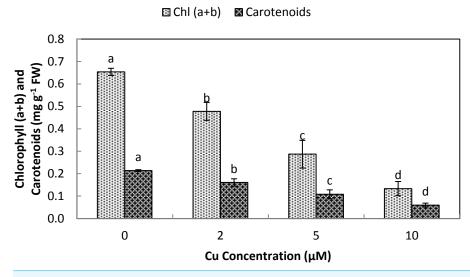


Figure 3 Impact of elevated levels of Cu^{2+} on pigment content (chl. a+b & carotenoids) of stone-head cabbage (*Brassica oleracea var. capitata*). 10-day-old seedlings of stone-head cabbage (*Brassica oleracea var. capitata*) were grown on a 25% Hoagland solution containing 0, 2, 5 and 10 μ M $CuCl_2$ in the root environment. Data on chlorophyll content (chl. a+b; mgg^{-1} FW) and carotenoid content (mgg^{-1} FW) represent the mean of 2 independent experiments with 9 measurements having 3 plants in each treatment (\pm SD). Means with different letters differ significantly at $p \leq 0.01$ (Student's t-test).

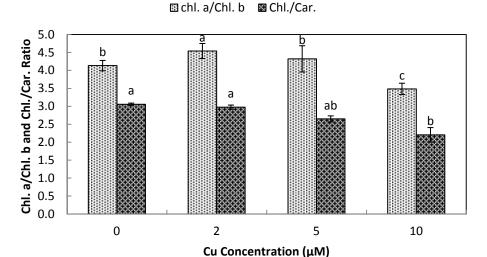


Figure 4 Impact of elevated levels of Cu^{2+} on pigment content (chl.a/chl.b & chl./car. ratio) of stone-head cabbage (*Brassica oleracea var. capitata*). 10-day-old seedlings of stone-head cabbage (*Brassica oleracea* var. *capitata*) were grown on a 25% Hoagland solution containing 0, 2, 5 and 10 μ M CuCl₂ in the root environment. Data on chlorophyll content (chl.a/chl.b and chl./carotenoid ratio) represent the mean of 2 independent experiments with 9 measurements having 3 plants in each treatment (\pm SD). Means with different letters differ significantly at $p \le 0.01$ (Student's t-test).

more, the exposure to $\geq 2 \,\mu\text{M} \,\text{Cu}^{2+}$ resulted in a solid raise in water-soluble non-protein thiol contents in the roots and to a smaller degree in the shoots at $10 \,\mu\text{M} \,\text{Cu}^{2+}$ (Fig. 5).

DISCUSSION

Cu exposure at elevated levels (<2 µM Cu²⁺) to stone-head cabbage significantly decreased the production of root and shoot biomass and raised the ratio of the shoot to the root. Copper contamination in the root environment generally results in retarded production of root and shoot biomass and a reduced photosynthetic activity. Moreover, it causes chlorosis, necrosis and bleaching of pigments (Yruela, 2005; Yruela, 2009; Sheldon & Menzies, 2005; Shahbaz et al., 2010a; Shahbaz et al., 2010b). In cabbage, the reduced production of biomass when exposed to elevated Cu levels coincided with decreased pigment contents (chl. a, b, carotenoids; Fig. 3) which may have resulted in reduced activity of photosynthesis and the dark respiration rate (Shahbaz et al., 2010a). It has been shown that Cu-toxicity damages chloroplasts either by inducing iron deficiency or by replacing Mg in the chlorophyll by Cu (*Pätsikkä et al.*, 2002; Küpper et al., 2003). Cu exposure at elevated levels not only decreased the pigment content but there was also a change in pigment composition. Chlorophyll a content decreased significantly faster than that of chlorophyll b and carotenoids, which resulted in a decreased chlorophyll a/b and chlorophyll/carotenoid ratio. Similar results were reported by Chu et al. (2006) in Trifolium repens L.

