

Trichostatin A increases the TIMP-1/MMP ratio to protect against osteoarthritis in an animal model of the disease

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Received September 7, 2015; Accepted June 7, 2016

DOI: 10.3892/mmr.2016.5523

Abstract. The histone deacetylase inhibitor trichostatin A (TSA) has been demonstrated to alleviate certain symptoms associated with osteoarthritis (OA). However, the exact mechanisms underlying this protective effect remain to be elucidated. The present study therefore examined the effects of TSA on the expression levels of interleukin-1 β (IL-1 β)-induced matrix metalloproteinases (MMPs) and tissue inhibitor of metalloproteinases-1 (TIMP-1) *in vitro* and *in vivo*. *In vitro*, reverse transcription-quantitative polymerase chain reaction was performed to investigate alterations in mRNA expression levels in TSA-treated chondrocytes in the presence or absence of IL-1 β ; in addition, protein expression and acetylation levels were assessed by western blotting. *In vivo*, TSA was administered to rats by intra-articular injection, following which the mRNA and protein expression levels were analyzed. In addition, macroscopic and histological observations were conducted. Chondrocytes treated with IL-1 β demonstrated increased mRNA and protein expression levels of MMP-1, MMP-3 and MMP-13, and decreased expression levels of TIMP-1 mRNA and protein; these alterations were significantly attenuated by TSA treatment. In addition, increased MMPs and decreased TIMP-1 expression levels were observed *in vivo* in the OA rat model. TSA treatment demonstrated *in vivo* efficacy through the attenuation of various OA-associated molecular and physiological changes. Taken together, the results of the present study suggest that TSA has potential therapeutic value for the treatment of OA.

Introduction

Osteoarthritis (OA), the predominant form of arthritis (1), is characterized by slow, progressive joint degeneration. It is a primary cause of continuous pain and chronic disability that restricts the normal daily lives of many elderly individuals (2-4). Although there are numerous risk factors for OA, including obesity, aging and trauma, the mechanisms underlying these associations remain to be elucidated (5-7). Previous studies have demonstrated that synthesis and catabolism disorders in chondrocytes are involved in the onset of OA (8,9). It is generally accepted that the loss of articular cartilage matrix components, including collagen type II and aggrecan, resulting from the activation of proteolytic enzymes, such as matrix metalloproteinases (MMPs), is a frequent characteristic of OA (10-14). MMPs are a family of endopeptidases that contribute to numerous physiological processes via the degradation of extracellular matrix components. MMP-1 and MMP-13, also referred to as collagenases, degrade all types of collagen, particularly those that supply mechanical strength to tissues, such as collagen type II in articular cartilage (14,15). MMP-3, also referred to as pro-MMP, is a type of stromelysin that activates other MMPs, including MMP-1 and MMP-13 (16). The biological activities of MMPs are antagonized by tissue inhibitors of metalloproteinases (TIMPs), such as TIMP-1 (17,18). Thus, an imbalance in the activities of MMPs and TIMPs is considered to be critical in OA progression (19,20).

Inflammatory cytokines, including interleukin (IL)-1 β , tumor necrosis factor- α and IL-6, are crucial for chondrocyte apoptosis and degradation of the extracellular matrix (21,22), and are thus involved in the pathogenesis and development of OA. A previous study has confirmed that IL-1 β induces upregulation of MMPs and downregulation of TIMPs, resulting in degradation of proteoglycans and collagen (21).

Trichostatin A (TSA), originally developed as an anti-fungal agent (23), is a non-selective histone deacetylase inhibitor (HDACi) (24,25). The acetylation of lysine residues on histones H3 and H4 opens the chromatin structure to promote active gene transcription (26-29). The regulation of inflammatory genes by TSA has attracted considerable attention. Previous studies have demonstrated that TSA exerts anti-OA and anti-rheumatoid arthritis effects via suppression of the expression of MMPs, cyclooxygenase-2, inducible nitric

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Key words: osteoarthritis, trichostatin A, matrix metalloproteinases, tissue inhibitors of metalloproteinases-1, acetylation

oxide synthase and a disintegrin and MMP with thrombospondin motifs (4,30). Furthermore, it has been revealed that the underlying anti-inflammatory mechanism may involve inhibition of the mitogen-activated protein kinase and nuclear factor- κ B (NF- κ B) signaling pathways, and subsequent upregulation of sirtuin 1 expression (31). However, the underlying mechanism by which TSA ameliorates the OA inflammatory response remains to be fully elucidated, and there is currently not enough evidence to determine whether the anti-OA effect of TSA involves altering the level of acetylation in chondrocytes. Therefore, the present study investigated the association between the anti-OA effects of TSA and the levels of acetylation in chondrocytes.

