Original Article

Risk factors of multidrug-resistant *Acinetobacter baumannii* recurrence after successful eradication in ventilated patients

Chiung-Yu Lin [a], *Yu-Mu Chen* [a], *Meng-Chih Lin* [a], *Yu-Ping Chang* [a], *Tung-Ying Chao* [a], *Chin-Chou Wang* [a], *Yuh-Chyn Tsai* [b], *Lien Shi Shen* [b], *Chin-Ling Li* [b], *An-Shen Lin* [a,*]

[a] Division of Pulmonary and Critical Care Medicine, Department of Internal Medicine, Kaohsiung Chang Gung Memorial Hospital, Chang Gung University College of Medicine, Kaohsiung, Taiwan
[b] Department of Respiratory Therapy, Kaohsiung Chang Gung Memorial Hospital, Chang Gung University College of Medicine, Kaohsiung, Taiwan

ARTICLE INFO

ABSTRACT

Background: Clinically, multidrug-resistant *Acinetobacter baumannii* (MDR-AB) recurrence is found in some patients although identified as successfully eradicated. We aim to discover the characteristics of patients with MDR-AB recurrence in the respiratory tract.
Methods: We retrospectively collected 106 chronic respiratory failure patients with MDR-AB harvest in pulmonary secretion culture.
Results: MDR-AB was successfully eradicated in 69 patients. Diabetes mellitus ($p = 0.030$, odds ratio [OR]: 2.7, 95% confidence interval [CI]: 1.1—6.4) and acute respiratory distress syndrome ($p = 0.001$, OR = 4.8, 95% CI: 1.8—12.7) reduce the MDR-AB eradication rate. Besides, a classification of colonization or infection was made beyond the 69 MDR-AB eradicated patients. In the colonization group, diabetes mellitus ($p = 0.009$; OR = 5.1, 95% CI: 1.5—17.6) is the only independent factor to increase the recurrence rate. Glycated hemoglobin level is also analyzed for each group to investigate diabetes control effect, but no significant difference found.
Conclusions: Diabetes mellitus is a risk factor of MDR-AB recurrence among MDR-AB-colonized patients; the impact of localized pneumonia patch in MDR-AB-infected patients requires further study to be clarified.

* *Corresponding author.* Pulmonary and Critical Care Medicine, Kaohsiung Chang Gung Memorial Hospital, 123, Dapi Rd., Niaosong Dist., Kaohsiung 833, Taiwan. Tel.: +886 7 7317123 ext. 8199; fax: +886 7 7322402.
    E-mail address: anshen@adm.cgmh.org.tw (A.-S. Lin).
    Peer review under responsibility of Chang Gung University.

At a glance commentary

Scientific background of the subject

The recurrence of multidrug-resistant *A. baumannii* (MDR-AB) after treatment is often seen, and it causes a great medical waste in medical centers.

What this study adds to the field

To discover the risk factors of MDR-AB recurrence gives assistance to future disease control policy formulation. Besides, the information may contribute in early detection of MDR-AB recurrence and thus preventing possible outbreaks.

Multidrug-resistant *A. baumannii* (MDR-AB) emerges as a cause of numerous global outbreaks via its strong adaptation [1–3], and it admittedly causes a considerable medical waste in most medical centers. The mechanisms of drug resistance acquisition in *A. baumannii* include obtaining mobile genetic elements, such as plasmids, transposons, or integrons from the surrounding [4–7]. Series of literatures contribute to the microbial agent selection against MDR-AB, including inhaled or intravenous colistin methanesulfonate, Sulbactam, or tigecycline [8–11]; However, few articles trace the disease after cured.

In clinical practice, recurrent MDR-AB respiratory tract acquisition is found in some cases although they are declared MDR-AB is successfully eradicated. The recurrent isolates easily cause an outbreak and threaten the critically ill hospitalized patients if not detected [12]. We suppose the risk factors of MDR-AB recurrence exist, and recognizing these risk factors is a benefit in clarifying the patients prone to MDR-AB recurrence. A new disease control policy may be established to detect early and control the recurrent MDR-AB. The purpose of this study is to discover the risk factors of pulmonary MDR-AB recurrence in chronic respiratory failure patients after successful eradication.

Methods

Setting and study population

Kaohsiung Chang Gung Memorial Hospital is a 2500-bed Teaching Medical Center in Southern Taiwan, and the Respiratory Care Center (RCC) is a stepwise weaning unit accumulating patients under ventilator dependence more than 21 days. Most of the patients come from internal medical, cardiologic, surgical, or neurological intensive care units. One who fails to wean his ventilator in 42 days after admitted to RCC will be transferred to out-hospital chronic facilities, or our ordinary wards, or even back to intensive care unit. The transferring indication is based on patients' medical condition.

The patients in the RCC in Kaohsiung Chang Gung Memorial Hospital between June 1, 2009, and December 31, 2011, were enrolled. As we know, there are some MDR-AB outbreaks in the period. The laboratory profile is more complete in the period as well. The inclusion criteria include age over 18 years old and harvested with MDR-AB in pulmonary secretions culture between June 1, 2009, and December 31, 2011. The exclusion criteria were patients without tracheal intubation or tracheostomy, without antibiotics therapy, with MDR-AB bacteremia or wound infection (either surgical or nonsurgical), and prior MDR-AB acquisition before the data collection.

