

# **The Human-Restricted Isoform of the α7 nAChR, CHRFAM7A: A Double-Edged Sword in Neurological and Inflammatory Disorders**

Simona Di Lascio <sup>1</sup>, Diego Fornasari <sup>1,2</sup> and Roberta Benfante <sup>1,2,3,\*</sup>

- <sup>1</sup> Department of Medical Biotechnology and Translational Medicine (BIOMETRA), Università degli Studi di Milano, 20129 Milan, Italy; simona.dilascio@unimi.it (S.D.L.); diego.fornasari@unimi.it (D.F.)
- <sup>2</sup> CNR Institute of Neuroscience, 20845 Vedano al Lambro, Italy
- <sup>3</sup> NeuroMi, Milan Center for Neuroscience, University of Milano Bicocca, 20126 Milan, Italy
- Correspondence: roberta.benfante@in.cnr.it

Abstract: CHRFAM7A is a relatively recent and exclusively human gene arising from the partial duplication of exons 5 to 10 of the  $\alpha$ 7 neuronal nicotinic acetylcholine receptor subunit ( $\alpha$ 7 nAChR) encoding gene, CHRNA7. CHRNA7 is related to several disorders that involve cognitive deficits, including neuropsychiatric, neurodegenerative, and inflammatory disorders. In extra-neuronal tissues,  $\alpha$ 7nAChR plays an important role in proliferation, differentiation, migration, adhesion, cell contact, apoptosis, angiogenesis, and tumor progression, as well as in the modulation of the inflammatory response through the "cholinergic anti-inflammatory pathway". CHRFAM7A translates the dup $\alpha$ 7 protein in a multitude of cell lines and heterologous systems, while maintaining processing and trafficking that are very similar to the full-length form. It does not form functional ion channel receptors alone. In the presence of CHRNA7 gene products, dup $\alpha$ 7 can assemble and form heteromeric receptors that, in order to be functional, should include at least two  $\alpha$ 7 subunits to form the agonist binding site. When incorporated into the receptor, in vitro and in vivo data showed that  $dup\alpha 7$ negatively modulated  $\alpha$ 7 activity, probably due to a reduction in the number of ACh binding sites. Very recent data in the literature report that the presence of the duplicated gene may be responsible for the translational gap in several human diseases. Here, we will review the studies that have been conducted on CHRFAM7A in different pathologies, with the intent of providing evidence regarding when and how the expression of this duplicated gene may be beneficial or detrimental in the pathogenesis, and eventually in the therapeutic response, to CHRNA7-related neurological and non-neurological diseases.

**Keywords:** CHRFAM7A; CHRNA7; human-specific gene; nicotinic receptor; cholinergic anti-inflammatory pathway; neurodegeneration; neuroinflammation

## 1. Introduction

*CHRFAM7A* (*dup* $\alpha$ 7) is a human-lineage-specific gene [1–3] that first appeared about 3.5 million years ago during evolution: this can be confirmed by its absence in other primates [4], and its absence in rodents excludes the possibility of gene loss [5]. It arose from the duplication of *CHRNA7* gene exons 5 to 10 (Figure 1A), encoding the  $\alpha$ 7 subunit of the neuronal nicotinic acetylcholine receptor (nAChR) on chromosome 15q13-q14, 1.6 Mb apart from *CHRNA7*, and has an inverted orientation with respect to the parental gene (Figure 1B) [2,6,7]. Here, the *CHRNA7*-derived cassette fused in-frame to a cluster (*FAM7A*) of seven exons (A to F), which are present four times on chromosome 15q13.3, located both upstream and downstream of *CHRNA7*, in the sense and antisense directions (Figure 1A). In particular, exons A, B, C, and E derive from the unc-51 like kinase 4 gene (*ULK4*), located at chromosome 3p22.1, whereas exons D and F are homologous to the *GOLGA8B* gene, which is located 2.5 Mb 3' from *CHRNA7* [8].



Citation: Di Lascio, S.; Fornasari, D.; Benfante, R. The Human-Restricted Isoform of the α7 nAChR, CHRFAM7A: A Double-Edged Sword in Neurological and Inflammatory Disorders. *Int. J. Mol. Sci.* 2022, 23, 3463. https://doi.org/ 10.3390/ijms23073463

Academic Editor: Patrizia Hrelia

Received: 24 January 2022 Accepted: 21 March 2022 Published: 22 March 2022

**Publisher's Note:** MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



**Copyright:** © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/).



**Figure 1.** The *CHRNA7* and *CHRFAM7A* locus on Chr. 15q13.3. (**A**) Scheme of the genomic structure of the ten-exon *CHRNA7* gene, flanked by four FAM7A cassettes. Numbers indicate exons (1 to 4 in blue, 5 to 10 in red). (**B**) *CHRNA7* exons 5 to 10 (red) are duplicated and fused in-frame with one *FAM7A* cassette (green), 1.6 Mb from, and in the opposite orientation to, the *CHRNA7* gene, giving rise to the hybrid *CHRFAM7A* gene (wild-type allele). (**C**) A deletion of two base pairs in exon 6 (2 bp del), generating the  $\Delta 2bp$  polymorphic allele, which is associated with *CHRFAM7A* gene inversion with the same orientation as the *CHRNA7* gene. (Created with BioRender.com (accessed on 23 January 2022)).

More recently, a polymorphism (c.497-498delTG; rs67158670) and a deletion of two base pairs ( $\Delta 2bp$ ) in exon 6, occurred in *CHRFAM7A* (dup $\Delta \alpha 7$ ; Figure 1C), which was determined as being associated with gene inversion, with the same orientation as the *CHRNA7* gene [6]. The *CHRFAM7A* gene varies in terms of its number of copies in each individual, with 5–10% of individuals being non-carriers of *CHRNA7* duplication and approximately 30% only having one copy [2,9,10]. The absence, as well as the presence of one or two copies of the gene, and the presence of  $\Delta 2bp$  deletion, results in six different genotypes (see Box 1). Moreover, the frequency of the  $\Delta 2bp$  allele varies in different ethnic groups (Table 1), and this polymorphism has been associated with an increased susceptibility to psychiatric disorders (see below).

**Table 1.** Allelic frequency of 2bp deletion (-/*TG*) from the 1000 Genomes Project. EAS = East Asian; EUR = European; AFR = African; AMR = Ad-mixed Americans; SAS = South Asian.

| Sample     |                         | Alleles    |            |
|------------|-------------------------|------------|------------|
| Population | Chromosome Sample Count | -          | CA         |
| EAS        | 1008                    | 0.66469997 | 0.33530000 |
| EUR        | 1006                    | 0.37770000 | 0.62230003 |
| AFR        | 1322                    | 0.08170000 | 0.91829997 |
| AMR        | 694                     | 0.44090000 | 0.55910003 |
| SAS        | 978                     | 0.48469999 | 0.51530004 |

Box 1. The six possible combinations of the CHRFAM7A genotype.

- CHRFAM7A null/null
- CHRFAM7A dupα7/null
- CHRFAM7A dupα7/dupα7
- CHRFAM7A  $dup\alpha7/dup\Delta\alpha7$
- CHRFAM7A  $dup\Delta\alpha7/dup\Delta\alpha7$
- CHRFAM7A  $dup\Delta\alpha7/null$

Recently, data in the literature have reported that the presence of the duplicated gene may be responsible for the translational gap in several human diseases [11–14]. However, it is important to address whether this isoform is indeed expressed in vivo. Moreover, clinical and pre-clinical data neither seem to indicate a unique role of CHRFAM7A, nor whether it is protective or toxic, because of confounding factors caused by the disease being studied, and the model (clinical or pre-clinical) used. In this review, we will summarize all of the biochemical studies containing evidence of the formation of a heteromeric receptor composed of CHRNA7 and CHRFAM7A encoded subunits for the first time. Moreover, we present all of the available data on the role of CHRFAM7A in different pathologies, with the intent of providing evidence regarding when and how the expression of this duplicated gene may be beneficial or detrimental in the pathogenesis, and eventually in the therapeutic response, to both neurological and non-neurological diseases. As a common trait, the deregulated expression of both genes (up- and down-regulated), due to the presence of the  $\Delta 2bp$  polymorphism in CHRFAM7A and/or the CNV of both genes, alter the CHRNA7/CHRFAM7A ratio, thus contributing to the beneficial/detrimental effects of dup $\alpha$ 7 on  $\alpha$ 7 nAChR function.

#### 2. Structure of Neuronal Nicotinic Receptors (nAChRs)

Nicotinic Acetylcholine Receptors (nAChRs) are part of the Cys-loop receptor superfamily. These receptors are transmembrane ion channels activated by neurotransmitters that are mainly expressed in the central and peripheral nervous systems and are responsible for the regulation and transmission of excitatory and inhibitory signals [15]. The ionotropic GABAergic receptors GABA<sub>A</sub> and GABA<sub>C</sub>, glycinergic receptors, serotonin (5-HTn) receptors, and the aforementioned nAChRs, which include muscle- and neuronalsubtypes, are the best-known members of the Cys-loop receptor superfamily [16]. Cys-loop receptors share a common pentameric structure, with their subunits being arranged in a pseudo-symmetrical rosette, forming a central pore [15,16]. Each subunit consists of an extracellular hydrophilic N-terminal domain that contains the binding site for the ligand, three hydrophobic transmembrane domains (M1–M3), a highly variable cytoplasmic domain between the different subunits, a fourth hydrophobic domain (M4), and an extracellular C-terminal domain (Figure 2). The M2 domains of the five subunits form the central pore and possess amino acids that are important for ion selectivity, permeability, and channel gating [16]. Nicotinic receptors are selective cationic channels for  $Na^+$ ,  $K^+$ , and  $Ca^{2+}$ , and have a high affinity for nicotine and the endogenous ligands choline (Ch) and acetylcholine (ACh); the relative affinity of the ligands varies according to the subunit composition [16]. The combination of nine  $\alpha$  ( $\alpha 2-\alpha 10$ ) and three  $\beta$  ( $\beta 2-\beta 4$ ) subunits form both hetero- and homo-pentamer receptors that have different structural, functional, and pharmacological properties (Figure 2). Homo-pentameric receptors have five identical orthosteric ACh binding sites that are located in the extracellular interfaces between two adjacent subunits (Figure 2), whereas hetero-pentameric receptors, which have two  $\alpha$ /three  $\beta$  and three  $\alpha$ /two  $\beta$  subunits, have two orthosteric binding sites that are located at the interfaces between the  $\alpha$  subunit and the  $\beta$  subunit (Figure 2).



**Figure 2.** Structure of nAChRs. On the left, each nAChR subunit is composed of an extracellular amino terminal portion, followed by three hydrophobic transmembrane domains (M1–M3), a large intracellular loop, a fourth transmembrane domain (M4), and an extracellular carboxy–terminus. In the middle, the pentameric arrangement of nAChR subunits is shown in an assembled receptor. The M2 transmembrane domain of the five subunits forms the central pore and possesses amino acids that are important for ion selectivity, permeability, and channel gating. On the right, five subunits can assemble to form homo- (five  $\alpha$  subunits) or hetero-pentameric ( $\alpha$  and  $\beta$  subunits) receptors. The orthosteric ligand binding site is formed between two  $\alpha$  subunits (in red) in homomeric receptors, and between the  $\alpha$  and  $\beta$  subunits in an heteromeric receptor. (Created with BioRender.com (accessed on 23 January 2022)).

nAChRs, as modulators of the release of neurotransmitters, play a fundamental role in cognitive functions, and their decline or dysfunction leads to neuropsychiatric and neurodegenerative diseases. Genetic and knock-out studies on mouse models have confirmed a link between the malfunction or mutation of these genes and pathologies such as epilepsy, schizophrenia, depression, and nicotine addiction [16,17].

### 2.1. The $\alpha$ 7 nAChR

 $\alpha$ 7 nAChR is a homo-pentamer whose subunit,  $\alpha$ 7, is encoded by the ten-exon CHRNA7 gene (Chr 15q13-q14), which allows the translation of a protein of about 57 kDa (Figure 3A). Exons 1–7 encode the N-terminal portion that contains the ACh binding site; exons 7–8 comprise the M1–M3 transmembrane regions; and exons 9–10 comprise the cytoplasmic loop, the fourth transmembrane domain (M4), and the C-terminal portion (Figure 3A).  $\alpha$ 7 nAChR exhibits a low affinity for nicotine, a high affinity for  $\alpha$ bungarotoxin ( $\alpha$ -Bgtx), and is highly permeable to Ca<sup>2+</sup>, thus suggesting that it plays a role not only as an ion channel, but also as a regulator of the calcium-activated signaling pathways [16]. The receptor is expressed in the brain, in excitatory and inhibitory neurons, both pre-synaptically, where it modulates the release of neurotransmitters, and post-synaptically, as well as in astrocytes and microglia [18-20]. In the periphery, it is expressed in neuroendocrine cells [21], sperm acrosome [22], epithelial cells [23], microvascular endothelial cells [24], pulmonary fibroblasts [25], bone marrow [26], chondrocytes [27], T and B lymphocytes [28,29], and macrophages [30]. Here, it plays an essential role in the "cholinergic anti-inflammatory" response [31-33]. Its nature as a modulator of intracellular calcium concentration, which is one of the most widespread second messengers, explains its almost ubiquitous expression. *CHRNA7* is related to several disorders with cognitive deficits, including schizophrenia [9], P50 auditory gating deficits [34], autism [35], epilepsy [36], bipolar disorder [37], attention deficit hyperactivity disorder (ADHD) [38], Down syndrome [39], Parkinson's disease (PD) [40], and Alzheimer's disease (AD) [41,42]. In extra-neuronal tissues,  $\alpha$ 7 nAChR plays important roles in proliferation, differentiation, migration, adhesion, cell contact, apoptosis, angiogenesis, and tumor progression [43–48].



**Figure 3.** The  $\alpha$ 7 and dup $\alpha$ 7 receptor. Relative protein products of *CHRNA7* and *CHRFAM7A*. (**A**), *CHRNA7* encodes the 503 aa  $\alpha$ 7 subunit: exons 1–7 encode the extracellular N-terminus domain, including the signal peptide and the acetylcholine binding domain; exons 7–10 encode the M1–M3 transmembrane domains, the long M3–M4 intracellular loop, the M4 transmembrane domain, and the small extra-cellular C-terminal domain. (**B**,**C**), *CHRFAM7A* encodes the dup $\alpha$ 7 subunit via two alternatively spliced mRNAs that differ in terms of the presence of exon B. Translation starting from exon B (**B**) results in a 413 aa protein that differs from  $\alpha$ 7 for the N-terminal domain, and a 27 amino acid-long peptide that is encoded by ExB–Ex6. Translation starting from exon 6 (**C**) results in a 322 aa protein that can be considered a truncated  $\alpha$ 7 subunit with a short N-terminus encoded by exons 6–7. (**D**), *CHRFAM7A* $\Delta$ 2*bp*. The presence of the  $\Delta$ 2*bp* polymorphism (-/TG) in exon 6 results in a short, truncated protein that is encoded by exons 8–6 because of the insertion of a translation stop codon and a receptor encoded by exons 6–10 (dup $\Delta \alpha$ 7), due to the translational start site in exon 6. This protein is similar to the one obtained in (**C**). (Created with BioRender.com (accessed on 23 January 2022)).

## 2.2. The dupa7 Receptor

The CHRFAM7A gene transcribes for two isoforms (isoform 1, NM\_139320.2/ NP\_647536.1; isoform 2, NM\_148911.1/NP\_683709.1) that differ in terms of the presence of exon B [49], due to alternative splicing. The transcript of isoform 1 presents two open reading frames (ORF) (Figure 3B); translation starting from exon B (ORF 1) results in a protein that is 412 amino acids (aa) long (46 KDa; dup $\alpha$ 7) and is different from the  $\alpha$ 7 nAChR subunit for the N-terminal domain which is formed by a 27 aa long peptide (Figure 3B), but it still includes the disulfide bridge and the vicinal cysteines. The resulting peptide will lose a putative glycosylation site and part of the agonist binding site. A shorter polypeptide (38 KDa) may putatively originate from the second ORF starting at exon 6 (Figure 3B). This translation start site is the only start site that exists in isoform 2 (Figure 3C) and results in the 38 KDa protein, which lacks the signal peptide and the entire binding site, but contains all of the  $\alpha$ 7 transmembrane domain sequences. The expression of both transcripts and proteins has been confirmed in both heterologous expression systems [50–52] and in vivo [49,50,53–56]. The  $\Delta 2bp$  deletion polymorphism (Figure 3D) introduces a stop codon in ORF 1, generating a truncated subunit that is formed by a short 40 aa long peptide that is codified by exon B-exon 5. It is possible to synthesize the 38 KDa protein (dup $\Delta \alpha$ 7) if translation restarts from exon 6, downstream from the  $\Delta 2bp$ polymorphism (Figure 3D) [51,52].

In leukocytes, a ~2 Kb long promoter, located upstream and that overlaps with part of exon D, drives *CHRFAM7A* expression [49].

Numerous attempts have been made to understand the nature and function of this duplicated gene. The homology of the full-length gene suggests that the duplicated gene may have a new function, but always in the context of cholinergic biology. The fact that this endogenous duplicated gene translates a subunit [49,53,57,58], reinforces the hypothesis that dup $\alpha$ 7 may interact with and regulate  $\alpha$ 7 nAChR.

