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OPEN Correlation of Gli1 and HER2 expression in gastric cancer: Identification of novel target

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HER2 becomes the standard of care for guiding adjuvant treatment of gastric cancer with trastuzumab in recent years. However, the usage of this target agent is still limited because of the resistance to trastuzumab or the negative expression of HER2 in tumor tissues. The Gli1 and HER2 both play an important role in the pathogenesis of gastric cancer. However, the correlation of them is still unclear. Here we found Gli1 and HER2 are highly expressed in gastric cancer tissues, and they are positively related. Next, we found Gli1 positive patients live a shorter survival time no matter HER2 positive or negative. Furthermore, univariate and multivariate analysis revealed that venous invasion, HER2 expression, Gli1 expression were independent prognostic factors for the survival time in gastric cancer. In addition, suppressing the expression level of Gli1 can decrease the cell viability and migration ability in cells and subcutaneous tumors. Finally, we found that HER2 may regulate Gli1 by Akt-mTOR-p70S6K pathway. Inhibit of HER2 and SMO have synergistic effect on reduction of cell viability. In conclusion, Gli1 is a favorable prognostic indicator in gastric cancer. As a novel target, Gli1 worth further study, especially in Her2-targeted therapy-resistant cancers.

Cancer is the leading cause of death among men aged 45 to 79 in recent 5 years in US¹. In 2012, there were 951,000 cases of gastric cancer worldwide, the highest regional rates were in Eastern/Southeastern Asia². H. pylori infection³, environmental problems and inherited genes all contribute to gastric oncogenesis. Nowadays, the complex therapies including surgery, chemotherapy and molecular targeted therapy have been used widely. However, advanced gastric cancers still face poor therapeutic effect and terrible drug resistance⁴⁻⁶. Therefore, it is very urgent to identify novel therapeutic targets to treat this fatal disease.

The Hedgehog (HH) pathway plays a crucial role in cell proliferation and metastasis in human tumors⁷⁻¹⁰. It has three legends, Sonic Hedgehog (SHH), Indian Hedgehog (IHH) and Desert Hedgehog (DHH) in classical HH signaling pathway. When the pathway is activated, the HHs bind to 12-transmembrane receptor Patched (PTCH) and then depresses it. The inhibition of PTCH resulted in the release the restraining of Smoothened (SMO, a G-coupled receptor-like protein). Then SMO translocates to the primary cilium and phosphorylate SUFU. Subsequently, Gli zinc-finger transcription factors, including Gli1, Gli2 and Gli3, translocate into nucleus and induce the regulatory expression of HH target genes¹¹⁻¹³. Emerging researches have suggested that Hedgehog pathway is closely associated with poor prognosis and drug resistance of cancer¹⁴⁻¹⁶.

Overexpression of human epidermal growth factor receptor 2 (HER2) in gastric cancer is related to poor outcome^{17,18}. Because the HER2 positive gastric cancer patients receive significant benefit from trastuzumab, testing the expression of HER2 before targeted therapy is necessary¹⁹. With the more and more severe condition of drug resistance of trastuzumab, the way to prolong survival time of gastric cancer patients is limited. New studies have suggested that the expression of Gli1 is related to prognosis in HER2 positive breast cancer, and Gli1 is regulated by HER2 via PI3K-Åkt pathway in esophageal adenocarcinoma^{20,21}. That is to say, inhibiting Hedgehog pathway, especially targets Gli1, may become one of the effective treatments of this lethal disease. However, only few investigations were done to explore the relationship of Gli1 and HER2 in gastric cancer. And there are fewer studies targeting Gli1 according to their relationship. In this study, we tested the expression level of Gli1 and HER2 in specimens, and analyzed their relationship through cell researches. We retrospectively evaluated the impact of Gli1 on the outcomes of gastric cancer patients. Furthermore, we validated the anti-tumor effects of down-regulation of Gli1 in gastric cancer cells and in heterologous subcutaneous caner mice, trying to identify a novel target for therapeutic intervention, especially in HER2-targeted therapy-resistant gastric cancer.

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Figure 1. Expression of Gli1 and HER2 in gastric cancer tissues. (**A**) Immunohistochemical staining of Gli1 and HER2 in 67 human gastric cancer tissues and surrounding tissues($200 \times$). The protein expression of Gli1 was negative (a), weak (b), positive (c), strong positive (d), and the protein expression of HER2 was negative (e), weak (f), positive (g), strong positive (h), respectively. (**B**) Staining scores analysis of Gli1 and HER2 expression in 67 cancer tissue samples and their surrounding tissues. ***P < 0.001.

| | HER2 positive | HER2 negative | P value |
|---------------|------------------|---------------|---------|
| Gli1 positive | 17 | 28 | 0.043* |
| Gli1 negative | 3 | 19 | |

Table 1. Statistics of HER2 and Gli1 expression in human gastric cancer tissues. *P < 0.05.

