# **Archival Report**

# Peripheral Blood Cytokines as Markers of **Longitudinal Change in White Matter** Microstructure Following Inpatient Treatment for **Opioid Use Disorders**

Eduardo R. Butelman, Yuefeng Huang, Sarah G. King, Pierre-Olivier Gaudreault, Ahmet O. Ceceli, Greg Kronberg, Flurin Cathomas, Panos Roussos, Scott J. Russo, Eric L. Garland, Rita Z. Goldstein, and Nelly Alia-Klein

#### **ABSTRACT**

BACKGROUND: Opioid use disorder (OUD) causes major public health morbidity and mortality. Although standardof-care treatment with medications for OUD (MOUDs) is available, there are few biological markers of the clinical process of recovery. Neurobiological aspects of recovery can include normalization of brain white matter (WM) microstructure, which is sensitive to cytokine signaling. Here, we determined whether blood-based cytokines can be markers of change in WM microstructure following MOUD.

METHODS: Inpatient individuals with heroin use disorder (iHUDs) (n = 21) with methadone or buprenorphine MOUD underwent magnetic resonance imaging (MRI) scans with diffusion tensor imaging (DTI) and provided ratings of drug cue-induced craving, arousal, and valence earlier in treatment (MRI1) and ≈14 weeks thereafter (MRI2). Healthy control participants (HCs) (n = 24) also underwent 2 MRI scans during a similar time interval. At MRI2, participants provided a peripheral blood sample for multiplex quantification of serum cytokines. We analyzed the correlation of a multitarget biomarker score (from a principal component analysis of 19 cytokines that differed significantly between iHUDs and HCs) with treatment-related change in DTI metrics (ΔDTI; MRI2 – MRI1).

RESULTS: The cytokine biomarker score was negatively correlated with ΔDTI metrics in frontal, frontoparietal, and corticolimbic WM tracts in iHUDs but not in HCs. Also, serum levels of specific cytokines in the cytokine biomarker score, including the interleukin-related oncostatin M (OSM), similarly correlated with ΔDTI metrics in iHUDs but not in HCs. Serum levels of other specific cytokines were negatively correlated with changes in cue-induced craving and arousal in the iHUDs.

CONCLUSIONS: Specific serum cytokines, studied alone or as a group, may serve as accessible biomarkers of WM microstructure changes and potential recovery in iHUDs undergoing treatment with MOUD.

https://doi.org/10.1016/j.bpsgos.2025.100480

Opioid use disorder (OUD) causes major morbidity and mortality (1,2) and constitutes a national epidemic. While treatment with medications for OUD (MOUDs) (e.g., methadone or buprenorphine) is the effective standard of care (2,3), many individuals with heroin use disorder (iHUDs) either do not receive treatment, drop out, or relapse (3,4). Insufficient knowledge about potential biomarkers of treatment outcomes (inclusive of recovery) of people receiving MOUD severely limits the development of personalized approaches and clinical stratification (5,6). Therefore, there is a critical unmet need for the identification of new and accessible biologically based markers to monitor change during treatment.

Chronic exposure to mu opioid receptor (MOR) agonists such as heroin, prescription opioids, morphine or fentanyl causes dysregulation of several peripheral and central targets, including cytokine systems (7-12). A small number of studies

have reported potential differences in cytokine profiles based on the type of MOR agonist exposure (for example, with multiple daily use of short-acting compounds such as heroin vs. stable daily dosing with an MOUD such as methadone) (13,14). Likely based on pharmacodynamic or pharmacokinetic factors, such differences need to be examined more fully for a broader set of cytokine targets. Cytokines are a pleiotropic group of signaling molecules that act on cognate receptors in peripheral blood mononuclear cells, other leukocytes (15), and central neurons and glia (8,16,17). For example, several studies have shown that specific cytokines affect glial and white matter (WM) functions and responses to different insults (18,19). Specific leukocyte subsets or cytokines from blood can also affect central neuroglial processes by migration through the blood-brain barrier (20-22). The major canonical cytokine families include interleukins (ILs), chemokines, growth factors,

and tumor necrosis factor-related proteins. We recently reported that serum analysis of a multiplex cytokine and inflammatory panel of proteins revealed significant differences in iHUDs (n = 21) receiving MOUD compared with healthy control participants (HCs) (n = 24) (23). Specifically, of the 19 cytokines (from different canonical families) that differed significantly between the groups, all but 3 exhibited iHUD > HC levels, and a multitarget cytokine biomarker score based on a principal component analysis (PCA) of these 19 cytokines robustly differentiated iHUDs from HCs, while adjusting for major demographic and clinical factors (23). Other studies have similarly shown changes in blood levels of several cytokines in iHUDs compared with HCs and changes in messenger RNA (mRNA) expression of several cytokine system genes, both in peripheral blood mononuclear cells and in the brain (9,15). These central and peripheral mechanisms may be broadly conserved because levels of several cytokines are altered in the blood of rats with extended-access fentanyl self-administration (24), and there are alterations in expression of several cytokinerelated genes in the striatum of mice that chronically selfadministered oxycodone (8). Importantly, preclinical studies have shown that the blood-brain barrier can also become functionally compromised after chronic MOR agonist exposure or withdrawal, potentially exacerbating neuroinflammatory effects (17,25). The brain's WM is composed primarily of myelinated neuronal tracts (axonal bundles), which underlie functional connectivity networks (26,27). Myelination is an active regulatory process based on interactions between glia, especially oligodendrocytes and oligodendrocyte progenitor cells (as well as microglia and astrocytes), and neurons (28,29) that affect neurotransmission and neuroplasticity (29). Crucially, myelination and neuroinflammation are governed by complex signaling networks, including cytokines and cognate receptors, as well as other proteins (30,31). Of relevance to HUD, a recent preclinical study showed that mu opioid agonist-mediated reward depended in part on interactions between neurons and oligodendrocytes (32). Also, there is robust mRNA expression of OPRM1 (the gene that encodes MORs) in brain glia of humans post mortem (11). Chronic exposure to MOR agonists causes transcriptional and functional changes not only in molecular targets in central nervous system neurons (11,33,34) but also in oligodendrocytes and other glia (11), especially for diverse cytokine and neuroinflammatory genes (11).

WM microstructure can be examined in living humans with diffusion tensor imaging (DTI) and related techniques (35,36). We and others have documented widespread deficits in WM microstructure in individuals with substance use disorders including HUD (encompassing reduced fractional anisotropy [FA]) (37-39) being correlated with enhanced disease severity (e.g., increased craving and longer periods of regular use) (37). FA a measure of restriction in water molecule diffusion to a primary direction, often considered as a measure of normal WM microstructural organization. Importantly, some of these WM microstructure impairments appear to be dynamic, recovering in time with treatment. Specifically, we showed that following inpatient treatment of approximately 14 weeks with an MOUD (daily maintenance with methadone or buprenorphine), FA increased, especially in frontal callosal projections, association tracts, and the genu and body of the corpus

callosum, and this increase was associated with reduced craving in iHUDs (40). Another recent longitudinal study of iHUDs (with 2 DTI scans separated by 8 months) found that prolonged abstinence was associated with an increase in mean FA in specific frontostriatal tracts, and this was correlated with a decrease in craving scores (41).

Brain levels of cytokines cannot be measured directly in living humans with currently available technologies. Furthermore, the passage of specific cytokines between the periphery and the brain depends on different mechanisms (42,43) and may vary dynamically during neuroinflammatory states (44). Therefore, it cannot be assumed that levels of cytokines from blood are directly reflective of their levels in the brain (or across different brain areas), and therefore research is needed to examine associations among blood levels of specific cytokines and change in brain WM. Here, we examined whether the longitudinal changes in WM microstructure found in iHUDs receiving MOUD would be correlated with readily accessible blood-based cytokine biomarkers. We hypothesized that levels of serum cytokines from a multiplex assay would be correlated with changes in WM microstructure and drug cue-induced subjective ratings (e.g., of craving, arousal, and valence) during MOUD treatment. The study was also intended to illuminate specific cytokine targets for future mechanistic evaluation of WM microstructure integrity in the context of OUD.

