



Therapeutic Effects of Mesenchymal Stromal Cell-Derived Small Extracellular Vesicles in Oxygen-Induced Multi-Organ Disease: A Developmental Perspective

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OPEN ACCESS

Edited by:

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Reviewed by:

Fabio Mosca, University of Milan, Italy Gianandrea Pasquinelli, University of Bologna, Italy

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Specialty section:

This article was submitted to Stem Cell Research, a section of the journal Frontiers in Cell and Developmental Biology

> Received: 28 December 2020 Accepted: 17 February 2021 Published: 16 March 2021

Citation:

Fernandez-Gonzalez A, Willis GR, Yeung V, Reis M, Liu X, Mitsialis SA and Kourembanas S (2021) Therapeutic Effects of Mesenchymal Stromal Cell-Derived Small Extracellular Vesicles in Oxygen-Induced Multi-Organ Disease: A Developmental Perspective. Front. Cell Dev. Biol. 9:647025. doi: 10.3389/fcell.2021.647025 Despite major advances in neonatal intensive care, infants born at extremely low birth weight still face an increased risk for chronic illness that may persist into adulthood. Pulmonary, retinal, and neurocognitive morbidities associated with preterm birth remain widespread despite interventions designed to minimize organ dysfunction. The design of therapeutic applications for preterm pathologies sharing common underlying triggers, such as fluctuations in oxygen supply or in the inflammatory state, requires alternative strategies that promote anti-inflammatory, pro-angiogenic, and trophic activities-ideally as a unitary treatment. Mesenchymal stem/stromal cell-derived extracellular vesicles (MEx) possess such inherent advantages, and they represent a most promising treatment candidate, as they have been shown to contribute to immunomodulation, homeostasis, and tissue regeneration. Current pre-clinical studies into the MEx mechanism of action are focusing on their restorative capability in the context of preterm birth-related pathologies, albeit not always with a multisystemic focus. This perspective will discuss the pathogenic mechanisms underlying the multisystemic lesions resulting from early-life disruption of normal physiology triggered by high oxygen exposures and pro-inflammatory conditions and introduce the application of MEx as immunomodulators and growth-promoting mediators for multisystem therapy.

Keywords: development, BPD, preterm, oxygen, inflammation, multiorgan, MEx

INTRODUCTION

The survival of premature infants born at extremely low birth weight (ELBW) has improved dramatically due to recent advances in neonatal care (Matthews et al., 2015). However, 40% of infants born before 28 weeks of gestational age (GA < 28) are at risk of one or more long-term complications, as they often require prolonged mechanical ventilation and exposure to high and variable oxygen tension (Younge et al., 2017). Preterm ELBW infants, even those who do not

require supplemental oxygen, are exposed to "perceived" hyperoxia that often causes direct tissue damage and/or triggers maladaptive physiological responses. Such disruption not only impacts lung growth but can also lead to multiorgan dysfunction including cardiac, retinal, and neurological deficits as well as gastrointestinal and renal abnormalities (Weinberger et al., 2002; Johnson et al., 2011). Bronchopulmonary dysplasia (BPD) and other morbidities associated with prematurity, such as retinopathy of prematurity (ROP) and neurodevelopmental disabilities, share a common pathogenesis (i.e., inflammation and oxygen toxicity) not adequately addressed with current established therapies. Therapeutic advances based on new concepts and less invasive techniques are being currently tested to prevent BPD injury and the consequences of postdelivery respiratory care but do not target the underpinnings of perinatal diseases that accompany BPD. Moreover, the contribution of placental, microbial, and immune factors to the extrapulmonary sequelae of prematurity is just being recognized, and it is prompting the search for new therapeutic options (Collins et al., 2017).

Mesenchymal stem/stromal cell (MSC)-derived extracellular vesicles (MEx) have shown considerable anti-inflammatory and regenerative capacity in preclinical animal models, (Lee et al., 2012; Sdrimas and Kourembanas, 2014; Willis et al., 2018b), and a safety study of stem cell-derived extracellular vesicles in preterm neonates at high risk for BPD is currently ongoing (NCT03857841). The MEx advantage lies in their potential to modify immune, vascular, and parenchymal processes precipitated by preterm birth and exacerbated by oxygen fluctuations. This article will incorporate existing knowledge of the outcomes of prematurity and BPD as relating to MEx treatments in preclinical models of neonatal diseases, provide new information and perspective on the topic, and conclude by advancing the postulation that MEx can serve as a unitary therapeutic vector against systemic multi-organ pathologies of prematurity with common underlying triggers.