Materials and methods

Reagents. TSA and IL-1 β were purchased from Sigma-Aldrich (St. Louis, MO, USA). 3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT), Dulbecco's modified Eagle's medium (DMEM), fetal bovine serum (FBS), and collagenase II were obtained from Gibco; Thermo Fisher Scientific, Inc. (Waltham, MA, USA).

Isolation and culture of rat chondrocytes. Rat cartilage was collected from the knee and hip joints of six 2-week-old female Sprague Dawley rats (weight, 35-45 g) from the Zhejiang Academy of Medical Sciences (Hangzhou, China). Rats were housed under pathogen-free conditions at room temperature (22-25°C) and 50% humidity, and has a 12-h light/dark cycle. Food and water were offered *ad libitum*. To isolate chondrocytes, rat cartilage was digested with 0.2% collagenase II (Sigma-Aldrich) in DMEM for 4 h at 37°C. Rat cartilage was subsequently filtered through a 150 μ m mesh filter and centrifuged at 300 x g for 5 min at 4°C, and chondrocytes were then collected and cultured in DMEM with 10% FBS, 100 U/ml penicillin and 100 μ g/ml streptomycin in a 5% CO₂ atmosphere at 37°C. Chondrocytes at passage 3 were used for all analyses, according to a previous study (32).

Determination of TSA concentration. To select the optimal TSA concentration for the *in vitro* and *in vivo* experiments, cell viability in the presence of TSA was evaluated using the Cell Counting kit-8 (CCK-8; Dojindo Molecular Technologies, Inc., Kumamoto, Japan) assay. Cells were seeded in 96-well plates at 6x10³ cells/well and allowed to adhere overnight. Cells were then cultured in various concentrations of TSA in diluted in serum-free DMEM for 24 h. The medium was then removed, and 10 μ l CCK-8 solution in 90 μ l fresh medium was added to each well. Following incubation for 3 h at 37°C the absorbance was measured at 450 nm using a microplate reader (Bio-Rad Laboratories, Inc., Hercules, CA, USA).

Treatment of rat chondrocytes. Chondrocytes were seeded in 6-well plates at 2x10⁵ cells/well. At 80% confluency, they were serum-starved for 12 h. Cells were then pre-treated with various concentrations of TSA for 2 h, and stimulated with IL-1 β (5 ng/ml) for 12 h or left unstimulated. Cells were harvested for mRNA expression analysis of MMPs and TIMP-1. For determination of total protein and acetylation levels of MMPs and TIMP-1, IL-1 β stimulation was maintained for 24 h.

Reverse transcription-quantitative polymerase chain reaction (RT-qPCR) analysis. Briefly, total RNA was extracted from cell and cartilage samples with TRIzol[®] reagent (Invitrogen; Thermo Fisher Scientific, Inc.). RNA (0.5 μ g) was reversed-transcribed to cDNA using the PrimeScript-RT reagent kit (Takara Bio, Inc., Otsu, Japan). cDNA samples were amplified and quantified by qPCR (ABI Prism 7500; Applied Biosystems; Thermo Fisher Scientific, Inc.) using the SYBR Premix Ex Taq (Takara Bio, Inc.). qPCR protocol cycling conditions were as follows: Holding stage, 95°C for 30 sec; cycling stage, 95°C for 5 sec, then 60°C for 30 sec, for a total of 40 cycles. Stage Primers for MMP-1, MMP-3, MMP-13, TIMP-1, and 18S ribosomal RNA (18S rRNA) genes were obtained from Sangon Biotech Co., Ltd. (Shanghai, China); the sequences are listed in Table I. 18S rRNA served as an internal control. Relative expression was calculated using the 2^{- $\Delta\Delta$ C_q} method (33).

Western blot analysis. Cell and cartilage samples were washed twice with ice-cold phosphate-buffered saline (PBS). Proteins were then extracted using radioimmunoprecipitation assay lysis buffer (Wuhan Boster Biological Technology, Ltd., Wuhan, China) for 30 min. Protein levels were quantified with the Pierce Bicinchoninic Acid Protein assay (Thermo Fisher Scientific, Inc.). Proteins (5 μ g) were resolved by 10% sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) at 110 V for 90 min, and transferred to polyvinylidene difluoride membranes. Following incubation in blocking buffer (5% non-fat milk) for 1 h at room temperature, the membranes were sectioned based on molecular weights, and probed with the following primary antibodies: Rabbit-anti β -actin (catalog no., DW130656; Hang Zhou Dawen Biotech Co., Ltd., Hangzhou, China), rabbit-anti MMP-1 (catalog no., BS1229; Bioworld Technology, Inc., Louis Park, MN, USA), rabbit-anti MMP-3 (catalog no., 17,873-1-AP; ProteinTech Group, Inc., Chicago, IL, USA), rabbit-anti MMP-13 (catalog no., BS6668; Bioworld Technology, Inc.), rabbit-anti TIMP-1 (catalog no., sc-5538; Santa Cruz Biotechnology, Inc., Dallas, TX, USA), and mouse anti-H3 (96C10, catalog no., #3638), mouse anti-H4 (L64C1, catalog no., #2935), rabbit anti-acetyl-H3 (Lys27, catalog no., #8173), and rabbit anti-acetyl-H4 (Lys8, catalog No., #2594; all Cell Signaling Technology, Inc., Danvers, MA, USA). All primary antibodies were diluted 1:1,000. The membranes were washed and incubated with horseradish peroxidase (HRP)-conjugated secondary goat anti-rabbit IgG antibody (catalog no., sc-2004; Santa Cruz Biotechnology, Inc.; 1:5,000) or HRP-conjugated secondary goat anti-mouse IgG antibody (catalog no., sc-2005; Santa Cruz Biotechnology, Inc.; 1:2,000) for 1 h at room temperature. Following washing, signals were detected using Enhanced Chemiluminescence (ECL; Thermo Fisher Scientific, Inc.) and densitometry was performed using the Quantity One version 4.6.2 software (Bio-Rad Laboratories Inc., Munich, Germany).