#### **METHODS AND MATERIALS**

#### **Participants and Diagnostic Procedures**

iHUDs (n=21) were recruited from an inpatient treatment facility in New York City, and the HCs (n=24) were recruited from the surrounding community. The study was approved by the Institutional Review Board of the Icahn School of Medicine at Mount Sinai; participants gave written informed consent. A highly trained study team supervised by clinical psychologists conducted participant evaluations that included the Mini-International Neuropsychiatric Interview (version 7.0.0 for DSM-5) (45) and the Addiction Severity Index (46). Medication exposure was recorded for all participants and did not include compounds with major immune effects (23).

Inclusion criteria for all participants included ages 18 to 65 years and having the capacity to understand and provide informed consent in English. Inclusion criteria for iHUDs included meeting DSM-5 criteria for OUD, with heroin as the drug of choice and the main reason for treatment. The iHUDs were inpatients receiving methadone (n = 17) or buprenorphine (n = 4) MOUD daily maintenance. Exclusion criteria for all participants included the following: 1) present or past history of a DSM-5 diagnosis of psychotic disorder or neurodevelopmental disorder; 2) history of head trauma with loss of consciousness (>30 minutes); 3) history of neurological disorders including seizures; 4) current use of any medication (with the exception of MOUD in the iHUDs) that may affect neurological functions; 5) current medical illness and/or evident infection including cardiovascular disease, as well as metabolic, endocrine, oncological, or autoimmune diseases and infectious diseases common in individuals with substance use disorders including Hepatitis B and C or HIV/AIDS (we did not exclude iHUDs for a history of other substance use

disorders [e.g., alcohol, cannabis, cocaine] or other psychiatric disorders with high rates of comorbidity with substance use disorders [e.g., depression, anxiety, posttraumatic stress disorder]); 6) magnetic resonance imaging (MRI) contraindications including any metallic implants, pacemaker device, or pregnancy; 7) a positive breathalyzer test for alcohol; and 8) MRI quality assurance, including the presence of incidental findings in the WM as indicated by a radiologist, insufficient quality of diffusion data, or an MRI session that did not include a diffusion sequence. Exclusion criteria for HCs included a lifetime history of any substance use disorders, including for alcohol, or a positive urine test for any drug of abuse. The HCs were recruited and assessed by the same study team with the same tools, but HCs did not complete scales specific to persons with HUD.

#### **Behavioral Variables**

Demographic (age, sex, race, body mass index) and clinical variables (sleep and drug use measures) are shown in Table 1. Because depression, anxiety, and stress exposure are comorbid with OUD and can be associated with change in cytokine mechanisms (22,47,48), participants completed the Beck Depression Inventory-II (49,50), Beck Anxiety Inventory (51), and Perceived Stress Scale (52). In the iHUDs, we also examined methadone/buprenorphine dose (either from documented report or self-report) and duration of current heroin abstinence from the daily interview. Age of first heroin use, age of onset of regular heroin use, and number of years of regular heroin use (excluding abstinence periods) were also assessed (Table 1). Following each of the 2 imaging scans (described below), we quantified self-reported ratings of cue-induced craving, valence, and arousal in response to drug-related pictures using a 5-point scale (with increasing numbers indicating greater ratings for these measures), as described previously (53).

## **Cytokine Assays**

Blood samples were obtained by venipuncture on the day of the second MRI scan (MRI2) or on the nearest practical date (mean interval between MRI2 and blood sample = 1.3 days across all participants; n = 45). The blood sample was obtained during the time range of 09:00 AM to 5:00 PM (i.e., at least 1 hour after the daily MOUD administration in the iHUDs). Samples were centrifuged (10 minutes; 1200g) within ≈1 hour of collection, and serum was stored at -80 °C. Serum samples were analyzed with the Olink Target 96 Inflammation panel (Olink), following manufacturer's instructions (34), by the Human Immune Monitoring Center at the Icahn School of Medicine at Mount Sinai. This validated panel measures 92 targets, including cytokines from different canonical families and other inflammation-related proteins (full target list at https://olink. com/products-services/target/inflammation/). The assay provides relative quantification in normalized protein expression (NPX) separately for each target on a log<sub>2</sub> scale; therefore a 1unit NPX change indicates a 2-fold difference in protein levels. Targets with ≥50% of samples below the limit of detection within either group were not analyzed (47). Of 92 targets in the multiplex, 14 were excluded for this reason. The remaining 78 targets, including individual values below the limit of detection as in previous studies (54), were analyzed (23). Also, individual outliers ( $>\pm3$  SDs from the group mean) were removed from the analyses and were replaced by imputation for the PCA (using missMDA in R; see below) (55).

#### **MRI Acquisition and DTI**

All participants were scanned twice (MRI1 and MRI2), as part of a larger study, as recently described (40). Briefly, MRI acquisition was performed using a Siemens 3T Skyra scanner (Siemens Healthineers AG) with a 32-channel head coil. Diffusion echo-planar sequence was acquired with opposite phase encoding along the left-right axis, monopolar diffusion encoding with 128 diffusion-weighted images (2  $\times$  64 for each encoding phase) at single-shell maximum  $b = 1500 \text{ s/mm}^2$ , 13 reference images at  $b = 0 \text{ s/mm}^2$ , field of view (FOV) = 882  $\times$ 1044 mm, repetition time (TR) = 3650 ms, echo time (TE) = 87 ms, bandwidth = 1485 Hz/px, 80° flip angle, multiband = 3, no in-plane acceleration, and 1.8-mm isometric voxel size, with a total acquisition time of approximately 7 minutes. Anatomical T1-weighted structural images were acquired using the following parameters: 3-dimensional magnetization-prepared rapid acquisition gradient-echo (MPRAGE) sequence with  $FOV = 256 \times 256 \times 179 \text{ mm}^3$ , 0.8-mm isotropic resolution, TR/ TE/inversion time = 2400/2.07/1000 ms, 8° flip angle with binomial (1, −1) fat saturation, 240-Hz/px bandwidth, 7.6 ms echo spacing, and in-plane acceleration (generalized autocalibrating partially parallel acquisitions) factor of 2.

# Statistical Analyses for Phenotypic Variables and Cue-Induced Subjective Ratings

Table 1 shows the distribution of sex and race across groups, analyzed with Fisher's exact tests. Demographic/clinical data (body mass index, age, sleep hours, depression, anxiety, and perceived stress) were compared across groups with Mann-Whitney U tests. Drug cue–induced subjective craving, arousal and valence ratings were analyzed as  $\Delta$ values (i.e., MRI2 – MRI1), where a negative  $\Delta$ value indicates a decrease in a specific rating from MRI1 to MRI2. Pearson correlations between levels of 19 cytokines (see below) and these  $\Delta$ drug cue–induced ratings were examined (53). Normality of variables was examined with the D'Agostino and Pearson test, followed by examination of quantile-quantile plots, if necessary. The overall alpha level of significance was set at the p = .05 level.

