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A bird's-eye view of deep learning in bioimage analysis

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1. Introduction

Ever since the introduction of digital scanning technologies in biological imaging [1-3], there has been a growing need for powerful computational methods to enable automated quantitative image analysis. Microscopy images potentially contain a wealth of information about the morphological, structural, and dynamical characteristics of tissues, cells, and molecules, which may go unnoticed even to the expert human eye [4-6]. However, designing computer algorithms to extract this information with high fidelity is a great challenge, as has been well recognized since the mid-1960s, after the first decade of serious attempts [7-9], and is still true today.

Automated bioimage analysis typically requires executing an intricate series of operations, which may involve image restoration [10-12] and registration [13-15], object detection [16-18], segmentation [17,19,20], and tracking [21-23], as well as downstream image or object classification [24-26], quantification [27-29], and visualization [30-32]. As attested by the just cited reviews and evaluations, a plethora of methods and tools have been developed for this purpose in the first half a century of computational bioimage analysis, based on what may now be considered traditional image processing and computer vision paradigms.

Recently, a major paradigm shift has occurred with the massive adoption of deep learning technologies [33–35], which are now rapidly replacing traditional data analysis approaches in virtually all fields of science, including bioimage analysis. In a matter of just

ABSTRACT

Deep learning of artificial neural networks has become the de facto standard approach to solving data analysis problems in virtually all fields of science and engineering. Also in biology and medicine, deep learning technologies are fundamentally transforming how we acquire, process, analyze, and interpret data, with potentially far-reaching consequences for healthcare. In this mini-review, we take a bird'seye view at the past, present, and future developments of deep learning, starting from science at large, to biomedical imaging, and bioimage analysis in particular.

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a few years, the scientific literature on deep learning has grown explosively, not only with research papers describing novel concepts, algorithms, software platforms, and applications, but also with an abundance of reviews and surveys exploring and commenting on the state of the art.

In this mini-review, we take stock and summarize the latest developments and the challenges ahead, starting from science at large, to biomedical imaging, and to bioimage analysis in particular. Rather than providing a technical introduction or an exhaustive review, we briefly discuss major trends in the past, present, and future of deep learning and their implications for bioimage analysis. Along the way, we mainly cite other reviews and surveys for further reading on specific subtopics.

2. Deep learning on the rise

Deep learning popularly refers to the use of artificial neural networks (ANNs) with multiple (ultimately many) layers of elementary computational cells (called "neurons" by analogy with neuronal cells in biological neural networks) to progressively extract higher-level representations of given input data in order to perform data analysis tasks [33,35,36]. It is a form of machine learning [34,37,38], a major branch of the field of artificial intelligence (AI) [39–41], which is concerned with the science and engineering of developing machines exhibiting characteristics associated with human intelligence. While deep learning is now

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taking the world by storm (Fig. 1), its road to success has been long and arduous.

2.1. A brief history of deep learning

The idea of using ANNs for data analysis dates back to the dawn of digital computing [42]. In the early 1940s, the first mathematical model of a biological neuron was proposed, providing "a tool for rigorous symbolic treatment of known nets and an easy method of constructing hypothetical nets of required properties" [43]. In order for the model to work, its parameters (weights) had to be set correctly, which initially was done manually. In the late 1950s, the perceptron became the first model capable of learning the weights from examples, illustrating "some of the fundamental properties of intelligent systems" [44]. However, by the end of the 1960s, it was clear that such models have severe limitations [45], and multilayer perceptrons are required for more complex tasks, but it was not obvious how to train them.

After the ensuing first "AI winter", from the late 1960s throughout the 1970s and some time beyond, interest in ANNs resurged in the mid-1980s with the (re)invention of the back-propagation algorithm [46]. Important advances were made in the 1990s, including the development of recurrent neural networks (RNNs) such as the long short-term memory (LSTM) for modeling data sequences [47], and successful applications of multilayer convolutional neural networks (CNNs) in image analysis [48]. But by the end of the millennium, due to unmet overinflated expectations created by AI-exploiting ventures, and successes in other areas of machine learning, interest in ANNs waned for the second time.

Until the mid-2000s it was generally believed that deep ANNs are very hard to train. This perception started to change when it was shown that a particular type of multilayer ANN called a deep belief network (DBN), where each layer is a restricted Boltzmann machine (RBM), can be efficiently trained by greedy layer-wise unsupervised learning [49]. Soon after, based on the same principle, algorithms for training deep autoencoders (AEs) were proposed [50], as well as other deep architectures [33]. By this time, deep learning began to clearly outperform competing machine learning technologies for various data analysis tasks. This became most evident in the 2012 edition of the ImageNet challenge on image classification, where a CNN called AlexNet won by a large margin [51]. Since then, deep learning has gained ground at an exponential rate, including in the biomedical domain, as covered in later sections of this article.

2.2. Driving forces behind deep learning

In recent years, deep learning has been well recognized as a breakthrough technology. So much so that in 2018, the Turing Award, given annually since 1966 by the Association for Computing Machinery (ACM) and generally considered to be the "Nobel Prize of Computing", was awarded to three highly influential researchers "for conceptual and engineering breakthroughs that have made deep neural networks a critical component of computing": Yoshua Bengio (University of Montreal), Geoffrey Hinton (University of Toronto & Google), and Yann LeCun (New York University & Facebook) [52].

Apart from groundbreaking research, two other factors have played an important role in the relatively recent rapid rise of deep learning [53]. Both relate directly to the very needs of deep learning algorithms to be successful. The first is the need for large data sets to properly train the likewise large numbers of neural network parameters. Compiling such data sets has been greatly facilitated since the turn of the millennium by the increasing digitization of the world, leading to the present era of "big data". The second is the need for large computing power to complete the required large numbers of iterations in the training process within reasonable time. More and more advanced computing power has become affordable even for individual researchers in the form of general purpose graphics processing units (GPUs).

Together, these advances have enabled the development of ever deeper neural networks, reaching ever higher accuracies and beating the state of the art in an ever growing number of applications. The widespread usage of deep learning has been further accelerated by the development of open-access software libraries and frameworks [54–57], greatly facilitating deep neural network (DNN) design and training even for non-computer scientists. Tech giants such as Google, Facebook, Apple, IBM, Intel, Microsoft, Amazon, Baidu, and many others invest heavily in deep learning, capitalizing on its potential and contributing to a world that is increasingly driven by DNNs, and it seems this is only the beginning [58].

2.3. Widespread impact of deep learning

The extraordinary power of deep learning in addressing intractable challenges has led to a competitive race for leadership among research groups, universities, companies, and even nations [59]. Every week, new papers appear, not seldom by researchers without a solid background in computer science, commenting on the impact of deep learning in their field, or claiming victory with DNNs in yet another application domain, often simply by exploiting existing software tools and network architectures. The past few years have seen a flood of reviews and surveys on the subject, in virtually all fields of science, often by authors or in journals the seasoned computer scientist had never heard of. Apparently, despite many remaining challenges requiring further research (Section 5), a methodology has emerged that is relatively easy to use and that everyone is eager to own.

By now, deep learning has become the go-to data analysis technology in domains as diverse as agriculture [60], bioinformatics [61], biometrics [62], computational biology [63], consumer analytics [64], cyber security [65], dentistry [66], drug discovery [67], education [68], face recognition [69], gaming [70], health informatics [71], high-energy physics [72], hydrology [73], genomics [74], linguistics [75], mobile multimedia [76], mobile networking [77], multimedia analytics [78], nanotechnology [79], natural language processing [80], precision medicine [81], remote sensing [82], renewable energy forecasting [83], robotics [84],



Fig. 1. Explosive growth of the scientific literature on deep learning and related topics. The graph shows the number of publications per year in the past decade, having the terms deep learning (DL), machine learning (ML), or artificial intelligence (AI) in the title, according to Google Scholar (GS) and Web of Science (WOS) around the time of submission of this article.

smart manufacturing [85], speech generation [86], surveillance [87], traffic control [88], video coding [89], and countless others [90].

3. Deep learning in biomedical imaging

A domain we focus on more specifically in this article is biomedical imaging (Fig. 2). Here we take biomedical imaging to be the broad, multidisciplinary field concerned with the acquisition, processing, visualization, and interpretation of structural and functional images of living organisms, whether for clinical or for research purposes. Celebrating a long history of its own [91], including multiple Nobel Prize winning revolutions [92], biomedical imaging has become a cornerstone of modern healthcare and life sciences, to the extent that today "a world without imaging is clearly not imaginable" [93]. For the sake of brevity in this mini-review, we roughly divide the field into medical imaging, pathological imaging, preclinical imaging, and biological imaging in the life sciences, and summarize the impact of deep learning on each.

3.1. Deep learning in medical imaging

In clinical practice, the screening, diagnosis, prognosis, and treatment of disease in the human body, all rely increasingly on advanced medical imaging technologies such as X-ray computed tomography (CT), magnetic resonance imaging (MRI), positron emission tomography (PET), single-photon emission computed tomography (SPECT), and ultrasound (US) imaging. Successful application of imaging technologies involves not only highfidelity image acquisition but also reliable image interpretation [94]. While medical imaging devices have improved substantially in recent decades in terms of sensitivity, efficiency, and image quality, for a long time image interpretation was done primarily by humans. But even experts are known to suffer from subjectivity, variability, and fatigue. These impediments can potentially be overcome by computational methods, and deep learning in particular has emerged as a key enabling technology for this purpose, as attested by many recent overview articles in the field [95–102].

The impact of deep learning has been reviewed more specifically in a wide range of medical imaging areas, including abdominal imaging [103], atherosclerosis imaging [104], structural and functional brain imaging [105,106], in-vivo cancer imaging [107], dermatological imaging [108], endoscopy [109], mammography [110], musculoskeletal imaging [111], nuclear imaging [112], ophthalmology [113], pulmonary imaging [114], thoracic imaging [115], as well as in radiotherapy [116], interventional radiology [117], and radiology in general [118–120]. The massive body of papers on deep learning in virtually all areas of medical imaging has inspired many to write primers [121-123], guides [124-126], white papers or roadmaps [127-129], and other commentaries [130–132]. There is now growing evidence that deep learning methods can perform on par with, if not better than, radiologists in specific tasks [133], though the latter will continue to play a critical role in integrating such methods in clinical workflows [127].

3.2. Deep learning in pathological imaging

Disease diagnosis and prognosis cannot always be performed solely using structural or functional in-vivo medical imaging, but often also require complementary ex-vivo pathological imaging of tissue, cell, and body fluid samples extracted from the body. Perhaps even more than in medical imaging, visual image interpretation in pathology has traditionally been the task of human experts. However, the increasing adoption of digital whole-slide imaging (WSI) into routine clinical practice in recent years has created unprecedented opportunities for computer-aided diagnosis (CAD) in pathology [134–137]. Here, too, deep learning is being rapidly and widely adopted for this purpose, as reported in many reviews [138–145].

Pathological imaging plays a prominent role especially in cancer diagnosis and prognosis, and the impact of deep learning has been reviewed in various areas of oncological pathology, including in histopathology [141], cytopathology [146], and hematopathology [147]. Deep learning in pathology has been surveyed more specifically for breast cancer [142,148], lung cancer [149,150], tumor pathology in many other forms of cancer [151], and cancer prognosis [152], with many opinion articles commenting on challenges and opportunities [153–157]. As in medical imaging, there is mounting evidence for the potential of deep learning to provide



Fig. 2. Impact of deep learning on biomedical imaging. The graphs show the number of peer-reviewed journal and selected conference proceedings publications on deep learning in different biomedical application areas, categorized by imaging modality (top, see text for abbreviations) and subject of study (bottom), ranked from most to least popular. Numbers were estimated from the PubMed database of the US National Library of Medicine, National Institutes of Health, around the time of submission of this article, by searching for publications having relevant terms in the title or abstract (Supplementary Data).

fast and reliable image analysis at a performance level of a seasoned pathologist, or to serve as a synergistic tool for the latter to improve accuracy and throughput [131].

3.3. Deep learning in preclinical imaging

Innovative clinical medical imaging technologies and procedures are usually the fruit of preclinical imaging research with animal models representing humans in studying responses to physiological and environmental changes. Modern small-animal based anatomical, functional, and molecular imaging research involves a wide range of well-established as well as more experimental imaging modalities, including micro versions of clinical scanners (μ CT, μ MRI, μ PET, μ SPECT, μ US), optical coherence tomography (OCT), fluorescence molecular tomography (FMT), bioluminescence imaging (BLI), photoacoustic (PA) and thermoacoustic (TA) imaging, multispectral imaging (MSI), and others [93,158,159]. The use of deep learning for automated analysis of such imaging data is relatively uncharted territory, but recent studies have reported first applications in translational molecular imaging experiments [160–164].

3.4. Deep learning in biological imaging

Even more fundamental to our understanding of disease processes and the homeostatic mechanisms maintaining life down to the cellular and molecular levels, is biological microscopy imaging, more succinctly also referred to as bioimaging. Revolutionary scientific discoveries and technological innovations in the past decades have spurred the development of a vast array of advanced light microscopy (LM), notably fluorescence microscopy (FM), as well as electron microscopy (EM) and scanning probe microscopy (SPM) imaging modalities that have proven key to much of the progress in modern biological research [165–170].

Of all biomedical imaging fields, bioimaging arguably faces the biggest challenges in automating visual image interpretation tasks, due to the lack of standard imaging protocols, the high variability of experimental conditions, and the sheer volume of the data produced. Whereas (pre)clinical medical imaging systems typically generate data sets of dozens of megabytes (MB), and digital pathology scanners yield data sets of tens to hundreds of gigabytes (GB), automated microscopes may easily produce on the order of terabytes (TB) of image data in a single experiment [171–174]. Here, the power of deep learning is increasingly leveraged not only to improve image formation [175–179], but also subsequent image analysis, discussed next.

4. Deep learning for bioimage analysis

First studies using ANNs for bioimage analysis date back to the late 1980s [180], soon after the popularization of the back-propagation algorithm. A review on future trends in microscopy around that time already commented that for complex visual tasks "a good deal of faith is now placed in electronic neural networks" [181]. Indeed, the use of ANNs caught on during the 1990s [182–184] and 2000s [185–187], but as in biomedical imaging at large, deep learning began to be massively adopted for bioimage analysis only in recent years [188–194]. We briefly discuss some of the common tasks in bioimage analysis (Fig. 3) where deep learning has been particularly successful.

