

Contents lists available at ScienceDirect

**Biochemistry and Biophysics Reports** 



journal homepage: www.elsevier.com/locate/bbrep

# Fundamental mechanisms of cell death for polycystic ovary syndrome

Ying-ying Li, Yi-qiu Peng, Yu-xi Yang, Ning Xu, Ting-juan Shi, Rui-xia Liu, Ying-yi Luan<sup>\*</sup>, Cheng-hong Yin<sup>\*\*</sup>

Department of Central Laboratory, Beijing Obstetrics and Gynecology Hospital, Capital Medical University. Beijing Maternal and Child Health Care Hospital, Beijing, 100026, China

ARTICLE INFO	A B S T R A C T		
Keywords:	Polycystic ovary syndrome (PCOS) is a common endocrine disorder in women of childbearing age with complex		
PCOS	symptoms and multiple hormone imbalances. Patients usually present with irregular menstruation, ovarian cysts,		
Apoptosis	and metabolic abnormalities. Current research has been found that multiple cell death programs may be involved		
Autophagy	in the occurrence of the disease. This article focuses on the mechanism between PCOS and six cell death pro-		
Ferroptosis	cesses including apoptosis, autophagy, ferroptosis, pyroptosis, NETosis and necroptosis. Ovarian granulosa cell		
Pyroptosis	apoptosis, autophagy, and ferroptosis play key roles in PCOS. In addition, pyroptosis and NETosis may also be		
NETosis	involved, but the specific mechanism needs further study. In general, a deeper understanding of these cell death		
Necroptosis	mechanisms will help develop innovative treatments for PCOS.		

## 1. Introduction

The history of cell death research can be traced back to the embryonic period of modern biology [1,2]. Scientists have gradually discovered that cell death is a highly regulated biological process. Among them, apoptosis, as the most classic form of programmed death, has completely changed our understanding of cell fate [3]. When a cell initiates the apoptotic program, it undergoes characteristic morphological changes: cell shrinkage, chromatin condensation, and eventually decomposition into apoptotic bodies that are cleared by neighboring cells or professional phagocytes. In the early 21st century, scientists discovered a unique iron-dependent death mode - ferroptosis. Unlike apoptosis, ferroptosis is more like an"oxidative disaster" of cells. When iron ion accumulation in cells triggers lipid peroxidation, the cell membrane system will suffer fatal damage. This mode of death is tightly connected to a range of metabolic irregularities, especially dysfunction of the glutathione system [4,5]. Autophagy, the breakdown of cellular components by lysosomes [6]. Pyroptosis, a form of inflammatory cell death triggered by infection or cellular stress [7]. Furthermore, several additional cell death programs have surfaced, such as NETotic cell death [8] and necroptosis [9]. These cell death processes are regulated by a complex regulatory network consisting of key molecules represented by

the caspase protease family and the Bcl-2 protein family [10-13]. When this system loses balance, it may cause major diseases such as tumors and autoimmune diseases [14,15]. Therefore, analyzing the mechanism of cell death is not only of theoretical significance, but also can provide new targets for disease treatment (see Table 1).

Polycystic ovary syndrome (PCOS) is a common endocrinopathy occurring in reproductive-age women [16]. Hyperandrogenism, polycystic ovaries, chronic anovulation, and metabolic aberrations are the common features in PCOS [17,18]. The development of PCOS is complex, influenced by a mix of genetic and environmental factors [19–24]. Recent research suggests that dysregulated cell death processes may serve as a critical link among these contributing factors. For instance, increased follicular atresia appears to be associated with imbalanced apoptosis; impaired autophagy may hinder normal follicular development; and insulin resistance may further disrupt these cellular pathways. Together, these disturbances contribute to ovarian dysfunction and are closely associated with metabolic abnormalities. A deeper understanding of the molecular mechanisms behind PCOS could offer valuable insights into its pathogenesis and open new avenues for clinical intervention.

https://doi.org/10.1016/j.bbrep.2025.102043

Received 28 December 2024; Received in revised form 16 April 2025; Accepted 6 May 2025

<sup>\*</sup> Corresponding author. Beijing Obstetrics and Gynecology Hospital, Capital Medical University. Beijing Maternal and Child Health Care Hospital, Beijing, 100026, China.

<sup>\*\*</sup> Corresponding author. Beijing Obstetrics and Gynecology Hospital, Capital Medical University. Beijing Maternal and Child Health Care Hospital, Beijing, 100026, China.

E-mail addresses: luanyingyi@mail.ccmu.edu.cn (Y.-y. Luan), yinchh@ccmu.edu.cn (C.-h. Yin).

<sup>2405-5808/© 2025</sup> Published by Elsevier B.V. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

#### Table 1

## Definition of cell death and its pathogenesis in PCOS.

Cell death	Year	Definition	Pathogenesis in PCOS	Reference
Apoptosis	1972	Apoptosis is currently best	The regulation of GCs' growth and	[3,50,51]
		identified by a	apoptosis during	
		morphological	development in the	
		changes in dying	ovarian follicles	
		cells.	relies heavily on the	
			of the PI3K/Akt	
			pathway, as well as	
			downstream pro-	
			as FOXO1, Bax,	
			caspase-9, and	
Autonhagy	1963	Autophagy is a	caspase-3. The abundance of	[6,61-64]
лиюрнаду		highly conserved	autophagy-related	.,
		eukarytotic cellular	genes ATG5, ATG7,	
		Through the	the ratio of	
		degradation of	autophagy marker	
		cytoplasmic	protein light chain 3B	
		proteins, and	significantly	
		macromolecules,	increased whereas	
		and the recycling of	the abundance of the	
		products,	SQSTM1/p62 was	
		autophagy plays	decreased in ovarian	
		important roles in cell survival and	granulosa cells from	
		maintenance.	r dob patients.	
Ferroptosis	2012	Ferroptosis is	The quantity and	[4,5,
		dependent upon intracellular iron.	location of polyunsaturated fatty	80-82]
		but not other	acids (PUFAs) in	
		metals, and is	PCOS determine the	
		biochemically,	peroxidation within	
		genetically distinct	cells, and to some	
		from apoptosis,	extent, determine	
		autophagy.	of iron metabolism in	
			regulating ferroptosis	
			has gained significant	
			studies investigating	
			ferroptosis-mediated	
Pyroptosis	2001	Cell pyroptosis is a	therapy for PCOS. TLR4, IL-6 and IL-18	[88
1 910 910 913	2001	programmed cell	levels in	104–106]
		death characterized	hyperandrogenism-	
		by a highly inflammatory	induced PCOS-like mice were highly	
		response.	increased, moreover,	
			pyroptosis factors	
			caspase-1, GSDMD.	
			IL-1 $\beta$ and IL-18 in the	
			ovary were activated	
NETosis	2004	NETosis is a form of	Research has shown	[109,120,
		regulated cell death	that women with	121]
		that involves the release of	PCOS have increased	
		neutrophil	and inflammatory	
		extracellular traps	cytokines, which are	
		by neutro	involved in the process of NFTosis	
			Some studies have	
			proposed that	
			NETosis may contribute to the	

Table 1 (continued)

Cell death
Necroptosis

## 2. Cell death and PCOS

### 2.1. Apoptosis and PCOS

Under steady-state conditions, caspase-dependent apoptosis accounts for approximately 90 % of cell turnover. Its homeostatic role is closely related to its non-inflammatory properties [25,26]. These pathways are activated by different types of death signals and exhibit distinct molecular mechanisms. The extrinsic pathway, also known as the death receptor pathway, is initiated by external signals binding to specific cell surface receptors [27-29]. In contrast, the mitochondrial pathway is more like the cell's "internal monitoring system ". When internal crises such as DNA damage occur, members of the pro-apoptotic protein Bcl-2 family break the balance with anti-apoptotic proteins [30,31]. This change causes mitochondria to release cytochrome c, which in turn activates the caspase cascade and completes programmed cell death. Sheena L.P. et al. identified three major regulatory mechanisms that orchestrate granulosa cell (GC) apoptosis: growth factors that increase cyclic AMP (cAMP) levels, which can promote apoptosis via granzyme B; mitochondrial pathways modulated by Bcl-2 family proteins, such as Bax and Bcl-2, that determine mitochondrial membrane integrity; and death receptor-mediated signaling triggered by TNFa and FasL-Fas interactions [32]. These pathways can be activated at multiple stages of follicular development, highlighting the central role of apoptosis in maintaining ovarian follicle homeostasis and regulating follicular atresia.