### **Quantification of Individual Cytokines and PCA**

NPX ( $\log_2$  scale) were analyzed with Wilcoxon's rank-sum tests for group differences; p values were corrected for multiple comparisons with the false discovery rate (FDR) (5% cutoff level) approach (56). After identifying 29 targets that showed significant group differences from the previous step (including FDR correction), data from the 19 targets from canonical cytokine families that differed significantly between the 2 groups (these were primarily elevated in iHUDs vs. HCs) were entered into a PCA for dimensionality reduction (47,57), using z scores to standardize values. Because PCA requires complete data, individual outliers (i.e., greater than  $\pm 3$  SDs of each group's mean) were replaced by multiple imputation (missMDA procedure in R) (55). The frequency of imputed values was

**Table 1. Demographic and Clinical Variables** 

Variable	iHUD, <i>n</i> = 21	HC, <i>n</i> = 24	Group Differences, Mann-Whitney U or Fisher's Exact Test
Age, Years	42.0 (38.5–45.5)	40.7 (36.0–45.4)	NS
Sex, Female/Male	4/17	9/15	Fisher's exact test, NS
Race, Black/Other/White	0/3/18	8/4/12	Fisher's exact test, p = .006
Education, Years	12.1 (11.2–12.9)	16.6 (15.3–17.9)	<i>U</i> = 45.5, <i>p</i> < .0001
Body Mass Index at MRI2	32.3 (29.4–35.3)	26.7 (24.9–28.6)	<i>U</i> = 110, <i>p</i> = .0009
Sleep Hours in Night Prior to MRI2	6.54 (5.6–7.5)	6.51 (5.9–7.2)	NS
PSS-10 Perceived Stress Scores at MRI2, Scale Range: 0-40	20.5 (17.7–23.3), n = 20	12.3 (9.5–16.0)	<i>U</i> = 104, <i>p</i> = .001
BDI-II Depression Scores at MRI2, Scale Range: 0-63	15.5 (9.9–21.2), n = 19	3.8 (1.8–5.7)	<i>U</i> = 78, <i>p</i> = .0001
BAI Anxiety Scores at MRI2, Scale Range: 0-63	14.1 (8.8–19.4), n = 20	2.8 (1.1–4.4)	<i>U</i> = 85.5, <i>p</i> = .0001
Age of Onset of First Use of Heroin, Years	23.6 (17.4–27.8), n = 20	NA	NA
Age of Onset of Regular Use of Heroin, Years	25.1 (21.2–28.9), n = 20	NA	NA
Years of Regular Heroin Use	8.5 (5.9–11.2), n = 19	NA	NA
Heroin Abstinence Duration, Days	198 (130–268)	NA	NA
Methadone Daily Dose, mg	91.9 (63–121), n = 17	NA	NA
Years of Regular Alcohol Use	12.3 (5.9–18.6), n = 17	13.2 (6.6–19.8), n = 18	NS

Values are presented as n or mean (95% CI). Differing ns reflect missing data for specific variables.

BAI, Beck Anxiety Inventory; BDI-II, Beck Depression Inventory-II; HC, healthy control participant; iHUD, individual with heroin use disorder; MRI2, second magnetic resonance imaging scan; NA, not applicable; NS, nonsignificant; PSS, Perceived Stress Scale.

relatively low. Specifically, 3 targets in the HC group (TNFRSF9, CCL19, and CXCL9) each had 1 imputed data point (of a total n per target = 24). In the iHUD group, 3 targets (IL-6, CCL19, and CXCL9) also each had 1 imputed value (of a total n per target = 21). Nineteen principal components were calculated in the PCA algorithm, using a 95% threshold for significance and 1000 Monte Carlo simulations (GraphPad Prism). The complete results for these participants were reported recently (23). Scores for the first PC (PC1) accounted for the greatest proportion of variability (40.9%), and the individual PC1 scores were regressed against the whole-brain WM changes (i.e.,  $\Delta$ DTI for the 4 metrics; see below).

## Analyses of Change in WM Metrics During Treatment

The preprocessing of the diffusion MRI data was conducted according to established pipelines with MRtrix3 (58) and FSL 6.0 toolboxes, as fully described (37,40). Tensor-based quantitative maps of diffusion metrics including FA, mean diffusivity (MD), axial diffusivity (AD), and radial diffusivity (RD) were generated and used to perform tract-based spatial statistics (39). Whole-brain voxelwise analyses across all participants were conducted by first creating a WM skeleton and registered to the MNI152 standard space. Separate whole-brain correlation analyses were then performed between the 4  $\Delta$ DTI metrics (i.e., FA, MD, AD, RD) and the mean-centered cytokine PC1 scores (Figure S1) to test for slope differences between the iHUD and HC groups. We also tested the correlations between cytokine PC1 scores and the 4 DTI metrics at MRI1 and MRI2 separately for completeness. Within-group correlations were performed when significant group differences in slopes were detected. Analysis was performed via the FSL tool "Randomise," a general linear model for nonparametric permutation inferences via 10,000 permutations. In a follow-up, we repeated the same analyses with NPX (on a log<sub>2</sub> scale) from the individual cytokines from each canonical family with the highest PC1 loadings (CCL7, OSM, CSF1, TNFRSF9) (23) (Figure S1). We also performed correlation analyses between the Δmaps of the 4 DTI metrics and intracranial volume (estimated by FreeSurfer from the individual T1-weighted structural scan), education, age, sex, and body mass index to rule out confounding factors. All DTI analyses accounted for multiple comparisons by using the threshold-free cluster enhancement correction (59), which enhances areas of signal that exhibit spatial contiguity, to better discriminate clusters.

### **RESULTS**

As shown in Table 1, the groups were well matched on age, sex, sleep hours the night before the second scan, and years of alcohol use but different on racial distribution, education (iHUD < HC), body mass index, depression, anxiety, and perceived stress scores (iHUD > HC).

#### **Cytokine Data**

As recently reported (23), 19 cytokines were significantly different between the iHUD and HC groups, and these were analyzed with PCA (also see the Supplement, Table S1 and Figure S1). Of these 19 cytokines, 16 had iHUD > HC levels, and 3 had the opposite profile (HC > iHUD).

### Correlations of Cytokine PC1 Scores With Longitudinal Changes in DTI Metrics

We found significant group differences only in correlations between cytokine PC1 scores and the changes in DTI metrics (i.e., MRI2 – MRI1  $\Delta$  scores) (Figure 1) but not with the absolute DTI metrics measured at either MRI1 or MRI2. Specifically, lower PC1 scores were correlated with overall increases in FA, MD, and AD following inpatient treatment across distributed WM anatomical connectivity networks that represent primarily frontal, frontoparietal, and corticolimbic tracts

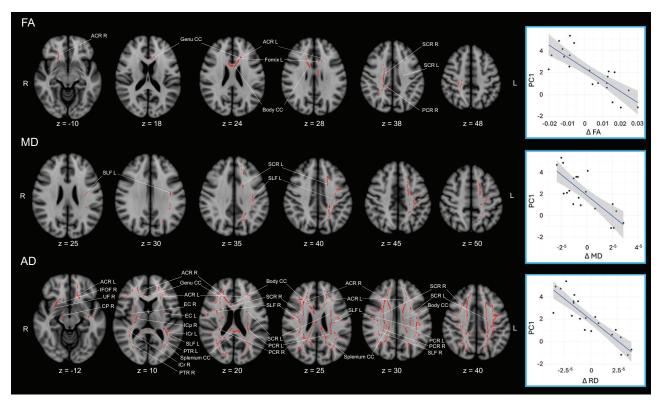


Figure 1. Cytokine first principal component (PC1) score correlations with change in diffusion tensor imaging (DTI) metrics across longitudinal scans (ΔDTI) (i.e., the second magnetic resonance imaging [MRI] scan – the first MRI scan). Binarized tracts that overlay the significant correlations between the PC1 score (of the 19 cytokines that showed group differences in individuals with heroin use disorder [iHUDs] vs. healthy control participants) and the changes in DTI metrics (fractional anisotropy [FA], mean diffusivity [MD], axial diffusivity [AD]; nonsignificant with radial diffusivity [RD]) (axial images). The negative correlations in the iHUD group are shown on the right (examining all significant voxels). ACR, anterior corona radiata; CC, corpus callosum; CP, midbrain cerebral peduncle; EC, external capsule; ICp, posterior limb of the internal capsule; ICr, retrolenticular part of the internal capsule; IFOF, inferior fronto-occipital fasciculus; L, left; PCR, posterior corona radiata; PTR, posterior thalamic radiation; R, right; SCR, superior corona radiata; SLF, superior longitudinal fasciculus; UF, uncinate fasciculus.