4.1. Deep learning for image enhancement

Many bioimage analysis tasks are greatly facilitated if the raw microscopy images are first enhanced by removing artifacts and restoring essential information as much as possible. Generally, a high signal-to-noise ratio (SNR) and spatial resolution are beneficial, but may not be achievable in a given experiment due to the required imaging speed and maximum allowable light exposure to avoid damaging the sample. Depending on the type of microscope used and the imaging conditions, different kinds of image enhancement operations may be applied, and deep learning has proven to be a powerful methodology for these. For instance, using well-registered pairs of low-quality and high-quality images, a CNN can be trained to perform denoising and recover resolution [175,179,200–204]. Similarly, trained with pairs of images from different imaging modalities, deep networks can predict fluorescent labels from transmitted-light microscopy images of unlabeled biological samples [179,195,205] (Fig. 4A), a technique referred to as cross-modality inference or transformation. Also, generative adversarial networks (GANs) have been shown to enable virtual refocusing of a two-dimensional (2D) fluorescence microscopy image onto a user-defined three-dimensional (3D) surface within a biological sample, correcting for sample drift, tilt, and other aberrations [178].

4.2. Deep learning for object detection

Another challenge central to many bioimage analysis tasks is to determine whether certain objects of interest are present in given microscopy images. Object detection often goes hand in hand with object localization and has been the subject of intense research for more than half a century [206]. The problem can be solved by extracting features from local image patches and performing classification on them. Here, too, traditional approaches have made way for deep learning in numerous applications, with two-stage region-proposal CNN-based (R-CNN) and unified vou-only-lookonce (YOLO) approaches and variants being most popular [207– 209]. In contrast with traditional object detection methods, which have found broad application in bioimage analysis for spotting intracellular particles [16,18,210,211], cell nuclei [17,26], and cellular events such as mitosis [212–214], deep learning approaches for these tasks have been explored since only recently. First results are promising [196,215-218] (Fig. 4B) but more extensive evaluations are needed to assess their general superiority.

4.3. Deep learning for image segmentation

One of the most ubiquitous tasks in bioimage analysis is the partitioning of images into meaningful segments for downstream quantification and statistical evaluation [17,19,26]. It is therefore no surprise that the bulk of literature on deep learning in many application areas of computer vision including bioimage analysis has focused on the potential for image segmentation [188,190,219-222]. Similar to object detection, image segmentation can be cast as a classification problem, this time down to the pixel level rather than the object level, which indeed is the approach taken by many deep-learning based methods. In particular, fully convolutional neural networks (FCNs) [223] such as U-Net [224], SegNet [225], DeepLab [226], and variants [227] (Fig. 4C) have become immensely popular for image segmentation. Deep learning methods have also begun to feature prominently in recent international competitions in bioimage analysis, including on segmentation of EM brain images [228], cell nuclei in FM images [229], cells in a variety of time-lapse microscopy images [23], and glandular structures in microscopy images of histological slides [230]. No doubt the future will see more and more deep-



Fig. 3. Common tasks in bioimage analysis. The ultimate goal is to gain knowledge of biological processes in health and disease by extracting relevant information from microscopy image or video recordings of these processes. Depending on the specific application, information extraction may involve image enhancement, object detection, image segmentation, object tracking, quantification, and classification, data visualization and analytics, and mathematical or statistical modeling. Deep learning is used increasingly in many of these tasks and we discuss several prominent ones in the main text. The diagram shows a typical order of tasks, with double-headed arrows indicating the possible interrelation and feedback between tasks, as well as the fact that any of them independently may also contribute to knowledge along the way, affecting other tasks. Modified from [6].

learning based methods dominating the charts in such evaluation studies.

4.4. Deep learning for object tracking

Characterizing real-life objects requires quantifying not only their spatial properties but also their temporal behavior. As advanced microscopes nowadays enable fully automated acquisition of time-lapse images of living cells and intracellular particles, this calls for robust computational methods capable of not only detecting and segmenting objects, but also tracking them over time in these images. Object tracking is generally considered to be "one of the most challenging computer vision problems" [231] and is a common task also in bioimage analysis [22,232–235]. The first international competition of particle tracking methods was held before deep learning broke through, but already led researchers to suggest the development of learning-based tracking methods [21]. Also, the continuing series of cell tracking challenges has seen the increasing use of deep learning methods for the problem [23]. As discussed in recent reviews, much of the work on deep learning for object tracking in bioimage analysis has focused on the spatial aspect of detection and segmentation [190,193], while the temporal aspect of data association and linking is typically still solved using traditional computer vision methods. First studies have appeared using DNNs to address both [236,237], as well as for subsequent trajectory analysis [198] (Fig. 4D), but the challenge remains to develop end-to-end deep-learning based cell and particle tracking methods [238].

4.5. Deep learning for object classification

The task of identifying images or objects therein as belonging to one of multiple predefined classes is a fundamental problem of computer vision in general [239–241] and a recurring theme also in bioimage analysis [242–244]. Traditionally, the problem has been addressed by extracting handcrafted image features and



Fig. 4. Examples of successful application of deep learning in bioimage analysis. **A**: Prediction of a fluorescence microscopy image (middle) from a bright-field microscopy image (left) compared to the truth (right) [195]. The image shows neurons in a culture of induced pluripotent stem cells differentiated toward the motor neuron lineage but containing other cell types as well. Fluorescent labels are TuJ1 (green) with Hoechst (blue) for the cell nuclei. The predicted image was obtained using a multiscale CNN inspired by U-Net. **B**: Detection of cells in various types of microscopy images [196]: Ki-67 stained bright-field microscopy image of neuroendocrine tumor tissue (top left), phase-contrast microscopy image of HeLa cervical cancer cells (top right), and H&E stained bright-field microscopy images of breast cancer tissue (bottom left) and human bone marrow tissue (bottom right). Detected cells are marked by yellow dots with green circles indicating the ground truth and were obtained using a structured regression model based on a fully residual CNN. C: Segmentation of neuronal axons (blue) and myelin sheaths (red) in a full scanning electron microscopy image slice of a rat spinal cord [197]. The segmentation was obtained using a CNN called AxonDeepSeg. **D**: Motion analysis of tracked breast cancer susceptibility gene BRCA2 particles in time-lapse fluorescence microscopy images [198]. Tracks were segmented into tracklets showing consistent motion (no switching between different dynamics states) using an LSTM network. Subsequent moment scaling spectrum (MSS) analysis of the tracklets yielded an estimate of the number of mobility classes (three in this case) and their associated parameters. Color coding indicates the value of the MSS slope per tracklet. **E**: Classification was done using a CNN called DeepLoc. A visualization (bottom) of the activations of the final convolutional layer of the network in 2D using t-distributed stochastic neighbor embedding (t-SNE) illustrates the power of the model to distinguis

using these together with given class labels to train classifiers such as support vector machines (SVM) or random forests (RF) [25,245,246]. But the capacity of deep CNN-based classifiers to learn relevant image features autonomously make them favorable over traditional approaches. Following their great success in the 2012 ImageNet challenge [51], CNN-based approaches have grown in popularity for image classification tasks across the board. In bioimage analysis, they have been shown to achieve expert-level performance in a wide range of cell classification and subcellular pattern recognition tasks [188,189,191] (Fig. 4E), although recent evaluations have revealed they do not necessarily outperform traditional approaches [244]. An issue in many studies is the lack of sufficient training data, which may be remedied by leveraging transfer learning [190] or crowd-sourcing strategies [247].

5. Summary and outlook

Deep learning has had a long history of discoveries, inventions, expectations, disappointments, rejections, revivals, successes, declines, recoveries, and breakthroughs, but is now widely accepted as the most powerful computing paradigm for big data analysis. The impact of deep learning on our daily lives is already unlike any other technology in the history of computer science, yet it seems we have seen only the proverbial tip of the iceberg. In biomedical imaging, DNNs are beginning to outperform human experts in a growing number of visual interpretation tasks, which is fueling fierce debates among professionals on the future ramifications for the field. Zeroing in on biological imaging, we have reviewed the use of deep learning approaches for common tasks in bioimage analysis, where they are now increasingly favored over traditional computer vision methods. Notwithstanding impressive achievements reported to date, many scientific and engineering challenges remain to further improve deep learning. In closing this mini-review, we touch on several important developments addressing these challenges that are relevant to bioimage analysis (see Table 1 for a quick summary of key research topics with references to reviews and commentaries for further reading).

5.1. Biological deep learning

Biology has always been a great source of inspiration for technology. Recognizing the unparalleled capacity of the brain in processing information, researchers in computer vision have exploited models of human vision from very early on [248,270,271]. Similarly, the idea of developing ANNs for data analysis was born out of research into the workings of biological neural networks (BNNs) [40,53,249]. In both cases, however, the ties between computer science and neuroscience have not remained strong, perhaps because "we simply do not have enough information about the brain to use it as a guide" [53]. But as long as human experts continue to be the gold standard in critical vision-based decision-making tasks, it seems there is still much to be gained from renewed interactions between the fields [40,248–250]. Bioimage analysis could play a pivotal role here, in a virtuous circle of helping to decipher BNNs at the microscopic level [272–274] and translating discoveries into improved ANNs for such studies [275–277].

5.2. Optimal deep learning

One of the key strengths of deep learning underlying its great success is that it automates the process of finding optimal feature descriptors given any data analysis task. While this eliminates the cumbersome handcrafting of descriptors, it leaves the user with the responsibility to design the right DNN architecture and tweak its hyperparameters to achieve satisfactory results. In practice, this may still require significant manual effort and yield suboptimal results. Notwithstanding the great arsenal of software toolkits for deep learning available today [54–57], there is still much room for the development of higher-level, user-friendly platforms that make it easier also for non-experts to adopt and use existing DNNs or to design and deploy their own solutions. The desire to further minimize human intervention in finding optimal solutions has given birth to the field of automated machine learning (AutoML). For deep learning, various neural architecture search (NAS) approaches have been proposed to automate the network engineering process [251,278,279]. First successful applications have recently been reported in medical imaging [252,280,281], suggesting NAS holds great potential also for bioimage analysis.

5.3. Economical deep learning

The most common form of deep learning is supervised learning, which requires input data with corresponding labeled output data. Especially in biomedical imaging applications, the output labels are typically obtained by expert manual annotation of the input data. However, as deep learning methods are notoriously data hungry, preparing a training data set this way can be extremely burdersome. Humans themselves largely learn in an unsupervised fashion, as they "discover the structure of the world by observing it, not by being told the name of every object" [35]. More economical deep learning approaches requiring less human input and/or training data are very much needed. Semi-supervised, weakly supervised, and unsupervised learning are important research topics [253,254,282] receiving increasing attention also in biomedical imaging [283–285]. An alternative approach is transfer learning between domains [255,278,286] which holds great promise for biomedical imaging as well [287–290]. Another strategy popular in bioimage analysis is to use high-fidelity simulated data as a surrogate for real data [256,291,292], which allows supervised learning with any number of images without requiring manual annotation [293-295].

5.4. Generalizable deep learning

In experimental evaluations of deep-learning methods, visual recognition tasks are typically framed as "closed set" problems, where the possible conditions in the test set are exactly the same as those in the training set. But in many applications, including in biomedical imaging, this is not very realistic. In practice, a more

Table 1

Overview of key reviews and commentaries for further reading on big research topi	ics
in deep learning (DL).	

Торіс	References
Biological DL	Neuro-inspired AI [40]
	Bio-inspired computer vision [248]
	Integrating DL and neuroscience [249]
	Biological vision and ANNs [250]
Optimal DL	User-friendly software platforms [56]
	Neural architecture search [251]
	AutoML in biomedical imaging [252]
Economical DL	Semi/weakly supervised learning [253]
	Unsupervised learning strategies [254]
	Transfer learning strategies [255]
	Data simulation and DL [256]
Generalizable DL	Open-set recognition [257]
	Domain adaptation [255]
Multimodal DL	Multimodal learning models [258]
	Data fusion strategies [259]
	Omics applications [260]
Efficient DL	Parallelization and distribution [261]
	Compression and acceleration [262]
	Biomedical imaging applications [263]
Explainable DL	Interpretable AI approaches [264]
	Visual analytics tools [265]
Responsible DL	On replacing radiologists [266]
	On replacing physicians [267]
	On replacing microscopists [268]
	Biomedical students on AI [269]

realistic scenario is that "incomplete knowledge of the world is present at training time, and unknown classes can be submitted to an algorithm during testing" [296]. This implies that current claims of superiority of machines over humans must be taken with a grain of salt, and that more generalizable or "open set" approaches to developing and evaluating deep-learning methods are needed. Open-set recognition (OSR) has been studied in the AI literature for some time [257,296,297] but has thus far received very little attention in biomedical imaging.

5.5. Multimodal deep learning

Nowadays, biomedical studies are hardly ever based on data from one imaging modality alone. Multiple, complementary imaging modalities are often used to obtain a more complete picture of the subject or sample under study. An example in bioimaging is the correlative recording of structural and functional image data, using electron and fluorescence microscopy, respectively [298–300]. But it does not stop there. Experiments typically also involve collecting genomic, proteomic, metabolomic, or other "omic" information [260,301,302], and in clinical studies additional data may come from electronic patient records. To take full advantage of all available information in such studies, powerful multimodal deep learning methods are required. This has been well recognized in various other fields [258,259,303] but deserves more attention in bioimaging and calls for an integrative approach to bioimage analysis and bioinformatics.

5.6. Efficient deep learning

The ever-growing volume of biomedical data sets and the increasing complexity of DNNs for improved analysis put proportionally higher demands on computing power. Training a deep network to achieve super-human performance, particularly in highly specialized domains such as biomedical imaging, essentially requires super-computing technology. To some extent this is provided by modern multicore GPUs, and more recent tensor processing units (TPUs), which enable single-machine parallelization. But more efficiency is often needed to finish network training within a desired time frame. Codesigning architectural, algorithmic, software, and hardware solutions to allow multi-machine parallelism and scalable distributed deep learning for this purpose is an ongoing engineering challenge [261,262,304]. Biomedical imaging at large will greatly benefit from such solutions, as they also facilitate exploiting data from multiple institutes in training DNNs without actually having to share the data, thus mitigating legal or ethical concerns [263,305,306].