Definition

MDR-AB is defined to be insusceptible to all of the following classes of antimicrobials: Aminoglycosides, antipseudomonal carbapenems, antipseudomonal fluoroquinolones, antipseudomonal penicillins, carbapenems, monobactams, quinolones, aminoglycosides, tetracycline, or trimethoprim/sulfamethoxazole [13,14]. Acute respiratory distress syndrome (ARDS) is defined as arterial oxygen tension to fraction of inspired oxygen ratio less than 200 [15].

The term "chronic respiratory failure" is defined as ventilator use more than 21 days, and the term "successful eradication" complies with our hospital's disease control and prevention policy that two sets of pulmonary secretion specimens in a row yield no MDR-AB in culture examinations. All the specimens should be specified for "MDR-AB detection", and the secretion quality should be approved by our laboratory inspector. Unqualified specimens, e.g., saliva-contaminated, are not counted. Once the MDR-AB is not eradicated in 4 weeks after antibiotics initiation, the treatment fails.

If the patients received aerosolized colistin therapy only, we sorted these patients into "colonization group". On the other hand, people are identified as "infection group" if they received any kinds of intravenous anti-MDR-AB antibiotics. This classification matches with the conventional management for MDR-AB pulmonary acquisition in our hospital. The patient's sputum culture is traced for 1 month after successful eradication. If one or more sets of sputum culture harvesting MDR-AB during the first 1 month, we define that MDR-AB recurs.

Study design

We recorded each patient's MDR-AB culture harvesting date, antibiotics initiation date, MDR-AB successful eradication date, MDR-AB recurrence date, and antibiotics duration; meanwhile, the interval between MDR-AB cultures harvesting date to the antibiotics initiation date is calculated.

We also collected the patients' basic information (age, gender, cigarette smoking history, and alcohol drinking history), comorbidities (diabetes mellitus, hypertension, coronary artery disease, chronic kidney disease (CKD), liver cirrhosis, chronic obstructive pulmonary disease, cerebral vascular accident, and cancer history), clinical features (ear temperature, pulse rate, respiration rate, blood pressure, fraction of inspiration oxygen, and ventilator days prior to MDR-AB treatment), and laboratory exam data (hemogram, blood gas analysis profiles, C-reactive protein, blood creatine level, sodium, and potassium). In addition, patients' disease severity was measured in Acute

Physiology and Chronic Health Evaluation scoring system (APACHE II) [16].

Initially, we compared the variables to look for the factors affecting MDR-AB eradication. A categorization was made on the MDR-AB-eradicated patients into colonization groups and infection groups. The variables were then analyzed to discover the risk factors of MDR-AB recurrence in each group (overall, colonization, and infection group).

The sputum culture examination after MDR-AB eradication is not a routine check-up, and we re-examine the sputum if any sign of airway infection (e.g., fever, radiologic patch, or purulent sputum). Our RCC patients may be transferred to respiratory care wards, kinds of chronic facilities adopting long-term ventilated patients, if they fail to wean the ventilators. For these patients, we can obtain their sputum specimen if they are brought back to our emergency depart to treat upper airway infection.

Owing to the finding of the diabetes's influence to MDR-AB recurrence, a secondary investigation of diabetes control is performed via analyzing glycated hemoglobin (HbA1c) level on each group. We calculated the HbA1c receiver operating characteristic (ROC) curve to find the cut-off point, and then a comparison between better sugar control and worsen control was made.

Statistical analysis

Variables were analyzed with Chi-square test, Fisher's exact test, Student's t-test, or the Mann–Whitney U-test. Continuous, normally distributed variables were presented as mean (standard deviation), whereas non-normally distributed variables were presented as median (interquartile range). Variables with $p < 0.05$ in univariate analysis were included into a multivariate logistic regression.

Ethics

This is a retrospective study, and the study protocol was approved by the Chang Gung Medical Foundation Institutional Review Board (Project No. 103-4706B).

Results

From June 1, 2009, to December 31, 2011, we enrolled 295 patients with MDR-AB harvest in pulmonary secretion culture examination. After filtered with exclusion criteria, 106 patients remained and 69 (65.1%) among these patients were MDR-AB successfully eradicated [Fig. 1]. Of the variables compared, diabetes mellitus (51% vs. 30.4%, $p = 0.034$) and ARDS (43.2% vs. 14.5%, $p = 0.001$) are the two factors to reduce the MDR-AB eradication rate. The p-values of both variables in multivariable analysis are <0.05 (diabetes: OR: 2.7, 95% CI: 1.1–6.4; ARDS OR: 4.8, 95% CI: 1.8–12.7) [Table 1]. There is no significant difference in APACHE II scores between the treatment failure group and the successful eradication group.

