



Imbalances in Copper or Zinc Concentrations Trigger Further Trace Metal Dyshomeostasis in Amyloid-Beta Producing *Caenorhabditis elegans*

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Alzheimer's Disease (AD), a progressive neurodegenerative disease characterized by the buildup of amyloid-beta (A β) plaques, is believed to be a disease of trace metal dyshomeostasis. Amyloid-beta is known to bind with high affinity to trace metals copper and zinc. This binding is believed to cause a conformational change in A β , transforming A β into a configuration more amenable to forming aggregations. Currently, the impact of A β -trace metal binding on trace metal homeostasis and the role of trace metals copper and zinc as deleterious or beneficial in AD remain elusive. Given that Alzheimer's Disease is the sixth leading cause of adult death in the U.S., elucidating the molecular interactions that characterize Alzheimer's Disease pathogenesis will allow for better treatment options. To that end, the model organism *C. elegans* is used in this study. *C. elegans*, a transparent nematode whose connectome has been fully established, is an amenable model to study AD phenomena using a multi-layered, interconnected approach. A β -producing and non-A β -producing *C. elegans* were individually supplemented with copper and zinc. On day 6 and day 9 after synchronization, the percent of worms paralyzed, concentration of copper, and concentration of zinc were measured in both groups of worms. This study demonstrates that dyshomeostasis of trace metals copper or zinc triggers further trace metal dyshomeostasis in A β -producing worms, while dyshomeostasis of copper or zinc triggers a return to equilibrium in non-A β -producing worms. This supports the characterization of Alzheimer's Disease as a disease of trace metal dyshomeostasis.

Keywords: Alzheimer's, amyloid-beta, copper, zinc, trace metal, dyshomeostasis, *Caenorhabditis elegans*, imbalances

INTRODUCTION

Alzheimer's Disease (AD) is the 6th leading cause of death in the U.S., with one in ten people age 65 or older having AD (Alzheimer's Association, 2021). As a progressive neurodegenerative disease, AD is characterized by extra-neuronal amyloid-beta plaques and intraneuronal tau neurofibrillary tangles which affect memory and cognition. Amyloid-beta plaques are aggregates of the amyloid-beta peptide (A β), a cleavage product of the amyloid precursor protein (APP). Buildup of A β causes neural death and neuroinflammation.

Neurodegenerative diseases such as Alzheimer's, Parkinson's, and Wilson's Diseases have been associated with metal dyshomeostasis, which often accompanies aging (Luo et al., 2011; Squitti, 2012; Singh et al., 2013). Metal dyshomeostasis occurs when metal levels increase or decrease beyond normal bounds. As important components of vitamins and enzymes, trace metals play a crucial role in neural and biochemical processes. When in homeostasis, these trace metals facilitate proper brain functioning and growth by protecting against reactive oxygen species (ROS), regulating gene expression, and activating enzymes. The dyshomeostasis of trace metals results in cellular damage and oxidative injury, induced by the formation of ROS (Grochowski et al., 2019).

Both trace metals copper and zinc play key roles in proper brain functioning. Copper is an essential trace element that plays a key role in energy production, free radicals scavenging, and neurotransmission (Singh et al., 2013). Zinc is another essential trace element that plays a key role in neurotransmission and redox regulation (Grochowski et al., 2019). Amyloid beta plaques have high affinity to trace metals copper and zinc and have thus been found to contain high concentrations of these trace metals (Bush et al., 1994; Atwood et al., 1998; Lovell et al., 1998; Sayre et al., 2000; Suh et al., 2000; Cherny et al., 2001; Dong et al., 2003; Miller et al., 2006; Mital et al., 2015; Ejaz et al., 2020). For instance, a 339% increase in Zn and a 466% increase in Cu were found in amyloid beta plaques of AD patients in comparison to healthy subjects (Leskovjan et al., 2009). The levels of copper and zinc in AD, however, remains controversial (Huang et al., 2000; Strausak et al., 2001; Cerpa et al., 2005; Kessler et al., 2005; Watt et al., 2010; Bagheri et al., 2018; Rana and Sharma, 2019). Some studies indicate copper deficiency in AD, suggesting a need for supplementation (Borchardt et al., 1999; Exley, 2006; Jiao and Yang, 2007; Kessler et al., 2008; Vural et al., 2010; Kaden et al., 2011; Exley et al., 2012; Xu et al., 2017), while others indicate copper excess in AD, suggesting a need for chelating agents (Cherny et al., 2001; Sparks et al., 2006; Hua et al., 2011; Luo et al., 2011; Ceccom et al., 2012; Eskici and Axelsen, 2012; Brewer, 2014; Squitti et al., 2014; Yu et al., 2015; Patel and Aschner, 2021). Similarly, some studies indicate zinc deficiency in AD (Kapaki et al., 1989; Molina et al., 1998; Brewer et al., 2010; Rivers-Auty et al., 2021), while others indicate zinc excess (Lovell et al., 1998; Religa et al., 2006; Bonda et al., 2011; Greenough et al., 2013; James et al., 2017). These conflicting findings could be partially due to differences in the brain regions in which copper and zinc were measured. With over 5 million Americans currently living with AD and nearly 14 million projected to be living with AD by 2050, better understanding the molecular mechanisms characterizing the involvement of copper and zinc dyshomeostasis in AD will allow for better treatment options and outcomes (Alzheimer's Association, 2021).

Caenorhabditis elegans, a non-parasitic nematode whose connectome has been fully established, is an advantageous model for studying the molecular mechanisms in Alzheimer's Disease (Caito et al., 2012). The nematode's simple nervous system and transparency allow for the study of the effects of AD on neuronal pathways and function. Roughly 38% of worm genes have a human ortholog, such as APP and tau, making *C. elegans*

an excellent *in vivo* model for the study of AD (Shaye and Greenwald, 2011). Since the toxic A β 42-peptide is expressed in muscle cells in *C. elegans* strain CL2006, A β aggregations result in the paralysis of *C. elegans*, thus allowing the extent of A β aggregation in response to different treatments to be viewed macroscopically (Saharia et al., 2016).