#### 2.3. The Heteromeric $\alpha$ 7/dup $\alpha$ 7 Receptor

*CHRFAM7A* gene products (mRNA and protein) are present in different brain areas in different proportions compared to *CHRNA7* (up to 10–20% of the  $\alpha$ 7 transcript) [1,8,44,50,55,59–61], as well as in at least thirty tissues in the periphery, including in leukocytes [11,45,49,53,54,56,57,62]. In leukocytes, *CHRFAM7A* expression overcomes that of *CHRNA7* by two orders of magnitude [49,50], and seminal electrophysiological studies [57] have failed to measure an ACh or a nicotine-evoked current, probably due to the missing ACh binding sites. On the other hand, the presence of the transmembrane domains in the translated dup $\alpha$ 7 subunits (in particular M2, forming the ion pore) indicates that a hypothetical dup $\alpha$ 7 receptor could trigger Ca<sup>2+</sup> influx in response to a specific ligand. An alternative hypothesis is that the dup $\alpha$ 7 subunit does not form homomeric receptors, but is instead assembled with conventional  $\alpha$ 7 subunits to form a new class of heteromeric receptors. The role of dup $\alpha$ 7 functions.

To address this issue, several groups have used both functional and structural approaches [50–52,58,63,64], most of which have been based on the heterologous expression of the  $\alpha$ 7 and dup $\alpha$ 7 proteins in oocytes and cell lines. In oocytes, the expression of dup $\alpha$ 7 alone has failed to produce any ACh-induced currents [50], an effect that has also been observed in mammalian cell lines [51,52,63], and that has been hypothesized as being due to the missing agonist binding sites and the lack of  $\beta$ 1 $\beta$ 2 loop, both of which are required for channel opening [65,66]. When the receptors were co-expressed, the increased concentration of dup $\alpha$ 7 ( $\alpha$ 7/dup $\alpha$ 7 ratio 1:5/1:10) reduced the nicotine- and ACh-induced  $\alpha$ 7 currents, but not receptor sensitivity to ACh [50–52], thus suggesting that dup $\alpha$ 7 has a dominant negative effect on the  $\alpha$ 7 current. The overexpression of the construct with the  $\Delta$ 2*bp* in exon 6, dup $\Delta\alpha$ 7, further reduced the current amplitude by an additional 10% [51]. This effect has not been replicated in the Neuro2a cell line, which may be due to poor dup $\alpha$ 7 translation [52], something that was recently confirmed in SH-SY5Y cells [58].

Confocal images using both an antibody, which detects both receptors, and FITC conjugated  $\alpha$ -Bungarotoxin ( $\alpha$ -Bgtx), an  $\alpha$ 7 antagonist that only binds to the N-terminal domain of the  $\alpha$ 7 receptor, demonstrated that this effect was due to a reduced number of  $\alpha$ 7 receptors at the plasma membrane [11,50,63,64]. The dup $\alpha$ 7 protein seemed to preferentially localize within the endoplasmic reticulum (ER) [50,52,64], making it likely that the dominant negative effect may be exerted by increased  $\alpha$ 7 receptor retention in the ER (Figure 4). This model does not exclude the possibility of a heteromeric receptor  $\alpha$ 7/dup $\alpha$ 7 being formed. In the presence of <sup>125</sup>I- $\alpha$ -Bgtx, surface receptor binding assays showed a reduced number of <sup>125</sup>I-α-Bgtx binding sites and an increase in the equilibrium dissociation constant (Kd), thus suggesting the formation of a less functional receptor [50,51]. On the other hand, higher CHRFAM7A expression versus CHRNA7 expression can result in a potentiation of the effect of a positive allosteric modulator that is able to bind to the transmembrane domains of the receptor (PNU-120596) [51]. This finding proved that a heteromeric receptor that incorporates CHRFAM7A subunits, and with properties that are different from those of  $\alpha$ 7 nAChR, can be formed, although this result has not been confirmed by other groups [50,58,63].



**Figure 4.** The heteromeric  $\alpha 7/\text{dup}\alpha 7$  receptor. Dup $\alpha 7$  can assemble and interact with  $\alpha 7$  subunits to form heteropentameric receptors, causing reduced function of the receptor at the cell membrane (dashed line), as demonstrated by a net reduction in the macroscopic current, that is inversely proportional to the amount of dup $\alpha 7$  and/or retention (dashed line) at the endoplasmic reticulum (ER). In order for the heteromeric receptor to be functional, it should include at least two  $\alpha 7$  subunits to form the agonist binding site. The possible stoichiometries of the heteromeric receptor are the result of experimental data and computational model simulations. (Created with BioRender.com (accessed on 23 January 2022)).

The putative assembly of the subunits was investigated using *Forster Resonance Energy Transfer* (FRET) *by Donor Recovery after Acceptor Photobleaching* (DRAP), and *FRET by Fluores-cence Lifetime Imaging Microscopy* (FLIM), in the Neuro2a [52], GH4C1 and RAW264.7 [64], and SH-SY5Y [58] cell lines. Both approaches revealed and confirmed the assembly of duplicated subunits with full-length  $\alpha$ 7, forming several types of hetero-pentamers between  $\alpha$ 7 and dup $\alpha$ 7/dup $\Delta\alpha$ 7 ( $\alpha$ 7 with  $\alpha$ 7;  $\alpha$ 7 with dup $\alpha$ 7;  $\alpha$ 7 with dup $\alpha$ 7; dup $\alpha$ 7; dup $\alpha$ 7 with dup $\alpha$ 7; dup $\alpha$ 7 with dup $\alpha$ 7; d

dup $\Delta \alpha$ 7 with dup $\Delta \alpha$ 7). According to these data, a limited number of dup $\alpha$ 7 subunits (mainly one) may be incorporated and are likely adjacent to a  $\alpha$ 7 subunit in the heteromeric receptor. Interestingly, the duplicated isoforms can interact with other nAChRs subunits, namely  $\alpha$ 4 and  $\alpha$ 3 [52].

The formation of  $\alpha 7/dup\alpha 7$  or  $\alpha 7/dup\Delta \alpha 7$  hetero-pentamers in cell lines does not imply that they are functional. This question has been addressed by means of the substituted cysteine accessibility method (SCAM), a technique that allows the measurement of altered ion channel function, following the substitution of key amino acids with cysteine, and treatment with sulfhydryl-specific reagents [52]. The leucine at the 247 -position resides nine residues downstream from the cytoplasmic beginning of the TM2 (9' position), which forms the ion channel pore of  $\alpha$ 7 and the duplicated isoform. A whole-patch clamp of Neuro2a cells, transfected with  $\alpha$ 7 along with substituted L9'C subunits ( $\alpha$ 7 L9'C; dup $\alpha$ 7L9'C; dup $\Delta \alpha$ 7L9'C), showed a reduction in the ACh-evoked inward current  $(I_{ACh})$  in the presence of the dup $\alpha$ 7 subunit isoforms that was similar to the reduction measured on cells expressing  $\alpha 7/\alpha 7L9'C$ . These data suggest that both dup $\alpha 7$  and dup $\Delta \alpha 7$ could form heteromeric functional receptors with  $\alpha$ 7. Despite the expression of dup $\alpha$ 7 and dup $\Delta \alpha 7$ , there was no effect on the ACh-evoked current amplitude in Neuro2a [52]. The inclusion of dup $\alpha$ 7 induced a higher sensitivity to a high concentration of choline compared to cells that were only expressing  $\alpha$ 7 or  $\alpha$ 7/dup $\Delta \alpha$ 7, whereas brief exposure to a physiological concentration of choline led to a faster desensitization of both the  $\alpha 7/dup\alpha 7$ and  $\alpha 7/dup\Delta \alpha 7$  receptors. Conversely, the inclusion of  $dup\Delta \alpha 7$  increased the sensitivity of the  $\alpha$ 7 nAChR to the full  $\alpha$ 7 agonist varenicline [52], which is commonly used for smoking cessation [67]. However, genetic studies did not reveal any significant association between dup $\Delta \alpha 7$  and smoking abstinence [67].

Patch clamp analyses to measure single-channel function, coupled with the electric fingerprinting strategy, have provided the first evidence of the actual formation of functional hetero-pentamers [63]. This strategy relies on the possibility of distinguishing the activity of the  $\alpha$ 7 subunit with three arginine substitutions at the intracellular M3–M4 loop region ( $\alpha$ 7LC,  $\alpha$ 7 low-conductance).  $\alpha$ 7LC incorporation generates multiple and discrete current amplitude clusters, each one corresponding to a different population of receptors with a fixed number of LC subunits. LC receptors, when expressed individually, do not display detectable currents at the single-channel level. Single channel currents activated by ACh and 1  $\mu$ M PNU-1205696, from cells expressing  $\alpha$ 7/ $\alpha$ 7LC, distribute into clusters of different amplitudes, something that is indicative of the formation of pentameric receptors that incorporate from zero up to three  $\alpha$ 7LC subunits. When co-expressed,  $\alpha$ 7LC + dup $\alpha$ 7, or the reverse combination dup $\alpha$ 7LC +  $\alpha$ 7, reveal that dup $\alpha$ 7 can assemble with  $\alpha$ 7, forming functional heteromeric receptors containing one, two, or three dup $\alpha$ 7 subunits (Figure 4). Mutagenesis analyses of the conserved Tyr-93 in the  $\alpha$ 7 subunit, which is important for ACh binding, in the context of the  $(\alpha 7)_2(dup\alpha 7)_3$  receptor, revealed that the heteromeric receptor has to contain at least two adjacent  $\alpha$ 7 subunits, and no more than three dup $\alpha$ 7 subunits to be functional (Figure 4) [63,68]. Moreover, the channel kinetics elicited by the ACh + PNU-120596 of the  $(\alpha 7)_2(dup\alpha 7)_3$  receptor does not differ from that of the  $\alpha 7$ homomeric receptor [63], as previously reported [51].

These findings have been confirmed by computational model studies [69] on all of the possible combinations of  $\alpha$ 7 and dup $\alpha$ 7 pentamers. The authors of this study have shown that receptors with dup $\alpha$ 7/ $\alpha$ 7 interfaces are more stable and less detrimental for ion conductance than dup $\alpha$ 7/dup $\alpha$ 7 interfaces, and that the most stable stoichiometry is 1: dup $\alpha$ 7/4:  $\alpha$ 7; however, functional heteromeric receptors can be formed with no more than three dup $\alpha$ 7 subunits (Figure 4). Moreover, the heteromeric receptors showed a very low affinity for  $\alpha$ -Bgtx, thus explaining the reduction observed in previous studies [50–52,70], and are less sensitive to A $\beta_{1-42}$ , which supports recent experimental data [12].

In conclusion, these studies demonstrated that the duplicated form of the *CHRNA7* gene, *CHRFAM7A*, translates for the dup $\alpha$ 7 (or dup $\Delta\alpha$ 7) protein in a multitude of cell lines and heterologous systems with processing and trafficking that are very similar to the

full-length form without affecting *CHRNA7* transcription. It does not form functional ion channel receptors alone. In the presence of *CHRNA7* gene products, dup $\alpha$ 7 can assemble and form heteromeric receptors that, in order to be functional, should include at least two  $\alpha$ 7 subunits to form the agonist binding site. When incorporated into the receptor, dup $\alpha$ 7 negatively modulates  $\alpha$ 7 activity, as demonstrated by a net reduction in the macroscopic current, which is inversely proportional to the amount of dup $\alpha$ 7 and is probably due to a reduction in the number of ACh binding sites.

This observation can be of fundamental importance, especially when considering the different dup $\alpha$ 7 and  $\alpha$ 7 expression levels in different tissues, and the modulation of their expression by various stimuli (IL-1 $\beta$ , LPS, nicotine, donepezil, HIV-1 gp120) [49,50,53,61], although evidence of the formation of a heteromeric dup $\alpha$ 7/ $\alpha$ 7 receptor in vivo is still missing. Recently, *CHRFAM7A* transgenic mice showed decreased  $\alpha$ -Bgtx at the neuro-muscular junction and in brain tissue compared to wild-type (WT) animals, showing for the first time that dup $\alpha$ 7 expression decreases  $\alpha$ 7 nAChR ligand binding in vivo [70,71]. Silencing *CHRFAM7A* expression in SH-SY5Y cells significantly increased  $\alpha$ 7 nAChR-induced neurotransmitter release, which was reduced by the overexpression of the dup $\alpha$ 7 protein [58].

The involvement of  $\alpha$ 7 nAChR in physiological mechanisms, and its association with neurological, neurodegenerative, and inflammatory disorders, confirmed the importance of understanding whether an heteromeric receptor is formed in vivo (i.e., in neurons), and how these isoforms are regulated and eventually deregulated in different pathologies and in response to therapies targeting the cholinergic system.

## 3. CHRNA7 and CHRFAM7A Role in Diseases

3.1. Schizophrenia and Neuropsychiatric Disorders

Chromosomal defects (deletion, duplication) and polymorphisms in the 15q13-q14 region are associated with several neuropsychiatric disorders, including schizophrenia [72–74], epilepsy [10,36,75,76], psychosis [77–79], intellectual disability (ID) [80,81], and autism [35,82]. A direct genetic association of CHRNA7 with these disorders is controversial; many markers (i.e., the dinucleotide marker D15S1360 in intron 2 of the CHRNA7 gene and other microsatellite markers) in the vicinity of the CHRNA7 gene at 15q13-q14, which have a strong association with schizophrenia, have been reported [74]. Other studies have failed to find any genetic linkage to this region [83–86], thus supporting the idea that schizophrenia is a genetically heterogeneous disease, and that the loci involved may differ depending on ethnicity [87,88]. A stronger linkage with CHRNA7 was obtained when the P50 auditorily evoked response deficits [89,90], an endophenotype of both schizophrenia and bipolar disorders, and nicotine, transiently normalized the P50 deficit in schizophrenic and bipolar disorder patients, thus confirming the involvement of a nicotinic receptor [91]. The finding that schizophrenic and depressed patients showed the highest prevalence of smoking [92,93] reinforced the hypothesis that smoking may be a form of "self-medication" for these patients [92,94]. Indeed, evidence from animal and human studies have indicated that targeting  $\alpha$ 7 nAChRs improves cognition, memory, and sensory gating deficits in schizophrenia [95–98]. It is worth noting that in peripheral blood lymphocytes, CHRFAM7A expression is lower in cotinine, in self-reported smokers versus in non-smokers, and is negatively correlated to cotinine (nicotine metabolite) levels, but not with the diagnosis of schizophrenia [99]. Conversely, individuals with two or three copies of the CHRFAM7A gene who also smoked showed a higher rate of smoking cessation success under varenicline therapy [67].

The presence of *CHRNA7* and *CHRFAM7A*, in this locus, could increase the probability that the deregulation (gene dosage; altered gene expression) of even one of the two genes could lead to the onset of neuropsychiatric pathologies [55,100,101]. The expression of  $\alpha$ 7 is reduced in multiple areas of postmortem brain specimens from schizophrenic patients and from patients with major psychiatric disorders [55,102–106], and 21 polymorphisms have been identified in the promoter region, most of which decrease tran-

scription and are strongly associated with schizophrenia and P50 deficits [34,107,108]. Polymorphisms in both the coding region and the introns that result in splice variants were identified in *CHRNA7* and *CHRFAM7A* genes [109], but none of them were associated with schizophrenia, thus reinforcing the assumption that the receptor is functionally normal and that the causative defect may rely on the expression level of *CHRNA7*. Deletions involving *CHRNA7* are rare but are strongly associated with schizophrenia [72,73]: these patients only have one copy of *CHRNA7* and two copies of *CHRFAM7A*, leading to an altered *CHRNA7/CHRFAM7A* ratio, whereas 15q13.3 duplication has been linked to childhood-onset schizophrenia [110]. A reduced *CHRNA7*:*CHRFAM7A* ratio, due to increased *CHRFAM7A* and/or reduced *CHRNA7* expression, has been reported in the prefrontal cortex of patients with bipolar disorder and schizophrenia [55,59].

Conversely, reduced *CHRFAM7A* expression has been measured in the peripheral blood lymphocytes of schizophrenic patients [111,112] and was determined to be related to illness severity. A recent study [100] examined the expression of *CHRFAM7A* in the peripheral blood lymphocytes of schizophrenic patients, and found a significant negative correlation between *CHRFAM7A* expression and a negative psychopathology score (SANS), but not with a positive score (SAPS). Pairwise analyses before and after antipsychotic treatment revealed an increase in *CHRFAM7A* gene expression during follow-ups compared to the baseline, thus suggesting that *CHRFAM7A* has a role in schizophrenia pathogenesis and treatment. The effect of antipsychotics on *CHRFAM7A* expression has not been found in previous studies [55]. This discrepancy may be explained by limitations in both studies: there is missing knowledge about *CHRNA7* expression (not detected in lymphocytes), the genotype of the analyzed patients in Kalmady's study, and the use of postmortem human brain samples in Kunii's analyses, where confounding factors, such as the antemortem use of antipsychotic and antidepressant drugs as well as of substance of abuse, may contribute to the changes observed in the patients in those studies.

Several studies have investigated whether the  $\Delta 2bp$  allelic variant of CHRFAM7A could confer susceptibility to schizophrenia [8,9,84,113–119]. Similar to CHRNA7, controversial data have been obtained, probably due to differences in ethnic groups, in the phenotype considered (schizophrenia, P50 deficit, antisaccade performance), and in small sample sizes. Sinkus and colleagues [9] reported that despite the deleted CHRFAM7A allele being more frequent in Caucasians than in African-Americans, its presence is significantly associated with schizophrenia in both ethnic groups, but the number of CHRFAM7A alleles [8], and the presence of at least one copy of the deleted allele [113,117], is sufficient to have a P50 sensory gating deficit. These data have not been confirmed by another study [55], which reported no differences in the allelic frequency of  $\Delta 2bp$  and no association with schizophrenia, but instead reported an association with decreased CHRFAM7A expression in all subjects, including in the control groups. Moreover, statistical analyses [109] revealed that  $\Delta 2bp$  is less frequent in individuals with CHRNA7 promoter variants [34], which are responsible for decreased  $\alpha$ 7 expression and are linked to schizophrenia. Other studies have linked this polymorphism to major depressive disorder [78], bipolar disorder [114], and deficits in episodic memory [116], but not to antisaccade performance [119], the latter two being endophenotypes of schizophrenia.