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Results

Expression levels of Gli1/HER2 in human gastric cancer tissues and paired adjacent tissues. We detected the expression level of Gli1 and HER2 in the 67 primary human gastric cancer tissues and paired adjacent healthy tissues by immunohistochemistry (IHC). The percentage and intensity scores were then multiplied to obtain a total staining score (Fig. 1A,B). We judged positive or negative according to the rule mentioned above. After IHC staining, staining scores of Gli1 and HER2 were calculated. The result showed the expression levels of Gli1 and HER2 were significantly higher in gastric cancer than in para-cancer tissues (P < 0.001 and P < 0.001, Fig. 1B). Thus, we concluded that Gli1 and HER2 were overexpressed in gastric cancer tissues. For further study, we investigated whether Gli1 correlates with HER2 expression in tissues. By IHC, we found Gli1 was positively expressed in 17 out of 20 HER2 positive tumor tissues, while in 28 out of 47 HER2 negative tumors tissues, fourfold table chi-square test showed the expression level of Gli1 and HER2 was related (P = 0.043, Table 1).

Association between Gli1/HER2 expression levels and the clinicopathological factors. We accessed whether the expression level of Gli1 or HER2 is associated with clinicopathological factors of postoperative gastric cancer patients (Table 2). Pearson χ^2 test analysis showed that expression level of Gli1 is correlated to the depth of tumor invasion (P < 0.001), lymph node metastasis (P = 0.021) and TNM staging (P = 0.003). And, HER2 had a significant relationship with depth of tumor invasion (P = 0.013), lymph node metastasis (P = 0.014) and TNM staging (P = 0.006). There is no correlation between Gli1/HER2 and other clinicpathological parameters such as age, gender or size of tumor (P > 0.05).

| | Gli1 | | | HER2 | | | |
|---------------------------|----------|----------|-----------|----------|----------|---------|--|
| | Negative | Positive | Р | Negative | Positive | Р | |
| Age(years) | | | | | | | |
| ≤ 60 | 9 | 16 | 0.670 | 17 | 8 | 0.767 | |
| >60 | 13 | 29 | | 30 | 12 | | |
| Gender | | | | | | | |
| Male | 18 | 32 | 0.344 | 35 | 15 | 0.963 | |
| Female | 4 | 13 | | 12 | 5 | | |
| Size(cm) | | | | | | | |
| ≤ 5 | 13 | 27 | 0.943 | 26 | 14 | 0.262 | |
| >5 | 9 | 18 | | 21 | 6 | | |
| Depth of tumor invasion | | | | | | | |
| T1-2 | 10 | 2 | <0.001*** | 12 | 0 | 0.013* | |
| T3-4 | 12 | 43 | | 35 | 20 | | |
| Degree of differentiation | | | | | | | |
| Well | 10 | 22 | 0.792 | 24 | 8 | 0.407 | |
| Poor | 12 | 23 | | 23 | 12 | | |
| Lymph node metastasis | | | | | | | |
| Yes | 11 | 35 | 0.021* | 28 | 18 | 0.014* | |
| No | 11 | 10 | | 19 | 2 | | |
| TNM staging | | | | | | | |
| I-II | 13 | 10 | 0.003** | 21 | 2 | 0.006** | |
| III-IV | 9 | 35 | | 26 | 18 | | |

Table 2. Association between Gli1/HER2 and clinic-pathological factors in 67 patients with gastric cancer.*P < 0.05, **P < 0.01, ***P < 0.001.

Gli1 expression links with gastrectomy patients' survival. To elaborate whether the expression levels of Gli1/HER2 in gastric cancer tissues exert an influence on overall survival of patients after gastrectomy, we analyzed survival curves according to target genes expression. In result, the survival of patients with positive Gli1 expression was worse than those with negative Gli1 expression (P < 0.001, Fig. 2A). Whereas the patients with negative HER2 got a modest survival benefit, which didn't reach a significant difference (P = 0.170, Fig. 2B). Based on the expression level of HER2, we further analyzed the association between Gli1 expression and survival of patients. As the outcome shows, no matter the expression of HER2 was positive or negative, the survival of gastric cancer patients with positive Gli1 expression is poorer than the patients with negative ones (P = 0.003, P = 0.045, Fig. 2C,D). Univariate analysis revealed that Gli1 expression, venous invasion, TNM staging, depth of tumor invasion, lymph node metastasis, size of tumor, differentiation, neural invasion were associated with an inferior survival duration (P < 0.05, Table 3). However, only venous invasion, HER2 expression, Gli1 expression were verified to be independent prognostic factors for the survival in gastric cancer patients after multivariate analysis (P < 0.05, Table 3), especially Gli1 expression (P < 0.001, Table 3).