Given that molecular mechanisms characterizing the interaction between copper, zinc, and amyloid-beta remain elusive, the present study aims to elucidate whether the dyshomeostasis of one trace metal induces the dyshomeostasis of other trace metals and of amyloid-beta in Alzheimer's Disease. It is hypothesized that increases in amyloid-beta aggregations are part of a failed protective homeostatic mechanism to bind excess trace metals copper and zinc. The present study newly shows that dyshomeostasis of trace metals copper or zinc triggers further trace metal dyshomeostasis in A β -producing worms while dyshomeostasis of copper or zinc triggers a return to equilibrium in non-A β -producing worms.

MATERIALS AND METHODS

Nematode Strains and Maintenance

Caenorhabditis elegans strains were received from the *Caenorhabditis* Genetics Center (CGC). The transgenic *C. elegans* strain CL2006, which expresses human A β _{1–42} in body-wall muscle cells, is characterized by progressive, adult-onset paralysis and a roller phenotype (Link, 1995). The *C. elegans* strain N2 represents the wild type. During two independent trials, worm strains were synchronized according to the following procedure: Adult hermaphrodite worms were transferred to fresh plates and allowed to lay eggs for 2–4 h. After removal of the adult parental worms, the synchronized progeny were allowed to reach adulthood, then later scored for paralysis (Fonte et al., 2002). The worms were propagated at 20°C on Nematode Growth Media (NGM) plates seeded with the bacterial strain OP50 and supplemented with either copper or zinc (Brenner, 1974).

Supplementation With Copper and Zinc

CuCl₂ was used to supplement the worms with copper. CuCl₂ stock solution was diluted into a live *E. coli* OP50 suspension, reaching a final concentration of 150 μ M, and was placed on the surface of the NGM plates. Once the worms reached adulthood (day 3), a group of synchronized CL2006 worms and a group of synchronized N2 worms were placed on the copper-supplemented plates. ZnSO₄ was used to supplement the worms with zinc. ZnSO₄ stock solution was diluted into a live *E. coli* OP50 suspension, reaching a final concentration of 500 μ M, and was placed on the surface of the NGM plates. Once the worms reached adulthood, a group of synchronized CL2006 worms and a group of synchronized N2 worms were placed on these zinc-supplemented plates.

Paralysis Assay

On days 6 and 9¹ after synchronization, 20 worms from the copper-supplemented and zinc-supplemented CL2006 and N2 groups were tested for paralysis. Paralysis indicates the extent of A β -aggregation development. The worms were tested for paralysis by tapping their noses with a platinum wire pick. Worms that moved their noses but failed to move their bodies were scored as "paralyzed" (Luo et al., 2011).

Lysis Procedure

On days 6 and 9, thirty worms from each of the four groups: (1) copper-supplemented CL2006, (2) zinc-supplemented CL2006, (3) copper-supplemented N2, (4) zinc-supplemented N2, were lysed in preparation for copper and zinc colorimetric assays. The following procedure is especially useful for dauer larvae lysis. Worms were spun in a centrifuge at 4,000 rpm for 1 min to a pellet. The supernatant was removed, and the pellet was washed in 1.5 mL of ice cold L15 buffer. The worms were centrifuged, and the supernatant was removed once again. Twenty-five microliters of the pellet was pipetted onto a glass slide. A 50 mm glass coverslip was added on top, and pressure was applied to the coverslip using a pipette. When viewed under a microscope, head disruption head could be visualized as pressure was applied with the pipette tip. Pressure continued to be applied until most of the worms were exploded. The contents on the coverslip and slide were washed off with 1 ml of cold L15 into a test tube. Finally, this L15-cell solution was pipetted vigorously 25 times to ensure the *C. elegans* were completely lysed.

Copper and Zinc Colorimetric Assays

To quantify the amount of copper in the *C. elegans* on days 6 and 9, a copper colorimetric assay (Elabsience) was applied to the lysed *C. elegans* solution. Similarly, to quantify the amount of zinc in the *C. elegans* on days 6 and 9, a zinc colorimetric assay (Elabsience) was applied to the lysed *C. elegans* solution. Once the standard wells were created for both assays, the percent transmittance of the standards and test groups was measured using a colorimeter. The percent transmittance was converted to ion content ($\mu\text{mol/L}$) as specified by the Elabsience assays.

Statistical Analysis

All values were expressed as mean \pm SEM. Statistical analysis involving two groups was conducted using a *t*-test. Statistical analysis involving more than two groups was conducted using a one-way analysis of variance (ANOVA) followed by a *post-hoc* analysis using Tukey test. The differences were considered to be significant at $p < 0.05$.

RESULTS

To elucidate the differences in trace metal homeostasis maintenance in amyloid-beta producing *C. elegans* compared to

non-amyloid-beta producing *C. elegans*, both strains of *C. elegans* were supplemented with copper and zinc individually.

Zinc Concentration Changes in Response to Copper Supplementation

When A β -producing *C. elegans* were supplemented with copper, the zinc concentration increased significantly ($p = 0.013$) from day 6 ($13.5 \pm 0.6 \mu\text{mol/L}$) to day 9 ($20.1 \pm 0.9 \mu\text{mol/L}$). Likewise, when wild-type worms were supplemented with copper, the zinc concentration increased significantly ($p = 0.041$) from day 6 ($16.0 \pm 0.9 \mu\text{mol/L}$) to day 9 ($19.3 \pm 0.6 \mu\text{mol/L}$, **Figure 1**). Additionally, the percent change in zinc content from day 6 to day 9 in A β -producing *C. elegans* (49% increase) was more than double the percent change in wild-type *C. elegans* (21% increase). This indicates that a high copper concentration results in a larger change in the zinc concentration in A β -producing *C. elegans* compared to non-A β -producing *C. elegans*.