MDR-AB recurred in 18 (26.1%) patients after 1 month [Table 2]; we analyzed the MDR-AB recurrence risk factors in 3 groups (overall patient group, MDR-AB colonization group, and MDR-AB infection group). For the overall MDR-AB
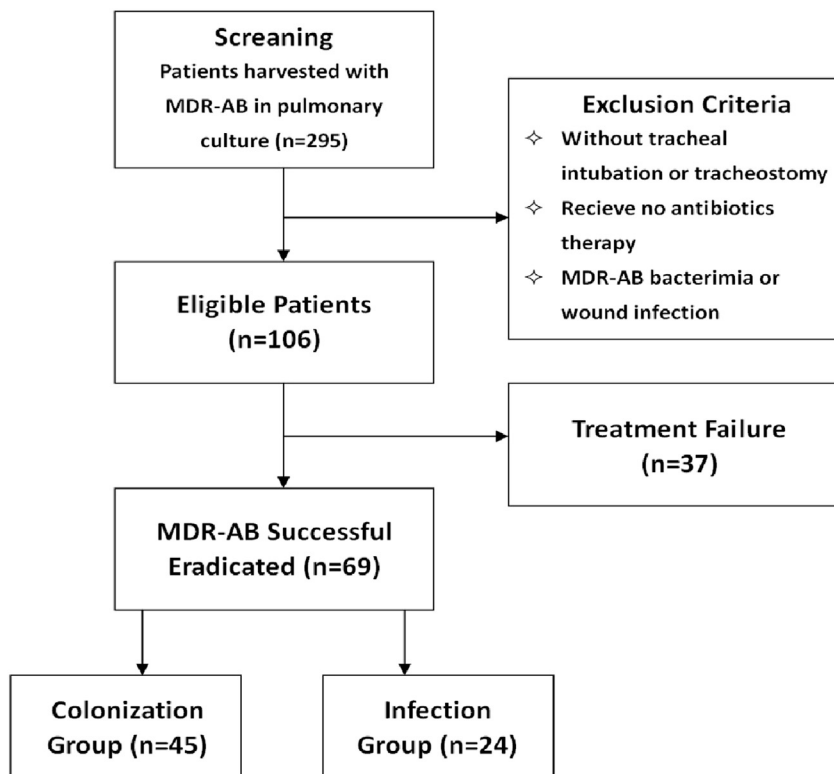

Fig. 1 – Multidrug-resistant *Acinetobacter baumannii* was successfully eradicated in 69 patients and a categorization is made on these patients into colonization or infection group.

Table 1 — Baseline factors to eradicate multidrug-resistant *Acinetobacter baumannii*.

Variables	Without eradication (n = 37)	Eradication (n = 69)	Univariate analysis P	Multivariate analysis P	OR (95% CI)
Age, mean (SD)	76.0 (10.9)	71.4 (14.0)	0.085		
Gender (male), n (%)	21 (56.8)	40 (58.0)	0.904		
APACHE II, median (IQR)[a]	20 (18.5—21)	20 (17—23)	0.839		
Comorbidities					
Diabetes, n (%)	19 (51.4)	21 (30.4)	0.034	0.030	2.7 (1.1—6.4)
Hypertension, n (%)	22 (59.5)	42 (60.9)	0.887		
Coronary artery disease, n (%)	6 (16.2)	15 (21.7)	0.496		
Chronic kidney disease, n (%)	6 (16.2)	14 (20.3)	0.609		
Liver cirrhosis, n (%)	3 (8.1)	1 (1.4)	0.121		
COPD, n (%)	8 (21.6)	12 (17.4)	0.596		
Cerebral vascular accident, n (%)	7 (18.9)	21 (30.4)	0.200		
Cancer, n (%)	5 (13.5)	14 (20.3)	0.386		
Past histories					
Cigarette smoking, n (%)	6 (16.2)	12 (17.4)	0.878		
Alcohol drinking, n (%)	3 (8.1)	6 (8.7)	1.000		
Pulmonary TB history, n (%)	2 (5.4)	6 (8.7)	0.710		
Laboratory profiles					
WBC, median (IQR)	11,300 (9150—17,400)	11,200 (8500—14,750)	0.327		
Hematocrit, mean (SD)	31.0 (4.6)	30.4 (4.7)	0.537		
Creatine, median (IQR)	0.77 (0.60—2.09)	0.86 (0.55—2.12)	0.866		
C-reactive protein, median (IQR)	59.6 (25.0—88.2)	60.1 (34.0—141.2)	0.276		
Clinical profiles					
Blood pH, mean (SD)	7.44 (0.07)	7.44 (0.07)	0.940		
FiO$_2$ (%), median (IQR)	40 (35—40)	40 (32.5—40)	0.308		
PaO$_2$/FiO$_2$ < 200, n (%)	16 (43.2)	10 (14.5)	0.001	0.001	4.8 (1.8—12.7)
Glasgow Coma Scale, median (IQR)	10 (3—11)	9.5 (3—11)	0.537		
Shock, n (%)	12 (32.4)	13 (18.8)	0.116		
Radiology					
Localized patch, n (%)[b]	26 (70.3)	41 (59.4)	0.270		

Abbreviations: SD: Standard deviation; IQR: Interquartile range; APACHE II: Acute Physiology and Chronic Health Evaluation scoring system; COPD: Chronic obstructive pulmonary disease; TB: Tuberculosis; WBC: White blood cell; OR: Odds ratio; CI: Confidence interval; FiO$_2$: Fraction of inspired oxygen.
[a] APACHE II score was measured on days of first pulmonary sputum collection.
[b] The radiologic character of localized patch is determined by at least one licensed chest or radiology doctor.

