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Prognostic and predictive value of extended *RAS* mutation and mismatch repair status in stage III colorectal cancer

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Adjuvant chemotherapy, colorectal cancer, mismatch repair, *RAS*, tegafur–uracil

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The prognostic and predictive value of KRAS gene mutations in stage III colorectal cancer is controversial because many recent clinical trials have not involved a surgery-alone arm. Additionally, data on the significance of extended RAS (KRAS/ NRAS) mutations in stage III cancer are not available. Hence, we undertook a combined analysis of two phase III randomized trials, in which the usefulness of adjuvant chemotherapy with tegafur-uracil (UFT) was evaluated, as compared with surgery alone. We determined the association of extended RAS and mismatch repair (MMR) status with the effectiveness of adjuvant chemotherapy. Mutations in KRAS exons 2, 3, and 4 and NRAS exons 2 and 3 were detected by direct DNA sequencing. Tumor MMR status was determined by immunohistochemistry. Total RAS mutations were detected in 134/304 (44%) patients. In patients with RAS mutations, a significant benefit was associated with adjuvant UFT in relapse-free survival (RFS) (hazard ratio = 0.49; P = 0.02) and overall survival (hazard ratio = 0.51; P = 0.03). In contrast, among patients without RAS mutations, there was no difference in RFS or overall survival between the adjuvant UFT group and surgery-alone group. We detected deficient DNA MMR in 23/ 304 (8%) patients. The MMR status was neither prognostic nor predictive for adjuvant chemotherapy. An interaction analysis showed that there was better RFS among patients treated with UFT with RAS mutations, but not for those without RAS mutations. Extended RAS (KRAS/NRAS) mutations are proposed as predictive indicators with respect to the efficacy of adjuvant UFT chemotherapy in patients with resected stage III colorectal cancer.

C olorectal cancer is the third leading cause of death from cancer in Japan. Surgical resection offers a potential cure for patients with colon cancer; however, some patients with completely resected cancers continue to die of metastatic relapse. In patients with stage III colon cancer, postoperative adjuvant chemotherapy is the international standard of care for improved survival. The guidelines at North America and Europe recommend 5-FU and folic acid (LV) or capecitabine combined with oxaliplatin as the first choice for adjuvant chemotherapy in stage III colon cancer.⁽¹⁻³⁾ In addition, radiotherapy combined with chemotherapy is recommended as a standard adjuvant therapy for rectal cancer. However, in Japan, clinical trials of postoperative adjuvant chemotherapy have focused mainly on oral fluoropyrimidine-based regimens in both colon and rectal cancers.⁽⁴⁻⁶⁾ Tegafur–uracil is a combination drug comprising tegafur, a prodrug of 5-FU, and uracil, an inhibitor of the 5-FU-degrading enzyme dihydropyrimidine dehydrogenase, in a molar ratio of 1:4. The non-inferiority of UFT/LV to 5-FU/LV as adjuvant chemotherapy for stage III colorectal cancer has been verified in several clinical trials.^(5,7) Therefore, at present, the UFT/LV regimen (five courses of 6 months of treatment consisting of UFT 300 mg/m²/day for 28 days plus a 7-day washout) has been widely adopted in Japan as standard adjuvant chemotherapy.

In metastatic colorectal cancer, *KRAS* mutation in exon 2 is a well-established biomarker that predicts a lack of benefit from treatment with anti-epidermal growth factor receptor mAbs.^(8–10) Moreover, patients with metastatic colorectal cancer harboring activating mutations in other *RAS* (*KRAS* exons 3 and 4 and *NRAS* exons 2, 3, and 4) genes were not shown to benefit from anti-epidermal growth factor receptor therapy in past clinical trials.^(11–13) However, the prognostic and predictive value of the *KRAS* mutation in exon 2 in stage III cancer is controversial because many recent clinical trials have not

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involved a surgery alone arm.⁽¹⁴⁻¹⁶⁾ Additionally, data on the significance of extended *RAS* (*KRAS* and *NRAS*) mutations in stage III cancer were not reported in these studies.