Copper Concentration Changes in Response to Zinc Supplementation

When A β -producing *C. elegans* were supplemented with zinc, the copper concentration increased significantly ($p = 0.022$) from day 6 ($27.8 \pm 4.9 \mu\text{mol/L}$) to day 9 ($58.6 \pm 3.7 \mu\text{mol/L}$). In contrast, when wild-type *C. elegans* were supplemented with zinc, the copper concentration decreased significantly ($p = 0.012$) from day 6 ($60.8 \pm 2.4 \mu\text{mol/L}$) to day 9 ($24.7 \pm 5.4 \mu\text{mol/L}$, **Figure 2**). In fact, the copper content on day 6 in mutant *C. elegans* was roughly equivalent to the copper content on day 9 in wild-type *C. elegans* ($p = 0.9$). Similarly, the copper content on day 9 in mutant *C. elegans* was roughly equivalent to the copper content on day 6 in wild-type *C. elegans* ($p = 0.9$). This indicates that a high zinc concentration through supplementation results in an increase in the copper content of A β -producing *C. elegans* that is roughly equal in magnitude to the decrease in copper content in non-A β -producing *C. elegans*.

Effect of Copper and Zinc Dyshomeostasis on A β Aggregations

To characterize the effect of imbalances in such trace metals on the extent of A β aggregations, the percent of worms paralyzed was measured in A β -producing *C. elegans* supplemented with copper or zinc. The percent of worms paralyzed significantly increased in both copper-supplemented mutant worms ($p = 0.0142$) from day 6 ($37 \pm 4\%$) to day 9 ($92 \pm 8\%$) and zinc-supplemented mutant worms ($p = 0.0187$) from day 6 ($67 \pm 0\%$) to day 9 ($87 \pm 4\%$), as shown in **Figure 3**. The change in percent paralyzed from day 6 to day 9 was larger for the copper-supplemented group (145% increase) compared to the zinc-supplemented group (31% increase). Also, the percent paralyzed was significantly higher on day 6 for the zinc-supplemented group compared to the copper-supplemented group ($p = 0.0098$), while there was no significant difference in percent of the worms paralyzed between the two groups on day 9 ($p = 0.3483$). Overall, high concentrations of both copper and zinc are positively correlated with increases in the percent of worms paralyzed.

¹Day 6 was chosen to ensure that the effects of supplementation, which occurred when worms reached adulthood on day 3, will be significant. Day 9 was chosen as the last date of data collection to ensure that dead worms were not mistaken for paralyzed worms.

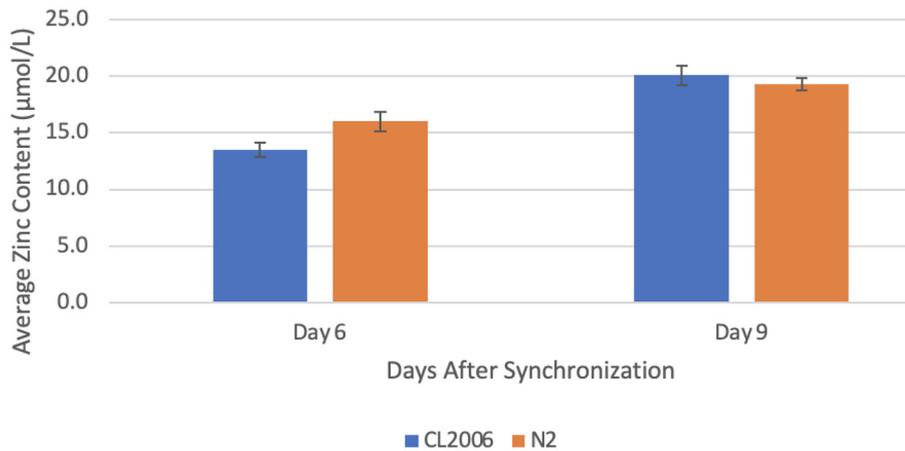


FIGURE 1 | Average zinc content in *C. elegans* supplemented with copper. When amyloid-beta producing *C. elegans* (CL2006) were supplemented with copper, a statistically significant ($p = 0.013$) increase in the average zinc content occurred from day 6 ($13.5 \pm 0.6 \mu\text{mol/L}$) to day 9 ($20.1 \pm 0.9 \mu\text{mol/L}$). Similarly, when non-amyloid-beta producing *C. elegans* (N2) were supplemented with copper, a statistically significant ($p = 0.041$) increase in the average zinc content occurred from day 6 ($16.0 \pm 0.9 \mu\text{mol/L}$) to day 9 ($19.3 \pm 0.6 \mu\text{mol/L}$). Values are mean \pm SEM and are representative of 2 experiments where 30 *C. elegans* were analyzed at each time point.