PREMATURITY AND ASSOCIATED ORGAN PATHOLOGY

BPD is a major cause of morbidity and mortality in surviving extremely preterm infants. Its incidence has increased between 2009 and 2012 (Stoll et al., 2015). BPD pathogenesis is triggered by prenatal factors, postnatal mechanical trauma, and hyperoxia during mechanical ventilation and is exacerbated by other stressors, including infection, inflammation, and pulmonary volume overload. The pathology of BPD has evolved during the past 50 years, thanks to the introduction of surfactant therapy (Jobe, 1999) and improvements in prenatal and neonatal care. Infants born during the late canalicular and early saccular stage of normal lung development are likely to develop BPD pathology that includes a reduced number of alveoli and secondary septa. The large decrease in surface area is associated with reduced and dysmorphic pulmonary microvasculature, leading to increased vascular resistance and pulmonary hypertension (Mourani et al., 2015). Survivors with severe BPD also manifest a deterioration of

lung function and a higher risk of developing chronic obstructive pulmonary disease later in life (Broström et al., 2010).

The pathologic processes driving abnormal lung growth affect the development of other systems as well. It is estimated that about 25–50% of surviving preterm infants suffer neurodevelopmental deficits such as cognitive deterioration (Larroque et al., 2008; Linsell et al., 2018), functional disability (Johnson et al., 2009), behavioral and psychological abnormalities that manifest later in life (Olsen et al., 2018), and, in a small proportion of cases, development of cerebral palsy and motor impairment (Jarjour, 2015).

An additional outcome of extreme preterm survival and supplemental oxygen is the occurrence of ROP (Silverman, 2004). Advanced ROP may lead to retinal detachment and significant visual impairment (Tin et al., 2001; Sommer et al., 2014). Although, clinically, ROP is considered a vascular disease precipitated by the premature exposure to high oxygen tension, there is evidence that neurons within the retina are also affected, causing a reduction in visual acuity, perception, and performance with increasing age (Leung et al., 2018).

Current therapies designed to treat and prevent BPD and other prematurity-related complications attempt to ameliorate acute stress reactions and rapidly improve organ function. An update on the current pharmacological therapies for BPD has been reported by Michael and colleagues (Michael et al., 2018). For example, the use of postnatal corticosteroids reduces inflammation and facilitates extubation in ventilatordependent preterm infants (Doyle et al., 2006). Milder ventilation approaches, targeted oxygen saturation levels, growth factors, antioxidants, and other drug therapies are also being considered as less invasive alternatives for the prevention and treatment of BPD (Collins et al., 2017). Anti-vascular endothelial growth factor (VEGF) and erythropoietin therapies are used in the treatment of ROP and preterm brain injury, respectively (Patel et al., 2016; Younge et al., 2017). However, these interventions have limited efficacy as they only target isolated organs and stages of the disease. More importantly, they may contribute to the development of a "new" and "evolving" pathophysiology of prematurity in which the long-term outcomes are difficult to predict. Preterm birth is associated with arrested structural or functional development of key organs, causing impairment that will likely persist into adulthood. Thus, the development of new protective strategies requires an understanding of the molecular and cellular vulnerabilities of developing organs during increased oxygenation and concomitant inflammation with the goal of finding common pathogenic mechanisms amenable to therapeutic intervention.

ORGAN DEVELOPMENT AND OXYGEN VULNERABILITY

The transition from *in utero* environment to extrauterine life in the preterm born neonate triggers a sudden rise of lung and systemic oxygen tension, and supplemental oxygen application intensifies their vulnerability to injury. Postnatal development of several organ systems including the lung, brain, gastrointestinal tract, and lymphoid organs occurs during defined and particularly vulnerable stages in humans and rodents (Picut and Parker, 2017). Oxygen fluctuations during this stage fuel an inflammatory response and cause the arrested growth of parenchyma as well as dysregulated angiogenic and immune responses which can be regarded as having a common mechanistic origin.