These studies seem to conclude that, overall, the mutations in the *CHRNA7/CHRFAM7A* "gene cluster" which are associated with neuropsychiatric disorders, fall into the *CHRNA7* promoter, the  $\Delta 2bp$  deletion of *CHRFAM7A* and, although very rare, deletion of the *CHRNA7* gene. Moreover, the deregulated expression of both genes (up- and down-regulated) could also contribute to altering the *CHRNA7/CHRFAM7A* ratio, thus contributing to an increased detrimental effect of dup $\alpha$ 7 on  $\alpha$ 7 nAChR function.

#### 3.2. Epilepsy and Neurodevelopmental Disorders

The impact of *CHRFAM7A* gene expression and the presence of the  $\Delta 2bp$  polymorphism has also been evaluated in several neurological disorders, including in epilepsy and autism spectrum disorders (ASD). A susceptibility locus for the common idiopathic

generalized epilepsies (IGEs) has been mapped to the 15q13-q14 region, with a strong association with microdeletions of Chr 15q13.3 [10,36,120–124]. Rozycka et al. [120] determine an association between the  $\Delta 2bp$  polymorphism and IGE. In a population-based study, they reported that the frequency of  $\Delta 2bp$  carriers was lower in the cohort of IGE patients versus in the cohort of healthy controls, thus providing a protective effect against IGE. This finding is in contrast to what was found in schizophrenic patients [8] and the authors were unable to exclude the possibility that the  $\Delta 2bp$  is in linkage disequilibrium (LD) with the true causative polymorphism contributing to IGE [120]. The protective effect could only be explained by considering the negative effect of dup $\alpha$ 7 when translated from a wild-type allele, something that is limited in  $\Delta 2bp$  allele carriers, and by considering the *CHRFAM7A*  $\Delta 2bp$  allele as a null allele that is unable to be translated into a functional protein. This would lead to an increase in functional  $\alpha$ 7 receptors.

The study by Rozycka [120] does not report any indication regarding CHRFAM7A expression levels. However, independent studies reported a 1.5 Mb microdeletion of the region encompassing the CHRNA7 gene in a small fraction of IGE patients [121,123–125], thus confirming that  $\alpha$ 7 nAChR may play a fundamental role in IGE pathogenesis. On the other hand, a 15q13.3 duplication involving CHRNA7 has been implicated as being a risk factor for attention deficit hyperactivity disorder (ADHD) [38]. The presence of CNV and the  $\Delta 2bp$  in the CHRFAM7A gene, along with the microduplication in the CHRNA7 gene, has been associated with phenotypic variation in a family with Tourette syndrome, ADHD, and obsessive-compulsive disorders (OCD) [126]. It is worth noting that the expression levels of duplicated genes do not always follow the gene copy number, as shown in brain samples with maternal 15q duplication [127], and for CHRNA7, in a neuronal cell model of 15q duplication [128], because of epigenetic alterations that can contribute to defining the CHRNA7/CHRFAM7A ratio. Indeed, significantly reduced CHRNA7 expression has been reported in the frontal cortex of individuals with Rett syndrome or with typical ASD [60]. It was hypothesized that MeCP2 modulates both CHRNA7 and CHRFAM7A expression through long-range chromatin interactions with the 15q11.2–13.3 region, which includes the Prader-Willi/Angelman syndrome's imprinting center (PWS-IC). MeCP2 deficiency caused by mutations, such as in Rett syndrome [129], or decreased expression, such as in autism samples [130], influences chromatin loop organization, thus leading to altered gene expression.

A lack of *CHRFAM7A* expression in the CD4+ T-lymphocytes of autosomal dominant nocturnal frontal lobe epilepsy (ADNFLE) patients [131], a form of epilepsy that is mainly associated with mutations in other nAChR subunits (*CHRNA2, CHRNA4 and CHRNB2*), suggests that there might be a link between the expression of *CHRFAM7A* and the occurrence of ADNFLE symptoms [131].

## 3.3. Inflammatory Diseases: The "Cholinergic Anti-Inflammatory Pathway"

Neural reflex circuits [132,133] regulate the immune system response to pathogens during the inflammatory process. The central nervous system (CNS) senses inflammation by means of specialized cells in the brain vasculature, choroid plexus, and circumventricular organs as well as through Toll-like receptors (TLRs) and cytokine receptors in the brain. The observation that vagal nerve stimulation attenuated the systemic inflammatory response in rats under endotoxemia, and that the inhibition of the pro-inflammatory cytokines was due to the acetylcholine stimulation of the nicotinic receptors expressed on macrophages, led to the discovery of the "cholinergic anti-inflammatory pathway (CAIP)" [134]. Later studies, reviewed in Hoover, 2017 [33], defined the anatomy of this inflammatory reflex and showed that  $\alpha$ 7 nAChR plays a major role in the vagal inhibition of the inflammatory response to endotoxemia [32], despite the presence of other nAChRs on immune cells [33,135].

Functional studies on the  $dup\alpha7$  gene in leukocytes revealed the essential role that it plays in CAIP activation [53]. In the macrophages, where  $dup\alpha7$  is expressed at higher levels compared to  $\alpha7$  [49], pro-inflammatory stimuli down-regulate its expression, which is counterbalanced by an increase in *CHRNA7* [49,53], with an NF- $\kappa$ B dependent mech-

anism. These findings incite the hypothesis that in physiological situations, the high *CHRFAM7A/CHRNA7* ratio allows for the formation of hetero-pentamers due to a large contribution of dup $\alpha$ 7, or due to the intracellular retention of  $\alpha$ 7 [31,50], thus contributing to blocking CAIP activation. In the presence of inflammatory signals, the down-regulation of *CHRFAM7A*, which is paralleled by increased *CHRNA7* expression, will result in the formation of functional  $\alpha$ 7 nAChRs on the cell membrane and activation of CAIP, subsequently dampening inflammation. Due to pathological status, the altered regulation of both subunits will lead to an altered *CHRFAM7A/CHRNA7* ratio, consequently deregulating CAIP response.

Uncontrolled systemic inflammation leads to sepsis [136], the leading cause of death in intensive care units (ICUs). One of the main causes of its pathogenesis may rely on an imbalance between excessive inflammation and anti-inflammatory mechanisms, and was reviewed in van der Poll et al. 2021 [137]. During an inflammatory challenge imposed by sepsis,  $\alpha$ 7 mRNA levels increased during acute illness and returned to the levels found in the controls once the patient recovered [138]. An inverse correlation between the CHRNA7 expression level and CAIP activity, and disease severity and mortality, was reported in a cohort of patients with sepsis [138] that had no deaths among patients expressing high levels of CHRNA7, thus suggesting that measuring the  $\alpha$ 7 expression level in PBMC may serve as a prognostic marker for sepsis. A later analysis [64] showed that patients showing a high CHRFAM7A/CHRNA7 ratio have a poor prognosis compared to patients expressing higher CHRNA7 levels. A similar conclusion was drawn by Baird and colleagues [139], who showed that CHRFAM7A expression was increased, and that CHRNA7 expression was decreased in Inflammatory Bowel Disease (IBD), thus suggesting that CHRFAM7A may be an unrecognized target for the development of therapeutics for IBD. These data are in accordance with previously reported increases of CHRFAM7A caused by the LPS in gut epithelial cells [56], suggesting a different role for dup $\alpha$ 7 in this tissue type.

 $\alpha$ 7 nAChR activation plays a protective role in osteoarthritis (OA), since its absence was associated with severe cartilage degradation in a murine meniscectomy  $\alpha$ 7 KO model, whereas  $\alpha$ 7 nAChR activation decreased the IL-1 $\beta$ -induced chondrocyte inflammation [27]. The chondrocytes of OA patients showed elevated *CHRFAM7A* expression [27], which was positively correlated with increased Metalloproteinase-3 (MMP-3) expression, thus suggesting it has a role in the pathophysiology of OA.  $\alpha$ 7 nAChR silencing, and treatment with specific agonists, revealed a role for  $\alpha$ 7 nAChR in controlling joint inflammation in a collagen-induced arthritis rat model of rheumatoid arthritis (RA) [140], and in the synovial tissues and fibroblast-like synoviocytes of patients with RA [62]. These tissues express  $\alpha$ 7 and dup $\alpha$ 7 receptors, but since there is no information regarding their expression levels in RA compared to in healthy tissues, no conclusions about the role of CHRFAM7A in RA can be drawn.

A protective role for dup $\alpha$ 7 has also been proposed. Cerebral ischemia/reperfusion (I/R) injury [141] is an inflammatory-related disorder that is caused by the abnormal activation of the nod-like receptor protein 3 (NLRP3) inflammasome, and microglia pyroptosis, following the disruption to blood supply after an ischemic event [142]. In a case-control study [141], in patients with cerebral I/R injury, the *CHRFAM7A* level was negatively related to the expression of inflammatory cytokines. These data are in accordance with the reported down-regulation of *CHRFAM7A* by inflammatory stimuli [53]. The *CHRFAM7A* overexpression in OGD/R-treated microglia human cell lines, a model of I/R injury, reverted the clinical observation by increasing cell proliferation, decreasing the release of pro-inflammatory cytokines, and promoting M1 to M2 microglia polarization [141], thus suggesting a protective role for *CHRFAM7A* in the attenuation of inflammatory injury of microglia.

CHRFAM7A is also down-regulated in hypertrophic scars (HTS) [143]. In a human HTS-like SCID mouse model, transfected CHRFAM7A played a positive role in the amelioration of HTS formation [143] by decreasing TGF- $\beta$  and CTFG expression and by increasing MMP-1 expression, and conversely to the I/R injury model, attenuating M2 macrophage polarization via the activation of the Notch pathway. An increase in *CHRFAM7A* expression has been measured in radiotherapy-induced lacrimal gland injury, resulting in the inhibition of the p38/JNK signaling pathway and oxidative stress [144].

An imbalanced inflammatory response is responsible for tissue damage and COVID-19 severity [145]. The cholinergic system has been proposed as one of the regulators of COVID-19 induced hypercytokinemia [146–150]. *CHRFAM7A* expression showed a slight tendency to decrease its level in a cohort of SARS-CoV-2 patients compared with healthy controls [14]. When patients were stratified according to disease severity, *CHRFAM7A* expression was significantly lower in critical COVID-19 patients. Again, this is in accordance with inflammatory-induced *CHRFAM7A* down-regulation [53]. Absence of *CHRFAM7A* expression was also associated with increased inflammatory biomarkers, including the C-reactive protein, lymphopenia, extension pulmonary lesions, and plasma viral load, as well as the enhanced expression of genes in the TNF signaling pathway [14]. Ongoing trials are evaluating the usefulness of nicotine to determine the risk of developing the SARS-CoV-2 infection and severe disease; however, these data have led to the hypothesis that individuals with no *CHRFAM7A* duplication, and/or who are carriers of  $\Delta 2bp$ , may develop symptomatic and severe COVID-19. This aspect is challenging and deserves further investigation.

All these studies suffer from the lack of *CHRNA7* expression analysis, making it difficult to draw conclusions about the significance of a *CHRFAM7A* increase in I/R injury. It is possible to hypothesize that the increase in *CHRFAM7A* is a step in the inflammation resolution process. In accordance with this hypothesis, is a study on the effect of donepezil [49], an acetylcholine esterase inhibitor (AChEi) used as a symptomatic therapy in AD. Besides its inhibitory effect, it has been proposed as an anti-inflammatory mechanism for this class of compounds [151]. In macrophages, donepezil synergizes with LPS to increase *CHRNA7* expression, resulting in the potentiation of CAIP. The parallel increase in *CHRFAM7A* expression may suggest a role in controlling excessive CAIP activation, with the consequence of an impaired response to inflammatory stimuli [31]. In COVID-19 patients, such deregulation in the cholinergic system may explain severe prognosis. In view of this, CHRFAM7A may be considered as a target for intervention in the fine modulation of CAIP and in the restoration of a homeostatic state.

In individuals with traumatic spinal cord injury (SCI), the presence of the  $\Delta 2bp$  polymorphism may affect clinical outcomes [13,152] depending on the severity of the injury. In severe SCI,  $\Delta 2bp$  carriers show higher levels of circulating inflammatory molecules (TNF- $\alpha$ , INF- $\gamma$ , IL-13, CCL11, IL-12p70, IL-8, CXCL10, CCL4, IL-12p40, IL-1 $\beta$ , IL-15 and IL-2), whereas in mild SCI, IL-15 is lower in  $\Delta 2bp$  carriers than it is  $\Delta 2bp$  non-carriers. Temporal variations in inflammatory mediator analyses have revealed that severe SCI  $\Delta 2bp$  carriers had higher levels of IL-8 and CCL2 during the acute phase post-SCI, but no variations were observed in mild SCI. These two molecules are important for macrophage and neutrophil migration and infiltration modulators, as theyplay an important role in the early phases of inflammation post-SCI. Moreover, neuropathic pain was positively associated with IL-13 in  $\Delta 2bp$  carriers, but only in severe SCI. Conversely,  $\Delta 2bp$  carriers with mild SCI showed that a higher risk of pressure ulcers was positively correlated with the circulating levels of IFN- $\gamma$ , CXCL10, and CCL4, and negatively associated with IL-12p70 levels.

Animal studies on the association between IL-13 and neuropathic pain produced opposite results [153], thus confirming the importance of including *CHRFAM7A* genotyping in human studies, and that the presence of dup $\Delta \alpha 7$  can modify the anti-inflammatory function of  $\alpha 7$  receptor after SCI and the response to  $\alpha 7$  ligands for inflammatory and pain control [154].

## Other Roles for CHRFAM7A

The higher expression of *CHRFAM7A* in macrophages compared to *CHRNA7* suggests other roles for dup $\alpha$ 7 in regulating monocyte/macrophage activation other than the regulation of  $\alpha$ 7 nAChR activity. The transgenic expression of *CHRFAM7A*, in a model of human

systemic inflammatory response syndrome (SIRS) due to severe injury and sepsis, showed an increase in the hematopoietic stem cell (HSC) reservoir in bone marrow, and increased immune cell mobilization, myeloid cell differentiation, and inflammatory monocytes in inflamed lungs [11]. These data confirmed the importance of CHRFAM7A in physiological, pathophysiological, and species-specific responses, and point to its role as a doubled-edge sword, being both protective and detrimental in an emergency myelopoiesis model upon injury. Indeed, limiting HSC exhaustion upon injury can increase the responsiveness to a second stimulus, thus improving, for example, survival.

The overexpression of *CHRFAM7A* in monocyte-like cell lines has shown reduced cell migration, chemotaxis to the monocyte chemo-attractant protein (MCP-1), and the inhibition of anchorage-independent colony formation [155]. The mechanism of this effect is still unknown and is in contrast with previous data from the same group [11,54], which reported that the increased expression of gene encoding for proteins favors cell migration, adhesion, and the formation of cell clusters upon *CHRFAM7A* overexpression. Other affected pathways tied to pro-inflammatory conditions [156,157] include Type L interferon and cancer pathways [54]. Unexpectedly, *CHRFAM7A* overexpression increased the CHRNA7 level [54], resulting in increased  $\alpha$ -Bgtx binding. This is in contrast with previously discussed data [50,52,63,64], but it corroborates the finding that *CHRNA7* and *CHRFAM7A* expression are co-regulated [49,53,56]. Thus, we cannot rule out that many of the direct effects of *CHRFAM7A* overexpression in the modulation of different signaling pathways are indeed the result of the modulation of the metabotropic function of  $\alpha$ 7 nAChR [158].

## 3.4. Neurodegenerative Diseases

Epidemiological [159] and in vitro studies indicate a protective role of nicotine assumption in Alzheimer's (AD) and Parkinson's (PD) disease against glutamate and  $\beta$ amyloid-related cytotoxicity [160], and on dopaminergic neurons via an anti-inflammatory mechanism that is mediated by the modulation of microglial activation and astrocyte apoptosis [161,162].  $\alpha$ 7 nAChR is expressed by microglial cells, where it plays important roles in controlling their activation [163,164], and several reports support the existence of a brain CAIP [18,165,166].

One of the hallmarks of AD and PD is the loss of cholinergic neurons and a selective decrease in the number of nicotinic receptors (reviewed in [160,167]) that is associated with an inflammatory state, which is generally caused by the hyper-activation of the microglia [168,169]. For these reasons, the cholinergic system is one of the therapeutic targets in AD. Acetylcholinesterase inhibitors (AChEi), such as galantamine or donepezil, are indeed effective, and ameliorate the cognitive symptoms of AD [170].

Genetic variations in the *CHRNA7* gene, or the altered expression and function of  $\alpha$ 7 nAChR have been associated with AD [41,160,171,172] and PD [173,174], and in the case of AD, the reduced  $\alpha$ 7 expression is correlated with beta-amyloid (A $\beta$ ) plaque deposition and cognitive impairments [41,160,175]. The high-affinity binding of A $\beta$  oligomers to  $\alpha$ 7 nAChR shows A $\beta$  concentration-dependent opposite effects. At physiological concentrations (pM to low nM range), A $\beta$  triggers a change in  $\alpha$ 7 to a desensitized conformation that still responds to agonists, while at higher concentrations (nM to low  $\mu$ M range), A $\beta$  acts as a negative modulator [176–179]. This results in the activation of neuroprotective (pM range) or neurotoxic (nM range) signaling pathways [167,177,180,181]. Moreover, A $\beta$ - $\alpha$ 7 interaction leads to intraneuronal accumulation of A $\beta$ <sub>1–42</sub> and A $\beta$ -induced tau protein phosphorylation [182,183].

CHRFAM7A CNV is over-represented in mild cognitive impairment (MCI) and lateonset AD patients [184–186]. Ordered-subset analysis (OSA) by age at AD onset confirmed CHRFAM7A as one of the loci that can confer the risk of AD [187] in a subset of patients. Here, reduced CHRFAM7A expression in human post-mortem temporal lobe tissue was correlated with the disease state (AD vs. controls) and with CNV [187]. In view of this, the increased functional  $\alpha$ 7 nAChR due to lower CHRFAM7A expression is able to sustain enhanced  $A\beta_{1-42}$  internalization and neuronal vulnerability, and therapies with  $\alpha$ 7 antagonists may be more effective in these AD patients.