(encompassing the genu and body of the corpus callosum, fornix, anterior and posterior corona radiata, and superior longitudinal fasciculus for  $\Delta FA$  and in the superior longitudinal fasciculus and superior corona radiata for ΔMD; correlations with  $\Delta AD$  were observed in similar regions, as well as some more posterior cortical tracts). No correlation was found between cytokine PC1 scores and the other DTI metric, ΔRD (the maximum 1 - p value was .58). In a follow-up, we found that cytokine PC1 scores were significantly correlated with  $\Delta$ MD in 3187 voxels, 2700 (84.7%) of which overlapped with the voxels with significant correlations with ΔAD (not shown). Therefore, this shows considerable spatial congruence for the correlation of cytokine PC1 scores and ΔMD or ΔAD. Other follow-up analyses within the HC and iHUD groups revealed that the slope differences mentioned above were driven by significant negative correlations in the iHUD group and null results in the HC group.

To explore possible specific targets involved in the findings mentioned above (as opposed to an overall cytokine PC1 score), we examined levels of the cytokines with the greatest PC1 loading from each canonical family for their correlations with  $\Delta DTI$  metrics (Figures 2–4; also see Table S1 and Figure S1). Among these representative targets, levels of OSM were negatively correlated with  $\Delta FA$  and  $\Delta AD$  (Figure 2), levels

of CSF1 were negatively correlated with  $\Delta$ MD and  $\Delta$ AD (Figure 3), and levels of MP3/CCL7 were negatively correlated with  $\Delta$ AD (Figure 4) in iHUDs only. These correlations occurred over largely overlapping regions. No significant correlations were observed with TNFRSF9 (not shown).

### Change in Cue-Induced Ratings (MRI2 - MRI1)

First, we examined whether changes in drug cue-induced ratings of craving, arousal, and valence were correlated with the overall cytokine PC1 score. Only cue-induced arousal showed a significant negative correlation with PC1 scores (Pearson's r = -0.46, uncorrected p = .034, did not survive FDR correction). However, we found that change in some cueinduced ratings was negatively correlated with levels of specific cytokines. Thus, change in cue-induced craving was negatively correlated with levels of CCL19 (Pearson's r = -0.67, uncorrected p = .001, FDR-corrected p = .019) (Figure 5A) such that a decrease in cue-induced craving (i.e., MRI2 - MRI1) was correlated with greater CCL19 levels. Also, change in cue-induced arousal was negatively correlated with levels of CCL2 (Pearson's r = -0.68, uncorrected p = .001, FDR-corrected p = .011) (Figure 5B) such that a decrease in cue-induced arousal was correlated with greater CCL2 levels. None of the correlations with changes in cue-induced valence

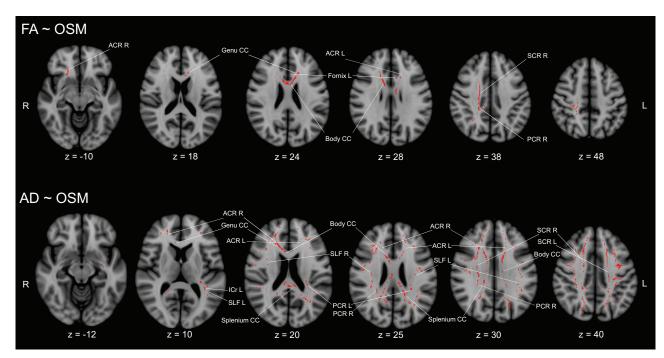


Figure 2. Correlation of serum OSM levels with change in diffusion tensor imaging metrics across longitudinal scans in the individuals with heroin use disorder (iHUDs) group. Binarized tracts for within-iHUD group negative correlations with serum OSM levels and ΔFA (top row) and ΔAD (bottom row). ACR, anterior corona radiata; AD, axial diffusivity; CC, corpus callosum; FA, fractional anisotropy; ICp, posterior limb of the internal capsule; L, left; PCR, posterior corona radiata; R, right; SCR, superior corona radiata; SLF, superior longitudinal fasciculus.

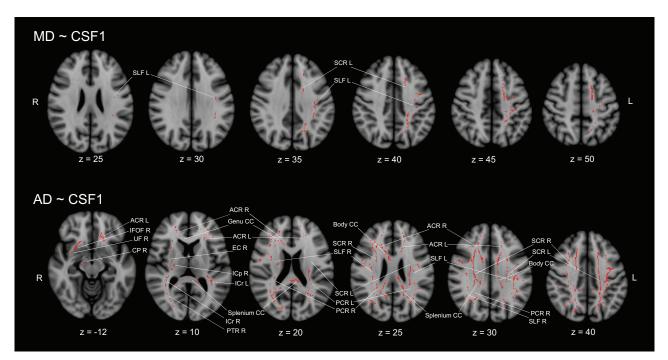


Figure 3. Correlation of serum CSF1 levels with change in diffusion tensor imaging metrics across longitudinal scans in the individuals with heroin use disorder (iHUDs) group. Binarized tracts for within-iHUD group negative correlations with serum CSF1 levels and ΔMD (top row) and ΔAD (bottom row). ACR, anterior corona radiata; AD, axial diffusivity; CC, corpus callosum; EC, external capsule; ICp, posterior limb of the internal capsule; ICr, retrolenticular part of the internal capsule; L, left; MD, mean diffusivity; PCR, posterior corona radiata; PTR, posterior thalamic radiation; R, right; SCR, superior corona radiata; SLF, superior longitudinal fasciculus.

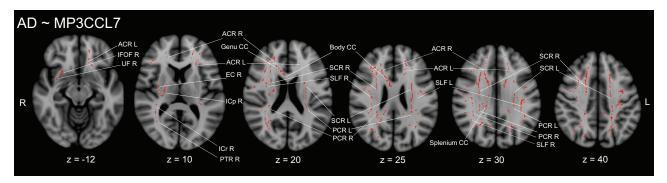


Figure 4. Correlation of MP3/CCL7 with change in diffusion tensor imaging metrics across longitudinal scans in the individuals with heroin use disorder (iHUDs) group. Binarized tracts for within-iHUD group negative correlations with serum MP3/CCL7 levels and ΔAD are shown. ACR, anterior corona radiata; AD, axial diffusivity; CC, corpus callosum; EC, external capsule; ICp, posterior limb of the internal capsule; ICr, retrolenticular part of the internal capsule; IFOF, inferior fronto-occipital fasciculus; L, left; PCR, posterior corona radiata; PTR, posterior thalamic radiation; R, right; SCR, superior corona radiata; SLF, superior longitudinal fasciculus; UF, uncinate fasciculus.

ratings survived FDR correction (full data are provided in Table S2). Neither PC1 scores nor levels of any of these 19 cytokines were correlated with cue-induced craving, arousal, or valence measured separately at either MRI1 or MRI2 after FDR correction (not shown).

#### **DISCUSSION**

The goal of this study was to examine the association between serum cytokine biomarkers and change in WM microstructure, together with drug cue–induced ratings, following  $\approx \! 14$  weeks of inpatient MOUD treatment in iHUDs compared with a similar interval in HCs. The main finding of this study is that a recently developed multitarget cytokine biomarker score (23) was correlated with  $\Delta DTI$  metrics in several WM tracts, especially in the iHUDs. To our knowledge, this is the first study to examine cytokine blood levels as biomarkers of WM change in iHUDs, thereby expanding approaches that have been used for some other neuropsychiatric disorders (60–62).

