



## Peptide-Drug Conjugates and Their Targets in Advanced Cancer Therapies

#### Paul Hoppenz, Sylvia Els-Heindl and Annette G. Beck-Sickinger\*

Faculty of Life Sciences, Institute of Biochemistry, Leipzig University, Leipzig, Germany

Cancer became recently the leading cause of death in industrialized countries. Even though standard treatments achieve significant effects in growth inhibition and tumor elimination, they cause severe side effects as most of the applied drugs exhibit only minor selectivity for the malignant tissue. Hence, specific addressing of tumor cells without affecting healthy tissue is currently a major desire in cancer therapy. Cell surface receptors, which bind peptides are frequently overexpressed on cancer cells and can therefore be considered as promising targets for selective tumor therapy. In this review, the benefits of peptides as tumor homing agents are presented and an overview of the most commonly addressed peptide receptors is given. A special focus was set on the bombesin receptor family and the neuropeptide Y receptor family. In the second part, the specific requirements of peptide-drug conjugates (PDC) and intelligent linker structures as an essential component of PDC are outlined. Furthermore, different drug cargos are presented including classical and recent toxic agents as well as radionuclides for diagnostic and therapeutic approaches. In the last part, boron neutron capture therapy as advanced targeted cancer therapy is introduced and past and recent developments are reviewed.

#### **OPEN ACCESS**

#### Edited by:

Jennifer L. Schaefer, University of Notre Dame, United States

#### Reviewed by:

Rakesh Kumar Tiwari, Chapman University, United States Kyeongsoon Park, Chung-Ang University, South Korea Piotr A. Mroz, Indiana University, United States

#### \*Correspondence:

Annette G. Beck-Sickinger abeck-sickinger@uni-leipzig.de

#### Specialty section:

This article was submitted to Medicinal and Pharmaceutical Chemistry, a section of the journal Frontiers in Chemistry

Received: 24 March 2020 Accepted: 03 June 2020 Published: 07 July 2020

#### Citation:

Hoppenz P, Els-Heindl S and Beck-Sickinger AG (2020) Peptide-Drug Conjugates and Their Targets in Advanced Cancer Therapies. Front. Chem. 8:571. doi: 10.3389/fchem.2020.00571 Keywords: peptide, GPCR, cancer, peptide-drug conjugate, boron neutron capture therapy

## NEED FOR TARGETED CANCER THERAPY

For a long time, cardiovascular diseases were the leading cause of death among middle-aged adults globally. This changed recently in high-income countries, where cancer is now responsible for twice as many deaths as cardiovascular diseases (Dagenais et al., 2019). In 2018, cancer was responsible for 9.6 million deaths and 18.1 million new cases were diagnosed globally (Bray et al., 2018). The most frequently occurring cancer in women is breast cancer and in men, prostate cancer is most common. Notably, the leading cause of death is lung cancer in both sexes (Siegel et al., 2019). It is predicted that these numbers will rise in the future and 27.5 million new cases of cancer will be diagnosed worldwide each year by 2040 (Bray et al., 2018). This trend clearly demonstrates that novel therapeutic approaches have to be developed and efforts have to be strengthened in the future to improve the efficacy of already existing treatments.

For the last 60 years, chemotherapy remained the trademark of cancer treatment (Gilman, 1963; DeVita and Chu, 2008). In this approach, highly cytotoxic drugs are systemically administered, which seek into rapidly dividing cancer cells. However, these chemotherapeutic drugs affect also healthy cells that exhibit high proliferation rates like intestinal epithelium cells. This peripheral toxicity is the reason for frequently occurring side effects, which can vary from hair loss,

1

anemia, bruising and bleeding to neuropathy and many more (Strebhardt and Ullrich, 2008). The occurrence of undesired effects depends highly on the patient and the applied chemotherapeutic drug. This holds not only true for chemotherapeutics, but rather for any therapeutically active molecule that is administered systemically without bearing any selectivity providing features.

One possibility to reduce side effects is to waive a systemically administered drug and only apply the tumor harming treatment locally to the affected tissue. Radiation therapy can provide this local harm even though the radiation doses are delivered to all cells within the irradiation area. Nevertheless, radiation therapy remains an important curative treatment modality, with more than 50% of the patients benefiting from this treatment (Barton et al., 2006; Chen and Kuo, 2017). In principle, highenergy radiation is used to destroy cancer cells by depositing energy that damages the genetic material and prevents further cell proliferation. The type of radiation and the chosen method for delivering the energy to the malignant tissue is highly dependent on the type of tumor. Radiation can mainly be divided into two classes: X-rays and gamma-rays belong to the group of electromagnetic radiation whereas electron, neutron, proton and hole atom irradiation can be assigned to particle radiation (Gianfaldoni et al., 2017). Even though radiation therapy is often used in combination with surgery and chemotherapeutic modalities to increase the anti-tumor effect, long term cancer survivors have a higher risk for developing second malignancies because the radiation does also harm genetic material of healthy tissue (Dracham et al., 2018). Thus, the delivered dose has to be as low as possible to prevent harm to healthy tissue but it has to be effective enough to eliminate cancer cells. Various approaches were developed to increase the radiation sensitivity of tumor cells, but all of them require the administration of drugs, which selectively seek into cancer cells. This can be achieved by exploiting biochemical characteristics, which mark cancer cells differently from healthy tissues (Hanahan and Weinberg, 2011). For example, the dysregulation of translational regulators (Bhat et al., 2015), altered epigenetic regulation mechanisms (Pfister and Ashworth, 2017), the overproduction of enzymes (DeBerardinis and Chandel, 2016) or changes in the cellular microenvironment such as a lower pH (Kato et al., 2013) can be facilitated for selective therapeutic drug delivery. Moreover, cancer cells frequently overexpress cell surface proteins, which bind ligands of different nature, allowing the targeting of tumor cells by various molecule classes (Vhora et al., 2014).

The idea of targeted therapy was envisioned over 100 years ago by the German scientist Paul Ehrlich, who was awarded with the Nobel Prize in Medicine in 1908. Originally, he proposed the concept of specifically killing pathogens, which cause diseases in the body, without harming the body itself. Ehrlich named the hypothetical agent, which could facilitate this "magic bullet" (Ehrlich et al., 1956). His idea evolved over the time and his concept was transferred into cancer therapy where malignant cells have to be eliminated. Therefore, apoptosis inducing agents are combined with molecules, which selectively bind tumor cells. Although this concept seems to be quite simple, the fine-tuning is fairly elaborate and still challenging nowadays.

Various approaches have been investigated to achieve a selective delivery of different effector molecules to cancer cells. In principle, all conjugates used for selective drug delivery consist of three modules (**Figure 1**). The first component is the carrier, which facilitates tumor targeting. Besides aptamers also small molecules and biologics such as peptides, proteins and antibodies have been investigated intensively to facilitate sufficient tumor selectivity (Yoo et al., 2019). The second component is the drug itself, which can induce a variety of biological functions, but in context of cancer treatment mostly cytotoxic molecules or radionuclides are used. The third module links the former two. In dependence of the used payload, either cleavable or non-cleavable linkers can be applied to enable a controlled drug release, if



necessary. Since all three components contribute to the overall biological efficacy and selectivity, they are described separately in more detail in the following sections.

## TUMOR-HOMING AGENTS: ANTIBODIES VS. PEPTIDES

The first component is the tumor-homing molecule. Multiple classes of molecules were investigated, including small molecules (Fitzgerald et al., 2016; Kue et al., 2016; Fernández et al., 2018), aptamers (Kaur et al., 2018; Kim et al., 2018), and polysaccharides (Choi et al., 2011). Even though these molecule classes led to promising results in preclinical studies, monoclonal antibodies (mAbs) evolved as fastest-growing drug class, with more than 30 mAbs being approved and more than 100 in clinical development over the past years (Carter and Lazar, 2018). So far, five antibodydrug conjugates (ADC) have received regulatory approval of the US Food and Drug Administration (FDA) and the European Medicines Agency (EMA) (Chau et al., 2019). All of them bind to cell surface receptors such as the CD22, CD30, CD33 or the human epidermal growth factor receptor 2 (HER2), which internalizes together with the ADC. After degradation of the ADC in the lysosome, the attached cytotoxic payload is released and the desired effect is induced. Although these conjugates reached the market and proved the feasibility of this approach, antibodies have serious limitations. Most mAbs do not penetrate into tumors. With a molecular weight of about 150 kDa, they are too big to diffuse efficiently into malignant tissue (Dreher et al., 2006; Jain and Stylianopoulos, 2010). Monoclonal antibodies can be immunogenic, even when they have been humanized and they tend to aggregate in excretory organs like liver and kidneys (Borsi et al., 2002; van Schouwenburg et al., 2010; Carrasco-Triguero et al., 2013). Moreover, the generation of mAbs is very expensive as well as time-consuming and non-selective payload conjugation can lead to reduced product homogeneity (Nejadmoghaddam et al., 2019).

Most of these drawbacks can be eliminated by using smaller biomolecules like peptides. In fact, peptide-drug conjugates (PDC) comprise several advantages as carrier molecules. Peptides with up to 50 amino acids can be easily synthesized in large scale to a reasonable price and modifications as nonnatural amino acids can be directly introduced in the synthesis process (Firer and Gellerman, 2012; Mäde et al., 2014). The straightforward synthesis allows a rational optimization of side chains and backbone structures, which can result in increased binding affinities and physicochemical properties can be directly influenced (Erak et al., 2018). Furthermore, peptides are considered to be rapidly cleaved by proteolytic enzymes and quickly cleared from the blood circulation by liver and kidney. Those pharmacodynamic properties can be modulated by different modification and stabilization approaches (Vlieghe et al., 2010). One of the most known concepts of peptide stabilization is lipidation, which involves the incorporation of fatty acids into the peptide (Zhang and Bulaj, 2012). Fatty acids bind to serum albumin, preventing proteolytic cleavage in the blood by proteases and leads thereby to a prolonged circulation time (Frokjaer and Otzen, 2005). The long-acting glucagon-like peptide-1 (GLP-1) receptor agonists liraglutide (Victoza<sup>®</sup>) (Guryanov et al., 2016) and semaglutide (Ozempic<sup>®</sup>) (Marso et al., 2016), which are used to treat type-2 diabetes and obesity, are recent examples for this approach. Peptides are generally considered safe, since they feature low immunogenicity and produce non-toxic metabolites (Ahrens et al., 2012). Furthermore, their low molecular weight leads to an enhanced penetration into solid tissues resulting in better anti-tumor effects (Firer and Gellerman, 2012; Hock et al., 2015).

Peptides used in PDCs can be divided into two categories: cellpenetrating peptides (CPPs) and cell-targeting peptides (CTPs). The uptake mechanisms of CPPs across the cell membrane are not fully understood yet. Some CPPs are reported to cross the cell membrane by an energy-dependent cellular process like endocytosis- or receptor-mediated uptake, whereas others use energy-independent non-endocytic translocation pathways (Derossi et al., 1996; Thorén et al., 2005). Nevertheless, a significant increase in drug delivery was reported for small toxophores as well as for proteins being attached to CPPs like the TAT peptide, Pep-1 or Transportan (Morris et al., 2001; Muratovska and Eccles, 2004; Duong and Yung, 2013). However, extensive application of CPPs is limited due to its low cell specificity (Regberg et al., 2012).

In contrast, CTPs are ideal carrier molecules as they possess the same ability as mAbs. They bind with high affinity to overexpressed receptors on the tumor cell surface without exhibiting the disadvantages of mAbs. However, the conjugation of payloads to the peptide molecules is more critical because receptor binding and selectivity can be affected due to the steric demand of the payload, which can interfere with receptor recognition. Therefore, extensive knowledge of the interaction between peptide and receptor is needed to introduce drug cargos rationally. Amino acids like lysine, cysteine, glutamate, serine, which are not involved in the receptor recognition, can be used directly for payload conjugation at the sidechain or not required positions can be exchanged by those to introduce possible modification sites (Mäde et al., 2014). Many peptides also allow simple N-terminal modification because their N-terminus is not involved in the receptor recognition. Ideally, peptide carriers contain multiple modification sides, which allow the incorporation of multiple payloads per peptide molecule. The increased payload loading can enhance the therapeutic effects due to the increased drug concentrations at the tumor site (Dubowchik et al., 2002; Böhme et al., 2016).

# PEPTIDE RECEPTORS AS TARGETS IN CANCER THERAPY

A major challenge in the development of novel and highly effective anti-cancer drugs is the selective drug-delivery to the tumor site while healthy tissue is spared. Cell surface receptors are of high interest in the targeted cancer therapy approach as they provide the desired properties to allow selective tumor targeting (**Table 1**) (Reubi, 2003; Vhora et al., 2014). One of these requirements is the ectopical overexpression in high amounts

Targeted receptor	Tumor expression	References
Integrin (ανβ3)	Activated endothelial cells and tumor cells (such as U87MG glioblastoma cells), ovarian cancer cells	Desgrosellier and Cheresh, 2010; Chen and Chen, 2011
EGFR	Lung, breast, bladder, and ovarian cancers	Li et al., 2005; Yarden and Pines, 2012
NPY (hY1R)	Breast cancer, Ewing sarcoma	Söll et al., 2001; Körner et al., 2008; Li et al., 2015
Bn receptors (GRPR)	Lung, prostate, breast, pancreatic, head/neck, colon, uterine, ovarian, renal cell, glioblastomas, neuroblastomas, gastrointestinal carcinoids, intestinal carcinoids, and bronchial carcinoids	Jensen et al., 2008; Sancho et al., 2011
Somatostatin (SSTR2)	Small cell lung, neuroendocrine tumor, prostate cancer, breast cancer, colorectal carcinoma, gastric cancer, hepatocellular carcinoma	Volante et al., 2008; Sun and Coy, 2011
GnHR-R	Ovarian, breast, prostate, lung cancer	Schally and Nagy, 1999; Limonta et al., 2012
VIP receptors	Endocrine tumors, colon, breast cancer	Reubi, 2003; Tang et al., 2014
MC1R	Melanoma tissues	Froidevaux and Eberle, 2002; Froidevaux et al., 2005
Neurotensin receptors (NTSR1)	Breast, colon, pancreatic, lung, prostate cancer	Kokko et al., 2003; Wu et al., 2012

TABLE 1 | Overview of peptide-binding receptors studied for anti-cancer drug delivery.

on the malignant tissue to facilitate a sufficient selectivity. A tumor-to-normal-cell expression ratio of 3:1 or higher is usually desired. Secondly, the amounts of overexpressed receptors have to be sufficient to ensure drug delivery in appropriate amounts to obtain the desired therapeutic effect (Reubi, 2003; Vrettos et al., 2018). The determination of these expression levels is not trivial since tissue sampling and processing is difficult to standardize. Expression levels can vary not only within biopsies, but also over the time course of the therapy and therefore each treatment plan has to be tailored for every patient.

Even though two peptide-drug conjugates have already been approved by the FDA for treatment and diagnosis of cancer, the development of novel peptide-drug conjugates targeting overexpressed receptors is still very challenging. In the following section, comprehensively studied peptide binding receptors will be presented, which are considered as promising targets in targeted cancer therapy.

#### Integrins

A frequently used peptide carrier is the tripeptide motif arginine-glycine-aspartic acid (RGD), which was identified in the 1980s within fibronectin that was known to bind integrins (Pierschbacher and Ruoslahti, 1984; Plow et al., 2000; Kapp et al., 2017). The membrane-bound receptor family appears as heterodimers of non-covalently connected  $\alpha$ - and  $\beta$ -subunits. Integrins link the extracellular matrix with the intracellular cytoskeleton to mediate cell adhesion, migration, and proliferation (Hynes, 1987; Schwartz et al., 1995). Since all of these processes are highly relevant for carcinogenesis, the integrin  $\alpha v\beta 3$  evolved as important anti-cancer target primarily because it is associated with tumor angiogenesis and metastasis (Cox et al., 2010). It is overexpressed also in activated endothelial cells, newborn vessels, and various other tumor cells but is not found in most adult epithelial cells (Cai and Chen, 2006; Desgrosellier and Cheresh, 2010).

Targeting of these cancer types is mostly accomplished by using the cyclic RGD variant c(RGDfK) owing to its improved affinity for the integrin receptors (Ryppa et al., 2008; Chen and Chen, 2011; Gilad et al., 2016a). Recently, internalizing RGD (iRGD) conjugates gained considerable attention due to its improved vascular and tumor-tissue permeability (Sugahara et al., 2009). The sequence CRGDKGPDC, in which the cysteines at the C- and N-terminus are connected through a disulfide bond, allowed a proteolytic cleavage after integrin binding. The cleaved peptide gains affinity for neuropilin-1 (NRP-1), which is then involved in a not fully understood internalization mechanism (Sugahara et al., 2009; Kadonosono et al., 2015). However, using this peptide as carrier allows the generation of smart delivery vehicles that facilitate an active and directed uptake of the drug into tumor cells (Cho et al., 2016).

## Epidermal Growth Factor Receptor (EGFR)

Another highly interesting transmembrane protein is the epidermal growth factor receptor (EGFR), which belongs to the ErbB family. Apart from EGFR/HER1 (ErbB-1), HER2/neu (ErbB-2), HER3 (ErbB-3), and HER4 (ErbB-4) belong also to this tyrosine kinase family (Wieduwilt and Moasser, 2008). EGFR was found to be overexpressed in very high amounts in various cancer types and is associated with a strongly enhanced proliferation rate of cancer cells (Yarden and Pines, 2012). Therefore, many monoclonal antibodies and small molecule inhibitors target the EGFR in a clinical setting, inhibiting EGFR and decelerating tumor growth (Seshacharyulu et al., 2012).

EGFR is also considered as a uptake system for targeted drug delivery owing to its internalization behavior upon ligand binding and receptor dimerization (Grandal and Madshus, 2008). Several short peptides have been identified by phage display libraries (Li et al., 2005; Ai et al., 2013). For example, the GE11 peptide (YHWYGYTPQNVI) was conjugated to liposomes (Zou et al., 2018), nanoparticles (Li et al., 2019), or toxophores like doxorubicin (Fang et al., 2017).