5.7. Explainable deep learning

A major point of criticism for which even the pioneers of deep learning had their early papers rejected by peers in computer science, is that the use of neural networks for any given perceptual task provides "no insight into how to design a vision system" [51]. Even today, many in the community still have a propensity for carefully hand-designed solutions based on a solid understanding of the nature of the task. But the reality is that "methods that require careful hand-engineering by a programmer who understands the domain do not scale as well as methods that replace the programmer with a powerful general-purpose learning procedure" [51]. Nevertheless, the call for more explainability and interpretability of deep learning methods is legitimate and receiving growing attention in many areas of AI research [307,308,264] including computer vision [309-311]. A host of visual analytics tools have been developed to dissect DNNs and uncover what they have actually learned [265,312,313]. Such tools have not yet found widespread application in bioimage analysis but could help practitioners better understand the predictions made by network models.

5.8. Responsible deep learning

Ultimately, the goal of developing computational image analysis methods for the biomedical domain, from fundamental biological imaging to clinical medical imaging, is to improve the efficacy of healthcare. But in order for biomedical professionals to be willing to transfer their responsibilities to machines, and for those whose health depends on their care to accept such transition, these methods need to be trustworthy enough. In this regard it seems we have not quite reached the tipping point. In the past few years, the question whether or when AI will replace human experts has been pondered in many areas of biomedical imaging [132,266-269,314,315]. It goes without saying that decision making in biomedicine is more critical and risk-averse than in most other technological domains. Much work remains to take deep learning to the level of transparency, adaptability, creativity, empathy, and responsibility normally required of biomedical specialists. That said, as deep learning methods are already achieving humancompetitive performance in specific subtasks and have only just begun showing their considerable potential, DNNs will increasingly play an integral role in biomedical procedures. Historically, "human-machine collaborations have performed better than either one alone" [266], and there are no compelling reasons to believe this will ever change.

CRediT authorship contribution statement

Erik Meijering: Conceptualization, Data curation, Formal analysis, Investigation, Methodology, Resources, Visualization.

Declaration of Competing Interest

The author declares he has no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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Appendix A. Supplementary data

Supplementary data to this article can be found online at https://doi.org/10.1016/j.csbj.2020.08.003.

References

- Tolles WE. The Cytoanalyzer an example of physics in medical research. Trans New York Acad Sci 1955;17:250–6. <u>https://doi.org/10.1111/j.2164-0947.1955.tb01204.x</u>.
- [2] Preston K. Machine techniques for automatic leukocyte pattern analysis. Ann N Y Acad Sci 1962;97:482–90. <u>https://doi.org/10.1111/j.1749-6632.1962.</u> <u>tb34658.x.</u>
- [3] Ledley RS. High-speed automatic analysis of biomedical pictures. Science 1964;146:216–23. <u>https://doi.org/10.1126/science.146.3641.216</u>.
- [4] Murphy RF, Meijering E, Danuser G. Special issue on molecular and cellular bioimaging. IEEE Trans Image Processing 2005;14:1233-6. <u>https://doi.org/ 10.1109/TIP.2005.855701</u>.
- [5] Danuser G. Computer vision in cell biology. Cell 2011;147:973–8. <u>https://doi.org/10.1016/j.cell.2011.11.001</u>.
- [6] Meijering E, Carpenter AE, Peng H, Hamprecht FA, Olivo-Marin JC. Imagining the future of bioimage analysis. Nat Biotechnol 2016;34:1250–5. <u>https://doi.org/10.1038/nbt.3722</u>.
- [7] Prewitt JMS, Mendelsohn ML. The analysis of cell images. Ann N Y Acad Sci 1966;128:1035–53. <u>https://doi.org/10.1111/j.1749-6632.1965.tb11715.x</u>.
- [8] Lipkin LE, Watt WC, Kirsch RA. The analysis, synthesis, and description of biological images. Ann New York Acad Sci 1966;128:984–1012. <u>https://doi. org/10.1111/j.1749-6632.1965.tb11712.x</u>.
- [9] Ingram M, Preston K. Automatic analysis of blood cells. Scientific American 1970;223:72–82. <u>https://doi.org/10.1038/scientificamerican1170-72</u>.
- [10] Sarder P, Nehorai A. Deconvolution methods for 3-D fluorescence microscopy images. IEEE Signal Processing Magazine 2006;23:32–45. <u>https://doi.org/ 10.1109/MSP.2006.1628876.</u>
- [11] Roels J, Aelterman J, Luong HQ, Lippens S, Pizurica A, Saeys Y, et al. An overview of state-of-the-art image restoration in electron microscopy. J Microsc 2018;271:239–54. <u>https://doi.org/10.1111/jmi.12716</u>.
- [12] Meiniel W, Olivo-Marin JC, Angelini ED. Denoising of microscopy images: a review of the state-of-the-art, and a new sparsity-based method. IEEE Trans Image Processing 2018;27:3842–56. <u>https://doi.org/10.1109/TIP. 2018.2819821</u>.
- [13] Tsai CL, Lister JP, Bjornsson CS, Smith K, Shain W, Barnes CA, et al. Robust, globally consistent and fully automatic multi-image registration and montage synthesis for 3-D multi-channel images. J Microsc 2011;243:154–71. <u>https://doi.org/10.1111/j.1365-2818.2011.03489.x</u>.
- [14] Lucotte B, Balaban RS. Motion compensation for in vivo subcellular optical microscopy. J Microsc 2014;254:9–12. <u>https://doi.org/10.1111/jmi.12116</u>.
- [15] Qu L, Long F, Peng H. 3-D registration of biological images and models: registration of microscopic images and its uses in segmentation and annotation. IEEE Signal Processing Magazine 2015;32:70–7. <u>https://doi.org/ 10.1109/MSP.2014.2354060</u>.
- [16] Stěpka K, Matula P, Matula P, Worz S, Rohr K, Kozubek M. Performance and sensitivity evaluation of 3D spot detection methods in confocal microscopy. Cytometry Part A 2015;87:759–72. <u>https://doi.org/10.1002/cyto.a.22692</u>.
- [17] Xing F, Yang L. Robust nucleus/cell detection and segmentation in digital pathology and microscopy images: a comprehensive review. IEEE Rev Biomed Eng 2016;9:234–63. <u>https://doi.org/10.1109/RBME.2016.2515127</u>.
- [18] Mabaso MA, Withey DJ, Twala B. Spot detection methods in fluorescence microscopy imaging: a review. Image Anal Stereol 2018;37:173–90. <u>https:// doi.org/10.5566/ias.1690</u>.
- [19] Meijering E. Cell segmentation: 50 years down the road. IEEE Signal Process Mag 2012;29:140-5. <u>https://doi.org/10.1109/MSP.2012.2204190</u>.
- [20] Beneš M, Zitová B. Performance evaluation of image segmentation algorithms on microscopic image data. J Microsc 2014;257:65–85. <u>https://doi.org/ 10.1111/jmi.12186</u>.

- [21] Chenouard N, Smal I, de Chaumont F, Maska M, Sbalzarini IF, Gong Y, Cardinale J, Carthel C, Coraluppi S, Winter M, Cohen AR, Godinez WJ, Rohr K, Kalaidzidis Y, Liang L, Duncan J, Shen H, Xu Y, Magnusson KEG, Jaldén J, Blau HM, Paul-Gilloteaux P, Roudot P, Kervrann C, Waharte F, Tinevez JY, Shorte SL, Willemse J, Celler K, van Wezel GP, Dan HW, Tsai YS, Ortiz de Solórzano C, Olivo-Marin JC, Meijering E. Objective comparison of particle tracking methods. Nat Methods 2014;11:281–9, https://doi.org/10.1038/nmeth.2808.
- [22] Manzo C, Garcia-Parajo MF. A review of progress in single particle tracking: from methods to biophysical insights. Rep Progress Phys 2015;78. <u>https://doi.org/10.1088/0034-4885/78/12/124601</u>.
- [23] Ulman V, Maska M, Magnusson KEG, Ronneberger O, Haubold C, Harder N, Matula P, Matula P, Svoboda D, Radojevic M, Smal I, Rohr K, Jaldén J, Blau HM, Dzyubachyk O, Lelieveldt B, Xiao P, Li Y, Cho SY, Dufour AC, Olivo-Marin JC, Reyes-Aldasoro CC, Solis-Lemus JA, Bensch R, Brox T, Stegmaier J, Mikut R, Wolf S, Hamprecht FA, Esteves T, Quel-has P, Demirel O, Malmström L, Jug F, Tomancak P, Meijering E, Mun-oz-Barrutia A, Kozubek M, Ortiz-de Solorzano C. An objective comparison of cell-tracking algorithms. Nat Methods 2017;14:1141-52, https://doi.org/10.1038/nmeth.4473.
- [24] Orlov N, Shamir L, Macura T, Johnston J, Eckley DM, Goldberg IG. WND-CHARM: multi-purpose image classification using compound image transforms. Pattern Recogn Lett 2008;29:1684–93. <u>https://doi.org/10.1016/ j.patrec.2008.04.013</u>.
- [25] Shamir L, Delaney JD, Orlov N, Eckley DM, Goldberg IG. Pattern recognition software and techniques for biological image analysis. PLoS Comput Biol 2010;6:. <u>https://doi.org/10.1371/journal.pcbi.1000974</u>e1000974.
- [26] Irshad H, Veillard A, Roux L, Racoceanu D. Methods for nuclei detection, segmentation, and classification in digital histopathology: a review-current status and future potential. IEEE Rev Biomed Eng 2014;7:97–114. <u>https://doi.org/10.1109/RBME.2013.2295804</u>.
- [27] Hamilton N. Quantification and its applications in fluorescent microscopy imaging. Traffic 2009;10:951–61. <u>https://doi.org/10.1111/j.1600-0854.2009.00938.x.</u>
- [28] Eliceiri KW, Berthold MR, Goldberg IG, Ibáñez L, Manjunath BS, Martone ME, Murphy RF, Peng H, Plant AL, Roysam B, Stuurman N, Swedlow JR, Tomancak P, Carpenter AE. Biological imaging software tools. Nat Methods 2012;9:697–710. <u>https://doi.org/10.1038/nmeth.2084</u>.
- [29] Skylaki S, Hilsenbeck O, Schroeder T. Challenges in long-term imaging and quantification of single-cell dynamics. Nat Biotechnol 2016;34:1137-44. <u>https://doi.org/10.1038/nbt.3713</u>.
- [30] Walter T, Shattuck DW, Baldock R, Bastin ME, Carpenter AE, Duce S, et al. Visualization of image data from cells to organisms. Nat Methods 2010;7: S26-41. <u>https://doi.org/10.1038/nmeth.1431</u>.
- [31] Long F, Zhou J, Peng H. Visualization and analysis of 3D microscopic images. PLoS Comput Biol 2012;8:. <u>https://doi.org/10.1371/journal.pcbi.1002519</u>e1002519.
- [32] Sailem HZ, Cooper S, Bakal C. Visualizing quantitative microscopy data: history and challenges. Crit Rev Biochem Mol Biol 2016;51:96–101. <u>https:// doi.org/10.3109/10409238.2016.1146222</u>.
- [33] Bengio Y. Learning deep architectures for Al. Foundations Trends Machine Learning 2009;2:1–127. <u>https://doi.org/10.1561/220000006</u>.
- [34] Arel I, Rose DC, Karnowski TP. Deep machine learning a new frontier in artificial intelligence research. IEEE Computational Intelligence Magazine 2010;5:13–8. <u>https://doi.org/10.1109/MCI.2010.938364</u>.
- [35] LeCun Y, Bengio Y, Hinton G. Deep learning. Nature 2015;521:436–44. https://doi.org/10.1038/nature14539.
- [36] Serre T. Deep learning: the good, the bad, and the ugly. Ann Rev Vision Sci 2019;5:399–426. <u>https://doi.org/10.1146/annurev-vision-091718-014951</u>.
- [37] Marx V. Machine learning, practically speaking. Nat Methods 2019;16:463-7. https://doi.org/10.1038/s41592-019-0432-9.
- [38] Baltrusaitis T, Ahuja C, Morency LP. Multimodal machine learning: a survey and taxonomy. IEEE Trans Pattern Anal Mach Intell 2019;41:423–43. <u>https:// doi.org/10.1109/TPAML2018.2798607</u>.
- [39] Tecuci G. Artificial intelligence. Wiley Interdisciplinary Reviews: Computational Statistics 2012;4:168–80. <u>https://doi.org/10.1002/wics.200</u>.
- [40] Hassabis D, Kumaran D, Summerfield C, Botvinick M. Neuroscience-inspired artificial intelligence. Neuron 2017;95:245–58. <u>https://doi.org/10.1016/j.neuron.2017.06.011</u>.
- [41] Herna'ndez-Orallo J. Evaluation in artificial intelligence: from task-oriented to ability-oriented measurement. Artificial Intelligence Rev 2017;48:397–447.
- [42] Schmidhuber J. Deep learning in neural networks: an overview. Neural Networks 2015;61:85–117. <u>https://doi.org/10.1016/j.</u> <u>neunet.2014.09.003.2018.02.016</u>.
- [43] McCulloch WS, Pitts W. A logical calculus of the ideas immanent in nervous activity. Bull Mathematical Biophys 1943;5:115–33. <u>https://doi.org/10.1007/ BF02478259</u>.
- [44] Rosenblatt F. The perceptron: a probabilistic model for information storage and organization in the brain. Psychol Rev 1958;65:386–408. <u>https://doi.org/ 10.1037/h0042519</u>.
- [45] Minsky ML, Papert SA. Perceptrons: An Introduction to Computational Geometry. Cambridge, MA, USA: The MIT Press; 1969.
- [46] Rumelhart DE, Hinton GE, Williams RJ. Learning representations by backpropagating errors. Nature 1986;323:533–6. <u>https://doi.org/10.1038/ 323533a0</u>.

