

Recent Progress on Therapeutic Vaccines for Breast Cancer

Lianru Zhang¹, Xipeng Zhou², Huizi Sha¹, Li Xie^{1*} and Baorui Liu^{1*}

¹ The Comprehensive Cancer Centre of Drum Tower Hospital, Medical School of Nanjing University & Clinical Cancer Institute of Nanjing University, Nanjing, China, ² Department of oncology, Yizheng People's Hospital, Yangzhou, China

Breast cancer remains the most frequently diagnosed malignancy worldwide. Advanced breast cancer is still an incurable disease mainly because of its heterogeneity and limited immunogenicity. The great success of cancer immunotherapy is paving the way for a new era in cancer treatment, and therapeutic cancer vaccination is an area of interest. Vaccine targets include tumor-associated antigens and tumor-specific antigens. Immune responses differ in different vaccine delivery platforms. Next-generation sequencing technologies and computational analysis have recently made personalized vaccination possible. However, only a few cases benefiting from neoantigen-based treatment have been reported in breast cancer, and more attention has been given to overexpressed antigen-based treatment, especially human epidermal growth factor 2-derived peptide vaccines. Here, we discuss recent advancements in therapeutic vaccines for breast cancer and highlight near-term opportunities for moving forward.

OPEN ACCESS

Edited by:

Vajihe Akbari, Isfahan University of Medical Sciences, Iran

Reviewed by:

Rodney Macedo Gonzales, Albert Einstein College of Medicine, United States Chiara Focaccetti, Università di Roma Tor Vergata, Italy

*Correspondence:

Baorui Liu baoruiliu@nju.edu.cn Li Xie sherry01130@163.com

Specialty section:

This article was submitted to Breast Cancer, a section of the journal Frontiers in Oncology

Received: 27 March 2022 Accepted: 11 May 2022 Published: 06 June 2022

Citation:

Zhang L, Zhou X, Sha H, Xie L and Liu B (2022) Recent Progress on Therapeutic Vaccines for Breast Cancer. Front. Oncol. 12:905832. doi: 10.3389/fonc.2022.905832 Keywords: breast cancer, cancer vaccines, cancer immunotherapy, clinical trials, concurrent therapies

INTRODUCTION

Breast cancer (BC) is the leading cause of cancer worldwide (1). Although there has been an increase in the overall survival rate in BC because of improvements in early-stage diagnosis and targeted therapies, almost all metastatic tumors develop drug resistance and cannot be cured. It is still a difficult problem to reduce the recurrence rate of early breast cancer and to prolong the survival time of advanced breast cancer. Immune-based interventions could be a beacon of hope to decrease morbidity and mortality of cancer. Although immune checkpoint inhibitors (ICIs) have been proven to increase the survival rate in lung cancer, melanoma, gastric cancer and so on, the indications of ICIs for the treatment of BC are only focused on first-line and neoadjuvant therapy for triple-negative breast cancer (TNBC) (2) to date.

The tumor microenvironment (TME) plays a crucial role in the recognition and prevention of cancer and early eradication. The TME may also interact with tumor cells and promote the progression of cancer. The immunoediting hypothesis describes the dynamic interaction between the immune system and tumor cells in three phases: elimination phase, equilibrium phase and escape phase (3). Tumor cells that avoid immune recognition and elimination steps enter the escape phase and present a clinically detectable tumor. The advantage of active immunotherapy is to develop a protective effect against tumor tissue, modifying the immune microenvironment and resetting the immune system to an antitumor surveillance status. Therapeutic cancer vaccines led by neoantigens are hotspots of active immunotherapy. Combination strategies with ICIs have shown clinical benefits in multiple types of cancer (4, 5). To date, only one vaccine named sipuleucel-T has

1

been approved by the FDA and is used to treat metastatic castration-resistant prostate cancer in a limited group of nearly asymptomatic patients (6). No BC vaccine has been approved for clinical use. BC is a heterogeneous disease and can be classified into 4 common groups: luminal A, luminal B, human epidermal growth factor 2 (HER2)-positive, and TNBC (7). BC is traditionally considered a poorly immunogenic tumor. However, recently published data on TNBC have shown that a significant number of tumor infiltrating lymphocytes infiltrate TNBC tissues (8), indicating that an immunotherapeutic approach may be suitable for this hard-to-treat malignancy. A series of clinical trials for TNBC vaccines are underway. In addition, increasing numbers of clinical trials are being conducted demonstrating that vaccination is capable of inducing an antitumor-specific response in BC. In this review, we discuss recent progress on therapeutic vaccines from the perspective of tumor development and clinical data, and a blueprint for personalized vaccines is also presented.

SPECTRUM OF VACCINE TARGETS

Therapeutic tumor antigens are divided into two main categories: tumor-associated antigens (TAAs) and tumor-specific antigens (TSAs) (9) (**Figure 1**). TAAs include tumor germline antigens, tumor differentiation antigens and overexpressed antigens (10). Tumor germline antigens, or cancer testis antigens, are expressed at high levels in the germinal cells of the testis, ovaries, and placenta and are not expressed in somatic cells under normal conditions (11). They are expressed in malignant cells of various cancer types, including BC. In BC, the expression of a number of cancer testis antigens has been reported, such as MAGE-A1 (12), NY-ESO1 (13) and KK-LC-1 (14). Serum antibodies against cancer testis antigens can be detected as useful biomarkers for predicting the clinical benefits of immunotherapy (14-16). Tumor differentiation antigens are proteins expressed in tumor cells and in normal tissue from which the tumor originates, such as Melan-A/Mart-1 (17), gp100 (18), PSA (19), CEA (20) and NY-BR-1 (21, 22). Overexpressed antigens are proteins expressed at low levels in normal cells and at high levels in cancer cells. The most common overexpressed antigens targeted in BC are HER2 (23), MUC-1 (24), hTERT (25) and survivin (26). TAA-based vaccines must be sufficiently immunogenic to activate the remaining low-affinity TAA-reactive T cells because central and peripheral immune tolerance mechanisms have removed T cells with strong TAA affinity.

TSAs are expressed specifically in tumor cells, mainly including oncoviral antigens and neoantigens (27). Neoantigens are products of genomic alterations and consist of simple point mutations that change single amino acids, frameshift insertion or deletion mutations, splice-site alterations that lead to exon skipping, structural alterations that lead to the formation of fusion proteins and other forms of collateral damage (28). Although there are thousands of genomic alterations in the process of tumor formation, only a handful of neoantigens succeed in eliciting antitumor immune responses. BC shows an intermediate genomic mutational load, and only a few cases benefiting from neoantigen-

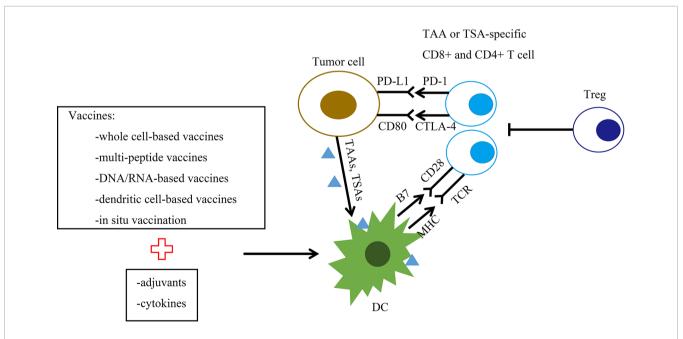


FIGURE 1 | Graphic representation of the therapeutic breast cancer vaccine platforms and their mechanism of action. The figure summarizes the spectrum of vaccine targets and vaccine platforms for breast cancer. Vaccine platforms are whole cell-based vaccines, multipeptide vaccines, DNA/RNA-based vaccines, dendritic cell (DC)-based vaccines and *in situ* vaccination. DCs present processed tumor-associated antigen (TAA) or tumor-specific antigen (TSA) to CD4+ and CD8+ T cells. This interaction generates TAA/TSA-specific effector T cells, leading to the killing of tumor cells. B7, B7 protein; CD28, T cell costimulatory molecule CD28; CTLA-4, cytotoxic T lymphocyte-associated antigen 4; DC, dendritic cell; MHC, major histocompatibility complex; PD-1, programmed cell death protein 1; PDL-1, programmed death ligand 1; TAAs, tumor-associated antigens; TSAs, tumor-specific antigens; TCR, T cell receptor; Treg, regulatory T cell.

based treatment have been reported in BC (29). Since TNBC is recognized as a potential suitable subtype for immunotherapy, clinical trials of neoantigens are enrolling TNBC patients to evaluate the safety and induction of specific T cell responses. Clinical trials using autologous dendritic cells (DCs) pulsed with tumor-specific neoantigen (NCT04105582) or neoantigen DNA vaccine administered with durvalumab (NCT03199040) or personalized synthetic long peptide (SLP) neoantigen vaccine administered with durvalumab and nab-paclitaxel (NCT03606967) are currently enrolling TNBC patients. The neoantigen prediction process includes identifying tumor-specific somatic mutations and predicting major histocompatibility complex (MHC)-binding epitopes. Whole-exome sequencing is performed using tumor biopsy specimens and nonmalignant tissue samples to identify tumor-specific somatic mutations (30, 31). Tumor and germline DNA are compared to exclude germline mutations, while RNA sequencing provides additional information on mutated genes and confirms the mutation calls (32-34). Owing to human leukocyte antigen (HLA) restriction, various algorithmbased computational approaches have been developed to predict the binding of a tumor antigen to MHC molecules (35, 36). Peptides predicted with a moderate-to-strong HLA-binding affinity (IC50 < 150 nmol/l) are considered more likely to induce CD8+ T cell responses. Mass spectrometry-based immunopeptidomics can be used to identify neoantigens or to validate those predicted by in silico strategies. Recently, a new strategy based on using signaling and antigen-presenting bifunctional receptor (SABR) libraries was developed, enabling the identification of specific TCR-pMHC interactions (37).