Animal study. A total of 30 4-week-old male Sprague Dawley rats (weight, 120-140 g), from the Animal Center at Zhejiang University (Hangzhou, China), were used in the present study. Experiments were conducted with the approval of the Zhejiang University Animal Care and Use Committee (Hangzhou, China). Rats were housed under pathogen-free conditions at

Table I. Primer sequences.

Gene	Forward	Reverse
18S rRNA	5'-TTGACGGAAGGGCACCA-3'	5'-CAGACAAATCGCTCCACCAA-3'
MMP-1	5'-GGAACAGATACGAAGAGGAAACA-3'	5'-TGTGGGAATCAGAGGTAGAAGA-3'
MMP-3	5'-GCATTGGCTGAGTGAAAGAGAC-3'	5'-ATGATGAACGATGGACAGATGA-3'
MMP-13	5'-TGAGAGTCATGCCAACAAATTC-3'	5'-CAGCCACGCATAGTCATGTAGA-3'
TIMP-1	5'-GGTTCCTGGCATAATCTGAG-3'	5'-ATCGCTCTGGTAGCCCTTCT-3'

rRNA, ribosomal RNA; MMP, matrix metalloproteinase; TIMP-1, tissue inhibitor of metalloproteinases-1.

room temperature (22-25°C) and 50% humidity, and had a 12-h light/dark cycle. Food and water were offered *ad libitum*. Rats were randomly divided into three groups: i) Control, ii) OA and iii) TSA-treated. Following anesthesia with 2% pentobarbital sodium (0.1 ml/100 g; Sigma-Aldrich) (34), rats in the OA and TSA-treated groups received an intra-articular injection of mono-iodoacetate (MIA; 1 mg in 50 μ l PBS; Sigma-Aldrich) into the knee joint (35). Rats in the control group were injected with 50 μ l PBS. Three days later, rats in the TSA-treated group were bilaterally intra-articularly injected into the knees with 50 μ l TSA (0.2 μ M), while the other groups received 50 μ l PBS injected intra-articularly. These injections were repeated once a week for four weeks. All of the rats were sacrificed seven days following the final injection. Mice were anaesthetized with 2% pentobarbital sodium (0.1 ml/100 g; Sigma-Aldrich) and cervical vertebrae were dislocated. Then femoral condyles were collected for gene and protein expression analysis and histological assessment.

Histological analysis. Specimens were fixed in 4% paraformaldehyde in 0.1 M PBS for 72 h (31) and decalcified with 10% buffered formic acid at room temperature for two months. Specimens were embedded in paraffin wax and cut into 5 mm sections. The sections were stained with Safranin-O and fast green (36). The Mankin scoring system was used for blinded histological assessment (36).

Statistical analysis. Data are expressed as the mean \pm standard deviation. Comparisons were made using an unpaired Student's *t*-test and one-way analysis of variance, followed by Dunnett's analysis. $P < 0.05$ was considered to indicate a statistically significant difference.

Results

Assessment of TSA concentrations. To select the optimal concentrations of TSA for *in vitro* and *in vivo* experiments, a CCK-8 assay was performed to evaluate various TSA concentrations. As presented in Fig. 1, TSA concentrations $< 0.4 \mu$ M exerted no significant effect on chondrocyte viability at 24 h (0.8 vs. 0 μ M TSA; $P = 0.03$), and therefore were selected for use in subsequent experiments.