## **G** Protein-Coupled Receptors (GPCRs)

G protein-coupled receptors form the biggest class of transmembrane proteins with more than 800 receptors. Approximately 15% of these are targeted by ca. 35% of all FDA approved drugs, demonstrating the tremendous pharmacological potential of this receptor class (Sriram and Insel, 2018). GPCRs share the common structure of seven transmembrane (TM) helices, which are linked by three intra- and three extracellular loop regions, an extracellular N-terminus, and an intracellular carboxyl-terminal domain (Rosenbaum et al., 2009). Classical



cleavable linker or just by endo-lysosomal degradation of the peptide-drug conjugate.

GPCR signaling is initiated by an extracellular ligand binding, which induces a conformational change in the receptor that initiates binding and activation of intracellular heterotrimeric G proteins ( $G\alpha_i$ ,  $G\alpha_q$ ,  $G\alpha_s$ ) (Weis and Kobilka, 2018). The physiological response depends strongly on the activated G protein. Since the induced signaling cascades control elementary processes within the cell, malfunctions in this system are often associated with cancerogenesis (Nieto Gutierrez and McDonald, 2018). In these cases, GPCRs are frequently found to be overexpressed, allowing the specific targeting of tumor cells with peptide-drug conjugates (Insel et al., 2018).

Besides the overexpression, GPCRs are known to desensitize after activation by intracellular phosphorylation mediated by GPCR kinases (GRKs). The subsequent recruitment of adaptor proteins such as arrestin induces the clathrin-mediated endocytosis (Moore et al., 2007). The entire receptor-ligand complex is internalized in this process and translocated to the lysosome. Some GPCRs are separated from the ligand in the endosome and recycle back to the cell membrane, where they can be reactivated and internalized. This mechanism can be exploited to actively accumulate drug molecules inside the cell making peptide-drug conjugates a highly interesting and suitable class of molecules as a cellular drug delivery system (Figure 2) (Vrettos et al., 2018). In this review, the gastrin-releasing peptide receptor and the human Y<sub>1</sub> receptor will be presented in more detail.

#### **NPY Receptor Family**

The human  $Y_1$  receptor (h $Y_1$ R) is a member of the neuropeptide Y (NPY) receptor family, which is comprised of three additional receptors, namely the Y<sub>2</sub> receptor (hY<sub>2</sub>R), the Y<sub>4</sub> receptor (hY<sub>4</sub>R), and the Y<sub>5</sub> receptor (hY<sub>5</sub>R) (Michel et al., 1998). The human Y receptors (hYR) are recognized and activated by the neuropeptide Y peptide hormone family including the 36-aminoacid NPY, peptide YY (PYY), and pancreatic polypeptide (PP) (Figure 3). All native peptides are composed of a flexible Nterminus, a C-terminal amphipathic  $\alpha$ -helix and an amidated C-terminus (Larhammar, 1996; Monks et al., 1996). The Y receptor/NPY hormone family forms a multi-ligand/multireceptor system in which NPY and PYY display high affinity for the hY<sub>1</sub>R, hY<sub>2</sub>R, and hY<sub>5</sub>R, whereas PP has the highest affinity for the hY<sub>4</sub>R (Pedragosa-Badia et al., 2013). However, all three peptides bind and activate all four receptors to some extent. While NPY is mainly expressed in the central nervous system (CNS) where it acts as a neurotransmitter regulating energy homeostasis and anxiety (Colmers and Bleakman, 1994;



FIGURE 3 | (A) Three-dimensional solution structure of human NPY determined by nuclear magnetic resonance spectroscopy (PDB: 1RON). Substituted amino acids in [F<sup>7</sup>,P<sup>34</sup>]NPY are indicated in blue. (B) Amino acid sequences of pNPY, hPY, hPP, and the hY<sub>1</sub>R-preferring [F<sup>7</sup>,P<sup>34</sup>]-NPY.

Wettstein et al., 1995), PYY and PP were found in peripheral tissues acting as hormones regulating the vasoconstriction (Michel and Rascher, 1995), insulin release (Wang et al., 1994), and gastrointestinal and renal epithelial secretion (Playford and Cox, 1996). The association with metabolic diseases like diabetes, obesity, hypertension, and dyslipidemia led to the development of various selective small molecules, which bind and activate only one of the Y receptors (Yulyaningsih et al., 2011; Tan et al., 2018; Yi et al., 2018). hY<sub>1</sub>R-selective molecules were particularly interesting, not only because inhibition of the hY<sub>1</sub>R leads to an anorexic effect (Brothers and Wahlestedt, 2010) but also because the hY1R was found to be present in certain tumor tissues. While ovarian sex cord-stromal tumors, nephroblastomas, gastrointestinal stromal tumors and testicular tumors were found to express the hY<sub>1</sub>R in combination with the hY<sub>2</sub>R, adrenal cortical tumors, Ewing sarcoma tumors, and renal cell carcinomas display exclusive hY1R expression (Körner and Reubi, 2007; Körner et al., 2008).

An outstanding  $hY_1R$  expression profile was identified in breast cancer by Reubi et al. (2001). He demonstrated that the  $hY_1R$  is expressed in very high density in 85% of investigated primary human breast tumors and in 100% of breast cancer-derived metastases (Reubi et al., 2001). In contrast to the previously described tumors, the non-neoplastic breast tissues showed no  $hY_1R$  expression. Instead, the  $hY_2R$  was predominantly found in non-malignant tissue. This switch in the expression profile from  $hY_2R$  in healthy tissue to  $hY_1R$  in breast tumors allows the selective targeting of these cancer cells if a  $hY_1R$ -preferring ligand is used (**Figure 3**).

The use of selective agonists is favored because agonists exploit the desensitization mechanism of the Y-receptors to deliver certain payloads to intracellular compartments. All Y receptors desensitize by clathrin-mediated endocytosis in complex with the bound ligand after activation. While the internalization process for the hY<sub>5</sub>R is relatively slow, the other Y-receptors internalize rapidly within few minutes (Gicquiaux et al., 2002). The recycling of the hY<sub>1</sub>R and hY<sub>2</sub>R back to the cell membrane has been observed by fast and slow endosomal routes (Gicquiaux et al., 2002; Mörl and Beck-Sickinger, 2015). These recycling processes are essential for an active drug accumulation inside the cell.

In order to use this drug shuttle system, many studies focused on the development of hY<sub>1</sub>R preferring agonists, which turned out to be challenging due to the great similarities between the receptors and native peptides (Zhang et al., 2011). Nevertheless, the shortened, C-terminally derived analog [Pro<sup>30</sup>, Nle<sup>31</sup>, Bpa<sup>32</sup>, Leu<sup>34</sup>]NPY(28-36) was identified as a hY<sub>1</sub>R-selective agonist in 2009 (Zwanziger et al., 2009). Another hY1R-selective agonist is the full length NPY analog [F<sup>7</sup>,P<sup>34</sup>]-NPY, which exhibits nanomolar potency at the hY1R and a highly reduced affinity for the hY<sub>2</sub>R (Söll et al., 2001). This derivative was already used in in vivo studies and serves as a first proof of concept for this receptor targeting approach. A fluorine-18 (<sup>18</sup>F)-labeled, fluoroglycosylated [F7,P34]-NPY analog was synthesized and enabled the visualization of hY1R-expressing MCF-7 tumor cells in a xenograft mice model (Hofmann et al., 2015). Furthermore, four breast tumor patients received a technetium-99m labeled [F<sup>7</sup>,P<sup>34</sup>]-NPY conjugate. While no significant peptide uptake was observed in healthy volunteers, the primary tumor in all four patients as well as the metastatic sites were clearly visualized by whole-body scintimammography (Khan et al., 2010). These studies demonstrated the tremendous potential of the  $hY_1R$  as a target in a selective drug delivery system for breast cancer.

#### **Bombesin Receptor Family**

The mammalian bombesin (Bn) receptor family consists of three GPCRs: the neuromedin B (NMB) receptor (NMBR or BB1-receptor), the gastrin-releasing peptide (GRP) receptor (GRPR or BB2-receptor) and the orphan bombesin receptor subtype 3 (BRS-3 or BB3-receptor) (Jensen et al., 2008). All three receptors are widely expressed in the CNS where they are associated with processes including satiety, thermoregulation, stress and fear responses (Roesler et al., 2006; González et al., 2008). They are also found in the gastrointestinal tract, where they are mainly involved in smooth muscle contraction and gastrin release (Uehara et al., 2011).

These receptors form together with their natural ligands a multi-ligand/multi-receptor system. While NMB binds with high affinity to the NMBR, GRP prefers the GRPR. The endogenous ligand of the BRS-3 could not be identified so far. Nevertheless, all three receptors are combined in one family because the amphibian 14-mer peptide homolog Bn (Sequence: Pyr-QRLGNQWAVGHLM-NH<sub>2</sub>), which was isolated from the skin of the European fire-bellied toad, binds and activates all three receptors (Anastasi et al., 1971; Erspamer et al., 1972). All bombesin-like peptides share two common features: their C-terminus is amidated and the last seven C-terminal amino acids are highly similar (McDonald et al., 1979; Erspamer, 1988; Kroog et al., 1995; Hellmich et al., 1997). The Bn receptors (BnR), especially the GRPR, have been extensively studied and found to be overexpressed in several human cancers including breast, colon, non-small cell lung cancer, gliomas, meningiomas, head/neck squamous cell, ovarian, pancreatic, and prostate cancers, and neuroblastomas (Table 2) (Gugger and Reubi, 1999;

**TABLE 2** | Incidence of bombesin receptor subtype expression in various human cancers (Reubi et al., 2002b).

Tumor type	n cases	Receptor incidence		
		NMBR	GRPR	BRS-3
Prostate carcinomas	12	0/12	12/12	0/12
Breast carcinomas	57	0/57	41/57	0/57
NE <sup>a</sup> GEP tumors				
Gastrinomas	5	0/5	5/5	0/5
Intestinal carcinoids	24	11/24	0/24	0/24
Thymic carcinoid	1	1/1	0/1	0/1
NE lung tumors				
Bronchial carcinoids	26	1/26	0/26	9/26
Small cell lung cancers	9	0/9	3/9	4/9
LCNEC	1	0/1	0/1	1/1
Renal cell carcinomas	16	0/16	6/16	4/16
Ewing sarcomas	10	0/10	0/10	2/10

<sup>a</sup>NE, neuroendocrine; GEP, gastroenteropancreatic; LCNEC, large cell neuroendocrine carcinoma.



Markwalder and Reubi, 1999; Reubi et al., 2002a,b; Moody et al., 2004; Pu et al., 2015; Moreno et al., 2016). Since the BnR are expressed in a number of common tumors, increasing interest rose not only to target the BnR for tumor localization and visualization but also to deliver cytotoxic agents (Schroeder et al., 2009; Sancho et al., 2011; Yu et al., 2013).

More than 200 reports were published focusing on the investigation of various Bn-like peptides conjugated primarily with radionuclides, including <sup>9m</sup>Tc, <sup>111</sup>In, <sup>67</sup>Ga, <sup>68</sup>Ga, <sup>64</sup>Cu, <sup>177</sup>Lu, <sup>90</sup>Y or <sup>213</sup>Bi, which were either used for tumor diagnostic or peptide receptor radionuclide therapy (PRRT) (Dash et al., 2015).

A number of these studies reported excellent visualization of BnR overexpressing tumors like prostate cancer in vivo as well as in humans (Scopinaro et al., 2002, 2004; van Essen et al., 2009). Thereby, radiolabeled BnR antagonists were found to be more suitable for tumor visualization applications then BnR agonists because they showed higher tumor uptake and better imaging properties (Ginj et al., 2006; Cescato et al., 2008; Mansi et al., 2013). This might be explained by better plasma stability of BnR antagonists compared to agonists, and their higher selectivity for the GRPR. In many studies the synthetic Bn peptide agonist [D-Phe<sup>6</sup>,  $\beta$ -Ala<sup>11</sup>, Phe<sup>13</sup>, Nle<sup>14</sup>]Bn(6–14) and its D-Tyr<sup>6</sup> analog were used due to their high affinity for the GRPR (Mantey et al., 1997; La Bella et al., 2002; Schroeder et al., 2009). However, the NMBR and the BRS-3 were bound with similar potencies (IC<sub>50</sub>, 0.3-2 nM) leading to off-target effects and reduced effective concentrations at the tumor side (La Bella et al., 2002). Thus, the potential tumor uptake is theoretically lower in comparison to a stable antagonist, which features comparable binding properties. Moreover, the rapid degradation of common Bn agonists in blood plasma reduces the potential uptake by tumor cells even further (Bläuenstein et al., 2004). A stable and selective GRPR-agonist could potentially feature similar or even better tumor uptake values compared to antagonists. Nevertheless, the development of GRPR-selective peptide agonists, which feature sufficient plasma stability is still challenging and was addressed only in few studies (Darker et al., 2001; Valverde et al., 2013). Recently, we could identify the peptide [D-Phe<sup>6</sup>,  $\beta$ -Ala<sup>11</sup>, NMe-Ala<sup>13</sup>, Nle<sup>14</sup>]Bn(6–14), which displays high activity at the GRPR, more than 4,000-fold selectivity and > 75% blood plasma stability after 24 h (Hoppenz et al., 2019) (**Figure 4**).

By using agonistic Bn peptides, the desensitization mechanism of the GRPR can be exploited for an active drug shuttling into the tumor cells. Following the clathrin-mediated internalization, the GRPR is separated from the endosome and recycles back to the cell membrane rapidly, where it can be activated again (Slice et al., 1998). Based on this approach, different bombesin conjugates were synthesized bearing toxophores (Stangelberger et al., 2006; Moody et al., 2007), siRNA (Rellinger et al., 2015), nanoparticles (Jafari et al., 2015), or liposomes (Accardo et al., 2012) demonstrating great results in preclinical and clinical studies.

#### Somatostatin Receptors (SSTRs)

Another very intensely studied receptor family is the somatostatin receptor (SSTR) family, which is comprised of five members (SSTR1-5). Their natural ligand is the disulfidecyclized oligopeptide somatostatin (SST), which occurs in two active isoforms, the SST-14 and SST-28 (Reisine and Bell, 1995). Both of these isoforms bind and activate the SSTRs, which are widely expressed in different tissues in the body including nervous, pituitary, kidney, lung and immune cells (Patel, 1999). The interaction of SST and its receptors not only controls the endocrine system and neurotransmission but also provides potent antisecretory and antiproliferative activities (Patel, 1999; Olias et al., 2004). Owing to these features, metabolically stabilized derivatives of somatostatin including octreotide (Sandostatin<sup>®</sup>), lanreotide, or pasireotide are directly used to treat growth hormone-producing tumors (Keskin and Yalcin, 2013). The SSTR2 and SSTR5 are thereby primarily targeted because they have been found in various neuroendocrine tumors

as well as other tumors such as breast, ovarian and lung cancer (Reubi et al., 1992; Volante et al., 2008). Besides the direct targeting of the SSTRs, peptide-drug conjugates were intensely studied to even further boost the antiproliferative effect of these peptides. For example, Paclitaxel (Shen et al., 2008), Doxorubicin (Nagy et al., 1998), and Camptothecin (Sun et al., 2008) were conjugated to octreotide analogs and provided impressive toxicities and selectivity for SSTR2 and SSTR5 expressing cells *in vivo*. Furthermore, the conjugation of <sup>111</sup>Indium to octreotide led to the first FDA approved peptide-drug conjugate (Octreoscan<sup>®</sup>), which can be used for diagnostic tumor imaging (Forssell-Aronsson et al., 2004).

## **Other Receptors**

In addition to the already introduced receptor targets, a number of other peptide receptors are also investigated as potential targets for anti-cancer drug delivery and will be covered briefly in this section.

The first example is the gonadotropin-releasing hormone receptor (GnRH-R), also known as luteinizing hormonereleasing hormone (LHRH) receptor, which is primarily expressed on gonadotrope cells in the pituitary but also found in lymphocytes, breast, ovary, and prostate (Harrison et al., 2004). The activation of the GnRH-R causes the release of folliclestimulating hormone (FSH) and luteinizing hormone (LH), which are known as gonadotropins. Over the past decade, the GnRH-R emerged as promising drug delivery systems owing to its ectopic overexpression in a variety of human tumors such as prostate, endometrial, epithelial ovarian, bladder, breast, lymphomas, and lung cancers (Halmos et al., 2000; Keller et al., 2005; Gründker et al., 2011). This receptor can be addressed by agonistic peptides or small molecules in cancer therapy (Gründker and Emons, 2017). For the generation of drug conjugates, the selective GnRH analog [D-Lys<sup>6</sup>]-GnRH-I is frequently used and reached as a doxorubicin derivative (AEZS-108) clinical phase III (Kovács et al., 2007; Yu et al., 2017).

Another example are the vasoactive intestinal peptide (VIP) receptors 1 and 2, which were found to be overexpressed in colon, breast, and endocrine tumors. The natural ligand VIP and its analogs are considered to be a valuable target for the molecular imaging of tumors and therapeutic interventions (Moody et al., 2007; Tang et al., 2014).

The melanocortin receptor 1 (MC1R) is also a highly attractive target as it has been found to be highly expressed in the majority of melanomas (Miao and Quinn, 2008). Since traditional chemotherapy treatment of metastatic melanoma is not very effective, MC1R targeting peptide-drugs are highly desired (Helmbach et al., 2001). Apart from the natural ligand  $\alpha$ -melanocyte-stimulating hormones ( $\alpha$ -MSH), the short agonistic peptide-drug conjugates (Froidevaux et al., 2005).

Furthermore, the neurotensin receptor 1 (NTSR1) was found to be overexpressed in breast, prostate, colorectal, lung, liver, and pancreas cancers among others. Since the metabolic stability of the natural ligand neurotensin was not sufficient for the use as peptide-drug conjugate, tetra-branched neurotensin peptides were synthesized and conjugated with various toxic agents such as 5-fluorodeoxyuridine (5-FdU), MTX and a chlorambucil alkylating agent (Falciani et al., 2010). Moreover, pseudoneurotensin derivatives [Lys<sup>8-9</sup>]NT(8-13) were used as carrier molecules targeting the NTSR1 (Kokko et al., 2003; Gaviglio et al., 2012).