Given the importance of apoptosis in follicular dynamics, its dysregulation can have profound implications in reproductive disorders such as PCOS. Granulosa cells are essential somatic cells that support oocyte maturation within the ovarian follicles. Among women diagnosed with PCOS, an increased number of follicular cysts is often observed [33,34]. These cysts are thought to arise from impaired follicular maturation, potentially resulting from altered granulosa cell function [35,36]. Indeed, several studies suggest a reduction in GC apoptosis in PCOS, which may contribute to the persistence of immature follicles and the development of polycystic ovarian morphology [37-40]. Furthermore, oxidative stress (OS) has been identified as a key pathological factor exacerbating GC dysfunction in PCOS. OS results from an imbalance between reactive oxygen species (ROS) and antioxidant defenses. Elevated ROS levels in PCOS patients may be attributed to hormonal imbalances such as reduced estrogen and progesterone levels. ROS overproduction can lead to mitochondrial damage and trigger intrinsic apoptotic pathways [41–44]. The PI3K/Akt signaling pathway plays a crucial role in regulating granulosa cell growth and apoptosis during folliculogenesis. Downstream apoptotic regulators, including FOXO1, Bax, caspase-9, and caspase-3, are critically involved in this process [45-47]. Among them, Bcl-2 serves as a key anti-apoptotic molecule. It forms homodimers that stabilize the mitochondrial membrane potential (MMP); however, under apoptotic stress, heterodimerization with Bax reduces MMP, increases mitochondrial permeability, and promotes cytochrome c release, thereby activating caspases [48,49]. Studies have demonstrated altered expression of these key factors in PCOS patients, including elevated levels of pro-apoptotic FOXO1, Bax, caspase-9, and caspase-3, as well as decreased expression of PI3K, Akt, and Bcl-2 [50, 51]. Taken together, these findings underscore that both insufficient apoptosis and excessive apoptosis can disrupt follicular development. Apoptotic mechanisms, operating at all stages of folliculogenesis, are crucial for maintaining a balance between follicle survival and atresia. Disruption of these finely tuned processes may lead to follicular arrest, anovulation, and the characteristic polycystic morphology observed in PCOS.

#### 2.2. Autophagy and PCOS

Autophagy is an important process for cells to maintain homeostasis, and its initiation is like a carefully choreographed molecular ballet. When cells sense nutrient deprivation, the inhibitory effect of the mTOR pathway is released, and the ULK complex including ULK1/2, ATG13 and other proteins is activated [52,53]. This key "start switch" recruits the PI3K complex composed of Beclin 1 and VPS34 to generate PI3P phospholipid molecules on specific membrane structures, laying the foundation for subsequent steps [54]. As the process progresses, the cell will form a unique cup-shaped structure - the phagophore. The maturation of this structure depends on two major protein modification systems: ATG12-ATG5-ATG16L1 complex system and LC3 protein lipidation system. It is particularly worth mentioning that LC3-II is like a "molecular tag" of the autophagosome, which not only promotes the extension of the membrane structure, but also selectively identifies the substances to be degraded through receptor proteins such as P62 [55-57]. These receptors can specifically mark damaged organelles or misfolded proteins to ensure that they are accurately transported to the autophagosome. Finally, the mature autophagosome fuses with the lysosome to form an autophagolysosome. This critical step requires the precise coordination of multiple proteins such as syntaxins and Rab GTPases [58]. Under the action of lysosomal enzymes, the encapsulated "cellular garbage" is completely degraded, and the released nutrients re-participate in cell metabolism, helping cells cope with various stress environments.

Building upon this fundamental role, recent research has increasingly linked dysregulated autophagy to reproductive disorders, particularly PCOS. Studies have found that there is an abnormal increase in autophagic activity in the granulosa cells of PCOS patients. This may lead to excessive degradation of essential cellular components, thereby impairing cell viability and function [59,60]. By comparing and analyzing the ovarian granulosa cells of PCOS and non-PCOS patients, the researchers found that the expression profile of autophagy-related genes changed: ATG5, ATG7 and BECN1 mRNA levels were significantly increased, the autophagy marker protein LC3 II/I ratio increased, and the autophagy substrate SQSTM1/p62 protein level decreased [61]. Moreover, HMGB1 levels in serum and follicular fluid of PCOS patients with combined insulin resistance are significantly increased [62,63], suggesting that it may be involved in the development of insulin resistance and related inflammatory responses. In-depth studies have found that HMGB1 plays an important regulatory role in the autophagic activity of ovarian granulosa cells. Specifically, it increases the ratio of LC3B-II to LC3B-I, increases the expression level of ATG7, and reduces the content of SQSTM1 protein [64]. Animal model studies provide more in-depth insights into the pathogenesis of PCOS. Xing Y et al. found

that the PCOS rat model induced by testosterone propionate (TP) showed that the expression levels of Beclin-1 and LC3 were significantly increased. After intervention with Guizhi Yikun Formula (GZYKF), the increase of Beclin-1 induced by TP was significantly inhibited [65], which indicate that GZYKF may inhibit the autophagy process by activating the TP53-AMPK signaling pathway and upregulating mTOR expression. Thus, autophagy dysregulation may interfere with follicular development by disrupting the signaling of key hormones such as insulin and androgens. These changes lead to increased follicular atresia and ovulatory dysfunction. It is worth noting that OS, as an important feature of PCOS, forms a complex interaction network with autophagy regulation: the increased ROS level in PCOS patients is accompanied by a decrease in antioxidant capacity, and autophagy dysfunction leads to the obstruction of the clearance of ROS-producing organelles, thus forming a vicious cycle of oxidative stress damage, insulin resistance and hyperandrogenism [66,67]. In conclusion, the amassed evidence underscores the significance of autophagy in the disrupted physiology observed in PCOS conditions [Fig. 1].

# 2.3. Ferroptosis and PCOS

Research on ferroptosis has made significant progress since it was discovered in 2012. This unique cell death mode relies on iron-mediated lipid peroxidation, and its regulatory network involves multiple key metabolic links [68]. Iron plays a central role among the major drivers of ferroptosis. It can participate in Fenton reactions, generating ROS and initiating lipid peroxidation. Transferrin receptor 1 (TFRC) and divalent metal transporter 1 (DMT1) are involved in iron uptake, while ferroportin (FPN) exports iron out of the cell [69,70]. Once inside the cell, iron catalyzes the formation of lipid hydroperoxides from polyunsaturated fatty acids (PUFAs) in the cell membrane, leading to disruption of membrane integrity and eventual cell death [71]. Glutathione (GSH) is a key weapon for cells to fight oxidative damage, and it can effectively neutralize ROS. However, when GSH stocks are low, whether synthesis is reduced or consumption is increased, it will damage the ability of cells to resist lipid peroxidation, leading to ferroptosis [72]. Glutathione peroxidase 4 (GPX4) is an enzyme that utilizes GSH to detoxify lipid hydroperoxides. When GPX4 is inhibited, these lipid peroxides accumulate in the cell, eventually triggering ferroptosis [73]. A key upstream regulator of GSH synthesis is the xc-antiporter system, which transports cystine into the cell in exchange for glutamate. Once cystine enters the cell, it is reduced to cysteine, an important precursor for GSH biosynthesis. Dysfunction or inhibition of the xc-system limits the availability of cysteine, thereby hindering GSH production and making cells more sensitive to ferroptosis [74]. Beyond metabolic pathways, several redox signaling networks also influence ferroptosis susceptibility. For instance, the Nrf2-Keap1 pathway modulates the expression of antioxidant and iron-regulating genes. Similarly, the tumor suppressor protein p53 has emerged as a key player in ferroptosis regulation by altering the transcription of genes involved in iron metabolism and redox homeostasis [75,76].

The significance of ferroptosis extends beyond cell death, playing a pivotal role in disease contexts where OS is a contributing factor. This is particularly relevant in the female reproductive system, where maintaining a balance between ROS and antioxidants is crucial for normal physiological function. Disruption of this equilibrium either through excessive ROS generation or antioxidant depletion has been implicated in the pathogenesis of various obstetric and gynecological disorders [77, 78]. A notable example is PCOS, where nearly half of patients are overweight and exhibit elevated levels of circulating free fatty acids, including arachidonic acid—a PUFA that alters mitochondrial distribution and increases ROS generation [41]. These metabolic imbalances heighten oxidative stress, thereby promoting lipid peroxidation. Under normal physiological conditions, the processes of oxygen radical reactions and lipid peroxidation are dynamically regulated, maintaining homeostasis in cellular, biochemical, and immune functions [79].



### Fig. 1. Autophagy and PCOS

The levels of autophagy-related genes, such as ATG5, ATG7, and BECN1 mRNA, were significantly increased in granulosa cells of PCOS patients, as were the proportions of the autophagy marker protein light chain 3B II. The autophagosomes was formed with phagocytes and autophagy substrates.