eradicated patients, diabetes mellitus (21.6% vs. 55.6%, $p = 0.009$, OR [95% CI]: 5.1 [1.5—17.6]) and localized pneumonia patch (51.0% vs. 88.9%, $p = 0.012$, OR [95% CI]: 8.2 [1.6—42.2]) were the two recurrence risk factors. Among the colonization group, diabetes (14.3% vs. 50.0%, $p = 0.017$) and ARDS (5.7% vs. 30.0%, $p = 0.031$) show a significant difference in univariate analysis [Table 3]. Both diabetes and ARDS were included into a multivariate logistic regression, and the multivariate p-value of diabetes and ARDS are 0.039 and 0.085, respectively. Diabetes becomes the only independent risk factor ($p = 0.039$; OR = 5.6, 95% CI: 1.09—29.03). As regards to the infection group, localized patch (50% vs. 100%, $p = 0.022$) passes the 0.05 threshold in univariate p-value [Table 3].

A secondary investigation of HbA1c is performed in each study group. A total of 29 and 16 pieces of HbA1c data are obtained in the raw eligible patients (diabetes number: 40) and successful eradication patients (diabetes number: 21). The HbA1c ROC curve is analyzed in both the overall group and the MDR-AB colonization group. Of the overall group, the cut-off point is 7.0 and the area under the curve (AUC) is 0.73. Nevertheless, HbA1c < 7.0 ($n = 7$) and HbA1c > 7.0 ($n = 9$) make no significant differences in respect to MDR-AB recurrence (25.0% vs. 62.5%, $p = 0.315$). There are only seven piece of data obtained in the colonization group; more case numbers are

required for further analysis in this sub-group. The information of antibiotics selection in each group is provided in Table 4.

Discussion

Acinetobacter baumannii is a Gram-negative *coccobacillus* and tolerates long periods in both moist and dry conditions [2,3], and the easy acquisition of drug resistance makes it a great challenge in disease control. *A. baumannii* has become a leading cause of nosocomial pneumonia and bacteremia globally [17,18]; Nevertheless, some people doubt its clinical influence on morbidity and mortality. Recent studies began to associate *A. baumannii* to increased mortality and prolonged intensive care unit days [19,20]. Falagas et al. conducted a nine-study systematic review and demonstrate the relationship between *A. baumannii* and increased mortality [21]. He advocates that clinicians should make every effort to combat them. In fact, physicians have invested much investigation in *A. baumannii* opposition, including antimicrobial agent selection and anti-infection measures such as contact precautions, meticulous environmental decontamination, hand washing, or even chlorhexidine baths [8—12,22,23]. MDR-AB sometimes recurs in respiratory tract even after successful eradication, and the

Table 2 – Recurrence for the overall MDR-AB-eradicated patients.

Variables	Without recurrence (n = 51)	Recurrence (n = 18)	Univariate analysis P	Multivariate analysis P	OR (95% CI)
Age, mean (SD)	70.7 (15.1)	73.7 (10.1)	0.603		
Gender (male), n (%)	31 (60.8)	9 (50.0)	0.426		
APACHE II, median (IQR)	20 (16–23)	21 (18–24)	0.195		
Chronic comorbidities					
Diabetes, n (%)	11 (21.6)	10 (55.6)	0.007	0.009	5.1 (1.5–17.6)
Hypertension, n (%)	29 (56.9)	13 (72.2)	0.251		
Coronary artery disease, n (%)	12 (23.5)	3 (16.7)	0.743		
Chronic kidney disease, n (%)	10 (19.6)	4 (22.2)	1.000		
Liver cirrhosis, n (%)	0 (0.0)	1 (5.6)	0.261		
COPD, n (%)	10 (19.6)	2 (11.1)	0.718		
Cerebral vascular accident, n (%)	13 (25.5)	8 (44.4)	0.133		
Cancer, n (%)	11 (21.6)	3 (16.7)	0.747		
Past histories					
Cigarette smoking, n (%)	10 (19.6)	2 (11.1)	0.718		
Alcohol drinking, n (%)	4 (7.8)	2 (11.1)	0.647		
Pulmonary TB history, n (%)	6 (11.8)	0 (0.0)	0.328		
Laboratory profiles					
WBC, median (IQR)	10,500 (8300–14,100)	11,650 (9000–17,050)	0.224		
Hematocrit, mean (SD)	30.3 (4.7)	30.8 (5.0)	0.703		
Creatine, median (IQR)	0.86 (0.59–1.82)	1.05 (0.46–3.60)	0.989		
C-reactive protein, median (IQR)	59.5 (29.0–139.5)	79.9 (37.0–205.5)	0.557		
Clinical profiles					
Blood pH, mean (SD)	7.44 (0.07)	7.43 (0.10)	0.563		
FiO_2, median (IQR) (%)	35 (35–40)	40 (35–40)	0.554		
$PaO_2/FiO_2 < 200$, n (%)	5 (11.1)	5 (20.8)	0.113		
Glasgow Coma Scale, median (IQR)	10 (7–10)	9.5 (7.5–11)	0.631		
Shock, n (%)	10 (20.0)	3 (16.7)	0.784		
Radiologic characteristics					
Localized patch, n (%)	26 (51.0)	16 (88.9)	0.005	0.012	8.2 (1.6–42.2)
Treatment					
Days from culture positive to antibiotics initiation, median (IQR)	3 (2–6)	3 (3–4)	0.743		
Antibiotics duration, median (IQR)	11 (9–14)	9.5 (6.75–19)	0.929		

