Oncosuppressive functions of decorin

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Abbreviations: AMPK α , AMP-activated protein kinase, α ; AP4, activating enhancer binding protein 4; ATG, autophagy-related gene; Bcl2, B-cell CLL/lymphoma 2; BRAF, B-Raf proto-oncogene; CLEAR, coordinated lysosomal expression and regulation; CXCL12, C-X-C motif chemokine 12; CXCR4, C-X-C chemokine receptor type 4; DYRK1, dual-specificity tyrosine-(Y)phosphorylation regulated kinase 1A; ECM, extracellular matrix; EGFR, epidermal growth factor receptor; ERK, extracellular regulated kinase; GSK-3 β , glycogen synthase kinase 3 β ; HIF-1 α , hypoxia inducible factor-1 α ; HGF, hepatocyte growth factor; IGF-1, insulin-like growth factor 1; IGF-IR, insulin-like growth factor 1 receptor; IgG, immunoglobulin G-like folds; IR-A, insulin receptor isoform A; IRS, insulin receptor substrate 1; LC3, microtubule-associated protein 1A/1B-light chain 3; LYNUS, lysosomal nutrient sensing; MAPK, mitogen activated protein kinase; MMP, matrix metalloproteinase; mTOR, mechanistic target of rapamycin; PDGFR, platelet derived growth factor receptor; Peg3, paternally expressed gene 3; PINK1, PTEN-induced putative kinase-1; PI3K, phosphoinositide 3 kinase; PKB/Akt, protein kinase B; PKC, protein kinase C; PGC-1a, peroxisome proliferator activated receptor γ co-activator-1 α ; Rac1, ras-related C3 botulinum toxin substrate 1; Rheb, Ras homolog enriched in brain; RhoA, ras homolog gene family, member A; ROCK1, rho-associated, coiled-coil-containing protein kinase 1; RRM, RNA recognition motif; RTK, receptor tyrosine kinase; Peg3, paternally expressed gene 3; p70S6K, ribosomal protein S6 kinase, 70kDa; SLRP, small leucine-rich proteoglycan; TIMP3, tissue inhibitor of metalloproteinases 3; TGF-β1, transforming growth factor β 1; TFEB, transcription factor EB; TSP1, thrombospondin 1; ULK1, unc-51 like autophagy activating kinase 1; VDAC, voltage-dependent anion channel; VEGF, vascular endothelial growth factor receptor; Vps34, vacuolar protein sorting 34.

The extracellular matrix is rapidly emerging as a prominent contributor to various fundamental processes of tumorigenesis. In particular, decorin, a member of the small leucine-rich proteoglycan gene family, is assuming a central role as a potent soluble tumor repressor. Decorin binds and antagonizes various receptor tyrosine kinases and inhibits downstream oncogenic signaling in several solid tumors. Among other functions, decorin evokes cell cycle arrest, apoptosis, and antimetastatic, and antiangiogenic programs. Recent work has revealed a paradigmatic shift in our understanding of the molecular mechanisms underlying its tumoricidal properties. Decorin adversely compromises the genetic signature of the tumor microenvironment and induces endothelial cell autophagy downstream of VEGFR2. Moreover, decorin selectively evokes destruction of tumor cell mitochondria downstream of Met through mitophagy. Acting as a partial agonist, decorin signals via proautophagic receptors and triggers procatabolic processes that parallel the classical tumoricidal properties of this multifaceted proteoglycan.

Introduction

Solid malignancies are complex entities that arise from intricate associations among a heterogeneous population of cells derived from several epigenetically and transcriptionally distinct lineages.^{1,2} The impressive assortment of recruited mesenchymal and inflammatory cells within the elaborate network of the extracellular matrix (ECM) is emerging as a critical entity defining chemotherapeutic responsiveness and clinical outcomes.³ The ECM acts as a bidirectional signaling hub, linking the local microenvironment with the tumor cells.⁴ This communicative epicenter provides instructional cues in the form of solid-phase ligands⁵ and/or soluble signals that are capable of modulating multiple aspects of tumorigenesis and angiogenesis.⁶⁻⁹

The diverse regulatory properties exerted by the multifunctional nature of ECM molecules are embodied and exemplified by the small leucine-rich proteoglycan (SLRP) gene family.^{10,11} Decorin, the prototypical SLRP of this 18-member strong clan, is composed of a singular N-terminal glycosaminoglycan chain of dermatan or chondroitin sulfate, 12 leucine-rich tandem repeats, and a C-terminal Ear domain.¹² Decorin was named for its function as an avid collagen-binding partner for fibrillogenesis,^{6,13} and regulates various biomechanical properties of collagen-containing tissue, including tendons and skin.¹⁴⁻¹⁷ Subsequent paradigm-shifting work demonstrated a strong affinity of decorin for various receptor tyrosine kinases (RTKs) that resulted in potent and sustained oncostasis and angiostasis. Moreover, decorin binds and sequesters numerous growth factors,¹⁸ multiple matrix constituents,¹⁹ and indirectly suppresses downstream signaling.¹⁹ Collectively, these studies revealed that

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decorin functions as a soluble tumor repressor that counteracts tumorigenic and angiogenic growth, and the protein has aptly been designated "a guardian from the matrix."¹⁹

Decorin is currently emerging as a multifaceted and multifunctional signaling molecule with roles beyond the tumor stroma. Pertinent examples include inflammatory responses,^{8,20,21} delayed hypersensitivity,²² wound healing,^{23,24} keratinocyte function,²⁵ hepatic healing,²⁶ asthma,²⁷ diabetic nephropathies,²⁸ myogenesis,²⁹ shaping hematopoietic stem cell niches,³⁰ convergent extension,³¹ and renal diseases.^{32,33} We are currently in the midst of a SLRP renaissance in which decorin is challenging established cancer biology precepts for tumorigenic and angiogenic suppression by matrix constituents.³⁴

In this review, we will evaluate the consequences of autophagic and mitophagic processes that occur downstream of proautophagic RTKs and impinge upon the tumor microenvironment and the tumor proper. Importantly, autophagic and mitophagic processes evoked by decorin are above the operative threshold for ambient homeostatic function, thus controlled or limited autophagy may result in revitalization of cellular processes. The balance between excessive and insufficient autophagy is crucial as either may result in a pathological state. We will critically assess these novel avenues and their unique interfaces with the well-established tumoricidal properties of this versatile proteoglycan and discuss potential therapeutic interventions for decorin bioactivity.

Localization of Decorin Within the Tumor and the Tumor Stroma

Understanding the biological effects of decorin on the tumor first requires a discussion of its expression patterns and localization within the tumor compartments. The degree of decorin expression in various types and grades of tumors has recently been reported and reveals several apparent discrepancies. Clinically, loss of decorin within the tumor microenvironment serves as a poor prognosticator of invasive breast cancer.^{12,35} Moreover, by querving the Human Protein Atlas, Bozoky et al.³⁶ demonstrated a marked reduction in decorin levels within the stroma of many solid tumors, including bladder, breast, cervical, colon, kidney, ovary, pancreas, prostate, rectal, skin, stomach, and testis. Additionally, decorin expression is significantly reduced in the stroma of low- and high-grade bladder carcinomas, but is high in the submucosa and deep tumor stroma.³⁷ Decorin expression is also decreased in multiple myeloma and monoclonal gammopathy of undetermined significance.^{38,39}

However, other studies report an increase in the amount of stromal decorin in cancer, primarily within colon⁴⁰⁻⁴² and breast^{43,44} carcinomas. Given the breadth of the decorin interactome with multiple matrix constituents,¹⁹ a role for decorin in orchestrating higher-order matrix assemblies and coordinating a desmoplastic reaction emerges. Formation of these collagen-rich structures (comprised of collagen II and IV microfibrils) and the propensity for sequestering potent antitumorigenic (e.g., decorin) and antiangiogenic factors (e.g., decorin and matrilin-1⁴⁵) into large complexes favor tumor suppression.^{7,19} We believe that

profuse amounts of decorin within the stromal compartments of these tumor types would negatively regulate the juxtapositioned RTKs expressed by the growing neoplasm in a paracrine fashion.

Moreover, and pertinent for tumor parenchyma, multiple studies demonstrate a complete loss of decorin expression by the tumor cells.^{19,46} Deficiencies of decorin have been found in multiple tumors including prostate carcinomas,⁴⁷ urothelial malignancies,⁴⁸ and hepatic carcinomas.⁴⁹ Interestingly, an unbiased deep transcriptome sequencing approach revealed that decorin expression is markedly decreased in hepatocellular carcinomas.⁵⁰ In the case of urothelial carcinomas, decorin mRNA expression is highly reduced in superficial⁵¹ and infiltrating tumors.⁵² Certain neoplasms have an avid proclivity for hypermethylation of the decorin promoter, thereby effectively silencing expression for unchecked tumor progression.¹⁹

Further evidence for the oncostatic role of decorin as a tumor repressor stems from strong genetic models in which decorin was unconditionally ablated.⁵³ Under conditions of a high-fat (i.e., Western) diet, decorin (*Dcn*)-null mice develop spontaneous intestinal tumors.⁵⁴ Moreover, compound knockout mice for *Dcn* and *Tp53* accede to aggressive lymphomas,⁵⁵ whereas reintroduction of decorin via adenoviral delivery or systemic administration significantly counters tumorigenic and angiogenic growth in a variety of solid tumors.^{46,56–60} Collectively, these data provide firm support for a tumor repressive role of decorin in a physiologically relevant setting.