However, excessive ROS can oxidize membrane-bound PUFAs and nucleic acids, producing lipid peroxidation byproducts such as malondialdehyde (MDA) and 4-hydroxynonenal (HNE). These reactive species compromise membrane fluidity and permeability, eventually altering cell structure and function [80–82]. The quantity and location of polyunsaturated fatty acids (PUFAs) in PCOS determine the extent of lipid



Fig. 2. Ferroptosis and PCOS

Total free fatty acid levels are elevated in PCOS patients, and the amount and location of polyunsaturated fatty acids (PUFA) determine the degree of intracellular lipid peroxidation and, to a certain extent, ferroptosis.

peroxidation within cells, and to some extent, determine ferroptosis. The role of iron metabolism in regulating ferroptosis has gained significant attention in recent studies investigating ferroptosis-mediated therapy for PCOS. A clinical study involving 149 women with PCOS and 108 healthy controls demonstrated significantly higher serum ferritin levels in the PCOS group, independent of obesity status, indicating the presence of systemic iron overload in these patients. Ovulatory dysfunction, a hallmark of PCOS, often leads to delayed menstruation, light menstrual flow, or even amenorrhea. Since regular menstruation is one of the primary mechanisms for iron loss in women of reproductive age, disruptions in menstrual patterns may contribute to iron retention. Indeed, studies have shown a correlation between elevated serum ferritin levels and the severity of menstrual irregularities in PCOS, suggesting that impaired menstrual blood loss may be associated with increased iron stores [83,84]. Moreover, the compensatory hyperinsulinemia seen in insulin-resistant PCOS patients may enhance tissue iron uptake and inhibit iron release from macrophages [85], further contributing to systemic iron overload. Iron overload can further disrupt glucose metabolism, exacerbate metabolic disorders, and worsen insulin resistance or hyperinsulinemia. These effects enhance the sensitivity of ovarian cells to luteinizing hormone (LH), disrupt the normal ovulation process, and may affect embryo implantation [86,87]. These interrelated mechanisms not only reveal the potential role of ferroptosis in the pathogenesis of PCOS, but also provide new ideas for therapeutic intervention [Fig. 2].

# 2.4. Pyroptosis and PCOS

Pyroptosis is a form of RCD driven by the activation of inflammasome, which is morphologically distinct from apoptosis. It has attracted much attention due to its association with innate immunity and disease [88–91]. In response to infection, cellular stress, or activation by specific signaling pathways, inflammasomes are assembled within the cytoplasm. These complexes typically consist of sensor proteins such as NLRP3, adaptor proteins, and the effector enzyme caspase-1 [92-94]. Once the inflammasome is formed, caspase-1 becomes activated and begins cleaving several target proteins, including the inactive precursors of pro-inflammatory cytokines interleukin-1 $\beta$  (IL-1 $\beta$ ) and interleukin-18 (IL-18). Caspase-1 processes these precursors into their active forms, which are then released from the dying cell. These cytokines, in turn, initiate a strong inflammatory response and attract immune cells to the site of cell death [95,96]. Concomitantly, caspase-1 also cleaves gasdermin D (GSDMD), releasing its N-terminal domain, which oligomerizes to form pores in the plasma membrane. This pore formation causes rapid cell swelling, membrane rupture, and the release of intracellular contents into the extracellular space—hallmarks of pyroptosis that further amplify inflammation and signal immune cell recruitment [97]. In conclusion, cell pyroptosis is integral to host immunity against microbial infections, as it helps to eliminate infected cells and initiate an inflammatory response [98-100]. However, excessive or dysregulated pyroptosis can contribute to various inflammatory diseases, including sepsis, inflammatory bowel disease, and neurodegenerative disorders [101,102].

Recent research has highlighted the relevance of pyroptosis in reproductive disorders, particularly in PCOS. Persistent hyperandrogenemia of PCOS promotes local inflammatory activation of the ovary, and the imbalanced inflammatory microenvironment leads to pyroptosis of GCs, further including a series of pathologies including follicular dysfunction and ovarian interstitial cell fibrosis [103]. These findings establish a potential mechanistic link between chronic inflammation and reproductive impairment in PCOS. Further supporting this connection, numerous studies have demonstrated elevated levels of inflammatory markers and mediators-such as CRP, TNF- $\alpha$ , IL-6, IL-8 and IL-18 in patients with PCOS. These cytokines are key mediators of the inflammatory response, and their elevated levels further support the involvement of systemic inflammation in the pathogenesis of PCOS

[104]. Under normal physiological conditions, tissue remodeling and controlled inflammation are essential for ovarian folliculogenesis. However, excessive or dysregulated inflammation can disrupt normal follicular development [105]. Notably, previous studies have observed significantly elevated expression of pyroptosis-related genes and proteins-such as caspase-1, NLRP3, and the cleaved form of gasdermin D (GSDMD-N)-in granulosa cells from PCOS patients. These molecular signatures confirm the activation of the NLRP3 inflammasome in PCOS and its central role in triggering pyroptotic cell death. Once activated, the NLRP3 inflammasome facilitates cleavage of pro-caspase-1 into its active form, leading to the maturation and release of IL-1 $\beta$  and IL-18, as well as the cleavage of GSDMD and subsequent pore formation in the cell membrane [106]. Although NLRP3 serves physiological roles in normal follicular development, its chronic or excessive activation has been implicated in the development of polycystic ovaries and the deterioration of ovarian function [107]. For instance, a study by Cai et al. reported significantly elevated levels of GSDMD and caspase-1 in the ovaries of PCOS model mice, along with evident pyroptosis of granulosa cells cultured in vitro—a process initiated by caspase-1 activation [108]. Building upon this, Wang et al. were the first to demonstrate that pyroptosis mediated via the NLRP3/GSDMD/caspase-1 axis leads to substantial granulosa cell loss in PCOS [103]. This finding underscores the pathological importance of inflammasome-induced pyroptosis in the depletion of functional ovarian cells. Importantly, inflammasome activation and the ensuing pyroptotic cascade not only damage local ovarian tissue but also amplify systemic inflammation. When sustained over time, this creates a vicious cycle: inflammation promotes pyroptosis, which further exacerbates the inflammatory response, ultimately impairing ovarian function and accelerating the progression of PCOS. Therefore, targeting the pyroptosis pathway may hold therapeutic potential for breaking this cycle and restoring reproductive health in affected individuals.

# 2.5. NETosis and PCOS

NETosis constitutes a regulated cell death mechanism noted for the liberation of neutrophil extracellular traps (NETs)by neutrophils, which are a type of white blood cell [109]. Neutrophils serve as integral elements of the immune system, pivotal in safeguarding the body against microbial infections. As a defense mechanism, NETosis is initiated when neutrophils encounter certain stimuli, such as pathogens, inflammatory signals, or immune complexes. When stimulated by external factors, neutrophils are activated and undergo a series of significant morphological changes: first, the DNA in the nucleus depolymerizes, and the nuclear membrane disintegrates, causing the nuclear contents to mix with the cytoplasm; then, these depolymerized chromatin binds to a variety of antimicrobial proteins such as histones, neutrophil elastase, and myeloperoxidase, eventually forming a mesh of neutrophil extracellular traps (NETs). After these special mesh structures are released outside the cells, they can not only serve as a physical barrier to capture pathogenic microorganisms, but also directly kill pathogens through a variety of antimicrobial proteins attached to their surface, showing a unique dual defense mechanism [110-113]. Therefore, NETs have a dual role: they can both remove pathogens and regulate inflammatory responses. However, NETosis is actually a double-edged sword although NETs play a protective role in anti-infection, excessive or uncontrolled NETs formation may cause tissue damage and aggravate the inflammatory response [114,115].

NETs released during NETosis are known to have both beneficial and detrimental effects. This dual nature of NETosis has drawn attention in the context of chronic inflammatory diseases. Studies have observed increased NETosis and NET formation in various inflammatory conditions, including autoimmune disorders [116,117]. One such condition potentially influenced by NETosis is PCOS, which is linked to chronic low-grade inflammation, marked by heightened concentrations of inflammatory markers circulating in the blood. Indeed, inflammation

contributes significantly to the onset and advancement of PCOS along with its related symptoms [118,119]. Emerging evidence suggests that NETosis may be involved in the underlying inflammatory processes of PCOS. Studies have indicated elevated concentrations of neutrophils and inflammatory cytokines in women with PCOS, implicating their involvement in the process of NETosis. Some studies have proposed that NETosis may contribute to the inflammatory environment observed in PCOS. Specifically, the heightened NETosis observed in women with PCOS is speculated to correlate with activated neutrophils and the consequent release of NETs. This process potentially contributes to inflammation and insulin resistance, mutual traits of PCOS [120,121]. Nevertheless, it is crucial to emphasize that the connection between PCOS and NETosis remains an active area of investigation. The precise mechanisms and direct relationships are not yet fully comprehended. Thus, ongoing studies are essential to clarify the exact role of NETosis in the advancement of PCOS and its potential clinical implications.

# 2.6. Necroptosis and PCOS

Necroptosis is a regulated form of necrotic cell death that serves as an alternative to apoptosis, particularly when apoptotic pathways are inhibited. Unlike apoptosis, which is typically immunologically silent, necroptosis is pro-inflammatory, as it disrupts the integrity of the plasma membrane and leads to the release of damage-associated molecular patterns (DAMPs) that activate the innate immune system [9]. Mechanistically, this form of cell death is primarily mediated through a signaling cascade initiated by tumor necrosis factor receptor 1 (TNFR1) or other death receptors such as TLR3 and TLR4 under specific conditions. Upon binding of TNF- $\alpha$  to TNFR1, a multiprotein signaling complex-Complex I-is formed, comprising TRADD, TRAF2/5, RIPK1, cIAP1/2, and LUBAC. Under normal circumstances, this pathway may lead to apoptosis; however, if apoptosis is inhibited, often due to caspase-8 suppression, RIPK1 interacts with RIPK3 to form a functional unit known as the necrosome. Within this complex, RIPK3 phosphorylates MLKL (mixed lineage kinase domain-like protein), which then oligomerizes and translocates to the plasma membrane. There, MLKL forms disruptive pores, leading to loss of cellular homeostasis, cell lysis, and inflammatory cell death [122-124]. Beyond its role in host defense-where necroptosis eliminates pathogen-infected cells that evade apoptosis-this mechanism has broader biological significance. In particular, it contributes to inflammatory processes by releasing DAMPs and amplifying immune responses. Interestingly, recent studies have highlighted the importance of necroptosis in reproductive biology, especially in regulating follicular cell death, which is essential for proper follicle development, growth, and ovulation.