Abbreviations: MDR-AB: Multidrug-resistant *Acinetobacter baumannii*; SD: Standard deviation; IQR: Interquartile range; APACHE II: Acute Physiology and Chronic Health Evaluation scoring system; COPD: Chronic obstructive pulmonary disease; TB: Tuberculosis; WBC: White blood cell; FiO_2: Fraction of inspired oxygen; OR: Odds ratio; CI: Confidence interval.

recurrent isolates being a great burden on disease control since they easily cause cross-contamination before detected.

This article investigated the risk factors of MDR-AB recurrence. We excluded patients without tracheal intubation or tracheostomy to make sure the entire specimens were collected from pulmonary secretion rather than saliva, and thus increasing the reliability of the microbial culture examination result. People with MDR-AB bacteremia or wound infection are excluded to confirm the recurrent MDR-AB isolates not related to systemic spreading. Although 65.1% people are MDR-AB eradicated, 26.1% develop MDR-AB re-acquisition after 1 month.

In addition, diabetes mellitus and ARDS are the two factors reducing MDR-AB eradication rate in our study. ARDS can be interpreted to be a more severe infectious status, and it is reasonable to have a lower cure rate. As regard to diabetes, we interestingly found that diabetes is a risk factor of treatment failure as well to the MDR-AB recurrence (either the overall patient group or the MDR-AB colonization group). Our hypothesis is that diabetes mellitus interferes with our immune system and raise difficulties in pathogen eradication. Previous studies have discovered the dysfunctions of diabetic polymorphonuclear cells and diabetic monocytes/macrophages in

chemotaxis, phagocytosis, and killing [24]. The declined *A. baumannii* population after antimicrobial agent therapy may not be totally eliminated but causing false-positive result in eradication. The residual MDR-AB pathogen re-grows after antibiotic treatment suspension. A multicenter case–control study in Korea shows that diabetes mellitus is independently associated with hospital-acquired pneumonia caused by carbapenem-resistant Gram-negative bacteria. In this study, *A. baumannii* is the major pathogens of hospital-acquired pneumonia and accounted for 72.1% of the total carbapenem-resistant Gram-negative bacteria isolates [25]. This Korea study may drop a hint of a possible relation between diabetes and higher MDR-AB pneumonia incidence. Nevertheless, the influence of diabetes vanishes (17.1% vs. 62.6%, $p = 0.235$) when we focus on the MDR-AB infection group. Our explanation is that people in the infection group received intravenous anti-MDR-AB agents rather than aerosolized colistin only; the antibiotics selection may somehow affect the clearance of MDR-AB. The higher clear effect may lower the residual MDR-AB and overshadow the diabetes influence.

Localized pneumonia patch in the chest plain film is another risk factor of MDR-AB recurrence in the overall

Variables	Without recurrence (n = 35, 16)[a]	Recurrence (n = 10, 8)[b]	Univariate analysis P	Multivariate analysis P	OR (95% CI)[d]
Table 3 – MDR-AB recurrence for colonization group and infection group.					
Age, mean (SD)					
Colonization	72.0 (13.7)	72.4 (10.7)	0.932		
Infection	67.7 (17.9)	75.3 (9.8)	0.279		
Gender (male), n (%)					
Colonization	22 (62.9)	7 (70.0)	0.677		
Infection	9 (56.3)	2 (25.0)	0.156		
APACHE II, median (IQR)					
Colonization	20 (15–23)	19.5 (18–22)	0.935		
Infection	20 (17–23)	23 (20–24)	0.140		
Chronic comorbidities					
Diabetes, n (%)					
Colonization	5 (14.3)	5 (50.0)	0.017	0.039	5.6 (1.09–29.03)
Infection	6 (17.1)	5 (62.6)	0.235		
Hypertension, n (%)					
Colonization	20 (57.1)	5 (50.0)	0.688		
Infection	9 (25.7)	8 (100.0)	0.054		
Coronary artery disease, n (%)					
Colonization	7 (20.0)	2 (20.0)	1.000		
Infection	5 (14.3)	1 (12.5)	0.319		
Chronic kidney disease, n (%)					
Colonization	9 (25.7)	1 (10.0)	0.292		
Infection	1 (2.9)	3 (37.5)	0.091		
Liver cirrhosis, n (%)					
Colonization	0 (0.0)	0 (0.0)	–		
Infection	0 (0.0)	1 (12.5)	0.333		
COPD, n (%)					
Colonization	6 (17.1)	2 (20.0)	0.835		
Infection	4 (11.4)	0 (0.0)	0.171		
Cerebral vascular accident, n (%)					
Colonization	11 (31.4)	5 (50.0)	0.279		
Infection	2 (5.7)	3 (37.5)	0.186		
Cancer, n (%)					
Colonization	9 (25.7)	2 (20.0)	0.711		
Infection	2 (5.7)	1 (12.5)	0.751		
Past histories					
Cigarette smoking, n (%)					
Colonization	9 (25.7)	2 (20.0)	0.711		
Infection	1 (2.9)	0 (0.0)	0.667		
Alcohol drinking, n (%)					
Colonization	4 (11.4)	2 (20.0)	0.402		
Infection	0 (0.0)	0 (0.0)	–		
Pulmonary TB history, n (%)					
Colonization	5 (14.3)	0 (0.0)	0.328		
Infection	1 (2.9)	0 (0.0)	0.667		
Laboratory profiles					
WBC, median (IQR)					
Colonization	10,500 (6875–12,675)	11,400 (8075–17,050)	0.662		
Infection	10,100 (8600–15,600)	12,350 (9750–19,700)	0.133		
Hematocrit, mean (SD)					
Colonization	30.3 (4.5)	30.7 (5.1)	0.764		
Infection	30.3 (5.2)	30.9 (5.2)	0.796		
Creatine, median (IQR)					
Colonization	0.86 (0.60–1.82)	0.82 (0.46–2.73)	0.754		
Infection	0.81 (0.56–1.81)	1.60 (0.35–4.08)	0.830		
C-reactive protein, median (IQR)					
Colonization	67.0 (23.0–139.5)	44.6 (29.1–104.7)	0.548		
Infection	54.1 (37.4–148.4)	153.1 (56.8–222.0)	0.159		
Clinical profiles					
Blood pH, mean (SD)					
Colonization	7.44 (0.06)	7.45 (0.12)	0.118		
Infection	7.44 (0.07)	7.39 (0.07)	0.118		