Lung

Contrary to what occurs in full-term human neonates that are born during the alveolar stage, rodents are born at a term during the saccular stage of lung development. From approximately postnatal (P) day 4-P21, the sacculae undergo thinning and reconstruction in a process termed alveolarization. Capillaries also grow superimposed to the process of alveolarization to create a network for blood supply and air gas exchange. The lungs of human preterm newborns experience an increase in arterial oxygen tension, even without supplemental oxygen, which leads to arrested alveolarization and vascularization. Preclinical studies have demonstrated that low antioxidant levels underlie the activation of transcription factors and pathways leading to endothelial and alveolar type II cell dysfunction and survival and the inactivation of surfactant (Stenmark and Abman, 2005). Exposure to hyperoxia also results in pulmonary accumulation of inflammatory cells and increased cytokine production (Ambalavanan et al., 2009).

Brain

In humans, the period of rapid brain growth and therefore greater vulnerability occurs during the last 3 months of pregnancy and continues postnatally with outgrowth and reorganization of the central nervous system. Similarly, although rodent brain development is generally completed at birth, cell apoptosis, neuronal pruning, and cell migration are still occurring by P7 and not completed until P14, while myelination progresses beyond P21, the equivalent of a 2-year-old human (Semple et al., 2013). Moreover, blood vessels in the rodent brain develop in parallel with the cortical parenchyma, due in part to their association with microglia during the first week of postnatal development (Mondo et al., 2020). A major pathological feature of preterm brain injury and early postnatal exposure to toxic levels of oxygen is the presence of white matter lesions. Oligodendrocyte precursors are highly sensitive to oxygen fluctuations (Back and Miller, 2014), and hyperoxia induces their apoptosis and reduces their proliferation, causing white matter deficiencies that persist well beyond the juvenile age (Schmitz et al., 2011). In addition, other cellular changes including abnormal glial maturation and phenotype (Schmitz et al., 2011; Leaw et al., 2017), cell death (Truttmann et al., 2020), and disrupted cortical development and neuronal maturation have been described in preclinical and preterm brain injury (Fletcher et al., 2017; Fleiss et al., 2020).

Retina

Hyperoxia also plays an initiating role in the pathogenesis of ROP in preterm infants and in experimental models of oxygen induced-retinopathy (OIR) (Vessey et al., 2011; Zin and Gole, 2013). Retinal vascularization in humans is completed at 36–40 weeks post-conceptual age. Preterm birth suddenly exposes the incompletely vascularized retina to hyperoxia, leading to vascular structural and functional compromise. Glial cells have been implicated in the genesis of retinal pathology in animal models of OIR, as they support the development of the vasculature and regulate cell apoptosis and neuronal metabolism and activation (Vessey et al., 2011; Shen et al., 2012). Müller cells express angiogenic growth factors such as VEGF and up-regulate the expression of glial fibrillary acidic protein (GFAP) as a hallmark of reactive gliosis in response to pathological oxygen tension. Microglia, like those in the brain, are generally considered immune cells, but they also contribute to the vascular and neuronal pathologic effects of oxygen-induced toxicity.

It is clear that perinatal oxidant stress, inflammation, and other toxic stimuli adversely impact the premature neonate cared for in modern neonatal intensive care units, and these result in acute and long-term injuries. The lack of regenerative therapies addressing the multiple organ pathologies occurring during this critical developmental stage highlights the need to consider a new, for the lack of a better term, "holistic" approach that can redirect normal development and enhance early organ restoration.

MEX AS A REGENERATIVE AGENT FOR PERINATAL PATHOLOGIES

Therapies based on the MSC secretome have gained increased attention as promising candidates for a multifaceted approach, impacting multiple pathological aspects of premature infant disease. MSCs, as most cells, secrete a plethora of heterogenous extracellular vesicles (EVs) of diverse biogenetic origins. The molecular composition and associated bioactive cargo of each EV class reflect both their biogenesis as well as the specific stimulus triggering their formation. EV biogenesis and nomenclature have been extensively reviewed by Yeung et al. (2019). On a gross level, EVs are categorized into three subclasses: small EVs (sEVs) (~30-150 nm in diameter) represent the class that includes the exosomes, i.e., EVs generated through the endosomal pathway; microvesicles ($\sim 100 \text{ nm} - 1 \mu \text{m}$ in diameter) are generated through budding from the plasma membrane, and apoptotic bodies (>1 μ m) are generated through decomposition of apoptotic blebs. These subclasses of vesicles, defined through their biogenesis and biophysical properties (such as size, density, and predominant protein markers), remain vaguely defined, especially in terms of functionality, due to limitations in EV isolation and the absence of universally accepted characterization methodologies. The International Society of Extracellular Vesicles spearheads the efforts to establish standardization in nomenclature, definitions, and methodology in this field (Lener et al., 2015; Théry et al., 2018; Witwer et al., 2019).