- [47] Hochreiter S, Schmidhuber J. Long short-term memory. Neural Computation 1997;9:1735–80. <u>https://doi.org/10.1162/neco.1997.9.8.1735</u>.
- [48] Lecun Y, Bottou L, Bengio Y, Haffner P. Gradient-based learning applied to document recognition. Proc IEEE 1998;86:2278–324. <u>https://doi.org/ 10.1109/5.726791.</u>
- [49] Hinton GE, Osindero S, Teh YW. A fast learning algorithm for deep belief nets. Neural Comput 2006;18:1527–54. <u>https://doi.org/10.1162/neco.2006.18.7.1527.</u>
- [50] Hinton GE, Salakhutdinov RR. Reducing the dimensionality of data with neural networks. Science 2006;313:504–7. <u>https://doi.org/10.1126/ science.1127647</u>.
- [51] Krizhevsky A, Sutskever I, Hinton GE. ImageNet classification with deep convolutional neural networks. Commun ACM 2017;60:84–90. <u>https://doi.org/10.1145/3065386</u>.
- [52] Savage N. Neural net worth. Commun ACM 2019;62:10–2. <u>https://doi.org/ 10.1145/3323872</u>.
- [53] Goodfellow I, Bengio Y, Courville A. Deep Learning. The MIT Press, Cambridge, MA, USA, 2016. https://www.deeplearningbook.org/.
- [54] Erickson BJ, Korfiatis P, Akkus Z, Kline T, Philbrick K. Toolkits and libraries for deep learning. J Digital Imaging 2017;30:400-5. <u>https://doi.org/10.1007/ s10278-017-9965-6</u>.
- [55] Pouyanfar S, Sadiq S, Yan Y, Tian H, Tao Y, Reyes MP, et al. A survey on deep learning: algorithms, techniques, and applications. ACM Comput Surv 2018;51:92. <u>https://doi.org/10.1145/3234150</u>.
- [56] Nguyen G, Dlugolinsky S, Bobák M, Tran V, Garćia A.L., Here-dia I, Maĺik P, Hluchý L. Machine learning and deep learning frameworks and libraries for large-scale data mining: a survey. Artificial Intelligence Rev 2019;52:77–124, https://doi.org/10.1007/s10462-018-09679-z.
- [57] Shrestha A, Mahmood A. Review of deep learning algorithms and architectures. IEEE Access 2019;7:53040–65. <u>https://doi.org/10.1109/</u> <u>ACCESS.2019.2912200.</u>
- [58] Sejnowski TJ. The Deep Learning Revolution. The MIT Press, Cambridge, MA, USA, 2018. https://www.amazon.com/dp/026203803X.
- [59] Williams MA. The artificial intelligence race: will Australia lead or lose?. J Proc Royal Soc New South Wales 2019;152:105–14. https://royalsoc.org.au/ council-members-section/435-v152-11007/s10462-016-9505-7.
- [60] Kamilaris A, Prenafeta-Boldú FX. Deep learning in agriculture: a survey. Computers and Electronics in Agriculture2018;147:70–90. https://doi.org/10. 1016/j.compag.
- [61] Min S, Lee B, Yoon S. Deep learning in bioinformatics. Briefings in Bioinformatics 2017;18:851–69. <u>https://doi.org/10.1093/bib/bbw068</u>.
- [62] Sundararajan K, Woodard DL. Deep learning for biometrics: a survey. ACM Computing Surveys 2018;51:65. <u>https://doi.org/10.1145/3190618</u>.
- [63] Angermueller C, Prnamaa T, Parts L, Stegle O. Deep learning for computational biology. Molecular Syst Biol 2016;12:878. <u>https://doi.org/10.15252/ msb.20156651</u>.
- [64] Zhang S, Yao L, Sun A, Tay Y. Deep learning based recommender system: a survey and new perspectives. ACM Comput Surv 2019;52:5. <u>https://doi.org/ 10.1145/3285029</u>.
- [65] Berman DS, Buczak AL, Chavis JS, Corbett CL. A survey of deep learning methods for cyber security. Information 2019;10:122. <u>https://doi.org/ 10.3390/info10040122</u>.
- [66] Hwang J, Jung YH, Cho BH, Heo MS. An overview of deep learning in the field of dentistry. Imaging Sci Dentistry 2019;49:1–7. <u>https://doi.org/10.5624/ isd.2019.49.1.1.</u>
- [67] Gawehn E, Hiss JA, Schneider G. Deep learning in drug discovery. Molecular Informatics 2016;35:3–14. <u>https://doi.org/10.1002/minf.201501008</u>.
- [68] Hernández-Blanco A, Herrera-Flores B, Tomás D, Navarro-Colorado B. A systematic review of deep learning approaches to educational data mining. Complexity 2019;2019:1306039. <u>https://doi.org/10.1155/2019/1306039</u>.
- [69] Guo G, Zhang N. A survey on deep learning based face recognition. Comp Vision Image Understanding 2019;189:. <u>https://doi.org/10.1016/j. cviu.2019.102805</u>102805.
- [70] Justesen N, Bontrager P, Togelius J, Risi S. Deep learning for video game playing. IEEE Trans Games 2020;12:1–20. <u>https://doi.org/10.1109/</u> TG.2019.2896986.
- [71] Ravi D, Wong C, Deligianni F, Berthelot M, Andreu-Perez J, Lo B, et al. Deep learning for health informatics. IEEE J Biomed Health Inf 2017;21:4–21. https://doi.org/10.1109/IBHI.2016.2636665.
- [72] Abdughani M, Ren J, Wu L, Yang JM, Zhao J. Supervised deep learning in high energy phenomenology: a mini review. Commun Theor Phys 2019;71:955. https://doi.org/10.1088/0253-6102/71/8/955.
- [73] Shen C. A transdisciplinary review of deep learning research and its relevance for water resources scientists. Water Resources Res 2018;54:8558–93. https://doi.org/10.1029/2018WR022643.
- [74] Eraslan G, Avsec Z, Gagneur J, Theis FJ. Deep learning: new computational modelling techniques for genomics. Nat Rev Genetics 2019;20:389–403. https://doi.org/10.1038/s41576-019-0122-6.
- [75] Monroe D. Deep learning takes on translation. Commun ACM 2017;60:12–4. https://doi.org/10.1145/3077229.
- [76] Ota K, Dao MS, Mezaris V, De Natale FGB. Deep learning for mobile multimedia: a survey. ACM Trans Multimedia Comput Commun Appl 2017;13:34. <u>https://doi.org/10.1145/3092831</u>.
- [77] Zhang C, Patras P, Haddadi H. Deep learning in mobile and wireless networking: a survey. IEEE Commun Surveys Tutorials 2019;21:2224–87. <u>https://doi.org/10.1109/COMST.2019.2904897</u>.

- [78] Zhang W, Yao T, Zhu S. Deep learning-based multimedia analytics: a review. ACM Trans Multimedia Comp Commun Appl 2019;15:2. <u>https://doi.org/ 10.1145/3279952</u>.
- [79] Sacha GM, Varona P. Artificial intelligence in nanotechnology. Nanotechnology 2013;24. <u>https://doi.org/10.1088/0957-4484/24/45/452002</u>.
- [80] Young T, Hazarika D, Poria S, Cambria E. Recent trends in deep learning based natural language processing. IEEE Comput Intelligence Magazine 2018;13:55–75. <u>https://doi.org/10.1109/MCI.2018.2840738</u>.
- [81] Parekh VS, Jacobs MA. Deep learning and radiomics in precision medicine. Expert Rev Precision Med Drug Devel 2019;4:59–72. <u>https://doi.org/10.1080/23808993.2019.1585805</u>.
- [82] Zhu XX, Tuia D, Mou L, Xia GS, Zhang L, Xu F, et al. Deep learning in remote sensing: a comprehensive review and list of resources. IEEE Geosci Remote Sens Mag 2017;5:8–36. <u>https://doi.org/10.1109/MGRS.2017.2762307</u>.
- [83] Wang H, Lei Z, Zhang X, Zhou B, Peng J. A review of deep learning for renewable energy forecasting. Energy Conversion Manag 2019;198. <u>https:// doi.org/10.1016/j.enconman.2019.111799</u>.
- [84] Pierson HA, Gashler MS. Deep learning in robotics: a review of recent research. Adv Robotics 2017;31:821–35. <u>https://doi.org/10.1080/</u> 01691864.2017.1365009.
- [85] Wang J, Ma Y, Zhang L, Gao RX, Wu D. Deep learning for smart manufacturing: methods and applications. J Manuf Syst 2018;48:144–56. <u>https://doi.org/10.1016/j.imsy.2018.01.003</u>.
- [86] Ling ZH, Kang SY, Zen H, Senior A, Schuster M, Qian XJ, et al. Deep learning for acoustic modeling in parametric speech generation: a systematic review of existing techniques and future trends. IEEE Signal Processing Magazine 2015;32:35–52. <u>https://doi.org/10.1109/MSP.2014.2359987</u>.
- [87] Sreenu G, Durai MAS. Intelligent video surveillance: a review through deep learning techniques for crowd analysis. J Big Data 2019;6:48. <u>https://doi.org/ 10.1186/s40537-019-0212-5</u>.
- [88] Nguyen H, Kieu LM, Wen T, Cai C. Deep learning methods in transportation domain: a review. IET Intelligent Trans Syst 2018;12:998–1004. <u>https://doi.org/10.1049/iet-its.2018.0064</u>.
- [89] Liu D, Li Y, Lin J, Li H, Wu F. Deep learning-based video coding: a review and a case study. ACM Comput Surv 2020;53:11. <u>https://doi.org/10.1145/3368405</u>.
- [90] Khamparia A, Singh KM. A systematic review on deep learning architectures and applications. Expert Syst 2019;36:. <u>https://doi.org/10.1111/ exsy.12400</u>e12400.
- [91] Robb RA. Biomedical imaging: past, present and predictions. Med Imaging Tech 2006;24:25–37. <u>https://doi.org/10.11409/mit.24.25</u>.
- [92] Wallyn J, Anton N, Akram S, Vandamme TF. Biomedical imaging: principles, technologies, clinical aspects, contrast agents, limitations and future trends in nanomedicines. Pharmaceutical Res 2019;36:78. <u>https://doi.org/10.1007/ s11095-019-2608-5</u>.
- [93] Weissleder R, Nahrendorf M. Advancing biomedical imaging. Proc National Acad Sci United States of America 2015;112:14424–8. <u>https://doi.org/ 10.1073/pnas.1508524112</u>.
- [94] Greenspan H, van Ginneken B, Summers RM. Deep learning in medical imaging: overview and future promise of an exciting new technique. IEEE Trans Medical Imaging 2016;35:1153–9. <u>https://doi.org/10.1109/</u> TML2016.2553401.
- [95] Litjens G, Kooi T, Bejnordi BE, Setio AAA, Ciompi F, Ghafoorian M, van der Laak JAWM, van Ginneken B, Sánchez CI. A survey on deep learning in medical image analysis. Med Image Anal 2017;42:60–88. <u>https://doi.org/10.1016/j.</u> media.2017.07.005.
- [96] Shen D, Wu G, Suk HI. Deep learning in medical image analysis. Ann Rev Biomed Eng 2017;19:221–48. <u>https://doi.org/10.1146/annurev-bioeng-071516-044442</u>.
- [97] Suzuki K. Overview of deep learning in medical imaging. Radiol Phys Tech 2017;10:257–73. <u>https://doi.org/10.1007/s12194-017-0406-5</u>.
- [98] Lee JG, Jun S, Cho YW, Lee H, Kim GB, Seo JB, et al. Deep learning in medical imaging: general overview. Korean J Radiol 2017;18:570–84. <u>https://doi.org/ 10.3348/kir.2017.18.4.570</u>.
- [99] Ker J, Wang L, Rao J, Lim T. Deep learning applications in medical image analysis. IEEE Access 2018;6:9375–89. <u>https://doi.org/10.1109/</u> ACCESS.2017.2788044.
- [100] Biswas M, Kuppili V, Saba L, Edla DR, Suri HS, Cuadrado-Godia E, Laird JR, Marinhoe RT, Sanches JM, Nicolaides A, Suri JS. State-of-the-art review on deep learning in medical imaging. Front Biosci 2019;24:392–426. <u>https://doi. org/10.2741/4725</u>.
- [101] Kaji S, Kida S. Overview of image-to-image translation by use of deep neural networks: denoising, super-resolution, modality conversion, and reconstruction in medical imaging. Radiol Phys Tech 2019;12:235–48. https://doi.org/10.1007/s12194-019-00520-y.
- [102] Chan HP, Samala RK, Hadjiiski LM, Zhou C. Deep learning in medical image analysis. Adv Exp Med Biol 2020;1213:3–21. <u>https://doi.org/10.1007/978-3-030-33128-3_1</u>.
- [103] Brattain LJ, Telfer BA, Dhyani M, Grajo JR, Samir AE. Machine learning for medical ultrasound: status, methods, and future opportunities. Abdominal Radiol 2018;43:786–99. <u>https://doi.org/10.1007/s00261-018-1517-0</u>.
- [104] Kolossváry M, De Cecco CN, Feuchtner G, Maurovich-Horvat P. Advanced atherosclerosis imaging by CT: radiomics, machine learning and deep learning. J Cardiovascular Computed Tomography 2019;13:274–80. <u>https:// doi.org/10.1016/i.jcct.2019.04.007</u>.
- [105] Nadeem MW, Ghamdi MAA, Hussain M, Khan MA, Khan KM, Almotiri SH, Butt SA. Brain tumor analysis empowered with deep learning: a review,

taxonomy, and future challenges. Brain Sci 2020;10:118. <u>https://doi.org/</u> 10.3390/brainsci10020118.