The presence of the  $\Delta 2bp$  polymorphism did not confer susceptibility to Alzheimer's disease in a case-control study [188]. Patients were stratified by three different *CHRFAM7A* genotypes (genotype 1: two wild-type alleles; genotype 2: one wild-type allele and one  $\Delta 2bp$  allele; genotype 3: two  $\Delta 2bp$  alleles) in four types of dementia, including AD, dementia with Lewy's bodies (DLB), Pick's disease (PiD), and vascular dementia (VD). The results showed that the *CHRFAM7A* genotype with two wild-type copies of the gene was over-represented in the AD, DLB, and PiD cases, but not in VD [189], which may be due to the heterogeneity of the underlying pathology [190]. This observation indicates that the  $\Delta 2bp$  polymorphism, as a protective factor against the development of these forms of dementia, which are associated with abnormal protein aggregations, and again, we have to hypothesize that this may occur if  $dup\Delta \alpha 7$  behaves as a null allele, results in increased  $\alpha 7$  nAChR assembly. The protective role of nicotine in AD and PD reinforces this hypothesis since nicotine, apart from stimulating  $\alpha 7$  nAChR, down-regulates the mRNA of *CHRFAM7A* [50].

These apparently controversial results may depend on multiple confounding factors, such as the age of onset, the duration of the disease from diagnosis, and the type of disease.

However, therapeutic approaches aimed at both stimulating or antagonizing  $\alpha$ 7 nAChR failed to translate into humans [191].

By using median ganglionic eminence (MGE) neuronal progenitors, obtained by the neuronal differentiation of human induced pluripotent stem cells (iPSC) from AD patients with 0 or 1 copies of *CHRFAM7A*, it has been shown [12,192] that dup $\alpha$ 7 decreases the probability of the  $\alpha$ 7 nAChR channel opening in the presence of the positive allosteric modulator (PAM) PNU 120,596 (PNU) with currents that desensitize faster [192], and this desensitization increases as a function of the *CHRFAM7A* dosage. These data suggest that the number of *CHRFAM7A* copies may affect the response to  $\alpha$ 7 PAM, agonists, and antagonists in different individuals. Moreover, dup $\alpha$ 7 mitigates A $\beta_{1-42}$  uptake at a higher concentration, as well as the  $\alpha$ 7-dependent A $\beta$ -induced inflammatory response, suggesting a protective role in AD during the A $\beta_{1-42}$  accumulation phase. At higher A $\beta$  concentrations, the presence of dup $\alpha$ 7 increases IL-1 $\beta$  and TNF- $\alpha$  release as an ultimate signal for the activation of an anti-inflammatory response. In this sense, the absence of *CHRFAM7A* is a risk factor for AD [192]. In this iPSC model,  $dup\Delta\alpha$ 7 behaves as a null allele [12], as its expression does not affect PNU-modulated  $\alpha$ 7 nAChR currents and A $\beta_{1-42}$  uptake.

Of greater interest, is that dup $\alpha$ 7 expression affects the response to AChEi (donepezil and rivastigmine) and encenicline, a selective  $\alpha$ 7 agonist used in AD therapy [12]. In *CHRFAM7A* non-carriers, the drugs reduced A $\beta_{1-42}$  uptake, and A $\beta_{1-42}$  induced toxicity and apoptosis. On the other hand, in *CHRFAM7A* carrier cells, the neurotoxicity caused by donepezil and encenicline was unchanged, and donepezil increased apoptosis. These data suggest that *CHRFAM7A* may affect drug response.

Non-carriers of the functional *CHRFAM7A* allele comprise 25% of the human population, whereas 75% are carriers, with no differences being observed between normal aged controls and AD patients, thus suggesting that *CHRFAM7A* is not associated with a disease phenotype, which is in contrast with the data discussed above [184–187,189]. A double-blind pharmacogenetics study [12] showed that non-carriers of the functional *CHRFAM7A* allele had a superior response to the AChEi, which is in agreement with the iPSC model, and this lasted over a 7-year observation period. The *CHRFAM7A* carriers did not demonstrate a treatment response effect.

To demonstrate the role of *CHRFAM7A* in Aβ-associated neuroinflammation, the same group [193] showed that dup $\alpha$ 7 also mitigates A $\beta_{1-42}$  uptake in iPSC-derived microglial-like cells, but unlike the iPSC-derived neurons, the presence of CHRFAM7A induced a high NF-κB-mediated innate immune response by prolonging its nuclear presence, resulting in microglia activation. In these cells, nicotine paradoxically increases the proinflammatory response to LPS in *CHRFAM7A* carrier cell lines, thus suggesting that the presence of dup $\alpha$ 7 antagonizes the anti-inflammatory role of homomeric  $\alpha$ 7. These data are in

line with earlier results obtained from the overexpression of *CHRFAM7A* in RAW264.7 murine macrophages [64], but not with the findings of other groups [141,143]. A limit of these studies is that most of the conclusions rely on data obtained from comparing a *CHRFAM7A* non-carrier cell line and its isogenic counterpart, which expresses *CHRFAM7A* under the control of a constitutive promoter. LPS, via an NF- $\kappa$ B dependent mechanism, and nicotine, down-regulate *CHRFAM7A* expression [50,53], an aspect that might lead to different conclusions, and it would be interesting to know whether A $\beta_{1-42}$  plays a similar role. Similar data on the LPS/nicotine synergism obtained in a native *CHRFAM7A* carrier cell line [193] may suffer from long-term LPS treatment (24 h), as it has been shown that 3–6 h of LPS down-regulates *CHRFAM7A* expression [53] that returns to basal level after 24 h, but no data on *CHRFAM7A* expression have been presented. On the other hand, long-term exposure to LPS may resemble a chronic inflammatory insult, such as that caused by A $\beta$  in AD, where an excessive inflammatory insult may lead to CAIP dysregulation.

Proteomic profiling in a *CHRFAM7A* transgenic mouse brain [71] showed that dup $\alpha$ 7 overexpression modulates the expression of proteins involved in the  $\alpha$ 7 nAChR signaling pathways, and that they are related to the pathogenesis of neurological and neuropsychiatric disorders, including Parkinson's and Alzheimer's disease. In particular, most of these genes are mitochondrial components, and are involved in the maintenance of the anti-oxidative response, whose deregulation is one of the main causes of neurodegenerative and neuropsychiatric disorders.

The HIV-1 (human immunodeficiency virus type-1) envelope protein gp120<sub>IIIB</sub> induces  $\alpha$ 7 nAChR in neuronal cells and in the brain, particularly in the striatum, the basal ganglia's primary input, and promotes cell death in a calcium-dependent manner, an effect abrogated by  $\alpha$ 7 antagonists [194]. Conversely, *CHRFAM7A* is down-regulated in some patients [61], thus increasing the *CHRNA7/CHRFAM7A* ratio.  $\alpha$ 7/dup $\alpha$ 7 deregulation may thus play a role in the development of HIV-1-associated neurocognitive disorders (HAND) [61].

#### 3.5. Cancer

The dysregulation of nAChRs gene expression and/or the dysfunction of these receptors may be involved in the development of lung tumors [45].  $\alpha$ 7 nAChR is overexpressed in squamous cells, but not in adenocarcinoma lung tumors, and is a determinant for the progression of this type of cancer. Nicotine further increases *CHRNA7* expression and function, mediating its proliferative, survival, and angiogenic effects. In both cell types, increased  $\alpha$ 5 and  $\beta$ 4 accompanies *dup* $\alpha$ 7 and  $\beta$ 3 down-regulation. In particular, reduced *CHRFAM7A* expression may facilitate the  $\alpha$ 7-mediated oncogenic process by relieving the negative effect of dup $\alpha$ 7. Indeed, its overexpression in two NSCLC cell lines (A549 and SK-MES-1) blocked nicotine- or NKK-induced tumor progression, both in vitro and in vivo, in an athymic mouse model implanted with A549dup $\alpha$ 7 or A549 xenografts [195]. These data suggest that dup $\alpha$ 7 can be considered as a therapeutic target in smoking-related cancers.

#### 4. Conclusions

Ever since its discovery in 1998, understanding the nature and function of the *CHRFAM7A* gene has become a great challenge to respond to the failure of translating  $\alpha$ 7-directed therapeutic approaches into humans, with treatments aimed at both stimulating or antagonizing  $\alpha$ 7 nAChR. Its assembly with the  $\alpha$ 7 subunit leads to the formation of heteromeric receptors, which show reduced ion conductance and a reduced affinity for  $\alpha$ -Bgtx, thus suggesting a negative role of dup $\alpha$ 7 on  $\alpha$ 7 nAChR function.

The great amount of data on *CHRFAM7A* involvement in neurological and nonneurological disorders reported here, and summarized in Table 2, showed a contrasting effect, being protective in some cases and related to a poor prognosis or a susceptibility gene in the pathogenesis of that disorder in others, but these studies have highlighted the impact of this gene in a variety of  $\alpha$ 7 nAChR-related disorders. These confounding results may suffer from being clinical studies based on small sample sizes and from the lack of expression levels detected from both *CHRNA7* and *CHRFAM7A* and/or *CHRFAM7A*  genotyping. Most importantly, the *CHRFAM7A* copy number and the presence of the  $\Delta 2bp$  polymorphism influence the quality of protein products, and therefore, the possible effect on  $\alpha$ 7 nAChR. Moreover, dup $\alpha$ 7 can play a different role depending on the tissue type, thus shifting the ionotropic feature of  $\alpha$ 7 nAChR to that of a metabotropic receptor, as indicated by the modulation of signaling pathways upon *CHRFAM7A* overexpression. One important issue to take into account is that the expression levels of *dup\alpha7* and *\alpha7* vary in different tissues and can be modulated by various stimuli. The deregulated expression of both genes (up- and down-regulated) thus contribute to altering the *CHRNA7/CHRFAM7A* ratio, paving the way to an increased detrimental effect of dup $\alpha$ 7 on  $\alpha$ 7 nAChR function.

**Table 2.** *CHRFAM7A* involvement in neurological and non-neurological disorders. ↓ Reduced expression; ↑ increased expression; =, unchanged; n.d., not determined; und, undetectable. AD, Alzheimer's disease; DLB, Dementia with Lewy's bodies; PiD, Pick's disease; PD, Parkinson's disease; HAND, HIV-1-associated neurocognitive disorder; HSC, hematopoietic stem cell; NSCLC, non-small cell lung carcinoma. Numbers in square brackets indicate references.

| Disease  | Expression Level/Genotype      |              | <b>Biological Effect</b>  | Therapeutic<br>Intervention                                       |
|--|--------------------------------|--------------|---|---|
|  | CHRNA7                         | CHRFAM7A     |   |   |
| Schizophrenia and<br>neuropsychiatric<br>disorders   |                                |              |   |   |
| Postmortem brain sample<br>from schizophrenic and<br>psychiatric patients<br>[55,59,72,73,102–106,109] | Ļ                              | ¢            | Decreased<br>CHRNA7/CHRFAM7A ratio  |   |
| Childhood-onset<br>schizophrenia [110]   | Gene duplication               | n.d.         | Increased<br>CHRNA7/CHRFAM7A ratio  |   |
| PBMC of schizophrenic patients [100,111,112]   | n.d.                           | Ļ            | Positive correlation with<br>illness severity; negative<br>correlation between<br><i>CHRFAM7A</i> expression and<br>negative psychopathology<br>score (SANS), but not with a<br>positive score (SAPS) | Antipsychotics<br>increase<br><i>CHRFAM7A</i><br>expression [100] |
| PBMC of schizophrenic patients [99]  | und                            | Ļ            | Lower CHRFAM7A<br>expression in smokers, not<br>associated with<br>diagnosis  |   |
| Association studies [9,113,117]  | 2 copies [9,113]<br>n.d. [117] | $\Delta 2bp$ | P50 sensory gating deficit  |   |
| Association studies [55]   | =                              | $\Delta 2bp$ | No differences in $\Delta 2bp$ allele<br>frequency between ethnic<br>groups; association with<br>reduced CHRFAM7A<br>expression in patients as well<br>as in<br>control group                         |   |
| Major depressive and<br>bipolar disorders, deficit in<br>episodic memory<br>[78,114,116]               | n.d.                           | $\Delta 2bp$ | SNP associated with<br>listed disorders   |   |

# Table 2. Cont.

| Disease  | Expression Level/Genotype |                      | <b>Biological Effect</b>  | Therapeutic<br>Intervention |
|--|---------------------------|----------------------|---|-----------------------------|
| Antisaccade<br>performance [119]   | n.d.                      | $\Delta 2bp$         | No association  |                             |
| Epilepsy and<br>Neurodevelopmental<br>disorders                                  |                           |                      |   |                             |
| Idiopathic generalized<br>epilepsies (IGEs) [120]                                | 2 copies                  | $\Delta 2bp$         | Frequency of Δ2 <i>bp</i> carriers<br>was lower in IGE patients'<br>cohort versus<br>healthy controls   |                             |
| Idiopathic generalized<br>epilepsies (IGEs)<br>[121,123–125]                     | 1.5 Mb microdeletion      | n.d.                 | Role in IGE pathogenesis  |                             |
| Genetic generalized<br>epilepsy [36]   | Missense mutations        | $\Delta 2bp$         | No association with Δ2bp<br>allele; association with<br>missense mutations<br>in CHRNA7   |                             |
| Attention Deficit<br>Hyperactivity<br>Disorder (ADHD) [38]                       | 15q13.3 duplication       | n.d.                 | Duplication involves<br>CHRNA7, considered a<br>risk factor   |                             |
| Tourette Syndrome,<br>ADHD, and obsessive<br>compulsive disorders<br>(OCD) [126] | Microduplication          | CNV and $\Delta 2bp$ | Phenotypic variation in a<br>family with all the listed<br>disorders; altered<br>CHRNA7/CHRFAM7A ratio  |                             |
| Rett syndrome [60]   | $\downarrow$              | n.d.                 | It was hypothesized that<br>MeCP2 modulates both<br>CHRNA7 and CHRFAM7A<br>expression by epigenetic<br>modifications  |                             |
| Autosomal dominant<br>nocturnal frontal lobe<br>epilepsy (ADNFLE) [131]          | n.d.                      | Not expressed        | CHRFAM7A is expressed in<br>PBMC of healthy individuals<br>but not in ADNFLE patients,<br>suggesting it can be an<br>important factor in ADNFLE<br>pathogenesis |                             |
| Inflammatory<br>diseases   |                           |                      |   |                             |
| Sepsis [138]   | High                      | High/low             | Altered<br>CHRNA7/CHRFAM7A ratio:<br>high CHRFAM7A/CHRNA7<br>ratio has a poor prognosis<br>compared with<br>patients expressing higher<br>CHRNA7 levels         | prognostic marker           |
| Inflammatory Bowel<br>Disease (IBD) [139]  | $\downarrow$              | Ť                    |   | dupα7<br>down-regulation    |
| Osteoarthritis (OA) [27]   | =                         | High/low             | CHRFAM7A expression<br>correlates with MMP-3 and<br>MMP-13 mRNA level in<br>human OA chondrocytes   | α7 agonists                 |

| Disease  | Expression Level/Genotype |                                  | <b>Biological Effect</b>  | Therapeutic<br>Intervention                                    |
|--|---------------------------|----------------------------------|---|--|
| Rheumatoid<br>arthritis (RA) [62]                      | Expressed                 | Expressed                        | α7 silencing/α7 agonists<br>reveal a role for CHRNA7 in<br>controlling joint<br>inflammation. Both<br>isoforms are<br>expressed in synovial tissues<br>from RA patients, but no<br>conclusions on the role of<br>CHRFAM7A can be drawn<br>due to the missing<br>healthy controls  |  |
| Cerebral<br>ischemia/reperfusion (I/R)<br>injury [141] | n.d.                      | Ļ                                | dupα7 has a protective role;<br>its expression is negatively<br>related to the expression of<br>inflammatory cytokines  | dupα7<br>up-regulation (see<br>OGD/R microglia<br>cells model) |
| Hypertrophic scars<br>(HTS) [143]                      | n.d.                      | $\downarrow$                     | Its expression ameliorates<br>HTS formation   | See animal model   |
| Radiotherapy-induced<br>lacrimal gland injury [144]    | n.d.                      | ¢                                | Inhibition of the p38/JNK<br>signaling pathway and<br>oxidative stress  |  |
| COVID-19 [14]  | und                       | Ļ                                | Reduction correlates<br>with disease<br>severity  |  |
| Spinal cord injury (SCI)<br>[13,152]                   | n.d.                      | $\Delta 2bp$                     | Its presence may<br>affect clinical<br>outcomes. Positively<br>correlated with high levels of<br>inflammatory molecules in<br>severe SCI  |  |
| Neurodegenerative<br>diseases                          |                           |                                  |   |  |
| MCI and late-onset AD patients [184–187]               | n.d.                      | Ļ                                | dupα7 has a protective role<br>as the<br>increased functional<br>α7nAChR, due to lower<br><i>CHRFAM7A</i> expression, will<br>sustain enhanced Aβ <sub>1-42</sub><br>internalization and neuronal<br>vulnerability  | α7 antagonists<br>dupα7<br>up-regulation                       |
| AD, DLB and PiD [189]                                  | n.d.                      | wt allele<br>over-represented    | $\Delta 2bp$ polymorphism as a protective factor  |  |
| AD: association<br>study [12]                          | 2 copies                  | 75% carriers<br>25% non-carriers | No differences between<br>normal aged and AD<br>patients in the <i>CHRFAM7A</i><br>carriers, thus suggesting that<br><i>CHRFAM7A</i> is not associated<br>with the disease phenotype.<br><i>CHRFAM7A</i> influences drug<br>response to AChEi, as<br>non-carriers showed higher<br>response to AChEi over a<br>7-year follow-up |  |