It is shown that the production of root biomass was more influenced than that of the shoot biomass production. The relatively higher reduction in the root biomass upon

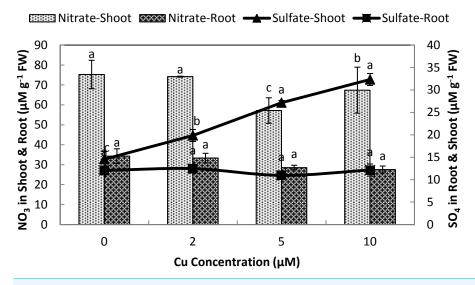


Figure 5 Impact of elevated levels of Cu^{2+} on nitrate and sulfate content of stone-head cabbage (*Brassica oleracea* var. *capitata*). 10-day-old seedlings of stone-head cabbage (*Brassica oleracea* var. *capitata*) were grown on a 25% Hoagland solution containing 0, 2, 5 and 10 μ M $CuCl_2$ in the root environment. Data on nitrate and sulfate content (μ mol g⁻¹ FW) represent the mean of 2 independent experiments with 9 measurements having 3 plants in each treatment (\pm SD). Means with different letters differ significantly at $p \le 0.01$ (Student's t-test).

exposure to metal contamination could be due the fact that roots come in direct contact with toxic metals (Cd, Cu). Toxic metal-induced hindered root growth also reduces the uptake of essential nutrients (*Sheldon & Menzies*, 2005).

Plants have evolved a tightly-controlled mechanism for the absorption, allocation and assimilation of sulfate under normal conditions. (Hawkesford & De Kok, 2006). Enhanced exposure of cabbage to Cu concentrations considerably affects the contents and allocation of sulfur compounds in the root and shoot of cabbage (Shahbaz et al., 2010a). The raised shoot sulfate contents might be attributed to Cu-toxicity induced upregulation of the sulfate suppliers in roots (Shahbaz et al., 2010a). To maintain rapid growth rates under stress conditions, Brassica species increase their demand for sulfur supply (Koralewska et al., 2008; Koralewska et al., 2009). Exposure of stone-head cabbage to elevated levels of Cu did not affect the nitrate contents in roots, however at $\geq 5 \,\mu M \, \text{Cu}^{2+}$ there was a significant decrease in the shoot nitrate contents. The reduced production of plant biomass at $\geq 5 \,\mu\text{M} \,\text{Cu}^{2+}$ could be attributed to reduced supply of nitrate in the shoot. The decrease in nitrate: sulfate ratio in the shoot of stone-head cabbage under elevated Cu levels may be attributed to enhanced sulfate contents, however the link between uptake rates of nitrate and sulfate is not evident yet (Stulen & De Kok, 2012). Since both nitrate and sulfate are involved in amino acid and protein synthesis, their uptake rates are related with growth rates (Stulen & De Kok, 2012).

Roots accumulated a slightly raised level of water-soluble non-protein thiols when compared with that of the shoot. Only a small proportion of the rise in thiol content might be attributed to a Cu-induced synthesis of phytochelatins in Chinese cabbage (Shahbaz et al., 2010a). Previous reports suggest that the formation of sulfur-rich metal-chelating compounds (i.e., water-soluble non-protein thiols) when disclosed to prospective toxic metals may perhaps require an enhanced demand for sulfur, viz. the absorption rate and incorporation of sulfate (Sirko & Gotor, 2007; Ernst et al., 2008). The possible significance of sulfur nutrition upon elevated copper exposure needs further investigation.

CONCLUSIONS

This investigation has shown that the elevated copper levels in the root surroundings proved toxic for stone-head cabbage. Copper exposure at $\geq 2~\mu M~Cu^{2+}$ negatively affected the plant biomass production and pigment contents. Furthermore, elevated Cu content considerably affected the concentration of sulfate and water-soluble non-protein thiol of stone-head cabbage which might be due to the induction of phytochelatins to detoxify excess copper.

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Competing Interests

The authors declare there are no competing interests.

Author Contributions

- Sajid Ali conceived and designed the experiments, wrote the paper, prepared figures and/or tables, reviewed drafts of the paper.
- Muhammad Shahbaz conceived and designed the experiments, performed the experiments, wrote the paper.
- Ahmad Naeem Shahzad and Ammara Fatima analyzed the data.
- Hafiz Azhar Ali Khan contributed reagents/materials/analysis tools, wrote the paper, prepared figures and/or tables, reviewed drafts of the paper.
- Moazzam Anees contributed reagents/materials/analysis tools.
- Muhammad Saleem Haider provided the facilities (laboratory and staff) for the research work.