- [106] Zhu G, Jiang B, Tong L, Xie Y, Zaharchuk G, Wintermark M. Applications of deep learning to neuro-imaging techniques. Front Neurol 2019;10:869. <u>https://doi.org/10.3389/fneur.2019.00869</u>.
- [107] Munir K, Elahi H, Ayub A, Frezza F, Rizzi A. Cancer diagnosis using deep learning: a bibliographic review. Cancers 2019;11:1235. <u>https://doi.org/ 10.3390/cancers11091235</u>.
- [108] Chan S, Reddy V, Myers B, Thibodeaux Q, Brownstone N, Liao W. Machine learning in dermatology: current applications, opportunities, and limitations. Dermatol Therapy 2020;10:365–86, https://doi.org/10.1007/s13555-020-00372-0.
- [109] Min JK, Kwak MS, Cha JM. Overview of deep learning in gastrointestinal endoscopy. Gut and Liver 2019;13:388–93. <u>https://doi.org/10.5009/ gnl18384</u>.
- [110] Zou L, Yu S, Meng T, Zhang Z, Liang X, Xie Y. A technical review of convolutional neural network-based mammographic breast cancer diagnosis. Comput Math Methods Med 2019;2019:6509357. <u>https://doi.org/10.1155/ 2019/6509357</u>.
- [111] Kijowski R, Liu F, Caliva F, Pedoia V. Deep learning for lesion detection, progression, and prediction of musculoskeletal disease. J Magnetic Resonance Imaging 2020;51:Forthcoming, https://doi.org/10.1002/jmri.27001.
- [112] Currie GM. Intelligent imaging: artificial intelligence augmented nuclear medicine. J Nuclear Medicine Tech 2019;47:217-22. <u>https://doi.org/10.2967/ inmt.119.232462.</u>
- [113] Ting DSW, Peng L, Varadarajan AV, Keane PA, Burlina PM, Chiang MF, Schmetterer L, Pasquale LR, Bressler NM, Webster DR, Abramoff M, Wong TY. Deep learning in ophthalmology: the technical and clinical considerations. Progress Retinal Eye Res 2019;72. <u>https://doi.org/10.1016/j. preteyeres.2019.04.003</u>.
- [114] Ma J, Song Y, Tian X, Hua Y, Zhang R, Wu J. Survey on deep learning for pulmonary medical imaging. Front Med 2020;14:Forthcoming, https://doi. org/10.1007/s11684-019-0726-4.
- [115] Chassagnon G, Vakalopolou M, Paragios N, Revel MP. Deep learning: definition and perspectives for thoracic imaging. Eur Radiol 2020;30:2021–30. <u>https://doi.org/10.1007/s00330-019-06564-3</u>.
- [116] Meyer P, Noblet V, Mazzara C, Lallement A. Survey on deep learning for radiotherapy. Comput Biol Med 2018;98:126–46. <u>https://doi.org/10.1016/j. compbiomed.2018.05.018</u>.
- [117] Chassagnon G, Vakalopolou M, Paragios N, Revel MP. Deep learning: definition and perspectives for thoracic imaging. Eur Radiol 2020;30:2021–30. <u>https://doi.org/10.1007/s00330-019-06564-3</u>.
- [118] McBee MP, Awan OA, Colucci AT, Ghobadi CW, Kadom N, Kansagra AP, et al. Deep learning in radiology. Acad Radiol 2018;25:1472–80. <u>https://doi.org/ 10.1016/j.acra.2018.02.018</u>.
- [119] Yasaka K, Abe O. Deep learning and artificial intelligence in radiology: current applications and future directions. PLoS Med 2018;15:e1002707. <u>https://doi. org/10.1371/iournal.pmed</u>.
- [120] Hosny A, Parmar C, Quackenbush J, Schwartz LH, Aerts HJWL. Artificial intelligence in radiology. Nat Rev Cancer 2018;18:500–10. <u>https://doi.org/</u> 10.1038/s41568-018-0016-5.
- [121] Chartrand G, Cheng PM, Vorontsov E, Drozdzal M, Turcotte S, Pal CJ, Kadoury S, Tang A. Deep learning: a primer for radiologists. Radiographics 2017;37:2113–31. <u>https://doi.org/10.1148/rg.2017170077</u>.
- [122] Montagnon E, Cerny M, Cadrin-Chênevert A, Hamilton V, Derennes T, Ilinca A, Vandenbroucke-Menu F, Turcotte S, Kadoury S, Tang A. Deep learning workflow in radiology: a primer. Insights Into Imaging 2020;11:22. <u>https:// doi.org/10.1186/s13244-019-0832-5</u>.
- [123] Do S, Song KD, Chung JW. Basics of deep learning: a radiologist's guide to understanding published radiology articles on deep learning. Korean J Radiol 2020;21:33–41. <u>https://doi.org/10.3348/kjr.2019.0312</u>.
- [124] England JR, Cheng PM. Artificial intelligence for medical image analysis: a guide for authors and reviewers. Am J Roentgenol 2019;212:513–9. <u>https:// doi.org/10.2214/AJR.18.20490</u>.
- [125] Soffer S, Ben-Cohen A, Shimon O, Amitai MM, Greenspan H, Klang E. Convolutional neural networks for radiologic images: a radiologist's guide. Radiology 2019;290:590–606. <u>https://doi.org/10.1148/radiol.2018</u> 180547.
- [126] Faes L, Liu X, Wagner SK, Fu DJ, Balaskas K, Sim DA, Bachmann LM, Keane PA, Denniston AK. A clinician's guide to artificial intelligence: how to critically appraise machine learning studies. Transl Vision Sci Tech 2020;9:7. <u>https:// doi.org/10.1167/tvst.9.2.7</u>.
- [127] Tang A, Tam R, Cadrin-Chênevert A, Guest W, Chong J, Barfett J, Chepelev L, Cairns R, Mitchell JR, Cicero MD, Poudrette MG, Jaremko JL, Reinhold C, Gallix B, Gray B, Geis R. Canadian Association of Radiologists white paper on artificial intelligence in radiology. Canadian Assoc Radiologists J 2018;69:120–35. https://doi.org/10.1016/i.carj.2018.02.002.
- [128] Langlotz CP, Allen B, Erickson BJ, Kalpathy-Cramer J, Bigelow K, Cook TS, et al. A roadmap for foundational research on artificial intelligence in medical imaging. Radiology 2019;291:781–91. <u>https://doi.org/10.1148/</u> radiol.2019190613.
- [129] European Society of Radiology (ESR). What the radiologist should know about artificial intelligence – an ESR white paper. Insights Into Imaging 2019;10:44. https://doi.org/10.1186/s13244-019-0738-2.
- [130] Saba L, Biswas M, Kuppili V, Cuadrado Godia E, Suri HS, Edla DR, Omerzu T, Laird JR, Khanna NN, Mavrogeni S, Protogerou A, Sfikakis PP, Viswanathan V,

Kitas GD, Nicolaides A, Gupta A, Suri JS. The present and future of deep learning in radiology. Eur J Radiol 2019;114:14–24. <u>https://doi.org/10.1016/j.eirad.2019.02.038</u>.

- [131] Kulkarni S, Seneviratne N, Baig MS, Khan AHA. Artificial intelligence in medicine: where are we now?. Acad Radiol 2020;27:62–70. <u>https://doi.org/ 10.1016/j.acra.2019.10.001</u>.
- [132] Pesapane F, Tantrige P, Patella F, Biondetti P, Nicosia L, Ianniello A, Rossi UG, Carrafiello G, Ierardi AM. Myths and facts about artificial intelligence: why machine- and deep-learning will not replace interventional radiologists. Med Oncol 2020;37:40. <u>https://doi.org/10.1007/s12032-020-01368-8</u>.
- [133] Shen J, Zhang CJP, Jiang B, Chen J, Song J, Liu Z, et al. Artificial intelligence versus clinicians in disease diagnosis: systematic review. JMIR Med Inforatics 2019;7:e10010. <u>https://doi.org/10.2196/10010</u>.
- [134] Bhargava R, Madabhushi A. Emerging themes in image informatics and molecular analysis for digital pathology. Annu Rev Biomed Eng 2016;18:387–412. <u>https://doi.org/10.1146/annurev-bioeng-112415-114722</u>.
- [135] Pantanowitz L, Sharma A, Carter AB, Kurc T, Sussman A. Saltz whole slide image analysis: an overview. Front Med 2019;6:264. <u>https://doi.org/10.3389/ fmed.2019.00264</u>.
- [136] Aeffner F, Zarella MD, Buchbinder N, Bui MM, Goodman MR, Hartman DJ, Lujan GM, Molani MA, Parwani AV, Lillard K, Turner OC, Vemuri VNP, Yuil-Valdes AG, Bowman D. Introduction to digital image analysis in whole-slide imaging: a white paper from the digital pathology association. J Pathol Informatics 2019;10:9. <u>https://doi.org/10.4103/jpi.jpi 82_18</u>.
- [137] Nam S, Chong Y, Jung CK, Kwak TY, Lee JY, Park J, Rho MJ, Go H. Introduction to digital pathology and computer-aided pathology. J Pathol Trans Med 2020;54:125–34. <u>https://doi.org/10.4132/iptm.2019.12.31</u>.
- [138] Janowczyk A, Madabhushi A. Deep learning for digital pathology image analysis: a comprehensive tutorial with selected use cases. J Pathol Informatics 2016;7:29. <u>https://doi.org/10.4103/2153-3539.186902</u>.
- [139] Madabhushi A, Lee G. Image analysis and machine learning in digital pathology: challenges and opportunities. Med Image Anal 2016;33:170–5. <u>https://doi.org/10.1016/j.media.2016.06.037</u>.
- [140] Zhong C, Han J, Borowsky A, Parvin B, Wang Y, Chang H. When machine vision meets histology: a comparative evaluation of model architecture for classification of histology sections. Med Image Anal 2017;35:530–43. <u>https:// doi.org/10.1016/j.media.2016.08.010</u>.
- [141] Komura D, Ishikawa S. Machine learning methods for histopathological image analysis. Computational Struct Biotech J 2018;16:34–42. <u>https://doi.org/10.1016/j.csbj.2018.01.001</u>.
- [142] Hamidinekoo A, Denton E, Rampun A, Honnor K, Zwiggelaar R. Deep learning in mammography and breast histology, an overview and future trends. Med Image Anal 2018;47:45–67. <u>https://doi.org/10.1016/j.media.2018.03.006</u>.
- [143] Wang S, Yang DM, Rong R, Zhan X, Xiao G. Pathology image analysis using segmentation deep learning algorithms. Am J Pathol 2019;189:1686–98. <u>https://doi.org/10.1016/j.ajpath.2019.05.007</u>.
- [144] Serag A, Ion-Margineanu A, Qureshi H, McMillan R, Saint Martin MJ, Diamond J, et al. Translational AI and deep learning in diagnostic pathology. Front Med 2019;6:185. <u>https://doi.org/10.3389/fmed.2019.00185</u>.
- [145] Dimitriou N, Arandjelovic J. Twenty years of digital pathology: an overview of the road travelled, what is on the horizon, and the emergence of vendorneutral archives. J Pathol Informatics 2018;9:40. <u>https://doi.org/10.4103/jpi. jpi 69 18</u>.
- [146] Landau MS, Pantanowitz L. Artificial intelligence in cytopathology: a review of the literature and overview of commercial landscape. J Am Soc Cytopathol 2019;8:230–41. <u>https://doi.org/10.1016/j.jasc.2019.03.003</u>.
- [147] El Achi H, Khoury JD. Artificial intelligence and digital microscopy applications in diagnostic hematopathology. Cancers 2020;12:797. <u>https:// doi.org/10.3390/cancers12040797</u>.
- [148] Saxena S, Gyanchandani M. Machine learning methods for computer-aided breast cancer diagnosis using histopathology: a narrative review. J Med Imaging Radiation Sci 2020;51:182–93. <u>https://doi.org/10.1016/j. jmir.2019.11.001</u>.
- [149] Wang S, Yang DM, Rong R, Zhan X, Fujimoto J, Liu H, et al. Artificial intelligence in lung cancer pathology image analysis. Cancers 2019;11:1673. <u>https://doi.org/10.3390/cancers11111673</u>.
- [150] Cong L, Feng W, Yao Z, Zhou X, Xiao W. Deep learning model as a new trend in computer-aided diagnosis of tumor pathology for lung cancer. J Cancer 2020;11:3615–22. <u>https://doi.org/10.7150/ica.43268</u>.
- [151] Jiang Y, Yang M, Wang S, Li X, Sun Y. Emerging role of deep learning-based artificial intelligence in tumor pathology. Cancer Commun 2020;40:154–66. https://doi.org/10.1002/cac2.12012.
- [152] Zhu W, Xie L, Han J, Guo X. The application of deep learning in cancer prognosis prediction. Cancers 2020;12:603. <u>https://doi.org/10.3390/ cancers12030603</u>.
- [153] Tizhoosh HR, Pantanowitz L. Artificial intelligence and digital pathology: challenges and opportunities. J Pathol Informatics 2018;9:38. <u>https://doi.org/ 10.4103/jpi.jpi 53 18</u>.
- [154] Niazi MKK, Parwani AV, Gurcan MN. Digital pathology and artificial intelligence. Lancet Oncol 2019;20:e253–61. <u>https://doi.org/10.1016/S1470-2045(19) 30154-8</u>.
- [155] Bera K, Schalper KA, Rimm DL, Velcheti V, Madabhushi A. Artificial intelligence in digital pathology – new tools for diagnosis and precision oncology. Nat Rev Clin Oncol 2019;16:703–15. <u>https://doi.org/10.1038/ s41571-019-0252-v</u>.