# Serum Levels of Specific Cytokines Are Correlated With WM Change With Treatment

We recently reported that, compared with HCs, iHUDs receiving chronic methadone or buprenorphine had robust dysregulation of serum cytokine levels (23). The current study shows that levels of specific cytokines (examined together as a multitarget score) were correlated with WM change following inpatient treatment. Thus, the cytokine PC1 score was negatively correlated with metrics of WM change with treatment in

iHUDs, as quantified by higher FA, MD, and AD (but not RD), even with a stringent whole-brain approach (59,63), in anterior, posterior, and superior corona radiata and genu of the corpus callosum (Figure 1).

The different DTI measures studied here are interrelated (64,65), and it is informative to interpret them together. In this study, results of the correlations with  $\Delta$ FA and  $\Delta$ AD were in the expected direction; however, the direction of correlations with  $\Delta$ MD were not (because higher FA and lower MD are commonly interpreted as representing more WM integrity and lower neuroinflammation) (66). Given the absence of correlations between the cytokine PC1 scores and RD, we interpret the correlations with MD (the average diffusivity of all directions, i.e., AD [ $\lambda$ 1] and RD [ $\lambda$ 2,  $\lambda$ 3]) as being driven by the correlation with AD, consistent with the direction of this correlation with FA. The follow-up analyses of representative cytokines from different canonical families included in the PCA (i.e., the IL-6 family member OSM, the growth factor CSF1, and the chemokine CCL7) largely recapitulated these results (Figures 2-4). Overall, the similar pattern of results for  $\Delta$ FA and  $\Delta$ MD (and  $\Delta$ AD) that we report here is an indicator of the caution needed when interpreting the levels of cytokine or group of cytokines (especially from serum), which are not necessarily indicative of overall neuroinflammation. Because different cytokines function in interdependent signaling networks (20,26) to support homeostasis and adaptation to change (allostasis), the direct impact of chronic drug use and/ or MOUD treatment on these targets needs to be further examined. Other factors that may impact interpretation are

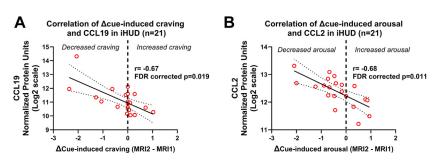


Figure 5. Correlations of changes in cue-induced craving or arousal (the second magnetic resonance imaging scan [MRI2] – the first MRI scan [MRI1]) with levels of specific serum cytokines in individuals with heroin use disorder (iHUDs), measured at the MRI2 stage. Pearson correlations are shown, with linear regression and 95% CI bands. Panel (A) shows change in cue-induced craving and CCL19 levels. Panel (B) shows change in cue-induced arousal and CCL2 levels. FDR, false discovery rate.

technical [e.g., the effect of crossing fibers or other heterogeneities on DTI metrics (67)], although commissural tracts such as the corpus callosum that were prominent in these correlations have predominantly parallel fiber organization, which makes them less vulnerable to such complexities (67).

Changes in WM microstructure in these and other tracts have previously been associated with neurocognitive dysfunction and disease severity in iHUDs (37,68,69), but the mechanisms that underlie WM impairment or recovery are not well understood. Although we cannot address the causality of the association between specific serum cytokines and changes in WM microstructure, some ancillary evidence may guide future studies. First, there is converging evidence that OSM and CSF1 mediate glial functions including myelination (70-73). Also, the transcription of genes encoding OSM and CSF1 (or that of cognate receptors) was elevated in postmortem ventral midbrain of iHUDs compared with HCs (11). Second, the chemokine CCL7 is a chemoattractant for macrophages and can promote neuroinflammation after trauma (74). More broadly, several studies have shown that central and peripheral cytokine/ immune mechanisms can be causally linked (17,75,76). Future studies should examine whether specific cytokines (functioning individually or in networks) are causally involved in changes in WM microstructure during treatment with MOUD and the process of recovery or whether they can function as biomarkers for personalized therapeutic approaches.

# Change in Cue-Induced Ratings (MRI2 - MRI1) Is Related to Cytokine Levels in the iHUDs

In exploratory analyses, we found that change in drug cueinduced subjective ratings in the iHUDs over the course of treatment (i.e., MRI2 - MRI1) was negatively correlated with serum levels of specific cytokines at the time of MRI2, suggesting that levels of these immune targets may be used as biomarkers of neurocognitive/emotional/motivational symptoms in OUD. Notably, we found a negative correlation between change in cue-induced craving and levels of the chemokine CCL19 (Figure 3A), a ligand for chemokine receptor CCR7 (77), which mediates heterologous desensitization of MORs in vitro (78). There was also a negative correlation between change in cue-induced arousal and levels of CCL2 (Figure 3B), the ligand for chemokine receptors CCR2 and CCR4 (79). Relatedly, a CCL2-induced mechanism has recently been implicated in the development of opioid withdrawal in a rodent model (17), and increased CCL2 blood levels have been causally associated with decreased FA in humans (76). Intriguingly, we recently reported that in these participants, serum levels of both CCL19 and CCL2 were greater in iHUDs than in HCs (23). Therefore, while these findings are apparently counterintuitive, two considerations may guide follow-up studies. First, it is unknown whether the timeline of specific cytokine serum levels is linear across the duration of MOUD treatment or recovery. Second, for simplicity, we only examined 4 representative members of canonical cytokine families for their correlations with ΔDTI measures. As mentioned above, cytokines work in complex functional networks (20). Therefore, other targets not examined here may more fully show how patterns of cytokine dysregulation in the iHUDs underlie these findings. Future multimodal longitudinal studies should examine larger serum cytokine

networks together with cue-induced craving/arousal and DTI measures in iHUDs.

#### **Limitations and Methodological Considerations**

This relatively small (low N) observational study should be replicated and expanded in a larger sample of iHUDs and HCs. Furthermore, obtaining blood samples several times, including at treatment onset, would allow future studies to examine whether cytokine/immune changes in iHUDs (15,23,80) could contribute to the long-term changes observed with treatment in WM microstructure and drug cue-induced craving or arousal. Also, as a means of dimensionality reduction, we focused here on the 19 cytokines in this multiplex assay that differed significantly between iHUDs and HCs (23); larger longitudinal studies could examine a broader array of cytokines at different stages in the OUD trajectory. Furthermore, the serum samples were obtained during daytime hours, but further circadian precision should be implemented in future studies (81). Future studies could also examine the influence of broader health features (e.g., diet and hydration) associated with inpatient treatment for substance use disorders, in addition to MOUD treatment or disease trajectory per se.

#### **Conclusions**

This is the first study to examine serum cytokine multiplex levels as biomarkers of longitudinal change in WM microstructure in iHUDs receiving chronic methadone or buprenorphine treatment. We found that cytokine PC1 scores, based on 19 cytokines that differed significantly between iHUDs and HCs (23), were negatively correlated with change in WM microstructure in specific tracts across ≈ 14 weeks of treatment with these MOUDs. This result was also observed when we examined representative cytokines (OSM, CSF1, and CCL7), thus implicating specific molecular targets in this association. Furthermore, exploratory analyses showed that levels of other cytokines (i.e., CCL19 and CCL2) were negatively correlated with change in cue-induced craving or arousal. Future studies should explore how specific cytokine targets (including those identified here) are functionally involved in changes in WM microstructure during recovery in iHUDs, potentially by using pharmacological manipulation (82-86) or neuroimaging studies of immune and inflammatory processes (76,87-89) in humans with OUD (82-86) or in preclinical models (90,91).