# Table 2. Cont.

# Table 2. Cont.

| Disease  | Expression Level/Genotype |                 | <b>Biological Effect</b>   | Therapeutic<br>Intervention |
|--|---------------------------|-----------------|--|-----------------------------|
| HIV-1 [61,194]   | ¢                         | Ļ               | Increased<br>CHRNA7/CHRFAM7A ratio.<br>dupα7 may play a protective<br>role against the development<br>of HAND  | dupα7<br>up-regulation      |
| Cancer   |                           |                 |  |                             |
| Squamous cell lung<br>tumor [45]   | ¢                         | Ļ               | dupα7 has a protective role,<br>as its down regulation<br>facilitates the oncogenic<br>properties of α7  | dupα7<br>up-regulation      |
| Cell and animal models   |                           |                 |  |                             |
| LPS stimulated<br>macrophages [49,53]  | $\uparrow$                | $\downarrow$    | Activation of CAIP   |                             |
| <i>CHRFAM7A</i><br>transgenic mice [70,71]   | =                         | Over-expression | Decrease in α7 nAChR<br>ligand binding at the<br>neuromuscular junction  |                             |
| <i>CHRFAM7A</i> silencing in SH-SY5Y cells [58]  | =                         | Over-expression | Increased α7<br>nAChR induced<br>neurotransmitter release  |                             |
| LPS stimulated gut<br>epithelial cells [56]  | Small changes             | Ļ               | CHRFAM7A/CHRNA7 ratio<br>increase varies depending on<br>gut epithelial cell line  |                             |
| OGD/R-treated<br>microglia human<br>cell lines (model of I/R<br>injury) [141]                          | n.d.                      | Over-expression | Attenuation of microglia<br>mediated-inflammatory<br>response by<br>increased cell proliferation,<br>decreased pro-inflammatory<br>cytokines, and promotion of<br>M1 to M2 microglia<br>polarization |                             |
| Human HTS-like SCID<br>mouse model [143]   | n.d.                      | Over-expression | Positive role in the<br>amelioration of HTS<br>formation by decreasing<br>TGF-β and CTFG expression<br>and increasing<br>MMP-1 expression  |                             |
| Donepezil-treated human<br>macrophages [49]  | ¢                         | Ļ               | The increased CHRFAM7A<br>expression suggested a role<br>in controlling excessive<br>CAIP activation   |                             |
| <i>CHRFAM7A</i> transgenic<br>model of human systemic<br>inflammatory response<br>syndrome (SIRS) [11] | n.d.                      | Over-expression | Increased HSC reservoir,<br>increased immune cell<br>mobilization, myeloid cell<br>differentiation   |                             |
| Monocyte-like cells<br>(THP-1) [54,155]  | ¢                         | Over-expression | Reduced cell migration and<br>chemotaxis to monocyte<br>chemo-attractant protein<br>(MCP-1); inhibition of<br>anchorage-<br>independent colony<br>formation; increased<br>α-Bgtx binding             |                             |

| Disease   | Expression Level/Genotype |                 | <b>Biological Effect</b>   | Therapeutic<br>Intervention |
|---|---------------------------|-----------------|--|-----------------------------|
| Medial ganglionic<br>eminence (MGE) neurons<br>derived from iPSCs<br>from AD<br>patients [12,192] | 2 copies                  | 1 сору          | dup $\alpha$ 7 mitigates A $\beta_{1-42}$<br>uptake at a higher<br>concentration, and a<br>$\alpha$ 7-dependent A $\beta$ -induced<br>inflammatory response,<br>suggesting a protective role<br>in AD during the phase of<br>A $\beta_{1-42}$ accumulation. The<br>absence of <i>CHRFAM7A</i><br>suggests a risk factor in AD.<br>On the other hand, dup $\alpha$ 7<br>expression affects the<br>response to AChEi |                             |
| AD patients' iPSC-derived<br>microglial-like cells [193]  | 2 copies                  | 1 сору          | dup $\alpha$ 7 mitigates A $\beta_{1-42}$<br>uptake and induces a high<br>NF- $\kappa$ B-mediated innate<br>immune response,<br>resulting in<br>microglia activation.<br>Nicotine increases the<br>proinflammatory response to<br>LPS in <i>CHRFAM7A</i> carrier<br>cell lines, thus suggesting<br>that the presence of dup $\alpha$ 7<br>antagonizes the homomeric<br>$\alpha$ 7 anti-inflammatory role           |                             |
| <i>CHRFAM7A</i><br>transgenic mice brains [71]  | =                         | Over-expression | Modulation of the<br>expression of<br>proteins involved in<br>$\alpha$ 7 nAChR<br>signaling pathways, and<br>related to the pathogenesis of<br>neurological and<br>neuropsychiatric disorders,<br>such as PD and AD,<br>including<br>anti-oxidative pathways   |                             |
| NSCLC cell lines (A549,<br>SK-MES-1) [195]  | =                         | Over-expression | Blocking nicotine- or<br>NKK-induced tumor<br>progression, in an athymic<br>mouse model implanted<br>with A549dupα7 or<br>A549 xenografts  |                             |

Table 2. Cont.

Many questions are still pending; in particular, the use of the *CHRFAM7A* expression level, which can be measured via blood samples, as a prognostic marker even in neurological disorders, or as therapeutic target, but whether to reduce or increase its expression, remains to be elucidated. On the other hand, computational model studies will be very important for the design of drugs targeting heteromeric receptors.

**Author Contributions:** Conceptualization, R.B., S.D.L. and D.F.; writing, R.B.; artworks, S.D.L.; writing-review and editing, R.B., S.D.L. and D.F. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

**Conflicts of Interest:** The authors declare no conflict of interest.

#### References

- Gault, J.; Robinson, M.; Berger, R.; Drebing, C.; Logel, J.; Hopkins, J.; Moore, T.; Jacobs, S.; Meriwether, J.; Choi, M.J.; et al. Genomic organization and partial duplication of the human alpha7 neuronal nicotinic acetylcholine receptor gene (CHRNA7). *Genomics* 1998, 52, 173–185. [CrossRef] [PubMed]
- Riley, B.; Williamson, M.; Collier, D.; Wilkie, H.; Makoff, A. A 3-Mb map of a large Segmental duplication overlapping the alpha7-nicotinic acetylcholine receptor gene (CHRNA7) at human 15q13-q14. *Genomics* 2002, 79, 197–209. [CrossRef] [PubMed]
- O'Bleness, M.; Searles, V.B.; Varki, A.; Gagneux, P.; Sikela, J.M. Evolution of genetic and genomic features unique to the human lineage. *Nat. Rev. Genet.* 2012, 13, 853–866. [CrossRef] [PubMed]
- 4. Locke, D.P.; Archidiacono, N.; Misceo, D.; Cardone, M.F.; Deschamps, S.; Roe, B.; Rocchi, M.; Eichler, E.E. Refinement of a chimpanzee pericentric inversion breakpoint to a segmental duplication cluster. *Genome Biol.* 2003, *4*, R50. [CrossRef]
- Locke, D.P.; Jiang, Z.; Pertz, L.M.; Misceo, D.; Archidiacono, N.; Eichler, E.E. Molecular evolution of the human chromosome 15 pericentromeric region. *Cytogenet. Genome Res.* 2005, 108, 73–82. [CrossRef]
- Flomen, R.H.; Davies, A.F.; Di Forti, M.; La Cascia, C.; Mackie-Ogilvie, C.; Murray, R.; Makoff, A.J. The copy number variant involving part of the alpha7 nicotinic receptor gene contains a polymorphic inversion. *Eur. J. Hum. Genet.* 2008, *16*, 1364–1371. [CrossRef]
- Szafranski, P.; Schaaf, C.P.; Person, R.E.; Gibson, I.B.; Xia, Z.; Mahadevan, S.; Wiszniewska, J.; Bacino, C.A.; Lalani, S.; Potocki, L.; et al. Structures and molecular mechanisms for common 15q13.3 microduplications involving CHRNA7: Benign or pathological? *Hum. Mutat.* 2010, *31*, 840–850. [CrossRef]
- 8. Sinkus, M.L.; Graw, S.; Freedman, R.; Ross, R.G.; Lester, H.A.; Leonard, S. The human CHRNA7 and CHRFAM7A genes: A review of the genetics, regulation, and function. *Neuropharmacology* **2015**, *96*, 274–288. [CrossRef]
- Sinkus, M.L.; Lee, M.J.; Gault, J.; Logel, J.; Short, M.; Freedman, R.; Christian, S.L.; Lyon, J.; Leonard, S. A 2-base pair deletion polymorphism in the partial duplication of the alpha7 nicotinic acetylcholine gene (CHRFAM7A) on chromosome 15q14 is associated with schizophrenia. *Brain Res.* 2009, 1291, 1–11. [CrossRef]
- Taske, N.L.; Williamson, M.P.; Makoff, A.; Bate, L.; Curtis, D.; Kerr, M.; Kjeldsen, M.J.; Pang, K.A.; Sundqvist, A.; Friis, M.L.; et al. Evaluation of the positional candidate gene CHRNA7 at the juvenile myoclonic epilepsy locus (EJM2) on chromosome 15q13-14. Epilepsy Res 2002, 49, 157–172. [CrossRef]
- Costantini, T.W.; Chan, T.W.; Cohen, O.; Langness, S.; Treadwell, S.; Williams, E.; Eliceiri, B.P.; Baird, A. Uniquely human CHRFAM7A gene increases the hematopoietic stem cell reservoir in mice and amplifies their inflammatory response. *Proc. Natl. Acad. Sci. USA* 2019, *116*, 7932–7940. [CrossRef] [PubMed]
- Szigeti, K.; Ihnatovych, I.; Birkaya, B.; Chen, Z.; Ouf, A.; Indurthi, D.C.; Bard, J.E.; Kann, J.; Adams, A.; Chaves, L.; et al. CHRFAM7A: A human specific fusion gene, accounts for the translational gap for cholinergic strategies in Alzheimer's disease. *EBioMedicine* 2020, 59, 102892. [CrossRef] [PubMed]
- Lin, M.; Huang, W.; Kabbani, N.; Theiss, M.M.; Hamilton, J.F.; Ecklund, J.M.; Conley, Y.P.; Vodovotz, Y.; Brienza, D.; Wagner, A.K.; et al. Effect of CHRFAM7A Δ2bp gene variant on secondary inflammation after spinal cord injury. *PLoS ONE* 2021, *16*, e0251110. [CrossRef] [PubMed]
- Courties, A.; Boussier, J.; Hadjadj, J.; Yatim, N.; Barnabei, L.; Péré, H.; Veyer, D.; Kernéis, S.; Carlier, N.; Pène, F.; et al. Regulation of the acetylcholine/α7nAChR anti-inflammatory pathway in COVID-19 patients. *Sci. Rep.* 2021, *11*, 11886. [CrossRef]
- 15. Thompson, A.J.; Lester, H.A.; Lummis, S.C. The structural basis of function in Cys-loop receptors. *Q. Rev. Biophys.* **2010**, *43*, 449–499. [CrossRef]
- 16. Fasoli, F.; Gotti, C. Structure of neuronal nicotinic receptors. Curr. Top. Behav. Neurosci. 2015, 23, 1–17. [CrossRef]
- 17. Schaaf, C.P. Nicotinic acetylcholine receptors in human genetic disease. Genet. Med. 2014, 16, 649–656. [CrossRef]
- Shytle, R.D.; Mori, T.; Townsend, K.; Vendrame, M.; Sun, N.; Zeng, J.; Ehrhart, J.; Silver, A.A.; Sanberg, P.R.; Tan, J. Cholinergic modulation of microglial activation by alpha 7 nicotinic receptors. J. Neurochem. 2004, 89, 337–343. [CrossRef]
- Gamage, R.; Wagnon, I.; Rossetti, I.; Childs, R.; Niedermayer, G.; Chesworth, R.; Gyengesi, E. Cholinergic modulation of glial function during aging and chronic neuroinflammation. *Front. Cell. Neurosci.* 2020, 14, 577912. [CrossRef]
- Patel, H.; McIntire, J.; Ryan, S.; Dunah, A.; Loring, R. Anti-inflammatory effects of astroglial α7 nicotinic acetylcholine receptors are mediated by inhibition of the NF-κB pathway and activation of the Nrf2 pathway. J. Neuroinflamm. 2017, 14, 192. [CrossRef]
- 21. Song, P.; Spindel, E.R. Basic and clinical aspects of non-neuronal acetylcholine: Expression of non-neuronal acetylcholine in lung cancer provides a new target for cancer therapy. *J. Pharmacol. Sci.* **2008**, *106*, 180–185. [CrossRef] [PubMed]
- 22. Son, J.H.; Meizel, S. Evidence suggesting that the mouse sperm acrosome reaction initiated by the zona pellucida involves an alpha7 nicotinic acetylcholine receptor. *Biol. Reprod.* 2003, *68*, 1348–1353. [CrossRef] [PubMed]

- Summers, A.E.; Whelan, C.J.; Parsons, M.E. Nicotinic acetylcholine receptor subunits and receptor activity in the epithelial cell line HT29. *Life Sci.* 2003, 72, 2091–2094. [CrossRef]
- Saeed, R.W.; Varma, S.; Peng-Nemeroff, T.; Sherry, B.; Balakhaneh, D.; Huston, J.; Tracey, K.J.; Al-Abed, Y.; Metz, C.N. Cholinergic stimulation blocks endothelial cell activation and leukocyte recruitment during inflammation. *J. Exp. Med.* 2005, 201, 1113–1123. [CrossRef] [PubMed]
- 25. Roman, J.; Ritzenthaler, J.D.; Gil-Acosta, A.; Rivera, H.N.; Roser-Page, S. Nicotine and fibronectin expression in lung fibroblasts: Implications for tobacco-related lung tissue remodeling. *FASEB J.* **2004**, *18*, 1436–1438. [CrossRef] [PubMed]
- Olofsson, P.S.; Katz, D.A.; Rosas-Ballina, M.; Levine, Y.A.; Ochani, M.; Valdés-Ferrer, S.I.; Pavlov, V.A.; Tracey, K.J.; Chavan, S.S. α7 nicotinic acetylcholine receptor (α7nAChR) expression in bone marrow-derived non-T cells is required for the inflammatory reflex. *Mol. Med.* 2012, *18*, 539–543. [CrossRef]
- Courties, A.; Do, A.; Leite, S.; Legris, M.; Sudre, L.; Pigenet, A.; Petit, J.; Nourissat, G.; Cambon-Binder, A.; Maskos, U.; et al. The role of the non-neuronal cholinergic system in inflammation and degradation processes in osteoarthritis. *Arthritis Rheumatol.* 2020, 72, 2072–2082. [CrossRef]
- Skok, M.; Grailhe, R.; Agenes, F.; Changeux, J.P. The role of nicotinic acetylcholine receptors in lymphocyte development. J. Neuroimmunol. 2006, 171, 86–98. [CrossRef]
- Skok, M.; Grailhe, R.; Changeux, J.P. Nicotinic receptors regulate B lymphocyte activation and immune response. *Eur. J. Pharmacol.* 2005, 517, 246–251. [CrossRef]
- 30. de Jonge, W.J.; Ulloa, L. The alpha7 nicotinic acetylcholine receptor as a pharmacological target for inflammation. *Br. J. Pharmacol.* **2007**, *151*, 915–929. [CrossRef]
- 31. Benfante, R.; Di Lascio, S.; Cardani, S.; Fornasari, D. Acetylcholinesterase inhibitors targeting the cholinergic anti-inflammatory pathway: A new therapeutic perspective in aging-related disorders. *Aging Clin. Exp. Res.* **2021**, *33*, 823–834. [CrossRef] [PubMed]
- Wang, H.; Yu, M.; Ochani, M.; Amella, C.A.; Tanovic, M.; Susarla, S.; Li, J.H.; Yang, H.; Ulloa, L.; Al-Abed, Y.; et al. Nicotinic acetylcholine receptor alpha7 subunit is an essential regulator of inflammation. *Nature* 2003, 421, 384–388. [CrossRef] [PubMed]
- Hoover, D.B. Cholinergic modulation of the immune system presents new approaches for treating inflammation. *Pharmacol. Ther.* 2017, 179, 1–16. [CrossRef] [PubMed]
- Leonard, S.; Gault, J.; Hopkins, J.; Logel, J.; Vianzon, R.; Short, M.; Drebing, C.; Berger, R.; Venn, D.; Sirota, P.; et al. Association of promoter variants in the alpha7 nicotinic acetylcholine receptor subunit gene with an inhibitory deficit found in schizophrenia. *Arch. Gen. Psychiatry* 2002, 59, 1085–1096. [CrossRef]
- 35. Bacchelli, E.; Battaglia, A.; Cameli, C.; Lomartire, S.; Tancredi, R.; Thomson, S.; Sutcliffe, J.S.; Maestrini, E. Analysis of CHRNA7 rare variants in autism spectrum disorder susceptibility. *Am. J. Med. Genet. A* 2015, *167A*, 715–723. [CrossRef]
- Damiano, J.A.; Mullen, S.A.; Hildebrand, M.S.; Bellows, S.T.; Lawrence, K.M.; Arsov, T.; Dibbens, L.; Major, H.; Dahl, H.H.; Mefford, H.C.; et al. Evaluation of multiple putative risk alleles within the 15q13.3 region for genetic generalized epilepsy. *Epilepsy Res.* 2015, *117*, 70–73. [CrossRef]
- 37. Masurel-Paulet, A.; Andrieux, J.; Callier, P.; Cuisset, J.M.; Le Caignec, C.; Holder, M.; Thauvin-Robinet, C.; Doray, B.; Flori, E.; Alex-Cordier, M.P.; et al. Delineation of 15q13.3 microdeletions. *Clin. Genet.* **2010**, *78*, 149–161. [CrossRef]
- Williams, N.M.; Franke, B.; Mick, E.; Anney, R.J.; Freitag, C.M.; Gill, M.; Thapar, A.; O'Donovan, M.C.; Owen, M.J.; Holmans, P.; et al. Genome-wide analysis of copy number variants in attention deficit hyperactivity disorder: The role of rare variants and duplications at 15q13.3. *Am. J. Psychiatry* 2012, *169*, 195–204. [CrossRef]
- 39. Deutsch, S.I.; Burket, J.A.; Benson, A.D. Targeting the α7 nicotinic acetylcholine receptor to prevent progressive dementia and improve cognition in adults with Down's syndrome. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **2014**, *54*, 131–139. [CrossRef]
- 40. Quik, M.; Zhang, D.; McGregor, M.; Bordia, T. Alpha7 nicotinic receptors as therapeutic targets for Parkinson's disease. *Biochem. Pharmacol.* 2015, 97, 399–407. [CrossRef]
- Ma, K.G.; Qian, Y.H. Alpha 7 nicotinic acetylcholine receptor and its effects on Alzheimer's disease. *Neuropeptides* 2019, 73, 96–106. [CrossRef] [PubMed]
- Tropea, M.R.; Li Puma, D.D.; Melone, M.; Gulisano, W.; Arancio, O.; Grassi, C.; Conti, F.; Puzzo, D. Genetic deletion of α7 nicotinic acetylcholine receptors induces an age-dependent Alzheimer's disease-like pathology. *Prog. Neurobiol.* 2021, 206, 102154. [CrossRef] [PubMed]
- Mucchietto, V.; Fasoli, F.; Pucci, S.; Moretti, M.; Benfante, R.; Maroli, A.; Di Lascio, S.; Bolchi, C.; Pallavicini, M.; Dowell, C.; et al. α9- and α7-containing receptors mediate the pro-proliferative effects of nicotine in the A549 adenocarcinoma cell line. *Br. J. Pharmacol.* 2018, 175, 1957–1972. [CrossRef]
- Pucci, S.; Fasoli, F.; Moretti, M.; Benfante, R.; Di Lascio, S.; Viani, P.; Daga, A.; Gordon, T.J.; McIntosh, M.; Zoli, M.; et al. Choline and nicotine increase glioblastoma cell proliferation by binding and activating α7- and α9- containing nicotinic receptors. *Pharmacol. Res.* 2021, 163, 105336. [CrossRef] [PubMed]
- Bordas, A.; Cedillo, J.L.; Arnalich, F.; Esteban-Rodriguez, I.; Guerra-Pastrián, L.; de Castro, J.; Martín-Sánchez, C.; Atienza, G.; Fernández-Capitan, C.; Rios, J.J.; et al. Expression patterns for nicotinic acetylcholine receptor subunit genes in smoking-related lung cancers. *Oncotarget* 2017, *8*, 67878–67890. [CrossRef]
- Hajiasgharzadeh, K.; Somi, M.H.; Sadigh-Eteghad, S.; Mokhtarzadeh, A.; Shanehbandi, D.; Mansoori, B.; Mohammadi, A.; Doustvandi, M.A.; Baradaran, B. The dual role of alpha7 nicotinic acetylcholine receptor in inflammation-associated gastrointestinal cancers. *Heliyon* 2020, 6, e03611. [CrossRef]