To further explore the influence of different Gli1 expression on other prognostic factors, we performed a subgroup analysis. The results showed that Gli1 expression significantly shortened the survival no matter the patients' age, the size of tumor, differentiation, venous invasion or neural invasion (P > 0.05, Fig. 3). For male patients, the survival time was worse in those with positive Gli1 expression than female patients. Meanwhile, Gli1 expression affected the survival in patients with stage T3-T4 (P=0.021, Fig. 3), positive lymph node metastasis (P=0.001, Fig. 3) and stage III-IV (P=0.012, Fig. 3), indicating that patients at the same depth of tumor invasion (stage T3-T4), lymph node metastasis (positive) or same TNM stage (stage III-IV), patients with Gli1 positive expression could have a significant poorer survival than others.

Gli1 depleted depresses the growth of gastric cancer *in vivo* and *in vitro*. As described above, the expression level of Gli1 is an independent prognostic factor of the survival duration of gastric cancer patients after operation. As a new target, Gli1 shows a cheerful prospect. To confirm the finding that decreasing the expression of Gli1 can further inhibit the tumorgenesis, we did studies both *in vivo* and *in vitro*. The efficiency of Gli1 knockdown in the two groups was shown in Fig. 4A. We transfected SGC7901 cells with control-shRNA or Gli1-shRNA. Compared with control group, the cells showed decreased cell viability and migration ability after knockdown Gli1 (Fig. 4B,C). Furthermore, we evaluated the effect of Gli1 in a mice model. We selected 3-4 week nude mice. We implanted the same number of SGC7901 gastric cancer cells, which treated differently with control-shRNA or Gli1-shRNA onto the subcutaneous sites of mice and measured tumor size twice a week. Then, we found that the subcutaneous tumors treated with Gli1 depletion grow much slower and relatively get a more steady weight (Fig. 4D–F). After stripping the subcutaneous tumors, we accessed the weight of tumors and finding the tumors derived from Gli1 depleted group were decreased markedly (P < 0.01, Fig. 4G).



Figure 2. Kaplan-Meier survival analysis in patients after gastrectomy. (**A**) 67 gastric cancer patients after operation were analyzed according to the Gli1 expression. In this 67 patients, Gli1 positive expression was 45 and negative expression was 22. (**B**) 67 gastric cancer patients after operation were analyzed according to the HER2 expression. HER2 positive expression was 20 and negative expression was 47. (**C**) 47 gastric cancer patients with negative HER2 expression after operation were analyzed according to the Gli1 expression. Gli1 positive expression was 28 and negative expression was 19 in patients. (**D**) 20 gastric cancer patients with positive HER2 expression after operation were analyzed according to the Gli1 expression. Gli1 positive expression was 3 in patients.

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Gli1 Crosstalk with HER2 through AKT-mTOR pathway. We found there is a significant association between Gli1 and HER2 expression in gastric cancer specimens. Thus, we further analyzed their relationship in cell lines. As a first step, we select a gastric cancer cell line, SGC7901 in our study. We treated SGC7901 with trastuzumab (the HER2 inhibitor) for 48 hours at the concentration of 10 or 20 µg/ml respectively. Compared with the control group, except SMO (Gli1 upstream target), the expression level of Gli1, p-AKT, p-mTOR and p-p70S6k (mTOR downstream target) decreased obviously (Fig. 5A,B). The quantitative statistics see Supplementary Fig. S1A,B. In order to further explore the result, we detected the association between Gli1 and Akt-mTOR, the downstream pathway of HER2. Some studies revealed there is a crosstalk between mTOR and Gli1²². We used rapamycin, the specific mTORC1 inhibitor, to treated SGC7901 for 48 hours and found the protein expression of Gli1 decreased distinctly, but SMO, the upstream of Gli1, was unchanged (Fig. 5C). The quantitative statistics see Supplementary Fig. S1C. To further confirm the result, SGC7901 was treated with $TNF\alpha$, which reported could enhance the expression of Gli1 via mTOR-p-7086K-Gli1 axis²². Western blot, proliferation and migration ability assays showed rapamysim could reverse the function of $TNF\alpha$, while cyclopamine (a canonical hedgehog pathway inhibitor target on SMO) could not (Fig. 5D-F). The transwell picture see Supplementary Fig. S1D. These researches suggested that HER2 may regulate Gli1 by Akt-mTOR-p-70S6K pathway, not canonical SMO-Gli1 pathway.