In this reproductive context, necroptosis has emerged as a potential pathological mechanism in PCOS. Increasing evidence indicates that GC necroptosis may play a pivotal role in the onset and progression of PCOS [125]. One key factor contributing to this process is oxidative stress, a known driver of GC apoptosis and follicular atresia. Sustained overproduction of reactive oxygen species (ROS) activates inflammatory signaling pathways, which, in turn, promote necroptosis in granulosa cells, potentially disrupting the follicular microenvironment. As necroptotic GCs perish, they deprive adjacent oocytes of vital nutrients, growth factors, and survival signals, rendering the oocytes more susceptible to further damage or even follicular system-mediated necroptosis [126]. As a result, the simultaneous necroptosis of GCs and oocytes contributes to follicular atresia, a hallmark of PCOS-related ovarian dysfunction. Moreover, elevated levels of ROS and pro-inflammatory cytokines have been shown to upregulate RIPK1 and RIPK3 expression in GCs, further amplifying necroptotic signaling pathways [127]. This suggests the presence of a self-reinforcing loop between oxidative stress, inflammation, and necroptosis, which may underlie the persistent follicular dysregulation observed in PCOS. Notably, a study by Wang et al. identified several genes-IL33, BCL2, PYGM, and TNFSF10-as potential diagnostic markers associated with

necroptosis in PCOS, highlighting the potential for targeting this pathway in future therapeutic strategies [125].

## 3. Conclusions

The key takeaway from these studies is the substantial contribution of regulatory forms of cell death to the development of PCOS. In PCOS, ovarian granulosa cell death is a central event in the pathogenesis. In PCOS, apoptotic and autophagic cell death programs appear to be particularly significant. These studies highlight their prevalence and suggest potential therapeutic avenues by targeting these specific cell death pathways. The latest data are encouraging a reevaluation of the significance of apoptosis and ferroptosis in PCOS, prompting a reconsideration of their respective roles in contributing to the syndrome [37-40,59-61]. Several studies have shown aletrations in apoptotic pathways in the ovaries of women with PCOS. These changes can affect follicular development, ovulation, and the overall balance of hormone production. Specifically, increased apoptosis has been observed in the granulosa cells, which surround and support the developing egg within the ovarian follicles. Granulosa cell apoptosis disrupts the normal development of follicles and promotes the formation of ovarian cysts in PCOS patients [128]. Studies have also found that women with PCOS often have abnormal lipid metabolism, manifested as disorders in specific lipid and fatty acid levels [129,130]. These lipid abnormalities may lead to lipid peroxidation, which may promote ferroptosis [5]. In addition, iron dysregulation has been observed in PCOS, which can increase the susceptibility to ferroptosis [87]. The above data suggest that preferentially targeting death receptors and apoptosis in the mitochondrial pathway is expected to be key to treating PCOS. However, considering the potential benefits, inhibition of ferroptosis may also be a promising complementary approach to treat this syndrome. In summary, in-depth study of the molecular complexity and physiological impact of apoptosis, autophagy, ferroptosis, pyroptosis, and NETosis is essential to understand their roles in PCOS. Designing targeted drug therapies against these specific cell death and immune processes is expected to more effectively treat this syndrome.

# 4. Prospect

Emerging evidence suggests that inflammation, oxidative stress, and regulated cell death pathways are intricately linked to the pathophysiology of PCOS, beyond the classic features of androgen excess and insulin resistance. Women with PCOS have persistent chronic low-grade inflammation, characterized by elevated levels of cytokines such as TNF- $\alpha$ , IL-6, and CRP, and are closely associated with metabolic and reproductive dysfunction [131,132]. At the same time, enhanced oxidative stress, manifested by increased ROS production and impaired antioxidant defenses, further amplifies inflammatory signaling and contributes to ovarian dysfunction, follicular atresia, and insulin resistance [133, 134].

It is worth noting that this oxidative stress-inflammatory microenvironment in PCOS patients may have an important impact on the activation and cross-regulation of multiple programmed cell death modes, including apoptosis, ferroptosis and pyroptosis. For instance, excessive ROS and lipid peroxidation may drive ferroptosis in granulosa cells, impairing follicular development [135], while inflammasome activation may trigger pyroptosis, exacerbating local ovarian inflammation [136]. Dysregulated apoptosis of granulosa or theca cells may also contribute to anovulation and the formation of cystic follicles [40]. Moreover, emerging studies suggest that metabolic stress and hyperandrogenemia may modulate the sensitivity of ovarian cells to different death modalities, further linking systemic metabolic disturbances with cellular homeostasis [137].

A deeper understanding of how oxidative stress and inflammation converge to regulate cell death pathways in PCOS may open new therapeutic avenues. Targeting this cross-talk—through antioxidants, antiinflammatory agents, or modulators of cell death—holds promise for restoring ovarian function and improving systemic metabolic outcomes. Future research should focus on delineating the molecular mechanisms underlying this interplay and identifying cell-type-specific vulnerabilities within the ovary and peripheral tissues.

#### CRediT authorship contribution statement

Ying-ying Li: Writing – original draft. Yi-qiu Peng: Investigation. Yu-xi Yang: Investigation. Ning Xu: Investigation. Ting-juan Shi: Investigation. Rui-xia Liu: Writing – review & editing. Ying-yi Luan: Writing – review & editing. Cheng-hong Yin: Writing – review & editing.

#### Ethics approval and consent to participate

Not applicable.

## **Consent for publication**

Not applicable.

## Funding

The research conducted for this study received support from the National Key Research and Development Program of China (Nos. 2016YFC1000100).

#### Declaration of competing interest

The authors declare that they have no conflict of interest.

#### Acknowledgements

Not applicable.

## Appendix A. Supplementary data

Supplementary data to this article can be found online at https://doi.org/10.1016/j.bbrep.2025.102043.

## Data availability

No data was used for the research described in the article.

#### References

- L. Galluzzi, J.M. Bravo-San Pedro, I. Vitale, et al., Essential versus accessory aspects of cell death: recommendations of the NCCD 2015, Cell Death Differ. 22 (1) (2015) 58–73, https://doi.org/10.1038/cdd.2014.137.
- [2] L. Galluzzi, I. Vitale, S.A. Aaronson, et al., Molecular mechanisms of cell death: recommendations of the nomenclature committee on cell death 2018, Cell Death Differ. 25 (3) (2018) 486–541, https://doi.org/10.1038/s41418-017-0012-4.
- [3] D. Hockenbery, Defining apoptosis, Am. J. Pathol. 146 (1) (1995) 16–19. PMID: 7856725.
- [4] S.J. Dixon, K.M. Lemberg, M.R. Lamprecht, et al., Ferroptosis: an iron-dependent form of nonapoptotic cell death, Cell 149 (5) (2012) 1060–1072, https://doi.org/ 10.1016/j.cell.2012.03.042.
- [5] B.R. Stockwell, J.P. Friedmann Angeli, H. Bayir, et al., Ferroptosis: a regulated cell death nexus linking metabolism, redox biology, and disease, Cell 171 (2) (2017) 273–285, https://doi.org/10.1016/j.cell.2017.09.021.
- [6] N. Mizushima, M. Komatsu, Autophagy: renovation of cells and tissues, Cell 147 (4) (2011) 728–741, https://doi.org/10.1016/j.cell.2011.10.026.
- [7] P. Yu, X. Zhang, N. Liu, L. Tang, C. Peng, X. Chen, Pyroptosis: mechanisms and diseases, Signal Transduct. Targeted Ther. 6 (1) (2021) 128, https://doi.org/ 10.1038/s41392-021-00507-5. Published 2021 Mar 29.
- [8] M. Inoue, M. Enomoto, M. Yoshimura, T. Mizowaki, Pharmacological inhibition of sodium-calcium exchange activates NADPH oxidase and induces infectionindependent NETotic cell death, Redox Biol. 43 (2021) 101983, https://doi.org/ 10.1016/j.redox.2021.101983.
- [9] A. Degterev, Z. Huang, M. Boyce, et al., Chemical inhibitor of nonapoptotic cell death with therapeutic potential for ischemic brain injury [published correction

appears in Nat Chem Biol. 2005 Sep;1(4):234], Nat. Chem. Biol. 1 (2) (2005) 112–119, https://doi.org/10.1038/nchembio711.