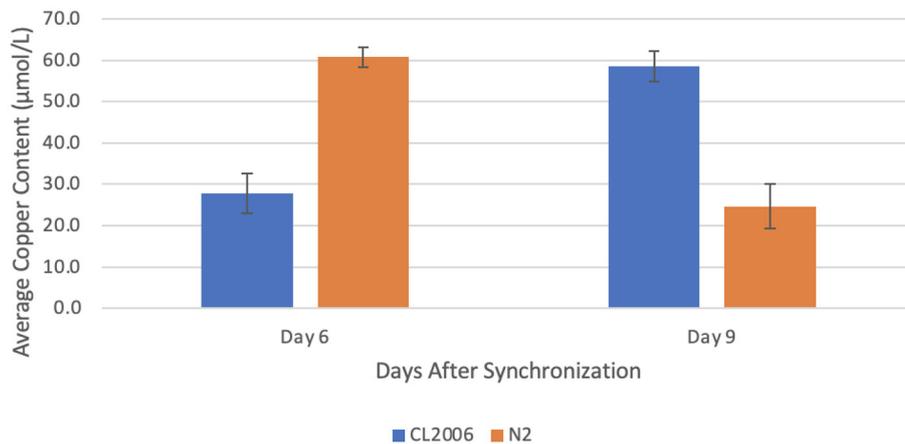


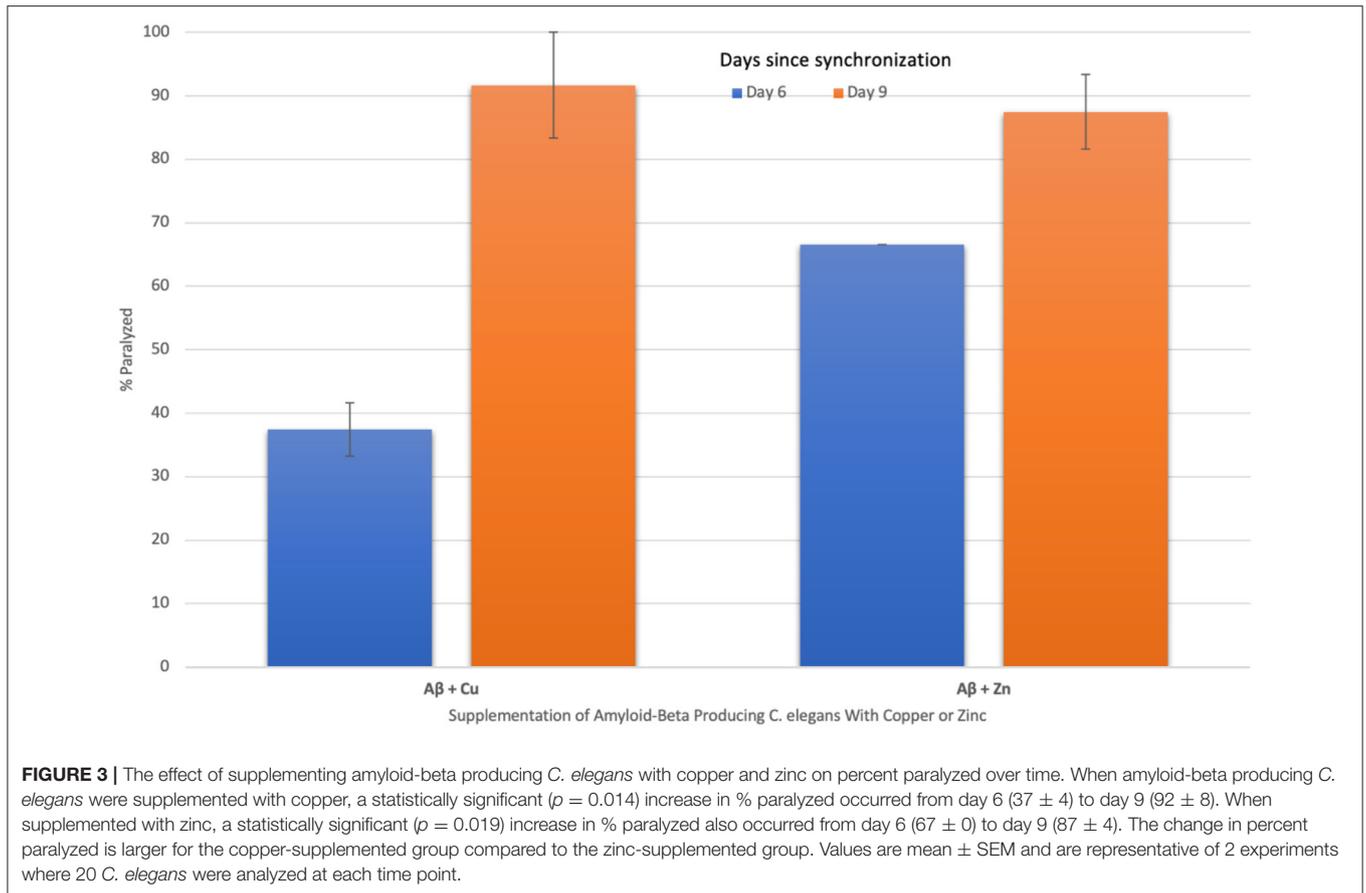
FIGURE 2 | Average copper content in *C. elegans* supplemented with zinc. When amyloid-beta producing *C. elegans* (CL2006) were supplemented with zinc, a statistically significant ($p = 0.022$) increase in the average copper content occurred from day 6 ($27.8 \pm 4.9 \mu\text{mol/L}$) to day 9 ($58.6 \pm 3.7 \mu\text{mol/L}$). In contrast, when non-amyloid-beta producing *C. elegans* (N2) were supplemented with zinc, a statistically significant ($p = 0.012$) decrease in the average copper content occurred from day 6 ($60.8 \pm 2.4 \mu\text{mol/L}$) to day 9 ($24.7 \pm 5.4 \mu\text{mol/L}$). Values are mean \pm SEM and are representative of 2 experiments where 30 *C. elegans* were analyzed at each time point.

DISCUSSION

Aging has been found to trigger copper and zinc dyshomeostasis (Myhre et al., 2013; McCord and Aizenman, 2014; Nuttall and Oteiza, 2014; Malavolta et al., 2015). While trace metals copper and zinc are crucial for normal functioning, excess copper, and zinc are highly damaging to proteins. Excess copper and zinc are known to bind with high affinity to A β , resulting in visible precipitation into an aggregated form (Bush et al., 1994; Huang et al., 1997, 2000; Atwood et al.,

2000; Kumar et al., 2016; Bagheri et al., 2018; Barykin et al., 2018). Therefore, it is of particular interest to determine how changes in the homeostasis of a given trace metal influence the homeostasis of other trace metals and the aggregation state of A β .