Decorin Promotes a Proautophagic Signaling Program in the Tumor Microenvironment

Intravital imaging of exogenously delivered near infraredlabeled decorin via tail vein injection demonstrated avid and exclusive targeting of orthotopic tumor xenografts.^{61,62} Importantly, decorin is not targeted to or retained by any other organ system and is subsequently secreted via the urine.⁶¹ Systemic delivery of decorin after establishment of triple-negative breast carcinoma orthotopic xenografts permitted high-resolution and simultaneous transcriptomic profiling of the host stromal compartment of mouse origin and the tumor parenchyma of human origin.⁵⁹ Unexpectedly, decorin evoked significant transcriptomic changes within the host-provided tumor microenvironment without significantly modulating the mRNA profile of the human breast carcinoma⁶³ Collectively, these changes reprogrammed the tumor stroma in a manner that disfavored tumorigenic growth and metastases.^{19,59}

Of the multitude of genes that are differentially expressed upon chronic decorin treatment, a small subset of targets have emerged that include a poorly-studied imprinted tumor suppressor known as Peg3.⁶³⁻⁶⁵ The induction of a tumor suppressor gene is clearly congruent with the antitumorigenic activity of decorin¹⁹ Furthermore, *PEG3* is epigenetically silenced (via promoter hypermethylation of the active allele) in multiple gynecologic and neural tumors.^{66,67} Our interest in pursuing Peg3 stemmed from its previously described role in the suppression of Wnt/ β -catenin signaling in a non-canonical manner,⁶⁸ which mirrored the bioactivity of decorin in a cervical carcinoma model.⁶¹ Our interest intensified when we found that Peg3 colocalized with subcellular structures highly reminiscent of autophagosomes upon decorin stimulation of macrovascular and microvascular endothelial cells used as a surrogate for the tumor stroma.⁶⁹ Further investigation using the specific autophagic markers Beclin 1 and LC, confirmed that these Peg3-positive structures were autophagosomes (**Fig. 1A**, **B**).⁷⁰ Intriguingly, Peg3 is required for the decorin-induced transcriptional activation and accumulation of Beclin 1 and LC3; moreover, Peg3 is required for maintenance of basal Beclin 1 levels in endothelial cells.^{70,71}

Mechanistically, decorin induces Peg3-dependent autophagy downstream of VEGFR2,⁷¹ the primary RTK for endothelial cell homeostasis. In contrast to the aforementioned function of decorin as a pan-RTK inhibitor, decorin acts as a partial VEGFR2 agonist for autophagic initiation (Fig. 1A).⁷¹ Decorin binds the VEGFR2 ectodomain (IgG domains 3–5) that partially overlaps with the binding site of VEGFA (IgG domains 1–3). By doing so, it activates the proautophagic AMPK α /Vps34 signaling arm^{72,73} and concurrently represses the antiautophagic PI3K/ Akt/mTOR/p70S6K pathway (Fig. 1A).^{74,75} Collectively, the net cellular output of these concerted signaling events promotes the formation of a Peg3–Beclin 1–LC3 ternary complex, induction of proautophagic gene targets, and concomitant disruption of the inhibitory Bcl-2–Beclin 1 complex.^{75,76} Decorin rapidly promotes activation of the catalytic core of the central energy sensor, AMPK α , at Thr172 in a VEGFR2-dependent manner within a nutrient-rich setting.⁷⁵

In theory, it is possible that decorin mediates activation of AMPK α by recruiting ULK1 (also known as ATG1) and promoting the formation of AMPK α –ULK1 heterodimers for the initiation of autophagy,^{77,78} with further and protracted antagonism of mTORC1 components (such as Raptor, Rheb, and G β L).⁷⁷ Notably, recent studies have elaborated an inhibitory function for EGFR and Akt signaling through phosphorylation and consequent inactivation of Beclin 1 (BSCN1) that is conducive to autophagic suppression and chemoresistance.^{73,79} As many RTKs share the core signaling machinery, it is possible that decorin abrogates phosphorylated Beclin 1 downstream of VEGFR2, thereby permitting autophagic activation and downstream supramolecular complex assembly.⁷⁵

Successful autophagy relies on positive flux, lysosomal fusion, and the successful formation of autophagolysosomes.^{80,81} Decorin may positively regulate transcription factor EB (TFEB), a crucial sensory node between autophagy and lysosomal formation.⁸² Intriguingly, TFEB is held inactive and sequestered

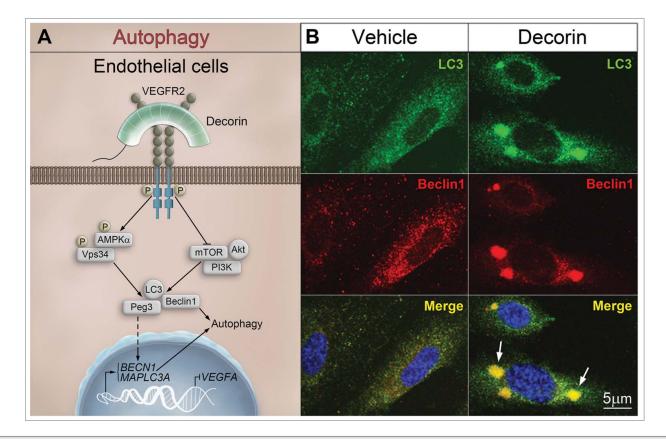


Figure 1. (**A**) Schematic representation of the downstream signaling events following binding of decorin to VEGFR2 and Peg-3–dependent endothelial cell autophagy. Please refer to the text for details. (**B**) Confocal laser microscopy of human umbilical vein endothelial cells (HUVECs) after treatment for 6 hours with vehicle or 200 nm human recombinant decorin. Please note the formation of autophagosomes, as identified by immunostaining for LC3 (green) or beclin 1 (red). The bottom row of panels demonstrates colocalization of autophagic markers (yellow), thereby confirming the identify of these subcellular structures. Nuclei are stained blue with DAPI. All images were taken with the same exposure, gain, and intensity. Scale bar = 5 μ m.

within the cytosol (at the lysosomal membrane) by inhibitory mTOR/LYNUS signaling.^{83,84} As decorin inactivates mTOR downstream of VEGFR2, in the presence of decorin TFEB may become dephosphorylated (either passively or actively) and translocate into the nucleoplasm for the activation of proautophagic targets (e.g., BECN1 and CLEAR network genes such as PPARGC1A),⁸² potentially in a Peg3-dependent manner.

One of the key tenets of decorin bioactivity is the suppression of rampant neovascularization.⁸⁵ Notably, biglycan, the closest member of the SLRP gene family, has opposite effects to decorin in both inflammation⁸⁶⁻⁸⁸ and angiogenesis,^{89,90} demonstrating specific bioactivities for individual SLRPs. As endothelial cells play a critical role in vascularization of an oxygen- and nutrientstarved solid tumor, we propose that increased autophagy may represent a novel mechanism by which decorin triggers a halt in migration, proliferation, and capillary morphogenesis,⁶⁹ and, ultimately, in angiogenesis—all key properties orchestrated by positive VEGFA signaling. This may be achieved by the concerted activation of VEGFR2/ULK1/AMPK α /Peg3/TFEB signaling for the repression of endothelial cell-derived VEGFA and/ or by rendering endothelial cells refractory to the aberrant angiogenic stimuli derived from the growing tumor cells.

In conclusion, endothelial cell autophagy heralds a paradigmatic shift in the role of decorin in tumorigenic and angiogenic suppression via RTKs. Indeed, autophagic induction in a tissuespecific manner may be a conserved mechanism of action for matrix-derived signals.^{34,91} Furthermore, these findings alter the established model of nutrient-dependent autophagic induction in favor of soluble signaling cues, present within the stroma, for the activation of the autophagic machinery.

Growth Suppression Via Pan-Rtk Inhibition in the Tumor

In the classic model, decorin exerts its ascribed tumoricidal properties by directly engaging a multitude of RTKs within the target-rich environment that constitutes the tumor parenchyma.^{19,92-95} Mechanistically, monomeric decorin⁹⁶ binds RTKs with high affinity and evokes receptor dimerization, transient autophosphorylation, caveolin-1-mediated internalization,⁹⁷ and eventual lysosomal degradation.^{19,97} Solid tumors that are dependent upon RTK signaling are severely suppressed following the introduction of decorin.^{35,46,56} In contrast to the response to naturally occurring receptor agonists, upon decorin exposure the unstructured intracellular tail of EGFR and Met acquires a unique phosphorylation signature,¹⁹ perhaps as a result of altered conformational states of the receptor ectodomain and transmembrane region⁹⁸ upon decorin binding. In contrast with active signaling agents, this phosphorylation pattern instructs cell cycle arrest, apoptosis, angiostasis, and protracted oncogene suppression (Fig. 2, left panel).¹⁹

A prime example is EGFR, in which decorin initiates a rapid phosphorylation that activates the MAPK signaling system.⁵⁸ This counterintuitively results in apoptosis and cell cycle arrest concomitant with the cleavage and subsequent activation of

caspase-3 and induction of p21^{WAF1}(p21), (Fig. 2, left panel), despite the fact that total cell surface EGFR levels are diminished by 50%.⁵⁸ A second example is the recently discovered Met receptor, which undergoes a strong Tyr phosphorylation signal on a phosphotyrosine array following decorin treatment;⁹⁹ this transient activation results in recruitment of c-Cbl and receptor downregulation. The transient nature of decorin-evoked receptor phosphorylation and the downstream transduction mechanisms represents a key tenet in cell signaling in which the duration, frequency, and strength of the signal combinatorially dictate cellular behaviors.¹⁰⁰ The oscillatory nature of the downstream signaling molecules (receptors, MAKP, PI3K/Akt/mTOR) framed within this conceptual scaffold may prove crucial for decorin-transduced signals and biological outcomes.

EGFR and Met are not the only RTKs responsible for decorin bioactivities. Several members of the EGFR family, such as the ErbB2/ErbB4 heterodimers, are also targeted by decorin⁶, although recent evidence now suggests direct ErbB4 antagonism.¹⁰¹ Many other RTKs have been identified, including IGF-IR, IR-A, and their ligands,^{62,102,103} PDGFR α^{49} and associated PDGFA ligand,¹⁰⁴ and VEGFR2.^{69,105,106} Notably, IGF-IR represents the only known exception where the receptor is not internalized and tagged for destruction by decorin binding;⁶² instead, decorin suppresses the IRS-1/Akt/ERK/p70S6K pathway, blocks migration, and prevents IGF-I–dependent localization of IGF-IR into caveosomes.³⁷

Suppression of Proliferative, Survival, and Migratory Signaling Pathways

Downstream of the robust binding events and receptor internalization and degradation, soluble decorin evokes potent and prolonged attenuation of several signaling pathways responsible for tumor cell proliferation, survival, and angiogenesis. Attenuation of Met results in the non-canonical and selective degradation of β -catenin and Myc^{61,85} with concurrent induction of p21.⁶¹ In the case of hepatocyte growth factor (HGF)/Met, this particular signaling system results in the direct stabilization and nuclear accumulation of B-catenin and transcriptional activation of β-catenin targets. Seemingly, this pathway functions independently of Wnt signaling via direct phosphorylation and inhibition of GSK-3B.¹⁰⁷ In contrast, binding of decorin to Met disrupts this signaling cascade⁶¹ in what appears to be a GSK-3β-independent manner, resulting in prompt 26S-proteasomal degradation and suppression of β -catenin (CTNNB1) expression (Fig. 2, left panel). Furthermore, Myc is also destabilized by increased phosphorylation on Thr58, a known phospho-acceptor site that designates Myc for degradation via the proteasome with concurrent suppression of MYC mRNA⁶¹ (Fig. 2, left panel). The increase in phosphorylated Myc at this position may be a result of derepressed GSK-3B downstream of attenuated Met signaling.⁶¹ However, the nuclear-localized priming kinase DYRK1 that is adept for phospho-transfer at Ser62 of Myc might work in concert with the GSK-3β-mediated phosphorylation of Myc at Thr58¹⁰⁸ that occurs downstream of decorin/Met binding.