- [10] T. Dragovich, C.M. Rudin, C.B. Thompson, Signal transduction pathways that regulate cell survival and cell death, Oncogene 17 (25) (1998) 3207–3213, https://doi.org/10.1038/sj.onc.1202587.
- [11] D. Denton, S. Kumar, Autophagy-dependent cell death, Cell Death Differ. 26 (4) (2019) 605–616, https://doi.org/10.1038/s41418-018-0252-y.
- [12] A.Z. Spitz, E. Gavathiotis, Physiological and pharmacological modulation of BAX, Trends Pharmacol. Sci. 43 (3) (2022) 206–220, https://doi.org/10.1016/j. tips.2021.11.001.
- [13] J. Kale, E.J. Osterlund, D.W. Andrews, BCL-2 family proteins: changing partners in the dance towards death, Cell Death Differ. 25 (1) (2018) 65–80, https://doi. org/10.1038/cdd.2017.186.
- [14] M. Raudenská, J. Balvan, M. Masařík, Cell death in head and neck cancer pathogenesis and treatment, Cell Death Dis. 12 (2) (2021) 192, https://doi.org/ 10.1038/s41419-021-03474-5. Published 2021 Feb 18.
- [15] X.X. Fan, H.D. Pan, Y. Li, R.J. Guo, E.L. Leung, L. Liu, Novel therapeutic strategy for cancer and autoimmune conditions: modulating cell metabolism and redox capacity, Pharmacol. Ther. 191 (2018) 148–161, https://doi.org/10.1016/j. pharmthera.2018.06.010.
- [16] R.J. Norman, D. Dewailly, R.S. Legro, T.E. Hickey, Polycystic ovary syndrome, Lancet 370 (9588) (2007) 685–697, https://doi.org/10.1016/S0140-6736(07) 61345-2.
- [17] R. Azziz, E. Carmina, Z. Chen, et al., Polycystic ovary syndrome, Nat. Rev. Dis. Primers 2 (2016) 16057, https://doi.org/10.1038/nrdp.2016.57. Published 2016 Aug 11.
- [18] D.A. Dumesic, S.E. Oberfield, E. Stener-Victorin, J.C. Marshall, J.S. Laven, R. S. Legro, Scientific statement on the diagnostic criteria, epidemiology, pathophysiology, and molecular genetics of polycystic ovary syndrome, Endocr. Rev. 36 (5) (2015) 487–525, https://doi.org/10.1210/er.2015-1018.
- [19] J.R. Givens, Familial polycystic ovarian disease, Endocrinol Metab. Clin. N. Am. 17 (4) (1988) 771–783. PMID: 3058473.
- [20] J.M. Vink, S. Sadrzadeh, C.B. Lambalk, D.I. Boomsma, Heritability of polycystic ovary syndrome in a Dutch twin-family study, J. Clin. Endocrinol. Metab. 91 (2006) 2100–2104, https://doi.org/10.1210/jc.2005-1494.
- [21] V. Padmanabhan, A. Veiga-Lopez, Developmental origin of reproductive and metabolic dysfunctions: androgenic versus estrogenic reprogramming, Semin. Reprod. Med. 29 (2011) 173–186, https://doi.org/10.1055/s-0031-1275519.
- [22] D.A. Dumesic, M.O. Goodarzi, G.D. Chazenbalk, D.H. Abbott, Intrauterine environment and polycystic ovary syndrome, Semin. Reprod. Med. 32 (2014) 159–165, https://doi.org/10.1055/s-0034-1371087.
- [23] I. Ek, P. Arner, M. Rydén, et al., A unique defect in the regulation of visceral fat cell lipolysis in the polycystic ovary syndrome as an early link to insulin resistance, Diabetes 51 (2002) 484–492, https://doi.org/10.2337/ diabetes.51.2.484.
- [24] V.T. Samuel, K.F. Petersen, G.I. Shulman, Lipid-induced insulin resistance: unravelling the mechanism, Lancet 375 (2010) 2267–2277, https://doi.org/ 10.1016/S0140-6736(10)60408-4.
- [25] J.F. Kerr, A.H. Wyllie, A.R. Currie, Apoptosis: a basic biological phenomenon with wide-ranging implications in tissue kinetics, Br. J. Cancer 26 (4) (1972) 239–257, https://doi.org/10.1038/bjc.1972.33.
- [26] N. Maghsoudi, Z. Zakeri, R.A. Lockshin, Programmed cell death and apoptosiswhere it came from and where it is going: from Elie Metchnikoff to the control of caspases, Exp. Oncol. 34 (3) (2012) 146–152. PMID: 2306998.
- [27] M.E. Guicciardi, Gregory J. Gores, Life and death by death receptors, FASEB J. 23 (2009) 1625–1637, https://doi.org/10.1096/fj.08-111005.
- [28] S. Fulda, K.M. Debatin, Death receptor signaling in cancer therapy, Curr. Med. Chem. Anticancer Agents 3 (2003) 253–262, https://doi.org/10.2174/ 1568011033482404.
- [29] D.R. Green, G. Kroemer, The pathophysiology of mitochondrial cell death, Sci. Technol. Humanit. 305 (2004) 626–629, https://doi.org/10.1126/ science.1099320.
- [30] G. Kroemer, L. Galluzzi, C. Brenner, Mitochondrial membrane permeabilisation in cell death, Physiol. Rev. 87 (2007) 99–163, https://doi.org/10.1152/ physrev.00013.2006.
- [31] N.N. Danial, S.J. Korsmeyer, Cell death: critical control points, Cell 116 (2) (2004) 205–219, https://doi.org/10.1016/s0092-8674(04)00046-7.
- [32] S.L.P. Regan, P.G. Knight, J.L. Yovich, Y. Leung, F. Arfuso, A. Dharmarajan, Granulosa cell apoptosis in the ovarian follicle-A changing view, Front. Endocrinol. 9 (2018) 61, https://doi.org/10.3389/fendo.2018.00061. Published 2018 Mar 2.
- [33] D. Dewailly, G. Robin, M. Peigne, C. Decanter, P. Pigny, S. Catteau-Jonard, Interactions between androgens, FSH, anti-Müllerian hormone and estradiol during folliculogenesis in the human normal and polycystic ovary, Hum. Reprod. Update 22 (6) (2016) 709–724, https://doi.org/10.1093/humupd/dmw027.
- [34] D. Dewailly, C.Y. Andersen, A. Balen, et al., The physiology and clinical utility of anti-Mullerian hormone in women [published correction appears in Hum Reprod Update. 2014 Sep-Oct;20(5):804], Hum. Reprod. Update 20 (3) (2014) 370–385, https://doi.org/10.1093/humupd/dmt062.
- [35] W. Niu, A.C. Spradling, Two distinct pathways of pregranulosa cell differentiation support follicle formation in the mouse ovary, Proc. Natl. Acad. Sci. U. S. A 117 (33) (2020) 20015–20026, https://doi.org/10.1073/pnas.2005570117.
- [36] A.L. Johnson, Ovarian follicle selection and granulosa cell differentiation, Poult. Sci. 94 (4) (2015) 781–785, https://doi.org/10.3382/ps/peu008.
- [37] T. Wang, J. Zhang, M. Hu, et al., Differential expression patterns of glycolytic enzymes and mitochondria-dependent apoptosis in PCOS patients with

endometrial hyperplasia, an early hallmark of endometrial cancer, in vivo and the impact of metformin in vitro, Int. J. Biol. Sci. 15 (3) (2019) 714–725, https://doi. org/10.7150/ijbs.31425. Published 2019 Jan 24.