In addition to TAAs and TSAs, multiple TME-targeting vaccinebased clinical trials are underway for patients with BC (38). Resident cells in the TME are likely more genomically stable than tumor cells. Pathological angiogenesis in the vascular TME can suppress effective immunotherapies. Multiple strategies targeting whole-cell endothelial cells (39), tumor blood vessel antigens (40), epidermal growth factor receptor (EGFR) (41), CD105 (42), platelet-derived growth factor receptor (PDGFR)- β (43) and vascular endothelial growth factor receptor (VEGFR) (44) have been tested in preclinical models of BC. A phase I study of pulsed DCs with tumor blood vessel antigens was completed recently (NCT02479230). Cancer-associated fibroblasts of the TME are vaccine targets as well. However, cancer-associated fibroblast vaccine strategies are all in the preclinical stage (45-47). Mads Hald Andersen et al. (48) designed an innovative investigational approach to target immune inhibitory pathways in the TME, modulating immune regulation. Therapeutic vaccination with long peptide epitopes is derived from proteins including indoleamine 2,3-dioxygenase (IDO), tryptophan 2,6-dioxygenase, arginase, and programmed death ligand 1 (PD-L1). Endogenous anti-regulatory T cells are activated because they recognize these peptides, and these pro-inflammatory cells are attracted to the TME, potentially altering tolerance to tumor antigens. Vaccinations against IDO or PD-L1 have been proven to be safe in clinical trials. Tryptophan 2,6-dioxygenase (TDO) is another enzyme involved in tryptophan degradation in the TME and is expressed in many cancers, including breast cancer, making it an interesting

target for therapeutic vaccinations against the TME for BC. Vaccines are also currently being developed to target gene products associated with epithelial-mesenchymal transition (EMT) and cancer cells with stem-like characteristics (49, 50).

VACCINE DELIVERY PLATFORMS

Diverse vaccine platforms have now been evaluated in clinical trials, including whole cell-based vaccines, multipeptide vaccines, DNA/RNA-based vaccines, dendritic cell-based vaccines and *in situ* vaccination (**Table 1**).

Whole Cell-Based Vaccines

Whole cell-based vaccines are derived from autologous or allogenic tumor cells (56). Immunizing BC patients with tumor cells isolated from the patient can circumvent the problems associated with antigen selection and epitope prediction. In addition, whole cellbased vaccines present the patient's immune system with a wide variety of TAAs as immunogens. However, whole cell-based vaccines have shown relatively poor immunogenic potential (57). The immunogenicity can be increased by engineering tumor cell lines to secrete granulocyte-macrophage colony stimulating factor (GM-CSF), combined with strong adjuvants or cytokines (58, 59). In addition, whole cell-based vaccines in combination with chemotherapy may also exert synergistic antitumor effects. Autologous tumor cell vaccines (ATCVs) present a unique set of antigens, such as particular point mutations or fusion gene products, from a given patient's own tumor (60-62). These antigens could help to launch a polyclonal response against a variety of tumor cells. However, the generation of ATCVs is patient specific with high complexity and high cost. Allogenic tumor cell vaccines, which typically contain two or three established human tumor cell lines, can be used as an alternative for the development of cell-based vaccines (56). In a phase I clinical trial enrolling 28 patients with stable metastatic breast cancer (mBC), the efficacy of a combination therapy using an allogenic GM-CSF-secreting BC vaccine along with chemotherapy was investigated (63). The vaccine was formulated from two HER2/ neu+ mammary adenocarcinoma BC cell lines, SKBR3 and T47D. This vaccine was administered either alone or in sequence with common chemotherapeutic agents, such as cyclophosphamide and doxorubicin. The results suggest that the vaccine alone or in sequence with low-dose chemotherapy could induce an effective immune response. In another phase I study, a human leukocyte antigen (HLA)-A2+, HER2/neu(+) allogeneic MDA-MB-231 BC cell line was modified to express the costimulatory molecule B7-1 (CD80) and used as a vaccine to treat stage IV BC patients (64). Although this immunization strategy proved to induce tumorspecific immune responses in a minority of patients, no significant tumor regression was observed. In a single-arm feasibility study, an allogeneic HER2+ GM-CSF-secreting BC vaccine was given with low-dose cyclophosphamide and weekly trastuzumab in 20 patients with HER2+ mBC (65). This vaccination regimen was safe and demonstrated clinical effects in terms of objective response rate (ORR), progression-free survival (PFS), and

TABLE 1 | Comparison of different vaccine platforms.

Vaccine platforms	Mechanisms	Advantages	Disadvantages	Ref
Whole cell- based vaccines	Whole tumor cell lysates can be prepared by hypochlorous acid, ultraviolet B ray-irradiation, repeat cycles of freezing and thawing or hyperthermia	All tumor cells express a wide range of tumor- associated antigens Gene sequencing and bioinformatics predictive screening are not required Diminishes the chance of tumor escape	Complex and expensive production The immunogenicity is relatively poor	(51)
Multipeptide vaccines	Peptide vaccines contain tumor-specific epitopes that can be taken up and processed by antigen-presenting cells to activate T cell immune responses	Stable Safe Can be inoculated repeatedly Long peptides can stimulate both CD4+ and CD8+ T cell responses	The immunogenicity of synthetic peptide-based vaccines can be significantly affected by the delivery process	(52)
DNA/RNA- Based Vaccines	<i>In vitro</i> transcribed RNA or plasmid DNA encoding cancer antigens is introduced into the body, and cancer antigens are expressed by the host to induce antitumor response	Rapid and inexpensive production Mimics viral infection DNA vaccines have flexible platform for molecule engineering RNA vaccines have intrinsic adjuvant properties	RNA vaccine is susceptible to extracellular degradation by RNAses DNA vaccine has theoretical risk of host genome integration, relatively low immunogenecity	(53)
Dendritic cell-based vaccines	DC cells are stimulated with cytokines <i>in vitro</i> to become mature DCs upregulating costimulatory molecules, and mature DCs loaded with antigens migrate to lymph nodes resulting in the subsequent specific immune responses	Bypass conventional antigen presentation pathways	Time-consuming personalized process Less practical Hard to preserve	(54)
In situ vaccination	Manipulation of intratumoral myeloid cells by injecting immunomodulators and local ablative therapies which are used to release tumor antigens from the therapy-killed tumor cells such as radiation or combination with vaccines	Simple and cost-effective Minimal side effects Minimizes immune escape Adjuvant delivery is feasible and flexible	Requirement for intratumoral injection	(55)

overall survival (OS), with a trend toward longer PFS and OS in HER2-specific T-cell responders.

Peptide Vaccines

The advantages of peptide vaccines include ease of synthesis and storage, safety, cost-effectiveness, and tolerable side effects. The great limitation for peptide-based vaccines is the possibility of insufficient immunogenicity, which makes a great need for a suitable adjuvant to produce an efficient response. The expression of antigen epitopes within the tumor bed can be heterogeneous, while the immune response may be limited to a few epitopes. Multipeptide vaccines formulated from MHC class I-restricted TAAs are being tested for their antigen-specific immune response in clinical trials (66-70). Peptides with epitopes can bind directly to MHC class I molecules on the surface of antigen-presenting cells without cross-presentation, but they often result in only low-level, short-lived responses without the help of CD4+ T cells. CD4+ T cells can enhance the tumoricidal activity of other antitumor effector cells, such as CD8+ T cells and macrophages. Some CD4+ subsets influence angiogenesis to facilitate the infiltration of CD8+ T cells, in addition to direct cytotoxic functions (71). There are attempts to activate both CD4+ and CD8+ T cells by using multivalent

synthetic long peptides (SLPs) containing both MHC class I and class II epitopes (72). SLP vaccines offer several advantages. They are not able to bind directly to MHC class I so that they have to be processed by DCs (73). SLP vaccines increase the duration of in vivo epitope presentation in the antigen-draining lymph node (74), which is shown to be important for clonal expansion (75) and for interferon- γ production by CD8+ T cells (76), and harbor both CD4+ and CD8+ T cell epitopes, ensuring a balanced CD4/ CD8 response. Some well-designed peptide vaccines will be discussed in the 4th part of this review. In addition, delivery systems have been applied to improve antitumor immunity. Among them, nanomaterials, such as liposomes, micelles, dendrimers, microneedles, proteins, polymer-based conjugates, the B-subunit of Shiga toxin (STxB), and polyactin A (PAA), are under investigation to convey and release antigens and immunostimulatory molecules (77).

DNA/RNA-Based Vaccines

DNA or RNA-based vaccines are easy to design and can encode multiple epitopes. DNA vaccines have good stability and can be rapidly and easily modified. Plasmid DNA vaccines can be integrated with additional immune modulators to elicit a maximal immune response (78). Most DNA-based cancer vaccine studies have targeted TAAs, such as HER2/neu and mammaglobin-A (MAM-A), in BC. The first clinical trial of a HER2/neu DNA vaccine evaluated the efficacy and tolerability of the vaccine in humans. The HER2/neu DNA vaccine was administered with low doses of interleukin-2 (IL-2) and GM-CSF in mBC patients in a pilot clinical trial, even though no significant T cell response was elicited (79). Currently, two phase I clinical trials of HER2/neu DNA vaccines are active (NCT00393783 and NCT00436254). The MAM-A DNA vaccine was also investigated in mBC in a phase I clinical trial. This vaccine was safe and succeeded in eliciting MAM-Aspecific CD8+ T cell responses. PFS was improved in vaccinated patients, although the sample size was low (n=14) (80). Additionally, a clinical trial using a neoantigen DNA vaccine to treat TNBC was launched (NCT03199040). RNA vaccines are designed to enter the cytosol and thus avoid safety concerns related to integration into the host cell genome. RNA-based vaccines have an inherent function through Toll-like receptor 3 (TLR3), TLR7 and TLR8 stimulation to provide an adjuvant effect. However, RNA is very unstable, so delivery systems such as nanoparticles and liposomes are challenging. Viral vectors can be used to deliver nucleic acid vaccines to enter the cytosol. However, the production of antibodies against viral vectors attenuates the efficiency. PANVAC (containing transgenes for CEA, MUC-1 and 3 T cell costimulatory molecules) is a well-studied poxviral vaccine. For the 12 mBC patients, 5 patients had stable disease (SD) by RESIST lasting \geq 4 months, with one patient having a complete response (CR) and remaining on study for ≥ 37 months (81).

Dendritic Cell-Based Vaccines

DCs are professional antigen-presenting cells that can process exogenous and endogenous antigens and present them to stimulate naïve T lymphocytes through the MHC I and II pathways. Therefore, DCs play a crucial role in the initiation of the primary response and induction of the antitumor-specific immune response. Most cancer vaccines are greatly dependent on the activation of DCs. Peptide-pulsed DCs have superiority in inducing antitumor responses compared to peptide vaccines with adjuvants (82). In a pilot study, autologous DCs were pulsed with HER2/neu- or MUC1-derived peptides to generate a DC-based vaccine. Ten patients suffering from advanced BC and ovarian cancer showed a strong immunogenic response with no side effects (83). A HER2 intracellular domain (ICD) proteincontaining DC vaccine was tested in disease-free BC patients. Six patients out of seven had circulating anti-HER2 ICD antibodies, and all patients were alive and disease free at 4.6-6.7 years of follow-up (84). Autologous DCs were also pulsed with patient-derived tumor cells or cell lysates to facilitate a strong immunogenic response (85-87). However, ex vivo generation of DCs is complicated, and it is costly and timeconsuming to generate the large number of DCs required for vaccination. The demanding production process of DC vaccines and lack of improvement in clinical benefits limit their application in the clinic.