- Vieira-Alves, I.; Coimbra-Campos, L.M.C.; Sancho, M.; da Silva, R.F.; Cortes, S.F.; Lemos, V.S. Role of the α7 Nicotinic Acetylcholine Receptor in the Pathophysiology of Atherosclerosis. *Front. Physiol.* 2020, *11*, 621769. [CrossRef]
- Zoli, M.; Pucci, S.; Vilella, A.; Gotti, C. Neuronal and extraneuronal nicotinic acetylcholine receptors. *Curr. Neuropharmacol.* 2018, 16, 338–349. [CrossRef]
- Maroli, A.; Di Lascio, S.; Drufuca, L.; Cardani, S.; Setten, E.; Locati, M.; Fornasari, D.; Benfante, R. Effect of donepezil on the expression and responsiveness to LPS of CHRNA7 and CHRFAM7A in macrophages: A possible link to the cholinergic anti-inflammatory pathway. J. Neuroimmunol. 2019, 332, 155–166. [CrossRef]
- 50. de Lucas-Cerrillo, A.M.; Maldifassi, M.C.; Arnalich, F.; Renart, J.; Atienza, G.; Serantes, R.; Cruces, J.; Sánchez-Pacheco, A.; Andrés-Mateos, E.; Montiel, C. Function of partially duplicated human α77 nicotinic receptor subunit CHRFAM7A gene: Potential implications for the cholinergic anti-inflammatory response. *J. Biol. Chem.* **2011**, *286*, 594–606. [CrossRef]
- 51. Araud, T.; Graw, S.; Berger, R.; Lee, M.; Neveu, E.; Bertrand, D.; Leonard, S. The chimeric gene CHRFAM7A, a partial duplication of the CHRNA7 gene, is a dominant negative regulator of α7\*nAChR function. *Biochem. Pharmacol.* 2011, 82, 904–914. [CrossRef] [PubMed]
- 52. Wang, Y.; Xiao, C.; Indersmitten, T.; Freedman, R.; Leonard, S.; Lester, H.A. The duplicated α7 subunits assemble and form functional nicotinic receptors with the full-length α7. *J. Biol. Chem.* **2014**, *289*, 26451–26463. [CrossRef] [PubMed]
- Benfante, R.; Antonini, R.A.; De Pizzol, M.; Gotti, C.; Clementi, F.; Locati, M.; Fornasari, D. Expression of the α7 nAChR subunit duplicate form (CHRFAM7A) is down-regulated in the monocytic cell line THP-1 on treatment with LPS. J. Neuroimmunol. 2011, 230, 74–84. [CrossRef] [PubMed]
- 54. Costantini, T.W.; Dang, X.; Yurchyshyna, M.V.; Coimbra, R.; Eliceiri, B.P.; Baird, A. A human-specific α7-nicotinic acetylcholine receptor gene in human leukocytes: Identification, regulation and the consequences of CHRFAM7A expression. *Mol. Med.* **2015**, 21, 323–336. [CrossRef]
- Kunii, Y.; Zhang, W.; Xu, Q.; Hyde, T.M.; McFadden, W.; Shin, J.H.; Deep-Soboslay, A.; Ye, T.; Li, C.; Kleinman, J.E.; et al. CHRNA7 and CHRFAM7A mRNAs: Co-localized and their expression levels altered in the postmortem dorsolateral prefrontal cortex in major psychiatric disorders. *Am. J. Psychiatry* 2015, 172, 1122–1130. [CrossRef]
- Dang, X.; Eliceiri, B.P.; Baird, A.; Costantini, T.W. CHRFAM7A: A human-specific α7-nicotinic acetylcholine receptor gene shows differential responsiveness of human intestinal epithelial cells to LPS. *FASEB J.* 2015, 29, 2292–2302. [CrossRef]
- Villiger, Y.; Szanto, I.; Jaconi, S.; Blanchet, C.; Buisson, B.; Krause, K.H.; Bertrand, D.; Romand, J.A. Expression of an alpha7 duplicate nicotinic acetylcholine receptor-related protein in human leukocytes. J. Neuroimmunol. 2002, 126, 86–98. [CrossRef]
- 58. Martín-Sánchez, C.; Alés, E.; Balseiro-Gómez, S.; Atienza, G.; Arnalich, F.; Bordas, A.; Cedillo, J.L.; Extremera, M.; Chávez-Reyes, A.; Montiel, C. The human-specific duplicated α7 gene inhibits the ancestral α7, negatively regulating nicotinic acetylcholine receptor-mediated transmitter release. *J. Biol. Chem.* 2021, 296, 100341. [CrossRef]
- De Luca, V.; Likhodi, O.; Van Tol, H.H.; Kennedy, J.L.; Wong, A.H. Regulation of alpha7-nicotinic receptor subunit and alpha7-like gene expression in the prefrontal cortex of patients with bipolar disorder and schizophrenia. *Acta Psychiatr. Scand.* 2006, 114, 211–215. [CrossRef]
- Yasui, D.H.; Scoles, H.A.; Horike, S.; Meguro-Horike, M.; Dunaway, K.W.; Schroeder, D.I.; Lasalle, J.M. 15q11.2-13.3 chromatin analysis reveals epigenetic regulation of CHRNA7 with deficiencies in Rett and autism brain. *Hum. Mol. Genet.* 2011, 20, 4311–4323. [CrossRef]
- Ramos, F.M.; Delgado-Vélez, M.; Ortiz, Á.; Báez-Pagán, C.A.; Quesada, O.; Lasalde-Dominicci, J.A. Expression of CHRFAM7A and CHRNA7 in neuronal cells and postmortem brain of HIV-infected patients: Considerations for HIV-associated neurocognitive disorder. J. Neurovirol. 2016, 22, 327–335. [CrossRef] [PubMed]
- van Maanen, M.A.; Stoof, S.P.; van der Zanden, E.P.; de Jonge, W.J.; Janssen, R.A.; Fischer, D.F.; Vandeghinste, N.; Brys, R.; Vervoordeldonk, M.J.; Tak, P.P. The alpha7 nicotinic acetylcholine receptor on fibroblast-like synoviocytes and in synovial tissue from rheumatoid arthritis patients: A possible role for a key neurotransmitter in synovial inflammation. *Arthritis Rheum.* 2009, 60, 1272–1281. [CrossRef] [PubMed]
- 63. Lasala, M.; Corradi, J.; Bruzzone, A.; Esandi, M.D.C.; Bouzat, C. A human-specific, truncated alpha7 nicotinic receptor subunit assembles with full-length alpha7 and forms functional receptors with different stoichiometries. *J. Biol. Chem.* **2018**, 293, 10707–10717. [CrossRef] [PubMed]
- 64. Maldifassi, M.C.; Martín-Sánchez, C.; Atienza, G.; Cedillo, J.L.; Arnalich, F.; Bordas, A.; Zafra, F.; Giménez, C.; Extremera, M.; Renart, J.; et al. Interaction of the α7-nicotinic subunit with its human-specific duplicated dupα7 isoform in mammalian cells: Relevance in human inflammatory responses. *J. Biol. Chem.* 2018, 293, 13874–13888. [CrossRef] [PubMed]
- 65. Lee, W.Y.; Sine, S.M. Principal pathway coupling agonist binding to channel gating in nicotinic receptors. *Nature* **2005**, *438*, 243–247. [CrossRef] [PubMed]
- 66. Bouzat, C.; Gumilar, F.; Spitzmaul, G.; Wang, H.L.; Rayes, D.; Hansen, S.B.; Taylor, P.; Sine, S.M. Coupling of agonist binding to channel gating in an ACh-binding protein linked to an ion channel. *Nature* **2004**, *430*, 896–900. [CrossRef]
- 67. Cameli, C.; Bacchelli, E.; De Paola, M.; Giucastro, G.; Cifiello, S.; Collo, G.; Cainazzo, M.M.; Pini, L.A.; Maestrini, E.; Zoli, M. Genetic variation in CHRNA7 and CHRFAM7A is associated with nicotine dependence and response to varenicline treatment. *Eur. J. Hum. Genet.* **2018**, *26*, 1824–1831. [CrossRef]
- Andersen, N.; Corradi, J.; Sine, S.M.; Bouzat, C. Stoichiometry for activation of neuronal α7 nicotinic receptors. *Proc. Natl. Acad. Sci. USA* 2013, 110, 20819–20824. [CrossRef]

- Liu, D.; de Souza, J.V.; Ahmad, A.; Bronowska, A.K. Structure, dynamics, and ligand recognition of human-specific CHRFAM7A (Dupα7) nicotinic receptor linked to neuropsychiatric disorders. *Int. J. Mol. Sci.* 2021, 22, 5466. [CrossRef]
- Chan, T.; Williams, E.; Cohen, O.; Eliceiri, B.P.; Baird, A.; Costantini, T.W. CHRFAM7A alters binding to the neuronal alpha-7 nicotinic acetylcholine receptor. *Neurosci. Lett.* 2019, 690, 126–131. [CrossRef]
- Jiang, Y.; Yuan, H.; Huang, L.; Hou, X.; Zhou, R.; Dang, X. Global proteomic profiling of the uniquely human CHRFAM7A gene in transgenic mouse brain. *Gene* 2019, 714, 143996. [CrossRef] [PubMed]
- 72. Stefansson, H.; Rujescu, D.; Cichon, S.; Pietiläinen, O.P.; Ingason, A.; Steinberg, S.; Fossdal, R.; Sigurdsson, E.; Sigmundsson, T.; Buizer-Voskamp, J.E.; et al. Large recurrent microdeletions associated with schizophrenia. *Nature* 2008, 455, 232–236. [CrossRef] [PubMed]
- 73. Stone, J.L.; O'Donovan, M.C.; Gurling, H.; Kirov, G.K.; Blackwood, D.H.R.; Corvin, A.; Craddock, N.J.; Gill, M.; Hultman, C.M.; Lichtenstein, P.; et al. Rare chromosomal deletions and duplications increase risk of schizophrenia. *Nature* 2008, 455, 237–241. [CrossRef]
- 74. Leonard, S.; Freedman, R. Genetics of chromosome 15q13-q14 in schizophrenia. *Biol. Psychiatry* **2006**, *60*, 115–122. [CrossRef] [PubMed]
- Elmslie, F.V.; Rees, M.; Williamson, M.P.; Kerr, M.; Kjeldsen, M.J.; Pang, K.A.; Sundqvist, A.; Friis, M.L.; Chadwick, D.; Richens, A.; et al. Genetic mapping of a major susceptibility locus for juvenile myoclonic epilepsy on chromosome 15q. *Hum. Mol. Genet.* 1997, *6*, 1329–1334. [CrossRef] [PubMed]
- 76. Neubauer, B.A.; Fiedler, B.; Himmelein, B.; Kämpfer, F.; Lässker, U.; Schwabe, G.; Spanier, I.; Tams, D.; Bretscher, C.; Moldenhauer, K.; et al. Centrotemporal spikes in families with rolandic epilepsy: Linkage to chromosome 15q14. *Neurology* 1998, *51*, 1608–1612. [CrossRef]
- 77. Joo, E.J.; Lee, K.Y.; Kim, H.S.; Kim, S.H.; Ahn, Y.M.; Kim, Y.S. Genetic association study of the alpha 7 nicotinic receptor (CHRNA7) with the development of schizophrenia and bipolar disorder in korean population. *Psychiatry Investig.* 2010, 7, 196–201. [CrossRef]
- Lai, I.C.; Hong, C.J.; Tsai, S.J. Association study of nicotinic-receptor variants and major depressive disorder. J. Affect. Disord. 2001, 66, 79–82. [CrossRef]
- 79. Stassen, H.H.; Bridler, R.; Hägele, S.; Hergersberg, M.; Mehmann, B.; Schinzel, A.; Weisbrod, M.; Scharfetter, C. Schizophrenia and smoking: Evidence for a common neurobiological basis? *Am. J. Med. Genet.* 2000, *96*, 173–177. [CrossRef]
- 80. Cooper, G.M.; Coe, B.P.; Girirajan, S.; Rosenfeld, J.A.; Vu, T.H.; Baker, C.; Williams, C.; Stalker, H.; Hamid, R.; Hannig, V.; et al. A copy number variation morbidity map of developmental delay. *Nat. Genet.* **2011**, *43*, 838–846. [CrossRef]
- Kaminsky, E.B.; Kaul, V.; Paschall, J.; Church, D.M.; Bunke, B.; Kunig, D.; Moreno-De-Luca, D.; Moreno-De-Luca, A.; Mulle, J.G.; Warren, S.T.; et al. An evidence-based approach to establish the functional and clinical significance of copy number variants in intellectual and developmental disabilities. *Genet. Med.* 2011, 13, 777–784. [CrossRef] [PubMed]
- 82. Marshall, C.R.; Noor, A.; Vincent, J.B.; Lionel, A.C.; Feuk, L.; Skaug, J.; Shago, M.; Moessner, R.; Pinto, D.; Ren, Y.; et al. Structural variation of chromosomes in autism spectrum disorder. *Am. J. Hum. Genet.* **2008**, *82*, 477–488. [CrossRef] [PubMed]
- Curtis, L.; Blouin, J.L.; Radhakrishna, U.; Gehrig, C.; Lasseter, V.K.; Wolyniec, P.; Nestadt, G.; Dombroski, B.; Kazazian, H.H.; Pulver, A.E.; et al. No evidence for linkage between schizophrenia and markers at chromosome 15q13-14. *Am. J. Med. Genet.* 1999, *88*, 109–112. [CrossRef]
- Iwata, Y.; Nakajima, M.; Yamada, K.; Nakamura, K.; Sekine, Y.; Tsuchiya, K.J.; Sugihara, G.; Matsuzaki, H.; Suda, S.; Suzuki, K.; et al. Linkage disequilibrium analysis of the CHRNA7 gene and its partially duplicated region in schizophrenia. *Neurosci. Res.* 2007, 57, 194–202. [CrossRef] [PubMed]
- 85. Neves-Pereira, M.; Bassett, A.S.; Honer, W.G.; Lang, D.; King, N.A.; Kennedy, J.L. No evidence for linkage of the CHRNA7 gene region in Canadian schizophrenia families. *Am. J. Med. Genet.* **1998**, *81*, 361–363. [CrossRef]
- Sanders, A.R.; Duan, J.; Levinson, D.F.; Shi, J.; He, D.; Hou, C.; Burrell, G.J.; Rice, J.P.; Nertney, D.A.; Olincy, A.; et al. No significant association of 14 candidate genes with schizophrenia in a large European ancestry sample: Implications for psychiatric genetics. *Am. J. Psychiatry* 2008, 165, 497–506. [CrossRef]
- Tsuang, D.W.; Skol, A.D.; Faraone, S.V.; Bingham, S.; Young, K.A.; Prabhudesai, S.; Haverstock, S.L.; Mena, F.; Menon, A.S.; Bisset, D.; et al. Examination of genetic linkage of chromosome 15 to schizophrenia in a large Veterans Affairs Cooperative Study sample. *Am. J. Med. Genet.* 2001, 105, 662–668. [CrossRef]
- 88. Tsuang, M.T.; Stone, W.S.; Faraone, S.V. Schizophrenia: A review of genetic studies. *Harv. Rev. Psychiatry* **1999**, *7*, 185–207. [CrossRef]
- Freedman, R.; Coon, H.; Myles-Worsley, M.; Orr-Urtreger, A.; Olincy, A.; Davis, A.; Polymeropoulos, M.; Holik, J.; Hopkins, J.; Hoff, M.; et al. Linkage of a neurophysiological deficit in schizophrenia to a chromosome 15 locus. *Proc. Natl. Acad. Sci. USA* 1997, 94, 587–592. [CrossRef]
- Freedman, R.; Olincy, A.; Ross, R.G.; Waldo, M.C.; Stevens, K.E.; Adler, L.E.; Leonard, S. The genetics of sensory gating deficits in schizophrenia. *Curr. Psychiatry Rep.* 2003, *5*, 155–161. [CrossRef]
- Leonard, S.; Adler, L.E.; Benhammou, K.; Berger, R.; Breese, C.R.; Drebing, C.; Gault, J.; Lee, M.J.; Logel, J.; Olincy, A.; et al. Smoking and mental illness. *Pharmacol. Biochem. Behav.* 2001, 70, 561–570. [CrossRef]
- 92. Glassman, A.H. Cigarette smoking: Implications for psychiatric illness. Am. J. Psychiatry 1993, 150, 546–553. [CrossRef] [PubMed]