#### **ACKNOWLEDGMENTS AND DISCLOSURES**

This work was supported by the National Institute on Drug Abuse (Grant Nos. U01DA053625 [to ERB], R01DA049547 [to NA-K], and R01DA047880 [to PR]), the National Center for Complementary and Integrative Health (Grant No. R01AT010627 [to RZG and ELG]), the National Institute of Mental Health (Grant Nos. R01MH104559 and R01MH127820 [to SJR]), and the National Institute on Aging (Grant No. R01AG067025 [to PR]).

We thank all the clinical coordinators of the Neuroimaging of Addictions and Related Conditions laboratory.

The authors report no biomedical financial interests or potential conflicts of interest.

#### **ARTICLE INFORMATION**

From the Neuropsychoimaging of Addictions and Related Conditions Research Program, Icahn School of Medicine at Mount Sinai, Departments

of Psychiatry and Neuroscience, New York, New York (ERB, YH, SGK, P-OG, AOC, GK, RZG, NA-K); Department of Psychiatry, Icahn School of Medicine at Mount Sinai, New York, New York (ERB, YH, SGK, P-OG, AOC, GK, PR, RZG, NA-K); Friedman Brain Institute, Icahn School of Medicine at Mount Sinai, New York, New York (FC, SJR); Nash Family Department of Neuroscience, Icahn School of Medicine at Mount Sinai, New York, New York (FC, SJR); Center for Disease Neurogenomics, Icahn School of Medicine at Mount Sinai, New York, New York (PR); Department of Genetics and Genomic Science, Icahn School of Medicine at Mount Sinai, New York, New York (PR); Mental Illness Research, Education, and Clinical Center (VISN 2 South), James J. Peters Veterans Affairs, Medical Center, Bronx, New York (PR); Center for Precision Medicine and Translational Therapeutics, James J. Peters Veterans Affairs, Medical Center, Bronx, New York (PR); Brain and Body Research Center, Icahn School of Medicine at Mount Sinai, New York, New York (SJR); Center of Affective Neuroscience, Icahn School of Medicine at Mount Sinai, New York, New York (SJR, RZG, NA-K); Department of Psychiatry, University of California San Diego, La Jolla, California (ELG); and Sanford Institute for Empathy and Compassion, University of California San Diego, La Jolla, California (ELG).

ERB, YH, and SGK contributed equally to this work.

Address correspondence to Nelly Alia-Klein, Ph.D., at nelly.alia-klein@mssm.edu, or Eduardo R. Butelman, Ph.D., at eduardo.butelman@mssm.edu.

Received Dec 13, 2024; revised Jan 24, 2025; accepted Feb 20, 2025. Supplementary material cited in this article is available online at https://doi.org/10.1016/j.bpsgos.2025.100480.

#### **REFERENCES**

- Butelman ER, Huang Y, Epstein DH, Shaham Y, Goldstein RZ, Volkow ND, Alia-Klein N (2023): Overdose mortality rates for opioids and stimulant drugs are substantially higher in men than in women: State-level analysis. Neuropsychopharmacology 48:1639–1647.
- Strang J, Volkow ND, Degenhardt L, Hickman M, Johnson K, Koob GF, et al. (2020): Opioid use disorder. Nat Rev Dis Primers 6:3.
- Kreek MJ, Reed B, Butelman ER (2019): Current status of opioid addiction treatment and related preclinical research. Sci Adv 5:eaax9140.
- Luo SX, Feaster DJ, Liu Y, Balise RR, Hu M-C, Bouzoubaa L, et al. (2024): Individual-level risk prediction of return to use during opioid use disorder treatment. JAMA Psychiatry 81:45–56.
- Perlis RH (2011): Translating biomarkers to clinical practice. Mol Psychiatry 16:1076–1087.
- Volkow ND, Koob G, Baler R (2015): Biomarkers in substance use disorders. ACS Chem Neurosci 6:522–525.
- Catale C, Bussone S, Lo Iacono L, Carola V (2019): Microglial alterations induced by psychoactive drugs: A possible mechanism in substance use disorder? Semin Cell Dev Biol 94:164–175.
- Zhang Y, Liang Y, Levran O, Randesi M, Yuferov V, Zhao C, Kreek MJ (2017): Alterations of expression of inflammation/immune-related genes in the dorsal and ventral striatum of adult C57BL/6J mice following chronic oxycodone self-administration: A RNA sequencing study. Psychopharmacology (Berl) 234:2259–2275.
- Butelman ER, Goldstein RZ, Nwaneshiudu CA, Girdhar K, Roussos P, Russo SJ, Alia-Klein N (2023): Neuroimmune mechanisms of opioid use disorder and recovery: Translatability to human studies, and future research directions. Neuroscience 528:102–116.
- Eisenstein TK (2019): The role of opioid receptors in immune system function. Front Immunol 10:2904.
- Wei J, Lambert TY, Valada A, Patel N, Walker K, Lenders J, et al. (2023): Single nucleus transcriptomics of ventral midbrain identifies glial activation associated with chronic opioid use disorder. Nat Commun 14:5610.
- Seney ML, Kim S-M, Glausier JR, Hildebrand MA, Xue X, Zong W, et al. (2021): Transcriptional alterations in dorsolateral prefrontal cortex and nucleus accumbens implicate neuroinflammation and synaptic remodeling in opioid use disorder. Biol Psychiatry 90:550–562.
- Sacerdote P, Franchi S, Gerra G, Leccese V, Panerai AE, Somaini L (2008): Buprenorphine and methadone maintenance treatment of heroin addicts preserves immune function. Brain Behav Immun 22:606–613.