In Situ Vaccination

In situ vaccination (ISV) refers to inducing and stimulating an immune response specially at the tumor site (88). ISV uses the

tumor itself as the antigen source and should be defined as a treatment process or strategy. There are several advantages of ISV. It is simple and cost-effective with minimal side effects, and it utilizes all tumor antigens in the tumor which minimizes immune escape. There is no need to identify antigens and adjuvant delivery is feasible and flexible. Besides, there is a great chance to obtain synergistic effect with other therapies (55). One limitation may be due to intratumoral injection, because some internal tumors will not be accessible to injection. As to breast cancer, the primary tumor is superficial, skin and regional lymph node recurrence is common. Therefore, breast cancer is quite accessible to injection, making it a good candidate for ISV.

Food and Drug Administraion (FDA) has approved a number of ISV-based cancer immunotherapies, such as Bacilus Calmette-Guerin (BCG) for in situ vaccination, toll-like receptor agonists for in situ vaccination, oncolytic virus for in situ vaccination, and in situ vaccination with cytokines and immune checkpoint blockade. ISV involves manipulation of intratumoral immune cells by injecting immunomodulators (89) and local ablative therapies which are used to release tumor antigens from the therapy-killed tumor cells (90). Besides, local treatment with vaccines and adjuvant is another option to provoke immune system in situ (91). The combination of ISV with other immnutherapies is likely to provide the optimal local control and systemic antitumor effect. Yokoi et al. treated mammary tumors with in situ immunomodulation consisting of intratumoral injections of Fms-like tyrosine kinase 3 receptor ligand to mobilize conventional type-1 dendritic cells (cDC1s), local irradiation to induce immunogenic tumor cell death, and TLR3/CD40 stimulation to activate cDC1s. Circulating effector T cells and CD8+ T cells infiltrated into metastatic brain lesions were increased and resistance to anti-PD-1 therapy was overcome, resulting in improved survival. Radiation can elicit systemic response which is known as abscopal effect, and the potential mechanism is to release tumor antigens in the process of ISV (92). Numerous clinical data supported the concept of radiation as an important part during in situ vaccine treatment (93-95), and clinical trials are underway investigating combination therapy of radiation with other immunotherapies (91). Combination therapy with noninvasive low intensity focused ultrasound and ablative radiation therapy was reported to generate an *in situ* tumor vaccine as well (96). Like radiation, heat (hyperthermia) has been used to damage targeted tumors and could be further combined with ISV (97). More approaches will be integrated into future multi-modality therapy.

Therapeutic Vaccines for Breast Cancer in Clinical Trials

The treatment for BC at different stages includes neoadjuvant therapy, adjuvant therapy for early BC, rescue therapy and maintenance therapy for advanced BC. Therapeutic vaccines for BC at different stages are summarized.

Neoadjuvant Setting

Disease at an early stage presents with a more intact immune system and a lower tumor burden, possibly affording vaccines the

potential to confer a more favorable outcome. Therapeutic vaccines in the neoadjuvant setting are the theoretically most likely method to optimize the immune microenvironment and improve prognosis. Cancer treatment starts with modulation of the microenvironment and promotion of antitumor immunity before any inhibition occurs to the immune system.

Mucin or MUC-1 is a transmembrane glycoprotein that is expressed in the lung, colon, breast, ovary, pancreas and other cancer tumor cells. MUC-1 is considered a promising candidate for vaccine development in BC. Tecemotide is a synthetic 27 amino acid lipopeptide used as an MUC-1 immunogen that is applied in clinical trials of prostate, NSCLC and colon cancer with promising effects. In a prospective, multicenter, randomized 2-arm academic phase II trial (ABCSG 34), tecemotide was added to neoadjuvant standard-of-care treatment in early BC patients. Approximately 400 patients with early BC were recruited into this trial. No significant difference was observed in residual cancer burden or overall pathological complete response (pCR) rates between the two groups. This trial demonstrated that MUC-1-based vaccination strategies are safe but did not show an improved treatment effect when added to standard treatment in the neoadjuvant setting (98). However, disease-free survival data are still premature and may provide further information. Interestingly, tumors which achieved a residual cancer burden (RCB) 0/I and a pCR had a higher concentration of intratumoural and stromal tumor-infiltrating lymphocytes in the pre-therapeutic biopsy than those which did not. Several ongoing studies address vaccines for BC in the neoadjuvant setting (NCT03387553, NCT02204098, NCT03564782, NCT03572361, NCT04144023). Further data are needed to determine whether neoadjuvant vaccine therapy can reduce the risk of recurrence and prolong relapsefree survival.

Adjuvant Setting

Further immune elimination of subclinical lesions is an important function of vaccines for BC after tumor resection. There have been a number of clinical studies of preventive vaccines in the field of adjuvant therapy.

In a pilot clinical trial of oxidized mannan–MUC-1 (M-FP) for the treatment of patients with stage II BC, the follow-up at 12-15 years showed that the recurrence rate was 12.5% (2/16) in the vaccine group compared with 60% (9/15) in the placebo group. M-FP also benefits the overall survival of stage II BC patients (99). In a phase II clinical trial (NCT02764333), a folate receptor alpha-based vaccine (TPIV200) was investigated in TNBC patients. In this trial, an immunologic response was elicited, and more data has not been exposed.

Peptide vaccines for HER2 have been explored in the adjuvant setting. The E75 peptide vaccine (nelipepimut-S), an HLA-A2/A3-restricted extracellular HER2-domain-derived peptide, is an MHC class I epitope (100). A series of trials in the adjuvant setting were conducted at approximately E75, demonstrating not only a good safety profile of the E75 peptide vaccine but also a superiority of immune response in BC patients with low HER2 expression than vaccinated patients with high levels of HER2 expression (101).

Mittendorf et al. further examined schedule optimization according to lymph node (LN) status and risk of disease recurrence in a phase I/II clinical trial (69). Analysis of disease-free survival (DFS) revealed that patients who had tumors with low HER2 expression (immunohistochemistry score 1+ or 2+ with fluorescence in situ hybridization negativity) and had positive lymph nodes benefited the most from vaccination therapy. In a phase I/II trial, 187 LNpositive and high-risk LN-negative breast cancer (IHC score 1-3) patients were evaluated in the adjuvant setting. E75 patients with GM-CSF versus placebo were administered to 108 patients with HLA-A2/3- and 79 HLA-A2/3-negative patients, respectively. The results concluded that the 5-year DFS was improved for those who received E75 with respect to controls (89.7% vs 80.2%, P=0.08) (102). Given these promising data, in phase III clinical trials, the study assessed the effects of vaccination with E-75 plus subcutaneous GM-CSF relative to placebo in LN+ BC patients with low expression of HER2 in the adjuvant setting (103). However, no significant difference was found in DFS between the vaccine group and the control group, leading to the termination of the trial. Future clinical trials should be carried out to study the combination of vaccines with other medications. Several studies were conducted combining traustuzumab plus E75 in hope of a synergistic effect of active immunotherapy and passive immunotherapy (104). In phase IIb, multicenter, randomized, single-blinded, controlled trial (NCT01570036), the efficacy of the combination with E-75 plus traustuzumab was evaluated in patients with HER2 low-expressing BC in the adjuvant setting. No significant difference in DFS was seen in the HER2 lowexpressing BC; however, significant clinical benefit was seen in patients with TNBC (105). These findings warrant further investigation in a phase III randomized trial. GP2 is a 9 amino acid-long peptide vaccine derived from the transmembrane domain of the HER/neu protein. It binds to the HLA-A2 molecule but has poor binding affinity compared to E75 (106). A phase II clinical trial was conducted to investigate GP2 vaccine efficacy in preventing recurrence in LN+ and high-risk LN- HER2 breast cancer patients (IHC 1+-3+) in the adjuvant setting. The results of the primary analysis did not show a significant difference in response to the vaccine compared to the control group in the rate of recurrence (70). However, patients who were vaccinated with GP2+GM-CSF had a significant increase in their delayed type hypersensitivity (DTH) reaction compared to pre-vaccination (p<0.001), the postvaccination response was significantly greater in vaccinated patients than in control patients (p<0.001). In addition, ex vivo immune responses were assessed by phenotypic clonal expansion assays and by T cell functional assays. The GP2+GM-CSF vaccine induced significant increase in both clonal expansion as well as improved CTL function compared to pre-vaccine levels while GM-CSF alone had no such effect. A post for a prospective, randomized, singleblinded, placebo-controlled, multicenter phase IIb clinical trial was presented during the 2020 San Antonio Breast Cancer Symposium (SABCS) on December 09, 2020. This trial was completed in 2018, and Kaplan-Meier analysis of DFS for patients treated with GP2 immunotherapy showed 100% survival (0% breast cancer recurrence, p=0.0338) in the HER2/neu-positive adjuvant setting after a median of 5 years of follow-up. Greenwich LifeSciences

announced an update of the GP2 phase III clinical trial design at the 2021 American Association for Cancer Research (AACR) annual meeting.

Metastatic Setting

Most mBC cannot be cured by surgery and is highly dependent on systemic therapy. Therapeutic vaccines can be used in combination with other therapies as part of rescue therapy, and other studies are exploring their value as maintenance therapy for advanced breast cancer.