## INTELLIGENT LINKER TECHNOLOGIES

Another very crucial aspect in the design of PDCs is the selected linker technology, which connects the carrier molecules and the payload. The linker structure has to provide distinct properties that ensure the greatest possible selectivity and biological efficacy of the payload (Figure 5). While, for example, radionuclides do not require the release after reaching the tumor cell, it is crucial for toxic payloads to be released to exploit their full potential. PDC prodrugs can fulfill this requirement by utilizing a linker structure, which controls the activity of the toxophore as long it is conjugated to the carrier peptide. As the tumor site is reached, the linker is selectively cleaved and the toxophore is released and thereby activated (Kratz et al., 2008). However, the linker must be stable during the circulation in the blood to avoid a premature release of the toxic agent, which would result in undesired peripheral toxicity. Different linker strategies have been reported for PDCs using various changes in the biological environment of cancer cells to facilitate the controlled release (Bildstein et al., 2011; Joubert et al., 2017).

One possibility to realize a selective release of toxophores is the use of pH-sensitive linkers. Since tumor cells grow and metabolize rapidly, tumor vessels are often unable to provide sufficient nutrients and oxygen. Anaerobic glycolysis inside the tumor cells produces lactic acid, which causes an acidification of the tumor tissue (pH 6.2-6.8) (Kato et al., 2013). A more significant difference in the pH-value was found in lysosomes, which features values between 4.5 and 6.0 (Ohkuma and Poole, 1978). Since agonistic PDCs are internalized into these compartments, a selective release of payloads can be achieved by using pH-sensitive linkers. The most frequently used pH-sensitive linker is a hydrazone bond, which is stable in neutral environments but easily hydrolyzed under weak acidic conditions. This concept has been demonstrated for antibodies (Ducry and Stump, 2010) as well as for peptides (Langer et al., 2001). Beside hydrazone bonds also imine, oxime, acetal, or cis-aconityl linkages were used (Chang et al., 2016).

Another approach uses redox-sensitive linkers. This is based on the high concentrations of antioxidants like glutathione (GSH) in the cytoplasm, which are 1,000-fold higher than in the plasma (15 mM intracellular vs. 15  $\mu$ M extracellular) (Meister and Anderson, 1983). The concentration of GSH in cancer cells was found to be even higher due to hypoxia, which is caused by the abnormal blood flow in tumor tissue (Bansal and Simon, 2018). Therefore, disulfide bonds have been extensively used in targeted drug delivery (Wang et al., 2012). These bonds can be readily integrated in PDCs by reaction of a cysteine-containing peptide with a thiol-including toxic agent or a prodrug (Saito et al., 2003). Recently, thioesters and azo-bond derivatives have



been investigated as cleavable linker structures (Chen et al., 2013; Medina et al., 2013).

An exquisite way to facilitate a selective linker cleavage is the use of structures, which can be cleaved by specific enzymes. For example esters and carbamates, which are either hydrolyzed by the low lysosomal pH or enzymatically by esterases and cytochrome P450 (Patterson et al., 1999; Cha et al., 2000). These structures can be introduced during peptide synthesis, but their extracellular stability needs to be carefully controlled because they are prone to hydrolysis in serum (Coin et al., 2007). However, great interest was gained by using short amino acid sequences as linkers, which are recognized explicitly by proteases being overexpressed in tumor tissues. Peptide bonds in these linkers are stable during circulation in plasma as the activity



of the exploited protease is reduced due to the unfavorably high pH in plasma (Ciechanover, 2005). A wide range of short peptide sequences have been identified, including the short Val-Cit (Szlachcic et al., 2016) and Gly-Phe-Leu-Gly (GFLG) linkers (Naqvi et al., 2010), which are cleaved by the protease cathepsin B. Another very interesting protease is legumain because it is the only asparaginyl endopeptidase in mammals featuring very high substrate specificity for the amino acid sequence Ala-Ala-Asn (Chen et al., 1997; Dall and Brandstetter, 2016). The recognition sequence is cleaved C-terminally after Asn allowing a traceless release of the attached payload if the cargo is conjugated to the C-terminus, while the N-terminus of the linker is coupled to the carrier (Stern et al., 2009; Bajjuri et al., 2011; Mai et al., 2017). Therefore, enzymatically cleavable linkers are ideal structures to utilize a controlled release of toxic payloads within tumor cells especially because legumain and cathepsin B were found to be overexpressed in solid tumors (Liu et al., 2003; Gondi and Rao, 2013).

## PAYLOADS IN TARGETED CANCER THERAPY

The heart of every PDC is the drug cargo. While the peptide carrier and the linker structure provide the selectivity of the PDC, the drug is the component that facilitates the actual purpose of the conjugates. In many cases, the term "drug" refers to cytotoxic (chemotherapeutic) anti-cancer agents, but a broad spectrum of therapeutically active moieties can be meant. Conjugation of radionuclide complexes to peptides can result in molecules, which can be used for cancer diagnostics or PRRT. Moreover, non-radioactive molecules like boron can be used as cargo to generate PDCs for boron neutron capture therapy (BNCT).

## **Chemotherapeutic Agents**

Currently, more than 250 FDA approved drugs (Cancer.gov) are used to treat malignant cancers. Many of these classical chemotherapeutic drugs feature strong pharmacological activity owing to their great cytotoxicity. However, a disadvantage of these small molecules is their uncontrolled toxicity, which results in severe side effects. By attaching these molecules to targetingmoieties like peptides can enhance their pharmacokinetic and increase the therapeutic window of the parent cytotoxic agent (Chen et al., 2014). The selected drugs must comply with certain design principles in order to serve as suitable compounds for the generation of a PDC. Most importantly, intrinsic functional groups have to be available to enable the attachment to the peptide carrier by a cleavable linker. Fortunately, many toxic agents provide hydroxy, carboxy, or amine groups that can be used (Figure 6). In some cases, these groups are necessary for their biological response, enabling the generation of inactive prodrugs, which have to be released without a trace to facilitate their action at the tumor side (Kratz et al., 2008). Furthermore, the drug has to be chosen to exhibit sufficient and potent cytotoxicity vs. the malignant cells, as drug resistance mechanisms of cancer cells are frequently found (Housman et al., 2014).

Chemotherapeutic agents used in PDCs can be classified by their general mode of action (Malhotra and Perry, 2003). The

first group of molecules bind and interact with the cellular DNA or DNA-protein complexes. Thus, the transcription and DNA replication are disturbed, leading to the induction of apoptosis. Highly potent molecules including metal complexes (Weidmann et al., 2014), camptothecin (CPT) (Wall and Wani, 1996), and anthracyclines like daunorubicin (DAU) or doxorubicin (DOX) (Minotti et al., 2004) are broadly applied in PDCs (Figure 6). For example, the SSTR2 targeting octreotide was conjugated to DOX or 2-pyrrolino-DOX by ester linkers and exhibited selective toxicity against receptor-expressing tumors in vivo (Nagy et al., 1998; Huang et al., 2000; Engel et al., 2005; Shen et al., 2008; Seitz et al., 2009). Moreover, the integrin-targeting cyclic peptides c(RGDfK) and c(RGDfS) were conjugated to CPT and the alkylating agent chlorambucil by a carbamate and ester linkage, demonstrating growth inhibition in cancer cell lines expressing the integrin αvβ3 (Gilad et al., 2016b). In addition, CPT was coupled to the Bn analog [D-Tyr<sup>6</sup>, β-Ala<sup>11</sup>, D-Phe<sup>13</sup>, Nle<sup>14</sup>]Bn(6-14) by a carbamate linker exhibiting great cytotoxicity in Bnreceptor overexpressing cells while cells without these receptors were not affected (Moody et al., 2006). Until now, the most progressed receptor-targeting PDC for selective chemotherapy is Zoptarelin Doxorubicin (AN-152, AEZS-108, Zoptrex<sup>TM</sup>), which is composed of a GnRH analog and doxorubicin conjugated through an ester bond with a glutaric acid spacer (Figure 7) (Nagy et al., 1996). This derivative reached clinical phase 3 for the treatment of endometrial cancer, and the results were disclosed in May 2017. Even though very encouraging results were obtained before, Zoptarelin Doxorubicin neither extended overall survival nor improved the safety profile compared to the classical chemotherapy with DOX (ZoptEC, 2013).

The second class of frequently used toxophores convey their cytotoxicity by inhibiting the DNA-biosynthesis. These antimetabolites, for example the nucleoside analog of deoxycytidine, gemcitabine (Galmarini et al., 2002), or the folate derivative methotrexate (MTX), which inhibits the enzyme dihydrofolate reductase (Chan and Cronstein, 2013) demonstrated great potency on cancer cells. Notably, MTX was specifically delivered to tumor cells by exploiting the hY<sub>1</sub>R-preferring peptide [F<sup>7</sup>,P<sup>34</sup>]-NPY. The modified conjugates [K<sup>4</sup>(GFLG-MTX),F<sup>7</sup>,P<sup>34</sup>]-NPY displayed high extracellular stability, paired with a fast uptake into cancer cells and the rapid release of MTX led to high cytotoxicity values in hY1R expressing tumor cells (Böhme and Beck-Sickinger, 2015). This potent cytotoxic effect was even further increased in MDA-MB-468 breast cancer cells by generating a double modified [K<sup>4</sup>(GFLG-MTX),F<sup>7</sup>,K<sup>18</sup>(GFLG-MTX),P<sup>34</sup>]-NPY conjugate (Böhme et al., 2016).

The third group of chemotherapeutics is formed by antimitotic agents, which act on microtubules. Drugs like paclitaxel (PTX), which inhibit microtubule depolymerization (Xiao et al., 2006), and vinca alkaloid analogs, which inhibit tubulin polymerization (Zhou and Rahmani, 1992), belong to this group. Even though PTX is highly hydrophobic and often associated with multidrug resistance owing to the P-glycoprotein-mediated efflux (Vargas et al., 2014; Barbuti and Chen, 2015), a PTX containing PDC reached phase 3 in clinical trials. The bloodbrain barrier (BBB)-penetrating peptide angiopep-2 was loaded

with three PTX molecules by ester linkers (ANG1005) (Régina et al., 2008). This highly loaded conjugate demonstrated higher brain uptake than free PTX and significant antitumor activity in vivo in glioblastoma-bearing mice (Régina et al., 2008). This conjugate also reached therapeutic concentrations in the tumor site and was well-tolerated in humans (Drappatz et al., 2013). Although no results of phase 2 were published yet, a phase 3 study was started in December 2018 (ANGLeD, 2018). Another very promising group of molecules is formed by kinesin spindle protein (KSP/Eg5/KIF11) inhibitors (KSPi), which affect the spindle formation during mitosis. When this process is impaired, cell cycle arrest is induced, which leads subsequently to apoptosis. Since these KSPis were found to be highly potent in various cancer types (Knight and Parrish, 2008; El-Nassan, 2013) their specificity has to be improved to transfer them into a clinical setting (Wakui et al., 2014). So far, only KSPi conjugated to antibodies were generated, which demonstrated high potency in vitro and in vivo (Lerchen et al., 2019).

## Radionuclides

Among chemotherapeutic drugs, radionuclides are the second major group of payloads in PDCs. They can be used for two main purposes related to cancer. The first field is their application in cancer diagnosis. Therefore, PDCs are labeled either with positron-emitting radioisotopes such as fluorine-18 (<sup>18</sup>F), copper-64 (<sup>64</sup>Cu), and gallium-68 (<sup>68</sup>Ga) to generate PET imaging agents or gamma-emitting radioisotopes including technetium-99m (99m Tc) and iodine-123 (123I), which can be used in single-photon emission computed tomography (SPECT). By binding to the targeted receptors on tumor cells, the malignant tissue can be precisely localized. The incorporation into the peptides is mostly facilitated by bifunctional chelating agents (BFCA), which provide the chelating group for the incorporation of the radiometals and a functional group allowing the conjugation to the peptide (Liu, 2008; Jamous et al., 2013). The most frequently applied BFCAs are diethylenetriaminepentaacetic acid (DTPA) and 1,4,7,10-tetraazacyclododecane-1,4,7,10-tetraacetic acid (DOTA) enabling the generation of PCDs with various radiometals and -lanthanides (León-Rodríguezn et al., 2008; Liu, 2008). Many reviews focused on the use of PDCs as molecular imaging agents and a huge variety of PDCs including a <sup>68</sup>Galabeled bombesin analog (RM2), [18F]Galacto-RGD, 99mTclabeled [Phe7,Pro34]NPY and many more were investigated in a clinical setting (Schottelius and Wester, 2009; Sun et al., 2017).

Besides the diagnostic approach, radionuclide-labeled PDCs can also be seen in a therapeutic setting, if  $\beta$ -emitting, Auger electron-emitting or  $\alpha$ -emitting nuclides are used (Karagiannis, 2007; Bhattacharyya and Dixit, 2011). The peptide receptor radionuclide therapy (PRRT) allows a directed and tissue-specific irradiation of tumor cells based on their overexpressed receptors. Many different radioactive metals are naturally or synthetically available and each of them features different properties. Therefore, the choice of the radionuclide strongly depends on the size of the targeted tumor or metastases (O'Donoghue et al., 1995).



The most commonly used radionuclides are indium-111 (<sup>111</sup>In), yttrium-90 (<sup>90</sup>Y), and lutetium-177 (<sup>177</sup>Lu) (Thundimadathil, 2012). While <sup>90</sup>Y emits  $\beta$ -radiation with tissue penetration depths of up to 11 mm, <sup>177</sup>Lu exhibits medium energy  $\beta$ -particles, which allow a tissue penetration range of max. 3 mm. Therefore, the use of <sup>90</sup>Y is preferred for pronounced tumors due to the potential "cross-fire" effects, which can compensate for frequently occurring receptor heterogeneity (Goffredo et al., 2011). Nevertheless,  $^{177}Lu$  was found to eradicate small metastases better than  $^{90}Y$  because  $^{177}Lu$  emits besides  $\beta$ -particles also low-energy Auger electrons and conversion electrons (CEs) that deposit their dose over a short distance (Michel et al., 2005; Hindié et al., 2016). Another advantage of

<sup>177</sup>Lu is its low abundant gamma radiation, which enables post-therapeutic dosimetry.

These features are also provided by <sup>111</sup>In, which is part of the first FDA approved radiopharmaceutical peptide-drug conjugate <sup>111</sup>In-DTPA-octreotide (Octreoscan<sup>®</sup>) (Figure 7) (Kwekkeboom et al., 2010). Even though the use of high doses of  $Octreoscan^{\mathbb{R}}$ in patients with metastasized neuroendocrine tumors led to symptom relief, the tumor size regression was unsatisfactory (Ochakovskava et al., 2001; Anthony et al., 2002). Therefore, Octreoscan<sup>®</sup> is only approved for diagnostic imaging of SSTRpositive tumors. To increase the biological effect of Auger electron-emitting isotopes, a delivery close to the nucleus has to be ensured. However, the most successful therapeutic peptide radiopharmaceutical so far is <sup>177</sup>Lu-DOTA-TATE. This conjugate has been investigated intensively as PRRT-agent in several clinical centers in Europe (Hirmas et al., 2018). Treatment with <sup>177</sup>Lu-DOTA-TATE resulted in longer progression-free survival and a significantly higher response rate than highdose octreotide long-acting repeatable among patients with advanced midgut neuroendocrine tumors (Strosberg et al., 2017). The European Commission approved <sup>177</sup>Lu-DOTA-TATE (Lutathera<sup>®</sup>) in October 2017, followed by the FDA in January 2018. This development demonstrates the enormous potential of the receptor-mediated drug delivery approach in cancer and is considered as a major milestone on the way toward peptide-based and personalized medicine.

## **BORON NEUTRON CAPTURE THERAPY**

Over the past decades, different approaches of targeted cancer therapy have been developed and reached the clinics. Even though these therapy approaches demonstrated excellent antitumor efficacy, the expected reduction in severe side effects was only partially achieved. Especially antibody-drug conjugates that target specific cell surface receptors display side effects including fever, nausea, infection, vomiting, stomatitis, and skin rashes (Hansel et al., 2010; Donaghy, 2016). These undesired unspecific effects are frequently mediated by the carrier molecules, which deliver their payload not only to the addressed tumor site but also to healthy tissues, which express the targeted markers as well. One possibility to overcome this issue is the use of nontoxic payloads, which have to be converted into their active form within the tumor cells. If the second step is also tumor-selective, the toxic payload will be generated only in tumor cells, whereas all healthy cells are not harmed because they contain only the inactive payload. This principle is used in a binary radiotherapy approach called boron neutron capture therapy (BNCT).

## Principles and General Requirements of BNCT

The idea of BNCT was already described by Locher (1936). In principle, BNCT offers the possibility to combine molecular drug targeting with the regional beam positioning of radiation therapy to achieve a double-selective therapeutic effect. This binary approach requires the accumulation of the non-radioactive isotope <sup>10</sup>B, which comprises 19.9% of the naturally occurring





boron in tumor cells. The subsequent local irradiation with thermal neutrons or epithermal neutrons (0.025 eV) induces the nuclear fission reaction [ $^{10}B(1n,\alpha)^7$ Li] resulting in an excited [ $^{11}B$ ]\* nuclei, which decays immediately in high linear energy transfer (LET) particles (Locher, 1936). Due to the limited path lengths of the generated  $\alpha$  and  $^7$ Li nuclei in tissues (5–8 µm), the destructive effects of those are limited to boron-containing cells (**Figure 8**) (Barth et al., 2005).

In order to achieve sufficient biological effects, the boron delivery agents must fulfill several critical requirements. (a) The uptake by tumor tissue has to be highly specific. To ensure no harm for non-neoplastic tissues, tumor to normal tissue and tumor to blood ratios of 3:1 and 5:1 are required, respectively. (b) Even though <sup>10</sup>B features a remarkably high neutron capture cross-section of 3,840 barns, which is more than 3,000-fold higher than the neutron capture cross-section of <sup>14</sup>N (1.82 barn) and <sup>1</sup>H (0.332 barn) (Schmitt et al., 1960), a boron amount of  $20 \,\mu g/g$  tumor is needed to achieve sufficient biological effects. This amount corresponds to about 10<sup>9</sup> <sup>10</sup>B atoms per cell, which could be lower if the boron is concentrated near or in the nucleus (Hawthorne, 1993; Hartman and Carlsson, 1994; Soloway et al., 1998). (c) The boron delivery agents have to retain within the tumor tissue over the period of neutron irradiation. (d) Simultaneously, the drug has to be rapidly cleared from the blood and healthy tissue. (e) The <sup>10</sup>B-loaded agents must be chemically and metabolically stable and (f) must not feature any systemic cytotoxicity. (g) Moreover, they should have an appropriate water solubility.

Each of these demanding perquisites and especially their combination challenge the development of suitable boron delivery agents, limiting the establishment of BNCT as a viable cancer treatment modality in the clinic so far (Kreiner et al., 2016).