- [38] G. Liu, S. Liu, G. Xing, F. Wang, IncRNA PVT1/MicroRNA-17-5p/PTEN Axis regulates secretion of E2 and P4, proliferation, and apoptosis of ovarian granulosa cells in PCOS [retracted in: mol ther nucleic acids. 2022 may 10;28:593], Mol. Ther. Nucleic Acids 20 (2020) 205–216, https://doi.org/10.1016/j. omtn.2020.02.007.
- [39] Q. Zheng, Y. Li, D. Zhang, et al., ANP promotes proliferation and inhibits apoptosis of ovarian granulosa cells by NPRA/PGRMC1/EGFR complex and improves ovary functions of PCOS rats, Cell Death Dis. 8 (10) (2017) e3145, https://doi.org/10.1038/cddis.2017.494. Published 2017 Oct 26.
- [40] W. Tan, F. Dai, D. Yang, et al., MiR-93-5p promotes granulosa cell apoptosis and ferroptosis by the NF-kB signaling pathway in polycystic ovary syndrome, Front. Immunol. 13 (2022) 967151, https://doi.org/10.3389/fimmu.2022.967151. Published 2022 Oct 19.
- [41] Y. Ma, L. Zheng, Y. Wang, Y. Gao, Y. Xu, Arachidonic acid in follicular fluid of PCOS induces oxidative stress in a human ovarian granulosa tumor cell line (KGN) and upregulates GDF15 expression as a response [published correction appears in front endocrinol (lausanne). 2022 oct 04;13:988767], Front. Endocrinol. 13 (2022) 865748, https://doi.org/10.3389/fendo.2022.865748. Published 2022 May 11.
- [42] M. Murri, M. Luque-Ramírez, M. Insenser, M. Ojeda-Ojeda, H.F. Escobar-Morreale, Circulating markers of oxidative stress and polycystic ovary syndrome (PCOS): a systematic review and meta-analysis, Hum. Reprod. Update 19 (3) (2013) 268–288, https://doi.org/10.1093/humupd/dms059.
- [43] Q. Lai, W. Xiang, Q. Li, et al., Oxidative stress in granulosa cells contributes to poor oocyte quality and IVF-ET outcomes in women with polycystic ovary syndrome, Front. Med. 12 (5) (2018) 518–524, https://doi.org/10.1007/s11684-017-0575-y.
- [44] J. Qiao, H.L. Feng, Extra- and intra-ovarian factors in polycystic ovary syndrome: impact on oocyte maturation and embryo developmental competence, Hum. Reprod. Update 17 (2011) 17–33, https://doi.org/10.1093/humupd/dmq032.
- [45] C.L. Hu, R.G. Cowan, R.M. Harman, S.M. Quirk, Cell cycle progression and activation of Akt kinase are required for insulin-like growth factor I-mediated suppression of apoptosis in granulosa cells, Mol. Endocrinol. 18 (2004) 326–338, https://doi.org/10.1210/me.2003-0178.
- [46] J.A. Baur, D.A. Sinclair, Therapeutic potential of resveratrol: the in vivo evidence, Nat. Rev. Drug Discov. 5 (2006) 493–506, https://doi.org/10.1038/nrd2060.
- [47] G.B. John, M.J. Shidler, P. Besmer, D.H. Castrillon, Kit signaling via PI3K promotes ovarian follicle maturation but is dispensable for primordial follicle activation, Dev. Biol. 331 (2009) 292–299, https://doi.org/10.1016/j. ydbio.2009.05.546.
- [48] W. Zheng, G. Nagaraju, Z. Liu, K. Liu, Functional roles of the phosphatidylinositol 3-kinases (PI3Ks) signaling in the mammalian ovary, Mol. Cell. Endocrinol. 356 (2012) 24–30, https://doi.org/10.1016/j.mce.2011.05.027.
- [49] T. Li, H. Mo, W. Chen, L. Li, Y. Xiao, J. Zhang, et al., Role of the PI3K-Akt signaling pathway in the pathogenesis of polycystic ovary syndrome, Reprod. Sci. 24 (2017) 646–655, https://doi.org/10.1177/1933719116667606.
- [50] Y. Gong, S. Luo, P. Fan, H. Zhu, Y. Li, W. Huang, Growth hormone activates PI3K/ Akt signaling and inhibits ROS accumulation and apoptosis in granulosa cells of patients with polycystic ovary syndrome, Reprod. Biol. Endocrinol. 18 (1) (2020) 121, https://doi.org/10.1186/s12958-020-00677-x. Published 2020 Dec 7.
- [51] Y. Gao, J. Chen, R. Ji, J. Ding, Y. Zhang, J. Yang, USP25 regulates the proliferation and apoptosis of ovarian granulosa cells in polycystic ovary syndrome by modulating the PI3K/AKT pathway via deubiquitinating PTEN, Front. Cell Dev. Biol. 9 (2021) 779718, https://doi.org/10.3389/ fcell.2021.779718, Published 2021 Nov 4.
- [52] N. Mizushima, The role of the Atg1/ULK1 complex in autophagy regulation, Curr. Opin. Cell Biol. 22 (2010) 132–139, https://doi.org/10.1016/j.ceb.2009.12.004.
- [53] D. Papinski, C. Kraft, Regulation of autophagy by signaling through the Atg1/ ULK1 complex, J. Mol. Biol. 428 (9 Pt A) (2016) 1725–1741.
- [54] R.C. Russell, Y. Tian, H. Yuan, H.W. Park, Y.Y. Chang, J. Kim, et al., ULK1 induces autophagy by phosphorylating Beclin-1 and activating VPS34 lipid kinase, Nat. Cell Biol. 15 (2013) 741–750, https://doi.org/10.1038/ncb2757.
- [55] J. Geng, D.J. Klionsky, The Atg 8 and Atg 12 ubiquitin-like conjugation systems in macroautophagy. 'Protein modifications: beyond the usual suspects' review series, EMBO Rep. 9 (2008) 859–864, https://doi.org/10.1038/embor.2008.163.
- [56] N. Mizushima, T. Noda, T. Yoshimori, Y. Tanaka, T. Ishii, M.D. George, et al., A protein conjugation system essential for autophagy, Nature 395 (1998) 395–398, https://doi.org/10.1038/26506.
- [57] Y. Ohsumi, Molecular dissection of autophagy: two ubiquitin-like systems, Nat. Rev. Mol. Cell Biol. 2 (2001) 211–216, https://doi.org/10.1038/35056522.
- [58] G. Kroemer, G. Marino, B. Levine, Autophagy and the integrated stress response, Mol. Cell 40 (2010) 280–293, https://doi.org/10.1016/j.molcel.2010.09.023.
- [59] J. Zhou, X. Peng, S. Mei, Autophagy in ovarian follicular development and Atresia, Int. J. Biol. Sci. 15 (4) (2019) 726–737, https://doi.org/10.7150/ ijbs.30369.
- [60] J.Y. Choi, M.W. Jo, E.Y. Lee, et al., The role of autophagy in follicular development and atresia in rat granulosa cells, Fertil. Steril. 93 (8) (2010) 2532–2537, https://doi.org/10.1016/j.fertnstert.2009.11.021.
- [61] X. Li, J. Qi, Q. Zhu, et al., The role of androgen in autophagy of granulosa cells from PCOS, Gynecol. Endocrinol. 35 (8) (2019) 669–672, https://doi.org/ 10.1080/09513590.2018.1540567.
- [62] F. Cirillo, C. Catellani, P. Lazzeroni, et al., HMGB1 is increased in adolescents with polycystic ovary syndrome (PCOS) and decreases after treatment with myo-

inositol (MYO) in combination with alpha-lipoic acid (ALA), Gynecol. Endocrinol. 36 (7) (2020) 588–593, https://doi.org/10.1080/09513590.2020.1725967.