Therapeutic vaccines for rescue therapy for mBC have rarely been reported. Wilms tumor 1 (WT-1) is a protein with transcription factor activity involved in the maintenance of tissue homeostasis, possibly as an oncogene in BC. In a phase I clinical trial, WT-I vaccination activated WT-1-specific cytotoxic T lymphocytes (CTLs) and resulted in cancer regression with a good safety profile in 2 patients with BC with overexpression of the WT-1 gene and HLA-A*2402-possibility (107). Yang et al (108) enrolled 10 patients with advanced cancers, including mBC, and treated them with a DC-based WT-1 vaccination. Two patients had a partial response (PR), and three patients had stable disease (SD) with a disease control rate up to 50%. WT-1-specific CTL responses were enhanced in patients. CEA is overexpressed in BC and has attracted much attention as a target of vaccines. In a pilot study, the recombinant PANVAC poxviral vaccine (containing transgenes for CEA and MUC-1 and three T cell costimulatory molecules) was tested in 12 heavily pretreated metastatic BC patients. One patient demonstrated a CR lasting >37 months, and 4 patients had SD lasting >4 months. The median time to progression (TTP) was 2.5 months, and the median OS was 13.7 months (81). In another open-label phase II clinical trial, 48 patients with mBC were enrolled to receive treatment with either docetaxel with PANVAC or docetaxel alone. The median PFS was 7.9 months in the vaccination group vs 3.9 months in the docetaxel alone group, but the difference was not significant (p=0.09) (109). There was also no statistical correlation seen between the generation of TAAspecific immune responses in peripheral blood mononuclear cells and time to progression in either group. Takahashi et al. (110) developed a novel regimen of personalized peptide vaccination (PPV), in which vaccine antigens were selected and administered from a pool of 31 different peptide candidates based on the preexisting immunoglobulin G (IgG) responses specific to peptides before vaccination. Based on previous results in cancer patients, they conducted a phase II study of PPV for metastatic recurrent breast cancer patients who had failed standard chemotherapies. Boosting of CTL and/or IgG responses was observed in most of the patients after vaccination. In addition, three CR cases and six PR cases were observed, irrespective of the BC subtypes. In a more recent early phase II study including 14 advanced metastatic triple-negative breast cancer (mTNBC) patients, the treatment protocol consisted of a weekly vaccination of mixed 19-peptide cancer vaccine monotherapy for 6 weeks. An increase in peptide-specific IgG was observed in all patients. The median OS was 11.5 months in all 14 patients and 24.4 months in the patients who completed the vaccination (111). Human telomerase reverse transcriptase (hTERT) is nearly universally overexpressed in human cancers and contributes critically to oncogenesis. A phase I clinical trial

was performed to evaluate the HLA-A2-restricted hTERT I540 peptide presented with keyhole limpet hemocyanin (KLH) by ex vivo-generated autologous DCs. hTERT-specific T lymphocytes were induced in 4/7 patients after vaccination. PR was seen in 1 patient in association with the induction of CD8+ tumor infiltrating lymphocytes (112). In conclusion, although no prospective large-sample studies have confirmed the efficacy of therapeutic vaccines in the rescue therapy of advanced BC, some studies have preliminary results suggesting their effectiveness and possible prospects.

Immunosurveillance using therapeutic vaccines to trigger active immunity when remission is achieved through rescue therapy such as radiotherapy or chemotherapy suggests novel opportunities for both therapeutic and prophylactic vaccine strategies for cancer treatment. MAM-A is overexpressed in 40-80% of breast tumors. Tiriveedhi et al. (80) enrolled 14 mBC patients with stable disease and treated them randomly with the MAM-A vaccine or placebo in a phase I clinical trial. Although this trial was not powered to evaluate PFS, improved PFS was seen in vaccinated patients. A significant increase in the frequency of MAM-A-specific CD8+ T cells (0.9% + 0.5% vs 3.8% + 1.2%; p<0.001) and an increase in the number of MAM-A-specific IFN γ -secreting T cells (41 + 32 vs 215 + 67 spots per million cells (spm); p<0.001) were observed. Increased Siayl-TN (STn) expression, which is a carbohydrate epitope found on a variety of glycoproteins, including MUC-1, has been proven to be associated with the progression and poor prognosis of BC (113). Miled et al. (114) conducted the largest phase III clinical trial in 1028 mBC patients across 126 centers. Patients were administered a vaccine made of STn conjugated to the carrier protein KLH versus placebo. Although clinically significant antibody titers specific for STn were produced in patients, no significant improvement in TTP or OS was observed (115). Ibrahim et al. conducted a subgroup analysis in which patients who were also on endocrine therapy (ET) had longer TTP and OS than the control group. Moreover, vaccinated patients on ET with higher antibody responses had longer OS (41.3 vs 25.4 months; p=0.0147). In an open-label prospective study, 19 patients with mBC refractory to at least one conventional therapy were treated with the hTERT peptide vaccine, and hTERT-specific CD8+ T cells were detected after vaccination in the peripheral blood of patients and exhibited effector functions in vitro, including proliferation, IFN-gamma production, and tumor lysis. In this small sample study, the median OS was significantly longer in patients who achieved an immune response to hTERT peptide than in patients who did not (116). All the results above suggest that therapeutic vaccines are a potentially feasible option for maintenance therapy of advanced BC, but no mature vaccine has been proven to be beneficial in a large-sample clinical trial. Another important issue that should be considered is that the essential immune capability to recognize and activate antigens should be conserved before vaccination.

FUTURE PERSPECTIVES

Tumor Stage Specific Vaccine Strategy

During cancer clonal evolution, both selection and neutral growth may progress simultaneously within the same tumor,

but both styles of tumor progression may alter dynamically over time (117). Metastatic BC shows an increase in mutational burden and clonal diversity compared to early BC because genomic alterations are acquired during the evolution of cancers from their early stages (118). A multitude of epigenetic mechanisms, including DNA methylation, chromatin remodeling and posttranslational modification of histones, contribute to diversity within tumors, and the heterogeneity becomes extensive. Intratumor heterogeneity (ITH) is a key factor contributing to the lethal outcome of cancer, therapeutic failure, and drug resistance. Some claim that tumors with high heterogeneity may generate neoantigens that attract immune cells (119), but others argue that immune cells provide selection pressure that shapes tumor heterogeneity. High heterogeneity tumors are associated with higher subclonal diversity, less immune cell infiltration, less activation of the immune response, and worse survival in BC (120). Immune-infiltrated tumor regions exhibit either HLA loss of heterozygosity (LOH) or depletion of expressed neoantigens, which will eventually make it increasingly difficult to treat mBC with an immune strategy (120-122). Finding the right target antigen and intervening at the right time are the most important issues of active immunotherapy. The continuous evolution of the immune microenvironment during tumorigenesis also suggests that different modes of treatment should be considered at different stages.

Neoantigen profiles keep changing while tumor-specific mutations change during tumorigenesis and progression. Therefore, individual immune status, clonal heterogeneity and stage of disease should be fully considered, and time specificity should be realized.

Universal Vaccines

Optimal antigens should be developed from publicly mutated genes or high frequency overexpressed genes that are shared by a number of patients. A punch of such public antigens that are consumed to cover most patients with one type of cancer can be used to develop public vaccines, also named universal vaccines (123). Universal vaccines have the great advantage of convenient production and reduction in cost. In addition, preprepared vaccines that can be quickly inoculated into patients also save time and are more practical. The efficacy of universal vaccines should be ensured. One important problem should be considered except for the restriction of MHC molecules. That is, the proportion of tumor antigen expression in the population. Although more than 900,000 neoantigens have been identified through a wide examination of 20 tumor histotypes, only 24 neoantigens among a tiny fraction of patients have the potential to become public vaccines (124). Therefore, it is more feasible to develop public vaccines based on TAAs. Public vaccines have broad coverage and can improve the immune surveillance function of individuals to prevent tumor metastasis and recurrence. It is theoretically more suitable for the stage of neoadjuvant and adjuvant therapy.

From a single genome point of view, improving the antitumor effect of tumor-specific T cells and memory T cells

is important for designing therapeutic vaccines. Personalized therapeutic vaccines targeting trunk or driver mutations are more effective and have a more comprehensive antitumor effect than those targeting companion or passenger mutations. In addition, the option of designing vaccines needs to be weighed between selecting a large number of target antigens to avoid immune escape and selecting antigens with good immunogenic potential.

Concurrent Therapies With Vaccination

Conventional therapies such as chemotherapy, radiotherapy and targeted therapy constantly promote the emergence of new subclones of tumor cells as a result of the pressure of clonal evolution, resulting in treatment failure. Immunotherapy, as a new therapeutic strategy, has a totally different effect on tumor heterogeneity from conventional therapy. However, patients who have received multiline conventional therapies can hardly benefit from immunotherapy. How to maximize the therapeutic effect of immunotherapy by rational arrangement of comprehensive therapy is an important direction in the future. In addition, how to exert antitumor effects of therapeutic vaccines synergistically with various therapeutic means is a hotspot. It was reported that sequential treatment with vaccine and PD-1 blockade was more effective than a simultaneous treatment regimen (125). In the PACIFIC trial, when durvalumab therapy was initiated within 14 days of completing chemoradiotherapy, better progression free survival was observed than when it was initiated after 14 days (126). Thus, timing is an important factor in obtaining abscopal effect and the optimal scheduling of vaccines, immunotherapy, radiation and chemotherapy needs to be clearly established, ideally through clinical trials. The TME is a major reason for the disappointing clinical results in addition to tumor-intrinsic resistance mechanisms, so an inflammatory TME is needed for sterile immunity (38). Except for what we have mentioned above about TME-targeting vaccines, in situ TME modulation strategies include stimulation of professional antigen presenting cells, combination with checkpoint inhibitors and depletion of regulatory T cells (Treg cells). PVX-410 (PVX) is a multipeptide vaccine targeting X-Box Binding Protein 1 (XBP1), and CD138 is overexpressed in TNBC. The synergistic effects of PVX-410 and ICI pembrolizumab will be evaluated in a clinical trial (NCT03362060) for TNBC. Another phase I clinical trial (NCT02826434) tested the synergistic effects of durvalumab and PVX-410 for TNBC. In this trial, the levels of CD8+ CTLs increased in patients 14 weeks after the first injection. The combination therapy strategy to work together with vaccines will include, but is not limited to, ICIs, antiangiogenic therapy, epigenetic regulation therapy, low intensity focused ultrasound (55) and conventional chemoradiotherapy. Cyclophosphamide to block Treg cells has been evaluated as a vaccine adjuvant in clinical trials (NCT03012100, NCT02938442). Several other ongoing trials are further assessing the application of various promising vaccination therapies in early and metastatic disease (Table 2).

TABLE 2 | Ongoing trials of tumor vaccine-based combination therapy for BCs (data from ClinicalTrials.gov).