The present study has found that in A β -producing *C. elegans*, imbalances in trace metals copper or zinc trigger further trace metal dyshomeostasis. When supplemented with copper, zinc levels increase significantly and when supplemented with zinc, copper levels increase significantly. Thus, an imbalance in



either trace metal causes a cascading effect resulting in further imbalances. This triggering of further trace metal dyshomeostasis might explain why the percent of worms paralyzed, which correlates to A β -aggregation levels, significantly increases in both copper and zinc supplemented mutant worms from day 6 to day 9.

However, in wild-type worms, dyshomeostasis of copper or zinc ultimately triggers a return to equilibrium. When copper levels increase through supplementation, zinc levels correspondingly increase from day 6 to day 9. In contrast, when zinc levels increase through supplementation, copper levels decrease from day 6 to day 9. Therefore, it is possible that through a negative feedback mechanism loop, an increase in copper triggers an increase in zinc which ultimately causes a decrease in copper and a return to equilibrium.

Increases in the concentration of zinc or copper, through supplementation, both result in increases in the percent of worms paralyzed, reflecting higher A β -aggregation levels, from day 6 to day 9. Given that trace metal levels naturally increase to a degree during the aging process, it is possible that in certain populations more prone to developing amyloidogenic diseases, trace metal levels dramatically increase during aging. Since A β avidly binds to trace metals copper and zinc, it is possible that when trace metal levels increase during the aging process, A β

levels increase in an effort to bind excess copper and zinc (Squitti et al., 2021). The binding of trace metals such as copper and zinc to A β is known to trigger an A β conformational shape change (Barykin et al., 2018; Kim et al., 2018; De Benedictis et al., 2019), thus transforming A β into a configuration more amenable to forming aggregations.

While both copper and zinc dyshomeostasis result in an increase in the percent of worms paralyzed over time, copper might have a stronger effect on the percent of worms paralyzed, reflecting A β aggregations, compared to zinc. The zinc supplementation concentration ($500 \mu\text{M ZnSO}_4$) was over three times higher than the copper supplementation concentration ($150 \mu\text{M CuCl}_2$); however, the percent of worms paralyzed on day 6 in the zinc supplemented group was only about 1.8 times higher than the copper supplemented group. The zinc supplementation (Kumar et al., 2016) and copper supplementation (Minniti et al., 2009) were chosen based on previous publications that found considerable changes in amyloid-beta aggregations, but did not measure whether dyshomeostasis of one trace metal triggers dyshomeostasis of other trace metals. Additionally, despite the higher zinc supplementation, there was no significant difference in the percent of worms paralyzed by day 9 when comparing the zinc supplemented group and the copper supplemented group. While

this could simply be due to natural age-related A β aggregation development caused by the inserted A β gene as the mutant *C. elegans* approach the end of their lifespan, more trials would be needed to better understand if there is a significant difference between the effect of copper vs. zinc on A β aggregations. It would be particularly useful to measure the percent of worms paralyzed when supplementing with the same concentration of CuCl₂ and ZnSO₄ in the future. Future work also includes measuring the levels of copper and zinc in A β -producing and non-A β -producing *C. elegans* without any supplementation to determine the copper and zinc homeostatic ranges.

Overall, the novelty of this study is the experimental demonstration that dyshomeostasis of trace metals copper or zinc triggers further trace metal dyshomeostasis in A β -producing worms, while dyshomeostasis of copper or zinc triggers a return to equilibrium in non-A β -producing worms. Future directions include determining how increases in amyloid-beta aggregations might be part of a failed protective homeostatic mechanism to bind excess trace metals copper and zinc. Additional future directions will include elucidating the mediating factors that facilitate A β -trace metal binding and testing the effects of trace metal chelators on A β levels.

REFERENCES

- Alzheimer's Association (2021). *Alzheimer's Disease Facts and Figures*. Alzheimer's Association. Available online at: <https://www.alz.org/alzheimers-dementia/facts-figures> (accessed August 8, 2021).
- Atwood, C. S., Moir, R. D., Huang, X., Scarpa, R. C., Bacarra, N. M., Romano, D. M., et al. (1998). Dramatic aggregation of Alzheimer abeta by Cu(II) is induced by conditions representing physiological acidosis. *J. Biol. Chem.* 273, 12817–12826. doi: 10.1074/jbc.273.21.12817
- Atwood, C. S., Scarpa, R. C., Huang, X., Moir, R. D., Jones, W. D., Fairlie, D. P., et al. (2000). Characterization of copper interactions with alzheimer amyloid beta peptides: identification of an attomolar-affinity copper binding site on amyloid beta1-42. *J. Neurochem.* 75, 1219–1233. doi: 10.1046/j.1471-4159.2000.0751219.x
- Bagheri, S., Squitti, R., Haertlé, T., Siotto, M., and Saboury, A. A. (2018). Role of copper in the onset of Alzheimer's disease compared to other metals. *Front. Aging Neurosci.* 9:446. doi: 10.3389/fnagi.2017.00446
- Barykin, E. P., Petrushanko, I. Y., Kozin, S. A., Telegin, G. B., Chernov, A. S., Lopina, O. D., et al. (2018). Phosphorylation of the amyloid-beta peptide inhibits zinc-dependent aggregation, prevents Na,K-ATPase inhibition, and reduces cerebral plaque deposition. *Front. Mol. Neurosci.* 11:302. doi: 10.3389/fnmol.2018.00302
- Bonda, D. J., Lee, H. G., Blair, J. A., Zhu, X., Perry, G., and Smith, M. A. (2011). Role of metal dyshomeostasis in Alzheimer's disease. *Metallomics* 3, 267–270. doi: 10.1039/c0mt00074d
- Borchardt, T., Camakaris, J., Cappai, R., Masters, C. L., Beyreuther, K., and Multhaup, G. (1999). Copper inhibits beta-amyloid production and stimulates the non-amyloidogenic pathway of amyloid-precursor-protein secretion. *Biochem. J.* 344(Pt 2), 461–467.
- Brenner, S. (1974). The genetics of *Caenorhabditis elegans*. *Genetics* 77, 71–94.
- Brewer, G. J. (2014). Alzheimer's disease causation by copper toxicity and treatment with zinc. *Front. Aging Neurosci.* 6:92. doi: 10.3389/fnagi.2014.00092
- Brewer, G. J., Kanzer, S. H., Zimmerman, E. A., Molho, E. S., Celmins, D. F., Heckman, S. M., et al. (2010). Subclinical zinc deficiency in Alzheimer's disease and Parkinson's disease. *Am. J. Alzheimers Dis. Other Dement.* 25, 572–575. doi: 10.1177/1533317510382283

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/supplementary material, further inquiries can be directed to the corresponding author/s.