## Past and Recently Developed Boron Delivery Agents

After the first efforts, during the 1940s and 1950s, using the simplest boron salts including disodium tetraborate and sodium pentaborate, it became clear that a selective and high boron accumulation in tumor cells is crucial for an application in BNCT (Farr et al., 1958). Around the 1960s, L-boronophenylalanine (BPA) (Snyder et al., 1958) and sodium mercaptoundecahydro*closo*-dodecaborate (BSH) (Soloway et al., 1967) were found to accumulate in the desired tissues in sufficient amount (**Figure 9A**).

BSH is a small hydrophilic molecule and has the advantage over BPA to deliver 11 boron atoms per molecule to the tumor. Although BSH has been applied for the treatment of glioblastomas demonstrating no toxic effects prior to irradiation, the efficacy has been limited due to low tumor boron concentrations (Kageji et al., 1997). To increase the boron concentration, various studies have been conducted in combination with other boron compounds (Barth et al., 2012).

Nowadays, BPA is the most applied drug in BNCT and is actively taken up into tumor cells by the L-type amino acid transporter system, which is highly expressed in tumor cells (Wittig et al., 2000). This results in higher boron concentrations within the tumor compared to BSH, which accumulates only passively in tumor tissue and cannot cross the cell membrane due to its net charge (Sköld et al., 2010). However, BPA has very limited water solubility (1.6 g/L) requiring the formulation as fructose complex, which demonstrated favorable biodistribution properties (Mori et al., 1989; Coderre et al., 1997). Even though BPA is considered to be a better boron delivery agent than BSH, it features some critical disadvantages. BPA consists of only a single boron atom per molecule requiring the administered of very high doses (500 mg/kg) and the rather modest tumor selectivity results in suboptimal BNCT efficiency (Luderer et al., 2015). Despite some promising results achieved by treating patients with recurrent head and neck cancer, high-grade gliomas and advanced melanomas, the development of novel boron delivery agents would lift this technology to the next level and support the implementation as advanced cancer treatment in clinics (Barth et al., 2018b). Although linear accelerator-based neutron beams have been developed, allowing the generation of neutrons in hospitals, boron delivery agents with high boron loading and excellent selectivity are still needed (Suzuki, 2019).

Multiple molecule classes have been investigated to find novel BNCT agents including low molecular weight compounds such as boronated nucleosides, amino acids, sugars, and porphyrin derivatives. Moreover, huge polymers like polyanionic and cationic polymers and polyamines, as well as lysosomes and nanoparticles filled with boron were designed to facilitate extensive boron loading (Luderer et al., 2015; Barth et al., 2018a). To increase the selectivity of these polymers, biomolecules including peptides, proteins, and antibodies were attached to these structures allowing the directed targeting of cancer cells. EGFR has been studied extensively as a target in BNCT owing to its elevated expression in gliomas. Heavily boronated polyamidoamine dendrimers (around 1,000 boron atoms) were linked to the natural ligand EGF (Capala et al., 1996; Yang et al., 2009a) and the monoclonal antibodies Cetuximab (Wu et al., 2004, 2007) and L8A4 (Yang et al., 2006, 2008). In vivo studies of the latter two revealed that the combination of both antibodies is recommended to achieve a reasonable increase in mean survival times after irradiation in rats bearing tumors expressing both isoforms (EGFR and ERGFRvIII) of the EGFR (Yang et al., 2009b).

Besides EGF, peptides have also been investigated as potential boron delivery systems. In a number of studies, carboranes were used due to their favored properties. These icosahedral, hydrophobic  $C_2B_{10}H_{12}$  clusters have a high boron content, while they occupy a rather small amount of space, slightly larger than a rotating phenyl ring (Scholz and Hey-Hawkins, 2011). Three different isomers exist, which are defined by the position of the carbon atoms in the cluster (**Figure 9B**). Moreover, they can be chemically functionalized to readily allow the facile conjugation to peptides by peptide chemistry and they display high biological stability and relatively low cytotoxicity (Valliant et al., 2002).

In 2011, dimeric integrin targeting PDCs were designed (Kimura et al., 2011), which featured *ortho*-carboranes as center and two cyclic c(RGDfK) peptides that were separately conjugated via a butanoic acid linker to each carbon atom in the cluster (**Figure 10A**). The resulting conjugate, named GPU-201, displayed a high integrin  $\alpha\nu\beta$ 3 binding affinity, dose-dependent tumor uptake in squamous cell carcinoma (SCCVII)-bearing mice and longer tumor retention time than BSH. Even though it showed a stronger tumor growth inhibition after irradiation compared to BSH alone, GPU-201 also demonstrated higher toxicity than BSH to both proliferating and quiescent cancer cells without irradiation (Masunaga et al., 2012).



The first boron delivery agent targeting cancer cells by their overexpressed GPCRs was proposed in 2003 (Schirrmacher et al., 2003). They used the SSTR2-addressing [Tyr<sup>3</sup>]-octreotate (TATE), which was N-terminally modified with the *ortho*carborane derivative 5,6-dicarba-*closo*-dodecaboranyl hexynoic acid. Subsequently, they attached a BSH via Michael-addition to the N-terminus of TATE, both derivatives showed no biological activity (Mier et al., 2004). Four years later, Betzel et al. succeeded in creating the first biologically active and boronated TATE analogs by using the ortho-carborane-containing building block 4-(O-methylencarboranyl)-benzoic acid (BBB1), which was coupled either directly to the N-terminus or by glycinylsarcosine spacers of different length (**Figure 10B**) (Betzel et al., 2008). Even though double carborane modified TATE conjugates were generated with suitable SSTR2 affinity, further pharmaceutical development of SSTR2-targeting, boronated peptides were not undertaken.

In addition to these PDCs, the hY<sub>1</sub>R preferring peptide  $[F^7, P^{34}]$ -NPY has been recently described as a suitable target in BNCT. An *ortho*-carboranyl propionic acid (Cpa)-containing amino acid Fmoc-Lys-N<sub> $\varepsilon$ </sub>(Cpa)-OH was incorporated at position 4 by solid-phase peptide synthesis (Ahrens et al., 2011). The modification led only to a slight loss of activity at the hY<sub>1</sub>R, but the hYR subtype-specific internalization was maintained. These results led to the development of the *ortho*-carborane building block 9-(carboxymethylthio)-1,2-dicarba-*closo*-dodecaborane(12) and a deoxygalactosyl-functionalized, charge-compensated cobalt bis(dicarbollide) building block,



which were introduced at position 4, 18, or 22 of  $[F^7, P^{34}]NPY$ by the side-chain of substituted lysines at the respective positions (Ahrens et al., 2015; Frank et al., 2016). A triple modified conjugate with 30 boron atoms per peptide molecule demonstrated nanomolar potency at the hY<sub>1</sub>R and selectivity against other hYR subtypes and the sufficient uptake into hY<sub>1</sub>R transfected HEK293 cells was proven. Recent studies showed that introduction of sugar moieties to heavily carborane-loaded hY<sub>1</sub>R selective analogs can enhance solubility and thereby enable the boron conjugation of up to 80 boron atoms (Worm et al., 2019) (**Figure 11**).

The concept of boron-containing peptides was also investigated for ghrelin analogs and GRPR analogs. A ghrelin receptor superagonist was modified with different carborane monoclusters and the meta-carborane building block with a mercaptoacetic acid linker demonstrated the best results (Worm et al., 2018). The GRPR-selective ligand [D-Phe<sup>6</sup>,  $\beta$ -Ala<sup>11</sup>, Ala<sup>13</sup>, Nle<sup>14</sup>]Bn(6–14) contained multiple bis-deoxygalactosyl-carborane building blocks, which allowed the generation of a peptide conjugate with up to 80 boron atoms per molecule (Hoppenz et al., 2020). These modifications did not influence

receptor activation but metabolic stability was increased and no intrinsic cytotoxicity was observed. Notably, undesired uptake into liver cells was suppressed by using L-deoxygalactosyl modified carborane building blocks.

Three studies reported the design and synthesis of CPPs modified with boron clusters. Conjugation of eight BSH clusters to the 11R peptide (8BSH-11R) facilitated the delivery of high amounts of boron (around 5,000 ppm/10<sup>6</sup> cells) into U87 $\Delta$ EGFR glioma cells (Michiue et al., 2014). In comparison to BSH, 8BSH-11R resulted in significantly stronger growth inhibition of U87 $\Delta$ EGFR glioma cells after neutron irradiation. In addition, accumulation of 8BSH-11R in implanted cells in mice was observed, while no uptake in healthy brain tissue was demonstrated.

In another study, an arginine-tripeptide (3R) was modified with BSH and a DOTA chelator to allow <sup>64</sup>Cu-radiolabeling and uptake quantification in glioma-bearing mice by PET imaging. For the labeled 3R conjugate, significantly higher tumor-tonormal-brain and tumor-to-blood radioisotope accumulation ratios compared to BSH-DOTA-64Cu were observed (Iguchi et al., 2015). Moreover, the mitochondrial targeting peptides KLA and RLA peptides were conjugated to dodecaborates (DB) in a very recent study (**Figure 10C**) (Nakase et al., 2019). It was shown that DB-RLA reached higher boron concentrations in C6 glioblastoma cells than BSH and the cell mortality rate was significantly higher than for BSH after neutron irradiation, for which no cytotoxicity was observed. The number of the conducted studies and the variety of applied approaches demonstrate the pronounced interest in developing suitable boron delivery agents.

## CONCLUSION

Today, targeted delivery of anticancer agents remains one of the most appealing methods for cancer treatment. Peptide receptors are promising targets owing to their high overexpression in various tumor types. The synthetic analogs of natural peptide ligands are of major interest for PDCs as they possess high target affinity and specificity, fast internalization rates, and low immunogenicity. Previous limitations including low *in vivo* stability can be overcome by peptide modification and

therapeutic cargos can be readily introduced to the peptides. PDCs have to compete with recently developed antibody-drug conjugates, but by now, they are rapidly catching up. More and

### REFERENCES

- Accardo, A., Salsano, G., Morisco, A., Aurilio, M., Parisi, A., Maione, F., et al. (2012). Peptide-modified liposomes for selective targeting of bombesin receptors overexpressed by cancer cells: a potential theranostic agent. *Int. J. Nanomed.* 7, 2007–2017. doi: 10.2147/IJN.S29242
- Ahrens, V. M., Bellmann-Sickert, K., and Beck-Sickinger, A. G. (2012). Peptides and peptide conjugates: therapeutics on the upward path. *Future Med. Chem.* 4, 1567–1586. doi: 10.4155/fmc.12.76
- Ahrens, V. M., Frank, R., Boehnke, S., Schütz, C. L., Hampel, G., Iffland, D. S., et al. (2015). Receptor-mediated uptake of boron-rich neuropeptide y analogues for boron neutron capture therapy. *ChemMedChem* 10, 164–172. doi: 10.1002/cmdc.201402368
- Ahrens, V. M., Frank, R., Stadlbauer, S., Beck-Sickinger, A. G., and Hey-Hawkins, E. (2011). Incorporation of ortho-carbaboranyl-Nepsilon-modified L-lysine into neuropeptide Y receptor Y1- and Y2-selective analogues. *J. Med. Chem.* 54, 2368–2377. doi: 10.1021/jm101514m
- Ai, S., Jia, T., Ai, W., Duan, J., Liu, Y., Chen, J., et al. (2013). Targeted delivery of doxorubicin through conjugation with EGF receptor-binding peptide overcomes drug resistance in human colon cancer cells. *Br. J. Pharmacol.* 168, 1719–1735. doi: 10.1111/bph.12055
- Anastasi, A., Erspamer, V., and Bucci, M. (1971). Isolation and structure of bombesin and alytesin, two analogous active peptides from the skin of the european amphibiansBombina and Alytes. *Experientia* 27, 166–167. doi: 10.1007/BF02145873
- ANGLED (2018). ANG1005 in Leptomeningeal Disease From Breast Cancer (ANGLED): Identifier: NCT03613181. Available online at: https://clinicaltrials. gov (accessed October 21, 2019).
- Anthony, L. B., Woltering, E. A., Espenan, G. D., Cronin, M. D., Maloney, T. J., and McCarthy, K. E. (2002). Indium-111-pentetreotide prolongs survival in gastroenteropancreatic malignancies. *Semin. Nucl. Med.* 32, 123–132. doi: 10.1053/snuc.2002.31769
- Bajjuri, K. M., Liu, Y., Liu, C., and Sinha, S. C. (2011). The legumain proteaseactivated auristatin prodrugs suppress tumor growth and metastasis without toxicity. *ChemMedChem* 6, 54–59. doi: 10.1002/cmdc.201000478

more conjugates containing classical chemotherapeutic agents progress into clinical studies and in January 2018, <sup>177</sup>Lu-DOTA-[Tyr<sup>3</sup>]-octreotate was approved by the FDA as the first PDC used in a therapeutic setting. This demonstrates the applicability of peptide-drug conjugates in future cancer treatment. Notably, the full potential of this approach is not exploited yet and PDC of the next generations are already in the pipeline. PDCs with intelligent linkers or non-radioactive payloads such as <sup>10</sup>boron might not only provide even better tumor selectivity but also higher efficacy. Therefore, the approach of peptide-based and receptor-mediated drug delivery will give new impulses to cancer therapies in the future.

## **AUTHOR CONTRIBUTIONS**

AB-S initiated the project. PH searched the data and wrote the manuscript. SE-H and AB-S revised the manuscript. All authors contributed to the article and approved the submitted version.

## FUNDING

The authors acknowledge support from the German Research Foundation (DFG) and Universität Leipzig within the program of Open Access Publishing.

- Bansal, A., and Simon, M. C. (2018). Glutathione metabolism in cancer progression and treatment resistance. J. Cell Biol. 217, 2291–2298. doi: 10.1083/jcb.201804161
- Barbuti, A. M., and Chen, Z.-S. (2015). Paclitaxel through the ages of anticancer therapy: exploring its role in chemoresistance and radiation therapy. *Cancers* 7, 2360–2371. doi: 10.3390/cancers7040897
- Barth, R. F., Coderre, J. A., Vicente, M. G. H., and Blue, T. E. (2005). Boron neutron capture therapy of cancer: current status and future prospects. *Clin. Cancer Res.* 11, 3987–4002. doi: 10.1158/1078-0432.CCR-05-0035
- Barth, R. F., Mi, P., and Yang, W. (2018a). Boron delivery agents for neutron capture therapy of cancer. *Cancer Commun.* 38:35. doi: 10.1186/s40880-018-0299-7
- Barth, R. F., Vicente, M. G. H., Harling, O. K., Kiger, W. S. III, Riley, K. J., et al. (2012). Current status of boron neutron capture therapy of high grade gliomas and recurrent head and neck cancer. *Radiat. Oncol.* 7:146. doi: 10.1186/1748-717X-7-146
- Barth, R. F., Zhang, Z., and Liu, T. (2018b). A realistic appraisal of boron neutron capture therapy as a cancer treatment modality. *Cancer Commun.* 38:36. doi: 10.1186/s40880-018-0280-5
- Barton, M. B., Frommer, M., and Shafiq, J. (2006). Role of radiotherapy in cancer control in low-income and middle-income countries. *Lancet Oncol.* 7, 584–595. doi: 10.1016/S1470-2045(06)70759-8
- Betzel, T., Hess, T., Waser, B., Reubi, J. C., and Roesch, F. (2008). Closo-borane conjugated regulatory peptides retain high biological affinity: synthesis of closoborane conjugated Tyr(3)-octreotate derivatives for BNCT. *Bioconjug. Chem.* 19, 1796–1802. doi: 10.1021/bc800101h
- Bhat, M., Robichaud, N., Hulea, L., Sonenberg, N., Pelletier, J., and Topisirovic, I. (2015). Targeting the translation machinery in cancer. *Nat. Rev. Drug Discov.* 14, 261–278. doi: 10.1038/nrd4505
- Bhattacharyya, S., and Dixit, M. (2011). Metallic radionuclides in the development of diagnostic and therapeutic radiopharmaceuticals. *Dalton Trans.* 40, 6112–6128. doi: 10.1039/c1dt10379b
- Bildstein, L., Dubernet, C., and Couvreur, P. (2011). Prodrug-based intracellular delivery of anticancer agents. Adv. Drug Deliv. Rev. 63, 3–23. doi: 10.1016/j.addr.2010.12.005