TSA decreases MMP and increases TIMP-1 expression levels in chondrocytes. RT-qPCR revealed that treatment of chondrocytes with IL-1 β decreased mRNA expression levels

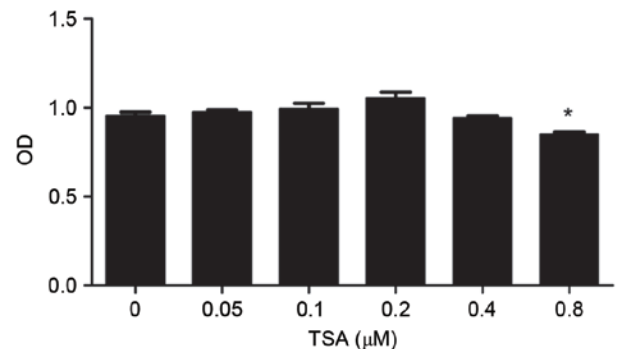


Figure 1. Determination of optimal TSA concentrations for *in vitro* and *in vivo* experiments. Chondrocytes were treated with increasing concentrations of TSA (0.05-0.8 μ M) for 24 h. Cell viability was determined using a cell counting kit-8 assay. * $P < 0.05$ vs. 0 μ M TSA. Data are presented as the mean \pm standard deviation. TSA, trichostatin A; OD, optical density.

of MMP-1, MMP-3 and MMP-13 (Fig. 2A). By contrast, mRNA expression levels of TIMP-1 increased following IL-1 β administration. Notably, IL-1 β -mediated regulation of each of these genes was significantly attenuated by increasing concentrations of TSA. While mRNA expression levels of MMP-1, MMP-3 and MMP-13 were not affected by treatment with TSA alone, TIMP-1 was significantly upregulated in cells incubated with TSA alone (0.05, 0.1, 0.2 vs. 0 μ M TSA; $P = 0.034$, < 0.001 , < 0.001 , respectively; Fig. 2B). Examinations of protein expression levels of MMPs and TIMP-1 were consistent with the RT-qPCR data. That is, TSA significantly attenuated IL-1 β -mediated regulation of MMP-1, MMP-3, MMP-13 and TIMP-1 protein expression levels (MMP-1, MMP-2, MMP-13 and TIMP-1 incubated with 0.2 vs. 0 μ M TSA; $P < 0.001$, $P = 0.004$, 0.006 and 0.004, respectively; Fig. 2C). Treatment of chondrocytes with TSA alone increased TIMP-1 protein expression levels, which was consistent with the transcriptional analysis ($P < 0.05$; Fig. 2D).

TSA treatment delays knee joint degeneration in the rat OA model and is accompanied by decreased MMP and increased TIMP-1 mRNA and protein expression levels in cartilage. The gross appearance of femoral condyles collected from the various treatment groups were examined (Fig. 3A). The condyle surface in the control group was intact and glossy. By contrast, the condyle surface of rats in the OA group revealed gross morphological alterations, including defects, osteophytes and tissue adhesions. Notably,

Table II. Histological scoring of articular cartilage.

Parameter	Control group	OA group	TSA-treated group
Structural changes	0.15±0.04	2.60±0.16 ^a	0.83±0.09 ^b
Cellular changes	0.19±0.03	2.04±0.26 ^a	0.86±0.15 ^b
Safranin-O staining	0.54±0.09	2.68±0.72 ^a	1.18±0.16 ^b
Tide mark	0.14±0.04	0.15±0.04	0.14±0.03
Sum of score	1.02±0.18	7.47±1.07 ^a	3.01±0.41 ^b

Mankin scores were positively correlated with OA severity. Using Mankin's histological scoring principles, a total score of 13 is possible. Structural changes, scored from normal to severely damaged (0-3); Cellular changes, scored from normal to severe hyperplasia (0-3); Safranin-O staining, scored from normal to non-staining (0-4); Tide mark, scored from normal to fuzzy (0-3). Data are expressed as the mean ± standard deviation. ^aP<0.05 vs. control; ^bP<0.05 vs. OA. OA, osteoarthritis; TSA, trichostatin A.