Drug Regimen	NCT.gov Identifier	Sample Size	Phase; Status	Population
Neoantigen DNA Vaccine Durvalumab	NCT03199040	10	I; ANR	Clinical Stage T1c-T4c, Any N, M0 TNBC Prior to Neoadjuvant Chemotherapy, with Residual Invasive BC after Neoadjuvant Therapy
VRP-HER2	NCT03632941	39	II; R	Advanced HER2-overexpressing BC
Pembrolizumab	110100002011	00	,	Adda lood 112112 of oloxip obouing 20
PVX-410	NCT03362060	20	I; ANR	HLA-A2 + Metastatic TNBC
Pembrolizumab				
Galinpepimut-S	NCT03761914	90	I/II; R	Advanced Tumors including Advanced TNBC
Pembrolizumab				
R07198457	NCT03289962	770	I; R	Advanced Tumors including Advanced TNBC
Atezolizumab				
PVX-410	NCT02826434	22	lb; ANR	HLA-A2 + Subjects Following Standard
Durvalumab				Treatment of Stage II or III TNBC
Hiltonol	NOTOCOLOGO	000		
Multiepitope Folate Receptor Alpha Peptide Vaccine	NCT03012100	280	ll; R	Stage I-III TNBC
Cyclophosphamide NeuVax Vaccine	NCT02297698	100	II; ANR	Stage I-III Noninflammatory, HER2+ High-risk BC
Trastuzumab	10102231030	100	11, 741111	Stage Fill Norin manimatory, TIEN2+ High-lisk DO
A Peptide Mimotope-based Vaccine P10s-PADRE with	NCT02938442	102	I/II; R	Stage I, II or III TNBC
MONTANIDE™ ISA 51 VG	10102000112	102	,	
Doxorubicin				
Cyclophosphamide				
Paclitaxel				
AE37 Peptide vaccine	NCT04024800	29	II; ANR	Advanced TNBC
Pembrolizumab				
Dendritic Cell Vaccine	NCT03387553	30	I; R	HER-2/neu Positive Invasive BC during
Neoadjuvant Chemotherapy				Neoadjuvant Therapy
Anti-HER2/HER3 Dendritic Cell Vaccine	NCT04348747	23	lla; NYR	Patients With Asymptomatic Brain Metastasis
Recombinant Interferon Alfa-2b				From TNBC or HER2+ BC
Celecoxib				
Pembrolizumab	NOT0000007	70		
Personalized Synthetic Long Peptide Vaccine	NCT03606967	70	ll; R	Advanced TNBC
Carboplatin Durvalumab				
Gemcitabine Hydrochloride				
Nab-paclitaxel				
Tremelimumab				
Multiepitope HER2 Peptide Vaccine TPIV100	NCT04197687	480	II; R	HER2 Positive, Stage II-III BC in Patients With
Pertuzumab				Residual Disease After Chemotherapy and
Trastuzumab				Surgery
pUMVC3-IGFBP2-HER2-IGF1R Plasmid DNA Vaccine	NCT04329065	16	II; R	BC during Neoadjuvant Therapy
Paclitaxel				
Trastuzumab				
Pertuzumab				
Brachyury-TRICOM	NCT04296942	65	l; R	Advacned BC
Entinostat M7824				
M7824 Ado-trastuzumab emtansine				
In Situ Vaccination With Flt3 L, Radiation, and Poly-ICLC	NCT03789097	56	I/II; R	Advanced, Measurable, Biopsy-accessible
Pembrolizumab	140100109091	50	<i>i</i> /11, 13	Cancers including BC
In Situ Vaccination	NCT02643303	58	I/II; ANR	Advanced, Measurable, Biopsy-accessible
Durvalumab	110102040000	50	<i>a</i> 11, 7 st MT 1	Cancers including BC
Tremelimumab				

ANR, active; not recruiting; NYR, not yet recruiting; R, recruiting.

CONCLUSION

In recent years, the application of therapeutic vaccines has been gradually accepted in the field of BC, but both the candidates and the efficacy need further study. Increasing attention has been

given to the use of therapeutic vaccines to modulate the immune microenvironment and fully mobilize the body's own immune system for active immunotherapy. However, the exploration of therapeutic vaccines for BC is still in the early stage and is bound to be long based on considering the stage of disease, personal immune status and clonal heterogeneity. Fully combining therapeutic vaccines with not only ICIs but also other multiple treatment methods may take great advantage in the future treatment of BC.

AUTHOR CONTRIBUTIONS

The corresponding authors are responsible for ensuring that the descriptions are accurate and agreed by all authors. Authors have contributed in multiple roles. LZ is responsible for writing original draft and literature search. XZ is responsible for literature search. LX is

REFERENCES

- Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, et al. Global Cancer Statistics 2020: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. *CA Cancer J Clin* (2021) 71:209–49. doi: 10.3322/caac.21660
- Kwapisz D. Pembrolizumab and Atezolizumab in Triple-Negative Breast Cancer. Cancer Immunol Immunother (2021) 70:607–17. doi: 10.1007/ s00262-020-02736-z
- Schreiber R, Old L, Smyth M. Cancer Immunoediting: Integrating Immunity's Roles in Cancer Suppression and Promotion. *Science (New York NY)* (2011) 331:1565–70. doi: 10.1126/science.1203486
- Ott PA, Hu Z, Keskin DB, Shukla SA, Sun J, Bozym DJ, et al. An Immunogenic Personal Neoantigen Vaccine for Patients With Melanoma. *Nature* (2017) 547:217–21. doi: 10.1038/nature22991
- Keskin DB, Anandappa AJ, Sun J, Tirosh I, Mathewson ND, Li S, et al. Neoantigen Vaccine Generates Intratumoral T Cell Responses in Phase Ib Glioblastoma Trial. *Nature* (2019) 565:234–9. doi: 10.1038/s41586-018-0792-9
- Kantoff PW, Higano CS, Shore ND, Berger ER, Small EJ, Penson DF, et al. Sipuleucel-T Immunotherapy for Castration-Resistant Prostate Cancer. N Engl J Med (2010) 363:411–22. doi: 10.1056/NEJMoa1001294
- Fragomeni SM, Sciallis A, Jeruss JS. Molecular Subtypes and Local-Regional Control of Breast Cancer. Surg Oncol Clin N Am (2018) 27:95–120. doi: 10.1016/j.soc.2017.08.005
- Tolba MF, Omar HA. Immunotherapy, an Evolving Approach for the Management of Triple Negative Breast Cancer: Converting non-Responders to Responders. *Crit Rev Oncol Hematol* (2018) 122:202–7. doi: 10.1016/j.critrevonc.2018.01.005
- Hollingsworth R, Jansen K. Turning the Corner on Therapeutic Cancer Vaccines. NPJ Vaccines (2019) 4:7. doi: 10.1038/s41541-019-0103-y
- Buonaguro L, Petrizzo A, Tornesello M, Buonaguro F. Translating Tumor Antigens Into Cancer Vaccines. *Clin Vaccine Immunol CVI* (2011) 18:23–34. doi: 10.1128/cvi.00286-10
- Lim S, Zhang Y, Zhang J. Cancer-Testis Antigens: The Current Status on Antigen Regulation and Potential Clinical Use. Am J Blood Res (2012) 2:29–35.
- Otte M, Zafrakas M, Riethdorf L, Pichlmeier U, Löning T, Jänicke F, et al. MAGE-A Gene Expression Pattern in Primary Breast Cancer. *Cancer Res* (2001) 61:6682–7.
- Vodolazhsky D, Kutilin D, Mogushkova K, Kit O. Specific Features of Transcription Activity of Cancer-Testis Antigens in Patients With Metastatic and Non-Metastatic Breast Cancer. Bull Exp Biol Med (2018) 165:382–5. doi: 10.1007/s10517-018-4175-x
- Kondo Y, Fukuyama T, Yamamura R, Futawatari N, Ichiki Y, Tanaka Y, et al. Detection of KK-LC-1 Protein, a Cancer/Testis Antigen, in Patients With Breast Cancer. *Anticancer Res* (2018) 38:5923–8. doi: 10.21873/ anticanres.12937
- Egland KA, Kumar V, Duray P, Pastan I. Characterization of Overlapping XAGE-1 Transcripts Encoding a Cancer Testis Antigen Expressed in Lung, Breast, and Other Types of Cancers. *Mol Cancer Ther* (2002) 1:441–50.

responsible for conceptualization and supervision. BL is responsible for review and editing for original draft and supervision. All authors contributed to the article and approved the submitted version.

FUNDING

This work was funded by a grant from the National Natural Science Foundation of China (Grant No. 81803093). The funding sources had no role in the study design, data collection, data analysis, data interpretation, or writing of the report.

- 16. Sakai Y, Kurose K, Sakaeda K, Abo H, Atarashi Y, Ide N, et al. A Novel Automated Immunoassay for Serum NY-ESO-1 and XAGE1 Antibodies in Combinatory Prediction of Response to Anti-Programmed Cell Death-1 Therapy in non-Small-Cell Lung Cancer. *Clinica chimica acta; Int J Clin Chem* (2021) 519:51–9. doi: 10.1016/j.cca.2021.04.008
- Kawakami Y, Eliyahu S, Sakaguchi K, Robbins P, Rivoltini L, Yannelli J, et al. Identification of the Immunodominant Peptides of the MART-1 Human Melanoma Antigen Recognized by the Majority of HLA-A2-Restricted Tumor Infiltrating Lymphocytes. J Exp Med (1994) 180:347–52. doi: 10.1084/jem.180.1.347
- Bakker A, Schreurs M, De Boer A, Kawakami Y, Rosenberg S, Adema G, et al. Melanocyte Lineage-Specific Antigen Gp100 is Recognized by Melanoma-Derived Tumor-Infiltrating Lymphocytes. J Exp Med (1994) 179:1005–9. doi: 10.1084/jem.179.3.1005
- Corman J, Sercarz E, Nanda N. Recognition of Prostate-Specific Antigenic Peptide Determinants by Human CD4 and CD8 T Cells. *Clin Exp Immunol* (1998) 114:166–72. doi: 10.1046/j.1365-2249.1998.00678.x
- Tsang K, Zaremba S, Nieroda C, Zhu M, Hamilton J, Schlom J. Generation of Human Cytotoxic T Cells Specific for Human Carcinoembryonic Antigen Epitopes From Patients Immunized With Recombinant Vaccinia-CEA Vaccine. J Natl Cancer Institute (1995) 87:982–90. doi: 10.1093/jnci/ 87.13.982
- Seil I, Frei C, Sültmann H, Knauer S, Engels K, Jäger E, et al. The Differentiation Antigen NY-BR-1 is a Potential Target for Antibody-Based Therapies in Breast Cancer. *Int J Cancer* (2007) 120:2635–42. doi: 10.1002/ ijc.22620
- Balafoutas D, Zur Hausen A, Mayer S, Hirschfeld M, Jaeger M, Denschlag D, et al. Cancer Testis Antigens and NY-BR-1 Expression in Primary Breast Cancer: Prognostic and Therapeutic Implications. *BMC Cancer* (2013) 13:271. doi: 10.1186/1471-2407-13-271
- Fisk B, Blevins T, Wharton J, Ioannides C. Identification of an Immunodominant Peptide of HER-2/Neu Protooncogene Recognized by Ovarian Tumor-Specific Cytotoxic T Lymphocyte Lines. J Exp Med (1995) 181:2109–17. doi: 10.1084/jem.181.6.2109
- Segal-Eiras A, Croce M. Breast Cancer Associated Mucin: A Review. Allergol Immunopathol (1997) 25:176–81.
- Vonderheide R, Hahn W, Schultze J, Nadler L. The Telomerase Catalytic Subunit is a Widely Expressed Tumor-Associated Antigen Recognized by Cytotoxic T Lymphocytes. *Immunity* (1999) 10:673–9. doi: 10.1016/s1074-7613(00)80066-7
- Andersen M, Pedersen L, Becker J, Straten P. Identification of a Cytotoxic T Lymphocyte Response to the Apoptosis Inhibitor Protein Survivin in Cancer Patients. *Cancer Res* (2001) 61:869–72.
- Peng M, Mo Y, Wang Y, Wu P, Zhang Y, Xiong F, et al. Neoantigen Vaccine: An Emerging Tumor Immunotherapy. *Mol Cancer* (2019) 18:128. doi: 10.1186/s12943-019-1055-6
- Mardis ER. Neoantigens and Genome Instability: Impact on Immunogenomic Phenotypes and Immunotherapy Response. *Genome Med* (2019) 11:71. doi: 10.1186/s13073-019-0684-0