Although the mechanisms by which MEx induce their therapeutic effect are only partially understood, they appear to promote tissue restoration through the modulation of immune cell phenotype in the inflamed tissue microenvironment. Our group has demonstrated that MEx suppress the levels of proinflammatory cytokines (IL6, TNF- α) and modulate the expression of anti-inflammatory markers (Cd206, Arginase-1) in proinflammatory (M1-like, LPS + IFN γ) and pro-remodeling (M2-like, IL4 + IL13) macrophages, respectively (Willis et al., 2018a). In the neonatal lung, a polarized "M2-like" macrophage phenotype is critical for the process of alveolarization and normal lung development (Jones et al., 2013). Immune system homeostasis, disrupted by high oxygen levels in murine models of BPD, appears to be restored by MEx treatment.

Paramount in the pursuance of MEx-based therapy for premature infant disease is the uniformity in MEx preparations. This emphasizes the need of reproducibility in the parental MSC clone characteristics, and factors such as age, gender, and health status should be taken into consideration when selecting MSC donors (Willis et al., 2017). MEx produced by either human umbilical cord Wharton's jelly or bone marrow MSCs have been used in preclinical studies, and they have been shown to be equally efficacious (Willis et al., 2018a).

Our group has extensively explored the role of MEx as vectors for lung-targeted therapy. MEx delivery suppressed pulmonary inflammation and the development of pulmonary hypertension and vascular remodeling in a model of hypoxia-induced lung injury (Lee et al., 2012) while preventing and reverting core features of pulmonary fibrosis, lung inflammation, and aberrant pulmonary morphology induced by bleomycin administration (Mansouri et al., 2019). Importantly, we have demonstrated a therapeutic effect of MEx in experimental BPD, ameliorating disrupted alveolarization and angiogenesis and restoring pulmonary function in adults. These actions were mostly mediated through the anti-inflammatory action of MEx on pulmonary macrophage number and phenotype (Willis et al., 2018a). Supporting results were also reported by other investigators in the field (Ahn et al., 2018; Braun et al., 2018; Chaubey et al., 2018).

MEx have been reported to restore myelination and functional outcome in experimental adult rodent models of demyelinating disease (Laso-García et al., 2018). Furthermore, MEx ameliorate white matter injury (Ophelders et al., 2016), inflammation, and gliosis (Drommelschmidt et al., 2017) and protect against impaired growth and altered cortical development from ischemic insults in models of neonatal preterm brain injury (Sisa et al., 2019). In addition, the therapeutic effect of MEx administration in ROP has been demonstrated in preclinical models. In OIR, MEx protect against retinal ischemia by preserving vascular flow and retinal thinning and enhancing functional recovery while decreasing inflammation and apoptosis. Importantly, these studies indicated that EVs are taken up by retinal neurons, retinal ganglion cells, and microglia (Moisseiev et al., 2017). Other investigators, using human placental amniotic membrane-derived MSCs, have also shown reduced pathological neovascularization in OIR (Kim et al., 2016). Collectively, MEx hold a great promise to potentially enhance the recovery of multiple organs and tissues. Their pleiotropic beneficial effect warrants their translation into clinical applications for ameliorating

multifactorial pathologies, underlying not only BPD but also other prematurity-associated diseases.