- Stephens, S.H.; Franks, A.; Berger, R.; Palionyte, M.; Fingerlin, T.E.; Wagner, B.; Logel, J.; Olincy, A.; Ross, R.G.; Freedman, R.; et al. Multiple genes in the 15q13-q14 chromosomal region are associated with schizophrenia. *Psychiatr. Genet.* 2012, 22, 1–14. [CrossRef] [PubMed]
- 94. Leonard, S.; Mexal, S.; Freedman, R. Smoking, genetics and schizophrenia: Evidence for self medication. *J. Dual. Diagn.* **2007**, *3*, 43–59. [CrossRef]
- Koike, K.; Hashimoto, K.; Takai, N.; Shimizu, E.; Komatsu, N.; Watanabe, H.; Nakazato, M.; Okamura, N.; Stevens, K.E.; Freedman, R.; et al. Tropisetron improves deficits in auditory P50 suppression in schizophrenia. *Schizophr. Res.* 2005, *76*, 67–72. [CrossRef]
- 96. Toyohara, J.; Hashimoto, K. α7 Nicotinic receptor agonists: Potential therapeutic drugs for treatment of cognitive impairments in schizophrenia and alzheimer's disease. *Open Med. Chem. J.* **2010**, *4*, 37–56. [CrossRef]
- 97. Mackowick, K.M.; Barr, M.S.; Wing, V.C.; Rabin, R.A.; Ouellet-Plamondon, C.; George, T.P. Neurocognitive endophenotypes in schizophrenia: Modulation by nicotinic receptor systems. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **2014**, *52*, 79–85. [CrossRef]
- Parikh, V.; Kutlu, M.G.; Gould, T.J. nAChR dysfunction as a common substrate for schizophrenia and comorbid nicotine addiction: Current trends and perspectives. *Schizophr. Res.* 2016, 171, 1–15. [CrossRef]
- Severance, E.G.; Dickerson, F.B.; Stallings, C.R.; Origoni, A.E.; Sullens, A.; Monson, E.T.; Yolken, R.H. Differentiating nicotineversus schizophrenia-associated decreases of the alpha7 nicotinic acetylcholine receptor transcript, CHRFAM7A, in peripheral blood lymphocytes. J. Neural Transm. 2009, 116, 213–220. [CrossRef]
- Kalmady, S.V.; Agrawal, R.; Venugopal, D.; Shivakumar, V.; Amaresha, A.C.; Agarwal, S.M.; Subbanna, M.; Rajasekaran, A.; Narayanaswamy, J.C.; Debnath, M.; et al. CHRFAM7A gene expression in schizophrenia: Clinical correlates and the effect of antipsychotic treatment. *J. Neural Transm.* 2018, 125, 741–748. [CrossRef]
- Bencherif, M.; Stachowiak, M.K.; Kucinski, A.J.; Lippiello, P.M. Alpha7 nicotinic cholinergic neuromodulation may reconcile multiple neurotransmitter hypotheses of schizophrenia. *Med. Hypotheses* 2012, 78, 594–600. [CrossRef] [PubMed]
- 102. Freedman, R.; Hall, M.; Adler, L.E.; Leonard, S. Evidence in postmortem brain tissue for decreased numbers of hippocampal nicotinic receptors in schizophrenia. *Biol. Psychiatry* **1995**, *38*, 22–33. [CrossRef]
- Guan, Z.Z.; Zhang, X.; Blennow, K.; Nordberg, A. Decreased protein level of nicotinic receptor alpha7 subunit in the frontal cortex from schizophrenic brain. *Neuroreport* 1999, 10, 1779–1782. [CrossRef] [PubMed]
- 104. Guillozet-Bongaarts, A.L.; Hyde, T.M.; Dalley, R.A.; Hawrylycz, M.J.; Henry, A.; Hof, P.R.; Hohmann, J.; Jones, A.R.; Kuan, C.L.; Royall, J.; et al. Altered gene expression in the dorsolateral prefrontal cortex of individuals with schizophrenia. *Mol. Psychiatry* 2014, 19, 478–485. [CrossRef]
- 105. Marutle, A.; Zhang, X.; Court, J.; Piggott, M.; Johnson, M.; Perry, R.; Perry, E.; Nordberg, A. Laminar distribution of nicotinic receptor subtypes in cortical regions in schizophrenia. *J. Chem. Neuroanat.* **2001**, *22*, 115–126. [CrossRef]
- Court, J.; Spurden, D.; Lloyd, S.; McKeith, I.; Ballard, C.; Cairns, N.; Kerwin, R.; Perry, R.; Perry, E. Neuronal nicotinic receptors in dementia with Lewy bodies and schizophrenia: Alpha-bungarotoxin and nicotine binding in the thalamus. *J. Neurochem.* 1999, 73, 1590–1597. [CrossRef]
- 107. Stephens, S.H.; Logel, J.; Barton, A.; Franks, A.; Schultz, J.; Short, M.; Dickenson, J.; James, B.; Fingerlin, T.E.; Wagner, B.; et al. Association of the 5'-upstream regulatory region of the alpha7 nicotinic acetylcholine receptor subunit gene (CHRNA7) with schizophrenia. *Schizophr. Res.* 2009, 109, 102–112. [CrossRef]
- Bertelsen, B.; Oranje, B.; Melchior, L.; Fagerlund, B.; Werge, T.M.; Mikkelsen, J.D.; Tümer, Z.; Glenthøj, B.Y. Association study of CHRNA7 promoter variants with sensory and sensorimotor gating in schizophrenia patients and healthy controls: A danish case-control study. *Neuromolecular. Med.* 2015, 17, 423–430. [CrossRef]
- Gault, J.; Hopkins, J.; Berger, R.; Drebing, C.; Logel, J.; Walton, C.; Short, M.; Vianzon, R.; Olincy, A.; Ross, R.G.; et al. Comparison of polymorphisms in the alpha7 nicotinic receptor gene and its partial duplication in schizophrenic and control subjects. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* 2003, 123B, 39–49. [CrossRef]
- 110. Zhou, D.; Gochman, P.; Broadnax, D.D.; Rapoport, J.L.; Ahn, K. 15q13.3 duplication in two patients with childhood-onset schizophrenia. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **2016**, *171*, 777–783. [CrossRef]
- Perl, O.; Strous, R.D.; Dranikov, A.; Chen, R.; Fuchs, S. Low levels of alpha7-nicotinic acetylcholine receptor mRNA on peripheral blood lymphocytes in schizophrenia and its association with illness severity. *Neuropsychobiology* 2006, *53*, 88–93. [CrossRef] [PubMed]
- 112. Perl, O.; Ilani, T.; Strous, R.D.; Lapidus, R.; Fuchs, S. The alpha7 nicotinic acetylcholine receptor in schizophrenia: Decreased mRNA levels in peripheral blood lymphocytes. *FASEB J.* **2003**, *17*, 1948–1950. [CrossRef] [PubMed]
- 113. Raux, G.; Bonnet-Brilhault, F.; Louchart, S.; Houy, E.; Gantier, R.; Levillain, D.; Allio, G.; Haouzir, S.; Petit, M.; Martinez, M.; et al. The -2 bp deletion in exon 6 of the 'alpha 7-like' nicotinic receptor subunit gene is a risk factor for the P50 sensory gating deficit. *Mol. Psychiatry* 2002, 7, 1006–1011. [CrossRef] [PubMed]
- 114. Hong, C.J.; Lai, I.C.; Liou, L.L.; Tsai, S.J. Association study of the human partially duplicated alpha7 nicotinic acetylcholine receptor genetic variant with bipolar disorder. *Neurosci. Lett.* **2004**, *355*, 69–72. [CrossRef]
- 115. Flomen, R.H.; Collier, D.A.; Osborne, S.; Munro, J.; Breen, G.; St Clair, D.; Makoff, A.J. Association study of CHRFAM7A copy number and 2 bp deletion polymorphisms with schizophrenia and bipolar affective disorder. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* 2006, 141B, 571–575. [CrossRef]

- 116. Dempster, E.L.; Toulopoulou, T.; McDonald, C.; Bramon, E.; Walshe, M.; Wickham, H.; Sham, P.C.; Murray, R.M.; Collier, D.A. Episodic memory performance predicted by the 2bp deletion in exon 6 of the "alpha 7-like" nicotinic receptor subunit gene. *Am. J. Psychiatry* 2006, *163*, 1832–1834. [CrossRef]
- 117. Flomen, R.H.; Shaikh, M.; Walshe, M.; Schulze, K.; Hall, M.H.; Picchioni, M.; Rijsdijk, F.; Toulopoulou, T.; Kravariti, E.; Murray, R.M.; et al. Association between the 2-bp deletion polymorphism in the duplicated version of the alpha7 nicotinic receptor gene and P50 sensory gating. *Eur. J. Hum. Genet.* 2013, 21, 76–81. [CrossRef]
- 118. Lai, I.C.; Hong, C.J.; Tsai, S.J. Association study of a nicotinic receptor variant with schizophrenic disorders. *Neuropsychobiology* **2001**, *43*, 15–18. [CrossRef]
- Petrovsky, N.; Schmechtig, A.; Flomen, R.H.; Kumari, V.; Collier, D.; Makoff, A.; Wagner, M.; Ettinger, U. CHRFAM7A copy number and 2-bp deletion polymorphisms and antisaccade performance. *Int. J. Neuropsychopharmacol.* 2009, 12, 267–273. [CrossRef]
- Rozycka, A.; Dorszewska, J.; Steinborn, B.; Lianeri, M.; Winczewska-Wiktor, A.; Sniezawska, A.; Wisniewska, K.; Jagodzinski, P.P. Association study of the 2-bp deletion polymorphism in exon 6 of the CHRFAM7A gene with idiopathic generalized epilepsy. DNA Cell Biol. 2013, 32, 640–647. [CrossRef]
- 121. Helbig, I.; Mefford, H.C.; Sharp, A.J.; Guipponi, M.; Fichera, M.; Franke, A.; Muhle, H.; de Kovel, C.; Baker, C.; von Spiczak, S.; et al. 15q13.3 microdeletions increase risk of idiopathic generalized epilepsy. *Nat. Genet.* **2009**, *41*, 160–162. [CrossRef] [PubMed]
- 122. Mullen, S.A.; Carvill, G.L.; Bellows, S.; Bayly, M.A.; Trucks, H.; Lal, D.; Sander, T.; Berkovic, S.F.; Dibbens, L.M.; Scheffer, I.E.; et al. Copy number variants are frequent in genetic generalized epilepsy with intellectual disability. *Neurology* 2013, *81*, 1507–1514. [CrossRef] [PubMed]
- 123. Mefford, H.C.; Muhle, H.; Ostertag, P.; von Spiczak, S.; Buysse, K.; Baker, C.; Franke, A.; Malafosse, A.; Genton, P.; Thomas, P.; et al. Genome-wide copy number variation in epilepsy: Novel susceptibility loci in idiopathic generalized and focal epilepsies. *PLoS Genet.* 2010, 6, e1000962. [CrossRef] [PubMed]
- 124. Dibbens, L.M.; Mullen, S.; Helbig, I.; Mefford, H.C.; Bayly, M.A.; Bellows, S.; Leu, C.; Trucks, H.; Obermeier, T.; Wittig, M.; et al. Familial and sporadic 15q13.3 microdeletions in idiopathic generalized epilepsy: Precedent for disorders with complex inheritance. *Hum. Mol. Genet.* **2009**, *18*, 3626–3631. [CrossRef] [PubMed]
- 125. Muhle, H.; Mefford, H.C.; Obermeier, T.; von Spiczak, S.; Eichler, E.E.; Stephani, U.; Sander, T.; Helbig, I. Absence seizures with intellectual disability as a phenotype of the 15q13.3 microdeletion syndrome. *Epilepsia* **2011**, *52*, e194–e198. [CrossRef] [PubMed]
- 126. Melchior, L.; Bertelsen, B.; Debes, N.M.; Groth, C.; Skov, L.; Mikkelsen, J.D.; Brøndum-Nielsen, K.; Tümer, Z. Microduplication of 15q13.3 and Xq21.31 in a family with Tourette syndrome and comorbidities. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* 2013, 162B, 825–831. [CrossRef]
- 127. Hogart, A.; Leung, K.N.; Wang, N.J.; Wu, D.J.; Driscoll, J.; Vallero, R.O.; Schanen, N.C.; LaSalle, J.M. Chromosome 15q11-13 duplication syndrome brain reveals epiGenet.ic alterations in gene expression not predicted from copy number. *J. Med. Genet.* 2009, 46, 86–93. [CrossRef]
- Meguro-Horike, M.; Yasui, D.H.; Powell, W.; Schroeder, D.I.; Oshimura, M.; Lasalle, J.M.; Horike, S. Neuron-specific impairment of inter-chromosomal pairing and transcription in a novel model of human 15q-duplication syndrome. *Hum. Mol. Genet.* 2011, 20, 3798–3810. [CrossRef]
- 129. Good, K.V.; Vincent, J.B.; Ausió, J. MeCP2: The Genet.ic driver of rett syndrome epiGenet.ics. *Front. Genet.* **2021**, *12*, 620859. [CrossRef]
- 130. Nagarajan, R.P.; Hogart, A.R.; Gwye, Y.; Martin, M.R.; LaSalle, J.M. Reduced MeCP2 expression is frequent in autism frontal cortex and correlates with aberrant MECP2 promoter methylation. *EpiGenetics* **2006**, *1*, e1–e11. [CrossRef]
- 131. Rozycka, A.; Dorszewska, J.; Steinborn, B.; Kempisty, B.; Lianeri, M.; Wisniewska, K.; Jagodzinski, P.P. A transcript coding for a partially duplicated form of α7 nicotinic acetylcholine receptor is absent from the CD4+ T-lymphocytes of patients with autosomal dominant nocturnal frontal lobe epilepsy (ADNFLE). *Folia Neuropathol.* 2013, *51*, 65–75. [CrossRef]
- 132. Tracey, K.J. The inflammatory reflex. Nature 2002, 420, 853. [CrossRef] [PubMed]
- Reardon, C.; Murray, K.; Lomax, A.E. Neuroimmune communication in health and disease. *Physiol. Rev.* 2018, 98, 2287–2316. [CrossRef] [PubMed]
- 134. Borovikova, L.V.; Ivanova, S.; Zhang, M.; Yang, H.; Botchkina, G.I.; Watkins, L.R.; Wang, H.; Abumrad, N.; Eaton, J.W.; Tracey, K.J. Vagus nerve stimulation attenuates the systemic inflammatory response to endotoxin. *Nature* 2000, 405, 458–462. [CrossRef] [PubMed]
- 135. Eduardo, C.C.; Alejandra, T.G.; Guadalupe, D.K.J.; Herminia, V.G.; Lenin, P.; Enrique, B.V.; Evandro, B.M.; Oscar, B.; Iván, G.M. Modulation of the extraneuronal cholinergic system on main innate response leukocytes. *J. Neuroimmunol.* 2019, 327, 22–35. [CrossRef] [PubMed]
- 136. Singer, M.; Deutschman, C.S.; Seymour, C.W.; Shankar-Hari, M.; Annane, D.; Bauer, M.; Bellomo, R.; Bernard, G.R.; Chiche, J.D.; Coopersmith, C.M.; et al. The third international consensus definitions for sepsis and septic shock (Sepsis-3). *JAMA* 2016, 315, 801–810. [CrossRef]
- 137. van der Poll, T.; Shankar-Hari, M.; Wiersinga, W.J. The immunology of sepsis. Immunity 2021, 54, 2450–2464. [CrossRef]