AUTHOR CONTRIBUTIONS

The author confirms being the sole contributor of this work and has approved it for publication.

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- Bush, A. I., Pettingell, W. H., Multhaup, G., d Paradis, M., Vonsattel, J. P., Gusella, J. F., et al. (1994). Rapid induction of Alzheimer A beta amyloid formation by zinc. *Science* 265, 1464–1467. doi: 10.1126/science.8073293
- Caito, S., Fretham, S., Martinez-Finley, E., Chakraborty, S., Avila, D., Chen, P., et al. (2012). Genome-wide analyses of metal responsive genes in *Caenorhabditis elegans*. *Front. Genet.* 3:52. doi: 10.3389/fgene.2012.00052
- Ceccom, J., Coslédan, F., Halley, H., Francès, B., Lassalle, J. M., and Meunier, B. (2012). Copper chelator induced efficient episodic memory recovery in a non-transgenic Alzheimer's mouse model. *PLoS ONE* 7:e43105. doi: 10.1371/journal.pone.0043105
- Cerpa, W., Varela-Nallar, L., Reyes, A. E., Minniti, A. N., and Inestrosa, N. C. (2005). Is there a role for copper in neurodegenerative diseases? *Mol. Aspects Med.* 26, 405–420. doi: 10.1016/j.mam.2005.07.011
- Cherny, R. A., Atwood, C. S., Xilinas, M. E., Gray, D. N., Jones, W. D., McLean, C. A., et al. (2001). Treatment with a copper-zinc chelator markedly and rapidly inhibits beta-amyloid accumulation in Alzheimer's disease transgenic mice. *Neuron* 30, 665–676. doi: 10.1016/s0896-6273(01)00317-8
- De Benedictis, C. A., Vilella, A., and Grabrucker, A. M. (2019). "The role of trace metals in Alzheimer's disease," in *Alzheimer's Disease*, ed T. Wisniewski (Brisbane, AU: Codon Publications), 85–106. doi: 10.15586/alzheimersdisease.2019.ch6
- Dong, J., Atwood, C. S., Anderson, V. E., Siedlak, S. L., Smith, M. A., Perry, G., et al. (2003). Metal binding and oxidation of amyloid-beta within isolated senile plaque cores: Raman microscopic evidence. *Biochemistry* 42, 2768–2773. doi: 10.1021/bi0272151
- Ejaz, H. W., Wang, W., and Lang, M. (2020). Copper toxicity links to pathogenesis of Alzheimer's disease and therapeutics approaches. *Int. J. Mol. Sci.* 21:7660. doi: 10.3390/ijms21207660
- Eskici, G., and Axelsen, P. H. (2012). Copper and oxidative stress in the pathogenesis of Alzheimer's disease. *Biochemistry* 51, 6289–6311. doi: 10.1021/bi3006169
- Exley, C. (2006). Aluminium and iron, but neither copper nor zinc, are key to the precipitation of beta-sheets of Abeta_{42} in senile plaque cores in Alzheimer's disease. *J. Alzheimers Dis.* 10, 173–177. doi: 10.3233/jad-2006-102-305
- Exley, C., House, E., Polwart, A., and Esiri, M. M. (2012). Brain burdens of aluminum, iron, and copper and their relationships with amyloid- β pathology in 60 human brains. *J. Alzheimers Dis.* 31, 725–730. doi: 10.3233/JAD-2012-120766