One of the genetic pathways involved in colorectal cancer is dMMR. Deficient DNA MMR attenuates protein expression (MSH2, MSH6, PMS2, and MLH1) leading to high-frequency microsatellite instability. Some reports suggest that dMMR may predict the response to adjuvant fluoropyrimidine-based chemotherapy.^(17–19) In contrast, the other study has shown that MMR status had no predictive value in respect to benefit from adjuvant chemotherapy in stage III colorectal cancer.⁽²⁰⁾

In this report, we undertook a combined analysis of two phase III randomized trials, NSAS-CC and NSAS-RC, in which adjuvant UFT chemotherapy was evaluated in the stage III colorectal cancer setting.^(4,21) These trials showed that postoperative adjuvant UFT therapy significantly improved RFS and OS in patients with stage III rectal cancer (RFS, HR = 0.66, P = 0.03; OS, HR = 0.60, P = 0.03), but not colon cancer (RFS, HR = 0.89, P = 0.56; OS, HR = 0.82, P = 0.39). In the current study, we determined the association of extended *RAS* and MMR status with the effectiveness of surgery plus adjuvant UFT chemotherapy, as compared with surgery alone.

Patients and Methods

Study design and treatment. The NSAS-CC and NSAS-RC studies were carried out as multicenter phase III randomized trials to examine the usefulness of postoperative adjuvant chemotherapy with UFT alone in patients with curatively resected stage III colon or rectal cancer, respectively. The study design and eligibility criteria have been reported previously.^(4,21) Enrolment of patients in the original trials occurred between October 1996 and April 2001. Patients were randomly assigned to receive either adjuvant chemotherapy with UFT, or no chemotherapy treatment within 6 weeks after surgery. In the UFT group, UFT (tegafur 400 mg/m²/day; Taiho Pharmaceutical Co., Tokyo, Japan) was given orally, twice daily for 5 days/week for 1 year. When this study was carried out, LV tablets could not be used because they had not been approved in Japan; therefore, UFT alone was used. The stage was classified according to the General Rules for Clinical and Pathological Studies on Cancer of the Colon, Rectum, and Anus.⁶ Cancers arising from the rectosigmoid colon were classified as rectal cancer. A diagnosis of recurrence was assessed by abdominal ultrasonography or computed tomography at 4-month intervals during the first 2 years and at 6-month intervals thereafter.

This study was carried out in accordance with the Declaration of Helsinki and was approved by the Institutional Review Board of the National Cancer Center Hospital (Tokyo, Japan).

KRAS/NRAS, BRAF, and *PIK3CA* mutation analysis. DNA was extracted from formalin-fixed, paraffin-embedded tumor samples. The mutation status of *KRAS* exon 2 (at codons 12 and 13), exon 3 (at codon 61), and exon 4 (at codon 146), *NRAS* exon 2 (at codons 12 and 13) and exon 3 (at codon 61), and *PIK3CA* exon 9 (at codons 542 and 545) and exon 20 (at codon 1047) was assessed by means of direct sequencing by the PCR method. Detection of *BRAF* V600E mutation was achieved using high-resolution melting analysis, which has been described in detail elsewhere.⁽²³⁾

Mismatch repair status determination. Tumor MMR status was determined by immunohistochemistry. A dMMR status was defined as the loss of tumor MSH2, MSH6, PMS2, or MLH1 protein expression. A proficient MMR status was defined by normal tumor MSH2, MSH6, PMS2, and MLH1 protein expression. Sections (5 µm thick) from paraffinembedded tissues were stained using an Autostainer (Dako, Glostrup, Denmark). Staining was carried out using antibodies to MSH2 (clone FE11, 1/200; Calbiochem, Tokyo, Japan), MSH6 (clone EPR3945, 1/200; GeneTex, Hsinchu City, Taiwan), PMS2 (clone A16-4, 1/200; BD Biosciences Pharmingen, Tokyo, Japan) and MLH1 (clone G168-278, 1/200; BD Biosciences Pharmingen). Mismatch repair protein loss was defined as abnormal (or absent) when nuclear staining of tumor cells was absent in the presence of positive staining in surrounding cells.

Statistical methods. Our primary objective for this analysis was to evaluate the effect of adjuvant chemotherapy with UFT, as compared with surgery alone, in relation to the presence (i.e., any KRAS exon 2, 3, or 4, or NRAS exon 2 or 3 mutations) or absence (i.e., neither *KRAS* nor *NRAS* mutation) of RAS mutation in tumor tissues from patients participating in NSAS-CC/NSAS-RC trials. Relapse-free survival was defined as the time from the date of surgery to recurrence or death from any cause. Overall survival was defined as the interval from the date of surgery to death from any cause or last follow-up. Hazard ratios were estimated using Cox proportional hazards models. A multivariate analysis adjusted for tumor stage, primary tumor location, and treatment was carried out. An interaction analysis was used to compare the treatment effect of UFT between subgroups with non-mutated RAS and RAS mutations, which was adjusted for tumor stage, primary tumor location, MMR status, and treatment.