- 138. Cedillo, J.L.; Arnalich, F.; Martín-Sánchez, C.; Quesada, A.; Rios, J.J.; Maldifassi, M.C.; Atienza, G.; Renart, J.; Fernández-Capitán, C.; García-Rio, F.; et al. Usefulness of α7 nicotinic receptor messenger RNA levels in peripheral blood mononuclear cells as a marker for cholinergic antiinflammatory pathway activity in septic patients: Results of a pilot study. *J. Infect. Dis.* 2015, 211, 146–155. [CrossRef]
- 139. Baird, A.; Coimbra, R.; Dang, X.; Eliceiri, B.P.; Costantini, T.W. Up-regulation of the human-specific CHRFAM7A gene in inflammatory bowel disease. *BBA Clin.* **2016**, *5*, 66–71. [CrossRef]
- 140. van Maanen, M.A.; Lebre, M.C.; van der Poll, T.; LaRosa, G.J.; Elbaum, D.; Vervoordeldonk, M.J.; Tak, P.P. Stimulation of nicotinic acetylcholine receptors attenuates collagen-induced arthritis in mice. *Arthritis Rheum.* **2009**, *60*, 114–122. [CrossRef]
- 141. Cao, X.; Wang, Y.; Gao, L. CHRFAM7A Overexpression attenuates cerebral ischemia-reperfusion injury via inhibiting microglia pyroptosis mediated by the NLRP3/Caspase-1 pathway. *Inflammation* **2021**, *44*, 1023–1034. [CrossRef] [PubMed]
- 142. Guo, Z.; Yu, S.; Chen, X.; Ye, R.; Zhu, W.; Liu, X. NLRP3 is involved in ischemia/reperfusion injury. CNS Neurol. Disord. Drug Targets 2016, 15, 699–712. [CrossRef] [PubMed]
- 143. Li, T.; Chen, W.; Zhang, Q.; Deng, C. Human-specific gene CHRFAM7A mediates M2 macrophage polarization via the Notch pathway to ameliorate hypertrophic scar formation. *Biomed. Pharmacother.* **2020**, *131*, 110611. [CrossRef] [PubMed]
- 144. Zhang, Y.; Qian, J.; Ren, H.; Meng, F.; Ma, R.; Xu, B. Human-specific CHRFAM7A protects against radiotherapy-induced lacrimal gland injury by inhibiting the p38/JNK signalling pathway and oxidative stress. *Int. J. Clin. Exp. Pathol.* 2017, 10, 9001–9011.
- 145. Mehta, P.; McAuley, D.F.; Brown, M.; Sanchez, E.; Tattersall, R.S.; Manson, J.J.; HLH Across Speciality Collaboration, UK. COVID-19: Consider cytokine storm syndromes and immunosuppression. *Lancet* **2020**, *395*, 1033–1034. [CrossRef]
- Changeux, J.P.; Amoura, Z.; Rey, F.A.; Miyara, M. A nicotinic hypothesis for COVID-19 with preventive and therapeutic implications. C. R. Biol. 2020, 343, 33–39. [CrossRef]
- 147. Farsalinos, K.; Angelopoulou, A.; Alexandris, N.; Poulas, K. COVID-19 and the nicotinic cholinergic system. *Eur. Respir. J.* **2020**, 56, 2001589. [CrossRef]
- 148. Gonzalez-Rubio, J.; Navarro-Lopez, C.; Lopez-Najera, E.; Lopez-Najera, A.; Jimenez-Diaz, L.; Navarro-Lopez, J.D.; Najera, A. Cytokine release syndrome (CRS) and nicotine in COVID-19 patients: Trying to calm the storm. *Front. Immunol.* 2020, 11, 1359. [CrossRef]
- 149. Bonaz, B.; Sinniger, V.; Pellissier, S. Targeting the cholinergic anti-inflammatory pathway with vagus nerve stimulation in patients with COVID-19? *Bioelectron. Med.* 2020, *6*, 15. [CrossRef]
- 150. Russo, P.; Bonassi, S.; Giacconi, R.; Malavolta, M.; Tomino, C.; Maggi, F. COVID-19 and smoking: Is nicotine the hidden link? *Eur. Respir. J.* **2020**, *55*, 2001116. [CrossRef]
- 151. Pohanka, M. Inhibitors of acetylcholinesterase and butyrylcholinesterase meet immunity. *Int. J. Mol. Sci.* **2014**, *15*, 9809–9825. [CrossRef]
- 152. Huang, W.; Kabbani, N.; Brannan, T.K.; Lin, M.K.; Theiss, M.M.; Hamilton, J.F.; Ecklund, J.M.; Conley, Y.P.; Vodovotz, Y.; Brienza, D.; et al. Association of a functional polymorphism in the CHRFAM7A gene with inflammatory response mediators and neuropathic pain after spinal cord injury. *J. Neurotrauma* 2019, *36*, 3026–3033. [CrossRef]
- 153. Kiguchi, N.; Sakaguchi, H.; Kadowaki, Y.; Saika, F.; Fukazawa, Y.; Matsuzaki, S.; Kishioka, S. Peripheral administration of interleukin-13 reverses inflammatory macrophage and tactile allodynia in mice with partial sciatic nerve ligation. *J. Pharmacol. Sci.* 2017, 133, 53–56. [CrossRef]
- 154. Hone, A.J.; McIntosh, J.M. Nicotinic acetylcholine receptors in neuropathic and inflammatory pain. *FEBS Lett.* **2018**, *592*, 1045–1062. [CrossRef]
- 155. Chan, T.W.; Langness, S.; Cohen, O.; Eliceiri, B.P.; Baird, A.; Costantini, T.W. CHRFAM7A reduces monocyte/macrophage migration and colony formation in vitro. *Inflamm. Res.* 2020, *69*, 631–633. [CrossRef]
- Kim, Y.H.; Lee, J.R.; Hahn, M.J. Regulation of inflammatory gene expression in macrophages by epithelial-stromal interaction 1 (Epsti1). *Biochem. Biophys. Res. Commun.* 2018, 496, 778–783. [CrossRef]
- 157. York, M.R.; Nagai, T.; Mangini, A.J.; Lemaire, R.; van Seventer, J.M.; Lafyatis, R. A macrophage marker, Siglec-1, is increased on circulating monocytes in patients with systemic sclerosis and induced by type I interferons and toll-like receptor agonists. *Arthritis Rheum.* **2007**, *56*, 1010–1020. [CrossRef]
- 158. Kabbani, N.; Nichols, R.A. Beyond the channel: Metabotropic signaling by nicotinic receptors. *Trends. Pharmacol. Sci.* **2018**, *39*, 354–366. [CrossRef]
- 159. Fratiglioni, L.; Wang, H.X. Smoking and Parkinson's and Alzheimer's disease: Review of the epidemiological studies. *Behav. Brain Res.* **2000**, *113*, 117–120. [CrossRef]
- 160. Schliebs, R.; Arendt, T. The cholinergic system in aging and neuronal degeneration. *Behav. Brain Res.* **2011**, 221, 555–563. [CrossRef]
- Park, H.J.; Lee, P.H.; Ahn, Y.W.; Choi, Y.J.; Lee, G.; Lee, D.Y.; Chung, E.S.; Jin, B.K. Neuroprotective effect of nicotine on dopaminergic neurons by anti-inflammatory action. *Eur. J. Neurosci.* 2007, *26*, 79–89. [CrossRef] [PubMed]
- 162. Liu, Y.; Zeng, X.; Hui, Y.; Zhu, C.; Wu, J.; Taylor, D.H.; Ji, J.; Fan, W.; Huang, Z.; Hu, J. Activation of α7 nicotinic acetylcholine receptors protects astrocytes against oxidative stress-induced apoptosis: Implications for Parkinson's disease. *Neuropharmacology* 2015, 91, 87–96. [CrossRef] [PubMed]
- Egea, J.; Buendia, I.; Parada, E.; Navarro, E.; León, R.; Lopez, M.G. Anti-inflammatory role of microglial alpha7 nAChRs and its role in neuroprotection. *Biochem. Pharmacol.* 2015, 97, 463–472. [CrossRef] [PubMed]

- King, J.R.; Gillevet, T.C.; Kabbani, N. A G protein-coupled alpha7 nicotinic receptor regulates signaling and TNF-alpha release in microglia. FEBS Open Biol. 2017, 7, 1350–1361. [CrossRef]
- De Simone, R.; Ajmone-Cat, M.A.; Carnevale, D.; Minghetti, L. Activation of alpha7 nicotinic acetylcholine receptor by nicotine selectively up-regulates cyclooxygenase-2 and prostaglandin E2 in rat microglial cultures. J. Neuroinflamm. 2005, 2, 4. [CrossRef]
- 166. Pavlov, V.A.; Parrish, W.R.; Rosas-Ballina, M.; Ochani, M.; Puerta, M.; Ochani, K.; Chavan, S.; Al-Abed, Y.; Tracey, K.J. Brain acetylcholinesterase activity controls systemic cytokine levels through the cholinergic anti-inflammatory pathway. *Brain Behav. Immun.* 2009, 23, 41–45. [CrossRef]
- Ballinger, E.C.; Ananth, M.; Talmage, D.A.; Role, L.W. Basal forebrain cholinergic circuits and signaling in cognition and cognitive decline. *Neuron* 2016, 91, 1199–1218. [CrossRef]
- 168. Heneka, M.T.; Kummer, M.P.; Latz, E. Innate immune activation in neurodegenerative disease. *Nat. Rev. Immunol.* 2014, 14, 463–477. [CrossRef]
- Kalkman, H.O.; Feuerbach, D. Modulatory effects of α7 nAChRs on the immune system and its relevance for CNS disorders. *Cell Mol. Life Sci.* 2016, 73, 2511–2530. [CrossRef]
- 170. Taylor, P. Development of acetylcholinesterase inhibitors in the therapy of Alzheimer's disease. *Neurology* **1998**, *51*, S30–S35, discussion S65–S67. [CrossRef]
- 171. Neri, M.; Bonassi, S.; Russo, P. Genet.ic variations in CHRNA7 or CHRFAM7 and susceptibility to dementia. *Curr. Drug Targets* **2012**, *13*, 636–643. [CrossRef] [PubMed]
- 172. Carson, R.; Craig, D.; McGuinness, B.; Johnston, J.A.; O'Neill, F.A.; Passmore, A.P.; Ritchie, C.W. Alpha7 nicotinic acetylcholine receptor gene and reduced risk of Alzheimer's disease. J. Med. Genet. 2008, 45, 244–248. [CrossRef] [PubMed]
- 173. Dziewczapolski, G.; Glogowski, C.M.; Masliah, E.; Heinemann, S.F. Deletion of the alpha 7 nicotinic acetylcholine receptor gene improves cognitive deficits and synaptic pathology in a mouse model of Alzheimer's disease. J. Neurosci. 2009, 29, 8805–8815. [CrossRef] [PubMed]
- 174. Quik, M.; Bordia, T.; Zhang, D.; Perez, X.A. Nicotine and nicotinic receptor drugs: Potential for Parkinson's disease and drug-induced movement disorders. *Int. Rev. Neurobiol.* **2015**, *124*, 247–271. [CrossRef]
- 175. Oddo, S.; Caccamo, A.; Green, K.N.; Liang, K.; Tran, L.; Chen, Y.; Leslie, F.M.; LaFerla, F.M. Chronic nicotine administration exacerbates tau pathology in a transgenic model of Alzheimer's disease. *Proc. Natl. Acad. Sci. USA* 2005, 102, 3046–3051. [CrossRef] [PubMed]
- 176. Puzzo, D.; Privitera, L.; Leznik, E.; Fà, M.; Staniszewski, A.; Palmeri, A.; Arancio, O. Picomolar amyloid-beta positively modulates synaptic plasticity and memory in hippocampus. *J. Neurosci.* **2008**, *28*, 14537–14545. [CrossRef] [PubMed]
- 177. Oz, M.; Lorke, D.E.; Yang, K.H.; Petroianu, G. On the interaction of β-amyloid peptides and α7-nicotinic acetylcholine receptors in Alzheimer's disease. *Curr. Alzheimer Res.* **2013**, *10*, 618–630. [CrossRef]
- 178. Lasala, M.; Fabiani, C.; Corradi, J.; Antollini, S.; Bouzat, C. Molecular modulation of human α7 Nicotinic receptor by amyloid-β peptides. *Front. Cell Neurosci.* **2019**, *13*, 37. [CrossRef]
- 179. Wang, H.Y.; Lee, D.H.; Davis, C.B.; Shank, R.P. Amyloid peptide Abeta(1-42) binds selectively and with picomolar affinity to alpha7 nicotinic acetylcholine receptors. *J. Neurochem.* 2000, 75, 1155–1161. [CrossRef]
- Dineley, K.T.; Westerman, M.; Bui, D.; Bell, K.; Ashe, K.H.; Sweatt, J.D. Beta-amyloid activates the mitogen-activated protein kinase cascade via hippocampal alpha7 nicotinic acetylcholine receptors: In vitro and in vivo mechanisms related to Alzheimer's disease. J. Neurosci. 2001, 21, 4125–4133. [CrossRef]
- Parri, H.R.; Hernandez, C.M.; Dineley, K.T. Research update: Alpha7 nicotinic acetylcholine receptor mechanisms in Alzheimer's disease. *Biochem. Pharmacol.* 2011, 82, 931–942. [CrossRef] [PubMed]
- 182. Nagele, R.G.; D'Andrea, M.R.; Anderson, W.J.; Wang, H.Y. Intracellular accumulation of beta-amyloid(1-42) in neurons is facilitated by the alpha 7 nicotinic acetylcholine receptor in Alzheimer's disease. *Neuroscience* **2002**, *110*, 199–211. [CrossRef]
- Wang, H.Y.; Li, W.; Benedetti, N.J.; Lee, D.H. Alpha 7 nicotinic acetylcholine receptors mediate beta-amyloid peptide-induced tau protein phosphorylation. J. Biol. Chem. 2003, 278, 31547–31553. [CrossRef] [PubMed]
- 184. Swaminathan, S.; Shen, L.; Kim, S.; Inlow, M.; West, J.D.; Faber, K.M.; Foroud, T.; Mayeux, R.; Saykin, A.J.; Initiative, A.S.D.N.; et al. Analysis of copy number variation in Alzheimer's disease: The NIALOAD/ NCRAD family study. *Curr. Alzheimer Res.* 2012, 9, 801–814. [CrossRef]
- 185. Swaminathan, S.; Huentelman, M.J.; Corneveaux, J.J.; Myers, A.J.; Faber, K.M.; Foroud, T.; Mayeux, R.; Shen, L.; Kim, S.; Turk, M.; et al. Analysis of copy number variation in Alzheimer's disease in a cohort of clinically characterized and neuropathologically verified individuals. *PLoS ONE* **2012**, *7*, e50640. [CrossRef]
- 186. Swaminathan, S.; Kim, S.; Shen, L.; Risacher, S.L.; Foroud, T.; Pankratz, N.; Potkin, S.G.; Huentelman, M.J.; Craig, D.W.; Weiner, M.W.; et al. Genomic copy number analysis in alzheimer's disease and mild cognitive impairment: An ADNI study. *Int. J. Alzheimers Dis.* 2011, 2011, 729478. [CrossRef]
- 187. Szigeti, K.; Kellermayer, B.; Lentini, J.M.; Trummer, B.; Lal, D.; Doody, R.S.; Yan, L.; Liu, S.; Ma, C.; Consortium, T.A.R.a.C. Ordered subset analysis of copy number variation association with age at onset of Alzheimer's disease. *J. Alzheimer's Dis.* 2014, 41, 1063–1071. [CrossRef]
- 188. Liou, Y.J.; Lai, I.C.; Hong, C.J.; Liu, H.C.; Liu, T.Y.; Tsai, S.J. Association analysis of the partially duplicated alpha7 nicotinic acetylcholine receptor Genet.ic variant and Alzheimer's disease. *Dement. Geriatr. Cogn. Disord.* 2001, 12, 301–304. [CrossRef]

- Fehér, A.; Juhász, A.; Rimanóczy, A.; Csibri, E.; Kálmán, J.; Janka, Z. Association between a Genet.ic variant of the alpha-7 nicotinic acetylcholine receptor subunit and four types of dementia. *Dement. Geriatr. Cogn. Disord.* 2009, 28, 56–62. [CrossRef]
   Laberta, G. Theorie and A. Starrawa, and the starrawa and the st
- 190. Iadecola, C. The pathobiology of vascular dementia. Neuron 2013, 80, 844-866. [CrossRef]
- Thomsen, M.S.; Hansen, H.H.; Timmerman, D.B.; Mikkelsen, J.D. Cognitive improvement by activation of alpha7 nicotinic acetylcholine receptors: From animal models to human pathophysiology. *Curr. Pharm. Des.* 2010, *16*, 323–343. [CrossRef] [PubMed]
- 192. Ihnatovych, I.; Nayak, T.K.; Ouf, A.; Sule, N.; Birkaya, B.; Chaves, L.; Auerbach, A.; Szigeti, K. iPSC model of CHRFAM7A effect on α7 nicotinic acetylcholine receptor function in the human context. *Transl. Psychiatry* **2019**, *9*, 59. [CrossRef] [PubMed]
- 193. Ihnatovych, I.; Birkaya, B.; Notari, E.; Szigeti, K. iPSC-derived microglia for modeling human-specific DAMP and PAMP responses in the context of alzheimer's disease. *Int. J. Mol. Sci.* **2020**, *21*, 9668. [CrossRef] [PubMed]
- 194. Ballester, L.Y.; Capó-Vélez, C.M.; García-Beltrán, W.F.; Ramos, F.M.; Vázquez-Rosa, E.; Ríos, R.; Mercado, J.R.; Meléndez, R.I.; Lasalde-Dominicci, J.A. Up-regulation of the neuronal nicotinic receptor α7 by HIV glycoprotein 120: Potential implications for HIV-associated neurocognitive disorder. J. Biol. Chem. 2012, 287, 3079–3086. [CrossRef]
- 195. Cedillo, J.L.; Bordas, A.; Arnalich, F.; Esteban-Rodríguez, I.; Martín-Sánchez, C.; Extremera, M.; Atienza, G.; Rios, J.J.; Arribas, R.L.; Montiel, C. Anti-tumoral activity of the human-specific duplicated form of α7-nicotinic receptor subunit in tobacco-induced lung cancer progression. *Lung Cancer* 2019, 128, 134–144. [CrossRef]