- Fonte, V., Kapulkin, W. J., Taft, A., Fluet, A., Friedman, D., and Link, C. D. (2002). Interaction of intracellular beta amyloid peptide with chaperone proteins. *Proc. Natl. Acad. Sci. U.S.A.* 99, 9439–9444. doi: 10.1073/pnas.152313999
- Greenough, M. A., Camakaris, J., and Bush, A. I. (2013). Metal dyshomeostasis and oxidative stress in Alzheimer's disease. *Neurochem. Int.* 62, 540–555. doi: 10.1016/j.neuint.2012.08.014
- Grochowski, C., Blicharska, E., Krukow, P., Jonak, K., Maciejewski, M., Szczepanek, D., et al. (2019). Analysis of trace elements in human brain: its aim, methods, and concentration levels. *Front. Chem.* 7:115. doi: 10.3389/fchem.2019.00115
- Hua, H., Münter, L., Harmeier, A., Georgiev, O., Multhaup, G., and Schaffner, W. (2011). Toxicity of Alzheimer's disease-associated A β peptide is ameliorated in a *Drosophila* model by tight control of zinc and copper availability. *Biol. Chem.* 392, 919–926. doi: 10.1515/BC.2011.084
- Huang, X., Atwood, C. S., Moir, R. D., Hartshorn, M. A., Vonsattel, J. P., Tanzi, R. E., et al. (1997). Zinc-induced Alzheimer's Abeta1-40 aggregation is mediated by conformational factors. *J. Biol. Chem.* 272, 26464–26470. doi: 10.1074/jbc.272.42.26464
- Huang, X., Cuajungco, M. P., Atwood, C. S., Moir, R. D., Tanzi, R. E., and Bush, A. I. (2000). Alzheimer's disease, beta-amyloid protein and zinc. *J. Nutr.* 130, 1488S–1492S. doi: 10.1093/jn/130.5.1488S
- James, S. A., Churches, Q. I., de Jonge, M. D., Birchall, I. E., Streltsov, V., McColl, G., et al. (2017). Iron, copper, and zinc concentration in A β plaques in the APP/PS1 mouse model of Alzheimer's disease correlates with metal levels in the surrounding neuropil. *ACS Chem. Neurosci.* 8, 629–637. doi: 10.1021/acschemneuro.6b00362
- Jiao, Y., and Yang, P. (2007). Mechanism of copper(II) inhibiting Alzheimer's amyloid beta-peptide from aggregation: a molecular dynamics investigation. *J. Phys. Chem. B* 111, 7646–7655. doi: 10.1021/jp0673359
- Kaden, D., Bush, A. I., Danzeisen, R., Bayer, T. A., and Multhaup, G. (2011). Disturbed copper bioavailability in Alzheimer's disease. *Int. J. Alzheimers Dis.* 2011:345614. doi: 10.4061/2011/345614
- Kapaki, E., Segditsa, J., and Papageorgiou, C. (1989). Zinc, copper and magnesium concentration in serum and CSF of patients with neurological disorders. *Acta Neurol. Scand.* 79, 373–378. doi: 10.1111/j.1600-0404.1989.tb03803.x
- Kessler, H., Pajonk, F. G., Bach, D., Schneider-Axmann, T., Falkai, P., Herrmann, W., et al. (2008). Effect of copper intake on CSF parameters in patients with mild Alzheimer's disease: a pilot phase 2 clinical trial. *J. Neural Transm.* 115, 1651–1659. doi: 10.1007/s00702-008-0136-2
- Kessler, H., Pajonk, F. G., Suppran, T., Falkai, P., Multhaup, G., and Bayer, T. A. (2005). The role of copper in the importance of copper in the pathophysiology of Alzheimer's disease. *Neurologist* 76, 581–585. doi: 10.1007/s00115-004-1849-6
- Kim, A. C., Lim, S., and Kim, Y. K. (2018). Metal ion effects on A β and tau aggregation. *Int. J. Mol. Sci.* 19:128. doi: 10.3390/ijms19010128
- Kumar, J., Barhydt, T., Awasthi, A., Lithgow, G. J., Killilea, D. W., and Kapahi, P. (2016). Zinc levels modulate lifespan through multiple longevity pathways in *Caenorhabditis elegans*. *PLoS ONE* 11:e0153513. doi: 10.1371/journal.pone.0153513
- Leskovan, A. C., Lanzirrotti, A., and Miller, L. M. (2009). Amyloid plaques in PSAPP mice bind less metal than plaques in human Alzheimer's disease. *Neuroimage* 47, 1215–1220. doi: 10.1016/j.neuroimage.2009.05.063
- Link, C. D. (1995). Expression of human beta-amyloid peptide in transgenic *Caenorhabditis elegans*. *Proc. Natl. Acad. Sci. U.S.A.* 92, 9368–9372. doi: 10.1073/pnas.92.20.9368
- Lovell, M. A., Robertson, J. D., Teesdale, W. J., Campbell, J. L., and Markesbery, W. R. (1998). Copper, iron and zinc in Alzheimer's disease senile plaques. *J. Neurol. Sci.* 158, 47–52. doi: 10.1016/s0022-510x(98)00092-6
- Luo, Y., Zhang, J., Liu, N., Luo, Y., and Zhao, B. (2011). Copper ions influence the toxicity of β -amyloid(1-42) in a concentration-dependent manner in a *Caenorhabditis elegans* model of Alzheimer's disease. *Sci. China Life Sci.* 54, 527–534. doi: 10.1007/s11427-011-4180-z
- Malavolta, M., Piacenza, F., Basso, A., Giacconi, R., Costarelli, L., and Mocchegiani, E. (2015). Serum copper to zinc ratio: relationship with aging and health status. *Mech. Ageing Dev.* 151, 93–100. doi: 10.1016/j.mad.2015.01.004
- McCord, M. C., and Aizenman, E. (2014). The role of intracellular zinc release in aging, oxidative stress, and Alzheimer's disease. *Front. Aging Neurosci.* 6:77. doi: 10.3389/fnagi.2014.00077
- Miller, L. M., Wang, Q., Telivala, T. P., Smith, R. J., Lanzirrotti, A., and Miklossy, J. (2006). Synchrotron-based infrared and X-ray imaging shows focalized accumulation of Cu and Zn co-localized with beta-amyloid deposits in Alzheimer's disease. *J. Struct. Biol.* 155, 30–37. doi: 10.1016/j.jsb.2005.09.004
- Minniti, A. N., Rebolledo, D. L., Grez, P. M., Fadic, R., Aldunate, R., Volitakis, I., et al. (2009). Intracellular amyloid formation in muscle cells of Abeta-transgenic *Caenorhabditis elegans*: determinants and physiological role in copper detoxification. *Mol. Neurodegener.* 4:2. doi: 10.1186/1750-1326-4-2
- Mital, M., Wezynfeld, N. E., Fraczyk, T., Wiloch, M. Z., Wawrzyniak, U. E., Bonna, A., et al. (2015). A functional role for A β in metal homeostasis? N-truncation and high-affinity copper binding. *Angew. Chem.* 54, 10460–10464. doi: 10.1002/anie.201502644
- Molina, J. A., Jiménez-Jiménez, F. J., Aguilar, M. V., Meseguer, I., Mateos-Vega, C. J., González-Muñoz, M. J., et al. (1998). Cerebrospinal fluid levels of transition metals in patients with Alzheimer's disease. *J. Neural Transm.* 105, 479–488. doi: 10.1007/s007020050071
- Myhre, O., Utkilen, H., Duale, N., Brunborg, G., and Hofer, T. (2013). Metal dyshomeostasis and inflammation in Alzheimer's and Parkinson's diseases: possible impact of environmental exposures. *Oxid. Med. Cell. Longev.* 2013:726954. doi: 10.1155/2013/726954
- Nuttall, J. R., and Oteiza, P. I. (2014). Zinc and the aging brain. *Genes Nutr.* 9:379. doi: 10.1007/s12263-013-0379-x
- Patel, R., and Aschner, M. (2021). Commonalities between copper neurotoxicity and Alzheimer's disease. *Toxics* 9:4. doi: 10.3390/toxics9010004
- Rana, M., and Sharma, A. K. (2019). Cu and Zn interactions with A β peptides: consequence of coordination on aggregation and formation of neurotoxic soluble A β oligomers. *Metallomics* 11, 64–84. doi: 10.1039/c8mt0203g
- Religa, D., Strozyk, D., Cherny, R. A., Volitakis, I., Haroutunian, V., Winblad, B., et al. (2006). Elevated cortical zinc in Alzheimer disease. *Neurology* 67, 69–75. doi: 10.1212/01.wnl.0000223644.08653.b5
- Rivers-Auty, J., Tapia, V. S., White, C. S., Daniels, M., Drinkall, S., Kennedy, P. T., et al. (2021). Zinc status alters Alzheimer's disease progression through NLRP3-dependent inflammation. *J. Neurosci.* 41, 3025–3038. doi: 10.1523/JNEUROSCI.1980-20.2020
- Saharia, K., Kumar, R., Gupta, K., Mishra, S., and Subramaniam, J. (2016). A novel way of amelioration of amyloid beta induced toxicity in *Caenorhabditis elegans*. *Ann. Neurosci.* 23, 149–154. doi: 10.1159/000449180
- Sayre, L. M., Perry, G., Harris, P. L., Liu, Y., Schubert, K. A., and Smith, M. A. (2000). *In situ* oxidative catalysis by neurofibrillary tangles and senile plaques in Alzheimer's disease: a central role for bound transition metals. *J. Neurochem.* 74, 270–279. doi: 10.1046/j.1471-4159.2000.0740270.x
- Shaye, D. D., and Greenwald, I. (2011). OrthoList: a compendium of *C. elegans* genes with human orthologs. *PLoS ONE* 6:e20085. doi: 10.1371/journal.pone.0020085
- Singh, I., Sagare, A. P., Coma, M., Perlmutter, D., Gelein, R., Bell, R. D., et al. (2013). Low levels of copper disrupt brain amyloid- β homeostasis by altering its production and clearance. *Proc. Natl. Acad. Sci. U.S.A.* 110, 14771–14776. doi: 10.1073/pnas.1302212110
- Sparks, D. L., Friedland, R., Petanceska, S., Schreurs, B. G., Shi, J., Perry, G., et al. (2006). Trace copper levels in the drinking water, but not zinc or aluminum influence CNS Alzheimer-like pathology. *J. Nutr. Health Aging* 10, 247–254.
- Squitti, R. (2012). Copper dysfunction in Alzheimer's disease: from meta-analysis of biochemical studies to new insight into genetics. *J. Trace Elements Med. Biol.* 26, 93–96. doi: 10.1016/j.jtemb.2012.04.012
- Squitti, R., Faller, P., Hureau, C., Granzotto, A., White, A. R., and Kepp, K. P. (2021). Copper imbalance in Alzheimer's disease and its link with the amyloid hypothesis: towards a combined clinical, chemical, and genetic etiology. *J. Alzheimers Dis.* 83, 23–41. doi: 10.3233/JAD-201556
- Squitti, R., Siotto, M., and Polimanti, R. (2014). Low-copper diet as a preventive strategy for Alzheimer's disease. *Neurobiol. Aging* 35(Suppl. 2), S40–S50. doi: 10.1016/j.neurobiolaging.2014.02.031
- Strausak, D., Mercer, J. F., Dieter, H. H., Stremmel, W., and Multhaup, G. (2001). Copper in disorders with neurological symptoms: Alzheimer's, Menkes, and Wilson diseases. *Brain Res. Bull.* 55, 175–185. doi: 10.1016/s0361-9230(01)00454-3
- Suh, S. W., Jensen, K. B., Jensen, M. S., Silva, D. S., Kesslak, P. J., Danscher, G., et al. (2000). Histochemically-reactive zinc in amyloid plaques, angiopathy, and