Inhibit of HER2 and SMO has synergistic effect on reduction of cell viability. Next step, we tested the impact of trastuzumab and Vismodegib (the SMO inhibitor) on the viability of SGC7901. After a 48 treatment, cells treated with trastuzumab reached a reduction plateau of cell viability at the concentration of $20 \,\mu$ g/ml (Fig. 5G). And the reduction plateau of Vismodegib was $20 \,\mu$ mol/l (Fig. 5H). Whereas, the reduction plateau of the dose response curve decreased obviously when we combined trastuzumab and Vismodegib to treat the cells

| | | | Univariate analysis | | Multivariate analysis | |
|--------------------------|-----------------|--------------------|---------------------|--------------------|-----------------------|--------------------|
| Factor | Group | Number of patients | HR (95% CI) | P value (log-rank) | HR (95% CI) | P value (log-rank) |
| 1.00 | \leq 60 years | 25 | 1 | | 1 | |
| Age | >60 years | 42 | 1.196 (0.693-2.064) | 0.520 | 1.360 (0.681–2.715) | 0.384 |
| Gender | Male | 50 | 1 | | 1 | |
| | Female | 17 | 1.320 (0.719–2.424) | 0.370 | 2.118 (0.894-5.019) | 0.088 |
| Size of tumor | \leq 5 cm | 38 | 1 | | 1 | |
| | >5 cm | 29 | 0.428 (0.246-0.745) | 0.003** | 1.089 (0.563-2.108) | 0.800 |
| Dopth of tumor invasion | T1-2 | 12 | 1 | | 1 | |
| Depth of tunior invasion | T3-4 | 55 | 0.183 (0.072-0.465) | <0.001*** | 1.138 (0.257-5.038) | 0.865 |
| Lymph node metastasis | Negative | 21 | 1 | | 1 | |
| | Positive | 46 | 0.123 (0.058-0.258) | <0.001*** | 0.222 (0.026-1.882) | 0.168 |
| Differentiation | Well | 36 | 1 | | 1 | |
| Differentiation | Poor | 31 | 2.255 (1.288-3.947) | 0.004** | 0.746 (0.315-1.766) | 0.505 |
| Vananaian | Negative | 38 | 1 | | 1 | |
| venous invasion | Positive | 29 | 0.361 (0.208-0.627) | <0.001*** | 0.357 (0.171-0.745) | 0.006** |
| Neural invasion | Negative | 42 | 1 | | 1 | |
| iveural invasion | Positive | 25 | 0.453 (0.262-0.781) | 0.004** | 0.516 (0.255-1.047) | 0.067 |
| TNM staging | I - II | 23 | 1 | | 1 | |
| | III - IV | 44 | 0.121 (0.060-0.245) | <0.001*** | 0.233 (0.022-2.423) | 0.223 |
| LIED2 ammassion | Negative | 47 | 1 | | 1 | |
| HER2 expression | Positive | 20 | 0.678 (0.385-1.194) | 0.178 | 2.200 (1.090-4.442) | 0.028* |
| CI II orprossion | Negative | 22 | 1 | | 1 | |
| GLITEXPIESSION | Positive | 45 | 0.332 (0.179–0.615) | <0.001*** | 0.210 (0.093-0.473) | <0.001*** |

Table 3. Results of univariate and multivariate analyses of gastrectomy patients' survival by Cox's proportionalhazard model.

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(Fig. 51). That is to say, inhibit Gli1 both via HER2–Akt–mTOR–p-70S6K and Hedgehog pathway has synergistic effect on gastric cancer cells.

Discussion

Gastric cancer still faces poor prognosis and low survival rate in the world, especially in Asia²³. Although the improvement of surgical techniques and the widely use of chemotherapies prolong the survival rate modestly, gastric cancer patients still face diverse side effects and severe drug-resistance²⁴. Testing the expression of HER2 becomes the standard of care for guiding adjuvant treatment of gastric cancer with trastuzumab^{19,25,26}. However, because of the resistance to HER2 inhibitor trastuzumab or the negative expression of HER2 in tumor tissues, the usage of this target agent is still limited²⁷⁻²⁹. So, identifying novel therapeutic targets in gastric cancer is essential.