- [156] Colling R, Pitman H, Oien K, Rajpoot N, Macklin P, in Histopathology Working Group CMPAI, Snead D, Sackville T, Verrill C. Artificial intelligence in digital pathology: a roadmap to routine use in clinical practice. J Pathol 2019;249:143–50, https://doi.org/10.1002/path.5310.
- [157] Acs B, Rantalainen M, Hartman J. Artificial intelligence as the next step towards precision pathology. J Internal Med 2020;288:62–81. <u>https://doi.org/ 10.1111/joim.13030</u>.
- [158] Stout DB, Zaidi H. Preclinical multimodality imaging in vivo. PET Clinics 2008;3:251–73. <u>https://doi.org/10.1016/j.cpet.2009.03.001</u>.
- [159] Alam IS, Steinberg I, Vermesh Ö, van den Berg NS, Rosenthal EL, van Dam GM, et al. Emerging intraoperative imaging modalities to improve surgical precision. Mol Imag Biol 2018;20:705–15. <u>https://doi.org/10.1007/s11307-018-1227-6.</u>
- [160] Xue Y, Chen S, Qin J, Liu Y, Huang B, Chen H. Application of deep learning in automated analysis of molecular images in cancer: a survey. Contrast Media Molecular Imaging 2017;2017:9512370. <u>https://doi.org/10.1155/2017/ 9512370</u>.
- [161] Choi H. Deep learning in nuclear medicine and molecular imaging: current perspectives and future directions. Nuclear Med Molecular Imaging 2018;52:109–18. <u>https://doi.org/10.1007/s13139-017-0504-7</u>.
- [162] Cook GJR, Goh V. What can artificial intelligence teach us about the molecular mechanisms underlying disease?. Eur J Nucl Med Mol Imaging 2019;46:2715–21. <u>https://doi.org/10.1007/s00259-019-04370-z</u>.
- [163] Rogers MA, Aikawa E. Cardiovascular calcification: artificial intelligence and big data accelerate mechanistic discovery. Nat Rev Cardiol 2019;16:261–74. <u>https://doi.org/10.1038/s41569-018-0123-8</u>.
- [164] Vaidya T, Agrawal A, Mahajan S, Thakur MH, Mahajan A. The continuing evolution of molecular functional imaging in clinical oncology: the road to precision medicine and radiogenomics. Molecular Diagnosis Therapy 2019;23:1–51. <u>https://doi.org/10.1007/s40291-018-0366-4</u>.
- [165] Ji N, Shroff H, Zhong H, Betzig E. Advances in the speed and resolution of light microscopy. Curr Opinion Neurobiol 2008;18:605-16. <u>https://doi.org/ 10.1016/j.conb.2009.03.009</u>.
- [166] Moerner WE. Microscopy beyond the diffraction limit using actively controlled single molecules. Jo Microsc 2012;246:213–20. <u>https://doi.org/ 10.1111/j.1365-2818.2012.03600.x</u>.
- [167] Timmermans FJ, Otto C. Review of integrated correlative light and electron microscopy. Rev Sci Instrum 2015;86:. <u>https://doi.org/10.1063/</u> <u>1.4905434</u>011501.
- [168] Dufrêne YF, Ando T, Garcia R, Alsteens D, Martinez-Martin D, Engel A, Gerber C, Müller DJ. Imaging modes of atomic force microscopy for application in molecular and cell biology. Nat Nanotechnol 2017;12:295–307. <u>https://doi.org/10.1038/nnano.2017.45</u>.
- [169] Sahi SJ, Hell SW, Jakobs S. Fluorescence nanoscopy in cell biology. Nat Rev Mol Cell Biol 2017;18:685–701. <u>https://doi.org/10.1038/nrm.2017.71</u>.
- [170] Smith D, Starborg T. Serial block face scanning electron microscopy in cell biology: applications and technology. Tissue Cell 2019;57:111–22. <u>https:// doi.org/10.1016/j.tice.2018.08.011</u>.
- [171] Pietzsch T, Saalfeld S, Preibisch S, Tomancak P. BigDataViewer: visualization and processing for large image data sets. Nat Methods 2015;12:481–3. <u>https://doi.org/10.1038/nmeth.3392</u>.
- [172] Bria A, Iannello G, Onofri L, Peng H. TeraFly: real-time three-dimensional visualization and annotation of terabytes of multi-dimensional volumetric images. Nat Methods 2016;13:192-4. <u>https://doi.org/10.1038/nmeth.3767</u>.
- [173] Peng H, Zhou J, Zhou Z, Bria A, Li Y, Kleissas DM, Drenkow NG, Long B, Liu X, Chen H. Bioimage informatics for big data. Adv Anatomy, Embryol Cell Biol 2016;219:263–72. <u>https://doi.org/10.1007/978-3-319-28549-8_10</u>.
- [174] Ouyang W, Zimmer C. The imaging tsunami: computational opportunities and challenges. Curr Opin Struct Biol 2018;17:105–13. <u>https://doi.org/ 10.1016/j.coisb.2017.07.011</u>.
- [175] Rivenson Y, Göröcs Z, Günaydin H, Zhang Y, Wang H, Ozcan A. Deep learning microscopy. Optica 2017;4:1437–43. <u>https://doi.org/10.1364/</u> OPTICA.4.001437.
- [176] Ouyang W, Aristov A, Lelek M, Hao X, Zimmer C. Deep learning massively accelerates super-resolution localization microscopy. Nat Biotechnol 2018;36:460–8. <u>https://doi.org/10.1038/nbt.4106</u>.
- [177] Zelger P, Kaser K, Rossboth B, Velas L, Schütz GJ, Jesacher A. Threedimensional localization microscopy using deep learning. Optics Express 2018;26:33166–79. <u>https://doi.org/10.1364/OE.26.033166</u>.
- [178] Wu Y, Rivenson Y, Wang H, Luo Y, Ben-David E, Bentolila LA, et al. Threedimensional virtual refocusing of fluorescence microscopy images using deep learning. Nat Methods 2019;16:1323–31. <u>https://doi.org/10.1038/s41592-019-0622-5</u>.
- [179] de Haan K, Rivenson Y, Wu Y, Ozcan A. Deep-learning-based image reconstruction and enhancement in optical microscopy. Proc IEEE 2020;108:30–50. <u>https://doi.org/10.1109/JPROC.2019.2949575</u>.
- [180] Wied GL, Bartels PH, Bibbo M, Dytch HE. Image analysis in quantitative cytopathology and histopathology. Hum Pathol 1989;20:549–71. <u>https://doi.org/10.1016/0046-8177(89)90245-1</u>.
- [181] Howie A. Future trends in microscopy. J Microsc 1989;155:419–35. <u>https:// doi.org/10.1111/j.1365-2818.1989.tb02900.x</u>.
- [182] Lerner B, Guterman H, Dinstein I, Romem Y. Human chromosome classification using multilayer perceptron neural network. Int J Neural Syst 1995;6:359–70. <u>https://doi.org/10.1142/s012906579500024x</u>.
- [183] Boland MV, Markey MK, Murphy RF. Automated recognition of patterns characteristic of subcellular struc- tures in fluorescence microscopy images.

Cytometry 1998;33:366–375, https://doi.org/10.1002/(SICI)1097-0320 (19981101)33:3<366::AID-CYTO12>3.0.CO; 2-R.

- [184] Sjöström PJ, Frydel BR, Wahlberg LU. Artificial neural network-aided image analysis system for cell counting. Cytometry 1999;36:18–26, https://doi.org/ 10.1002/(SICI)1097-0320(19990501)36:1<18::AID-CYTO3>3.0.CO;2-J.
- [185] Arámbula Coíso F, Vega L, Herrera Becerra A, Prieto Mélendez C, Corkidi G. Automatic identification of metaphase spreads and nuclei using neural networks. Med Biol Eng Computing 2001;39:391–6. <u>https://doi.org/10.1007/</u> bf02345296.
- [186] Nattkemper TW, Twellmann T, Ritter H, Schubert W. Human vs machine: evaluation of fluorescence micrograph. Computers Biol Med 2003;33:31–43. https://doi.org/10.1016/s0010-4825(02)00060-4.
- [187] Huang K, Murphy RF. From quantitative microscopy to automated image understanding. J Biomed Opt 2004;9:893–912. <u>https://doi.org/10.1117/ 1.1779233</u>.
- [188] Xing F, Xie Y, Su H, Liu F, Yang L. Deep learning in microscopy image analysis: a survey. IEEE Trans Neural Networks Learn Syst 2018;29:4550-68. <u>https:// doi.org/10.1109/TNNLS.2017.2766168</u>.
- [189] Wang H, Shang S, Long L, Hu R, Wu Y, Chen N, et al. Biological image analysis using deep learning-based methods: literature review. Digital Med 2018;4:157–65. <u>https://doi.org/10.4103/digm.digm_16_18</u>.
- [190] Moen E, Bannon D, Kudo T, Graf W, Covert M. Van Valen Deep learning for cellular image analysis. Nat Methods 2019;16:1233–46. <u>https://doi.org/ 10.1038/s41592-019-0403-1</u>.
- [191] von Chamier L, Laine RF, Henriques R. Artificial intelligence for microscopy: what you should know. Biochem Soc Trans 2019;47:1029-40. <u>https://doi.org/10.1042/BST20180391</u>.
- [192] Gupta A, Harrison PJ, Wieslander H, Pielawski N, Kartasalo K, Partel G, Solorzano L, Suveer A, Klemm AH, Spjuth O, Sintorn IM, Wahlby C. Deep learning in image cytometry: a review. Cytometry Part A 2019;95:366–80. https://doi.org/10.1002/cyto.a.23701.
- [193] Sun J, Tárnok A, Su X. Deep learning-based single-cell optical image studies. Cytometry Part A 2020;97:226–40. <u>https://doi.org/10.1002/cyto.a.23973</u>.
- [194] Zinchuk V, Grossenbacher-Zinchuk O. Machine learning for analysis of microscopy images: a practical guide. Curr Protocols Cell Biol 2020;86. https://doi.org/10.1002/cpcb.101.
- [195] Christiansen EM, Yang SJ, Ando DM, Javaherian A, Skibinski G, Lipnick S, Mount E, Oeil A, Shah K, Lee AK, Goyal P, Fedus W, Poplin R, Esteva A, Berndl M, Rubin LL, Nelson P, Finkbeiner S. In silico labeling: predicting fluorescent labels in unlabeled images. Cell 2018;173:792–803. <u>https://doi.org/10.1016/ i.ccll.2018.03.040</u>.
- [196] Xie Y, Xing F, Shi X, Kong X, Su H, Yang L. Efficient and robust cell detection: a structured regression approach. Med Image Anal 2018;44:245–54. <u>https:// doi.org/10.1016/j.media.2017.07.003</u>.
- [197] Zaimi A, Wabartha M, Herman V, Antonsanti PL, Perone CS, Cohen-Adad J. AxonDeepSeg: automatic axon and myelin segmentation from microscopy data using convolutional neural networks. Sci Rep 2018;8:3816. <u>https://doi.org/10.1038/s41598-018-22181-4</u>.
- [198] Arts M, Smal I, Paul MW, Wyman C, Meijering E. Particle mobility analysis using deep learning and the moment scaling spectrum. Sci Rep 2019;9:17160. <u>https://doi.org/10.1038/s41598-019-53663-8</u>.
- [199] Kraus OZ, Grys BT, Ba J, Chong Y, Frey BJ, Boone C, Andrews BJ. Automated analysis of high-content microscopy data with deep learning. Molecular Syst Biol 2017;13:924. <u>https://doi.org/10.15252/msb.20177551</u>.
- [200] Dong C, Loy CC, He K, Tang X. Image super-resolution using deep convolutional networks. IEEE Trans Pattern Analysis Machine Intelligence 2016;38:295–307. <u>https://doi.org/10.1109/TPAMI.2015.2439281</u>.
- [201] McCann MT, Jin KH, Unser M. Convolutional neural networks for inverse problems in imaging: a review. IEEE Signal Processing Magazine 2017;34:85–95. <u>https://doi.org/10.1109/MSP.2017.2739299.</u>
- [202] Lucas A, Iliadis M, Molina R, Katsaggelos AK. Using deep neural networks for inverse problems in imaging: beyond analytical methods. IEEE Signal Processing Magazine 2018;35:20–36. <u>https://doi.org/10.1109/ MSP.2017.2760358</u>.
- [203] Weigert M, Schmidt U, Boothe T, Müller A, Dibrov A, Jain A, Wilhelm B, Schmidt D, Broaddus C, Culley S, Rocha-Martins M, Segovia-Miranda F, Norden C, Henriques R, Zerial M, Solimena M, Rink J, Tomancak P, Royer L, Jug F, Myers EW. Content-aware image restoration: pushing the limits of fluorescence microscopy. Nat Methods 2018;15:1090–7. <u>https://doi.org/ 10.1038/s41592-018-0216-7</u>.
- [204] Ishii S, Lee S, Urakubo H, Kume H, Kasai H. Generative and discriminative model-based approaches to microscopic image restoration and segmentation. Microscopy 2020;69:79–91. <u>https://doi.org/10.1093/jmicro/ dfaa007</u>.
- [205] Ounkomol C, Seshamani S, Maleckar MM, Collman F, Johnson GR. Label-free prediction of three-dimensional fluorescence images from transmitted-light microscopy. Nat Methods 2018;15:917–20. <u>https://doi.org/10.1038/s41592-018-0111-2</u>.
- [206] Andreopoulos A, Tsotsos JK. 50 years of object recognition: directions forward. Computer Vision Image Understanding 2013;117:827–91. <u>https:// doi.org/10.1016/j.cviu.2013.04.005</u>.
- [207] Liu L, Ouyang W, Wang X, Fieguth P, Chen J, Liu X, et al. Deep learning for generic object detection: a survey. Int J Computer Vision 2020;128:261–318. <u>https://doi.org/10.1007/s11263-019-01247-4</u>.