- Zhang X, Kim S, Hundal J, Herndon JM, Li S, Petti AA, et al. Breast Cancer Neoantigens Can Induce CD8(+) T-Cell Responses and Antitumor Immunity. *Cancer Immunol Res* (2017) 5:516–23. doi: 10.1158/2326-6066.Cir-16-0264
- Hwang S, Kim E, Lee I, Marcotte E. Systematic Comparison of Variant Calling Pipelines Using Gold Standard Personal Exome Variants. Sci Rep (2015) 5:17875. doi: 10.1038/srep17875
- Xu C. A Review of Somatic Single Nucleotide Variant Calling Algorithms for Next-Generation Sequencing Data. *Comput Struct Biotechnol J* (2018) 16:15–24. doi: 10.1016/j.csbj.2018.01.003
- 32. Karasaki T, Nagayama K, Kuwano H, Nitadori J, Sato M, Anraku M, et al. Prediction and Prioritization of Neoantigens: Integration of RNA Sequencing Data With Whole-Exome Sequencing. *Cancer Sci* (2017) 108:170–7. doi: 10.1111/cas.13131
- Smart A, Margolis C, Pimentel H, He M, Miao D, Adeegbe D, et al. Intron Retention is a Source of Neoepitopes in Cancer. Nat Biotechnol (2018) 36:1056–8. doi: 10.1038/nbt.4239
- 34. Rathe S, Popescu F, Johnson J, Watson A, Marko T, Moriarity B, et al. Identification of Candidate Neoantigens Produced by Fusion Transcripts in Human Osteosarcomas. Sci Rep (2019) 9:358. doi: 10.1038/s41598-018-36840-z
- Hombrink P, Hassan C, Kester M, De Ru A, Van Bergen C, Nijveen H, et al. Discovery of T Cell Epitopes Implementing HLA-Peptidomics Into a Reverse Immunology Approach. J Immunol (Baltimore Md. 1950) (2013) 190:3869–77. doi: 10.4049/jimmunol.1202351
- Rammensee HG, Singh-Jasuja H. HLA Ligandome Tumor Antigen Discovery for Personalized Vaccine Approach. *Expert Rev Vaccines* (2013) 12:1211–7. doi: 10.1586/14760584.2013.836911
- Joglekar A, Leonard M, Jeppson J, Swift M, Li G, Wong S, et al. T Cell Antigen Discovery via Signaling and Antigen-Presenting Bifunctional Receptors. Nat Methods (2019) 16:191–8. doi: 10.1038/s41592-018-0304-8
- Gordon B, Gadi V. The Role of the Tumor Microenvironment in Developing Successful Therapeutic and Secondary Prophylactic Breast Cancer Vaccines. *Vaccines* (2020) 8:529. doi: 10.3390/vaccines8030529
- Yan HX, Cheng P, Wei HY, Shen GB, Fu LX, Ni J, et al. Active Immunotherapy for Mouse Breast Cancer With Irradiated Whole-Cell Vaccine Expressing VEGFR2. Oncol Rep (2013) 29:1510–6. doi: 10.3892/ or.2013.2282
- Zhao X, Bose A, Komita H, Taylor JL, Chi N, Lowe DB, et al. Vaccines Targeting Tumor Blood Vessel Antigens Promote CD8(+) T Cell-Dependent Tumor Eradication or Dormancy in HLA-A2 Transgenic Mice. J Immunol (2012) 188:1782–8. doi: 10.4049/jimmunol.1101644
- Lu Y, Wei YQ, Tian L, Zhao X, Yang L, Hu B, et al. Immunogene Therapy of Tumors With Vaccine Based on Xenogeneic Epidermal Growth Factor Receptor. *J Immunol* (2003) 170:3162–70. doi: 10.4049/jimmunol.170.6.3162
- Wood LM, Pan ZK, Guirnalda P, Tsai P, Seavey M, Paterson Y. Targeting Tumor Vasculature With Novel Listeria-Based Vaccines Directed Against CD105. *Cancer Immunol Immunother* (2011) 60:931–42. doi: 10.1007/ s00262-011-1002-x
- Kaplan CD, Kruger JA, Zhou H, Luo Y, Xiang R, Reisfeld RA. A Novel DNA Vaccine Encoding PDGFRbeta Suppresses Growth and Dissemination of Murine Colon, Lung and Breast Carcinoma. *Vaccine* (2006) 24:6994–7002. doi: 10.1016/j.vaccine.2006.04.071
- 44. Jin D, Yu X, Chen B, Li Z, Ding J, Zhao X, et al. Combined Immunotherapy of Breast Cancer With EGF and VEGF Vaccines From DNA Shuffling in a Mouse Model. *Immunotherapy* (2017) 9:537–53. doi: 10.2217/imt-2017-0004
- 45. Sung Kim T, Cohen EP. Immunity to Breast Cancer in Mice Immunized With Fibroblasts Transfected With a cDNA Expression Library Derived From Small Numbers of Breast Cancer Cells. *Cancer Gene Ther* (2005) 12:890–9. doi: 10.1038/sj.cgt.7700853
- 46. Kim TS, Jung MY, Cho D, Cohen EP. Prolongation of the Survival of Breast Cancer-Bearing Mice Immunized With GM-CSF-Secreting Syngeneic/ Allogeneic Fibroblasts Transfected With a cDNA Expression Library From Breast Cancer Cells. Vaccine (2006) 24:6564–73. doi: 10.1016/ j.vaccine.2006.06.012
- 47. Xia Q, Zhang FF, Geng F, Liu CL, Xu P, Lu ZZ, et al. Anti-Tumor Effects of DNA Vaccine Targeting Human Fibroblast Activation Protein Alpha by

Producing Specific Immune Responses and Altering Tumor Microenvironment in the 4T1 Murine Breast Cancer Model. *Cancer Immunol Immunother* (2016) 65:613–24. doi: 10.1007/s00262-016-1827-4

- Andersen M. The T-Win[®] Technology: Immune-Modulating Vaccines. Semin Immunopathol (2019) 41:87–95. doi: 10.1007/s00281-018-0695-8
- Palena C, Polev D, Tsang K, Fernando R, Litzinger M, Krukovskaya L, et al. The Human T-Box Mesodermal Transcription Factor Brachyury is a Candidate Target for T-Cell-Mediated Cancer Immunotherapy. *Clin Cancer Res* (2007) 13:2471–8. doi: 10.1158/1078-0432.Ccr-06-2353
- Quaglino E, Conti L, Cavallo F. Breast Cancer Stem Cell Antigens as Targets for Immunotherapy. *Semin Immunol* (2020) 47:101386. doi: 10.1016/ j.smim.2020.101386
- Chiang CL, Benencia F, Coukos G. Whole Tumor Antigen Vaccines. Semin Immunol (2010) 22:132–43. doi: 10.1016/j.smim.2010.02.004
- Liu W, Tang H, Li L, Wang X, Yu Z, Li J. Peptide-Based Therapeutic Cancer Vaccine: Current Trends in Clinical Application. *Cell Prolif* (2021) 54: e13025. doi: 10.1111/cpr.13025
- Supabphol S, Li L, Goedegebuure SP, Gillanders WE. Neoantigen Vaccine Platforms in Clinical Development: Understanding the Future of Personalized Immunotherapy. *Expert Opin Investig Drugs* (2021) 30:1–13. doi: 10.1080/13543784.2021.1896702
- Perez CR, De Palma M. Engineering Dendritic Cell Vaccines to Improve Cancer Immunotherapy. *Nat Commun* (2019) 105408. doi: 10.1038/s41467-019-13368-y
- Sheen MR, Fiering S. In Situ Vaccination: Harvesting Low Hanging Fruit on the Cancer Immunotherapy Tree. Wiley Interdiscip Rev Nanomed Nanobiotechnol (2019) 11:e1524. doi: 10.1002/wnan.1524
- González FE, Gleisner A, Falcón-Beas F, Osorio F, López MN, Salazar-Onfray F. Tumor Cell Lysates as Immunogenic Sources for Cancer Vaccine Design. *Hum Vaccin Immunother* (2014) 10:3261–9. doi: 10.4161/ 21645515.2014.982996
- Sanjay S, Patel JM, Bozeman EN, Imasuen IE, Sara H, Danielle D, et al. Allogeneic Tumor Cell Vaccines: The Promise and Limitations in Clinical Trials. *Hum Vaccines Immunother* (2013) 10:52–63. doi: 10.4161/hv.26568
- Emens LA. Breast Cancer Vaccines: Maximizing Cancer Treatment by Tapping Into Host Immunity. *Endocr Related Cancer* (2005) 12:1. doi: 10.1677/erc.1.00671
- Simons JW, Sacks N. Granulocyte-Macrophage Colony-Stimulating Factortransduced Allogeneic Cancer Cellular Immunotherapy: The GVAX Vaccine for Prostate Cancer. Urol Oncol Semin Orig Invest (2006) 24:419– 24. doi: 10.1016/j.urolonc.2005.08.021
- 60. Dranoff G, Jaffee E, Lazenby A, Golumbek P, Levitsky H, Brose K, et al. Vaccination With Irradiated Tumor Cells Engineered to Secrete Murine Granulocyte-Macrophage Colony-Stimulating Factor Stimulates Potent, Specific, and Long-Lasting Anti-Tumor Immunity. *Proc Natl Acad Sci* USA (1993) 90:3539–43. doi: 10.1073/pnas.90.8.3539
- Kurtz SL, Ravindranathan S, Zaharoff D. Current Status of Autologous Breast Tumor Cell-Based Vaccines. *Expert Rev Vaccines* (2014) 13:1439–45. doi: 10.1586/14760584.2014.969714
- 62. Anderson K, Erick T, Chen M, Daley H, Campbell M, Colson Y, et al. The Feasibility of Using an Autologous GM-CSF-Secreting Breast Cancer Vaccine to Induce Immunity in Patients With Stage II-III and Metastatic Breast Cancers. Breast Cancer Res Treat (2022). doi: 10.1007/s10549-022-06562-y
- 63. Emens LA, Asquith JM, Leatherman JM, Kobrin BJ, Petrik S, Laiko M, et al. Timed Sequential Treatment With Cyclophosphamide, Doxorubicin, and an Allogeneic Granulocyte-Macrophage Colony-Stimulating Factor-Secreting Breast Tumor Vaccine: A Chemotherapy Dose-Ranging Factorial Study of Safety and Immune Activation. J Clin Oncol (2009) 27:5911. doi: 10.1200/ JCO.2009.23.3494
- 64. Dols A, Smith JW, Meijer SL, Fox BA, Hu HM, Walker E, et al. Vaccination of Women With Metastatic Breast Cancer, Using a Costimulatory Gene (CD80)-Modified, HLA-A2-Matched, Allogeneic, Breast Cancer Cell Line: Clinical and Immunological Results. *Hum Gene Ther* (2003) 14:1117–23. doi: 10.1089/104303403322124828
- 65. Chen G, Gupta R, Petrik S, Laiko M, Leatherman J, Asquith J, et al. A Feasibility Study of Cyclophosphamide, Trastuzumab, and an Allogeneic GM-CSF-Secreting Breast Tumor Vaccine for HER2+ Metastatic Breast