The Hedgehog pathway is important in the oncogenesis of many human tumors^{7–10}. Promoting Gli zinc-finger transcription factors translocate into nucleus, especially Gli1 is associated with tumor proliferation, metastasis and drug resistance^{14–16}. In our study, we detected 67 gastric cancer tissues and showed Gli1 was related to several clinicopathological factors, such as depth of tumor invasion, lymph node metastasis and TNM staging.

Our study showed the expression levels of Gli1 and HER2 were significantly higher in gastric cancer than in para-cancer tissues. As reported, there is a significant association between the expression of Gli1 and HER2²¹. Activation of Hedgehog pathway is a multiple process and the level of Gli1 affected by several signal pathway through the crosstalk between them. A potential SMO-independent crosstalk is identified between PI3K-Akt-mTOR and Gli1^{21,30-33}. Meanwhile, PI3K-Akt is the main downstream of HER2. Thus, HER2 may regulate the expression level of Gli1 by this uncanonical pathway. To confirm this hypothesis, at first, we investigated the effect of HER2 inhibitor. After adding trastuzumab to gastric cancer cells, the expression level of Gli1 decreased while SMO remained unchanged. This suggests HER2 may regulate Gli1 via SMO-independent way. Then we tested the expression of p-Akt, p-mTOR, p-p70S6k with the same treatment, and found they all decreased significantly. This means HER2 regulates its main downstream pathway PI3K-Akt-mTOR^{34,35}. Then we added rapamycin to gastric cancer cells, the expression level of p-p70S6k and Gli1 decreased while SMO remained unchanged. This means mTOR interacts with Gli1 via the crosstalk. In conclusion, HER2-Akt-mTOR-S6K-Gli1 axis may be one of the pathways which HER2 regulates Gli1.

Since the interaction between Gli1 and HER2, we assessed the influence of Hedgehog pathway and HER2 inhibitors exerted on the viability of cells. Our research found the viability of cells was reduced after treated with trastuzumab or vismodegib distinctively. Moreover, the combination of trastuzumab and vismodegib has a synthesis impact on depressing cell viability.

Some previous studies considered that the high expression level of Gli1 and HER2 indicates a poor prognosis^{17,18,36,37}. Whereas, in survival analysis of the 67 patients after surgery according to gastric cancer tissues IHC outcome, the differential expression of Gli1 related to different survival time (P < 0.05) while the HER2 didn't (P = 0.170). Besides, higher Gli1 expression also leads to poorer survival time in HER2-positve patients, which

| Number of patients | | | HR (95% CI) | P value | P value for interaction | |
|--------------------|--------------|--------|-------------|---------------------|----------------------------|------|
| Age | | | | | | 0.86 |
| | ≤60 years | 9/16 | | 0.304 (0.114-0.806) | 0.017 | |
| | >60 years | 13/29 | | 0.342 (0.153-0.766) | 0.009 | |
| Gender | | | | | | 0.65 |
| | Male | 18/32 | | 0.290 (0.144-0.587) | 0.001 | |
| | Female | 4/13 | | 0.408 (0.111-1.500) | 0.177 | |
| Size of tumor | | | | | | 0.60 |
| | ≤5 cm | 14/24 | | 0.331 (0.139-0.789) | 0.013 | |
| | >5 cm | 8/21 | | 0.228 (0.076-0.686) | 0.008 | |
| Depth of tumo | r invasion | | | | | 0.39 |
| | T1-2 | 10/2 + | | 28.907 (0.002->50) | 0.481 | |
| | Т3-4 | 12/43 | | 0.425 (0.205-0.880) | 0.021 | |
| Lymph node n | netastasis | | | | | 0.41 |
| | Negative | 11/10 | | 0.492 (0.138-1.752) | 0.274 | |
| | Positive | 11/35 | | 0.258 (0.112-0.594) | 0.001 | |
| Differentiation | | | | | | 0.65 |
| | Poor | 12/24 | | 0.244 (0.103-0.582) | 0.001 | |
| | Well | 10/21 | | 0.333 (0.122-0.910) | 0.032 | |
| Venous invasi | on | | | | | 0.66 |
| | Negative | 14/24 | | 0.335 (0.140-0.801) | 0.014 | |
| | Positive | 8/21 | <u> </u> | 0.248 (0.090-0.682) | 0.007 | |
| Neural invasio | n | | | | | 0.09 |
| | Negative | 15/27 | | 0.419 (0.192-0.912) | 0.028 | |
| | Positive | 7/18 _ | | 0.094 (0.020-0.434) | 0.002 | |
| TNM staging | | | | | | 0.36 |
| | Stage I-II | 10/13 | | 0.674 (0.217-2.094) | 0.495 | |
| | Stage III-IV | 9/35 | | 0.348 (0.152-0.795) | 0.012 | |
| | | + | | | | |
| | | 0.0 | 2 0.1 1 | 10 50 | | |
| | | GIIT(- |) better | Gli1(+) better | | |