- Franchi S, Moschetti G, Amodeo G, Sacerdote P (2019): Do all opioid drugs share the same immunomodulatory properties? A review from animal and human studies. Front Immunol 10:2914.
- Re GF, Jia J, Xu Y, Zhang Z, Xie ZR, Kong D, et al. (2022): Dynamics and correlations in multiplex immune profiling reveal persistent immune inflammation in male drug users after withdrawal. Int Immunopharmacol 107:108696.
- Kruyer A, Angelis A, Garcia-Keller C, Li H, Kalivas PW (2022): Plasticity in astrocyte subpopulations regulates heroin relapse. Sci Adv 8: eabo7044
- Zhu Y, Yan P, Wang R, Lai J, Tang H, Xiao X, et al. (2023): Opioidinduced fragile-like regulatory T cells contribute to withdrawal. Cell 186:591–606.e23.
- Boutou A, Roufagalas I, Politopoulou K, Tastsoglou S, Abouzeid M, Skoufos G, et al. (2024): Microglia regulate cortical remyelination via TNFR1-dependent phenotypic polarization. Cell Rep 43:114894.
- Andreadou M, Ingelfinger F, De Feo D, Cramer TLM, Tuzlak S, Friebel E, et al. (2023): IL-12 sensing in neurons induces neuroprotective CNS tissue adaptation and attenuates neuroinflammation in mice. Nat Neurosci 26:1701–1712.
- Becher B, Spath S, Goverman J (2017): Cytokine networks in neuroinflammation. Nat Rev Immunol 17:49–59.
- Cathomas F, Lin H-Y, Chan KL, Li L, Parise LF, Alvarez J, et al. (2024): Circulating myeloid-derived MMP8 in stress susceptibility and depression. Nature 626:1108–1115.
- Ménard C, Pfau ML, Hodes GE, Russo SJ (2017): Immune and neuroendocrine mechanisms of stress vulnerability and resilience. Neuropsychopharmacology 42:62–80.
- Butelman ER, Huang Y, Cathomas F, Gaudreault P-O, Roussos P, Russo SJ, et al. (2024): Serum cytokines and inflammatory proteins in individuals with heroin use disorder: Potential mechanistically based biomarkers for diagnosis. Transl Psychiatry 14:414.
- Marchette RCN, Carlson ER, Said N, Koob GF, Vendruscolo LF (2023): Extended access to fentanyl vapor self-administration leads to addiction-like behaviors in mice: Blood chemokine/cytokine levels as potential biomarkers. Addict Neurosci 5:100057.
- Sharma HS, Sjöquist P-O, Ali SF (2010): Alterations in blood-brain barrier function and brain pathology by morphine in the rat. Neuroprotective effects of antioxidant H-290/51. Acta Neurochir Suppl 106:61–66.
- Goldsmith DR, Bekhbat M, Mehta ND, Felger JC (2023): Inflammationrelated functional and structural dysconnectivity as a pathway to psychopathology. Biol Psychiatry 93:405–418.
- Khelfaoui H, Ibaceta-Gonzalez C, Angulo MC (2024): Functional myelin in cognition and neurodevelopmental disorders. Cell Mol Life Sci 81:181.
- Baaklini CS, Ho MFS, Lange T, Hammond BP, Panda SP, Zirngibl M, et al. (2023): Microglia promote remyelination independent of their role in clearing myelin debris. Cell Rep 42:113574.
- Bacmeister CM, Huang R, Osso LA, Thornton MA, Conant L, Chavez AR, et al. (2022): Motor learning drives dynamic patterns of intermittent myelination on learning-activated axons. Nat Neurosci 25:1300–1313.
- Anderson WD, Vadigepalli R (2022): Neuroinflammation, glia, and cytokines: Networks of networks. In: Jaeger D, Jung R, editors. Encyclopedia of Computational Neuroscience. New York, NY: Springer, 2281–2287.
- 31. Kennedy RH, Silver R (2015): Neuroimmune signaling: Cytokines and the CNS. In: Pfaff DW, Volkow ND, Rubenstein J, editors. Neuroscience in the 21st Century. Berlin, Germany: Springer, 1–41.
- Yalçın B, Pomrenze MB, Malacon K, Drexler R, Rogers AE, Shamardani K, et al. (2024): Myelin plasticity in the ventral tegmental area is required for opioid reward. Nature 630:677–685.
- Anderson SAR, Michaelides M, Zarnegar P, Ren Y, Fagergren P, Thanos PK, et al. (2013): Impaired periamygdaloid-cortex prodynorphin is characteristic of opiate addiction and depression. J Clin Invest 123:5334–5341.
- 34. Okvist A, Fagergren P, Whittard J, Garcia-Osta A, Drakenberg K, Horvath MC, et al. (2011): Dysregulated postsynaptic density and

- endocytic zone in the amygdala of human heroin and cocaine abusers. Biol Psychiatry 69:245–252.
- Basser PJ, Mattiello J, LeBihan D (1994): MR diffusion tensor spectroscopy and imaging. Biophys J 66:259–267.
- Le Bihan D, Breton E, Lallemand D, Grenier P, Cabanis E, Laval-Jeantet M (1986): MR imaging of intravoxel incoherent motions: Application to diffusion and perfusion in neurologic disorders. Radiology 161:401–407.
- Gaudreault PO, King SG, Malaker P, Alia-Klein N, Goldstein RZ (2023): Whole-brain white matter abnormalities in human cocaine and heroin use disorders: Association with craving, recency, and cumulative use. Mol Psychiatry 28:780–791.
- King SG, Gaudreault PO, Malaker P, Kim JW, Alia-Klein N, Xu J, Goldstein RZ (2022): Prefrontal-habenular microstructural impairments in human cocaine and heroin addiction. Neuron 110:3820–3832.e4.
- Yeh P-H, Simpson K, Durazzo TC, Gazdzinski S, Meyerhoff DJ (2009): Tract-Based Spatial Statistics (TBSS) of diffusion tensor imaging data in alcohol dependence: Abnormalities of the motivational neurocircuitry. Psychiatry Res 173:22–30.
- Gaudreault P-O, King SG, Huang Y, Ceceli AO, Kronberg G, Alia-Klein N, Goldstein RZ (2024): Frontal white matter changes and craving recovery in inpatients with heroin use disorder. JAMA Netw Open 7:e2451678.
- Lu L, Yang W, Zhang X, Tang F, Du Y, Fan L, et al. (2022): Potential brain recovery of frontostriatal circuits in heroin users after prolonged abstinence: A preliminary study. J Psychiatr Res 152:326–334.
- Yang J, Ran M, Li H, Lin Y, Ma K, Yang Y, et al. (2022): New insight into neurological degeneration: Inflammatory cytokines and blood-brain barrier. Front Mol Neurosci 15:1013933.
- Banks WA, Lynch JL, Price TO (2008): Cytokines and the blood-brain barrier. In: The Neuroimmunological Basis of Behavior and Mental Disorders. Boston, MA: Springer, 3–17.
- Konsman JP (2022): Cytokines in the brain and neuroinflammation: We didn't starve the fire! Pharmaceuticals (Basel) 15:140.
- 45. Sheehan DV, Lecrubier Y, Sheehan KH, Amorim P, Janavs J, Weiller E, et al. (1998): The Mini-International Neuropsychiatric Interview (M.I.N.I.): The development and validation of a structured diagnostic psychiatric interview for DSM-IV and ICD-10. J Clin Psychiatry 59(suppl 20):22–33.
- McLellan AT, Kushner H, Metzger D, Peters R, Smith I, Grissom G, et al. (1992): The Fifth Edition of the Addiction Severity Index. J Subst Abuse Treat 9:199–213.
- Costi S, Morris LS, Collins A, Fernandez NF, Patel M, Xie H, et al. (2021): Peripheral immune cell reactivity and neural response to reward in patients with depression and anhedonia. Transl Psychiatry 11:565.
- Liang J, Xu Y, Gao W, Sun Y, Zhang Y, Shan F, Xia Q (2024): Cytokine profile in first-episode drug-naïve major depressive disorder patients with or without anxiety. BMC Psychiatry 24:93.
- Beck AT, Steer RA, Ball R, Ranieri W (1996): Comparison of beck depression inventories -IA and -II in psychiatric outpatients. J Pers Assess 67:588–597.
- Butelman ER, Bacciardi S, Maremmani AGI, Darst-Campbell M, Correa da Rosa J, Kreek MJ (2017): Can a rapid measure of self-exposure to drugs of abuse provide dimensional information on depression comorbidity? Am J Addict 26:632–639.
- Beck AT, Epstein N, Brown G, Steer RA (1988): An inventory for measuring clinical anxiety: Psychometric properties. J Consult Clin Psychol 56:893–897.
- Cohen S, Kamarck T, Mermelstein R (1983): A global measure of perceived stress. J Health Soc Behav 24:385–396.
- Huang Y, Ceceli AO, Kronberg G, King S, Malaker P, Parvaz MA, et al. (2024): Association of cortico-striatal engagement during cue reactivity, reappraisal, and savoring of drug and non-drug stimuli with craving in heroin addiction. Am J Psychiatry 181:153–165.
- Struglics A, Larsson S, Lohmander LS, Swärd P (2022): Technical performance of a proximity extension assay inflammation biomarker panel with synovial fluid. Osteoarthr Cartil Open 4:100293.
- 55. Josse J, Husson F (2016): missMDA: A package for handling missing values in multivariate data analysis. J Stat Soft 70:1–31.