Between-group differences in patient characteristics were assessed with the use of the Wilcoxon rank sum test for continuous variables and Fisher's exact test of association for categorical variables. Survival curves for RFS and OS were estimated using the Kaplan–Meier method, and differences were evaluated with the log–rank test. All *P*-values are twosided. *P*-values of <0.05 were considered to indicate statistical significance, and 95% CI were calculated. All statistical analyses were carried out with the use of SAS software (version 9.4; SAS Institute, Cary, NC, USA).

Results

Patient characteristics. Of the 610 patients (334 with colon cancer and 276 with rectal cancer) who underwent randomization in the original NSAS-CC/RC trials, we collected 324 tumor samples (178 colon and 146 rectum; Fig. 1). During a median follow-up of 74.8 months (range, 8.8-176.3 months), there were 115 RFS events and 90 deaths. The status of RAS and MMR was determined in 304 (94%) out of the 324 patients. Of the 304 patients, 134 (44%) were identified as having tumors with mutated RAS (any mutations in exons 2, 3, or 4 of KRAS, or in exons 2 or 3 of NRAS), and 170 (56%) had non-mutated RAS (no KRAS and NRAS mutations). KRAS exon 2 mutations were detected in 39% of tumors, with 29% detected in codon 12 and 10% in codon 13. The following additional mutation rates were also observed: 0% for KRAS exon 3; 2% for KRAS exon 4; 5% for NRAS exon 2; and 2% for NRAS exon 3 (Table S1). Mutations of both KRAS and NRAS occurred in eight patients; specifically, mutations in KRAS exon 2 and NRAS exon 2 (n = 7), along with KRAS exon 2 and NRAS exon 3 (n = 1). Overall, RAS mutations were detected in 82/152 (54%) of the surgery alone group and in 88/152 (58%) of the adjuvant UFT group. Seven cases (4%) displayed a BRAF exon 15 (V600E) mutation in those patients





without RAS mutations who could be evaluated for BRAF status. Mutations in BRAF exon 15 were mutually exclusive of KRAS and NRAS mutations. PIK3CA mutations were detected in 14% of tumors, with 10% detected in exon 9 and 4% in exon 20. Table 1 summarizes the baseline clinicopathologic characteristics of patients according to RAS mutation status. There was no difference according to gender, age, primary tumor location, histopathologic grade, T stage, N stage, tumor stage, treatment (surgery alone or adjuvant UFT), or MMR status between the non-mutated and RAS mutated groups. Deficient DNA MMR was detected in 14/170 (8%) of the nonmutated group and 9/134 (7%) of the RAS mutated group. Among dMMR tumors, those showing loss of MSH2, MSH6, PMS2, and MLH1 protein expression were 9, 4, 7, and 11, respectively (multiple protein loss was observed in some tumors).

Prognostic value of *RAS* **and MMR status on efficacy of surgery alone.** Among patients treated with surgery alone, there was no significant difference in RFS or OS between patients without and with *RAS* mutations (5-year RFS, 64.6% and 61.4%, respectively; HR = 1.23; 95% CI, 0.73–2.06; P = 0.43; 5-year OS, 77.3% and 70.0%, respectively; HR = 1.46; 95% CI, 0.82–2.59; P = 0.19). Mismatch repair status also had no impact on RFS (HR = 0.75; 95% CI, 0.27–2.07; P = 0.58) or OS (HR = 0.93; 95% CI, 0.33–2.58; P = 0.89). Multivariate analysis using a Cox proportional hazards model revealed that both *RAS* and MMR statuses were not prognostic factors (Table S2).