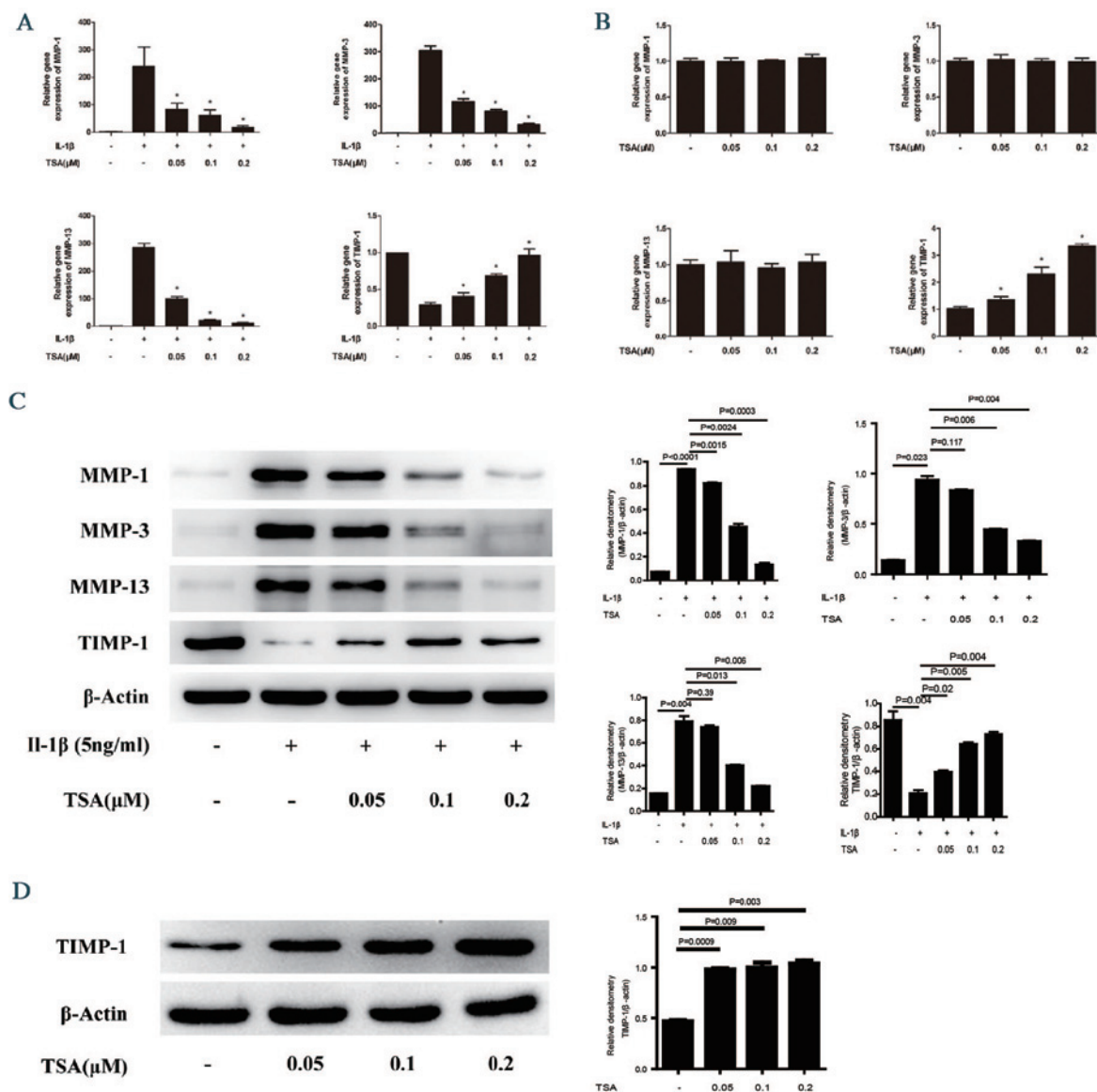


Figure 2. Effect of TSA on MMP-1, MMP-3, MMP-13, and TIMP-1 mRNA and protein expression levels in chondrocytes. (A) Chondrocytes were pretreated with increasing concentrations of TSA for 2 h, followed by stimulation with 5 ng/ml IL-1 β for 12 h. ^aP<0.05 vs. IL-1 β alone. (B) Chondrocytes were treated with increasing concentrations of TSA for 12 h. ^aP<0.05 vs. untreated. Total RNA was isolated and reverse transcription-quantitative polymerase chain reaction was performed to determine relative mRNA expression levels. Levels were calculated using the $2^{-\Delta\Delta C_t}$ method and expressed as the mean ± SD. (C) Chondrocytes were pretreated with increasing concentrations of TSA for 2 h, followed by stimulation with 5 ng/ml IL-1 β for 24 h. (D) Chondrocytes were treated with increasing concentrations of TSA for 24 h. Proteins were harvested for western blot, and bands were quantified using densitometry and quantity one software. Data are expressed as the mean ± SD. TSA, trichostatin A; MMP, matrix metalloproteinase; TIMP-1, tissue inhibitor of metalloproteinases-1; IL-1 β , interleukin-1 β ; SD, standard deviation.