Figure 3. Subgroup analysis for the influence factor of survival duration of gastrectomy patients according to Gli1 expression. There were 9 Gli1 negative patients and 16 Gli1 positive patients in the gastric cancer patients after operation who aged under 60 years old. The expression level of Gli1 was associated with the survival duration in patients under 60 years old (P = 0.017), so as in patients over 60 years old (P = 0.009). The same in other scale such as gender, the size of tumor, depth of tumor invasion, lymph node metastasis, differentiation, venous invasion, neural invasion and TNM staging.

is in line with the study Liu done in breast cancer²⁰. Interestingly, the survival time of patients also decreased when they had a high expression of Gli1, if their basal HER2 expression is negative. It means, Gli1 expression is related to the survival time of patients. No matter the expression level of HER2, when the Gli1 expression higher, the prognosis of gastric cancer patients becomes poorer. To further explore the influence factors of the survival duration of gastric cancer patients after operation, we conducted univariate analysis and multivariate analysis, which showed venous invasion, HER2 expression, Gli1 expression were independent prognostic factors for the survival in gastric cancer patients, especially Gli1 expression. Subgroup analysis indicated that patients with Gli1 positive expression could have a significant poorer survival than others at the same depth of tumor invation (stage T3-T4), lymph node metastasis (positive) or same TNM stage (stage III-IV). In conclusion, Gli1 is an important prognosis factor of gastric cancer.

Furthermore, we confirmed it *in vitro* and *in vivo*. Firstly, we investigated the ability of cell proliferation and migration after knockdown of Gli1 in SGC7901 cells. And, the ability of cells were restricted distinctly. It explained the role Gli1 plays in the tumor growth and metastasis³⁸⁻⁴⁰. Then, 3-4 week nude mice were selected. And cells treated with control-shRNA or Gli1-shRNA were injected into subcutaneous tissues respectively. We considered the basal expression of HER2 in nude mice was similar. As expected, mice injected with Gli1-shRNA cells got a slower tumor growth and a steady weight relatively. Thus, we conclude that decreasing the Gli1 expression leads a benefit no matter the expression level of HER2. It suggests that the synthetic therapy of Gli1 suppressor and trastuzumab may act as an effective therapy of gastric cancer.

Taken together, the novel finding in this study, HER2 relates the expression of Gli1, and HER2 may regulates Gli1 in SMO-independent pathway via HER2-Akt-mTOR-S6K-Gli1 axis. There was negative influence of Gli1 on survival, and the impact is independent on the expression of HER2. Inhibiting the expression level of Gli1 leads to a great benefit *in vitro* and *in vivo*. It suggests that Gli1 may act as a novel gastric cancer suppressor. In HER2-positive patients, Gli1 inhibitor combined with trastuzumab may be an ideal choice. Meanwhile, if the HER2 expression is negative, the patients also can benefit from the use of Gli1 inhibitor. These evidence may highlight a new therapeutic strategies to gastric cancer by Gli1 inhibitor applied.



Figure 4. Knockdown of Gli1 suppressing gastric tumor growth *in vitro* and *in vivo*. (**A**) SGC7901 cells were transfected with negative control shRNA (CTL) or Gli1 shRNA (Gli1-KD). The protein was analyzed by Western blot. (**B**) MTT assay was used to test the proliferation ability of cells transfected with negative control shRNA (CTL) or Gli1 shRNA (Gli1-KD), presented as mean \pm S.E.M. (n = 3). (**C**) Transwell assay was used to test the migration ability of cells transfected negative control shRNA (CTL) or Gli1 shRNA (Gli1-KD). Representative photographs were presented (X100) and the relative number of migratory cells were counted as mean \pm S.E.M. (n = 5). (D-G) SGC7901 cells transfected with negative control shRNA (CTL) or Gli1 shRNA (Gli1-KD) stably were transplanted into nude mice (n = 8). (**D**) The volume of the tumors was measured twice a week during the indicated 3 weeks. The tumor mass of each group was presented as mean \pm S.E.M. (n = 8). (**E**) The weight of mice was measured twice a week during the indicated 3 weeks. The weight of mice in each group was presented as mean \pm S.E.M. (n = 8). (**F**) Tumor was tested by immunohistochemical staining of Gli1 in each group after mice killed. (**G**) The weight of tumor mass in each group was also presented. Data was presented as mean \pm S.E.M. (n = 8). **P* < 0.05, ***P* < 0.01.