Predictive value of *RAS* and MMR status on efficacy of adjuvant UFT. As shown in Figure 2, when comparing patients with extended *RAS* mutation who received adjuvant UFT to those who received surgery alone, there were significant improvements in RFS (5-year RFS, 76.5% and 61.4%, respectively; HR = 0.49; 95% CI, 0.27–0.91; P = 0.02; Fig. 2a) and OS (5-year OS, 84.4% and 70.0%, respectively; HR = 0.51; 95% CI, 0.26–0.97; P = 0.03; Fig. 2c). In contrast, among patients without *RAS* mutation, there was no significant difference in RFS (5-year RFS, 63.5% and 64.6%, respectively; HR = 0.89; 95% CI, 0.55–1.46; P = 0.67; Fig. 2b) or OS (5-year OS, 82.9% and 77.3%, respectively; HR = 0.94; 95% CI, 0.51–1.71; P = 0.83; Fig. 2d) between the adjuvant UFT chemotherapy plus surgery and surgery alone groups. There was a trend

Table 1. Clinicopathologic characteristics in patients with stage III colorectal cancer according to *RAS* mutation (n = 304)

Variable, n (%)	No <i>RAS</i> mutation	<i>RAS</i> mutation	P-
	(<i>n</i> = 170)	(<i>n</i> = 134)	value
Gender			
Male	99 (58)	69 (51)	0.24
Female	71 (42)	65 (49)	
Age, year			
Median (range)	61 (36–74)	59 (32–75)	0.28
Primary tumor location			
Colon	92 (54)	72 (54)	0.94
Rectum	78 (46)	62 (46)	
Histopathologic grade			
Well differentiated	37 (22)	36 (27)	0.22
Moderately	120 (71)	90 (67)	
differentiated			
Poorly differentiated	10 (6)	3 (2)	
Mucinous	3 (1)	5 (4)	
T stage			
T1–3	113 (66)	86 (64)	0.67
T4	57 (34)	48 (36)	
N stage			
N1	135 (79)	112 (84)	0.52
N2	26 (15)	18 (13)	
N3	9 (6)	4 (3)	
Tumor stage			
Illa	135 (79)	111 (83)	0.45
IIIb	35 (21)	23 (17)	
Treatment			
Surgery alone	82 (48)	70 (52)	0.48
Adjuvant UFT	88 (52)	64 (48)	
MMR			
pMMR	156 (92)	125 (93)	0.63
dMMR	14 (8)	9 (7)	

dMMR, deficient DNA mismatch repair; MMR, mismatch repair; pMMR, proficient mismatch repair; UFT, tegafur–uracil.

toward improved survival among patients with only *KRAS* exon 2 mutations treated with UFT as compared to those managed by surgery alone, but the difference was not statistically



Fig. 2. Kaplan–Meier estimates of relapse-free survival (RFS) and overall survival (OS) in colorectal cancer patients, according to treatment group. (a,b) RFS in patients with extended *RAS* mutation (a) and with no *RAS* mutation (b). (c,d) OS in patients with extended *RAS* mutation (c), and with no *RAS* mutation (d). Adj, adjuvant; CI, confidence interval; HR, hazard ratio; UFT, tegafur–uracil.

significant (RFS, HR = 0.57; 95% CI, 0.30–1.07; P = 0.07; OS, HR = 0.61; 95% CI, 0.31–1.19; P = 0.14). There was no statistical significance in RFS or OS between the adjuvant UFT and surgery-alone group for patients with dMMR tumors (RFS, HR = 1.05; 95% CI, 0.26–4.20; P = 0.95; OS, HR = 0.90; 95% CI, 0.20–4.05; P = 0.89) or proficient MMR tumors (RFS, HR = 0.66; 95% CI, 0.44–1.02; P = 0.06; OS, HR = 0.70; 95% CI, 0.45–1.01; P = 0.11). In addition, among patients with *BRAF* or *PIK3CA* mutations, there was no statistically significant difference in OS between the adjuvant UFT group and the surgery-alone group. The benefit of adjuvant UFT in relation to OS is shown in Figure 3. The use of adjuvant UFT was associated with significantly better survival for patients in subgroups with male gender, rectal cancer, stage IIIb disease, and any evidence of *RAS* mutation.