(continued on next page)

Table 3 – (continued)					
Variables	Without recurrence $(n = 35, 16)$[a]	Recurrence $(n = 10, 8)$[b]	Univariate analysis P	Multivariate analysis P	OR (95% CI)[d]
FiO_2 (%), median (IQR)					
Colonization	35 (30–40)	40 (35–40)	0.261		
Infection	35 (37.5–40)	40 (32.5–47.5)	0.551		
$PaO_2/FiO_2 < 200$, n (%)					
Colonization	2 (5.7)	3 (30.0)	0.031		
Infection	3 (8.6)	2 (25.0)	0.593		
Glasgow Coma Scale, median (IQR)					
Colonization	10 (7–11)	7.25 (8.5–10.25)	0.631		
Infection	8.5 (7–10)	11 (6.75–11)	0.188		
Shock, n (%)					
Colonization	8 (22.9)	2 (20.0)	0.848		
Infection	2 (5.7)	1 (12.5)	0.751		
Radiologic characteristics					
Localized patch, n (%)					
Colonization	18 (51.4)	8 (80.0)	0.297		
Infection	8 (50)	8 (100)	0.022[c]		
Treatment					
Days from culture positive to antibiotics initiation, median (IQR)					
Colonization	3 (2–6)	3 (2–4)	0.429		
Infection	3 (2–5)	4 (3–4)	0.432		
Antibiotics duration, median (IQR)					
Colonization	11 (9–14)	9.5 (6.75–11.25)	0.099		
Infection	11 (9–20)	14 (7–22)	0.877		

Abbreviations: MDR-AB: Multidrug-resistant *Acinetobacter baumannii*; SD: Standard deviation; IQR: Interquartile range; APACHE II: Acute Physiology and Chronic Health Evaluation scoring system; COPD: Chronic obstructive pulmonary disease; TB: Tuberculosis; WBC: White blood cell; FiO_2: Fraction of inspired oxygen; OR: Odds ratio; CI: Confidence interval.
[a] The number of "colonization group" and "infection group" in without recurrence section is 35 and 16, respectively.
[b] The number of "colonization group" and "infection group" in recurrence section is 10 and 8, respectively.
[c] Localized patch is the only risk factor of recurrence in the infection group; however, there is no variation in the recurrence group. More subjects for the infection group are required to clarify this impact.
[d] The ORs are obtained from the univariate analysis, instead of multiple logistic regression.

patient group. The localized patches probably represent a higher pathogen burden, and the pathogen may not be fully eliminated under inadequate treatment course or drug concentration. For the infection group, localized patch (univariate analysis, $p = 0.022$) is the only risk factor of recurrence in the infection group. However, there is no variation in the recurrence group. More subjects for the infection group are required to clarify this impact.

Diabetes and ARDS ($PaO_2/FiO_2 < 200$) shows a significant difference in univariate analysis in the "colonization group" for MDR-AB recurrence, but diabetes mellitus becomes the only independent risk factor after multivariate analysis. A certain relationship between diabetes and ARDS may account for the result, and diabetes is the dominate factor while comparing with ARDS. In fact, the relationship has been widely discussed; some people believe the intersection of

hyperglycemia, metabolic abnormalities, inflammation, or therapeutic agents contributing to the relationship [26].