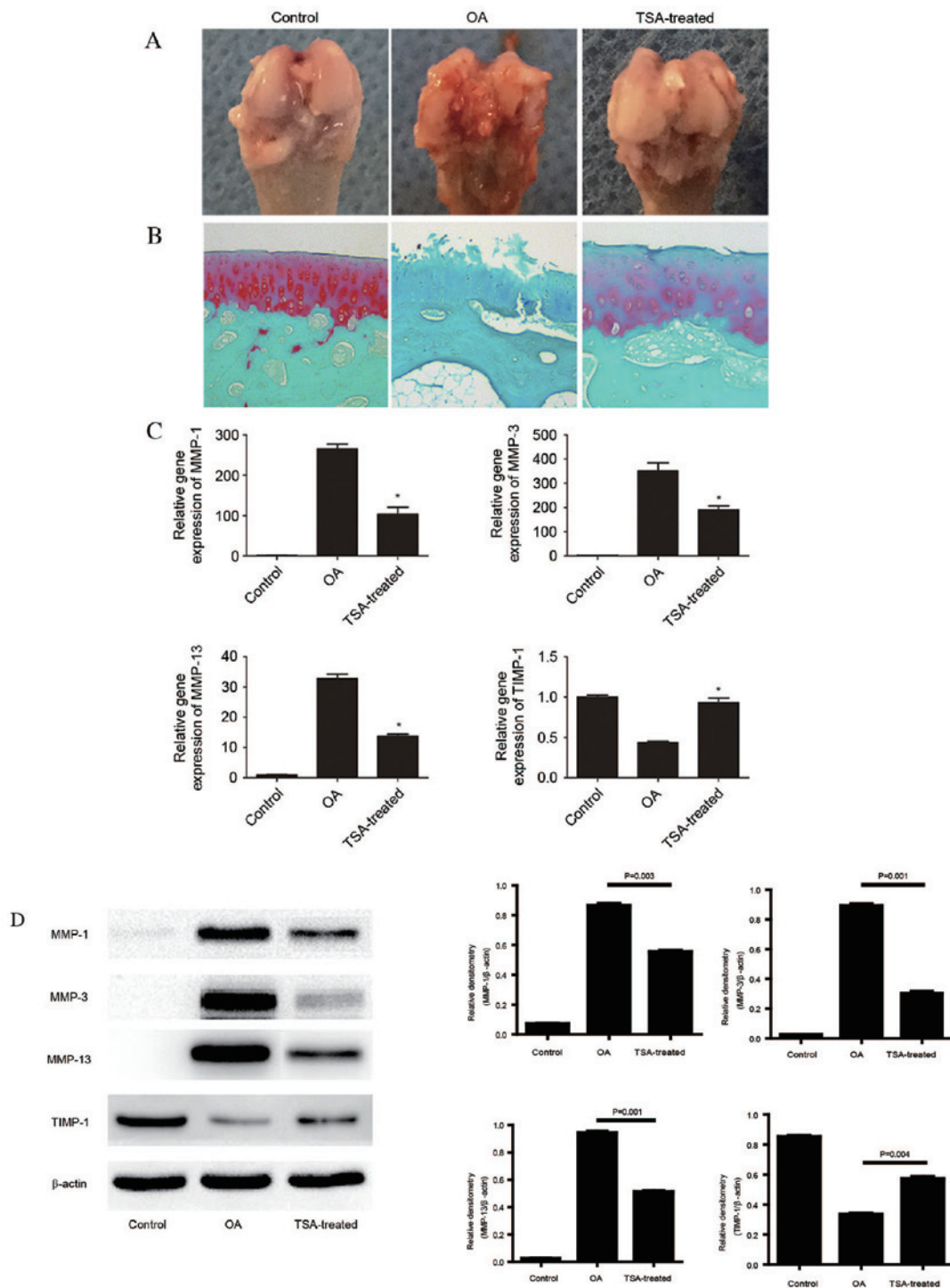


Figure 3. Effect of TSA on the rat OA model and on MMP-1, MMP-3, MMP-13 and TIMP-1 mRNA and protein expression levels in cartilage. An *in vivo* study was performed using Sprague Dawley rats, which were randomly divided into three groups (n=10/group): Control, OA and OA with TSA treatment. (A) Representative images of macroscopic observations. (B) Safranin-O/fast green staining (magnification, x100). (C) Total RNA was isolated from cartilage, and reverse transcription-quantitative polymerase chain reaction was performed to determine relative mRNA expression levels. Levels were calculated using the $2^{-\Delta\Delta C_q}$ method and expressed as the mean \pm SD. * $P < 0.05$ vs. OA. (D) Proteins were extracted from cartilage for western blot analysis, and bands were quantified using densitometry and quantity one software. Data are expressed as the mean \pm SD. TSA, trichostatin A; OA, osteoarthritis; MMP, matrix metalloproteinase; TIMP-1, tissue inhibitor of metalloproteinases-1; SD, standard deviation.

a marked reversion to normal morphology was observed in the TSA-treated group. Morphological analysis was accompanied by Safranin-O/fast green staining of cartilage (Fig. 3B). Compared with the control group, Safranin-O staining was largely absent in the OA group (demonstrated

by the absence of red staining in the OA group image). TSA administration markedly restored Safranin-O staining, indicative of cartilage retention, although not to the level of the control group. TSA treatment markedly attenuates numerous changes associated with OA progression, as measured by the