- Bläuenstein, P., Garayoa, E. G., Rüegg, D., Blanc, A., Tourw,é, D., Beck-Sickinger, A. G., et al. (2004). Improving the tumor uptake of 99mTc-labeled neuropeptides using stabilized peptide analogues. *Cancer Biother. Radiopharm.* 19, 181–188. doi: 10.1089/108497804323071959
- Böhme, D., and Beck-Sickinger, A. G. (2015). Controlling toxicity of peptide-drug conjugates by different chemical linker structures. *ChemMedChem* 10, 804–814. doi: 10.1002/cmdc.201402514
- Böhme, D., Krieghoff, J., and Beck-Sickinger, A. G. (2016). Double methotrexatemodified neuropeptide Y analogues express increased toxicity and overcome drug resistance in breast cancer cells. *J. Med. Chem* 59, 3409–3417. doi: 10.1021/acs.jmedchem.6b00043
- Borsi, L., Balza, E., Bestagno, M., Castellani, P., Carnemolla, B., Biro, A., et al. (2002). Selective targeting of tumoral vasculature: comparison of different formats of an antibody (L19) to the ED-B domain of fibronectin. *Int. J. Cancer* 102, 75–85. doi: 10.1002/ijc.10662
- Bray, F., Ferlay, J., Soerjomataram, I., Siegel, R. L., Torre, L. A., and Jemal, A. (2018). Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J. Clin.* 68, 394–424. doi: 10.3322/caac.21492
- Brothers, S. P., and Wahlestedt, C. (2010). Therapeutic potential of neuropeptide Y (NPY) receptor ligands. *EMBO Mol. Med.* 2, 429–439. doi: 10.1002/emmm.201000100
- Cai, W., and Chen, X. (2006). Anti-angiogenic cancer therapy based on integrin  $\alpha\nu\beta3$  antagonism. ACAMC 6, 407–428. doi: 10.2174/187152006778226530
- Capala, J., Barth, R. F., Bendayan, M., Lauzon, M., Adams, D. M., Soloway, A. H., et al. (1996). Boronated epidermal growth factor as a potential targeting agent for boron neutron capture therapy of brain tumors. *Bioconjug. Chem.* 7, 7–15. doi: 10.1021/bc950077q
- Carrasco-Triguero, M., Yi, J.-H., Dere, R., Qiu, Z. J., Lei, C., Li, Y., et al. (2013). Immunogenicity assays for antibody-drug conjugates: case study with adotrastuzumab emtansine. *Bioanalysis* 5, 1007–1023. doi: 10.4155/bio.13.64
- Carter, P. J., and Lazar, G. A. (2018). Next generation antibody drugs: pursuit of the 'high-hanging fruit'. Nat. Rev. Drug Discov. 17, 197–223. doi: 10.1038/nrd.2017.227
- Cescato, R., Maina, T., Nock, B. A., Nikolopoulou, A., Charalambidis, D., Piccand, V., et al. (2008). Bombesin receptor antagonists may be preferable to agonists for tumor targeting. J. Nucl. Med. 49, 318–326. doi: 10.2967/jnumed.107.045054
- Cha, S. W., Gu, H. K., Lee, K. P., Lee, M. H., Han, S. S., and Jeong, T. C. (2000). Immunotoxicity of ethyl carbamate in female BALB/c mice: role of esterase and cytochrome P450. *Toxicol. Lett.* 115, 173–181. doi:10.1016/S0378-4274(00)00176-4
- Chan, E. S. L., and Cronstein, B. N. (2013). Mechanisms of action of methotrexate. Bull Hosp Jt Dis. 71, 5–8.
- Chang, M., Zhang, F., Wei, T., Zuo, T., Guan, Y., Lin, G., et al. (2016). Smart linkers in polymer-drug conjugates for tumor-targeted delivery. J. Drug Target 24, 475–491. doi: 10.3109/1061186X.2015.1108324
- Chau, C. H., Steeg, P. S., and Figg, W. D. (2019). Antibody–drug conjugates for cancer. *Lancet* 394, 793–804. doi: 10.1016/S0140-6736(19)31774-X
- Chen, H. H. W., and Kuo, M. T. (2017). Improving radiotherapy in cancer treatment: promises and challenges. *Oncotarget* 8, 62742–62758. doi: 10.18632/oncotarget.18409
- Chen, J., Zhao, M., Feng, F., Sizovs, A., and Wang, J. (2013). Tunable thioesters as "reduction" responsive functionality for traceless reversible protein PEGylation. J. Am. Chem. Soc. 135, 10938–10941. doi: 10.1021/ja405261u
- Chen, J. M., Dando, P. M., Rawlings, N. D., Brown, M. A., Young, N. E., Stevens, R. A., et al. (1997). Cloning, isolation, and characterization of mammalian legumain, an asparaginyl endopeptidase. *J. Biol. Chem.* 272, 8090–8098. doi: 10.1074/jbc.272.12.8090
- Chen, K., and Chen, X. (2011). Integrin targeted delivery of chemotherapeutics. *Theranostics* 1, 189–200. doi: 10.7150/thno/v01p0189
- Chen, Z., Zhang, P., Cheetham, A. G., Moon, J. H., Moxley, J. W., and Lin, Y. A., et al. (2014). Controlled release of free doxorubicin from peptide-drug conjugates by drug loading. *J. Control. Release* 191, 123–130. doi: 10.1016/j.jconrel.2014.05.051
- Cho, H.-J., Lee, S.-J., Park, S.-J., Paik, C. H., Lee, S.-M., Kim, S., et al. (2016). Activatable iRGD-based peptide monolith: Targeting, internalization, and fluorescence activation for precise tumor imaging. *J. Control. Release* 237, 177–184. doi: 10.1016/j.jconrel.2016.06.032

- Choi, K. Y., Yoon, H. Y., Kim, J.-H., Bae, S. M., Park, R.-W., Kang, Y. M., et al. (2011). Smart nanocarrier based on PEGylated hyaluronic acid for cancer therapy. ACS Nano 5, 8591–8599. doi: 10.1021/nn202070n
- Ciechanover, A. (2005). Intracellular protein degradation: from a vague idea, through the lysosome and the ubiquitin-proteasome system, and onto human diseases and drug targeting (Nobel lecture). Angew. Chem. Int. Ed Engl. 44, 5944–5967. doi: 10.1002/anie.200501428
- Coderre, J. A., Elowitz, E. H., Chadha, M., Bergland, R., Capala, J., Joel, D. D., et al. (1997). Boron neutron capture therapy for glioblastoma multiforme using pboronophenylalanine and epithermal neutrons: trial design and early clinical results. J. Neurooncol. 33, 141–152. doi: 10.1023/A:1005741919442
- Coin, I., Beyermann, M., and Bienert, M. (2007). Solid-phase peptide synthesis: from standard procedures to the synthesis of difficult sequences. *Nat. Protoc.* 2, 3247–3256. doi: 10.1038/nprot.2007.454
- Colmers, W. F., and Bleakman, D. (1994). Effects of neuropeptide Y on the electrical properties of neurons. *Trends Neurosci.* 17, 373–379. doi:10.1016/0166-2236(94)90046-9
- Cox, D., Brennan, M., and Moran, N. (2010). Integrins as therapeutic targets: lessons and opportunities. *Nat. Rev. Drug Discov.* 9, 804–820. doi: 10.1038/nrd3266
- Dagenais, G. R., Leong, D. P., Rangarajan, S., Lanas, F., Lopez-Jaramillo, P., Gupta, R., et al. (2019). Variations in common diseases, hospital admissions, and deaths in middle-aged adults in 21 countries from five continents (PURE): a prospective cohort study. *Lancet* 395, 785–794. doi: 10.1016/S0140-6736(19)32007-0
- Dall, E., and Brandstetter, H. (2016). Structure and function of legumain in health and disease. *Biochimie* 122, 126–150. doi: 10.1016/j.biochi.2015.09.022
- Darker, J. G., Brough, S. J., Heath, J., and Smart, D. (2001). Discovery of potent and selective peptide agonists at the GRP-preferring bombesin receptor (BB2). *J. Pept. Sci.* 7, 598–605. doi: 10.1002/psc.359
- Dash, A., Chakraborty, S., Pillai, M. R. A., and Knapp, F. F. R. (2015). Peptide receptor radionuclide therapy: an overview. *Cancer Biother. Radiopharm.* 30, 47–71. doi: 10.1089/cbr.2014.1741
- DeBerardinis, R. J., and Chandel, N. S. (2016). Fundamentals of cancer metabolism. Sci. Adv. 2:e1600200. doi: 10.1126/sciadv.1600200
- Derossi, D., Calvet, S., Trembleau, A., Brunissen, A., Chassaing, G., and Prochiantz, A. (1996). Cell internalization of the third helix of the Antennapedia homeodomain is receptor-independent. J. Biol. Chem. 271, 18188–18193. doi: 10.1074/jbc.271.30.18188
- Desgrosellier, J. S., and Cheresh, D. A. (2010). Integrins in cancer: biological implications and therapeutic opportunities. *Nat. Rev. Cancer* 10, 9–22. doi: 10.1038/nrc2748
- DeVita, V. T., and Chu, E. (2008). A history of cancer chemotherapy. *Cancer Res.* 68, 8643–8653. doi: 10.1158/0008-5472.CAN-07-6611
- Donaghy, H. (2016). Effects of antibody, drug and linker on the preclinical and clinical toxicities of antibody-drug conjugates. *MAbs* 8, 659–671. doi: 10.1080/19420862.2016.1156829
- Dracham, C. B., Shankar, A., and Madan, R. (2018). Radiation induced secondary malignancies: a review article. *Radiat. Oncol. J.* 36, 85–94. doi: 10.3857/roj.2018.00290
- Drappatz, J., Brenner, A., Wong, E. T., Eichler, A., Schiff, D., Groves, M. D., et al. (2013). Phase I study of GRN1005 in recurrent malignant glioma. *Clin. Cancer Res.* 19, 1567–1576. doi: 10.1158/1078-0432.CCR-12-2481
- Dreher, M. R., Liu, W., Michelich, C. R., Dewhirst, M. W., Yuan, F., and Chilkoti, A. (2006). Tumor vascular permeability, accumulation, and penetration of macromolecular drug carriers. *J. Natl. Cancer Inst.* 98, 335–344. doi: 10.1093/jnci/djj070
- Dubowchik, G. M., Firestone, R. A., Padilla, L., Willner, D., Hofstead, S. J., Mosure, K., et al. (2002). Cathepsin B-labile dipeptide linkers for lysosomal release of doxorubicin from internalizing immunoconjugates: model studies of enzymatic drug release and antigen-specific *in vitro* anticancer activity. *Bioconjug. Chem.* 13, 855–869. doi: 10.1021/bc025536j
- Ducry, L., and Stump, B. (2010). Antibody-drug conjugates: linking cytotoxic payloads to monoclonal antibodies. *Bioconjug. Chem.* 21, 5–13. doi: 10.1021/bc9002019
- Duong, H. H. P., and Yung, L.-Y. L. (2013). Synergistic co-delivery of doxorubicin and paclitaxel using multi-functional micelles for cancer treatment. *Int. J. Pharm.* 454, 486–495. doi: 10.1016/j.ijpharm.2013.06.017

- Ehrlich, P., Himmelweit, F., Marquardt, M., and Dale, H. H. (1956). *The Collected Papers of Paul Ehrlich: In Four Volumes Including A Complete Bibliography.* London: Pergamon Press.
- El-Nassan, H. B. (2013). Advances in the discovery of kinesin spindle protein (Eg5) inhibitors as antitumor agents. *Eur. J. Med. Chem.* 62, 614–631. doi: 10.1016/j.ejmech.2013.01.031
- Engel, J. B., Schally, A. V., Halmos, G., Baker, B., Nagy, A., and Keller, G. (2005). Targeted therapy with a cytotoxic somatostatin analog, AN-238, inhibits growth of human experimental endometrial carcinomas expressing multidrug resistance protein MDR-1. *Cancer* 104, 1312–1321. doi: 10.1002/cncr.21327
- Erak, M., Bellmann-Sickert, K., Els-Heindl, S., and Beck-Sickinger, A. G. (2018). Peptide chemistry toolbox - transforming natural peptides into peptide therapeutics. *Bioorg. Med. Chem.* 26, 2759–2765. doi: 10.1016/j.bmc.2018.01.012
- Erspamer, V. (1988). Discovery, isolation, and characterization of bombesin-like peptides. Ann. N.Y Acad. Sci. 547, 3–9. doi: 10.1111/j.1749-6632.1988.tb23870.x
- Erspamer, V., Falconieri Erspamer, G., Inselvini, M., and Negri, L. (1972). Occurrence of bombesin and alytesin in extracts of the skin of three European discoglossid frogs and pharmacological actions of bombesin on extravascular smooth muscle. *Br. J. Pharmacol.* 45, 333–348. doi: 10.1111/j.1476-5381.1972.tb08087.x
- Falciani, C., Lelli, B., Brunetti, J., Pileri, S., Cappelli, A., Pini, A., et al. (2010). Modular branched neurotensin peptides for tumor target tracing and receptormediated therapy: a proof-of-concept. *Curr. Cancer Drug Targets* 10, 695–704. doi: 10.2174/156800910793605875
- Fang, Y., Yang, W., Cheng, L., Meng, F., Zhang, J., and Zhong, Z. (2017). EGFR-targeted multifunctional polymersomal doxorubicin induces selective and potent suppression of orthotopic human liver cancer *in vivo. Acta Biomater* 64, 323–333. doi: 10.1016/j.actbio.2017.10.013
- Farr, L. E., Robertson, J. S., Stickley, E. E., Bagnall, H. J., Easterday, O. D., and Kahle, W. (1958). "Revent advances in neutron capture therapy," in *Proceedings on the Second United Nations International Conference on the Peaceful Uses of Atomic Energy: Held in Geneva 1 September-13 September 1958*, ed. Naciones Unidas. International Conference (Geneva: United Nationals), 451–456.
- Fernández, M., Javaid, F., and Chudasama, V. (2018). Advances in targeting the folate receptor in the treatment/imaging of cancers. *Chem. Sci.* 9, 790–810. doi: 10.1039/C7SC04004K
- Firer, M. A., and Gellerman, G. (2012). Targeted drug delivery for cancer therapy: the other side of antibodies. J. Hematol. Oncol. 5, 70–86. doi: 10.1186/1756-8722-5-70
- Fitzgerald, K. A., Malhotra, M., Gooding, M., Sallas, F., Evans, J. C., Darcy, R., et al. (2016). A novel, anisamide-targeted cyclodextrin nanoformulation for siRNA delivery to prostate cancer cells expressing the sigma-1 receptor. *Int. J. Pharm.* 499, 131–145. doi: 10.1016/j.ijpharm.2015.12.055
- Forssell-Aronsson, E., Bernhardt, P., Nilsson, O., Tisell, L.-E., Wängberg, B., and Ahlman, H. (2004). Biodistribution data from 100 patients i.v. injected with 111In-DTPA-D-Phe1-octreotide. *Acta Oncol.* 43, 436–442. doi: 10.1080/02841860410030670
- Frank, R., Ahrens, V. M., Boehnke, S., Beck-Sickinger, A. G., and Hey-Hawkins, E. (2016). Charge-compensated metallacarborane building blocks for conjugation with peptides. *Chembiochem* 17, 308–317. doi: 10.1002/cbic.201500569
- Froidevaux, S., Calame-Christe, M., Tanner, H., and Eberle, A. N. (2005). Melanoma targeting with DOTA-alpha-melanocyte-stimulating hormone analogs: structural parameters affecting tumor uptake and kidney uptake. J. Nucl. Med. 46, 887–895.
- Froidevaux, S., and Eberle, A. N. (2002). Homologous regulation of melanocortin-1 receptor (MC1R) expression in melanoma tumor cells *in vivo. J. Recept. Signal Transduct. Res.* 22, 111–121. doi: 10.1081/RRS-120014590
- Frokjaer, S., and Otzen, D. E. (2005). Protein drug stability: a formulation challenge. Nat. Rev. Drug Discov. 4, 298–306. doi: 10.1038/nrd1695
- Galmarini, C. M., Mackey, J. R., and Dumontet, C. (2002). Nucleoside analogues and nucleobases in cancer treatment. *Lancet Oncol.* 3, 415–424. doi: 10.1016/S1470-2045(02)00788-X
- Gaviglio, L., Gross, A., Metzler-Nolte, N., and Ravera, M. (2012). Synthesis and *in vitro* cytotoxicity of cis,cis,transdiamminedichloridodisuccinatoplatinum(IV)-peptide bioconjugates. *Metallomics* 4, 260–266. doi: 10.1039/c2mt00171c

- Gianfaldoni, S., Gianfaldoni, R., Wollina, U., Lotti, J., Tchernev, G., and Lotti, T. (2017). An overview on radiotherapy: from its history to its current applications in dermatology. *Open Access Maced. J. Med. Sci.* 5, 521–525. doi:10.3889/oamjms.2017.122
- Gicquiaux, H., Lecat, S., Gaire, M., Dieterlen, A., Mély, Y., Takeda, K., et al. (2002). Rapid internalization and recycling of the human neuropeptide Y Y(1) receptor. J. Biol. Chem. 277, 6645–6655. doi: 10.1074/jbc.M107224200
- Gilad, Y., Noy, E., Senderowitz, H., Albeck, A., Firer, M. A., and Gellerman, G. (2016a). Dual-drug RGD conjugates provide enhanced cytotoxicity to melanoma and non-small lung cancer cells. *Biopolymers* 106, 160–171. doi: 10.1002/bip.22800
- Gilad, Y., Noy, E., Senderowitz, H., Albeck, A., Firer, M. A., and Gellerman, G. (2016b). Synthesis, biological studies and molecular dynamics of new anticancer RGD-based peptide conjugates for targeted drug delivery. *Bioorg. Med. Chem.* 24, 294–303. doi: 10.1016/j.bmc.2015.12.020
- Gilman, A. (1963). The initial clinical trial of nitrogen mustard. Am. J. Surg. 105, 574–578. doi: 10.1016/0002-9610(63)90232-0
- Ginj, M., Zhang, H., Waser, B., Cescato, R., Wild, D., Wang, X., et al. (2006). Radiolabeled somatostatin receptor antagonists are preferable to agonists for *in vivo* peptide receptor targeting of tumors. *Proc. Natl. Acad. Sci. U.S.A.* 103, 16436–16441. doi: 10.1073/pnas.0607761103
- Goffredo, V., Paradiso, A., Ranieri, G., and Gadaleta, C. D. (2011). Yttrium-90 (90Y) in the principal radionuclide therapies: an efficacy correlation between peptide receptor radionuclide therapy, radioimmunotherapy and transarterial radioembolization therapy. Ten years of experience (1999-2009). Crit. Rev. Oncol. Hematol. 80, 393–410. doi: 10.1016/j.critrevonc.2011.01.012
- Gondi, C. S., and Rao, J. S. (2013). Cathepsin B as a cancer target. *Expert Opin. Ther. Targets* 17, 281–291. doi: 10.1517/14728222.2013.740461
- González, N., Moody, T. W., Igarashi, H., Ito, T., and Jensen, R. T. (2008). Bombesin-related peptides and their receptors: recent advances in their role in physiology and disease states. *Curr. Opin. Endocrinol. Diabetes Obes.* 15, 58–64. doi: 10.1097/MED.0b013e3282f3709b
- Grandal, M. V., and Madshus, I. H. (2008). Epidermal growth factor receptor and cancer: control of oncogenic signalling by endocytosis. J. Cell. Mol. Med. 12, 1527–1534. doi: 10.1111/j.1582-4934.2008.00298.x
- Gründker, C., and Emons, G. (2017). The role of gonadotropin-releasing hormone in cancer cell proliferation and metastasis. *Front. Endocrinol.* 8:187. doi: 10.3389/fendo.2017.00187
- Gründker, C., Ernst, J., Reutter, M. D., Ghadimi, B. M., and Emons, G. (2011). Effective targeted chemotherapy using AEZS-108 (AN-152) for LHRH receptor-positive pancreatic cancers. *Oncol. Rep.* 26, 629–635. doi: 10.3892/or.2011.1340
- Gugger, M., and Reubi, J.-C. (1999). Gastrin-releasing peptide receptors in nonneoplastic and neoplastic human breast. Am. J. Pathol. 155, 2067–2076. doi: 10.1016/S0002-9440(10)65525-3
- Guryanov, I., Bondesan, A., Visentini, D., Orlandin, A., Biondi, B., Toniolo, C., et al. (2016). Innovative chemical synthesis and conformational hints on the lipopeptide liraglutide. *J. Pept. Sci.* 22, 471–479. doi: 10.1002/ psc.2890
- Halmos, G., Arencibia, J. M., Schally, A. V., Davis, R., and Bostwick, D. G. (2000). High incidence of receptors for luteinizing hormone-releasing hormone (LHRH) and LHRH receptor gene expression in human prostate cancers. J. Urol. 163, 623–629. doi: 10.1016/S0022-5347(05)67947-5
- Hanahan, D., and Weinberg, R. A. (2011). Hallmarks of cancer: the next generation. *Cell* 144, 646–674. doi: 10.1016/j.cell.2011.02.013
- Hansel, T. T., Kropshofer, H., Singer, T., Mitchell, J. A., and George, A. J. T. (2010). The safety and side effects of monoclonal antibodies. *Nat. Rev. Drug Discov.* 9, 325–338. doi: 10.1038/nrd3003
- Harrison, G. S., Wierman, M. E., Nett, T. M., and Glode, L. M. (2004). Gonadotropin-releasing hormone and its receptor in normal and malignant cells. *Endocr. Relat. Cancer* 11, 725–748. doi: 10.1677/erc.1.00777
- Hartman, T., and Carlsson, J. (1994). Radiation dose heterogeneity in receptor and antigen mediated boron neutron capture therapy. *Radiat. Oncol.* 31, 61–75. doi: 10.1016/0167-8140(94)90414-6
- Hawthorne, M. F. (1993). The role of chemistry in the development of boron neutron capture therapy of cancer. Angew. Chem. Int. Ed. Engl. 32, 950–984. doi: 10.1002/anie.199309501