Owing to the finding of diabetes influence on MDR-AB eradication and recurrence, we tried to discover the relationship between MDR-AB eradication and sugar control. HbA1c was chosen as the standard to characterize sugar control, because we think HbA1c is objective and quantitative. The ROC curve made a cut-point of HbA1c = 7.0 (AUC: 0.73), but $p > 0.05$ between HbA1c < 7.0 and HbA1c > 7.0. We did not disclaim the importance of diabetes control; on the other hand, more case numbers are required to confirm its influence. Another possible explanation is that HbA1c can only represent the sugar control in the past 3 months rather than the "current sugar control status" during antimicrobial therapy. However, it is difficult to characterize current sugar control status; it is also greatly affected by the operator of the

Table 4 – Comparing the initial antibiotics selection in each group.			
Variables	Without eradication, n (%)	Eradication, n (%)	Recurrence
Colonization group			
Aerosolized colistin only ($n = 64$)	19 (29.7)	45 (70.3)	10
Infection group			
Intravenous colistin ($n = 1$)	1 (100.0)	0 (0.0)	–
Tigecycline ($n = 9$)	7 (77.8)	2 (22.2)	1
Sulbactam ($n = 4$)	0 (0.0)	4 (100.0)	1
Aerosolized colistin + tigecycline ($n = 23$)	9 (39.1)	14 (60.9)	6
Aerosolized colistin + sulbactam ($n = 5$)	1 (20.0)	4 (80.0)	0

fingertip testing (e.g., insufficiency time between the food intake and fingertip testing).

The treatment on MDR-AB-colonized patients is controversial, and some physicians even advocate that antibiotics are not necessary in colonized cases. In our experience, MDR-AB colonization makes a huge medical waste since they may cause another outbreak if the lack of strict hand washing policies. Besides these, patients are often rejected from local chronic facilities and thus seriously reduced the bed availability in medical centers. In our opinion, the information for MDR-AB colonization in our study still provides certain use in practice if intensive MDR-AB eradication is demanded by clinical physicians.

To the best of our knowledge, our research is the first one to study the MDR-AB recurrence. A more aggressive disease control policy, prolonged duration of contact precautions, or meticulous environmental decontamination is considered on those infected by MDR-AB. Besides, we propose a higher standard eradication criteria and intensive pulmonary secretion culture check-up on these patients to prevent a possible cross-infection to other hosts.

There are limitations to this study. First, we excluded patients without antibiotics treatment in our study since they are usually identified to be MDR-AB-colonized. This group did not receive intensive pulmonary secretion culturing exam or laboratory exams and will easily cause bias. We, therefore, miss the information of MDR-AB recurrence in spontaneously MDR-AB-eradicated patients. A well-designed prospective study in the future may be required to solve the situation. Second, we do not perform molecular examination due to equipment limitation and failed to distinguish the recurrence MDR-AB from the origin strain or cross-infected strain. Last, most people enrolled in our study received aerosolized colistin rather than intravenous colistin; it is because the high proportion of old age or CKD patients in RCC. Concerning the possibility of colistin-related kidney injury [27], physicians seldom prescribe intravenous colistin unless MDR-AB bacteremia or high disease severity. However, this concept is challenged nowadays since some debate that colistin give a relatively high clinical cure rates and over-estimated in the incidence of kidney injury [28].

Conclusion

Diabetes mellitus and ARDS are risk factors to affect MDR-AB eradication. For the overall MDR-AB-eradicated patients, diabetes mellitus and localized pneumonia patch are the two risk factors. As regards to the MDR-AB colonization group, diabetes mellitus is the only independent factor to increase the recurrence rate. The localized patch in the infection group and the influence of HbA1c require further investigation. Recognizing the risk factors contributes to early detection of disease recurrence and preventing the possible cross-contamination to other hosts.

Source of support

Nil.

REFERENCES

[1] Villegas MV, Hartstein AI. *Acinetobacter* outbreaks, 1977–2000. Infect Control Hosp Epidemiol 2003;24:284–95.

[2] Wendt C, Dietze B, Dietz E, Rüden H. Survival of *Acinetobacter baumannii* on dry surfaces. J Clin Microbiol 1997;35:1394–7.

[3] Hirai Y. Survival of bacteria under dry conditions; from a viewpoint of nosocomial infection. J Hosp Infect 1991;19:191–200.

[4] Perez F, Hujer AM, Hujer KM, Decker BK, Rather PN, Bonomo RA. Global challenge of multidrug-resistant *Acinetobacter baumannii*. Antimicrob Agents Chemother 2007;51:3471–84.

[5] Seifert H, Boullion B, Schulze A, Pulverer G. Plasmid DNA profiles of *Acinetobacter baumannii*: clinical application in a complex endemic setting. Infect Control Hosp Epidemiol 1994;15:520–8.

[6] Devaud M, Kayser FH, Bächi B. Transposon-mediated multiple antibiotic resistance in *Acinetobacter* strains. Antimicrob Agents Chemother 1982;22:323–9.

[7] Segal H, Thomas R, Gay Elisha B. Characterization of Class 1 integron resistance gene cassettes and the identification of a novel IS-like element in *Acinetobacter baumannii*. Plasmid 2003;49:169–78.

[8] Kuo SC, Lee YT, Yang SP, Chen CP, Chen TL, Hsieh SL, et al. Eradication of multidrug-resistant *Acinetobacter baumannii* from the respiratory tract with inhaled colistin methanesulfonate: a matched case-control study. Clin Microbiol Infect 2012;18:870–6.