Methods

Patient information and follow up. A total of 67 surgically resected gastric carcinoma cases were obtained from Suzhou Municipal Hospital (Suzhou, China) from January 2008 to December 2010. All patients meet the criterial: (1)Tumor was confirmed to be gastric adenocarcinomar by pathological examination. (2)None of them received radiotherapy or chemotherapy before operation. (3) Everyone has complete clinical data and was available to be follow up. (4) All patients written informed consent. After surgery, each patient received a follow-up regularly. Until 60 months, the patients who remained alive record as 60 months.



Figure 5. HER2 regulates Gli1 via Akt-mTOR pathway, combination treatment of trastuzumab and vismodegib obtained a synergistic effect. (A) The expression of SMO and Gli1 was investigated via Western blotting after SGC7901 treated with Trastuzumab (the HER2 inhibitor) for 48 h. (B) The expression of p-Akt, p-mTOR, p-70S6K was tested by Western blotting after SGC7901 treated with Trastuzumab for 48 h. (C) The expression of p-70S6K, SMO and Gli1 was detected via Western blotting after SGC7901 treated with rapamycin (The mTORC1 inhibitor) for 48 h. (D) The expression of Gli1 was detected via Western blotting after SGC7901 were treated with TNF α (5ng/ml), TNF α (5ng/ml) and cyclopamine (1 μ M) or TNF α (5 ng/ml) and rapamycin $(0.5 \mu M)$ for 24 h. (E) MTT assay was used to test the proliferation ability of cells treated with TNF α (5ng/ ml), TNF α (5ng/ml) and cyclopamine (1 μ M) or TNF α (5 ng/ml) and rapamycin (0.5 μ M) for 24 h, 48 h and 72 h, presented as mean \pm S.E.M. (n = 3) (**P < 0.01). (F) Transwell assay was used to test the migration ability of cells treated with TNF α (5ng/ml) (T), TNF α (5ng/ml) and cyclopamine (1 μ M) (T + C) or TNF α (5 ng/ml) and rapamycin $(0.5 \mu M)$ (T + R) for 24 h. (G) Trastuzumab reduced the viability of SGC7901 significantly. The cells treatED with different concentrations of trastuzumab for 48 h. (H) Vismodegib(the SMO inhibitor) reduced the viability of SGC7901 significantly. The cells treatment with different concentrations of vismodegib for 48 h. (I) The combined or single treatment of trastuzumab and vismodegib for SGC7901 was analyzed. **P* < 0.05, ***P* < 0.01, ****P* < 0.001.

Preparation of human tissue samples. Paired human gastric cancer tissues and adjacent normal tissues were collected from surgically resected patients at the Department of General Surgery, the Suzhou Municipal Hospital (Suzhou, China) from 2008 to 2010. The study was approved by the Independent Ethics Committee of Suzhou Municipal Hospital. Meanwhile, we confirm that all experiments were performed in accordance with relevant guidelines and regulations.

Immunohistochemistry. Fix the surgical specimens with formalin and embed them with paraffin. Then the tissues were cut into 5μ m. After dewaxing and rehydrating, sections were incubated with the polyclonal antibodies of Gli1 or HER2 (Cell Signaling Technology, USA; 1:200 dilution) at the room temperature for 2 or 3 hours. The process was performed with the tissue staining kit (Zhongshan Biotechnology, Beijing, China) following the manufacturer's protocol. Two researchers got the outcome of immunostaining respectively. Five 200 × random regions were analyzed and classified into five levels according to the percentage of positively staining cells per section: absent, $0 \sim 5\%$; 1, $6 \sim 25\%$; 2, $26 \sim 50\%$; 3, $51 \sim 75\%$; 4, >75%. We deemed the staining intensity as follows: 0 (negative); 1 (weak); 2 (moderate); 3 (strong). The percentage and intensity scores were then multiplied to obtain

a total score (staining score = percentage score × intensity score). For the staining score, 0 was deemed as (–), $1 \sim 4$ as (+), $5 \sim 8$ as (++), $9 \sim 12$ as (+++). Eventually, we obtained a final average score, and consider (–) or (+) as negative, (++) or (+++) as positive⁴¹.

Cell culture. Human gastric cancer cell lines SGC7901, obtained from the Cell Bank of the Chinese Academy of Sciences (Shanghai, China), were grown in RPMI Medium 1640 (Hyclone) supplemented with 10% fetal bovine serum (Gibco) and 1% penicillin/streptomycin (Gibco) and cultured at 37 °C calorstat with 5% CO₂.