Cancer. Cancer Immunol Res (2014) 2:949-61. doi: 10.1158/2326-6066.Cir-14-0058

- 66. Zaks TZ, Rosenberg SA. Immunization With a Peptide Epitope (P369-377) From HER-2/Neu Leads to Peptide-Specific Cytotoxic T Lymphocytes That Fail to Recognize HER-2/Neu+ Tumors. *Cancer Res* (1998) 58:4902–8.
- Knutson KL, Schiffman K, Cheever MA, Disis ML. Immunization of Cancer Patients With a HER-2/Neu, HLA-A2 Peptide, P369-377, Results in Short-Lived Peptide-Specific Immunity. *Clin Cancer Res* (2002) 8:1014–8.
- 68. Carmichael MG, Benavides LC, Holmes JP, Gates JD, Mittendorf EA, Ponniah S, et al. Results of the First Phase 1 Clinical Trial of the HER-2/ Neu Peptide (GP2) Vaccine in Disease-Free Breast Cancer Patients: United States Military Cancer Institute Clinical Trials Group Study I-04. *Cancer* (2010) 116:292–301. doi: 10.1002/cncr.24756
- Mittendorf EA, Clifton GT, Holmes JP, Clive KS, Patil R, Benavides LC, et al. Clinical Trial Results of the HER-2/Neu (E75) Vaccine to Prevent Breast Cancer Recurrence in High-Risk Patients: From US Military Cancer Institute Clinical Trials Group Study I-01 and I-02. *Cancer* (2012) 118:2594–602. doi: 10.1002/cncr.26574
- Mittendorf EA, Ardavanis A, Litton JK, Shumway NM, Hale DF, Murray JL, et al. Primary Analysis of a Prospective, Randomized, Single-Blinded Phase II Trial Evaluating the HER2 Peptide GP2 Vaccine in Breast Cancer Patients to Prevent Recurrence. *Oncotarget* (2016) 7:66192–201. doi: 10.18632/ oncotarget.11751
- Miggelbrink A, Jackson J, Lorrey S, Srinivasan E, Waibl Polania J, Wilkinson D, et al. CD4 T-Cell Exhaustion: Does it Exist and What are its Roles in Cancer? *Clin Cancer Res* (2021) 27:5742–52. doi: 10.1158/1078-0432.Ccr-21-0206
- 72. Zhang H, Hong H, Li D, Ma S, Di Y, Stoten A, et al. Comparing Pooled Peptides With Intact Protein for Accessing Cross-Presentation Pathways for Protective CD8+ and CD4+ T Cells. J Biol Chem (2009) 284:9184–91. doi: 10.1074/jbc.M809456200
- Melief C, van der Burg S. Immunotherapy of Established (Pre)Malignant Disease by Synthetic Long Peptide Vaccines. *Nat Rev Cancer* (2008) 8:351– 60. doi: 10.1038/nrc2373
- 74. Bijker M, Van Den Eeden S, Franken K, Melief C, van der Burg S, Offringa R. Superior Induction of Anti-Tumor CTL Immunity by Extended Peptide Vaccines Involves Prolonged, DC-Focused Antigen Presentation. *Eur J Immunol* (2008) 38:1033–42. doi: 10.1002/eji.200737995
- Lefrançois L, Marzo A, Williams K. Sustained Response Initiation is Required for T Cell Clonal Expansion But Not for Effector or Memory Development *In Vivo. J Immunol (Baltimore Md. 1950)* (2003) 171:2832–9. doi: 10.4049/jimmunol.171.6.2832
- 76. Obst R, Van Santen H, Melamed R, Kamphorst A, Benoist C, Mathis D. Sustained Antigen Presentation can Promote an Immunogenic T Cell Response, Like Dendritic Cell Activation. *Proc Natl Acad Sci USA* (2007) 104:15460–5. doi: 10.1073/pnas.0707331104
- Li WH, Li YM. Chemical Strategies to Boost Cancer Vaccines. Chem Rev (2020) 120:11420–78. doi: 10.1021/acs.chemrev.9b00833
- Lopes A, Bastiancich C, Bausart M, Ligot S, Lambricht L, Vanvarenberg K, et al. New Generation of DNA-Based Immunotherapy Induces a Potent Immune Response and Increases the Survival in Different Tumor Models. *J Immunother Cancer* (2021) 9. doi: 10.1136/jitc-2020-001243
- 79. Norell H, Poschke I, Charo J, Wei W, Erskine C, Piechocki M, et al. Vaccination With a Plasmid DNA Encoding HER-2/Neu Together With Low Doses of GM-CSF and IL-2 in Patients With Metastatic Breast Carcinoma: A Pilot Clinical Trial. *J Trans Med* (2010) 8:53. doi: 10.1186/ 1479-5876-8-53
- Tiriveedhi V, Tucker N, Herndon J, Li L, Sturmoski M, Ellis M, et al. Safety and Preliminary Evidence of Biologic Efficacy of a Mammaglobin-a DNA Vaccine in Patients With Stable Metastatic Breast Cancer. *Clin Cancer Res* (2014) 20:5964–75. doi: 10.1158/1078-0432.Ccr-14-0059
- Mohebtash M, Tsang K, Madan R, Huen N, Poole D, Jochems C, et al. A Pilot Study of MUC-1/CEA/TRICOM Poxviral-Based Vaccine in Patients With Metastatic Breast and Ovarian Cancer. *Clin Cancer Res* (2011) 17:7164–73. doi: 10.1158/1078-0432.Ccr-11-0649
- Dissanayake D, Murakami K, Tran MD, Elford AR, Millar DG, Ohashi PS. Peptide-Pulsed Dendritic Cells Have Superior Ability to Induce Immune-Mediated Tissue Destruction Compared to Peptide With Adjuvant. *PLoS One* (2014) 9:e92380. doi: 10.1371/journal.pone.0092380

- Kugler A, Stuhler G, Walden P, Zöller G, Zobywalski A, Brossart P, et al. Regression of Human Metastatic Renal Cell Carcinoma After Vaccination With Tumor Cell-Dendritic Cell Hybrids. *Nat Med* (2000) 6:332–6. doi: 10.1038/73193
- Morse M, Hobeika A, Osada T, Niedzwiecki D, Marcom P, Blackwell K, et al. Long Term Disease-Free Survival and T Cell and Antibody Responses in Women With High-Risk Her2+ Breast Cancer Following Vaccination Against Her2. J Trans Med (2007) 5:42. doi: 10.1186/1479-5876-5-42
- Avigan D, Vasir B, Gong J, Borges V, Wu Z, Uhl L, et al. Fusion Cell Vaccination of Patients With Metastatic Breast and Renal Cancer Induces Immunological and Clinical Responses. *Clin Cancer Res* (2004) 10:4699–708. doi: 10.1158/1078-0432.Ccr-04-0347
- Gelao L, Criscitiello C, Esposito A, De Laurentiis M, Fumagalli L, Locatelli M, et al. Dendritic Cell-Based Vaccines: Clinical Applications in Breast Cancer. *Immunotherapy* (2014) 6:349–60. doi: 10.2217/imt.13.169
- Zhang P, Yi S, Li X, Liu R, Jiang H, Huang Z, et al. Preparation of Triple-Negative Breast Cancer Vaccine Through Electrofusion With Day-3 Dendritic Cells. *PLoS One* (2014) 9:e102197. doi: 10.1371/ journal.pone.0102197
- Pierce R, Campbell J, Pai S, Brody J, Kohrt H. In-Situ Tumor Vaccination: Bringing the Fight to the Tumor. Hum Vaccines Immunother (2015) 11:1901–9. doi: 10.1080/21645515.2015.1049779
- Yokoi T, Oba T, Kajihara R, Abrams S, Ito F. Local, Multimodal Intralesional Therapy Renders Distant Brain Metastases Susceptible to PD-L1 Blockade in a Preclinical Model of Triple-Negative Breast Cancer. *Sci Rep* (2021) 11:21992. doi: 10.1038/s41598-021-01455-4
- Savage T, Pandey S, Guha C. Postablation Modulation After Single High-Dose Radiation Therapy Improves Tumor Control via Enhanced Immunomodulation. Clin Cancer (2020) 26:910–21. doi: 10.1158/1078-0432.Ccr-18-3518
- Cadena A, Cushman T, Anderson C, Barsoumian H, Welsh J, Cortez M. Radiation and Anti-Cancer Vaccines: A Winning Combination. *Vaccines* (2018) 6:9. doi: 10.3390/vaccines6010009
- 92. Demaria S, Ng B, Devitt M, Babb J, Kawashima N, Liebes L, et al. Ionizing Radiation Inhibition of Distant Untreated Tumors (Abscopal Effect) is Immune Mediated. *Int J Radiat Oncol Biol Phys* (2004) 58:862–70. doi: 10.1016/j.ijrobp.2003.09.012
- Hiniker S, Chen D, Reddy S, Chang D, Jones J, Mollick J, et al. A Systemic Complete Response of Metastatic Melanoma to Local Radiation and Immunotherapy. *Trans Oncol* (2012) 5:404–7. doi: 10.1593/tlo.12280
- 94. Grimaldi A, Simeone E, Giannarelli D, Muto P, Falivene S, Borzillo V, et al. Abscopal Effects of Radiotherapy on Advanced Melanoma Patients Who Progressed After Ipilimumab Immunotherapy. *Oncoimmunology* (2014) 3: e28780. doi: 10.4161/onci.28780
- 95. Theurich S, Rothschild S, Hoffmann M, Fabri M, Sommer A, Garcia-Marquez M, et al. Local Tumor Treatment in Combination With Systemic Ipilimumab Immunotherapy Prolongs Overall Survival in Patients With Advanced Malignant Melanoma. *Cancer Immunol Res* (2016) 4:744–54. doi: 10.1158/2326-6066.Cir-15-0156
- 96. Skalina K, Singh S, Chavez C, Macian F, Guha C. Low Intensity Focused Ultrasound (LOFU)-Mediated Acoustic Immune Priming and Ablative Radiation Therapy for *in Situ* Tumor Vaccines. *Sci Rep* (2019) 9:15516. doi: 10.1038/s41598-019-51332-4
- 97. Ito A, Honda H, Kobayashi T. Cancer Immunotherapy Based on Intracellular Hyperthermia Using Magnetite Nanoparticles: A Novel Concept of "Heat-Controlled Necrosis" With Heat Shock Protein Expression. Cancer Immunol Immunother CII (2006) 55:320-8. doi: 10.1007/s00262-005-0049-y
- 98. Singer CF, Pfeiler G, Hubalek M, Bartsch R, Stöger H, Pichler A, et al. Efficacy and Safety of the Therapeutic Cancer Vaccine Tecemotide (L-BLP25) in Early Breast Cancer: Results From a Prospective, Randomised, Neoadjuvant Phase II Study (ABCSG 34). Eur J Cancer (2020) 132:43–52. doi: 10.1016/j.ejca.2020.03.018
- 99. Vassilaros S, Tsibanis A, Tsikkinis A, Pietersz GA, Mckenzie IF, Apostolopoulos V. Up to 15-Year Clinical Follow-Up of a Pilot Phase III Immunotherapy Study in Stage II Breast Cancer Patients Using Oxidized Mannan-MUC1. *Immunotherapy* (2013) 5:1177–82. doi: 10.2217/ imt.13.126