[9] Ho YH, Wang LS, Chao HJ, Chang KC, Su CF. Successful treatment of meningitis caused by multidrug-resistant *Acinetobacter baumannii* with intravenous and intrathecal colistin. J Microbiol Immunol Infect 2007;40:537–40.

[10] Lee CH, Tang YF, Su LH, Chien CC, Liu JW. Antimicrobial effects of varied combinations of meropenem, sulbactam, and colistin on a multidrug-resistant *Acinetobacter baumannii* isolate that caused meningitis and bacteremia. Microb Drug Resist 2008;14:233–7.

[11] Chuang YC, Cheng CY, Sheng WH, Sun HY, Wang JT, Chen YC, et al. Effectiveness of tigecycline-based versus colistin-based therapy for treatment of pneumonia caused by multidrug-resistant *Acinetobacter baumannii* in a critical setting: a matched cohort analysis. BMC Infect Dis 2014;14:102.

[12] Dijkshoorn L, Nemec A, Seifert H. An increasing threat in hospitals: multidrug-resistant *Acinetobacter baumannii*. Nat Rev Microbiol 2007;5:939–51.

[13] Wei HM, Hsu YL, Lin HC, Hsieh TH, Yen TY, Lin HC, et al. Multidrug-resistant Acinetobacter baumannii infection

among neonates in a neonatal intensive care unit at a medical center in central Taiwan. J Microbiol Immunol Infect 2015;48:531–9.

[14] Michalopoulos A, Fotakis D, Virtzili S, Vletsas C, Raftopoulou S, Mastora Z, et al. Aerosolized colistin as adjunctive treatment of ventilator-associated pneumonia due to multidrug-resistant Gram-negative bacteria: a prospective study. Respir Med 2008;102:407–12.

[15] Bernard GR, Artigas A, Brigham KL, Carlet J, Falke K, Hudson L, et al. The American-European Consensus Conference on ARDS. Definitions, mechanisms, relevant outcomes, and clinical trial coordination. Am J Respir Crit Care Med 1994;149(3 Pt 1):818–24.

[16] Knaus WA, Draper EA, Wagner DP, Zimmerman JE. APACHE II: a severity of disease classification system. Crit Care Med 1985;13:818–29.

[17] Kanafani ZA, Kara L, Hayek S, Kanj SS. Ventilator-associated pneumonia at a tertiary-care center in a developing country: incidence, microbiology, and susceptibility patterns of isolated microorganisms. Infect Control Hosp Epidemiol 2003;24:864–9.

[18] Gaynes R, Edwards JR, National Nosocomial Infections Surveillance System. Overview of nosocomial infections caused by gram-negative bacilli. Clin Infect Dis 2005;41:848–54.

[19] Fagon JY, Chastre J, Hance AJ, Montravers P, Novara A, Gibert C. Nosocomial pneumonia in ventilated patients: a cohort study evaluating attributable mortality and hospital stay. Am J Med 1993;94:281–8.

[20] Cisneros JM, Reyes MJ, Pachón J, Becerril B, Caballero FJ, García-Garmendía JL, et al. *Bacteremia* due to *Acinetobacter baumannii*: epidemiology, clinical findings, and prognostic features. Clin Infect Dis 1996;22:1026–32.

[21] Falagas ME, Bliziotis IA, Siempos II. Attributable mortality of *Acinetobacter baumannii* infections in critically ill patients: a systematic review of matched cohort and case-control studies. Crit Care 2006;10:R48.

[22] Boyce JM, Pittet D, Healthcare Infection Control Practices Advisory Committee. Society for healthcare epidemiology of America, association for professionals in infection control, infectious diseases society of America, hand hygiene task force. Guideline for hand hygiene in health-care settings: recommendations of the healthcare infection control practices advisory committee and the HICPAC/SHEA/APIC/IDSA hand hygiene task force. Infect Control Hosp Epidemiol 2002;23(12 Suppl.):S3–40.

[23] Pittet D. The lowbury lecture: behaviour in infection control. J Hosp Infect 2004;58:1–13.

[24] Geerlings SE, Hoepelman AI. Immune dysfunction in patients with diabetes mellitus (DM). FEMS Immunol Med Microbiol 1999;26:259–65.

[25] Kim T, Chong YP, Park SY, Jeon MH, Choo EJ, Chung JW, et al. Risk factors for hospital-acquired pneumonia caused by carbapenem-resistant gram-negative bacteria in critically ill patients: a multicenter study in Korea. Diagn Microbiol Infect Dis 2014;78:457–61.

[26] Honiden S, Gong MN. Diabetes, insulin, and development of acute lung injury. Crit Care Med 2009;37:2455–64.

[27] Kwon JA, Lee JE, Huh W, Peck KR, Kim YG, Kim DJ, et al. Predictors of acute kidney injury associated with intravenous colistin treatment. Int J Antimicrob Agents 2010;35:473–7.

[28] Spapen H, Jacobs R, Van Gorp V, Troubleyn J, Honoré PM. Renal and neurological side effects of colistin in critically ill patients. Ann Intensive Care 2011;1:14.