



## Draft Genome Sequence of the Archiascomycetous Yeast Saitoella complicata

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The draft genome sequence of the archiasomycetous yeast *Saitoella complicata* was determined. The assembly of newly and previously sequenced data sets resulted in 104 contigs (total of 14.1 Mbp;  $N_{50}$ , 239 kbp). On the newly assembled genome, a total of 6,933 protein-coding sequences (7,119 transcripts, including alternative splicing forms) were identified.

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The subphylum *Taphrinomycotina* (*Archiascomycetes*) is the earliest ascomycetous lineage that diverged before the separation of the subphyla *Pezizomycotina* (*Euascomycetes*, filamentous ascomycetes) and *Saccharomycotina* (*Hemiascomycetes*, budding ascomycetous yeasts) (1, 2). The anamorphic and saprobic budding yeast *Saitoella complicata* is a member of the *Taphrinomycotina*, which was isolated from Himalayan soil (3). Interestingly, *S. complicata* shares some characteristics with both ascomycetous and basidiomycetous yeasts (3, 4).

We previously attempted to assemble the genome sequence of *S. complicata* using 454 (Roche) sequences (5) and Illumina pairedend read pairs (6). Although these previous assemblies were of a large number of small contigs, at 7,981 contigs (13.0 Mbp) (5) and 1,800 contigs (14.2 Mbp) (6), respectively, we found that the amino acid sequences of protein-coding genes identified on the contigs showed the highest similarity to proteins of *Pezizomycotina* (5, 6).

To elucidate the detailed characteristics of the *S. complicata* genomic DNA sequences, we have refined the genome assembly with additional sequencing of mate-paired DNA libraries of this species. We generated a total of 11.4 million paired-end read pairs (700-bp insert and 100 bp in length) and a total of 23.7 million mate-paired read pairs (6.2 million 3-kb-, 6.2 million 5-kb-, 5.3 million 10-kb-, and 6.0 million 15-kb-long-insert read pairs), respectively, using Illumina HiSeq and MiSeq sequencers. The read pairs were dereplicated by Fulcrum (7) and assembled using the SPAdes assembler (8). The assembly of the dereplicated read pairs by using 21 to 89 bp for the *k*-mer size option yielded a set of 104 contigs of  $\geq 1$  kb, whose total size and  $N_{50}$  are 14.1 Mb and 239 kb, respectively.

Using Augustus (9), a gene prediction software based on the alignment of expressed sequences to the genome, we have determined coding sequences (CDSs) of the genes expressed on the assembled genome of *Saitoella* according to the gene model of *Aspergillus nidulans*, which is thought to have some taxonomic proximity to *Saitoella*. Based on the exon coordinates mapped by a total of 89.3 million RNA sequencing (RNA-seq) paired-end

read pairs (100 bp in length) uniquely mapped to the genome by BLAT (10), Augustus identified 6,933 protein-coding genes (7,119 transcripts, including alternative splicing forms) on the *Saitoella* genome. All this computational work was done on the NIG Supercomputer system (11).

**Nucleotide sequence accession numbers.** The DNA sequences have been deposited in DDBJ under the accession numbers BACD03000001 to BACD03000104.

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## REFERENCES

- Nishida H, Sugiyama J. 1994. Archiascomycetes: detection of a major new lineage within the *Ascomycota*. Mycoscience 35:361–366. http:// dx.doi.org/10.1007/BF02268506.
- Liu Y, Leigh JW, Brinkmann H, Cushion MT, Rodriguez-Ezpeleta N, Philippe H, Lang BF. 2009. Phylogenomic analyses support the monophyly of *Taphrinomycotina*, including *Schizosaccharomyces* fission yeasts. Mol Biol Evol 26:27–34. http://dx.doi.org/10.1093/molbev/msn221.
- Goto S, Sugiyama J, Hamamoto M, Komagata K. 1987. Saitoella, a new anamorph genus in the *Cryptococcaceae* to accommodate two Himalayan yeast isolates formerly identified as *Rhodotorula glutinis*. J Gen Appl Microbiol 33:75–85. http://dx.doi.org/10.2323/jgam.33.75.
- Sugiyama J, Fukagawa M, Chiu S, Komagata K. 1985. Cellular carbohydrate composition, DNA base composition, ubiquinone systems, and diazonium blue B color test in the genera *Rhodosporidium*, *Leucosporidium*, *Rhodotorula* and related basidiomycetous yeasts. J Gen Appl Microbiol 31:519–550. http://dx.doi.org/10.2323/jgam.31.519.
- Nishida H, Hamamoto M, Sugiyama J. 2011. Draft genome sequencing of the enigmatic yeast *Saitoella complicata*. J Gen Appl Microbiol 57: 243–246. http://dx.doi.org/10.2323/jgam.57.243.
- 6. Nishida H, Matsumoto T, Kondo S, Hamamoto M, Yoshikawa H. 2014. The early diverging ascomycetous budding yeast *Saitoella complicata* has three histone deacetylases belonging to the Clr6, Hos2, and Rpd3 lineages. J Gen Appl Microbiol **60**:7–12. http://dx.doi.org/10.2323/jgam.60.7.

- Burriesci MS, Lehnert EM, Pringle JR. 2012. Fulcrum: condensing redundant reads from high-throughput sequencing studies. Bioinformatics 28:1324–1327. http://dx.doi.org/10.1093/bioinformatics/bts123.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to singlecell sequencing. J Comput Biol 19:455–477. http://dx.doi.org/10.1089/ cmb.2012.0021.
- Stanke M, Diekhans M, Baertsch R, Haussler D. 2008. Using native and syntenically mapped cDNA alignments to improve *de novo* gene finding. Bioinformatics 24:637–644. http://dx.doi.org/10.1093/bioinformatics/ btn013.
- Kent WJ. 2002. BLAT—the BLAST-Like Alignment Tool. Genome Res 12:656–664. http://dx.doi.org/10.1101/gr.229202.
- Ogasawara O, Mashima J, Kodama Y, Kaminuma E, Nakamura Y, Okubo K, Takagi T. 2013. DDBJ new system and service refactoring. Nucleic Acids Res 41:D25–D29. http://dx.doi.org/10.1093/nar/gks1152.