- 100. Kawashima I, Hudson SJ, Tsai V, Southwood S, Takesako K, Appella E, et al. The Multi-Epitope Approach for Immunotherapy for Cancer: Identification of Several CTL Epitopes From Various Tumor-Associated Antigens Expressed on Solid Epithelial Tumors. *Hum Immunol* (1998) 59:1–14. doi: 10.1016/s0198-8859(97)00255-3
- Peoples G,E. Clinical Trial Results of a HER2/neu (E75) Vaccine to Prevent Recurrence in High-Risk Breast Cancer Patients. J Clin Oncol (2005) 23:7536–45. doi: 10.1200/JCO.2005.03.047
- 102. Mittendorf EA, Clifton GT, Holmes JP, Schneble E, Van Echo D, Ponniah S, et al. Final Report of the Phase I/II Clinical Trial of the E75 (Nelipepimut-S) Vaccine With Booster Inoculations to Prevent Disease Recurrence in High-Risk Breast Cancer Patients. Ann Oncol (2014) 25:1735–42. doi: 10.1093/annonc/mdu211
- 103. Mittendorf EA, Lu B, Melisko M, Hiller JP, Bondarenko I, Brunt AM, et al. Efficacy and Safety Analysis of Nelipepimut-S Vaccine to Prevent Breast Cancer Recurrence: A Randomized, Multicenter, Phase III Clinical Trial. *Clin Cancer Res* (2019) 25:4248–54. doi: 10.1158/1078-0432.CCR-18-2867
- 104. Gall VA, Philips AV, Na Q, Clise-Dwyer K, Mittendorf EA. Trastuzumab Increases HER2 Uptake and Cross-Presentation by Dendritic Cells. Cancer Res (2017) 77:5374–83. doi: 10.1158/0008-5472.CAN-16-2774
- 105. Clifton G, Hale D, Vreeland T, Hickerson A, Litton J, Alatrash G, et al. Results of a Randomized Phase IIb Trial of Nelipepimut-S + Trastuzumab Versus Trastuzumab to Prevent Recurrences in Patients With High-Risk HER2 Low-Expressing Breast Cancer. *Clin Cancer Res* (2020) 26:2515–23. doi: 10.1158/1078-0432.Ccr-19-2741
- 106. Ayoub NM, Al-Shami KM, Yaghan RJ. Immunotherapy for HER2-Positive Breast Cancer: Recent Advances and Combination Therapeutic Approaches. Breast Cancer: Targets Ther Volume (2019) 11:53–69. doi: 10.2147/ BCTT.S175360
- 107. Oka Y, Tsuboi A, Taguchi T, Osaki T, Kyo T, Nakajima H, et al. Induction of WT1 (Wilms' Tumor Gene)-Specific Cytotoxic T Lymphocytes by WT1 Peptide Vaccine and the Resultant Cancer Regression. *Proc Natl Acad Sci* USA (2004) 101:13885–90. doi: 10.1073/pnas.0405884101
- 108. Zhang W, Lu X, Cui P, Piao C, Xiao M, Liu X, et al. Phase I/II Clinical Trial of a Wilms' Tumor 1-Targeted Dendritic Cell Vaccination-Based Immunotherapy in Patients With Advanced Cancer. Cancer Immunol Immunother (2019) 68:121–30. doi: 10.1007/s00262-018-2257-2
- 109. Heery CR, Ibrahim NK, Arlen PM, Mohebtash M, Murray JL, Koenig K, et al. Docetaxel Alone or in Combination With a Therapeutic Cancer Vaccine (PANVAC) in Patients With Metastatic Breast Cancer: A Randomized Clinical Trial. JAMA Oncol (2015) 11087–95. doi: 10.1001/ jamaoncol.2015.2736
- 110. Takahashi R, Toh U, Iwakuma N, Takenaka M, Otsuka H, Furukawa M, et al. Feasibility Study of Personalized Peptide Vaccination for Metastatic Recurrent Triple-Negative Breast Cancer Patients. Breast Cancer Res (2014) 16:R70. doi: 10.1186/bcr3685
- 111. Toh U, Sakurai S, Saku S, Takao Y, Okabe M, Iwakuma N, et al. Early Phase II Study of Mixed 19-Peptide Vaccine Monotherapy for Refractory Triple-Negative Breast Cancer. *Cancer Sci* (2020) 111:2760–9. doi: 10.1111/ cas.14510
- 112. Vonderheide R, Domchek S, Schultze J, George D, Hoar K, Chen D, et al. Vaccination of Cancer Patients Against Telomerase Induces Functional Antitumor CD8+ T Lymphocytes. *Clin Cancer* (2004) 10:828–39. doi: 10.1158/1078-0432.ccr-0620-3
- 113. Kinney A, Sahin A, Vernon S, Frankowski R, Annegers J, Hortobagyi G, et al. The Prognostic Significance of Sialyl-Tn Antigen in Women Treated With Breast Carcinoma Treated With Adjuvant Chemotherapy. *Cancer* (1997) 80:2240–9. doi: 10.1002/(sici)1097-0142(19971215)80:12<2240::aidcncr4>3.0.co;2-y
- 114. Miles D, Roché H, Martin M, Perren TJ, Cameron DA, Glaspy J, et al. Phase III Multicenter Clinical Trial of the Sialyl-TN (STn)-Keyhole Limpet

Hemocyanin (KLH) Vaccine for Metastatic Breast Cancer. Oncologist (2011) 16:1092–100. doi: 10.1634/theoncologist.2010-0307

- 115. Ibrahim N, Murray J, Zhou D, Mittendorf E, Sample D, Tautchin M, et al. Survival Advantage in Patients With Metastatic Breast Cancer Receiving Endocrine Therapy Plus Sialyl Tn-KLH Vaccine: *Post Hoc* Analysis of a Large Randomized Trial. *J Cancer* (2013) 4:577–84. doi: 10.7150/jca.7028
- Domchek S, Recio A, Mick R, Clark C, Carpenter E, Fox K, et al. Telomerase-Specific T-Cell Immunity in Breast Cancer: Effect of Vaccination on Tumor Immunosurveillance. *Cancer Res* (2007) 67:10546–55. doi: 10.1158/0008-5472.Can-07-2765
- 117. Mcgranahan N, Swanton C. Clonal Heterogeneity and Tumor Evolution: Past, Present, and the Future. *Cell* (2017) 168:613–28. doi: 10.1016/ j.cell.2017.01.018
- Bertucci F, Ng CKY, Patsouris A, Droin N, Piscuoglio S, Carbuccia N, et al. Genomic Characterization of Metastatic Breast Cancers. *Nature* (2019) 569:560–4. doi: 10.1038/s41586-019-1056-z
- Heemskerk B, Kvistborg P, Schumacher T. The Cancer Antigenome. EMBO J (2013) 32:194–203. doi: 10.1038/emboj.2012.333
- 120. Mcdonald K, Kawaguchi T, Qi Q, Peng X, Asaoka M, Young J, et al. Tumor Heterogeneity Correlates With Less Immune Response and Worse Survival in Breast Cancer Patients. *Ann Surg Oncol* (2019) 26:2191–9. doi: 10.1245/ s10434-019-07338-3
- 121. Jager E, Ringhoffer M, Altmannsberger M, Arand M, Karbach J, Jager D, et al. Immunoselection *In Vivo*: Independent Loss of MHC Class I and Melanocyte Differentiation Antigen Expression in Metastatic Melanoma. *Int J Cancer* (1997) 71:142–7. doi: 10.1002/(sici)1097-0215(19970410)71:2<142:: aid-ijc3>30.co;2-0
- 122. Rosenthal R, Cadieux EL, Salgado R, Bakir MA, Moore DA, Hiley CT, et al. Neoantigen-Directed Immune Escape in Lung Cancer Evolution. *Nature* (2019) 567:479–85. doi: 10.1038/s41586-019-1032-7
- Armistead PM. Cellular Therapy Against Public Neoantigens. J Clin Invest (2019) 129:506–8. doi: 10.1172/jci126116
- 124. Charoentong P, Finotello F, Angelova M, Mayer C, Efremova M, Rieder D, et al. Pan-Cancer Immunogenomic Analyses Reveal Genotype-Immunophenotype Relationships and Predictors of Response to Checkpoint Blockade. *Cell Rep* (2017) 18:248-62. doi: 10.1016/ j.celrep.2016.12.019
- 125. Kim Y, Kang S, Shin H, Kim T, Yu B, Kim J, et al. Sequential and Timely Combination of a Cancer Nanovaccine With Immune Checkpoint Blockade Effectively Inhibits Tumor Growth and Relapse. Angewandte Chemie (International Ed English) (2020) 59:14628–38. doi: 10.1002/anie.202006117
- 126. Antonia S, Villegas A, Daniel D, Vicente D, Murakami S, Hui R, et al. Durvalumab After Chemoradiotherapy in Stage III Non-Small-Cell Lung Cancer. N Engl J Med (2017) 377:1919–29. doi: 10.1056/NEJMoa1709937

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2022 Zhang, Zhou, Sha, Xie and Liu. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.