- [63] F. Cirillo, C. Catellani, C. Sartori, et al., CFTR and FOXO1 gene expression are reduced and high mobility group box 1 (HMGB1) is increased in the ovaries and serum of women with polycystic ovarian syndrome, Gynecol. Endocrinol. 35 (10) (2019) 842–846, https://doi.org/10.1080/09513590.2019.1599349.
- [64] C. Zhang, J. Hu, W. Wang, et al., HMGB1-induced aberrant autophagy contributes to insulin resistance in granulosa cells in PCOS, FASEB J 34 (7) (2020) 9563–9574, https://doi.org/10.1096/fj.202000605RR.
- [65] Y. Xing, Y.X. Liu, X. Liu, et al., Effects of Gui Zhu Yi Kun formula on the P53/ AMPK pathway of autophagy in granulosa cells of rats with polycystic ovary syndrome, Exp. Ther. Med. 13 (6) (2017) 3567–3573, https://doi.org/10.3892/ etm.2017.4384.
- [66] D. Cheng, B. Zheng, Y. Sheng, Z. Zeng, Z. Mo, The roles of autophagy in the genesis and development of polycystic ovary syndrome, Reprod. Sci. 30 (10) (2023) 2920–2931, https://doi.org/10.1007/s43032-023-01255-3.
- [67] A.K. Yadav, P.K. Yadav, G.R. Chaudhary, et al., Autophagy in hypoxic ovary, Cell. Mol. Life Sci. 76 (17) (2019) 3311–3322, https://doi.org/10.1007/s00018-019-03122-4.
- [68] X. Jiang, B.R. Stockwell, M. Conrad, Ferroptosis: mechanisms, biology and role in disease, Nat. Rev. Mol. Cell Biol. 22 (4) (2021) 266–282, https://doi.org/ 10.1038/s41580-020-00324-8.
- [69] M. Conrad, D.A. Pratt, The chemical basis of ferroptosis, Nat. Chem. Biol. 15 (2019) 1137–1147, https://doi.org/10.1038/s41589-019-0408-1.
- [70] M. Gao, P. Monian, N. Quadri, R. Ramasamy, X. Jiang, Glutaminolysis and transferrin regulate ferroptosis, Mol. Cell 59 (2015) 298–308, https://doi.org/ 10.1016/j.molcel.2015.06.011.
- [71] S. Doll, et al., ACSL4 dictates ferroptosis sensitivity by shaping cellular lipid composition, Nat. Chem. Biol. 13 (2017) 91–98, https://doi.org/10.1038/ nchembio.2239.
- [72] F. Ursini, M. Maiorino, M. Valente, L. Ferri, C. Gregolin, Purification from pig liver of a protein which protects liposomes and biomembranes from peroxidative degradation and exhibits glutathione peroxidase activity on phosphatidylcholine hydroperoxides, Biochim. Biophys. Acta 710 (1982) 197–211, https://doi.org/ 10.1016/0005-2760(82)90150-3.
- [73] W.S. Yang, et al., Regulation of ferroptotic cancer cell death by GPX4, Cell 156 (2014) 317–331, https://doi.org/10.1016/j.cell.2013.12.010.
- [74] S.J. Dixon, et al., Pharmacological inhibition of cystine-glutamate exchange induces endoplasmic reticulum stress and ferroptosis, eLife 3 (2014), https://doi. org/10.7554/eLife.02523.
- [75] L. Jiang, et al., Ferroptosis as a p53-mediated activity during tumour suppression, Nature 520 (2015) 57–62, https://doi.org/10.1038/nature14344.
- [76] S.J. Wang, et al., Acetylation is crucial for p53-mediated ferroptosis and tumor suppression, Cell Rep 17 (2016) 366–373, https://doi.org/10.1016/j. celrep.2016.09.022.
- [77] O. Fainaru, B. Almog, I. Pinchuk, M.J. Kupferminc, D. Lichtenberg, A. Many, Active labour is associated with increased oxidisibility of serum lipids ex vivo, BJOG 109 (8) (2002) 938–941, https://doi.org/10.1111/j.1471-0528.2002.01494.x.
- [78] T.J. Mocatta, C.C. Winterbourn, T.E. Inder, B.A. Darlow, The effect of gestational age and labour on markers of lipid and protein oxidation in cord plasma, Free Radic. Res. 38 (2) (2004) 185–191, https://doi.org/10.1080/ 10715760310001646048.
- [79] X. Lin, Y. Dai, X. Tong, et al., Excessive oxidative stress in cumulus granulosa cells induced cell senescence contributes to endometriosis-associated infertility, Redox Biol. 30 (2020) 101431, https://doi.org/10.1016/j.redox.2020.101431.
- Biol. 30 (2020) 101431, https://doi.org/10.1016/j.redox.2020.101431.
  [80] P. Zhang, D. Konja, Y. Zhang, Y. Wang, Communications between mitochondria and endoplasmic reticulum in the regulation of metabolic homeostasis, Cells 10 (9) (2021) 2195, https://doi.org/10.3390/cells10092195.
- [81] M. Jaganjac, L. Milkovic, N. Zarkovic, K. Zarkovic, Oxidative stress and regeneration, Free Radic. Biol. Med. 181 (2022) 154–165, https://doi.org/ 10.1016/j.freeradbiomed.2022.02.004.
- [82] J. Li, B. Jia, Y. Cheng, Y. Song, Q. Li, C. Luo, Targeting molecular mediators of ferroptosis and oxidative stress for neurological disorders, Oxid. Med. Cell. Longev. 2022 (2022) 3999083, https://doi.org/10.1155/2022/3999083. Published 2022 Jul 22.
- [83] L. Mahoney-Sánchez, H. Bouchaoui, S. Ayton, D. Devos, J.A. Duce, J. C. Devedjian, Ferroptosis and its potential role in the physiopathology of Parkinson's Disease, Prog. Neurobiol. 196 (2021) 101890, https://doi.org/ 10.1016/j.pneurobio.2020.101890.
- [84] M.A. Martínez-García, M. Luque-Ramírez, J.L. San-Millán, H.F. Escobar-Morreale, Body iron stores and glucose intolerance in premenopausal women: role of hyperandrogenism, insulin resistance, and genomic variants related to inflammation, oxidative stress, and iron metabolism, Diabetes Care 32 (8) (2009) 1525–1530, https://doi.org/10.2337/dc09-0420.
- [85] H.F. Escobar-Morreale, M. Luque-Ramírez, Role of androgen-mediated enhancement of erythropoiesis in the increased body iron stores of patients with polycystic ovary syndrome, Fertil. Steril. 95 (5) (2011) 1730, https://doi.org/ 10.1016/j.fertnstert.2011.01.038, 5.e1.
- [86] M. Luque-Ramírez, F. Álvarez-Blasco, M. Alpañés, H.F. Escobar-Morreale, Role of decreased circulating hepcidin concentrations in the iron excess of women with the polycystic ovary syndrome, J. Clin. Endocrinol. Metab. 96 (3) (2011) 846–852, https://doi.org/10.1210/jc.2010-2211.
- [87] H.F. Escobar-Morreale, Iron metabolism and the polycystic ovary syndrome, Trends Endocrinol. Metabol. 23 (10) (2012) 509–515, https://doi.org/10.1016/j. tem.2012.04.003.

- [88] N. Kayagaki, J.D. Webster, K. Newton, Control of cell death in health and disease, Annu. Rev. Pathol. 19 (2024) 157–180, https://doi.org/10.1146/annurevpathmechdis-051022-014433.
- [89] M. Kurokawa, S. Kornbluth, Caspases and kinases in a death grip, Cell 138 (2009) 838–854, https://doi.org/10.1016/j.cell.2009.08.021.
- [90] J.F. Kerr, A.H. Wyllie, A.R. Currie, Apoptosis: a basic biological phenomenon with wide-ranging implications in tissue kinetics, Br. J. Cancer 26 (1972) 239–257, https://doi.org/10.1038/bjc.1972.33.
- [91] B.T. Cookson, M.A. Brennan, Pro-inflammatory programmed cell death, Trends Microbiol. 9 (2001) 113–114, https://doi.org/10.1016/S0966-842X(00)01936-3.
- [92] D. Frank, J.E. Vince, Pyroptosis versus necroptosis: similarities, differences, and crosstalk, Cell Death Differ. 26 (2019) 99–114, https://doi.org/10.1038/s41418-018-0212-6.
- [93] D.N. Jackson, A.L. Theiss, Gut bacteria signaling to mitochondria in intestinal inflammation and cancer, Gut Microbes 11 (2020) 285–304, https://doi.org/ 10.1080/19490976.2019.1592421.
- [94] S.M. Man, R. Karki, T.D. Kanneganti, AIM2 inflammasome in infection, cancer, and autoimmunity: role in DNA sensing, inflammation, and innate immunity, Eur. J. Immunol. 46 (2016) 269–280, https://doi.org/10.1002/eji.201545839.
- [95] P. Kelk, A. Johansson, R. Claesson, L. Hanstrom, S. Kalfas, Caspase 1 involvement in human monocyte lysis induced by Actinobacillus actinomycetemcomitans leukotoxin, Infect. Immun. 71 (2003) 4448–4455, https://doi.org/10.1128/ IAI.71.8.4448-4455.2003.
- [96] P. Li, et al., Mice deficient in IL-1 beta-converting enzyme are defective in production of mature IL-1 beta and resistant to endotoxic shock, Cell 80 (1995) 401–411, https://doi.org/10.1016/0092-8674(95)90490-5.
- [97] F. Martinon, K. Burns, J. Tschopp, The inflammasome: a molecular platform triggering activation of inflammatory caspases and processing of proIL-beta, Mol. Cell 10 (2002) 417–426, https://doi.org/10.1016/S1097-2765(02)00599-3.
- [98] V.A. Rathinam, K.A. Fitzgerald, Inflammasome complexes: emerging mechanisms and effector functions, Cell 165 (2016) 792–800, https://doi.org/10.1016/j. cell.2016.03.046.
- [99] I. Jorgensen, Y. Zhang, B.A. Krantz, E.A. Miao, Pyroptosis triggers pore-induced intracellular traps (PITs) that capture bacteria and lead to their clearance by efferocytosis, J. Exp. Med. 213 (2016) 2113–2128, https://doi.org/10.1084/ jem.20151613.
- [100] H. Hara, et al., The NLRP6 inflammasome recognizes lipoteichoic acid and regulates Gram-positive pathogen infection, Cell 175 (2018) 1651–1664.e14, https://doi.org/10.1016/j.cell.2018.09.047.
- [101] R.C. Coll, K. Schroder, P. Pelegrín, NLRP3 and pyroptosis blockers for treating inflammatory diseases, Trends Pharmacol. Sci. 43 (8) (2022) 653–668, https:// doi.org/10.1016/j.tips.2022.04.003.
- [102] S.K. Hsu, C.Y. Li, I.L. Lin, et al., Inflammation-related pyroptosis, a novel programmed cell death pathway, and its crosstalk with immune therapy in cancer treatment, Theranostics 11 (18) (2021) 8813–8835, https://doi.org/10.7150/ thno.62521. Published 2021 Aug 12.
- [103] D. Wang, Y. Weng, Y. Zhang, et al., Exposure to hyperandrogen drives ovarian dysfunction and fibrosis by activating the NLRP3 inflammasome in mice, Sci. Total Environ. 745 (2020) 141049, https://doi.org/10.1016/j. scitoteny 2020 141049
- [104] J. Adams, Z. Liu, Y.A. Ren, et al., Enhanced inflammatory transcriptome in the granulosa cells of women with polycystic ovarian syndrome, J. Clin. Endocrinol. Metab. 101 (9) (2016) 3459–3468, https://doi.org/10.1210/jc.2015-4275.
- [105] Y. Li, Q. Zheng, D. Sun, et al., Dehydroepiandrosterone stimulates inflammation and impairs ovarian functions of polycystic ovary syndrome, J. Cell. Physiol. 234 (5) (2019) 7435–7447, https://doi.org/10.1002/jcp.27501.
- [106] Y. Zhang, X. Xie, Y. Ma, et al., Cyproterone acetate mediates IRE1α signaling pathway to alleviate pyroptosis of ovarian granulosa cells induced by hyperandrogen, Biology (Basel) 11 (12) (2022) 1761, https://doi.org/10.3390/ biology11121761. Published 2022 Dec 4.
- [107] Wei H, Zhang Z, Zhang S, et al. Resveratrol improves follicular development in PCOS rats by inhibiting the inflammatory response and pyroptosis of granulosa cells. Biol. Reprod. Published online November 15, 2024. doi:10.1093/biolre/ ioae160.
- [108] Z. Cai, S. He, R. Liu, L. Zhou, L. Zhao, Plumbagin rescues the granulosa cell's pyroptosis by reducing WTAP-mediated N6-methylation in polycystic ovary syndrome, J. Ovarian Res. 15 (1) (2022) 126, https://doi.org/10.1186/s13048-022-01058-1. Published 2022 Dec 3.
- [109] A.B. Guimarães-Costa, M.T. Nascimento, G.S. Froment, et al., Leishmania amazonensis promastigotes induce and are killed by neutrophil extracellular traps, Proc. Natl. Acad. Sci. U. S. A 106 (16) (2009) 6748–6753, https://doi.org/ 10.1073/pnas.0900226106.
- [110] Q. Remijsen, T. Vanden Berghe, E. Wirawan, et al., Neutrophil extracellular trap cell death requires both autophagy and superoxide generation, Cell Res. 21 (2) (2011) 290–304, https://doi.org/10.1038/cr.2010.150.
- [111] B.G. Yipp, B. Petri, D. Salina, et al., Infection-induced NETosis is a dynamic process involving neutrophil multitasking in vivo, Nat. Med. 18 (9) (2012) 1386–1393, https://doi.org/10.1038/nm.2847.
- [112] N. Dwivedi, M. Radic, Citrullination of autoantigens implicates NETosis in the induction of autoimmunity, Ann. Rheum. Dis. 73 (3) (2014) 483–491, https:// doi.org/10.1136/annrheumdis-2013-203844.
- [113] Q. Remijsen, T.W. Kuijpers, E. Wirawan, S. Lippens, P. Vandenabeele, T. Vanden Berghe, Dying for a cause: NETosis, mechanisms behind an antimicrobial cell death modality, Cell Death Differ. 18 (4) (2011) 581–588, https://doi.org/ 10.1038/cdd.2011.1.