Predictive value of *RAS* status according to primary tumor location. With regard to the primary tumor location, adjuvant UFT was associated with a significant benefit in rectal cancer patients with *RAS* mutations in terms of both RFS (5-year RFS, 76.7% and 50.0%, respectively; HR = 0.36; 95% CI, 0.15–0.89; P = 0.02) and OS (5-year OS, 86.7% and 62.5%, respectively; HR = 0.28; 95% CI, 0.09–0.80; P = 0.01), although the difference was not statistically significant in relation to colon cancer patients (5-year RFS, 73.5% and 71.1%; HR = 0.75; 95% CI, 0.33–1.68; P = 0.49; 5-year OS, 82.4% and 76.3%; HR = 0.80; 95% CI, 0.33–1.91; P = 0.61, respectively; Table 2). In colon cancer patients without *RAS* mutations, there was a trend toward shorter survival in the adjuvant UFT group compared to the surgery alone group. However, in the case of the adjuvant UFT group, the HR of recurrence or death in those patients with *RAS* mutations as opposed to patients without *RAS* mutations was 0.65 (95% CI, 0.30–1.41) in colon cancer patients, and 0.65 (95% CI, 0.26–1.63) in rectal cancer patients. Therefore, the effects of UFT on patients, either with or without *RAS* mutation, were similar for both colon and rectal cancer.

Interaction analysis showed that, regarding primary tumor location, tumor stage, and MMR status, RFS was better among patients with *RAS* mutations treated with UFT (HR = 0.60; 95% CI, 0.33–1.08; P = 0.09), but not in patients with non-mutated *RAS* (HR = 0.80; 95% CI, 0.49–1.31; P = 0.37; Table S3).

Discussion

In this report, we undertook a combined analysis of two phase III randomized trials and determined the association of extended *RAS* (*KRAS*/*NRAS*) and MMR status with the effectiveness of surgery plus adjuvant UFT chemotherapy,

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Fig. 3. Forest plots of hazard ratios for overall survival time in selected colorectal cancer patient groups. dMMR, deficient DNA mismatch repair; pMMR, proficient mismatch repair; UFT, tegafur–uracil.

 Table 2. Treatment efficacy results according to RAS mutation status in patients with stage III colorectal cancer

	No RAS mutation (all)		RAS mutation (all)	
Parameter	Surgery alone (n = 82)	UFT (<i>n</i> = 88)	Surgery alone (n = 70)	UFT (<i>n</i> = 64)
5-year RFS, % HR (95% Cl) <i>P</i> -value 5-year OS, % HR (95% Cl) <i>P</i> -value	64.6 0.89 (0.55–1.46) 0.67 77.3 0.94 (0.51–1.71) 0.83	63.5 82.9	61.4 0.49 (0.27–0.91) 0.02 70.0 0.51 (0.26–0.97) 0.03	76.5 84.4
Parameter	No RAS mutation (colon)		RAS mutation (colon)	
	Surgery alone (n = 45)	UFT (n = 47)	Surgery alone (n = 38)	UFT (n = 34)
5-year RFS, % HR (95% Cl) <i>P</i> -value 5-year OS, % HR (95% Cl) <i>P</i> -value	71.1 1.35 (0.66–2.73) 0.40 77.8 1.44 (0.66–3.15) 0.34	59.5 74.5	71.1 0.75 (0.33–1.68) 0.49 76.3 0.8 (0.33–1.91) 0.61	73.5 82.4
Parameter	No <i>RAS</i> mutation (rectum)		RAS mutation (rectum)	
	Surgery alone (n = 37)	UFT (n = 41)	Surgery alone (n = 32)	UFT (<i>n</i> = 30)
5-year RFS, % HR (95% Cl) <i>P</i> -value 5-year OS, % HR (95% Cl) <i>P</i> -value	56.8 0.55 (0.27–1.12) 0.09 75.7 0.48 (0.20–1.13) 0.08	68.0 92.6	50.0 0.36 (0.15–0.89) 0.02 62.5 0.28 (0.09–0.80) 0.01	76.7 86.7

CI, confidence interval; HR, hazard ratio; OS, overall survival; RFS, relapse free survival.

compared with surgery alone, in stage III colorectal cancer patients. In our patient group, we observed that *KRAS/NRAS* mutations were significantly associated with the benefit of adjuvant UFT chemotherapy. However, in those patients with tumors containing only *KRAS* exon 2 mutations, the difference was not statistically significant. In contrast, among patients with non-mutated *KRAS/NRAS*, there was no significant difference in RFS and OS between the surgery-alone and adjuvant UFT groups. To our knowledge, we are the first group to show that extended *RAS* mutations, not only those in *KRAS* exon 2, affect survival in stage III colorectal cancer patients treated with adjuvant fluoropyrimidine-based chemotherapy in comparison with no chemotherapy.