- Hellmich, M. R., Battey, J. F., and Northup, J. K. (1997). Selective reconstitution of gastrin-releasing peptide receptor with Gαq. Proc. Natl. Acad. Sci. U.S.A. 94, 751–756. doi: 10.1073/pnas.94.2.751
- Helmbach, H., Rossmann, E., Kern, M. A., and Schadendorf, D. (2001). Drug-resistance in human melanoma. *Int. J. Cancer* 93, 617–622. doi: 10.1002/ijc.1378
- Hindié, E., Zanotti-Fregonara, P., Quinto, M. A., Morgat, C., and Champion, C. (2016). Dose Deposits from 90Y, 177Lu, 111In, and 161Tb in micrometastases of various sizes: implications for radiopharmaceutical therapy. *J. Nucl. Med* 57, 759–764. doi: 10.2967/jnumed.115.170423
- Hirmas, N., Jadaan, R., and Al-Ibraheem, A. (2018). Peptide receptor radionuclide therapy and the treatment of gastroentero-pancreatic neuroendocrine tumors: current findings and future perspectives. *Nucl. Med. Mol. Imaging* 52, 190–199. doi: 10.1007/s13139-018-0517-x
- Hock, M. B., Thudium, K. E., Carrasco-Triguero, M., and Schwabe, N. F. (2015). Immunogenicity of antibody drug conjugates: bioanalytical methods and monitoring strategy for a novel therapeutic modality. *AAPS J.* 17, 35–43. doi: 10.1208/s12248-014-9684-6
- Hofmann, S., Maschauer, S., Kuwert, T., Beck-Sickinger, A. G., and Prante, O. (2015). Synthesis and *in vitro* and *in vivo* evaluation of an (18)F-labeled neuropeptide Y analogue for imaging of breast cancer by PET. *Mol. Pharm.* 12, 1121–1130. doi: 10.1021/mp500601z
- Hoppenz, P., Els-Heindl, S., and Beck-Sickinger, A. G. (2019). Identification and stabilization of a highly selective gastrin-releasing peptide receptor agonist. J. Pept. Sci. 25:e3224. doi: 10.1002/psc.3224
- Hoppenz, P., Els-Heindl, S., Kellert, M., Kuhnert, R., Saretz, S., Lerchen, H.-G., et al. (2020). A selective carborane-functionalized gastrin-releasing peptide receptor agonist as boron delivery agent for boron neutron capture therapy. J. Org. Chem. 85, 1446–1457. doi: 10.1021/acs.joc.9b02406
- Housman, G., Byler, S., Heerboth, S., Lapinska, K., Longacre, M., Snyder, N., et al. (2014). Drug resistance in cancer: an overview. *Cancers* 6, 1769–1792. doi: 10.3390/cancers6031769
- Huang, C. M., Wu, Y. T., and Chen, S. T. (2000). Targeting delivery of paclitaxel into tumor cells via somatostatin receptor endocytosis. *Chem. Biol.* 7, 453–461. doi: 10.1016/S1074-5521(00)00131-9
- Hynes, R. (1987). Integrins: a family of cell surface receptors. *Cell* 48, 549–554. doi: 10.1016/0092-8674(87)90233-9
- Iguchi, Y., Michiue, H., Kitamatsu, M., Hayashi, Y., Takenaka, F., Nishiki, T., et al. (2015). Tumor-specific delivery of BSH-3R for boron neutron capture therapy and positron emission tomography imaging in a mouse brain tumor model. *Biomaterials* 56, 10–17. doi: 10.1016/j.biomaterials.2015.03.061
- Insel, P. A., Sriram, K., Wiley, S. Z., Wilderman, A., Katakia, T., McCann, T., et al. (2018). GPCRomics: GPCR expression in cancer cells and tumors identifies new, potential biomarkers and therapeutic targets. *Front. Pharmacol.* 9:431. doi: 10.3389/fphar.2018.00431
- Jafari, A., Salouti, M., Shayesteh, S. F., Heidari, Z., Rajabi, A. B., Boustani, K., et al. (2015). Synthesis and characterization of Bombesinsuperparamagnetic iron oxide nanoparticles as a targeted contrast agent for imaging of breast cancer using MRI. *Nanotechnology* 26, 75101–75112. doi: 10.1088/0957-4484/26/7/075101
- Jain, R. K., and Stylianopoulos, T. (2010). Delivering nanomedicine to solid tumors. Nat. Rev. Clin. Oncol. 7, 653–664. doi: 10.1038/nrclinonc.2010.139
- Jamous, M., Haberkorn, U., and Mier, W. (2013). Synthesis of peptide radiopharmaceuticals for the therapy and diagnosis of tumor diseases. *Molecules* 18, 3379–3409. doi: 10.3390/molecules18033379
- Jensen, R. T., Battey, J. F., Spindel, E. R., and Benya, R. V. (2008). International Union of pharmacology. LXVIII. Mammalian bombesin receptors: nomenclature, distribution, pharmacology, signaling, and functions in normal and disease states. Proc. Natl. Acad. Sci. U.S.A. 60, 1–42. doi: 10.1124/pr.107.07108
- Joubert, N., Denevault-Sabourin, C., Bryden, F., and Viaud-Massuard, M.-C. (2017). Towards antibody-drug conjugates and prodrug strategies with extracellular stimuli-responsive drug delivery in the tumor microenvironment for cancer therapy. *Eur. J. Med. Chem.* 142, 393–415. doi: 10.1016/j.ejmech.2017.08.049
- Kadonosono, T., Yamano, A., Goto, T., Tsubaki, T., Niibori, M., Kuchimaru, T., et al. (2015). Cell penetrating peptides improve tumor delivery of cargos

through neuropilin-1-dependent extravasation. J. Control. Release 201, 14–21. doi: 10.1016/j.jconrel.2015.01.011

- Kageji, T., Nakagawa, Y., Kitamura, K., Matsumoto, K., and Hatanaka, H. (1997). Pharmacokinetics and boron uptake of BSH (Na2B12H11SH) in patients with intracranial tumors. J. Neurooncol. 33, 117–130. doi: 10.1023/A:1005785718533
- Kapp, T. G., Rechenmacher, F., Neubauer, S., Maltsev, O. V., Cavalcanti-Adam, E. A., Zarka, R., et al. (2017). A comprehensive evaluation of the activity and selectivity profile of ligands for rgd-binding integrins. *Sci. Rep.* 7:e39805. doi: 10.1038/srep39805
- Karagiannis, T. C. (2007). Comparison of different classes of radionuclides for potential use in radioimmunotherapy. *Hell. J. Nucl. Med.* 10, 82–88.
- Kato, Y., Ozawa, S., Miyamoto, C., Maehata, Y., Suzuki, A., Maeda, T., et al. (2013). Acidic extracellular microenvironment and cancer. *Cancer Cell Int.* 13, 89–97. doi: 10.1186/1475-2867-13-89
- Kaur, H., Bruno, J. G., Kumar, A., and Sharma, T. K. (2018). Aptamers in the therapeutics and diagnostics pipelines. *Theranostics* 8, 4016–4032. doi: 10.7150/thno.25958
- Keller, G., Schally, A. V., Gaiser, T., Nagy, A., Baker, B., Halmos, G., et al. (2005). Receptors for luteinizing hormone releasing hormone (LHRH) expressed in human non-Hodgkin's lymphomas can be targeted for therapy with the cytotoxic LHRH analogue AN-207. *Eur. J. Cancer.* 41, 2196–2202. doi: 10.1016/j.ejca.2005.06.018
- Keskin, O., and Yalcin, S. (2013). A review of the use of somatostatin analogs in oncology. Onco. Targets. Ther. 6, 471–483. doi: 10.2147/OTT.S39987
- Khan, I. U., Zwanziger, D., Böhme, I., Javed, M., Naseer, H., Hyder, S. W., et al. (2010). Breast-cancer diagnosis by neuropeptide Y analogues: from synthesis to clinical application. *Angew. Chem. Int. Ed. Engl.* 49, 1155–1158. doi: 10.1002/anie.200905008
- Kim, M., Kim, D.-M., Kim, K.-S., Jung, W., and Kim, D.-E. (2018). Applications of cancer cell-specific aptamers in targeted delivery of anticancer therapeutic agents. *Molecules* 23:830. doi: 10.3390/molecules23040830
- Kimura, S., Masunaga, S.-,i., Harada, T., Kawamura, Y., Ueda, S., Okuda, K., et al. (2011). Synthesis and evaluation of cyclic RGD-boron cluster conjugates to develop tumor-selective boron carriers for boron neutron capture therapy. *Bioorg. Med. Chem.* 19, 1721–1728. doi: 10.1016/j.bmc.2011.01.020
- Knight, S. D., and Parrish, C. A. (2008). Recent progress in the identification and clinical evaluation of inhibitors of the mitotic kinesin KSP. *Curr. Top. Med. Chem.* 8, 888–904. doi: 10.2174/156802608784911626
- Kokko, K. P., Hadden, M. K., Orwig, K. S., Mazella, J., and Dix, T. A. (2003). In vitro analysis of stable, receptor-selective neurotensin8-13 analogues. J. Med. Chem. 46, 4141–4148. doi: 10.1021/jm0300633
- Körner, M., and Reubi, J. C. (2007). NPY receptors in human cancer: a review of current knowledge. *Peptides* 28, 419–425. doi: 10.1016/j.peptides.2006.08.037
- Körner, M., Waser, B., and Reubi, J. C. (2008). High expression of neuropeptide Y1 receptors in ewing sarcoma tumors. *Clin. Cancer Res.* 14, 5043–5049. doi: 10.1158/1078-0432.CCR-07-4551
- Kovács, M., Vincze, B., Horváth, J. E., and Seprodi, J. (2007). Structureactivity study on the LH- and FSH-releasing and anticancer effects of gonadotropin-releasing hormone (GnRH)-III analogs. *Peptides* 28, 821–829. doi: 10.1016/j.peptides.2007.01.003
- Kratz, F., Müller, I. A., Ryppa, C., and Warnecke, A. (2008). Prodrug strategies in anticancer chemotherapy. *ChemMedChem* 3, 20–53. doi: 10.1002/cmdc.200700159
- Kreiner, A. J., Bergueiro, J., Cartelli, D., Baldo, M., Castell, W., Asoia, J. G., et al. (2016). Present status of accelerator-based BNCT. *Rep. Pract. Oncol. Radiother*. 21, 95–101. doi: 10.1016/j.rpor.2014.11.004
- Kroog, G. S., Jensen, R. T., and Battey, J. F. (1995). Mammalian bombesin receptors. *Med. Res. Rev.* 15, 389–417. doi: 10.1002/med.2610150502
- Kue, C. S., Kamkaew, A., Burgess, K., Kiew, L. V., Chung, L. Y., and Lee, H. B. (2016). Small molecules for active targeting in Cancer. *Med. Res. Rev* 36, 494–575. doi: 10.1002/med.21387
- Kwekkeboom, D. J., Kam, B. L., van Essen, M., Teunissen, J. J. M., van Eijck, C. H. J., Valkema, R., et al. (2010). Somatostatin-receptor-based imaging and therapy of gastroenteropancreatic neuroendocrine tumors. *Endocr. Relat. Cancer* 17, 53–73. doi: 10.1677/ERC-09-0078
- La Bella, R., García Garayoa, E., Langer, M., Bläuenstein, P., Beck-Sickinger, A. G., and Schubiger, P. A. (2002). *In vitro* and *in vivo* evaluation of a 99mTc(I)-labeled

bombesin analogue for imaging of gastrin releasing peptide receptor-positive tumors. *Nucl. Med. Biol.* 29, 553–560. doi: 10.1016/S0969-8051(02)00314-1

- Langer, M., Kratz, F., Rothen-Rutishauser, B., Wunderli-Allenspach, H., and Beck-Sickinger, A. G. (2001). Novel peptide conjugates for tumor-specific chemotherapy. J. Med. Chem. 44, 1341–1348. doi: 10.1021/jm001065f
- Larhammar, D. (1996). Evolution of neuropeptide Y, peptide YY and pancreatic polypeptide. *Regul. Pept.* 62, 1–11. doi: 10.1016/0167-0115(95)00169-7
- León-Rodríguez, L. M., de, and Kovacs, Z. (2008). The synthesis and chelation chemistry of DOTA-peptide conjugates. *Bioconjug. Chem.* 19, 391–402. doi: 10.1021/bc700328s
- Lerchen, H.-G., Stelte-Ludwig, B., Berndt, S., Sommer, A., Dietz, L., Rebstock, A.-S., et al. (2019). Antibody-prodrug conjugates with KSP inhibitors and legumain-mediated metabolite formation. *Chemistry* 25, 8208–8213. doi: 10.1002/chem.201900441
- Li, C., Cai, G., Song, D., Gao, R., Teng, P., Zhou, L., et al. (2019). Development of EGFR-targeted evodiamine nanoparticles for the treatment of colorectal cancer. *Biomater. Sci.* 7, 3627–3639. doi: 10.1039/C9BM00613C
- Li, J., Tian, Y., and Wu, A. (2015). Neuropeptide Y receptors: a promising target for cancer imaging and therapy. *Regen. Biomater.* 2, 215–219. doi: 10.1093/rb/rbv013
- Li, Z., Zhao, R., Wu, X., Sun, Y., Yao, M., Li, J., et al. (2005). Identification and characterization of a novel peptide ligand of epidermal growth factor receptor for targeted delivery of therapeutics. *FASEB J.* 19, 1978–1985. doi: 10.1096/fj.05-4058com
- Limonta, P., Montagnani Marelli, M., Mai, S., Motta, M., Martini, L., and Moretti, R. M. (2012). GnRH receptors in cancer: from cell biology to novel targeted therapeutic strategies. *Endocr. Rev.* 33, 784–811. doi: 10.1210/er.2012-1014
- Liu, C., Sun, C., Huang, H., Janda, K., and Edgington, T. (2003). Overexpression of legumain in tumors is significant for invasion/metastasis and a candidate enzymatic target for prodrug therapy. *Cancer Res.* 63, 2957–2964.
- Liu, S. (2008). Bifunctional coupling agents for radiolabeling of biomolecules and target-specific delivery of metallic radionuclides. *Adv. Drug Deliv. Rev.* 60, 1347–1370. doi: 10.1016/j.addr.2008.04.006
- Locher, L. G. (1936). Biological effects and therapeutic possibilities of neutrons. Am. J. Roentgenol. 36, 1–13.
- Luderer, M. J., de La Puente, P., and Azab, A. K. (2015). Advancements in tumor targeting strategies for boron neutron capture therapy. *Pharm. Res.* 32, 2824–2836. doi: 10.1007/s11095-015-1718-y
- Mäde, V., Els-Heindl, S., and Beck-Sickinger, A. G. (2014). Automated solid-phase peptide synthesis to obtain therapeutic peptides. *Beilstein J. Org. Chem.* 10, 1197–1212. doi: 10.3762/bjoc.10.118
- Mai, C.-W., Chung, F. F.-L., and Leong, C.-O. (2017). Targeting legumain as a novel therapeutic strategy in cancers. *Curr. Drug Targets* 18, 1259–1268. doi: 10.2174/1389450117666161216125344
- Malhotra, V., and Perry, M. C. (2003). Classical chemotherapy: mechanisms, toxicities and the therapeutic window. *Cancer Biol. Ther.* 2, 2–4. doi: 10.4161/cbt.199
- Mansi, R., Fleischmann, A., Mäcke, H. R., and Reubi, J. C. (2013). Targeting GRPR in urological cancers-from basic research to clinical application. *Nat. Rev. Urol.* 10, 235–244. doi: 10.1038/nrurol.2013.42
- Mantey, S. A., Weber, H. C., Sainz, E., Akeson, M. A., Ryan, R. R., Pradhan, T. K., et al. (1997). Discovery of a high affinity radioligand for the human orphan receptor, bombesin receptor subtype 3, which demonstrates that it has a unique pharmacology compared with other mammalian bombesin receptors. *J. Biol. Chem.* 272, 26062–26071. doi: 10.1074/jbc.272.41.26062
- Markwalder, R., and Reubi, J.-C. (1999). Gastrin-releasing peptide receptors in the human prostate: relation to neoplastic transformation. *Cancer Res.* 59, 1152–1159.
- Marso, S. P., Bain, S. C., Consoli, A., Eliaschewitz, F. G., Jódar, E., Leiter, L. A., et al. (2016). Semaglutide and cardiovascular outcomes in patients with type 2 diabetes. N. Engl. J. Med. 375, 1834–1844. doi: 10.1056/NEJMoa1607141
- Masunaga, S.-,i., Kimura, S., Harada, T., Okuda, K., Sakurai, Y., Tanaka, H., et al. (2012). Evaluating the usefulness of a novel 10b-carrier conjugated with cyclic RGD peptide in boron neutron capture therapy. *World J. Oncol.* 3, 103–112. doi: 10.4021/wjon477w
- McDonald, T. J., Jörnvall, H., Nilsson, G., Vagne, M., Ghatei, M., Bloom, S. R., et al. (1979). Characterization of a gastrin releasing peptide from porcine

non-antral gastric tissue. *Biochem. Biophys. Res. Commun.* 90, 227–233. doi: 10.1016/0006-291X(79)91614-0