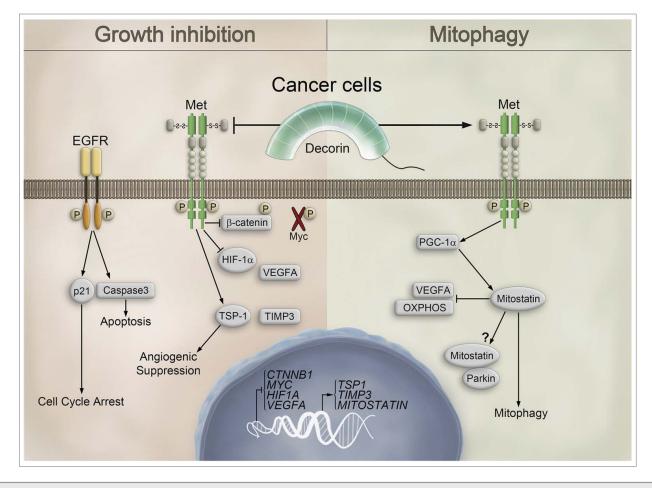


Figure 2. Schematic representations delineating the classic growth inhibitory functions (left panel) and novel promitophagic activities (right panel) of decorin in a tumor cell. Please refer to the text for a full mechanistic description.

Interestingly, decorin evokes strong nuclear translocation and subsequent degradation of Myc, concomitant with p21 accumulation.⁶¹ Additionally, transcriptional induction of p21 (also known as CDKN1A) may be linked to inactivation and destruction of the Met/ β -catenin/Myc signaling axis as AP4, a gene target of Myc,¹⁰⁹ actively represses *CDKN1A* expression (Fig 2, left panel).

Broad clinical implications of decorin-mediated suppression of the Met/ β -catenin axis have recently emerged from highthroughput transcriptomic screening.¹¹⁰ Basal mammary carcinomas driven by Wnt/ β -catenin and HGF/Met signaling have a unique genetic fingerprint known as the "Wnt-Met" signature, in which aberrant β -catenin activity drives self-renewal programs and Met suppresses differentiation commitments.¹¹⁰ It is plausible that clinical cases expressing the Wnt-Met signature would greatly benefit from receiving decorin, or related SLRPs, as an adjuvant protein-based therapy. The expected results of such treatment would include a reduction in the capacity for tumor self-renewal and the induction of a more differentiated tumor phenotype with decreased metastatic capacity. Personalized genomics combined with an understanding of matrix-derived tumor repressors may represent an important therapeutic option in the future. Indeed, molecular therapies targeting this system greatly alleviate tumor burden and increase overall survival. Moreover, a similar genetic signature was found for synergistic cooperativity between Myc and Her2/Neu (ErbB2) for stem-like breast cancer cell phenotypes without the requisite epithelial-to-mesenchymal transition discussed elsewhere.^{3,111,112} Therefore, decorin may suppress stem-like progenitors that would otherwise permit enhanced malignant states.

Intriguingly, as part of the newly described "Wnt-Met" signature, Wnt/ β -catenin signaling actively drives expression of CXCL12, a critical chemokine for tumor migration and metastasis.¹¹⁰ Moreover, HGF acting via the Akt/Rac1/PKC ζ arm evokes CXCR4 expression in breast carcinoma.¹¹³ Based on these findings, perturbation of the HGF/Met and Wnt/ β -catenin axis by decorin may significantly nullify the CXCR4/CXCL12 chemotactic system and thereby provide a molecular basis for the observed antimetastatic role of this multifunctional proteoglycan. Alternatively, the antimetastatic properties of decorin may be linked to the tumor suppressive function of MMP8 via antagonism of miR-21 and induction of MMP8.¹¹⁴

Suppression of Angiogenic Signaling Pathways

The cellular and molecular mechanisms responsible for governing and orchestrating tumor neovascularization are becoming more clearly defined. Moreover, several endogenous cues, chiefly soluble and matrix-derived in nature,¹¹⁵⁻¹¹⁸ that potently impede rampant tumor angiogenesis are being discovered. The literature surrounding the role of decorin in mediating angiogenic responses reflects the inherent intricacies of this vital developmental process. Perhaps the most striking example of complexity in decorin-mediated angiogenesis stems from studies on the normally developing cornea, in where dichotomous roles have been found;^{119,120} however, literature on the role of decorin in mediating tumor angiogenesis favors an antiangio-genic role.^{12,19,106,121} Furthermore, the observation that angiosarcomas exhibit a total lack of stromal decorin whereas hemangiomas have abundant decorin expression¹²² implies an inverse relationship between vascularized tumor malignancy and decorin expression.

Mechanistically, decorin directly suppresses the HGF/ Met signaling axis that ultimately inhibits VEGFA-mediated angiogenesis.^{85,123} Under normoxic conditions, decorin transcriptionally silences a potent combination of proangiogenic transcription factors, including hypoxia inducible factor-1 α (HIF-1 α), β -catenin, and Myc, downstream of engaging Met (Fig. 2, left panel).⁸⁵ Moreover, decorin noncanonically promotes the degradation of HIF-1a protein in a manner dependent on Von-Hipple Lindau tumor suppressor protein (pVHL).⁸⁵ Within the extracellular milieu, decorin attenuates the liberation of matrix-bound VEGFA by inhibiting the expression and activity of MMP-2 and MMP-9, which depend on competent β-catenin for sufficient transactivation (Fig. 2, left panel). Decorin also promotes the induction and secretion of well-known antiangiogenic effectors such as TSP-1 and TIMP3.85 Intriguingly, decorin promotes rapid secretion of TSP-1 by inhibiting the RhoA/ROCK1 signaling cascade¹²⁴ for early tempering of the initial angiogenicity of the tumor environment. Importantly, decorin significantly abrogates the HGF/Met signaling axis in vivo in the well-established matrigel plug assay, providing firm mechanistic evidence for decorin-mediated angiostasis. In essence, decorin subverts HGF signaling through Met and thus reduces vascularization and vessel density of the malignancy.^{19,85}

The implications of attenuating HIF-1 α and triggering the rapid release of TSP-1 under normoxic conditions open various possibilities of reprogramming the tumor parenchyma and tumor stroma that favor continued tumorigenic growth. It is plausible that decorin disrupts early vascularization events by quelling the angioplasticity of the stroma and repressing potent proangiogenic factors within the tumor proper. Collectively, inhibition of several key EGFR- and Met-mediated pathways including migration, proliferation, survival, and angiogenesis underlies many of the antioncogenic properties of soluble decorin.

New Antioncogenic Properties: Tumor Cell Mitophagy

Fulfilling its role as "a guardian from the matrix," decorin antagonizes tumorigenic progression indirectly by evoking endothelial autophagy and directly by circumventing the angiogenicity of the tumor proper via growth inhibition, suppression of proangiogenic promoters, and secretion of antiangiogenic factors. A novel mechanism that underlies and potentially unifies the classic tumoricidal effects evoked by decorin has been recently delineated.¹²⁵ Functionally akin with VEGFR2 (see above), decorin is a partial agonist of Met for the induction of tumor cell mitochondrial autophagy (mitophagy) (Fig. 2, right panel). At the core of this novel regulatory paradigm is a poorly understood decorin-inducible tumor suppressor gene known as mitostatin.^{126,127} Mitostatin (a mitochondrial protein with oncostatic activity, also known as tricoplein),¹²⁸ embodies all known characteristics of a conventional tumor suppressor while residing at the mitochondria,¹²⁷ possibly at specialized interfaces between the mitochondria and endoplasmic reticulum.¹²⁸

Downstream of Met signaling, decorin elicits rapid post-transcriptional regulation of mitostatin mRNA through PGC-1 α^{125} , a master regulator of mitochondrial biogenesis.¹²⁹ The prompt stabilization of mitostatin mRNA is coordinated by its direct binding to the C-terminal RNA-recognition motif (RRM) of PGC-1 α , which is dependent on arginine methylation of PGC-1 α by PRMT1.¹²⁵ Methylation of RNA binding proteins is commonly required for interactions between this polybasic domain and mRNA target binding.¹³⁰ Interruption of the RRM of PGC-1 α ablates the induction and accumulation of mitostatin protein.¹²⁵ Therefore, we have delineated an operative and mechanistic role for PGC-1 α , a crucial factor for BRAF-mediated oncogenesis,^{131,132} in stabilizing and permitting induction of a decorin-evoked tumor suppressor gene for mitophagic induction (**Fig. 2**, right panel).

Silencing of mitostatin abrogates the ability of breast carcinoma cells to undergo canonical or decorin-evoked mitophagy, as measured by oxidative phosphorylation (OXPHOS) complex turnover, voltage-dependent anion channel (VDAC) activity, and mtDNA depletion¹²⁵ (**Fig. 2**, right panel). An antitumorigenic consequence of mitophagic induction is demonstrated by the inability of decorin to suppress tumor-derived VEGFA in the absence of mitostatin¹²⁵ (**Fig. 2**, right panel). Therefore, mitophagy and decorin-evoked angiostasis may be functionally linked through mitostatin.