In the Quick and Simple and Reliable (QUASAR) trial, in which postoperative colorectal cancer patients were assigned to either adjuvant 5-FU/LV chemotherapy or no chemotherapy, the risk of recurrence was significantly higher for *KRAS* mutant compared to *KRAS* wild-type tumors.⁽²⁴⁾ However, 91% of patients in that trial had stage II disease, and OS was not evaluated. Additionally, the Kirsten Ras in Colorectal Cancer Collaborative Group (RASCAL) study reported an increased risk of recurrence and death linked to *KRAS* mutations.⁽²⁵⁾ However, this study was based on a large collection of patients enrolled in different studies from various countries. Our data are based on the analysis of a homogenous, prospective cohort that was treated and followed according to the highest clinical standards.

We observed that MMR status was neither prognostic nor predictive for adjuvant chemotherapy. This result differed from other studies reporting that dMMR predicts response to adjuvant fluoropyrimidine-based chemotherapy.^(17–19) Eight percent of tumors were dMMR in our study, which is less than that observed in previous reports, (11-14%).^(17,19,26) The reason may relate to differences in primary tumor location. Hutchins *et al.* showed that 26% of right-sided colon, 3% of left-sided colon, and 1% of rectal tumors were dMMR.⁽²⁴⁾ In our study, nearly half (47%) of all tumors consisted of rectal cancer. Overall, the small number of dMMR tumors in this study may have reduced the power to detect prognostic and predictive value in respect to MMR status. With regard to primary tumor location, adjuvant UFT was associated with a significant improvement in rectal cancer, but not colon cancer. We hypothesize that the benefit of adjuvant UFT was small for colon cancer patients because the outcome of surgery alone is better in colon cancer than rectal cancer. However, the HR of recurrence or death in patients with *RAS* mutations, as opposed to patients without *RAS* mutations, was equal in both colon and rectal cancer patients (both 0.65). Hence, adjuvant UFT chemotherapy improved survival in colon cancer patients, as well as in rectal cancer patients, if *RAS* mutations were present in tumors.

Today, the standard of care at most centers is fluorouracil or capecitabine combined with oxaliplatin (FOLFOX or CapeOX regimen) for all patients with stage III disease. However, some patients are harmed due to toxicity resulting from oxaliplatin treatment, with persistent neuropathy sometimes taking place.⁽²⁷⁾ Our study showed the potential that patients without *RAS* mutations may not need any adjuvant chemotherapy. Although it may not be realistic to undertake a new surgeryalone trial in the future, it is a welcome change for many patients if the non-inferiority of surgery alone relative to surgery plus adjuvant chemotherapy is shown in patients without *RAS* mutations in clinical trials.

An explanation why *RAS* mutations are associated with the effectiveness of UFT is unknown. However, Maus *et al.* showed that mutant *KRAS* status was associated with a lower expression of TS, an enzyme related to 5-FU sensitivity.⁽²⁸⁾ Inhibition of TS is considered to be the main mechanism for the activity of 5-FU. Laboratory studies have shown that acquired resistance to 5-FU has been associated with increased TS expression.^(29,30) Therefore, *RAS* mutations may affect the metabolic pathway of fluoropyrimidines.

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In conclusion, *KRAS/NRAS* mutations are predictive indicators, with respect to the efficacy of adjuvant UFT chemotherapy, in patients with resected stage III colorectal cancer. Further validations of these findings and elucidation of the underlying mechanisms are warranted in future.

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Disclosure Statement

The authors have no conflict of interest.

Abbreviations

5-fluorouracil
confidence interval
deficient DNA mismatch repair
hazard ratio
leucovorin
MutL homolog 1
mismatch repair
MutS homolog
National Surgical Adjuvant Study of Colon Cancer
National Surgical Adjuvant Study of Rectal Cancer
overall survival
PMS1 homolog 2, mismatch repair system component
relapse-free survival
thymidylate synthase
tegafur–uracil

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Supporting Information

Additional Supporting Information may be found online in the supporting information tab for this article:

Table S1. KRAS, NRAS, BRAF, and PIK3CA mutation status in patients with stage III colorectal cancer.

Table S2. Multivariate analysis for relapse-free survival in patients with stage III colorectal cancer treated with surgery alone.

Table S3. Relapse-free survival in patients with stage III colorectal cancer: interaction with RAS mutation and treatment arm.