- Medina, S. H., Chevliakov, M. V., Tiruchinapally, G., Durmaz, Y. Y., Kuruvilla, S. P., and Elsayed, M. E. H. (2013). Enzyme-activated nanoconjugates for tunable release of doxorubicin in hepatic cancer cells. *Biomaterials* 34, 4655–4666. doi: 10.1016/j.biomaterials.2013.02.070
- Meister, A., and Anderson, M. E. (1983). Glutathione. Annu. Rev. Biochem. 52, 711–760. doi: 10.1146/annurev.bi.52.070183.003431
- Miao, Y., and Quinn, T. P. (2008). Peptide-targeted radionuclide therapy for melanoma. *Crit. Rev. Oncol. Hematol.* 67, 213–228. doi: 10.1016/j.critrevonc.2008.02.006
- Michel, M. C., Beck-Sickinger, A., Cox, H., Doods, H. N., Herzog, H., Larhammar, D., et al. (1998). XVI. International union of pharmacology recommendations for the nomenclature of neuropeptide Y, peptide YY, and pancreatic polypeptide receptors. *Pharmacol. Rev.* 50, 143–150.
- Michel, M. C., and Rascher, W. (1995). Neuropeptide Y: a possible role in hypertension? *J. Hypertens.* 13, 385–395. doi: 10.1097/00004872-199504000-00003
- Michel, R. B., Andrews, P. M., Rosario, A. V., Goldenberg, D. M., and Mattes, M. J. (2005). 177Lu-antibody conjugates for single-cell kill of B-lymphoma cells *in vitro* and for therapy of micrometastases *in vivo*. *Nucl. Med. Biol.* 32, 269–278. doi: 10.1016/j.nucmedbio.2005.01.003
- Michiue, H., Sakurai, Y., Kondo, N., Kitamatsu, M., Bin, F., Nakajima, K., et al. (2014). The acceleration of boron neutron capture therapy using multilinked mercaptoundecahydrododecaborate (BSH) fused cell-penetrating peptide. *Biomaterials* 35, 3396–3405. doi: 10.1016/j.biomaterials.2013. 12.055
- Mier, W., Gabel, D., Haberkorn, U., and Eisenhut, M. (2004). Conjugation of the closo-Borane Mercaptoundecahydrododecaborate (BSH) to a tumour selective peptide. Z. Anorg. Allg. Chem. 630, 1258–1262. doi: 10.1002/zaac.200400064
- Minotti, G., Menna, P., Salvatorelli, E., Cairo, G., and Gianni, L. (2004). Anthracyclines: molecular advances and pharmacologic developments in antitumor activity and cardiotoxicity. *Pharmacol. Rev.* 56, 185–229. doi: 10.1124/pr.56.2.6
- Monks, S. A., Karagianis, G., Howlett, G. J., and Norton, R. S. (1996). Solution structure of human neuropeptide Y. J. Biomol. NMR. 8, 379–390. doi: 10.1007/BF00228141
- Moody, T. W., Mantey, S. A., Fuselier, J. A., Coy, D. H., and Jensen, R. T. (2007). Vasoactive intestinal peptide-camptothecin conjugates inhibit the proliferation of breast cancer cells. *Peptides* 28, 1883–1890. doi: 10.1016/j.peptides.2007.04.017
- Moody, T. W., Mantey, S. A., Pradhan, T. K., Schumann, M., Nakagawa, T., Martinez, A., et al. (2004). Development of high affinity camptothecinbombesin conjugates that have targeted cytotoxicity for bombesin receptor-containing tumor cells. *J. Biol. Chem.* 279, 23580–23589. doi: 10.1074/jbc.M401938200
- Moody, T. W., Sun, L.-C., Mantey, S. A., Pradhan, T., Mackey, L. V., Gonzales, N., et al. (2006). *In vitro* and *in vivo* antitumor effects of cytotoxic camptothecin-bombesin conjugates are mediated by specific interaction with cellular bombesin receptors. *J. Pharmacol. Exp. Ther.* 318, 1265–1272. doi: 10.1124/jpet.106.104141
- Moore, C. A. C., Milano, S. K., and Benovic, J. L. (2007). Regulation of receptor trafficking by GRKs and arrestins. *Annu. Rev. Physiol.* 69, 451–482. doi: 10.1146/annurev.physiol.69.022405.154712
- Moreno, P., Ramos-Alvarez, I., Moody, T. W., and Jensen, R. T. (2016). Bombesin related peptides/receptors and their promising therapeutic roles in cancer imaging, targeting and treatment. *Expert Opin. Ther. Targets* 20, 1055–1073. doi: 10.1517/14728222.2016.1164694
- Mori, Y., Suzuki, A., Yoshino, K., and Kakihana, H. (1989). Complex formation of p-boronophenylalanine with some monosaccharides. *Pigment. Cell Res.* 2, 273–277. doi: 10.1111/j.1600-0749.1989.tb00203.x
- Mörl, K., and Beck-Sickinger, A. G. (2015). Intracellular trafficking of neuropeptide Y receptors. *Prog. Mol. Biol. Transl. Sci.* 132, 73–96. doi: 10.1016/bs.pmbts.2015.02.011
- Morris, M. C., Depollier, J., Mery, J., Heitz, F., and Divita, G. (2001). A peptide carrier for the delivery of biologically active proteins into mammalian cells. *Nat. Biotechnol.* 19, 1173–1176. doi: 10.1038/nbt1201-1173

- Muratovska, A., and Eccles, M. R. (2004). Conjugate for efficient delivery of short interfering RNA (siRNA) into mammalian cells. *FEBS Lett.* 558, 63–68. doi: 10.1016/S0014-5793(03)01505-9
- Nagy, A., Schally, A. V., Armatis, P., Szepeshazi, K., Halmos, G., Kovacs, M., et al. (1996). Cytotoxic analogs of luteinizing hormone-releasing hormone containing doxorubicin or 2-pyrrolinodoxorubicin, a derivative 500-1000 times more potent. *Proc. Natl. Acad. Sci. U.S.A.* 93, 7269–7273. doi: 10.1073/pnas.93.14.7269
- Nagy, A., Schally, A. V., Halmos, G., Armatis, P., Cai, R. Z., Csernus, V., et al. (1998). Synthesis and biological evaluation of cytotoxic analogs of somatostatin containing doxorubicin or its intensely potent derivative, 2-pyrrolinodoxorubicin. *Proc. Natl. Acad. Sci. U.S.A.* 95, 1794–1799. doi: 10.1073/pnas.95.4.1794
- Nakase, I., Katayama, M., Hattori, Y., Ishimura, M., Inaura, S., Fujiwara, D., et al. (2019). Intracellular target delivery of cell-penetrating peptide-conjugated dodecaborate for boron neutron capture therapy (BNCT). *Chem. Commun.* 55, 13955–13958. doi: 10.1039/C9CC03924D
- Naqvi, S. A. R., Matzow, T., Finucane, C., Nagra, S. A., Ishfaq, M. M., Mather, S. J., et al. (2010). Insertion of a lysosomal enzyme cleavage site into the sequence of a radiolabeled neuropeptide influences cell trafficking *in vitro* and *in vivo*. *Cancer Biother. Radiopharm.* 25, 89–95. doi: 10.1089/cbr.2009.0666
- Nejadmoghaddam, M.-R., Minai-Tehrani, A., Ghahremanzadeh, R., Mahmoudi, M., Dinarvand, R., and Zarnani, A.-H. (2019). Antibody-drug conjugates: possibilities and challenges. *Avicenna J. Med. Biotechnol.* 11, 3–23.
- Nieto Gutierrez, A., and McDonald, P. H. (2018). GPCRs: emerging anti-cancer drug targets. Cell. Signal 41, 65–74. doi: 10.1016/j.cellsig.2017.09.005
- Ochakovskaya, R., Osorio, L., Goldenberg, D. M., and Mattes, M. J. (2001). Therapy of disseminated B-cell lymphoma xenografts in severe combined immunodeficient mice with an anti-CD74 antibody conjugated with (111)indium, (67)gallium, or (90)yttrium. *Clin. Cancer Res.* 7, 1505–1510.
- O'Donoghue, J. A., Bardiès, M., and Wheldon, T. E. (1995). Relationships between tumor size and curability for uniformly targeted therapy with beta-emitting radionuclides. *J. Nucl. Med.* 36, 1902–1909.
- Ohkuma, S., and Poole, B. (1978). Fluorescence probe measurement of the intralysosomal pH in living cells and the perturbation of pH by various agents. *Proc. Natl. Acad. Sci. U.S.A.* 75, 3327–3331. doi: 10.1073/pnas.75.7.3327
- Olias, G., Viollet, C., Kusserow, H., Epelbaum, J., and Meyerhof, W. (2004). Regulation and function of somatostatin receptors. *J. Neurochem.* 89, 1057–1091. doi: 10.1111/j.1471-4159.2004.02402.x
- Patel, Y. C. (1999). Somatostatin and its receptor family. Front. Neuroendocrinol. 20, 157–198. doi: 10.1006/frne.1999.0183
- Patterson, L. H., McKeown, S. R., Robson, T., Gallagher, R., Raleigh, S. M., and Orr, S. (1999). Antitumour prodrug development using cytochrome P450 (CYP) mediated activation. *Anticancer. Drug Des.* 14, 473–486.
- Pedragosa-Badia, X., Stichel, J., and Beck-Sickinger, A. G. (2013). Neuropeptide Y receptors: how to get subtype selectivity. *Front. Endocrinol.* 4:5. doi: 10.3389/fendo.2013.00005
- Pfister, S. X., and Ashworth, A. (2017). Marked for death: targeting epigenetic changes in cancer. *Nat. Rev. Drug Discov.* 16, 241–263. doi: 10.1038/nrd.2016.256
- Pierschbacher, M. D., and Ruoslahti, E. (1984). Cell attachment activity of fibronectin can be duplicated by small synthetic fragments of the molecule. *Nature* 309, 30–33. doi: 10.1038/309030a0
- Playford, R. J., and Cox, H. M. (1996). Peptide YY and neuropeptide Y: two peptides intimately involved in electrolyte homeostasis. *Trends Pharmacol. Sci.* 17, 436–438. doi: 10.1016/S0165-6147(96)01008-5
- Plow, E. F., Haas, T. A., Zhang, L., Loftus, J., and Smith, J. W. (2000). Ligand binding to integrins. J. Biol. Chem. 275, 21785–21788. doi: 10.1074/jbc.R000003200
- Pu, F., Qiao, J., Xue, S., Yang, H., Patel, A., Wei, L., et al. (2015). GRPR-targeted protein contrast agents for molecular imaging of receptor expression in cancers by MRI. Sci. Rep. 5:16214. doi: 10.1038/srep16214
- Regberg, J., Srimanee, A., and Langel, U. (2012). Applications of cell-penetrating peptides for tumor targeting and future cancer therapies. *Pharmaceuticals* 5, 991–1007. doi: 10.3390/ph5090991
- Régina, A., Demeule, M., Ch,é, C., Lavallée, I., Poirier, J., Gabathuler, R., et al. (2008). Antitumour activity of ANG1005, a conjugate between paclitaxel and

the new brain delivery vector Angiopep-2. Br. J. Pharmacol. 155, 185-197. doi: 10.1038/bjp.2008.260

- Reisine, T., and Bell, G. I. (1995). Molecular biology of somatostatin receptors. *Endocr. Rev.* 16, 427–442. doi: 10.1210/edrv-16-4-427
- Rellinger, E. J., Romain, C., Choi, S., Qiao, J., and Chung, D. H. (2015). Silencing gastrin-releasing peptide receptor suppresses key regulators of aerobic glycolysis in neuroblastoma cells. *Pediatr. Blood Cancer* 62, 581–586. doi: 10.1002/pbc.25348
- Reubi, J.-C. (2003). Peptide receptors as molecular targets for cancer diagnosis and therapy. *Endocr. Rev.* 24, 389–427. doi: 10.1210/er.2002-0007
- Reubi, J.-C., Gugger, M., and Waser, B. (2002a). Co-expressed peptide receptors in breast cancer as a molecular basis for *in vivo* multireceptor tumour targeting. *Eur. J. Nucl. Med. Mol. Imaging* 29, 855–862. doi: 10.1007/s00259-002-0794-5
- Reubi, J.-C., Wenger, S., Schmuckli-Maurer, J., Schaer, J.-C., and Gugger, M. (2002b). Bombesin receptor subtypes in human cancers: detection with the universal radioligand (125)I-[D-TYR(6), beta-ALA(11), PHE(13), NLE(14)] bombesin(6-14). *Clin. Cancer Res.* 8, 1139–1146.
- Reubi, J. C., Gugger, M., Waser, B., and Schaer, J. C. (2001). Y(1)-mediated effect of neuropeptide Y in cancer: breast carcinomas as targets. *Cancer Res.* 61, 4636–4641.
- Reubi, J. C., Laissue, J., Krenning, E., and Lamberts, S. W. J. (1992). Somatostatin receptors in human cancer: incidence, characteristics, functional correlates and clinical implications. *J. Steroid Biochem. Mol. Biol.* 43, 27–35. doi: 10.1016/0960-0760(92)90184-K
- Roesler, R., Henriques, J. A. P., and Schwartsmann, G. (2006). Gastrinreleasing peptide receptor as a molecular target for psychiatric and neurological disorders. CNS Neurol. Disord. Drug Targets 5, 197–204. doi: 10.2174/187152706776359673
- Rosenbaum, D. M., Rasmussen, S. G. F., and Kobilka, B. K. (2009). The structure and function of G-protein-coupled receptors. *Nature* 459, 356–363. doi: 10.1038/nature08144
- Ryppa, C., Mann-Steinberg, H., Fichtner, I., Weber, H., Satchi-Fainaro, R., Biniossek, M. L., et al. (2008). *In vitro* and *in vivo* evaluation of doxorubicin conjugates with the divalent peptide E-c(RGDfK)2 that targets integrin alphavbeta3. *Bioconjug. Chem.* 19, 1414–1422. doi: 10.1021/bc80 0117r
- Saito, G., Swanson, J. A., and Lee, K.-D. (2003). Drug delivery strategy utilizing conjugation via reversible disulfide linkages: role and site of cellular reducing activities. *Adv. Drug Deliv. Rev.* 55, 199–215. doi: 10.1016/S0169-409X(02)00179-5
- Sancho, V., Di Florio, A., Moody, T. W., and Jensen, R. T. (2011). Bombesin receptor-mediated imaging and cytotoxicity: review and current status. *Curr. Drug Deliv.* 8, 79–134. doi: 10.2174/156720111793663624
- Schally, A. V., and Nagy, A. (1999). Cancer chemotherapy based on targeting of cytotoxic peptide conjugates to their receptors on tumors. *Eur. J. Endocrinol.* 141, 1–14. doi: 10.1530/eje.0.1410001
- Schirrmacher, E., Schirrmacher, R., Beck, C., Mier, W., Trautman, N., and Rösch, F. (2003). Synthesis of a Tyr3-octreotate conjugated closo-carborane [HC2B10H10]: a potential compound for boron neutron capture therapy. *Tetrahedron Lett.* 44, 9143–9145. doi: 10.1016/j.tetlet.2003.10.048
- Schmitt, H. W., Block, R. C., and Bailey, R. L. (1960). Total neutron cross section of B10 in the thermal neutron energy range. *Nucl. Phys.* 17, 109–115. doi: 10.1016/0029-5582(60)90105-X
- Scholz, M., and Hey-Hawkins, E. (2011). Carbaboranes as pharmacophores: properties, synthesis, and application strategies. *Chem. Rev.* 111, 7035–7062. doi: 10.1021/cr200038x
- Schottelius, M., and Wester, H.-J. (2009). Molecular imaging targeting peptide receptors. *Methods* 48, 161–177. doi: 10.1016/j.ymeth.2009.03.012
- Schroeder, R. P. J., van Weerden, W. M., Bangma, C., Krenning, E. P., and Jong, M. D. (2009). Peptide receptor imaging of prostate cancer with radiolabelled bombesin analogues. *Methods* 48, 200–204. doi: 10.1016/j.ymeth.2009.04.002
- Schwartz, M. A., Schaller, M. D., and Ginsberg, M. H. (1995). Integrins: emerging paradigms of signal transduction. Annu. Rev. Cell Dev. Biol. 11, 549–599. doi: 10.1146/annurev.cb.11.110195.003001
- Scopinaro, F., Varvarigou, A. D., Ussof, W., Vincentis, G., de, Sourlingas, T. G., Evangelatos, G. P., et al. (2002). Technetium labeled bombesin-like peptide: preliminary report on breast cancer uptake in patients. *Cancer Biother. Radiopharm.* 17, 327–335. doi: 10.1089/10849780260179297