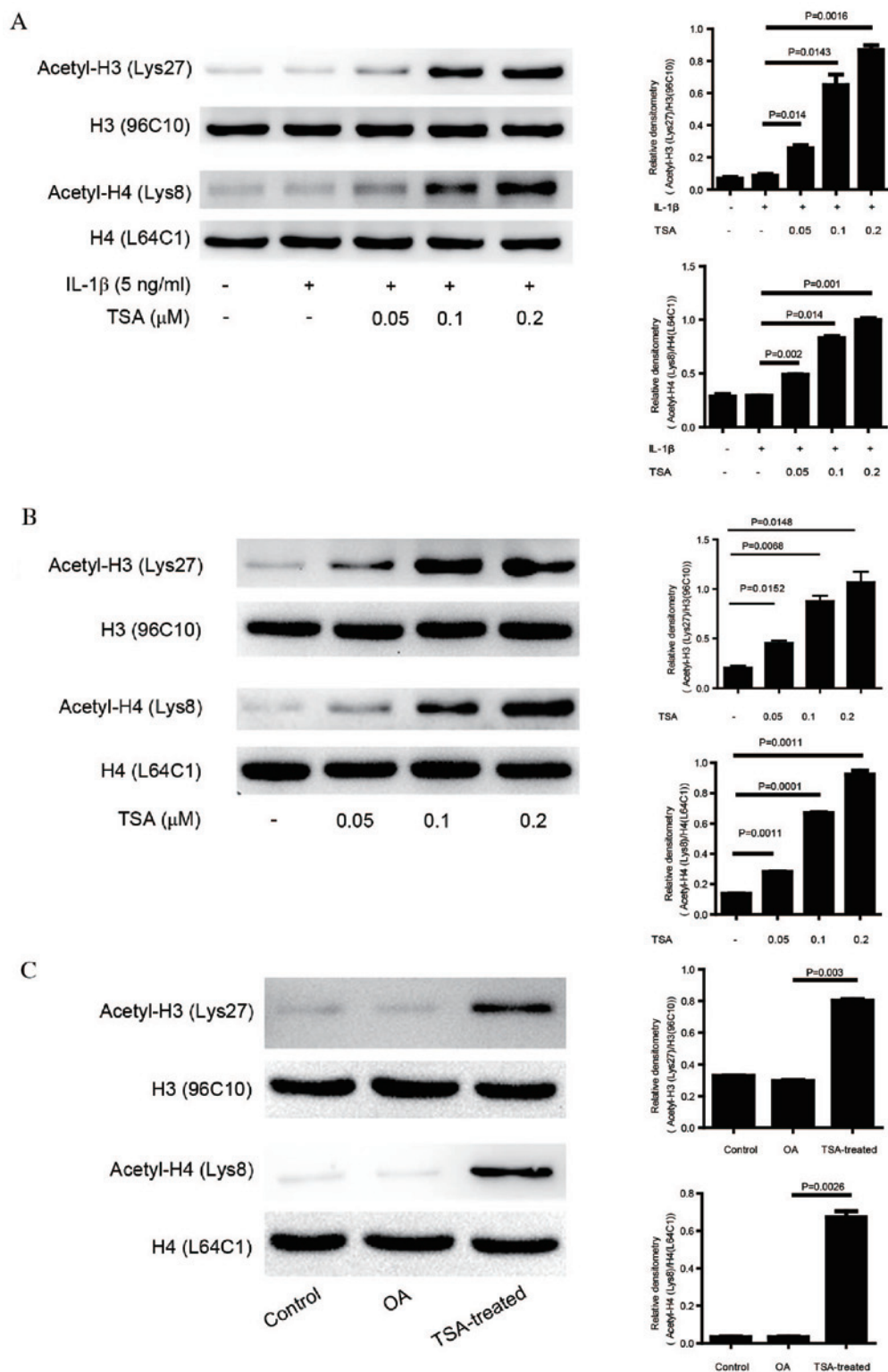


Figure 4. Effect of TSA on acetylation levels in chondrocytes and cartilage. (A) Chondrocytes were pretreated with increasing concentrations of TSA for 2 h, followed by stimulation with 5 ng/ml IL-1 β for 24 h. (B) Chondrocytes were treated with increasing concentrations of TSA for 24 h. (C) Proteins were extracted from the cartilage of control, OA, and OA with TSA treatment rats. Western blot protein bands were quantified using densitometry and quantity one software. Data are expressed as the mean \pm standard deviation. TSA, trichostatin A; IL-1 β , interleukin-1 β ; OA, osteoarthritis; H, histone.

Mankin scoring system for histological analyses (Table II). Structural and cellular changes were analyzed and assigned histological scores. The sum of the scores was significantly increased in the OA group (7.47 ± 1.07) compared with the

control (1.02 ± 0.18 ; $P=0.005$). Notably, and consistent with our earlier findings (4,30), TSA treatment promoted reversion back to normal, with a sum of 3.01 ± 0.41 ($P=0.003$ vs. OA group).