Mitophagy is initially evoked following the depolarization of mitochondria.¹³³ Loss of mitochondrial membrane potential is recognized by Parkin, an E3-ubiquitin ligase that is implicated in recessive forms of neurodegenerative disease, such as Parkinson's disease.¹³³ This signal permits discrimination of healthy from failing mitochondria.¹³⁴ As a very early harbinger of mitophagic induction, decorin triggers depolarization of the mitochondrial membrane analogous to that induced by the protonophore FCCP.¹²⁵ Interestingly, cytosolic calcium fluxes are

reported for mitophagy, concomitant with depolarization.¹³⁴ As soluble decorin promotes oscillations of cytosolic calcium in an EGFR-dependent manner,^{135,136} this release may precede and play a role in depolarizing the mitochondria. Furthermore, mitostatin may play a role in coordinating calcium release and subsequent mitochondrial depolarization, as it clusters with mitochondrial-associated membranes and interacts with mitofusion-2.¹²⁸

Mitostatin may interact with, or even function as, an intracellular mitochondrial receptor for Parkin recruitment (Fig. 2, right panel). This scenario appears plausible as Parkin promotes mitophagy^{133,137} and respiratory chain turnover *in vivo*,¹³⁸ mimicking the effects of decorin/mitostatin signaling. Parkin interacts with PINK1, a master mitophagic kinase that senses mitochondrial distress (e.g., loss of membrane potential) and permits activation of Parkin and downstream ubiquitination of target proteins for mitophagic progression.^{139,140} As both Parkin ¹⁴¹ and mitostatin¹²⁸ interact with mitofusion-2, a quaternary complex may exist among PINK1, Parkin, mitostatin, and mitofusion-2 for mitophagic initiation in response to traditional stimuli (e.g., FCCP, CCCP, nutrient deprivation) or decorin.

Currently, no crystal structure of mitostatin exists¹²⁶; however, an in silico analysis of the primary structure revealed an internal domain that shares homology with the DnaJ family of molecular chaperones. Intriguingly, selective destruction of mitochondrial proteins, primarily those associated with the outer mitochondrial membrane, promotes selective respiratory chain component turnover and mitophagy in a PINK1/Parkin-dependent manner.¹³⁴ It appears that mitophagy depends on an association between p62 (also known as sequestosome) and VDAC.¹³⁷ Decorin promotes loss of VDAC in a mitostatin-dependent manner.¹²⁵ Further, long-lived pools of p62/sequestosome promote mammary tumorigenesis via abnormal ErbB2, Akt, and β-catenin activation,¹⁴² whereas mitochondrial turnover promotes degradation of p62/sequestosome.¹⁴³ As such, mitostatin may elicit mitophagy in a PINK1/Parkin-dependent fashion, resulting in the turnover of downstream targets involved in this process (e.g., via p62/sequestosome). Tantalizingly, given that mitostatin may function as a molecular chaperone, additional decorin targets including Myc, β-catenin, and HIF-1α may be targeted for degradation via mitostatin during the course of mitophagic signaling and progression.

Therefore, the induction of mitophagy within the tumor by soluble decorin via mitostatin may account for the molecular outcomes and biological manifestations of decorin, such as inhibition of tumorigenic growth and rampant tumor angiogenesis.

Conclusions and Perspectives

Our increased understanding of the biofunctionality of decorin parallels our evolving and expanding comprehension of the fundamental mechanisms underlying molecular and cellular oncology. Originally characterized as a collagen binding factor and key regulator of fibrillogenesis, decorin has recently emerged as the frontrunner for a novel class of soluble and matrix-derived tumor repressors that potently antagonizes RTK signaling.⁷ The decorin interactome encompasses a broad and diverse repertoire of binding partners that effectively quell the tumor microenvironment and antagonize tumor angiogenesis and metastasis by multiple mechanisms.¹⁹ Original studies investigating decorin as a potent tumoricidal molecule focused on the multitude of interactions between decorin and RTK-enriched tumor cells and downstream antioncogenic and antiangiogenic effects. However, a paradigmatic shift has recently emerged with the demonstration that decorin affects the transcriptomic profile of the tumor microenvironment without significantly perturbing the genetic signature of the tumor itself.⁵⁹ This discovery heralded a new interest in decorin biology to understand the mechanisms operative within the stroma and how these signals interface with the known effects of decorin on the biology of the tumor proper. As such, a new regulatory mechanism has emerged that posits decorin as a procatabolic agent that activates the conserved autophagic machinery.³⁴ Acting as a partial agonist, decorin engages a new class of proautophagic signaling receptors-VEGFR2 for endothelial cell autophagy and Met for tumor cell mitophagythat are activated following decorin binding. Importantly, induction of autophagy and mitophagy may be required for the underlying antitumorigenic effects of decorin on a variety of tumors, including cell cycle arrest, apoptosis, angiogenesis, and metastasis. Such a model connects the secreted extracellular matrix component^{20,144} with complex intracellular metabolic and bioenergetic systems.

Therefore decorin, related SLRPs, and matrix components may be of great clinical interest as advanced chemotherapeutic modalities that could be genetically matched with the patient's individual cancer. Matrix-derived therapies may prove to be valuable armaments in the continued war against cancer.

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References

- Hanahan D, Coussens LM. Accessories to the crime: functions of cells recruited to the tumor microenvironment. Cancer Cell 2012; 21:309-22; PMID:22439926; http://dx.doi.org/10.1016/j.ccr.2012.02.022
- Hanahan D, Winberg RA. Hallmarks of cancer: the next generation. Cell 2011; 144:646-74; PMID:21376230; http://dx.doi.org/10.1016/j.cell.2011.02.013
- Karnoub AE, Dash AB, Vo AP, Sullivan A, Brooks MW, Bell GW, Richardson AL, Polyak K, Tubo R, Weinberg RA. Mesenchymal stem cells within tumor stroma promote breast cancer metastasis. Nature 2007; 449:557-65; PMID:17914389; http://dx.doi. org/10.1038/nature06188
- 4. Curran CS, Keely PJ. Breast tumor and stromal cell response to TGF-b and hypoxia in matrix deposition.

Matrix Biol 2013; 32:95-105; PMID:23262216; http://dx.doi.org/10.1016/j.matbio.2012.11.016

- Hynes RO. Cell-matrix adhesion in vascular development. J Thromb Haemost 2007; 5 (Suppl.1):32-40; PMID:17635706; http://dx.doi.org/10.1111/j.1538-7836.2007.02569.x
- 6. Goldoni S, Iozzo RV. Tumor microenvironment: modulation by decorin and related molecules

harboring leucine-rich tandem motifs. Int J Cancer 2008; 123:2473-9; PMID:18798267; http://dx.doi. org/10.1002/ijc.23930

- Iozzo RV, Sanderson RD. Proteoglycans in cancer biology, tumour microenvironment and angiogenesis. J Cell Mol Med 2011; 15:1013-31; PMID:21155971; http://dx.doi.org/10.1111/j.1582-4934.2010.01236.x
- Merline R, Nastase MV, Iozzo RV, Schaefer L. Small Leucine-rich proteoglycans: multifunctional signaling effectors. In: Karamanos N, ed. Extracellular Matrix: Pathobiology and Signaling. Berlin: Walter de Gruytier Gmbh and Co.; 2012:185-96
- Grahovac J, Wells A. Matrikine and matricellular regulators of EGF receptor signaling on cancer cell migration. Lab Invest 2014; 94:31-40; PMID:24247562; http://dx. doi.org/10.1038/labinvest.2013.132
- Schaefer L, Iozzo RV. Small leucine-rich proteoglycans, at the crossroad of cancer growth and inflammation. Curr Opin Genet Dev 2012; 22:56-7; PMID:22326829; http://dx.doi.org/10.1016/j.gde. 2011.12.002
- Iozzo RV, Schaefer L. Proteoglycans in health and disease: novel regulatory signaling mechanisms evoked by the small leucine-rich proteoglycans. FEBS J 2010; 277:3864-75; PMID:20840584; http://dx.doi.org/ 10.1111/j.1742-4658.2010.07797.x
- Iozzo RV, Goldoni S, Berendsen A, Young MF. Small leucine-rich proteoglycans. In: Mecham RP, ed. Extracellular Matrix: An Overview, Heidelberg, Germany: Springer; 2011:197-231.
- Zhang G, Chen S, Goldoni S, Calder BW, Simpson HC, Owens RT, McQuillan DJ, Young MF, Iozzo RV, Birk DE. Genetic evidence for the coordinated regulation of collagen fibrillogenesis in the cornea by decorin and biglycan. J Biol Chem 2009; 284:8888-97; PMID:19136671; http://dx.doi.org/10.1074/jbc. M806590200
- Zhang G, Ezura Y, Chervoneva I, Robinson PS, Beason DP, Carine ET, Soslowsky LJ, Iozzo RV, Birk DE. Decorin regulates assembly of collagen fibrils and acquisition of biomechanical properties during tendon development. J Cell Biochem 2006; 98:1436-49; PMID:16518859; http://dx.doi.org/10.1002/jcb.20776
- Dunkman AA, Buckley MR, Mienaltowski MJ, Adams SM, Thomas SJ, Satchell L, Kumar A, Pathmanathan L, Beason DP, Iozzo RV, et al. Decorin expression is important for age-related changes in tendon structure and mechanical properties. Matrix Biol 2013; 32:3-13; PMID:23178232; http://dx.doi.org/ 10.1016/j.matbio.2012.11.005
- Chen SC, Young MF, Chakravarti S, Birk DE. Interclass small leucine-rich repeat proteoglycan interactions regulate collagen fibrillogenesis and corneal stromal assembly. Matrix Biol 2014; 35:103-11; PMID:24447998; http://dx.doi.org/10.1016/j.matbio. 2014.01.004
- Reese SP, Underwood CJ, Weiss JA. Effects of decorin proteoglycan on fibrillogenesis, ultrastructure, and mechanics of type I collagen gels. Matrix Biol 2013; 32:414-23; PMID:23608680; http://dx.doi. org/10.1016/j.matbio.2013.04.004
- Takeuchi Y, Kodama Y, Matsumoto T. Bone matrix decorin binds transforming growth factor-b and enhances its bioactivity. J Biol Chem 1994; 269:32634-8; PMID:7798269
- Neill T, Schaefer L, Iozzo RV. Decorin, a guardian from the matrix. Am J Pathol 2012; 181:380-7; PMID:22735579; http://dx.doi.org/10.1016/j.ajpath. 2012.04.029
- Merline R, Moreth K, Beckmann J, Nastase MV, Zeng-Brouwers J, Tralhão JG, Lemarchand P, Pfeilschifter J, Schaefer RM, Iozzo RV et al. Signaling by the matrix proteoglycan decorin controls inflammation and cancer through PDCD4 and microRNA-21. Sci Signal 2011; 4:ra75; PMID:22087031; http://dx. doi.org/10.1126/scisignal.2001868