Transfection of gastric cancer cell lines. SGC7901 cell line stably expressing Gli1-specific shRNA or scrambled control-shRNA were constructed using a lentiviral shRNA technique. The sequences specific for human Gli1 (5'-CUCCACAGGCAUACAGGAU-3') selected to inhibit the target gene expression were synthesized by Gene Pharma (Shanghai, China). The lentivirus was titrated to $1 * 10^9$ TU/ml. SGC7901 cells were cultured at the concentration of $5 * 10^4$ /well in a six-well dish for 24 h, then we added lentivirus to the culture medium. After transfection, puromycin was used to screening cells 72 hour later to form the stable cell line. The efficiency of Gli1-shRNA was detected by fluorescence microscope and Western blot.

Protein extraction and western blotting. Whole protein extracted were lysed 30 min with RIPA lysis buffer (SigmaAldrich) according to manufacturer's protocol. Protein collected were stored at -80 °C or used for the next immunoblot. Each sample was separated by SDS-PAGE and transferred to the nitrocellulose membranes (Schleicher & Schuell). Then, the bands incubated with the polyclonal antibody (Cell Signaling Technology) overnight after blocking with 5% non-fat milk for 1 hour at room temperature, and incubated conjugated second antibodies for 1 hour at room temperature. The images of bands were performed by chemiluminescence and results were quantified by ImageJ software.

Cell viability assay. Cell viability was determined by MTT assays kit (Amresco, USA) according to the manufacturer's recommendations. Cells seeded in 96-well plates and added MTT solution mixed with complete medium. After incubation for 4 hours, solution was exchanged of $150 \,\mu$ l DMSO. Set aside for ten minutes into the atmosphere of $37 \,^{\circ}$ C and measure the absorbance at 490 nm. Each treatment had five replicate wells.

Cell migration assays. The ability of cell migration was evaluated with 24-well transwell plate (Corning Incorporated, USA). In each well, cells resuspended in serum-free RPMI 1640 were plated in the upper chambers. And the lower chambers were added $500 \,\mu$ l RPMI 1640 mixed with 10% FBS. Cells that had invaded were stained with 0.5% crystal violet after 24 h incubation. Then, the cells were recorded as pictures via microscopy ($200 \times$). Quantify the level of migration by counting the invaded cells in five random regions per specimen.

Xenografts. 16 Nude mice (BALB/c, SPF grade, 16-18 g, 3-5 weeks old, male), were purchased from Shanghai SLRC laboratory Animal Co (Shanghai, China) and housed in a pathogen-free and 12h light/dark cycle room. All nude mice were injected gastric cancer cells subcutaneously ($5*10^6$ cells per mouse). All nude mice were divided into 2 groups (n = 8 mice per group) after feed 3 days according the weight and recorded the day as 0. Then, they were injected with control-shRNA and Gli1-shRNA cells respectively. We recorded body weight and tumor size twice a week. Meanwhile. Mice were killed at the day 22, and tumor were removed for further detecting. All animal experimental procedures were approved by the Animal Ethics Committee of the Suzhou Municipal Hospital (Suzhou, China). Meanwhile, we confirm that all experiments were performed in accordance with relevant guidelines and regulations.

Statistical analysis. All experiments presented were repeated for three times. Results were expressed as means \pm S.E.M. Student's t-test was used to test the two groups. While the analysis of IHC were performed by chi-square statistical test. Survival durations were calculated using the Kaplan–Meier method and compared by the log-rank test. All parameters that were discovered to be significant on univariate analysis by the Cox proportional hazard model were then went into multivariate survival analysis. *P* < 0.05 was considered to suggest a statistically significant difference. The bars were draw with graphpad prism 5, and pictures were combined by Microsoft power point (PPT).

Availability of data and materials. Data are stored by the corresponding author of this paper and are available upon request.

Consent for Publication. All the individuals provided written informed consent prior to enrolling in the study, and the study was approved by the Ethics Committees of Suzhou Municipal Hospital.

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Author Contributions

X.Y.S. carried out literature research, experimental studies and data acquisition, participated in the study design and drafted the manuscript. X.Y.K. and Z.P. performed the clinic cases collection and carried out the data acquisition, participated in the study design and helped to draft the manuscript. L.P.Z. and L.Y.W. participated in the design of the study and performed the statistical analyses. L.Y.W. and L.J.X. helped to western blots, animal research and data acquisition. C.L.Z. proposed the study and participated in its design and helped to draft, and assisted writing the manuscript. All authors had already read and approved the final manuscript.

Additional Information

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