Considering that the pathobiology of the preterm newborn involves multiple organ systems that share common mechanisms (i.e., inflammation precipitated by oxygen toxicity), we considered it essential to explore the impact of systemic administration of MEx on postnatal lung, brain, and eyes using a model of multisystemic injury induced by hyperoxia exposure (see Figure 1A legend for section "Materials and Methods"). As demonstrated previously by our group (Willis et al., 2018a), hyperoxia (Hyrx) exposure to postnatal day 1 (P1) mice caused disrupted lung alveolarization and vascularization that were prevented with a single dose of MEx (Figure 1B). We now report that analysis of the retina in the same injured animals showed restoration of the retinal structure, decreased gliosis, and re-establishment of microglial homeostasis (Figure 1C) produced by Hyrx exposure, emphasizing the synergistic action of systemic MEx on immune and glial cell activation. In addition, white matter in the brain (Figure 1D) was preserved while astrogliosis, microglial response, and aberrant neuronal cortical pattern caused by Hyrx were reverted to normoxic (Nrmx) levels by MEx treatment, indicating a beneficial effect on myelination, gliosis, and neuronal density as the brain repairs and matures.

THE REGENERATIVE CAPACITY OF MEX TREATMENT FOR POSTNATAL OXYGEN-INDUCED MULTIPLE ORGAN SYSTEM INJURY: PERSPECTIVES FROM NEWBORN MEDICINE

These and previously reported results indicate that, in addition to lung, the retina and brain, known to be also vulnerable to the detrimental effects of hyperoxia exposure, can benefit from an early systemic administration of MEx. Some pathologic mechanisms involve the activation of immune cells and supportive glial cells that are critical for the maintenance of the vasculature and tissue homeostasis and suggest common pathways and processes amenable for therapeutic intervention (Figure 2). Although the detailed molecular mechanisms of MEx action are still the focus of intensive investigation, the beneficial effects MEx bestow on perinatal pathologies seem to principally rely on their immunomodulatory actions on tissue-resident immune cells. Alveolar macrophages in the lung and microglia in the brain revert to their anti-inflammatory polarization state in response to MEx, favoring remodeling and tissue repair. In addition, MEx effects on brain and retina seem to suggest that, directly or indirectly through parenchymal glial cells, MEx promote normal vascular function and neuronal integrity. Although the major systemic action of MEx appears to be the modulation of inflammatory states precipitated by oxygen toxicity, we should note that their impact on myelination has been reported, in certain studies, to be associated with their ability to



FIGURE 1 | Mesenchymal stromal cell (MSC)-extracellular vesicle (MEx) treatment reduces systemic cellular and inflammatory responses. (A) Schematic showing the experimental design. Neonatal mice were exposed to hyperoxia (Hyrx; 75% O₂) from postnatal day 1 (P1) to P7 and returned to room air (Nrmx) from P7 to P14 (this exposure modality impacts both alveolarization and myelination, as these processes progress postnatally). Hyrx mice were compared with age-matched control mice that remain in Nrmx conditions. The treated mice (Hyrx + MEx) received a single IV dose of MEx (50 μl corresponding to the product generated by 0.5 × 10⁶ MSCs injected via the superficial temporal vein) at P4 and were sacrificed immediately after Hyrx exposure (P7, period of maximal vulnerability for microglial activation and cortical migration) or after an additional 7 days in Nrmx (P14). MEx belong to a subset of "small" EVs that includes exosomes and were derived from either human umbilical cord Wharton's jelly or bone marrow MSCs and purified in accordance with the 2018 Minimal Information for Studies of Extracellular Vesicles (Théry et al., 2018). Lungs, brains, and retinas were harvested, fixed, and processed for immunohistochemical and immunofluorescence analyses. (B) Disrupted alveolarization and vascularization in P14 Hyrx lungs was prevented with a single dose of MEx as shown in hematoxylin and eosin and von Willebrand factor-stained sections, respectively. Scale bar = 100 µm. (C) Hyperoxic injury of P14 retinal layers was prevented by MEx. Hyrx decreased retinal thickness as assessed with toluidine blue staining as a ratio of outer nuclear layer (ONL) thickness/ONL-ganglion cell layer (GCL) distance and shown to be restored with a single dose of MEx at P4 (Hyrx + MEx). Data are shown as mean ± SEM; n = 4 per group. *P < 0.05 vs. Nrmx (ANOVA followed by Tukey's comparison test). Hyrx-induced gliosis at P14 is shown by GFAP immunofluorescence labeling of Müller cell bodies that are close to the inner nuclear layer with projections into the outer retina. MEx administration reverted Müller cell activation to Nrmx levels. Microglia activation and invasion into the ONL in P14 Hyrx retinas are depicted by increased ionized calcium binding adaptor molecule (Iba-1) immunofluorescence. MEx treatment restores microglial morphology and prevented invasion from injury. Scale bar = 25 µm. (D) Schematic depicting the cortical layers from which images were obtained. Myelin basic protein immunofluorescence showing marked staining decrease in the external capsule (ec) from P14 Hyrx mice and amelioration of white matter loss after a single dose of MEx at P4. Astrogliosis, as denoted by GFAP immunofluorescence staining at P14, was increased in early-exposed Hyrx mice but attenuated in the brains of MEx-treated littermates, particularly in areas in close association with white matter corresponding to the ec. Large Iba-1 positive cell bodies and thickened processes in P7 Hyrx cortex appear activated in comparison to Nrmx microglial cells. MEx normalized microglial morphological appearance in treated-Hyrx brain. Neuronal nuclear protein immunofluorescence staining of cortical P7 Hyrx brains, showing a less defined neuronal labeling pattern. MEx treatment restored cortical lamination after injury to Nrmx levels. Scale bar = 50 µm. GCL, ganglion cell layer; INL, inner nuclear layer; IPL, inner plexiform layer; IV, intravenous; ONL, outer nuclear layer; OS/IS, photoreceptor outer and inner segments junction; OPL, outer plexiform layer.