- Frey T, Schroeder N, Manon-Jensen T, Iozzo RV, Schaefer L. Biological interplay between proteoglycans and their innate immune receptors in inflammation. FEBS J 2013; 280:2165-79; PMID:23350913; http:// dx.doi.org/10.1111/febs.12145
- Seidler DG, Mohamed NA, Bocian C, Stadtmann A, Hermann S, Schäfers K, Schäfers M, Iozzo RV, Zarbock A, Götte M. The role for decorin in delayed-type hypersensitivity. J Immunol 2011; 187:6108-99; PMID:22043007; http://dx.doi.org/ 10.4049/jimmunol.1100373
- Järveläinen H, Puolakkainen P, Pakkanen S, Brown EL, Höök M, Iozzo RV, Sage H, Wight TN. A role for decorin in cutaneous wound healing and angiogenesis. Wound Rep Reg 2006; 14:443-52; http://dx. doi.org/10.1111/j.1743-6109.2006.00150.x
- Baghy K, Iozzo RV, Kovalszky I. Decorin-TGFb axis in hepatic fibrosis and cirrhosis. J Histochem Cytochem 2012; 60:262-8; PMID:22260996
- Nikolovska K, Renke JK, Jungmann O, Grobe K, Iozzo RV, Zamfir AD, Seidler DG. A decorin-deficient matrix affects skin chondroitin/dermatan sulfate levels and keratinocyte function. Matrix Biol 2014; 35:91-102; PMID:24447999; http://dx.doi.org/ 10.1016/j.matbio.2014.01.003
- Baghy K, Dezsó K, László V, Fullár A, Péterfia B, Paku S, Nagy P, Schaff Z, Iozzo RV, Kovalszky I. Ablation of the decorin gene enhances experimental hepatic fibrosis and impairs hepatic healing in mice. Lab Invest 2011; 91:439-51; PMID:20956977; http://dx.doi.org/10.1038/labinvest.2010.172
- Marchica CL, Pinelli V, Borges M, Zummer J, Narayanan V, Iozzo RV, Ludwig MS. A role for decorin in a murine model of allergen-induced asthma. Am J Physiol Lung Cell Mol Physiol 2011; 300:863-73; http://dx.doi.org/10.1152/ajplung.00300.2009
- Merline R, Lazaroski S, Babelova A, Tsalastra-Greul W, Pfeilschifter J, Schluter KD, Gunther A, Iozzo RV, Schaefer RM, Schaefer L. Decorin deficiency in diabetic mice: aggravation of nephropathy due to overexpression of profibrotic factors, enhanced apoptosis and mononuclear cell infiltration. J Physiol Pharmacol 2009; 60 (suppl 4):5-13; PMID:20083846
- Brandan E, Gutierrez J. Role of skeletal muscle proteoglycans during myogenesis. Matrix Biol 2013; 32:289-97; PMID:23583522; http://dx.doi.org/ 10.1016/j.matbio.2013.03.007
- Ichii M, Frank MB, Iozzo RV, Kincade PW. The canonical Wnt pathway shapes niches supportive of hematopoietic stem/progenitor cells. Blood 2012; 119:1683-92; PMID:22117039; http://dx.doi.org/ 10.1182/blood-2011-07-369199
- Zoeller JJ, Pimtong W, Corby H, Goldoni S, Iozzo AE, Owens RT, Ho S-Y, Iozzo RV. A central role for decorin during vertebrate convergent extension. J Biol Chem 2009; 284:11728-37; PMID:19211552; http://dx.doi.org/10.1074/jbc.M808991200
- Schaefer L. Small leucine-rich proteoglycans in kidney disease. J Am Soc Nephrol 2011; 22:1200-7; PMID:21719787; http://dx.doi.org/10.1681/ASN. 2010050570
- Hsieh LT, Nastase MV, Zeng-Brouwers J, Iozzo RV, Schaefer L. Soluble biglycan as a biomarker of inflammatory renal diseases. Int J Biochem Cell Biol 2014; 54C:223-35; http://dx.doi.org/10.1016/j.biocel.2014. 07.020
- Neill T, Schaefer L, Iozzo RV. Instructive roles of extracellular matrix on autophagy. Am J Pathol 2014; 184:2146-53; PMID:24976620; http://dx.doi.org/ 10.1016/j.ajpath.2014.05.010
- Goldoni S, Seidler DG, Heath J, Fassan M, Baffa R, Thakur ML, Owens RA, McQuillan DJ, Iozzo RV. An anti-metastatic role for decorin in breast cancer. Am J Pathol 2008; 173:844-55; PMID:18688028; http://dx.doi.org/10.2353/ajpath.2008.080275
- Bozoky B, Savchenko A, Guven H, Ponten F, Klein G, Szekely L. Decreased decorin expression in the

tumor microenvironment. Cancer Med 2014; 3:485-91; PMID:24634138; http://dx.doi.org/10.1002/ cam4.231

- Iozzo RV, Buraschi S, Genua M, Xu S-Q, Solomides CC, Peiper SC, Gomella LG, Owens RT, Morrione A. Decorin antagonizes IGF receptor I (IGF-IR) function by interfering with IGF-IR activity and attenuating downstream signaling. J Biol Chem 2011; 286:34712-21; PMID:21840990; http://dx.doi.org/ 10.1074/jbc.M111.262766
- Stock C, Jungmann O, Seidler DG. Decorin and chondroitin-6 sulfate inhibit B16V melanoma cell migration and invasion by cellular acidification. J Cell Physiol 2011; 226:2641-50; PMID:21792923; http://dx.doi.org/10.1002/jcp.22612
- 39. Kristensen IB, Pedersen L, Ro TD, Christensen JH, Lyng MB, Rasmussen LM, Ditzel HJ, Borset M, Abildgaard N. Decorin is down-regulated in multiple myeloma and MGUS bone marrow plasma and inhibits HGF-induced myeloma plasma cell viability and migration. Eur J Haematol 2013; 91:196-200; PMID:23607294; http://dx.doi.org/10.1111/ejh.12125
- Iozzo RV, Sampson PM, Schmitt G. Neoplastic modulation of extracellular matrix:stimulation of chondroitin sulfate proteoglycan and hyaluronic acid synthesis in co-cultures of human colon carcinoma and smooth muscle cells. J Cell Biochem 1989; 39:355-78; PMID:2722966; http://dx.doi.org/ 10.1002/jcb.240390403
- Iozzo RV, Bolender RP, Wight TN. Proteoglycan changes in the intercellular matrix of human colon carcinoma. Lab Invest 1982; 47:124-38; PMID:7109538
- Iozzo RV, Wight TN. Isolation and characterization of proteoglycans synthesized by human colon and colon carcinoma. J Biol Chem 1982; 257:11135-44; PMID:7107648
- Brown LF, Guidi AJ, Schnitt SJ, van de Water L, Iruela-Arispe ML, Yeo T-K, Tognazzi K, Dvorak HF. Vascular stroma formation in carcinoma in situ, invasive carcinoma, and metastatic carcinoma of the breast. Clin Cancer Res 1999; 5:1041-56; PMID:10353737
- Leygue E, Snell L, Dotzlaw H, Troup S, Hiller-Hitchcock T, Murphy LC, Roughley PJ, Watson PH. Lumican and decorin are differentially expressed in human breast carcinoma. J Pathol 2000; 192:313-20; PMID:11054714; http://dx.doi.org/10.1002/1096-9896(200011)192:3%3c313::AID-PATH694%3e3. 0.CO;2-B
- Foradori MJ, Chen Q, Fernandez CA, Harper J, Li X, Tsang PC, Langer R, Moses MA. Matrilin-1 is an inhibitor of neovascularization. J Biol Chem 2014; 289:14301-9; PMID:24692560; http://dx.doi.org/ 10.1074/jbc.M113.529982
- 46. Boström P, Sainio A, Kakko T, Savontaus M, Söderström M, Järveläinen H. Localization of decorin gene expression in normal human breast tissue and in benign and malignant tumors of the human breast. Histochem Cell Biol 2013; 139:161-71; PMID:23007289; http://dx.doi.org/10.1007/s00418-012-1026-0
- Henke A, Grace OC, Ashley GR, Stewart GD, Riddick ACP, Yeun H, O'Donnell M, Anderson RA, Thomson AA. Stromal expression of decorin, semaphorin6D, SPARC, Sprouty 1 and Tsukushi in developing prostate and decreased levels of decorin in prostate cancer. PLoS One 2012; 7:e4251; http://dx. doi.org/10.1371/journal.pone.0042516
- Sainio A, Nyman M, Lund R, Vuorikoski S, Boström P, Laato M, Boström PJ, Järveläinen H. Lack of decorin expression by human bladder cancer cells offers new tools in the therapy of urothelial malignancies. PLoS One 2013; 8:e76190; PMID:24146840; http://dx.doi.org/10.1371/journal.pone.0076190
- Horvath Z, Kovalszky I, Fullar A, Kiss K, Schaff Z, Iozzo RV, Baghy K. Decorin deficiency promotes

hepatic carcinogenesis. Matrix Biol 2014; 35:194-205; PMID:24361483; http://dx.doi.org/10.1016/j. matbio.2013.11.004