- [208] Jiao L, Zhang F, Liu F, Yang S, Li L, Feng Z, et al. A survey of deep learningbased object detection. IEEE Access 2019;7:128837–68. <u>https://doi.org/ 10.1109/ACCESS.2019.2939201</u>.
- [209] Zhao ZQ, Zheng P, Xu ST, Wu X. Object detection with deep learning: a review. IEEE Trans Neural Networks Learning Systems 2019;30:3212–32. https://doi.org/10.1109/TNNLS.2018.2876865.
- [210] Smal I, Loog M, Niessen W, Meijering E. Quantitative comparison of spot detection methods in fluorescence microscopy. IEEE Trans Med Imaging 2010;29:282–301. <u>https://doi.org/10.1109/TMI.2009.2025127</u>.
- [211] Ruusuvuori P, Aijö T, Chowdhury S, Garmendia-Torres C, Selinummi J, Birbaumer M, Dudley AM, Pelkmans L, Yli-Harja O. Evaluation of methods for detection of fluorescence labeled subcellular objects in microscope images. BMC Bioinformatics 2010;11:248. <u>https://doi.org/10.1186/1471-2105-11-248</u>.
- [212] Roux L, Racoceanu D, Loménie N, Kulikova M, Irshad H, Klossa J, Capron F, Genestie C, Le Naour G, Gurcan MN. Mitosis detection in breast cancer histological images: an ICPR 2012 contest. J Pathology Informatics 2013;4:8. https://doi.org/10.4103/2153-3539.112693.
- [213] Liu AA, Lu Y, Chen M, Su YT. Mitosis detection in phase contrast microscopy image sequences of stem cell populations: a critical review. IEEE Trans Big Data 2017;3:443–57. <u>https://doi.org/10.1109/TBDATA.2017.2721438</u>.
- [214] Li C, Wang X, Liu W, Latecki LJ. DeepMitosis: mitosis detection via deep detection, verification and segmentation networks. Med Image Anal 2018;45:121-33. <u>https://doi.org/10.1016/j.media.2017.12.002</u>.
- [215] Oktay AB, Gurses A. Automatic detection, localization and segmentation of nano-particles with deep learning in microscopy images. Micron 2019;120:113-9. <u>https://doi.org/10.1016/i.micron.2019.02.009</u>.
- [216] Phan HTH, Kumar A, Feng D, Fulham M, Kim J. Unsupervised two-path neural network for cell event detection and classification using spatiotemporal patterns. IEEE Trans Med Imaging 2019;38:1477–87. <u>https://doi.org/ 10.1109/TMI.2018.2885572</u>.
- [217] Yuan P, Rezvan A, Li X, Varadarajan N, Van Nguyen H. Phasetime: deep learning approach to detect nuclei in time lapse phase images. J Clinical Med 2019;8:1159. <u>https://doi.org/10.3390/jcm8081159</u>.
- [218] Wang EK, Zhang X, Pan L, Cheng C, Dimitrakopoulou-Strauss A, Li Y, et al. Multi-path dilated residual network for nuclei segmentation and detection. Cells 2019;8:499. <u>https://doi.org/10.3390/cells8050499</u>.
- [219] Araujo FHD, Silva RRV, Ushizima DM, Rezende MT, Carneiro CM, Campos Bianchi AG, Medeiros FNS. Deep learning for cell image segmentation and ranking. Computerized Med Imaging Graphics 2019;72:13–21. <u>https://doi.org/10.1016/j.compmedimag.2019.01.003</u>.
- [220] Hesamian MH, Jia W, He X, Kennedy P. Deep learning techniques for medical image segmentation: achievements and challenges. J Digit Imaging 2019;32:582–96. <u>https://doi.org/10.1007/s10278-019-00227-x</u>.
- [221] Guo Y, Liu Y, Georgiou T, Lew MS. A review of semantic segmentation using deep neural networks. Int J Multimedia Information Retrieval 2018;7:87–93. https://doi.org/10.1007/s13735-017-0141-z.
- [222] Garcia-Garcia A, Orts-Escolano S, Oprea S, Villena-Martinez V, Martinez-Gonzalez P, Garcia-Rodriguez J. A survey on deep learning techniques for image and video semantic segmentation. Appl Soft Computing 2018;70:41–65. <u>https://doi.org/10.1016/j.asoc.2018.05.018</u>.
- [223] Long J, Shelhamer E, Darrell T. Fully convolutional networks for semantic segmentation. IEEE Trans Pattern Anal Machine Intelligence 2017;39:640–51. <u>https://doi.org/10.1109/TPAMI.2016.2572683</u>.
- [224] Falk T, Mai D, Bensch R, C, ic ek O., Abdulkadir A, Marrakchi Y, Böhm A, Deubner J, Jäckel Z, Seiwald K, Dovzhenko A, Tietz O, Dal Bosco C, Walsh S, Saltukoglu D, Tay TL, Prinz M, Palme K, Simons M, Diester I, Brox T, Ronneberger O. U-Net: deep learning for cell counting, detection, and morphometry. Nat Methods 2019;16:67–70, https://doi.org/10.1038/ s41592-018-0261-2.
- [225] Badrinarayanan V, Kendall A, Cipolla R. SegNet: a deep convolutional encoder-decoder architecture for image segmentation. IEEE Trans Pattern Anal Machine Intelligence 2017;39:2481–95. <u>https://doi.org/10.1109/</u> <u>TPAMI.2016.2644615</u>.
- [226] Chen LC, Papandreou G, Kokkinos I, Murphy K, Yuille AL. DeepLab: semantic image segmentation with deep convolutional nets, atrous convolution, and fully connected CRFs. IEEE Trans Pattern Anal Mach Intell 2018;40:834–48. https://doi.org/10.1109/TPAML2017.2699184.
- [227] Jiao L, Zhao J. A survey on the new generation of deep learning in image processing. IEEE Access 2019;7:172231–63. <u>https://doi.org/10.1109/</u> ACCESS.2019.2956508.
- [228] Arganda-Carreras I, Turaga SC, Berger DR, Cires, an D, Giusti A, Gambardella LM, Schmidhuber J, Laptev D, Dwivedi S, Buhmann JM, Liu T, Seyedhosseini M, Tasdizen T, Kamentsky L, Burget R, Uher V, Tan X, Sun C, Pham TD, Bas E, Uzunbas MG, Cardona A, Schindelin J, Seung HS. Crowdsourcing the creation of image segmentation algorithms for connectomics. Front Neuroanatomy 2015;9:142, https://doi.org/10.3389/fnana.2015.00142.
- [229] Caicedo JC, Goodman A, Karhohs KW, Cimini BA, Ackerman J, Haghighi M, et al. Data science bowl. Nat Methods 2018;2019(16):1247–53. <u>https://doi.org/10.1038/s41592-019-0612-7</u>.
- [230] Sirinukunwattana K, Pluim JPW, Chen H, Qi X, Heng PA, Guo YB, Wang LY, Matuszewski BJ, Bruni E, Sanchez U, Böhm A, Ronneberger O, Cheikh BB, Racoceanu D, Kainz P, Pfeiffer M, Urschler M, Snead DRJ, Rajpoot NM. Gland segmentation in colon histology images: the glas challenge contest. Med Image Anal 2017;35:489–502, https://doi.org/10.1016/j.media.2016.08.008.

- [231] Smeulders AWM, Chu DM, Cucchiara R, Calderara S, Dehghan A, Shah M. Visual tracking: an experimental survey. IEEE Trans Pattern Anal Machine Intelligence 2014;36:1442–68. <u>https://doi.org/10.1109/TPAMI.2013.230</u>.
- [232] Meijering E, Dzyubachyk O, Smal I, van Cappellen WA. Tracking in cell and developmental biology. Seminars Cell Devel Biol 2009;20:894–902. <u>https:// doi.org/10.1016/i.semcdb.2009.07.004</u>.
- [233] Hilsenbeck O, Schwarzfischer M, Skylaki S, Schauberger B, Hoppe PS, Loeffler D, Kokkaliaris KD, Hastreiter S, Skylaki E, Filipczyk A, Strasser M, Buggenthin F, Feigelman JS, Krumsiek J, van der Berg AJJ, Endele M, Etzrodt M, Marr C, Theis FJ, Schroeder T. Software tools for single-cell tracking and quantification of cellular and molecular properties. Nat Biotechnol 2016;34:703–6. https://doi.org/10.1038/nbt.3626.
- [234] Shen H, Tauzin LJ, Baiyasi R, Wang W, Moringo N, Shuang B, et al. Single particle tracking: from theory to biophysical applications. Chem Rev 2017;117:7331–76. <u>https://doi.org/10.1021/acs.chemrev.6b00815</u>.
- [235] Ma Y, Wang X, Liu H, Wei L, Xiao L. Recent advances in optical microscopic methods for single-particle tracking in biological samples. Analytical Bioanalytical Chem 2019;411:4445–63. <u>https://doi.org/10.1007/s00216-019-01638-z</u>.
- [236] Lugagne JB, Lin H, Dunlop MJ. DeLTA: automated cell segmentation, tracking, and lineage reconstruction using deep learning. PLoS Comput Biol 2020;16: e1007673. <u>https://doi.org/10.1371/journal.pcbi.1007673</u>.
- [237] Wang J, Su X, Zhao L, Zhang J. Deep reinforcement learning for data association in cell tracking. Front Bioeng Biotechnol 2020;8:298. <u>https://doi.org/10.3389/fbioe.2020.00298</u>.
- [238] Yao Y, Smal I, Grigoriev I, Akhmanova A. Meijering Deep learning method for data association in parti- cle tracking. Bioinformatics 2020;36:Forthcoming. https://doi.org/10.1093/bioinformatics/btaa597.
- [239] Guo Y, Liu Y, Oerlemans A, Lao S, Wu S, Lew MS. Deep learning for visual understanding: a review. Neurocomputing 2016;187:27–48. <u>https://doi.org/ 10.1016/j.neucom.2015.09.116</u>.
- [240] Rawat W, Wang Z. Deep convolutional neural networks for image classification: a comprehensive review. Neural Computation 2017;29:2352–449. <u>https://doi.org/10.1162/NECO_a_00990</u>.
- [241] Voulodimos A, Doulamis N, Doulamis A, Protopapadakis E. Deep learning for computer vision: a brief review. Comput Intelligence Neurosci 2018;2018:7068349. <u>https://doi.org/10.1155/2018/7068349</u>.
- [242] Kraus OZ, Frey BJ. Computer vision for high content screening. Crit Rev Biochem Mol Biol 2016;51:102–9. <u>https://doi.org/10.3109/</u> 10409238.2015.1135868.
- [243] Inés A, Domínguez C, Heras J, Mata E, Pascual V. Deep-Clas4Bio: connecting bioimaging tools with deep learning frameworks for image classification. Computers Biol Med 2019;108:49–56, https://doi.org/10.1016/j. compbiomed.2019.03.026.
- [244] Shifat-E-Rabbi M, Yin X, Fitzgerald CE, Rohde GK. Cell image classification: a comparative overview. Cytometry Part A 2020;97:347–62. <u>https://doi.org/ 10.1002/cyto.a.23984</u>.
- [245] Sommer C, Gerlich DW. Machine learning in cell biology teaching computers to recognize phenotypes. J Cell Sci 2013;126:5529–39. <u>https:// doi.org/10.1242/ics.123604.</u>
- [246] Caicedo JC, Cooper S, Heigwer F, Warchal S, Qiu P, Molnar C, et al. Dataanalysis strategies for image-based cell profiling. Nat Methods 2017;14:849-63. <u>https://doi.org/10.1038/nmeth.4397</u>.
- [247] Sullivan DP, Winsnes CF, Åkesson L, Hjelmare M, Wiking M, Schutten R, et al. Deep learning is combined with massive-scale citizen science to improve large-scale image classification. Nat Biotechnol 2018;36:820–8. <u>https://doi.org/10.1038/nbt.4225</u>.
- [248] Medathati NVK, Neumann H, Masson GS, Kornprobst P. Bio-inspired computer vision: towards a synergistic approach of artificial and biological vision. Computer Vision Image Understanding 2016;150:1–30. <u>https://doi.org/10.1016/j.cviu.2016.04.009</u>.
- [249] Marblestone AH, Wayne G, Kording KP. Toward an integration of deep learning and neuroscience. Front Computational Neurosci 2016;10:94. https://doi.org/10.3389/fncom.2016.00094.
- [250] Kriegeskorte N. Deep neural networks: a new framework for modeling biological vision and brain information processing. Ann Rev Vision Sci 2015;1:417–46. <u>https://doi.org/10.1146/annurev-vision-082114-035447.</u>
- [251] Elsken T, Metzen JH, Hutter F. Neural architecture search: a survey. J Machine Learning Res 2019;20:1–21. http://jmlr.org/papers/v20/18-598. html.
- [252] Faes L, Wagner SK, Fu DJ, Liu X, Korot E, Ledsam JR, Back T, Chopra R, Pontikos N, Kern C, Moraes G, Schmid MK, Sim D, Balaskas K, Bachmann LM, Denniston AK, Keane PA. Automated deep learning design for medical image classification by health-care professionals with no coding experience: a feasibility study. The Lancet Digital Health 2019;1:e232–42. <u>https://doi.org/10.1016/S2589-7500(19)30108-6</u>.
- [253] Zhang M, Zhou Y, Zhao J, Man Y, Liu B, Yao R. A survey of semi- and weakly supervised semantic segmentation of images. Artificial Intelligence Rev 2020;53:4259–88. <u>https://doi.org/10.1007/s10462-019-09792-7</u>.
- [254] Wilson G, Cook DJ. A survey of unsupervised deep domain adaptation. ACM Trans Intelligent Syst Tech 2020;11:51. <u>https://doi.org/10.1145/3400066</u>.
- [255] Weiss K, Khoshgoftaar TM, Wang DD. A survey of transfer learning. J Big Data 2016;3:9, https://doi.org/10.1186/s40537-016-0043-6.
- [256] Kozubek M. When deep learning meets cell image synthesis. Cytometry Part A 2020;97:222–5. <u>https://doi.org/10.1002/cyto.a.23957</u>.