Supplemental Information

Supplemental information for this article can be found online at http://dx.doi.org/10.7717/peerj.1119#supplemental-information.

REFERENCES

- Brun LA, Maillet J, Hinsiger P, Pépin N. 2001. Evaluation of copper availability to plants in copper-contaminated vineyard soils. *Environmental Pollution* 111:293–302 DOI 10.1016/S0269-7491(00)00067-1.
- Burkhead J, Reynolds KAG, Abdel-Ghany SE, Cohu CM, Pilon M. 2009. Copper homeostasis. *New Phytologist* **182**(4):799–816 DOI 10.1111/j.1469-8137.2009.02846.x.
- **Butt MS, Sharif K, Bajwa BE, Aziz A. 2005.** Hazardous effects of sewage on the environment: focus on heavy metals and chemical composition of soil and vegetables. *Management of Environmental Quality* **16**:338–346 DOI 10.1108/14777830510601217.
- Chu L, Liu DY, Wang YB, Ding JH, Wang LL. 2006. Separate and combined effects of Cu and Cd on seedling growth and active oxygen metabolism system of *Trifolium repens* L. *Frontiers in Bioscience* 11:2861–2867 DOI 10.2741/2014.
- **Dach J, Starmans D. 2005.** Heavy metals balance in Polish and Dutch agronomy: actual state and provisions for the future. *Agriculture Ecosystems and Environment* **107**:309–316 DOI 10.1016/j.agee.2005.02.017.
- **De Kok LD, Buwalda F, Bosma W. 1988.** Determination of cysteine and its accummulation in spinach leaf tissue upon exposure to excess sulfur. *Journal of Plant Physiology* **133(4)**:502–505 DOI 10.1016/S0176-1617(88)80045-2.
- **De Vos CHR, Ten Bookum TM, Vooijs R, Schat H, De Kok LD. 1993.** Effect of copper on fatty acid composition and peroxidation of lipids in roots of copper tolerant and sensitive *Silenecucubalus*. *Plant Physiology and Biochemistry* **31(2)**:151–158.
- Ernst WHO, Krauss GJ, Verkleij GAC, Wesenberg D. 2008. Interaction of heavy metals with the sulphur metabolism in angiosperms from an ecological point of view. *Plant Cell and Environment* 31(1):123–143.
- FAO. 2013. FAOSTAT Domains. Available at http://faostat3.fao.org/download/Q/QC/E.
- **Hawkesford MJ, De Kok LJ. 2006.** Managing sulfur metabolism in plants. *Plant Cell and Environment* **29(3)**:382–395 DOI 10.1111/j.1365-3040.2005.01470.x.
- **Inouhe M. 2005.** Phytochelatins. *Brazilian Journal of Plant Physiology* **17(1)**:65–78 DOI 10.1590/S1677-04202005000100006.
- **Kopsell DE, Kopsell DA. 2007.** Copper. In: Barker AV, Pilbeam DJ, eds. *Handbook of plant nutrition*. Boca Raton: Taylor and Francis Group, 293–328.
- Koralewska A, Buchner P, Stuiver CEE, Posthumus FS, Kopriva S, Hawkesford MJ, De Kok LJ. 2009. Expression and activity of sulfate transporters and APS reductase in curly kale in response to sulfate deprivation and re-supply. *Journal of Plant Physiology* 166(2):168–179 DOI 10.1016/j.jplph.2008.03.005.
- Koralewska A, Stuiver CEE, Posthumus FS, Kopriva S, Hawkesford MJ, De Kok LJ. 2008.

 Regulation of sulfate uptake, expression of the sulfate transporters Sultr1;1 and Sultr1;2, and APS reductase in Chinese cabbage (*Brassica pekinensis*) as affected by atmospheric H₂S nutrition and sulfate deprivation. *Functional Plant Biology* **35(4)**:318–327 DOI 10.1071/FP07283.
- Küpper H, Šetlík E, Šetliková N, Ferimazova M, Küpper FC. 2003. Copper-induced inhibition of photosynthesis: limiting steps of *in vivo* copper chlorophyll formation in *Scenedesmus quadricauda*. Functional Plant Biology 30(12):1187–1196 DOI 10.1071/FP03129.
- **Lichtenthaler HK. 1987.** Chlorophylls and carotenoids: pigments of the photosynthetic biomembranes. *Methods in Enzymology* **148**:350–382 DOI 10.1016/0076-6879(87)48036-1.