- degenerating neurons of Alzheimer's diseased brains. *Brain Res.* 852, 274–278. doi: 10.1016/s0006-8993(99)02096-x
- Vural, H., Demirin, H., Kara, Y., Eren, I., and Delibas, N. (2010). Alterations of plasma magnesium, copper, zinc, iron and selenium concentrations and some related erythrocyte antioxidant enzyme activities in patients with Alzheimer's disease. *J. Trace Elements Med. Biol.* 24, 169–173. doi: 10.1016/j.jtemb.2010.02.002
- Watt, N. T., Whitehouse, I. J., and Hooper, N. M. (2010). The role of zinc in Alzheimer's disease. *Int. J. Alzheimers Dis.* 2011:971021. doi: 10.4061/2011/971021
- Xu, J., Church, S. J., Patassini, S., Begley, P., Waldvogel, H. J., Curtis, M. A., et al. (2017). Evidence for widespread, severe brain copper deficiency in Alzheimer's dementia. *Metallomics* 9, 1106–1119. doi: 10.1039/c7mt00074j
- Yu, J., Luo, X., Xu, H., Ma, Q., Yuan, J., Li, X., et al. (2015). Identification of the key molecules involved in chronic copper exposure-aggravated memory impairment in transgenic mice of Alzheimer's disease using proteomic analysis. *J. Alzheimers Dis.* 44, 455–469. doi: 10.3233/JAD-141776

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