Molecular analyses of *in vivo* samples were performed. Results from RT-qPCR revealed that mRNA expression levels of MMP-1, MMP-3 and MMP-13 were significantly upregulated, and TIMP-1 was downregulated, in cartilage from OA rats compared with control rats (Fig. 3C). Consistent with the *in vitro* observations, TSA treatment significantly attenuated the OA-associated alterations in expression levels of each of these genes ($P < 0.05$). Alterations in protein expression levels of MMPs and TIMP-1 were consistent with the RT-qPCR data ($P < 0.05$; Fig. 3D).

TSA increases acetylation levels in chondrocytes and cartilage. Data from the *in vitro* and *in vivo* experiments indicate that TSA may be beneficial in the treatment of OA. As TSA is an HDACi, alterations in levels of acetylation in cells (Fig. 4A and B) and in the OA animal model (Fig. 4C) were examined. Levels of acetylation at H3K27 and H4K8 were not significantly altered in cells treated with IL-1 β or in cartilage from the OA group, indicating that acetylation at these specific residues is not involved in OA progression. Notably, TSA administration significantly increased acetylation at these residues, providing evidence of functional TSA treatment ($P < 0.05$).

Discussion

The results of the present study revealed that TSA significantly attenuated the IL-1 β -induced upregulation of MMP-1, MMP-3 and MMP-13, and the downregulation of TIMP-1. In addition, the level of acetylation increased in a TSA dose-dependent manner, independent of IL-1 β treatment. Consistent with these observations *in vitro*, increased expression levels of MMP-1, MMP-3 and MMP-13, and decreased TIMP-1 expression levels, were detected in the OA rat model compared with control rats. These findings suggest that the altered MMP/TIMP-1 ratio is an important contributing factor in OA progression. In addition, TSA treatment demonstrated a beneficial effect *in vivo*, possibly via an increase in TIMP-1 expression levels. Furthermore, levels of acetylated histones, H3 and H4 were increased in the TSA-treated group compared with the OA and control groups. These observations *in vitro* and *in vivo* suggest that TSA treatment alters the MMP/TIMP-1 ratio to protect against OA. These conclusions are supported by the function of TIMP-1 as an important inhibitor of MMP activity during the process of cartilage degeneration (37). MMPs degrade various extracellular matrix components, including collagen type II, which is a key pathological feature of OA (38).

In the present study, the role of histone acetylation in OA progression and following TSA treatment was examined. Analyses were limited to acetylation of H3K27 and H4K8. Acetylation of these residues was not identified to be significantly altered *in vitro* by IL-1 β treatment or *in vivo* in the OA rat model. These findings suggest that altered histone acetylation of these specific residues is not involved in OA development and progression. Alternatively, alterations in acetylation of these residues may occur only at key genes involved in OA and, as such, may not be observable at the global level when probed by western blot analysis. However, increased acetylation of the two residues following TSA treatment *in vitro* and *in vivo* was observed. In addition, while TSA treatment increased TIMP-1

expression levels, it had no direct effect on expression levels of MMP-1, MMP-3 or MMP-13. This suggests that TSA attenuates the OA-associated increase in MMPs via upregulating TIMP-1, rather than by directly affecting MMP transcription. Previous studies have demonstrated that histone hyperacetylation is associated with anti-inflammatory properties (39) via altered gene expression (40). A similar mechanism of action may underlie the anti-OA effect of TSA that was observed in the present study. However, this hypothesis requires additional investigation. In future experiments, analysis of a larger panel of histone acetylation modifications and chromatin immunoprecipitation would be performed to determine which genes are marked by histone modifications. This would facilitate the determination of the precise underlying mechanism of action of TSA in the protection against OA.

The results of the present study revealed that TSA treatment does not significantly alter transcript levels of MMP-1, MMP-3 and MMP-13. These findings suggest that MMP transcription in this setting is not directly linked to histone acetylation. Consistent with this, a previous study demonstrated that the IL-1 β -mediated production of MMPs primarily occurs via the NF- κ B signaling pathway (41). However, in our previous study on the effect of TSA on the NF- κ B signaling pathway in chondrocytes treated with IL-1 β , no significant alterations in NF- κ B activity were observed (32). Additional experiments are therefore required to determine the exact underlying mechanism by which the anti-OA effects of TSA are exerted. In conclusion, the results of the present study demonstrated that TSA protects against OA formation in chondrocytes *in vitro* and in an *in vivo* OA rat model. It is proposed that TSA functions via the upregulation of TIMP-1 and subsequent inhibition of MMP activity. The findings of the present study suggest that TSA may be a potential therapy for the treatment of OA, and that histone acetylation may be an important contributing factor in this disease.

Acknowledgements

The present study was supported by the National Natural Science Foundation of China (grant nos. 81201429 and 81371996).

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