- Duncan MB. Extracellular matrix transcriptome dynamics in hepatocellular carcinoma. Matrix Biol 2013; 32:393-8; PMID:23727079; http://dx.doi.org/ 10.1016/j.matbio.2013.05.003
- Dyrskjøt L, Kruhøffer M, Thykjaer T, Marcussen N, Jensen JL, Møller K, Ørntoft TF. Gene expression in the urinary bladder: a common carcinoma in situ gene expression signature exists disregarding histopathological classification. Cancer Res 2004; 64:4040-8; PMID:15173019; http://dx.doi.org/10.1158/0008-5472.CAN-03-3620
- Sanchez-Carbayo M, Socci ND, Lozano J, Saint F, Cordon-Cardo C. Defining molecular profiles of poor outcome in patients with invasive bladder cancer using oligonucleotide microarrays. J Clin Oncol 2006; 24:778-9; PMID:16432078; http://dx.doi.org/ 10.1200/JCO.2005.03.2375
- Danielson KG, Baribault H, Holmes DF, Graham H, Kadler KE, Iozzo RV. Targeted disruption of decorin leads to abnormal collagen fibril morphology and skin fragility. J Cell Biol 1997; 136:729-43; PMID:9024701; http://dx.doi.org/10.1083/jcb.136. 3.729
- 54. Bi X, Tong C, Dokendorff A, Banroft L, Gallagher L, Guzman-Hartman G, Iozzo RV, Augenlicht LH, Yang W. Genetic deficiency of decorin causes intestinal tumor formation through disruption of intestinal cell maturation. Carcinogenesis 2008; 29:1435-40; PMID:18550571; http://dx.doi.org/10.1093/carcin/ bgn141
- 55. Iozzo RV, Chakrani F, Perrotti D, McQuillan DJ, Skorski T, Calabretta B, Eichstetter I. Cooperative action of germline mutations in decorin and p53 accelerates lymphoma tumorigenesis. Proc Natl Acad Sci U S A 1999; 96:3092-7; PMID:10077642; http:// dx.doi.org/10.1073/pnas.96.6.3092
- Reed CC, Waterhouse A, Kirby S, Kay P, Owens RA, McQuillan DJ, Iozzo RV. Decorin prevents metastatic spreading of breast cancer. Oncogene 2005; 24:1104-10; PMID:15690056; http://dx.doi.org/10.1038/sj. onc.1208329
- Reed CC, Gauldie J, Iozzo RV. Suppression of tumorigenicity by adenovirus-mediated gene transfer of decorin. Oncogene 2002; 21:3688-95; PMID:12032837; http://dx.doi.org/10.1038/sj.onc. 1205470
- Seidler DG, Goldoni S, Agnew C, Cardi C, Thakur ML, Owens RA, McQuillan DJ, Iozzo RV. Decorin protein core inhibits in vivo cancer growth and metabolism by hindering epidermal growth factor receptor function and triggering apoptosis via caspase-3 activation. J Biol Chem 2006; 281:26408-18; PMID:16835231; http://dx.doi.org/10.1074/jbc. M602853200
- Buraschi S, Neill T, Owens RT, Iniguez LA, Purkins G, Vadigepalli R, Evans B, Schaefer L, Peiper SC, Wang Z et al. Decorin protein core affects the global gene expression profile of the tumor microenvironment in a triple-negative orthotopic breast carcinoma xenograft model. PLoS One 2012; 7:e45559; PMID:23029096; http://dx.doi.org/10.1371/journal. pone.0045559
- Li X, Pennisi A, Yaccoby S. Role of decorin in the antimyeloma effects of osteoblasts. Blood 2008; 112:159-68; PMID:18436739; http://dx.doi.org/ 10.1182/blood-2007-11-124164
- Buraschi S, Pal N, Tyler-Rubinstein N, Owens RT, Neill T, Iozzo RV. Decorin antagonizes Met receptor activity and downregulates b-catenin and Myc levels. J Biol Chem 2010; 285:42075-85; PMID:20974860; http://dx.doi.org/10.1074/jbc.M110.172841
- Morrione A, Neill T, Jozzo RV. Dichotomy of decorin activity on the insulin-like growth factor-I system. FEBS J 2013; 280:2138-49; PMID:23351020; http://dx.doi.org/10.1111/febs.12149

- Nikitovic D, Aggelidakis J, Young MF, Iozzo RV, Karamanos NK, Tzanakakis GN. The biology of small leucine-rich proteoglycans in bone pathophysiology. J Biol Chem 2012; 287:33926-33; PMID:22879588; http://dx.doi.org/10.1074/jbc. R112.379602
- 64. Kuroiwa Y, Kaneko-Ishino T, Kagitani F, Kohda T, Li L-L, Tada M, Suzuki R, Yokoyama M, Shiroishi T, Wakana S et al. Peg3 imprinted gene on proximal chromosome 7 encodes for a zinc finger protein. Nature Genet 1996; 12:186-90; PMID:8563758; http://dx.doi.org/10.1038/ng0296-186
- 65. Yamaguchi A, Taniguchi M, Hori O, Ogawa S, Tojo N, Matsuoka N, Miyake S, Kasai K, Sugimoto H, Tamatani M et al. Peg3/Pw1 is involved in p53-mediated cell death pathway in brain ischemia/hypoxia. J Biol Chem 2002; 277:623-9; PMID:11679586; http://dx.doi.org/10.1074/jbc.M107435200
- 66. Kohda T, Asai A, Kuroiwa Y, Kobayashi S, Aisaka K, Nagashima G, Yoshida MC, Kondo Y, Kagiyama N, Kirino T et al. Tumour suppressor activity of human imprinted gene PEG3 in a glioma cell line. Genes Cells 2001; 6:237-47; PMID:11260267; http://dx. doi.org/10.1046/j.1365-2443.2001.00412.x
- Dowdy SC, Gostout BS, Shridhar V, Wu X, Smith DI, Podratz KC, Jiang S-W. Biallelic methylation and silencing of paternally expressed gene 3 (PEG3) in gynecologic cancer cell lines. Gynecol Oncol 2005; 99:126-34; PMID:16023706; http://dx.doi.org/ 10.1016/j.ygyno.2005.05.036
- Jiang X, Yu Y, Yang HW, Agar NYR, Frado L, Johnson MD. The imprinted gene PEG3 inhibits Wnt signaling and regulates glioma growth. J Biol Chem 2010; 285:8472-80; PMID:20064927; http://dx.doi.org/10.1074/jbc.M109.069450
- Buraschi S, Neill T, Goyal A, Poluzzi C, Smythies J, Owens RT, Schaefer L, Torres AT, Jozzo RV. Decorin causes autophagy in endothelial cells via Peg3. Proc Natl Acad Sci USA 2013; 110:E2582-E2591; PMID:23798385; http://dx.doi.org/10.1073/ pnas.1305732110
- Purushothaman A, Babitz A, Sanderson RD. Heparanase enhances the insulin receptor signaling pathway to activate extracellular signal-regulated kinase in multiple myeloma. J Biol Chem 2012; 287:41288-96; PMID:23048032; http://dx.doi.org/10.1074/jbc. M112.391417
- Neill T, Torres AT, Buraschi S, Iozzo RV. Decorin has an appetite for endothelial cell autophagy. Autophagy 2013; 9:1626-8; PMID:23989617; http:// dx.doi.org/10.4161/auto.25881
- Funderburk SF, Wang QJ, Yue Z. The Beclin 1-VPS34 complex- at the crossroads of autophagy and beyond. Trends Cell Biol 2010; 20:355-62; PMID:20356743; http://dx.doi.org/10.1016/j. rcb.2010.03.002
- Wang RC, Wei Y, An Z, Zou Z, Xiao G, Bhagat G, White M, Reichelt J, Levine B. Akt-mediated regulation of autophagy and tumorigenesis through Beclin 1 phosphorylation. Science 2012; 338:956-9; PMID:23112296; http://dx.doi.org/10.1126/ science.1225967
- 74. Alers S, Löffler AS, Wesselborg S, Stork B. Role of AMPK-mTOR-Ulk1/2 in the regulation of autophagy: crosstalk, shortcuts, and feedbacks. Mol Cell Biol 2012; 32:2-11; PMID:22025673; http://dx.doi.org/ 10.1128/MCB.06159-11
- Goyal A, Neill T, Owens RT, Schaefer L, Iozzo RV. Decorin activates AMPK, an energy sensor kinase, to induce autophagy in endothelial cells. Matrix Biol 2014; 34:46-54; PMID:24472739; http://dx.doi.org/ 10.1016/j.matbio.2013.12.011
- Pattingre S, Levine B. Bcl⁻² inhibition of autophagy: a new route to cancer? Cancer Res 2006; 66:2885-8; PMID:16540632; http://dx.doi.org/10.1158/0008-5472.CAN-05-4412
- 77. Kim J, Kundu M, Viollet B, Guan K-L. AMPK and mTOR regulate autophagy through direct

phopshorylation of Ulk1. Nat Cell Biol 2011; 13:132-41; PMID:21258367; http://dx.doi.org/ 10.1038/ncb2152