- [257] Geng C, Huang SJ, Chen S. Recent advances in open set recognition: a survey. IEEE Trans Pattern Anal Machine Intelligence 2020;42:Forthcoming, <u>https://doi.org/10.1109/TPAMI.2020.2981604</u>.
- [258] Guo W, Wang J, Wang S. Deep multimodal representation learning: a survey. IEEE Access 2019;7:63373–94. <u>https://doi.org/10.1109/ACCESS.2019.2916887</u>.
- [259] Gao J, Li P, Chen Z, Zhang J. A survey on deep learning for multimodal data fusion. Neural Comput 2020;32:829–64. <u>https://doi.org/10.1162/ neco a 01273</u>.
- [260] Zhang Z, Zhao Y, Liao X, Shi W, Li K, Zou Q, Peng S. Deep learning in omics: a survey and guideline. Briefings Functional Genomics 2019;18:41–57. <u>https:// doi.org/10.1093/bfgp/ely030</u>.
- [261] Ben-Nun T, Hoefler T. Demystifying parallel and distributed deep learning: an in-depth concurrency analysis. ACM Computing Surveys 2019;52:65. <u>https:// doi.org/10.1145/3320060</u>.
- [262] Deng L, Li G, Han S, Shi L, Xie Y. Model compression and hardware acceleration for neural networks: a comprehensive survey. Proc IEEE 2020;108:485–532. <u>https://doi.org/10.1109/JPROC.2020.2976475</u>.
- [263] Chang K, Balachandar N, Lam C, Yi D, Brown J, Beers A, et al. Distributed deep learning networks among institutions for medical imaging. J Am Med Informatics Assoc 2018;25:945–54. <u>https://doi.org/10.1093/jamia/ocy017</u>.
- [264] Adadi A, Berrada M. Peeking inside the black-box: a survey on explainable artificial intelligence (XAI). IEEE Access 2018;6:52138–60. <u>https://doi.org/ 10.1109/ACCESS.2018.2870052</u>.
- [265] Chatzimparmpas A, Martins RM, Jusufi I, Kerren A. A survey of surveys on the use of visualization for interpreting machine learning models. Information Visualization 2020;19:207–33. <u>https://doi.org/10.1177/1473871620904671.</u>
- [266] Langlotz CP. Will artificial intelligence replace radiologists?. Radiology: Artificial Intelligence 2019;1:e190058. https://doi.org/10.1148/ryai. 2019190058.
- [267] Ahuja AS. The impact of artificial intelligence in medicine on the future role of the physician. PeerJ 2019;7:e7702. <u>https://doi.org/10.7717/peeri.7702</u>.
- [268] Granter SR, Beck AH, Papke DJ. AlphaGo, deep learning, and the future of the human microscopist. Arch Pathol Lab Med 2017;141:619–21. <u>https://doi.org/ 10.5858/arpa.2016-0471-ED</u>.
- [269] Pinto Dos Santos D, Giese D, Brodehl S, Chon SH, Staab W, Kleinert R, et al. Medical students' attitude towards artificial intelligence: a multicentre survey. Eur Radiol 2019;29:1640–6. <u>https://doi.org/10.1007/s00330-018-5601-1</u>.
- [270] Stockham TG. Image processing in the context of a visual model. Proc IEEE 1972;60:828-42. <u>https://doi.org/10.1109/proc.1972.8782</u>.
- [271] Granrath DJ. The role of human visual models in image processing. Proc IEEE 1981;69:552–61. <u>https://doi.org/10.1109/PROC.1981.12024</u>.
- [272] Meijering E. Neuron tracing in perspective. Cytometry Part A 2010;77:693–704. <u>https://doi.org/10.1002/cyto.a.20895</u>.
- [273] Donohue DE, Ascoli GA. Automated reconstruction of neuronal morphology: an overview. Brain Res Rev 2011;67:94–102. <u>https://doi.org/10.1016/j. brainresrev.2010.11.003</u>.
- [274] Peng H, Hawrylycz M, Roskams J, Hill S, Spruston N, Meijering E, et al. BigNeuron: large-scale 3D neuron reconstruction from optical microscopy images. Neuron 2015;87:252–6. <u>https://doi.org/10.1016/j.neuron.2015.06.036</u>.
- [275] Fakhry A, Peng H, Ji S. Deep models for brain EM image segmentation: novel insights and improved performance. Bioinformatics 2016;32:2352–8. https://doi.org/10.1093/bioinformatics/btw165.
- [276] Zhou Z, Kuo HC, Peng H, Long F. DeepNeuron: an open deep learning toolbox for neuron tracing. Brain Informatics 2018;5:3. <u>https://doi.org/10.1186/ s40708-018-0081-2</u>.
- [277] Urakubo H, Bullmann T, Kubota Y, Oba S, Ishii S. UNI-EM: an environment for deep neural network-based automated segmentation of neuronal electron microscopic images. Sci Rep 2019;9:19413. <u>https://doi.org/10.1038/s41598-019-55431-0</u>.
- [278] Wong C, Houlsby N, Lu Y, Gesmundo A. Transfer learning with neural AutoML. Adv Neural Information Processing Syst 2018;31:8366–75. https:// papers.nips.cc/paper/8056-transfer-learning-with-neural-automl.
- [279] Liu C, Chen LC, Schroff F, Adam H, Hua W, Yuille AL, Fei-Fei L. Auto-DeepLab: hierarchical neural architecture search for semantic image segmentation. Computer Vision Pattern Recognition 2019;32:82–92. <u>https://doi.org/ 10.1109/CVPR.2019.00017</u>.
- [280] Weng Y, Zhou T, Li Y, Qiu X. NAS-Unet: neural architecture search for medical image segmentation. IEEE Access 2019;7:44247–57. <u>https://doi.org/10.1109/</u> <u>ACCESS.2019.2908991</u>.
- [281] Kwasigroch A, Grochowski M, Mikołajczyk A. Neural architecture search for skin lesion classification. IEEE Access 2020;8:9061–71. <u>https://doi.org/ 10.1109/ACCESS.2020.2964424</u>.
- [282] Zhou ZH. A brief introduction to weakly supervised learning. Natl Sci Rev 2018;5:44–53. <u>https://doi.org/10.1093/nsr/nwx106</u>.
- [283] Liu Q, Yu L, Luo L, Dou Q, Heng PA. Semi-supervised medical image classification with relation-driven self-ensembling model. IEEE Trans Med Imaging 2020;39:Forthcoming, https://doi.org/10.1109/TMI.2020.2995518.
 [284] Qu H, Wu P, Huang Q, Yi J, Yan Z, Li K, Riedlinger GM, De S, Zhang S, Metaxas
- [284] Qu H, Wu P, Huang Q, Yi J, Yan Z, Li K, Riedlinger GM, De S, Zhang S, Metaxas DN. Weakly supervised deep nuclei segmen- tation using partial points annotation in histopathology images. IEEE Transactions on Medical Imaging 2020;39:Forthcoming. https://doi.org/10.1109/TMI.2020.3002244.
- [285] Shi X, Su H, Xing F, Liang Y, Qu G, Yang L. Graph temporal ensembling based semi-supervised convolutional neural network with noisy labels for

histopathology image analysis. Med Image Anal 2020;60. <u>https://doi.org/</u> 10.1016/j.media.2019.101624.

- [286] Bengio Y. Deep learning of representations for unsupervised and transfer learning. Proc Machine Learning Res 2012;27:17–36. http://proceedings.mlr. press/ v27/bengio12a.html.
- [287] Perone CS, Ballester P, Barros RC, Cohen-Adad J. Unsupervised domain adaptation for medical imaging segmentation with self-ensembling. NeuroImage 2019;194:1–11. <u>https://doi.org/10.1016/j.</u> neuroimage.2019.03.026.
- [288] Wang P, Wang J, Li Y, Li L, Zhang H. Adaptive pruning of transfer learned deep convolutional neural network for classification of cervical pap smear images. IEEE Access 2020;8:50674–83. <u>https://doi.org/10.1109/</u> ACCESS.2020.2979926.
- [289] Shin HC, Roth HR, Gao M, Lu L, Xu Z, Nogues I, et al. Deep convolutional neural networks for computer-aided detection: CNN architectures, dataset characteristics and transfer learning. IEEE Trans Med Imaging 2016;35:1285–98. <u>https://doi.org/10.1109/TMI.2016.2528162</u>.
- [290] Abbas A, Abdelsamea MM, Gaber MM. DeTrac: transfer learning of class decomposed medical images in convolutional neural networks. IEEE Access 2020;8:74901–13. <u>https://doi.org/10.1109/ACCESS.2020.2989273</u>.
- [291] Ulman V, Svoboda D, Nykter M, Kozubek M, Ruusuvuori P. Virtual cell imaging: a review on simulation methods employed in image cytometry. Cytometry Part A 2016;89:1057-72. <u>https://doi.org/10.1002/cyto.a.23031</u>.
- [292] Svoboda D, Ulman V. MitoGen: a framework for generating 3D synthetic time-lapse sequences of cell populations in fluorescence microscopy. IEEE Trans Med Imaging 2017;36:310–21. <u>https://doi.org/10.1109/ TMI.2016.2606545</u>.
- [293] Rasti P, Huaman R, Riviere C, Rousseau D. Supervised machine learning for 3D microscopy without manual annotation: application to spheroids. Proc SPIE 2018;10677:1067728. <u>https://doi.org/10.1117/12.2303706</u>.
- [294] Dunn KW, Fu C, Ho DJ, Lee S, Han S, Salama P, et al. DeepSynth: threedimensional nuclear segmentation of biological images using neural networks trained with synthetic data. Sci Rep 2019;9:18295. <u>https://doi. org/10.1038/s41598-019-54244-5</u>.
- [295] Yao R, Qian J, Huang Q. Deep-learning with synthetic data enables automated picking of cryo-EM particle images of biological macromolecules. Bioinformatics 2020;36:1252–9. <u>https://doi.org/10.1093/bioinformatics/btz728</u>.
- [296] Scheirer WJ, de Rezende Rocha A, Sapkota A, Boult TE. Toward open set recognition. IEEE Trans Pattern Anal Machine Intelligence 2013;35:1757–72. https://doi.org/10.1109/TPAML2012.256.
- [297] Rudd EM, Jain LP, Scheirer WJ, Boult TE. The extreme value machine. IEEE Trans Pattern Anal Mach Intell 2017;40:762–8. <u>https://doi.org/10.1109/ TPAMI.2017.2707495</u>.
- [298] Modla S, Czymmek KJ. Correlative microscopy: a powerful tool for exploring neurological cells and tissues. Micron 2011;42:773–92. <u>https://doi.org/ 10.1016/j.micron.2011.07.001</u>.
- [299] Kobayashi S, Iwamoto M, Haraguchi T. Live correlative lightelectron microscopy to observe molecular dynamics in high resolution. Microscopy 2016;65:296–308. <u>https://doi.org/10.1093/jmicro/dfw024</u>.

- [300] Hauser M, Wojcik M, Kim D, Mahmoudi M, Li W, Xu K. Correlative superresolution microscopy: new dimensions and new opportunities. Chem Rev 2017;117:7428–56. <u>https://doi.org/10.1021/acs.chemrev.6b00604</u>.
- [301] Mamoshina P, Vieira A, Putin E, Zhavoronkov A. Applications of deep learning in biomedicine. Molecular Pharmaceutics 2016;13:1445–54. <u>https://doi.org/ 10.1021/acs.molpharmaceut.5b00982</u>.
- [302] Ching T, Himmelstein DS, Beaulieu-Jones BK, Kalinin AA, Do BT, Way GP, et al. Opportunities and obstacles for deep learning in biology and medicine. J R Soc Interface 2018;15. <u>https://doi.org/10.1098/rsif.2017.0387</u>.
- [303] Ramachandram D, Taylor GW. Deep multimodal learning: a survey on recent advances and trends. IEEE Signal Process Mag 2017;34:96–108. <u>https://doi. org/10.1109/MSP.2017.2738401</u>.
- [304] Sze V, Chen YH, Yang TJ, Emer JS. Efficient processing of deep neural networks: a tutorial and survey. Proc IEEE 2017;105:2295–329. <u>https://doi. org/10.1109/IPROC.2017.2761740</u>.
- [305] Remedios SW, Roy S, Bermudez C, Patel MB, Butman JA, Landman BA, et al. Distributed deep learning across multisite datasets for generalized CT hemorrhage segmentation. Med Phys 2020;47:89–98. <u>https://doi.org/ 10.1002/mp.13880</u>.
- [306] Mi H, Xu K, Feng D, Wang H, Zhang Y, Zheng Z, et al. Collaborative deep learning across multiple data centers. Sci China Inf Sci 2020;63:182102. <u>https://doi.org/10.1007/s11432-019-2705-2</u>.
- [307] Gilpin LH, Bau D, Yuan BZ, Bajwa A, Specter M, Kagal L. Explaining explanations: an overview of interpretability of machine learning. Data Sci Adv Analytics 2018;5:80–9. <u>https://doi.org/10.1109/DSAA.2018.00018</u>.
- [308] Hagras H. Toward human-understandable, explainable AI. Computer 2018;51:28–36. <u>https://doi.org/10.1109/MC.2018.3620965</u>.
- [309] Zhang QS, Zhu SC. Visual interpretability for deep learning: a survey. Front Information Tech Electronic Eng 2018;19:27–39. <u>https://doi.org/10.1631/ FITEE.1700808</u>.
- [310] Escalante HJ, Escalera S, Guyon I, Baró X, Güc Jütürk Y, Güc Jü U, van Gerven M (Eds). Explainable and Interpretable Models in Computer Vision and Machine Learning. Springer Nature, Cham, Switzerland, 2018, https://link. springer.com/book/10.1007/978-3-319-98131-4.
- [311] Samek W, Montavon G, Vedaldi A, Hansen LK, Müller KR (Eds). Explainable Al: Interpreting, Explaining and Visualizing Deep Learning. Springer Nature, Cham, Switzerland, 2019, https://link.springer.com/book/10.1007/978-3-030-28954-6.
- [312] Choo J, Liu S. Visual analytics for explainable deep learning. IEEE Comput Graphics Appl 2018;38:84–92. <u>https://doi.org/10.1109/MCG.2018.042731661</u>.
- [313] Hohman F, Kahng M, Pienta R, Chau DH. Visual analytics in deep learning: an interrogative survey for the next frontiers. IEEE Trans Visualization Computer Graphics 2019;25:2674–93. <u>https://doi.org/10.1109/TVCG.2018.2843369</u>.
- [314] Shah NR. Health care in 2030: will artificial intelligence replace physicians?. Ann Intern Med 2019;170:407-8. <u>https://doi.org/10.7326/M19-0344</u>.
- [315] Korot E, Wagner SK, Faes L, Liu X, Huemer J, Ferraz D, Keane PA, Balaskas K. Will AI replace ophthalmologists?. Trans Vision Sci Tech 2020;9:2. <u>https://doi.org/10.1167/tvst.9.2.2</u>.