promote oligodendrocyte differentiation and maturation (Otero-Ortega et al., 2018).

Following early systemic administration, MEx can be delivered to injured developing tissues and improve organ development through anti-inflammatory, angiogenic, and pro-survival mechanisms. This early intervention would require the development of reliable biomarkers of disease and better brain and lung imaging tools, thus allowing the identification of patients likely to develop BPD and associated illnesses as early as possible. Most current investigations of live cell MSC-based or MEx-based therapeutic approaches are focused on evaluating the safety and efficacy of these treatments after preterm birth. However, efficient MEx therapies should also consider the effects on the host immune system. As preterm neonates are more skewed toward a tolerogenic development and are more susceptible to infections, the immunomodulatory impact of MEx treatment has to be considered when applying those therapies in compromised newborns with risk of infection. Efforts must be made to minimize contamination and aid consistency in MEx preparations. Finally, given the systemic effect of MEx, the advancement of in vivo tracing methods will help elucidate their interaction with target organs and help decipher the contribution of circulating EVs on systemic signaling and immunomodulation during the critical period

surrounding preterm birth and the development of lung and multiorgan complications.

CONCLUSION

This perspective advances the postulation that common triggers and pathways underlying dysregulated angiogenesis, growth, and immune system development converge in precipitating systemic neonatal injury in susceptible organs, including but not limited to the lung, brain, and retina. This demands new approaches to simultaneously address the potential pathogenic mechanisms of BPD and associated pathologies of prematurity. Numerous reports of preclinical studies, including those presented here, highlight the protective potential of MEx treatment against such early-life injuries. A phase I clinical study on the effects of MEx treatment in BPD is currently in progress, and basic research studies defining the molecular and cellular mechanisms responsible for the protective effects of MEx are actively underway. We are confident that future research in MEx therapies based on these strategies will lead to a more holistic approach for effectively preventing or treating complications of prematurity.

DATA AVAILABILITY STATEMENT

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

ETHICS STATEMENT

The animal study was reviewed and approved by the Boston Children's Hospital Animal Care and Use Committee.

AUTHOR CONTRIBUTIONS

AF-G and GW participated in study design and execution, data collection, and data analysis. VY and MR participated in

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study execution and data analysis. XL participated in study execution and technical assistance. SAM and SK contributed to study design, supervision of study execution, manuscript writing, data analysis, and final article editing and approval. All authors contributed to the article and approved the submitted version.

FUNDING

This work was supported in part by NIH grants RO1 HL146128, R01 HL055454, and R21 AI134025 (SK), the Charles H. Hood Foundation Major Grants Initiative to Advance Child Health (SK) and a United Therapeutics Corp. Sponsored Research Grant (SAM & SK).

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Conflict of Interest: SAM and SK are named inventors on intellectual property licensed by Boston Children's Hospital to United Therapeutics Corp.

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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