- Benjamini Y, Hochberg Y (1995): Controlling the false discovery rate: A practical and powerful approach to multiple testing. J R Stat Soc B 57:289–300.
- Jolliffe IT, Cadima J (2016): Principal component analysis: A review and recent developments. Philos Trans A Math Phys Eng Sci 374: 20150202
- Tournier J-D, Smith R, Raffelt D, Tabbara R, Dhollander T, Pietsch M, et al. (2019): MRtrix3: A fast, flexible and open software framework for medical image processing and visualisation. Neuroimage 202:116137.
- Smith SM, Nichols TE (2009): Threshold-free cluster enhancement: Addressing problems of smoothing, threshold dependence and localisation in cluster inference. Neuroimage 44:83–98.
- Sammer G, Neumann E, Blecker C, Pedraz-Petrozzi B (2022): Fractional anisotropy and peripheral cytokine concentrations in outpatients with depressive episode: A diffusion tensor imaging observational study. Sci Rep 12:17450.
- Kitzbichler MG, Aruldass AR, Barker GJ, Wood TC, Dowell NG, Hurley SA, et al. (2021): Peripheral inflammation is associated with micro-structural and functional connectivity changes in depressionrelated brain networks. Mol Psychiatry 26:7346–7354.
- Breit S, Mazza E, Poletti S, Benedetti F (2023): White matter integrity and pro-inflammatory cytokines as predictors of antidepressant response in MDD. J Psychiatr Res 159:22–32.
- Smith SM, Johansen-Berg H, Jenkinson M, Rueckert D, Nichols TE, Miller KL, et al. (2007): Acquisition and voxelwise analysis of multi-subject diffusion data with tract-based spatial statistics. Nat Protoc 2:499–503.
- Curran KM, Emsell L, Leemans A (2016): Quantitative DTI measures.
  In: Van Hecke W, Emsell L, Sunaert S, editors. Diffusion Tensor Imaging. New York, NY: Springer, 65–87.
- Alexander AL, Lee JE, Lazar M, Field AS (2007): Diffusion tensor imaging of the brain. Neurotherapeutics 4:316–329.
- Sbardella E, Tona F, Petsas N, Pantano P (2013): DTI measurements in multiple sclerosis: Evaluation of brain damage and clinical implications. Mult Scler Int 2013:671730.
- Figley CR, Uddin MN, Wong K, Kornelsen J, Puig J, Figley TD (2021): Potential pitfalls of using fractional anisotropy, axial diffusivity, and radial diffusivity as biomarkers of cerebral white matter microstructure. Front Neurosci 15:799576.
- Bora E, Yücel M, Fornito A, Pantelis C, Harrison BJ, Cocchi L, et al. (2012): White matter microstructure in opiate addiction. Addict Biol 17:141–148.
- Liu H, Li L, Hao Y, Cao D, Xu L, Rohrbaugh R, et al. (2008): Disrupted white matter integrity in heroin dependence: A controlled study utilizing diffusion tensor imaging. Am J Drug Alcohol Abuse 34:562–575.
- Houben E, Hellings N, Broux B (2019): Oncostatin M, an underestimated player in the central nervous system. Front Immunol 10:1165.
- Houben E, Janssens K, Hermans D, Vandooren J, Van den Haute C, Schepers M, et al. (2020): Oncostatin M-induced astrocytic tissue inhibitor of metalloproteinases-1 drives remyelination. Proc Natl Acad Sci U S A 117:5028–5038.
- Glezer I, Rivest S (2010): Oncostatin M is a novel glucocorticoiddependent neuroinflammatory factor that enhances oligodendrocyte precursor cell activity in demyelinated sites. Brain Behav Immun 24:695–704.
- Marzan DE, Brügger-Verdon V, West BL, Liddelow S, Samanta J, Salzer JL (2021): Activated microglia drive demyelination via CSF1R signaling. Glia 69:1583–1604.
- Xue J, Zhang Y, Zhang J, Zhu Z, Lv Q, Su J (2021): Astrocyte-derived CCL7 promotes microglia-mediated inflammation following traumatic brain injury. Int Immunopharmacol 99:107975.
- Kiss MG, Mindur JE, Yates AG, Lee D, Fullard JF, Anzai A, et al. (2023): Interleukin-3 coordinates glial-peripheral immune crosstalk to incite multiple sclerosis. Immunity 56:1502–1514.e8.
- Yang Y, Yao Z, Huo L (2024): Causal effect of circulating cytokines on MRI markers of cerebral small vessel disease: A Mendelian randomization study. Cytokine 182:156713.
- Yoshida R, Imai T, Hieshima K, Kusuda J, Baba M, Kitaura M, et al. (1997): Molecular cloning of a novel human CC chemokine EBI1-ligand chemokine that is a specific functional ligand for EBI1, CCR7. J Biol Chem 272:13803–13809.

- Szabo I, Chen XH, Xin L, Adler MW, Howard OM, Oppenheim JJ, Rogers TJ (2002): Heterologous desensitization of opioid receptors by chemokines inhibits chemotaxis and enhances the perception of pain. Proc Natl Acad Sci U S A 99:10276–10281.
- Deshmane SL, Kremlev S, Amini S, Sawaya BE (2009): Monocyte chemoattractant protein-1 (MCP-1): An overview. J Interferon Cytokine Res 29:313–326.
- Chan YY, Yang SN, Lin JC, Chang JL, Lin JG, Lo WY (2015): Inflammatory response in heroin addicts undergoing methadone maintenance treatment. Psychiatry Res 226:230–234.
- 81. Zielinski MR, Gibbons AJ (2022): Neuroinflammation, sleep, and circadian rhythms. Front Cell Infect Microbiol 12:853096.
- 82. Mogali S, Askalsky P, Madera G, Jones JD, Comer SD (2021): Minocycline attenuates oxycodone-induced positive subjective responses in non-dependent, recreational opioid users. Pharmacol Biochem Behav 209:173241.
- Metz VE, Jones JD, Manubay J, Sullivan MA, Mogali S, Segoshi A, et al. (2017): Effects of ibudilast on the subjective, reinforcing, and analgesic effects of oxycodone in recently detoxified adults with opioid dependence. Neuropsychopharmacology 42:1825– 1832.
- Arout CA, Waters AJ, MacLean RR, Compton P, Sofuoglu M (2019): Minocycline does not affect experimental pain or addiction-related outcomes in opioid maintained patients. Psychopharmacology (Berl) 236:2857–2866.

- Cooper ZD, Johnson KW, Vosburg SK, Sullivan MA, Manubay J, Martinez D, et al. (2017): Effects of ibudilast on oxycodone-induced analgesia and subjective effects in opioid-dependent volunteers. Drug Alcohol Depend 178:340–347.
- Cooper ZD, Johnson KW, Pavlicova M, Glass A, Vosburg SK, Sullivan MA, et al. (2016): The effects of ibudilast, a glial activation inhibitor, on opioid withdrawal symptoms in opioid-dependent volunteers. Addict Biol 21:895–903.
- Saxton RA, Glassman CR, Garcia KC (2023): Emerging principles of cytokine pharmacology and therapeutics. Nat Rev Drug Discov 22:21–37.
- 88. Sugimoto K, Kakeda S, Watanabe K, Katsuki A, Ueda I, Igata N, et al. (2018): Relationship between white matter integrity and serum inflammatory cytokine levels in drug-naive patients with major depressive disorder: Diffusion tensor imaging study using tract-based spatial statistics. Transl Psychiatry 8:141.
- Sakamoto S, Zhu X, Hasegawa Y, Karma S, Obayashi M, Alway E, Kamiya A (2021): Inflamed brain: Targeting immune changes and inflammation for treatment of depression. Psychiatry Clin Neurosci 75:304–311.
- Shin SH, Kim E-K, Lee K-Y, Kim H-S (2019): TNF-α antagonist attenuates systemic lipopolysaccharide-induced brain white matter injury in neonatal rats. BMC Neurosci 20:45.
- Gardner C, Magliozzi R, Durrenberger PF, Howell OW, Rundle J, Reynolds R (2013): Cortical grey matter demyelination can be induced by elevated pro-inflammatory cytokines in the subarachnoid space of MOG-immunized rats. Brain 136:3596–3608.