- Maas FM, Hoffmann I, Van Harmelen MJ, De Kok LJ. 1986. Refractometric determination of sulphate and other anions in plants separated by High-Performance Liquid Chromatography. *Plant and Soil* 91(1):129–132 DOI 10.1007/BF02181825.
- Pätsikkä E, Aroan EM, Tyystjärvi E. 1998. Increase in the quantum yield of photoinhibition contributes to copper toxicity *in vivo*. *Plant Physiology* 117(2):619–627 DOI 10.1104/pp.117.2.619.
- Pätsikkä E, Kairavuo MF, Šerešen EM, Tyystjärvi E. 2002. Excess copper predisposes photosystem II to photoinhibition *in vivo* by outcompeting iron and causing decrease in leaf chlorophyll. *Plant Physiology* **129**(3):1359–1367 DOI 10.1104/pp.004788.
- Pilon M, Abdel-Ghany SE, Cohu CM, Gogolin KA, Ye H. 2006. Copper cofactor delivery in plant cells. *Current Opinion in Plant Biology* 9:256–263 DOI 10.1016/j.pbi.2006.03.007.
- Shahbaz M, Tseng MH, Stuiver CEE, Koralewska A, Posthumus FS, Venema JH, Parmar S, Hawkesford MJ, De Kok LJ. 2010a. Copper exposure interferes with the regulation of the uptake, distribution and metabolism of sulfate in Chinese cabbage. *Journal of Plant Physiology* 167(6):438–446 DOI 10.1016/j.jplph.2009.10.016.
- Shahbaz M, Tseng MH, Stuiver CEE, Posthumus FS, Parmar S, Koralewska A, Hawkesford MJ, De Kok LJ. 2010b. Impact of copper exposure on physiological functioning of Chinese cabbage (*Brassica pekinensis*). In: Sequi P, Ferri D, Rea E, Montemurro AV, Fornado F, eds. 18th international symposium of CIEC. Fertilitas Agrorum, 318–324.
- Sheldon A, Menzies NW. 2005. The effect of copper toxicity on growth and morphology of Rhodes grass (*Chlorisgayana*) in solution culture. *Plant and Soil* 278(1–2):341–349 DOI 10.1007/s11104-005-8815-3.
- **Sirko A, Gotor C. 2007.** Molecular links between metals in the environment and plant sulfur metabolism. In: Hawkesford MJ, De Kok J, eds. *Sulfur in plants—an ecological perspective*. Springer, 169–195.
- Stulen I, De Kok LJ. 2012. Exploring interactions between sulfate and nitrate uptake at a whole plant level. In: De Kok LJ, Tausz M, Hawkesford MJ, Hoefgen R, McManus MT, Norton RM, Rennenberg H, Saito K, Schnug E, Tabe L, eds. Sulfur metabolism in plants: mechanisms and application to food security, and response to climate change. Dordrecht: Springer, 1–8. Available at www.fao.org (http://faostat3.fao.org/download/Q/QC/E) (accessed 23 June 2015).
- Yang L, Stulen I, De Kok LJ. 2006. Impact of sulfate nutrition on utilization of atmospheric SO₂ as a sulfur source for Chinese cabbage. *Journal of Plant Nutrition and Soil Science* 169:529–534 DOI 10.1002/jpln.200520574.
- **Younas M, Shahzad F, Afzal S, Khan MI, Ali K. 1998.** Assessment of Cd, Ni, Cu and Pb pollution in Lahore, Pakistan. *Environment International* **24**:761–766

 DOI 10.1016/S0160-4120(98)00068-3.
- **Yruela I. 2005.** Copper in plants. *Brazilian Journal of Plant Physiology* **17(1)**:145–156 DOI 10.1590/S1677-04202005000100012.
- **Yruela I. 2009.** Copper in plants: acquisition, transport and interactions. *Functional Plant Biology* **36**(5):409–430 DOI 10.1071/FP08288.