- [114] S. Sangaletti, C. Tripodo, C. Chiodoni, et al., Neutrophil extracellular traps mediate transfer of cytoplasmic neutrophil antigens to myeloid dendritic cells toward ANCA induction and associated autoimmunity [published correction appears in Blood. 2022 Dec 15;140(24):2646-2647], Blood 120 (15) (2012) 3007–3018, https://doi.org/10.1182/blood-2012-03-416156.
- [115] H.R. Thiam, S.L. Wong, R. Qiu, et al., NETosis proceeds by cytoskeleton and endomembrane disassembly and PAD4-mediated chromatin decondensation and nuclear envelope rupture, Proc. Natl. Acad. Sci. U. S. A 117 (13) (2020) 7326–7337, https://doi.org/10.1073/pnas.1909546117.
- [116] E. Frangou, D. Vassilopoulos, J. Boletis, D.T. Boumpas, An emerging role of neutrophils and NETosis in chronic inflammation and fibrosis in systemic lupus erythematosus (SLE) and ANCA-associated vasculitides (AAV): implications for the pathogenesis and treatment, Autoimmun. Rev. 18 (8) (2019) 751–760, https://doi.org/10.1016/j.autrev.2019.06.011.
- [117] E. Rudnicka, K. Suchta, M. Grymowicz, et al., Chronic low grade inflammation in pathogenesis of PCOS, Int. J. Mol. Sci. 22 (7) (2021) 3789, https://doi.org/ 10.3390/ijms22073789. Published 2021 Apr 6.
- [118] S.A. Dabravolski, N.G. Nikiforov, A.H. Eid, et al., Mitochondrial dysfunction and chronic inflammation in polycystic ovary syndrome, Int. J. Mol. Sci. 22 (8) (2021) 3923, https://doi.org/10.3390/ijms22083923. Published 2021 Apr 10.
- [119] A.C. Özay, Ö.E. Özay, The importance of inflammation markers in polycystic ovary syndrome, Rev. Assoc. Med. Bras. 67 (3) (2021) 411–417, https://doi.org/ 10.1590/1806-9282.20200860, 1992.
- [120] S. He, X. Mao, H. Lei, et al., Peripheral blood inflammatory-immune cells as a predictor of infertility in women with polycystic ovary syndrome, J. Inflamm. Res. 13 (2020) 441–450, https://doi.org/10.2147/JIR.S260770. Published 2020 Aug 18.
- [121] N. Casares, et al., Caspase-dependent immunogenicity of doxorubicin-induced tumor cell death, J. Exp. Med. 202 (2005) 1691–1701, https://doi.org/10.1084/ jem.20050915.
- [122] J. Zhao, S. Jitkaew, Z. Cai, et al., Mixed lineage kinase domain-like is a key receptor interacting protein 3 downstream component of TNF-induced necrosis, Proc. Natl. Acad. Sci. U. S. A. 109 (14) (2012) 5322–5327, https://doi.org/ 10.1073/pnas.1200012109.
- [123] Z. Cai, S. Jitkaew, J. Zhao, et al., Plasma membrane translocation of trimerized MLKL protein is required for TNF-induced necroptosis [published correction appears in Nat Cell Biol. 2014 Feb;16(2):200], Nat. Cell Biol. 16 (1) (2014) 55–65, https://doi.org/10.1038/ncb2883.
- [124] X. Chen, W. Li, J. Ren, et al., Translocation of mixed lineage kinase domain-like protein to plasma membrane leads to necrotic cell death, Cell Res. 24 (1) (2014) 105–121, https://doi.org/10.1038/cr.2013.171.
- [125] M. Wang, K. An, J. Huang, R. Mprah, H. Ding, A novel model based on necroptosis to assess progression for polycystic ovary syndrome and identification of potential therapeutic drugs, Front. Endocrinol. 15 (2024 May 24) 1432697, https://doi. org/10.3389/fendo.2024.1432697 [published correction appears in.
- [126] G.R. Chaudhary, P.K. Yadav, A.K. Yadav, et al., Necroptosis in stressed ovary, J. Biomed. Sci. 26 (1) (2019) 11, https://doi.org/10.1186/s12929-019-0504-2. Published 2019 Jan 21.
- [127] K.H. Tsui, P.H. Wang, L.T. Lin, C.J. Li, DHEA protects mitochondria against dual modes of apoptosis and necroptosis in human granulosa HO23 cells, Reproduction 154 (2) (2017) 101–110, https://doi.org/10.1530/REP-17-0016.
- [128] H.X. Dong, Q. Wang, Z. Wang, et al., Impact of low frequency electro-acupuncture on glucose and lipid metabolism in unmarried PCOS women: a randomized controlled trial, Chin. J. Integr. Med. 27 (10) (2021) 737–743, https://doi.org/ 10.1007/s11655-021-3482-z.
- [129] M. Dastorani, E. Aghadavod, N. Mirhosseini, et al., The effects of vitamin D supplementation on metabolic profiles and gene expression of insulin and lipid metabolism in infertile polycystic ovary syndrome candidates for in vitro fertilization, Reprod. Biol. Endocrinol. 16 (1) (2018) 94, https://doi.org/ 10.1186/s12958-018-0413-3. Published 2018 Oct 4.
- [130] L. Zhang, F. Wang, D. Li, Y. Yan, H. Wang, Transferrin receptor-mediated reactive oxygen species promotes ferroptosis of KGN cells via regulating NADPH oxidase 1/PTEN induced kinase 1/acyl-CoA synthetase long chain family member 4 signaling, Bioengineered 12 (1) (2021) 4983–4994, https://doi.org/10.1080/ 21655979.2021.1956403.
- [131] F. González, Inflammation in Polycystic Ovary Syndrome: underpinning of insulin resistance and ovarian dysfunction, Steroids 77 (4) (2012) 300–305, https://doi. org/10.1016/j.steroids.2011.12.003.
- [132] H.F. Escobar-Morreale, Polycystic ovary syndrome: definition, aetiology, diagnosis and treatment, Nat. Rev. Endocrinol. 14 (5) (2018) 270–284, https:// doi.org/10.1038/nrendo.2018.24.
- [133] T. Srnovršnik, I. Virant-Klun, B. Pinter, Heavy metals and essential elements in association with oxidative stress in women with polycystic ovary syndrome-A systematic review, Antioxidants 12 (7) (2023) 1398, https://doi.org/10.3390/ antiox12071398. Published 2023 Jul 7.
- [134] A.J. Duleba, A. Dokras, Is PCOS an inflammatory process? Fertil. Steril. 97 (1) (2012) 7–12, https://doi.org/10.1016/j.fertnstert.2011.11.023.

- [135] X. Li, Y. Lin, X. Cheng, et al., Ovarian ferroptosis induced by androgen is involved in Jack and the second s
- cytokines promote an inflammatory cascade in PCOS patients via altering the

follicular microenvironment, Front. Immunol. 12 (2021) 685724, https://doi. org/10.3389/fimmu.2021.685724. Published 2021 May 17.

[137] Y.H. Shen, S. Peng, T. Zhu, M.J. Shen, Mechanisms of granulosa cell programmed cell death and follicular atresia in polycystic ovary syndrome, Physiol. Res. 74 (1) (2025) 31-40, https://doi.org/10.33549/physiolres.935485.