- Scopinaro, F., Vincentis, G., de, Corazziari, E., Massa, R., Osti, M., Pallotta, N., et al. (2004). Detection of colon cancer with 99mTc-labeled bombesin derivative (99mTc-leu13-BN1). *Cancer Biother. Radiopharm.* 19, 245–252. doi: 10.1089/108497804323072020
- Seitz, S., Schally, A. V., Treszl, A., Papadia, A., Rick, F., Szalontay, L., et al. (2009). Preclinical evaluation of properties of a new targeted cytotoxic somatostatin analog, AN-162 (AEZS-124), and its effects on tumor growth inhibition. *Anticancer. Drugs* 20, 553–558. doi: 10.1097/CAD.0b013e32832d190b
- Seshacharyulu, P., Ponnusamy, M. P., Haridas, D., Jain, M., Ganti, A. K., and Batra, S. K. (2012). Targeting the EGFR signaling pathway in cancer therapy. *Expert Opin. Ther. Targets* 16, 15–31. doi: 10.1517/14728222.2011.648617
- Shen, H., Hu, D., Du, J., Wang, X., Liu, Y., Wang, Y., et al. (2008). Paclitaxeloctreotide conjugates in tumor growth inhibition of A549 human non-small cell lung cancer xenografted into nude mice. *Eur. J. Pharmacol.* 601, 23–29. doi: 10.1016/j.ejphar.2008.10.035
- Siegel, R. L., Miller, K. D., and Jemal, A. (2019). Cancer statistics, 2019. CA Cancer J. Clin. 69, 7–34. doi: 10.3322/caac.21551
- Sköld, K., Gorlia, T., Pellettieri, L., Giusti, V., H.-,Stenstam, B., and Hopewell, J. W. (2010). Boron neutron capture therapy for newly diagnosed glioblastoma multiforme: an assessment of clinical potential: an assessment of clinical potential. Br. J. Radiol. 83, 596–603. doi: 10.1259/bjr/56953620
- Slice, L. W., Yee, H. F., and Walsh, J. H. (1998). Visualization of internalization and recycling of the gastrin releasing peptide receptor-green fluorescent protein chimera expressed in epithelial cells. *Recept Channels* 6, 201–212.
- Snyder, H. R., Reedy, A. J., and Lennarz, W. J. (1958). Synthesis of aromatic boronic acids. aldehydo boronic acids and a boronic acid analog of tyrosine 1. J. Am. Chem. Soc. 80, 835–838. doi: 10.1021/ja01537a021
- Söll, R. M., Dinger, M. C., Lundell, I., Larhammer, D., and Beck-Sickinger, A. G. (2001). Novel analogues of neuropeptide Y with a preference for the Y1-receptor. *Eur. J. Biochem.* 268, 2828–2837. doi: 10.1046/j.1432-1327.2001.02161.x
- Soloway, A. H., Hatanaka, H., and Davis, M. A. (1967). Penetration of brain and brain tumor. VII. Tumor-binding sulfhydryl boron compounds. J. Med. Chem. 10, 714–717. doi: 10.1021/jm00316a042
- Soloway, A. H., Tjarks, W., Barnum, B. A., Rong, F.-G., Barth, R. F., Codogni, I. M., et al. (1998). The chemistry of neutron capture therapy. *Chem. Rev.* 98, 1515–1562. doi: 10.1021/cr941195u
- Sriram, K., and Insel, P. A. (2018). G protein-coupled receptors as targets for approved drugs: how many targets and how many drugs? *Mol. Pharmacol.* 93, 251–258. doi: 10.1124/mol.117.111062
- Stangelberger, A., Schally, A. V., Letsch, M., Szepeshazi, K., Nagy, A., Halmos, G., et al. (2006). Targeted chemotherapy with cytotoxic bombesin analogue AN-215 inhibits growth of experimental human prostate cancers. *Int. J. Cancer* 118, 222–229. doi: 10.1002/ijc.21292
- Stern, L., Perry, R., Ofek, P., Many, A., Shabat, D., and Satchi-Fainaro, R. (2009). A novel antitumor prodrug platform designed to be cleaved by the endoprotease legumain. *Bioconjug. Chem.* 20, 500–510. doi: 10.1021/bc800448u
- Strebhardt, K., and Ullrich, A. (2008). Paul Ehrlich's magic bullet concept: 100 years of progress. Nat. Rev. Cancer 8, 473–480. doi: 10.1038/nrc2394
- Strosberg, J., El-Haddad, G., Wolin, E., Hendifar, A., Yao, J., Chasen, B., et al. (2017). Phase 3 trial of 177Lu-dotatate for midgut neuroendocrine tumors. N. Engl. J. Med. 376, 125–135. doi: 10.1056/NEJMoa1607427
- Sugahara, K. N., Teesalu, T., Karmali, P. P., Kotamraju, V. R., Agemy, L., Girard, O. M., et al. (2009). Tissue-penetrating delivery of compounds and nanoparticles into tumors. *Cancer Cell* 16, 510–520. doi: 10.1016/j.ccr.2009. 10.013
- Sun, L.-C., and Coy, D. H. (2011). Somatostatin receptor-targeted anti-cancer therapy. Curr. Drug Deliv. 8, 2–10. doi: 10.2174/156720111793663633
- Sun, L.-C., Mackey, L. V., Luo, J., Fuselier, J. A., and Coy, D. H. (2008). Targeted chemotherapy using a cytotoxic somatostatin conjugate to inhibit tumor growth and metastasis in nude mice. *Clin. Med. Oncol.* 2, 491–499. doi: 10.4137/CMO.S970
- Sun, X., Li, Y., Liu, T., Li, Z., Zhang, X., and Chen, X. (2017). Peptide-based imaging agents for cancer detection. Adv. Drug Deliv. Rev. 110-111, 38–51. doi: 10.1016/j.addr.2016.06.007
- Suzuki, M. (2019). Boron neutron capture therapy (BNCT): a unique role in radiotherapy with a view to entering the accelerator-based BNCT era. *Int. J. Clin. Oncol.* 25, 43–50. doi: 10.1007/s10147-019-01480-4

- Szlachcic, A., Zakrzewska, M., Lobocki, M., Jakimowicz, P., and Otlewski, J. (2016). Design and characteristics of cytotoxic fibroblast growth factor 1 conjugate for fibroblast growth factor receptor-targeted cancer therapy. *Drug Des. Devel. Ther.* 10, 2547–2560. doi: 10.2147/DDDT.S105896
- Tan, C. M. J., Green, P., Tapoulal, N., Lewandowski, A. J., Leeson, P., and Herring, N. (2018). The role of neuropeptide Y in cardiovascular health and disease. *Front. Physiol.* 9:01281. doi: 10.3389/fphys.2018.01281
- Tang, B., Yong, X., Xie, R., Li, Q.-W., and Yang, S.-M. (2014). Vasoactive intestinal peptide receptor-based imaging and treatment of tumors. *Int. J. Oncol.* 44, 1023–1031. doi: 10.3892/ijo.2014.2276
- Thorén, P. E. G., Persson, D., Lincoln, P., and Nordén, B. (2005). Membrane destabilizing properties of cell-penetrating peptides. *Biophys. Chem.* 114, 169–179. doi: 10.1016/j.bpc.2004.11.016
- Thundimadathil, J. (2012). Cancer treatment using peptides: current therapies and future prospects. J. Amino Acids. 2012: 967347. doi: 10.1155/2012/967347
- Uehara, H., González, N., Sancho, V., Mantey, S. A., Nuche-Berenguer, B., Pradhan, T., et al. (2011). Pharmacology and selectivity of various natural and synthetic bombesin related peptide agonists for human and rat bombesin receptors differs. *Peptides* 32, 1685–1699. doi: 10.1016/j.peptides.2011.06.017
- Valliant, J. F., Guenther, K. J., King, A. S., Morel, P., Schaffer, P., Sogbein, O. O., et al. (2002). The medicinal chemistry of carboranes. *Coord. Chem. Rev.* 232, 173–230. doi: 10.1016/S0010-8545(02)00087-5
- Valverde, I. E., Bauman, A., Kluba, C. A., Vomstein, S., Walter, M. A., and Mindt, T. L. (2013). 1,2,3-Triazoles as amide bond mimics: Triazole scan yields protease-resistant peptidomimetics for tumor targeting. *Angew. Chem. Int. Ed. Engl.* 52, 8957–8960. doi: 10.1002/anie.201303108
- van Essen, M., Krenning, E. P., Kam, B. L. R., de Jong, M., Valkema, R., and Kwekkeboom, D. J. (2009). Peptide-receptor radionuclide therapy for endocrine tumors. *Nat. Rev. Endocrinol.* 5, 382–393. doi: 10.1038/nrendo.2009.105
- van Schouwenburg, P. A., Bartelds, G. M., Hart, M. H., Aarden, L., Wolbink, G. J., and Wouters, D. (2010). A novel method for the detection of antibodies to adalimumab in the presence of drug reveals "hidden" immunogenicity in rheumatoid arthritis patients. J. Immunol. Methods 362, 82–88. doi: 10.1016/j.jim.2010.09.005
- Vargas, J. R., Stanzl, E. G., Teng, N. N. H., and Wender, P. A. (2014). Cellpenetrating, guanidinium-rich molecular transporters for overcoming efflux-mediated multidrug resistance. *Mol. Pharm.* 11, 2553–2565. doi: 10.1021/mp500161z
- Vhora, I., Patil, S., Bhatt, P., Gandhi, R., Baradia, D., and Misra, A. (2014). Receptor-targeted drug delivery: current perspective and challenges. *Ther. Deliv.* 5, 1007–1024. doi: 10.4155/tde.14.63
- Vlieghe, P., Lisowski, V., Martinez, J., and Khrestchatisky, M. (2010). Synthetic therapeutic peptides: science and market. *Drug Discov. Today* 15, 40–56. doi: 10.1016/j.drudis.2009.10.009
- Volante, M., Rosas, R., Allìa, E., Granata, R., Baragli, A., Muccioli, G., et al. (2008). Somatostatin, cortistatin and their receptors in tumours. *Mol. Cell. Endocrinol.* 286, 219–229. doi: 10.1016/j.mce.2007.12.002
- Vrettos, E. I., Mezo, G., and Tzakos, A. G. (2018). On the design principles of peptide-drug conjugates for targeted drug delivery to the malignant tumor site. *Beilstein J. Org. Chem.* 14, 930–954. doi: 10.3762/bjoc.14.80
- Wakui, H., Yamamoto, N., Kitazono, S., Mizugaki, H., Nakamichi, S., Fujiwara, Y., et al. (2014). A phase 1 and dose-finding study of LY2523355 (litronesib), an Eg5 inhibitor, in Japanese patients with advanced solid tumors. *Cancer Chemother. Pharmacol.* 74, 15–23. doi: 10.1007/s00280-014-2467-z
- Wall, M. E., and Wani, M. C. (1996). Camptothecin and taxol: from discovery to clinic. J. Ethnopharmacol. 51, 239–254. doi: 10.1016/0378-8741(95) 01367-9
- Wang, J., Li, S., Luo, T., Wang, C., and Zhao, J. (2012). Disulfide linkage: a potent strategy in tumor-targeting drug discovery. *Curr. Med. Chem.* 19, 2976–2983. doi: 10.2174/092986712800672030
- Wang, Z. L., Bennet, W. M., Wang, R. M., Ghatei, M. A., and Bloom, S. R. (1994). Evidence of a paracrine role of neuropeptide-Y in the regulation of insulin release from pancreatic islets of normal and dexamethasone-treated rats. *Endocrinology* 135, 200–206. doi: 10.1210/endo.135.1.8013354
- Weidmann, A. G., Komor, A. C., and Barton, J. K. (2014). Targeted chemotherapy with metal complexes. *Comments Mod. Chem. A Comments Inorg. Chem.* 34, 114–123. doi: 10.1080/02603594.2014.890099

- Weis, W. I., and Kobilka, B. K. (2018). The molecular basis of G protein-coupled receptor activation. Annu. Rev. Biochem. 87, 897–919. doi: 10.1146/annurev-biochem-060614-033910
- Wettstein, J. G., Earley, B., and Junien, J. L. (1995). Central nervous system pharmacology of neuropeptide Y. *Pharmacol. Ther.* 65, 397–414. doi: 10.1016/0163-7258(95)98598-K
- Wieduwilt, M. J., and Moasser, M. M. (2008). The epidermal growth factor receptor family: biology driving targeted therapeutics. *Cell. Mol. Life Sci.* 65, 1566–1584. doi: 10.1007/s00018-008-7440-8
- Wittig, A., Sauerwein, W. A., and Coderre, J. A. (2000). Mechanisms of transport of p -borono-phenylalanine through the cell membrane *in vitro. Radiat. Res.* 153, 173–180. doi: 10.1667/0033-7587(2000)153[0173:motopb]2.0.co;2
- Worm, D. J., Els-Heindl, S., Kellert, M., Kuhnert, R., Saretz, S., Koebberling, J., et al. (2018). A stable meta-carborane enables the generation of boronrich peptide agonists targeting the ghrelin receptor. *J. Pept. Sci.* 24:e3119. doi: 10.1002/psc.3119
- Worm, D. J., Hoppenz, P., Els-Heindl, S., Kellert, M., Kuhnert, R., Saretz, S., et al. (2019). Selective neuropeptide Y conjugates with maximized carborane loading as promising boron delivery agents for boron neutron capture therapy. J. Med. Chem. 63, 2358–2371. doi: 10.1021/acs.jmedchem.9b01136
- Wu, G., Barth, R. F., Yang, W., Chatterjee, M., Tjarks, W., Ciesielski, M. J., et al. (2004). Site-specific conjugation of boron-containing dendrimers to anti-EGF receptor monoclonal antibody cetuximab (IMC-C225) and its evaluation as a potential delivery agent for neutron capture therapy. *Bioconjug. Chem.* 15, 185–194. doi: 10.1021/bc0341674
- Wu, G., Yang, W., Barth, R. F., Kawabata, S., Swindall, M., Bandyopadhyaya, A. K., et al. (2007). Molecular targeting and treatment of an epidermal growth factor receptor-positive glioma using boronated cetuximab. *Clin. Cancer Res.* 13, 1260–1268. doi: 10.1158/1078-0432.CCR-06-2399
- Wu, Z., Martinez-Fong, D., Trédaniel, J., and Forgez, P. (2012). Neurotensin and its high affinity receptor 1 as a potential pharmacological target in cancer therapy. *Front. Endocrinol.* 3:184. doi: 10.3389/fendo.2012.00184
- Xiao, H., Verdier-Pinard, P., Fernandez-Fuentes, N., Burd, B., Angeletti, R., Fiser, A., et al. (2006). Insights into the mechanism of microtubule stabilization by Taxol. *Proc. Natl. Acad. Sci. U.S.A.* 103, 10166–10173. doi: 10.1073/pnas.0603704103
- Yang, W., Barth, R. F., Wu, G., Huo, T., Tjarks, W., Ciesielski, M., et al. (2009a). Convection enhanced delivery of boronated EGF as a molecular targeting agent for neutron capture therapy of brain tumors. *J. Neurooncol.* 95, 355–365. doi: 10.1007/s11060-009-9945-x
- Yang, W., Barth, R. F., Wu, G., Kawabata, S., Sferra, T. J., Bandyopadhyaya, A. K., et al. (2006). Molecular targeting and treatment of EGFRvIII-positive gliomas using boronated monoclonal antibody L8A4. *Clin. Cancer Res.* 12, 3792–3802. doi: 10.1158/1078-0432.CCR-06-0141
- Yang, W., Barth, R. F., Wu, G., Tjarks, W., Binns, P., and Riley, K. (2009b). Boron neutron capture therapy of EGFR or EGFRvIII positive gliomas using either boronated monoclonal antibodies or epidermal growth factor as molecular targeting agents. *Appl. Radiat. Isot.* 67, 328–331. doi: 10.1016/j.apradiso.2009.03.030
- Yang, W., Wu, G., Barth, R. F., Swindall, M. R., Bandyopadhyaya, A. K., Tjarks, W., et al. (2008). Molecular targeting and treatment of composite EGFR

and EGFRvIII-positive gliomas using boronated monoclonal antibodies. *Clin. Cancer Res.* 14, 883–891. doi: 10.1158/1078-0432.CCR-07-1968

- Yarden, Y., and Pines, G. (2012). The ERBB network: at last, cancer therapy meets systems biology. *Nat. Rev. Cancer* 12, 553–563. doi: 10.1038/ nrc3309
- Yi, M., Li, H., Wu, Z., Yan, J., Liu, Q., Ou, C., et al. (2018). A promising therapeutic target for metabolic diseases: neuropeptide Y receptors in humans. *Cell. Physiol. Biochem.* 45, 88–107. doi: 10.1159/000486225
- Yoo, J., Park, C., Yi, G., Lee, D., and Koo, H. (2019). Active targeting strategies using biological ligands for nanoparticle drug delivery systems. *Cancers* 11:640. doi: 10.3390/cancers11050640
- Yu, S. S., Athreya, K., Liu, S. V., Schally, A. V., Tsao-Wei, D., Groshen, S., et al. (2017). A phase II trial of AEZS-108 in castration- and taxane-resistant prostate cancer. *Clin. Genitourin. Cancer* 15, 742–749. doi: 10.1016/j.clgc.2017.06.002
- Yu, Z., Ananias, H. J., Carlucci, G., Hoving, H. D., Helfrich, W., Dierckx, Rudi, A. J. O., et al. (2013). An update of radiolabeled bombesin analogs for gastrin-releasing peptide receptor targeting. *Curr. Pharm. Des.* 19, 3329–3341. doi: 10.2174/1381612811319180015
- Yulyaningsih, E., Zhang, L., Herzog, H., and Sainsbury, A. (2011). NPY receptors as potential targets for anti-obesity drug development. *Br. J. Pharmacol.* 163, 1170–1202. doi: 10.1111/j.1476-5381.2011.01363.x
- Zhang, L., Bijker, M. S., and Herzog, H. (2011). The neuropeptide Y system: pathophysiological and therapeutic implications in obesity and cancer. *Pharmacol. Ther.* 131, 91–113. doi: 10.1016/j.pharmthera.2011.03.011
- Zhang, L., and Bulaj, G. (2012). Converting peptides into drug leads by lipidation. Curr. Med. Chem. 19, 1602–1618. doi: 10.2174/0929867127999 45003
- Zhou, X. J., and Rahmani, R. (1992). Preclinical and clinical pharmacology of vinca alkaloids. *Drugs* 44, 1–16. doi: 10.2165/00003495-199200444-00002
- ZoptEC (ZoptEC) (2013). Zoptarelin Doxorubicin (AEZS 108) as Second Line Therapy for Endometrial Cancer (ZoptEC): Identifier: NCT01767155. Available online at: https://clinicaltrials.gov/ (accessed October 21, 2019).
- Zou, Y., Xia, Y., Meng, F., Zhang, J., and Zhong, Z. (2018). GE11-directed functional polymersomal doxorubicin as an advanced alternative to clinical liposomal formulation for ovarian cancer treatment. *Mol. Pharm.* 15, 3664–3671. doi: 10.1021/acs.molpharmaceut.8b00024
- Zwanziger, D., Böhme, I., Lindner, D., and Beck-Sickinger, A. G. (2009). First selective agonist of the neuropeptide Y1-receptor with reduced size. J. Pept. Sci. 15, 856–866. doi: 10.1002/psc.1188

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2020 Hoppenz, Els-Heindl and Beck-Sickinger. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.