- Lee JW, Park S, Takahashi Y, Wang H-G. The association of AMPK with ULK1 regulates autophagy. PLoS One 2010; 5:e15394; PMID:21072212; http:// dx.doi.org/10.1371/journal.pone.0015394
- 79. Wei Y, Zou Z, Becker N, Anderson M, Sumpter R, Xiao G, Kinch L, Koduru P, Christudass CS, Veltri RW et al. EGFR-mediated beclin 1 phosphorylation in autophagy suppression, tumor progression, and tumor chemoresistance. Cell 2013; 154:1269-84; PMID:24034250; http://dx.doi.org/10.1016/j.cell. 2013.08.015
- Levine B, Kroemer G. Autophagy in the pathogenesis of disease. Cell 2008; 132:27-42; PMID:18191218; http://dx.doi.org/10.1016/j.cell.2007.12.018
- Choi AMK, Ryter SW, Levine B. Autophagy in human health and disease. New Engl J Med 2013; 368:651-62; PMID:23406030; http://dx.doi.org/ 10.1056/NEJMra1205406
- Settembre C, Di Malta C, Polito VA, Arencibia MG, Vetrini F, Erdin S, Huynh T, Medina D, Colella P, Sardiello M et al. TFEB links autophagy to lysosomal biogenesis. Science 2011; 332:1429-33; PMID:21617040; http://dx.doi.org/10.1126/science. 1204592
- Roczniak-Ferguson A, Petit CS, Froehlich F, Qian S, Ky J, Angarola B, Walther TC, Ferguson SM. The transcription factor TFEB links mTORC1 signaling to transcriptional control of lysosome homeostasis. Sci Signal 2012; 5:ra42; PMID:22692423; http://dx.doi. org/10.1126/scisignal.2002790
- Settembre C, Zoncu R, Medina DL, Vetrini F, Erdin S, Erdin S, Huynh T, Ferron M, Karsenty G, Vellard MC et al. A lysosome-to-lysosome signaling mechanism senses and regulates the lysosome via mTOR and TFEB. EMBO J 2012; 31:1095-108; PMID:22343943; http://dx.doi.org/10.1038/emboj. 2012.32
- Neill T, Painter H, Buraschi S, Owens RT, Lisanti MP, Schaefer L, Iozzo RV. Decorin antagonizes the angiogenic network. Concurrent inhibition of Met, hipoxia inducible factor-1a and vascular endothelial growth factor A and induction of thrombospondin-1 and TIMP3. J Biol Chem 2012; 287:5492-506; PMID:22194559; http://dx.doi.org/10.1074/jbc. M111.283499
- Moreth K, Iozzo RV, Schaefer L. Small leucine-rich proteoglycans orchestrate receptor crosstalk during inflammation. Cell Cycle 2012; 11:2084-91; PMID:22580469; http://dx.doi.org/10.4161/cc. 20316
- Moreth K, Frey H, Hubo M, Zeng-Brouwers J, Nastase MV, Hsieh LT, Haceni R, Pfeilschifter J, Iozzo RV, Schaefer L. Biglycan-triggered TLR-2- and TLR-4-signaling exacerbates the pathophysiology of ischemic acute kidney injury. Matrix Biol 2014; 35:143-51; PMID:24480070; http://dx.doi.org/10.1016/j. matbio.2014.01.010
- Zeng-Brouwers J, Beckmann J, Nastase MV, Iozzo RV, Schaefer L. De novo expression of circulating biglycan evokes an innate inflammatory tissue response via MyD88/TRIF pathways. Matrix Biol 2014; 35:132-42; PMID:24361484; http://dx.doi. org/10.1016/j.matbio.2013.12.003
- Yamamoto K, Ohga N, Hida Y, Maishi N, Kawamoto T, Kitayama K, Akiyama K, Osawa T, Kondoh M, Matsuda K et al. Biglycan is a specific marker and an autocrine angiogenic factor of tumour endothelial cells. Brit J Cancer 2012; 106:1214-23; PMID:22374465; http://dx.doi.org/ 10.1038/bjc.2012.59
- Berendsen AD, Pinnow EL, Maeda A, Brown AC, McCartney-Francis N, Kram V, Owens RT, Robey PG, Holmbeck K, de Castro LF et al. Biglycan modulates angiogenesis and bone formation during fracture healing. Matrix Biol 2014; 35:223-31; PMID:24373744; http://dx.doi.org/10.1016/j.matbio.2013.12.004

- Poluzzi C, Casulli J, Goyal A, Mercer TJ, Neill T, Iozzo RV. Endorepellin evokes autophagy in endothelial cells. J Biol Chem 2014; 289:16114-28; PMID:24737315; http://dx.doi.org/10.1074/jbc.M114. 556530
- Moscatello DK, Santra M, Mann DM, McQuillan DJ, Wong AJ, Iozzo RV. Decorin suppresses tumor cell growth by activating the epidermal growth factor receptor. J Clin Investig 1998; 101:406-12; PMID:9435313; http://dx.doi.org/10.1172/JCI846
- Iozzo RV, Moscatello D, McQuillan DJ, Eichstetter I. Decorin is a biological ligand for the epidermal growth factor receptor. J Biol Chem 1999; 274:4489-92; PMID:9988678; http://dx.doi.org/10.1074/ jbc.274.8.4489
- Santra M, Reed CC, Iozzo RV. Decorin binds to a narrow region of the epidermal growth factor (EGF) receptor, partially overlapping with but distinct from the EGF-binding epitope. J Biol Chem 2002; 277:35671-81; PMID:12105206; http://dx.doi.org/ 10.1074/jbc.M205317200
- Santra M, Eichstetter I, Iozzo RV. An anti-oncogenic role for decorin: downregulation of ErbB2 leads to growth suppression and cytodifferentiation of mammary carcinoma cells. J Biol Chem 2000; 275:35153-61; PMID:10942781; http://dx.doi.org/10.1074/jbc. M006821200
- Goldoni S, Owens RT, McQuillan DJ, Shriver Z, Sasisekharan R, Birk DE, Campbell S, Iozzo RV. Biologically active decorin is a monomer in solution. J Biol Chem 2004; 279:6606-12; PMID:14660661; http://dx.doi.org/10.1074/jbc.M310342200
- Zhu J-X, Goldoni S, Bix G, Owens RA, McQuillan D, Reed CC, Iozzo RV. Decorin evokes protracted internalization and degradation of the EGF receptor via caveolar endocytosis. J Biol Chem 2005; 280:32468-79; PMID:15994311; http://dx.doi.org/ 10.1074/jbc.M503833200
- Lemmon MA, Schlessinger J. Cell signaling by receptor tyrosine kinases. Cell 2010; 141:1117-34; PMID:20602996; http://dx.doi.org/10.1016/j.cell. 2010.06.011
- Goldoni S, Humphries A, Nyström A, Sattar S, Owens RT, McQuillan DJ, Ireton K, Iozzo RV. Decorin is a novel antagonistic ligand of the Met receptor. J Cell Biol 2009; 185:743-54; PMID:19433454; http://dx.doi.org/10.1083/jcb. 200901129
- Belov AA, Mohammadi M. Grb2, a double-edged sword of receptor tyrosine kinase signaling. Sci Signal 2012; 5:e49; http://dx.doi.org/10.1126/scisignal. 2003576
- 101. Minor KH, Bournat JC, Toscano N, Giger RJ, Davies SJA. Decorin, erythroblastic leukaemia viral oncogene homologue B4 and signal transducer and activator of transcription 3 regulation of semaphorin 3A in central nervous system scar tissue. Brain 2011; 134:1140-55; PMID:21115466; http://dx.doi.org/10.1093/brain/ awq304
- Schönherr E, Sunderkötter C, Iozzo RV, Schaefer L. Decorin, a novel player in the insulin-like growth factor system. J Biol Chem 2005; 280:15767-72; PMID:15701628; http://dx.doi.org/10.1074/jbc. M500451200
- Morcavallo A, Buraschi S, Xu S-Q, Belfiore A, Schaefer L, Iozzo RV, Morrione A. Decorin differentially modulates the activity of insulin receptor isoform A ligands. Matrix Biol 2014; 35:82-90; PMID:24389353; http://dx.doi.org/10.1016/j.matbio. 2013.12.010
- Baghy K, Horváth Z, Regös E, Kiss K, Schaff Z, Iozzo RV, Kovalszky I. Decorin interferes with plateletderived growth factor receptor signaling in experimental hepatocarcinogenesis. FEBS J 2013; 280:2150-64; PMID:23448253; http://dx.doi.org/10.1111/febs. 12215
- 105. Lala N, Gannareddy VG, Cloutier-Bosworth A, Lala PK. Mechanisms in decorin regulation of vascular

endothelial growth factor-induced human trophoblast migration and acquisition of endothelial phenotype. Biol Reprod 2012; 87:(article 59)1-14.

- 106. Khan GA, Girish GV, Lala N, DiGuglielmo GM, Lala PK. Decorin is a novel VEGFR-2-binding antagonist for the human extravillous trophoblast. Mol Endocrinol 2011; 25:1431-43; PMID:21659473; http://dx.doi.org/10.1210/me.2010-0426
- 107. Purdie CA, Harrison DJ, Peter A, Dobbie L, White S, Howie SEM, Salter DM, Bird CC, Wyllie AH, Hooper ML et al. Tumour incidence, spectrum and ploidy in mice with a large deletion in the p53 gene. Oncogene 1994; 9:603-9; PMID:8290271
- Becker W. Emerging role of DYRK family protein kinases as regulators of protein stability in cell cycle control. Cell Cycle 2012; 11:3389-94; PMID:22918246; http://dx.doi.org/10.4161/cc.21404
- Yoshida Y, Wang I-C, Yoder HM, Davidson NO, Costa RH. The forkhead box M1 transcription factor contributes to the development and growth of mouse colorectal cancer. Gastroenterology 2007; 132:1420-31; PMID:17408638; http://dx.doi.org/10.1053/j. gastro.2007.01.036
- 110. Holland JD, Gyorffy B, Vogel R, Eckert K, Valenti G, Fang L, Lohneis P, Elezkurtaj S, Ziebold U, Birchmeier W. Combined Wnt/β-catenin, Met, and CXCL12/CXCR4 signals characterize basal breast cancer and predict disease outcome. Cell Rep 2013; 5:1214-27; PMID:24290754; http://dx.doi.org/ 10.1016/j.celrep.2013.11.001
- 111. Nair R, Roden DL, Teo WS, McFarland A, Junankar S, Ye S, Nguyen A, Yang J, Nikolic I, Hui M et al. c-Myc and Her2 cooperate to drive a stem-like phenotype with poor prognosis in breast cancer. Oncogene 2014; 33:3992-4002; PMID:24056965; http://dx. doi.org/10.1038/onc.2013.368
- 112. Walz S, Lorenzin F, Morton J, Wiese KE, von EB, Herold S, Rycak L, Dumay-Odelot H, Karim S, Bartkuhn M et al. Activation and repression by oncogenic MYC shape tumour-specific gene expression profiles. Nature 2014; 511:483-7; PMID:25043018; http:// dx.doi.org/10.1038/nature13473
- 113. Huang S, Ouyang N, Lin L, Chen L, Wu W, Su F, Yao Y, Yao H. HGF-induced PKCzeta activation increases functional CXCR4 expression in human breast cancer cells. PLoS One 2012; 7:e29124; PMID:22242160; http://dx.doi.org/10.1371/journal. pone.0029124
- 114. Soria-Valles C, Gutiérrez-Fernández A, Guiu M, Mari B, Fueyo A, Gomis RR, López-Otin C. The anti-metastatic activity of collagenase-2 in breast cancer cells is mediated by a signaling pathway involving decorin and miR-21. Oncogene 2014; 33:3054-63; PMID:23851508; http://dx.doi.org/10.1038/onc. 2013.267
- 115. Goyal A, Poluzzi C, Willis AC, Smythies J, Shellard A, Neill T, Iozzo RV. Endorepellin affects angiogenesis by antagonizing diverse VEGFR2- evoked signaling pathways: transcriptional repression of HIF-1a and VEGFA and concurrent inhibition of NFAT1 activation. J Biol Chem 2012; 287:43543-56; PMID:23060442; http://dx.doi.org/10.1074/jbc. M112.401786
- 116. Goyal A, Pal N, Concannon M, Paulk M, Doran M, Poluzzi C, Sekiguchi K, Whitelock JM, Neill T, Iozzo RV. Endorepellin, the angiostatic module of perlecan, interacts with both the a2b1 integrin and vascular endothelial growth factor receptor 2 (VEGFR2). J Biol Chem 2011; 286:25947-62; PMID:21596751; http://dx.doi.org/10.1074/jbc.M111.243626
- Folkman J. Antiangiogenesis in cancer therapy endostatin and its mechanisms of action. Exp Cell Res 2006; 312:594-607; PMID:16376330; http://dx.doi. org/10.1016/j.yexcr.2005.11.015
- Kyriakides TR, MacLauchlan S. The role of thrombospondins in wound healing, ischemia, and the foreign body reaction. J Cell Commun Signal 2009; 3:

215-25; PMID:19844806; http://dx.doi.org/ 10.1007/s12079-009-0077-z

- 119. Schönherr E, Sunderkotter C, Schaefer L, Thanos S, Grässel S, Oldberg Å, Iozzo RV, Young MF, Kresse H. Decorin deficiency leads to impaired angiogenesis in injured mouse cornea. J Vasc Res 2004; 41:499-508; PMID:15528932; http://dx.doi.org/10.1159/ 000081806
- 120. Mohan RR, Tovey JCK, Sharma A, Schultz G, Cowden JW, Tandon A. Targeted decorin gene therapy delivered with adeno-associated virus effectively retards corneal neovascularization in vivo. PLoS One 2011; 6:e26432; PMID:22039486; http://dx.doi.org/ 10.1371/journal.pone.0026432
- 121. Grant DS, Yenisey C, Rose RW, Tootell M, Santra M, Iozzo RV. Decorin suppresses tumor cell-mediated angiogenesis. Oncogene 2002; 21:4765-77; PMID:12101415; http://dx.doi.org/10.1038/sj.onc. 1205595
- 122. Salomäki HH, Sainio AO, Söderström M, Pakkanen S, Laine J, Järveläinen HT. Differential expression of decorin by human malignant and benign vascular tumors. J Histochem Cytochem 2008; 56:639-46; http://dx.doi.org/10.1369/jhc.2008.950287
- 123. Peschard P, Ishiyama N, Lin T, Lipkowitz S, Park M. A conserved DpYR Motif in the juxtamembrane domain of the Met receptor family forms an atypical c-Cbl/Cbl-b tyrosine kinase binding domain binding site required for suppression of oncogenic activation. J Biol Chem 2004; 279:29565-71; PMID:15123609; http://dx.doi.org/10.1074/jbc.M403954200
- 124. Neill T, Jones HR, Crane-Smith Z, Owens RT, Schaefer L, Iozzo RV. Decorin induces rapid secretion of thrombospondin-1 in basal breast carcinoma cells via inhibition of Ras homolog gene family, member A/Rho-associated coiled-coil containing protein kinase 1. FEBS J 2013; 280:2353-68; PMID:23350987; http://dx.doi.org/10.1111/febs. 12148
- 125. Neill T, Torres A, Burchard J, Owens RT, Hoek J, Baffa R, Iozzo RV. Decorin induces mitophagy in breast carcinoma cells via peroxisome proliferator-activated receptor g coactivator-1a (PGC-1a) and mitostatin. J Biol Chem 2014; 289:4952-68; PMID:24403067; http://dx.doi.org/10.1074/jbc. M113.512566
- 126. Vecchione A, Fassan M, Anesti V, Morrione A, Goldoni S, Baldassarre G, Byrne D, D'Arca D, Palazzo JP, Lloyd J et al. MITOSTATIN, a putative tumor suppressor on chromosome 12q24.1, is downregulated in human bladder and breast cancer. Oncogene 2009; 28:257-69; PMID:18931701; http://dx.doi.org/ 10.1038/onc.2008.381
- 127. Fassan M, D'Arca D, Letko J, Vecchione A, Gardiman MP, McCue P, Wildemore B, Rugge M, Shupp-Byrne D, Gomella LG et al. Mitostatin is down-regulated in human prostate cancer and suppresses the invasive phenotype of prostate cancer cells. PLoS One 2011; 6:e19771; PMID:21573075; http://dx.doi.org/ 10.1371/journal.pone.0019771
- Cerqua C, Anesti V, Pyakurel A, Liu D, Naon D, Wiche G, Baffa R, Dimmer KS, Scorrano L. Trichoplein/mitostatin regulates endoplasmic reticulummitochondria juxtaposition. EMBO Reports 2010; 11:854-60; PMID:20930847; http://dx.doi.org/ 10.1038/embor.2010.151
- Ventura-Clapier R, Garnier A, Veksler W. Transcriptional control of mitochondrial biogenesis: the central role of PGC-1a. Cardiovasc Res 2008; 79:208-17; PMID:18430751; http://dx.doi.org/10.1093/cvr/ cvn098
- Teyssier C, Ma H, Emter R, Kralli A, Stallcup MR. Activation of nuclear receptort coactivator PGC-1a by arginine methylation. Genes Dev 2005; 19:1466-73; PMID:15964996; http://dx.doi.org/10.1101/gad. 1295005
- 131. Haq R, Shoag J, Andreu-Perez P, Yokoyama S, Edelman H, Rowe GC, Frederick DT, Hurley AD,

Nellore A, Kung AL et al. Oncogenic BRAF regulates oxidative metabolism via PGC1a and MITF. Cancer Cell 2013; 23:302-15; PMID:23477830; http://dx. doi.org/10.1016/j.ccr.2013.02.003

- 132. Vazquez F, Lim J-H, Chim H, Bhalla K, Girnun G, Pierce K, Clish CB, Granter SR, Widlund HR, Spiegelman BM et al. PGC1a expression defines a subset of human melanoma tumors with increased mitochondrial capacity and resistance to oxidative stress. Cancer Cell 2013; 23:287-301; PMID:23416000; http://dx.doi.org/10.1016/j.ccr.2012.11.020
- Dagda R, Cherra SJI, Kulich SM, Tandon A, Park D, Chu CT. Loss of PINK1 function promotes mitophagy through effects on oxidative stress and mitochondrial fission. J Biol Chem 2009; 284:13843-55; PMID:19279012; http://dx.doi.org/10.1074/jbc. M808515200
- 134. Narendra D, Walker JE, Youle R. Mitochondrial quality control mediated by PINK1 and Parkin: links to parkinsonism. Cold Spring Harb Perspect Biol 2012; 4:pii: a011338; PMID:23125018
- Patel S, Santra M, McQuillan DJ, Iozzo RV, Thomas AP. Decorin activates the epidermal growth factor receptor and elevates cytosolic Ca²⁺ in A431 cells. J Biol Chem 1998; 273:3121-4; PMID:9452417; http://dx.doi.org/10.1074/jbc.273.6.3121

- 136. Csordás G, Santra M, Reed CC, Eichstetter I, McQuillan DJ, Gross D, Nugent MA, Hajnóczky G, Iozzo RV. Sustained down-regulation of the epidermal growth factor receptor by decorin. A mechanism for controlling tumor growth in vivo. J Biol Chem 2000; 275:32879-87; http://dx.doi.org/10.1074/jbc. M005609200
- 137. Geisler S, Holmstrom KM, Skujat D, Fiesel FC, Rothfuss OC, Kahle PJ, Springer W. PINK1/Parkinmediated mitophagy is dependent on VDAC1 and p62/SQSTM1. Nat Cell Biol 2010; 12:119-31; PMID:20098416; http://dx.doi.org/10.1038/ncb2012
- 138. Vincow ES, Merrihew G, Thomas RE, Shulman NJ, Beyer RP, MacCoss MJ, Pallanck LJ. The PINK1-Parkin pathway promotes both mitophagy and selective respiratory chain turnover in vivo. Proc Natl Acad Sci U S A 2013; 110:6400-5; PMID:23509287; http://dx.doi.org/10.1073/pnas.1221132110
- 139. Iguchi M, Kujuro Y, Okatsu K, Koyano F, Kosako H, Kimura M, Suzuki N, Uchiyama S, Tanaka K, Matsuda N. Parkin-catalyzed ubiquitin-ester transfer is triggered by PINK1-dependent phosphorylation. J Biol Chem 2013; 288:22019-32; PMID:23754282; http://dx.doi.org/10.1074/jbc.M113.467530
- 140. Rakovic A, Grunewald A, Kottwitz J, Bruggemann N, Pramstaller PP, Lohmann K, Klein C. Mutations in

PINK1 and Parkin impair ubiquitination of Mitofusins in human fibroblasts. PLoS One 2011; 6:e16746; PMID:21408142; http://dx.doi.org/10.1371/journal. pone.0016746

- 141. Poole AC, Thomas RE, Yu S, Vincow ES, Pallanck L. The mitochondrial fusion-promoting factor mitofusin is a substrate of the PINK1/parkin pathway. PLoS One 2010; 5:e10054; PMID:20383334; http://dx. doi.org/10.1371/journal.pone.0010054
- Cai-McRae X, Zhong H, Karantza V. Sequestosome 1/p62 facilitates HER2-induced mammary tumorigenesis through multiple signaling pathways. Oncogene 2015: in press; PMID:25088198
- Komatsu M, Ichimura Y. Physiological significance of selective degradation of p62 by autophagy. FEBS Lett 2010; 584:1374-8; PMID:20153326; http://dx.doi. org/10.1016/j.febslet.2010.02.017
- 144. Bolton K, Segal D, McMillan J, Jowett J, Heilbronn L, Abberton K, Zimmet P, Chisholm D, Collier G, Walder K. Decorin is a secreted protein associated with obesity and type 2 diabetes. Int J Obes 2008; 32:1113-21; http://dx.doi.org/10.1038/ijo.2008.41