



Reshaping the Preterm Heart: Shifting Cardiac Renin-Angiotensin System Towards Cardioprotection in Rats Exposed to Neonatal High-Oxygen Stress

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BACKGROUND: Approximately 10% of infants are born preterm. Preterm birth leads to short and long-term changes in cardiac shape and function. By using a rat model of neonatal high-oxygen (80%O₂) exposure, mimicking the premature hyperoxic transition to the extrauterine environment, we revealed a major role of the renin-angiotensin system peptide Angio II (angiotensin II) and its receptor AT1 (angiotensin receptor type 1) on neonatal O₂-induced cardiomyopathy. Here, we tested whether treatment with either orally active compounds of the peptides Angio-(1–7) or alamandine included in cyclodextrin could prevent postnatal cardiac remodeling and the programming of cardiomyopathy induced by neonatal high-O₂ exposure.

METHODS: Sprague-Dawley pups were exposed to room air or 80% O₂ from postnatal day 3 (P3) to P10. Neonatal rats were treated orally from P3 to P10 and assessed at P10 and P28. Left ventricular (LV) shapes were characterized by tridimensional computational atlases of ultrasound images in addition to histomorphometry.

RESULTS: At P10, high O₂-exposed rats presented a smaller, globular and hypertrophied LV shape versus controls. Treatment with cyclodextrin–Angio-(1–7) significantly improved LV function in the O₂-exposed neonatal rats and slightly changed LV shape. Cyclodextrin–alamandine and cyclodextrin–Angio-(1–7) treatments similarly reduced hypertrophy at P10 as well as LV remodeling and dysfunction at P28. Both treatments upregulated cardiac angiotensin-converting enzyme 2 in O₂-exposed rats at P10 and P28.

CONCLUSIONS: Our findings demonstrate LV remodeling changes induced by O₂-stress and the potential benefits of treatments targeting the cardioprotective renin-angiotensin system axis, supporting the neonatal period as an important window for interventions aiming at preventing cardiomyopathy in people born preterm. (**Hypertension**. 2022;**79**:1789–1803. DOI: 10.1161/HYPERTENSIONAHA.122.19115.) • **Supplemental Material**

Key Words: angiotensin-converting enzyme ■ cardiomyopathies ■ cyclodextrins ■ echocardiography ■ premature birth ■ renin-angiotensin system

Globally, ≈10% of all infants are born less than 37 gestational weeks, and 1% are extremely preterm (<28 weeks).¹ Over the past few decades, the rapid translation of scientific findings in this field into clinical practice has stimulated significant improvements in perinatal care. As a direct consequence, nearly 90% of

these infants now survive, including those born the most premature. As the first generation of survivors reaches adulthood, long-term health consequences have started to be revealed.^{2–4}

Although the causes and risk factors of preterm birth are multiple, common clinical cardiovascular implications

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NOVELTY AND RELEVANCE

What Is New?

Orally active compounds of the peptides angiotensin-(1–7) or alamandine prevented postnatal cardiomyocyte hypertrophy and the programming of cardiomyopathy in rats exposed to neonatal high-O₂.

What Is Relevant?

Neonatal rats exposed to high-O₂ have different cardiac remodeling patterns associated with a higher predisposition to develop cardiomyopathy into adulthood, which were prevented by both treatments and more significantly modified by angiotensin-(1–7).

Both treatments stimulated the upregulation of cardiac ACE (angiotensin-converting enzyme) 2, which may balance renin-angiotensin system and prevent the deleterious effects of angiotensin II/AT1 (angiotensin receptor type 1) on heart remodeling and programming of cardiomyopathy in this model.

Clinical/Pathophysiological Implications?

Our findings demonstrate the potential benefits of treatments targeting the cardioprotective renin-angiotensin system axis and support the neonatal period as an important window for interventions aiming at preventing cardiomyopathy in people born preterm.

Nonstandard Abbreviations and Acronyms

ACE	angiotensin-converting enzyme
Angio II	angiotensin II
Angio-(1–7)	angiotensin-(1–7)
AT1	angiotensin receptor type 1
AT2	angiotensin receptor type 2
E/E'	mitral valve Doppler E wave/ tissue Doppler E' wave ratio
LV	left ventricle
MPI	myocardium performance index
RAS	renin-angiotensin system
RV	right ventricle

beyond the neonatal period are now shown as early as childhood.² Changes in heart shape and function are evidence of postnatal cardiac remodeling, which is one of the first signs of cardiovascular adaptations to preterm birth stress.^{5–8} The abrupt hemodynamic shift to a circulatory overload, plus systemic oxidative stress triggered by the early exposure to higher oxygen (O₂) concentrations in the extrauterine environment, are conditions imposed on all preterm newborns at a time when the heart is undergoing an accelerated growth phase.^{9–11} As a result, the hearts of people born preterm are reshaped, with ventricular changes also evident as they grow older.^{5,8} In addition, they show a reduced myocardial efficiency and ventricular systolic dysfunction during physiological exercise stress.¹²

We have previously reported similar findings in a well-established rat model of prematurity-related complications caused by transient neonatal high-O₂ exposure (80% O₂) from 3 to 10 days of life,^{13–16} an age at which cardiomyocyte proliferation is ongoing,¹⁷ mimicking the oxidant stress burst present during preterm birth. By using this model, we first described the progression

of left ventricular (LV) remodeling and dysfunction in male juvenile (28 days old) rats before any blood pressure changes, which only occur after 6 to 7 weeks in this model.^{18,19} Furthermore, we demonstrated that aging and hemodynamic overload predisposed O₂-exposed rats to a rapid transition to heart failure.¹⁸ Hence, clinical and experimental findings underline an inefficient myocardium adaption, being a relevant process contributing to increased risk of heart disease, which has been consistently observed in the preterm-born population.²⁴ However, there is still a knowledge gap on key mechanisms promoting adverse myocardium adaptations and, most importantly, on therapies capable of preventing adverse cardiac remodeling and diseases in preterm-born individuals.

The renin-angiotensin system (RAS) has been shown by us and others to exert an intriguing dueling role in preterm birth, particularly in neonatal heart development.^{18,20,21} RAS consists of 2 antagonistic branches: one prohypertensive and fibrogenic axis composed of ACE (angiotensin-converting enzyme), Angio II (angiotensin II), and its receptors AT1 (angiotensin receptor type 1) and AT2 (angiotensin receptor type 2), known as the ACE/Angio II/AT1-AT2 axis, and the counter regulatory axis with antifibrogenic and cardioprotective properties composed of ACE2, Angio-(1–7) and its receptor Mas, known as the ACE2/Angio-(1–7)/Mas axis. In addition, a newly discovered branch of the system has been added and is formed by alamandine and its receptor MrgD, which is also associated with cardioprotective effects.²² At birth and early postnatal days (P), ACE, the enzyme cleaving Angio I to form Angio II, and both Angio II receptors AT1 and AT2, are physiologically upregulated in hearts of neonatal rats and further exacerbated in those exposed to hyperoxia.²⁰ These main components of the prohypertensive ACE/Angio II/AT1 axis remain upregulated as they grow older and significantly contribute to the establishment of LV

remodeling and dysfunction in this model, which can be prevented by short-term neonatal treatment with losartan.²⁰ Despite being effective at preventing the developmental programming of cardiac dysfunction in our rat model, clinically, the use of losartan and other RAS blockers is contraindicated in newborns due to the high risk of cardiac and renal malformations.^{23,24} Therefore, in this study, we hypothesized that alternative RAS-related therapies that could potentially shift the RAS towards its cardioprotective axis, without blocking the physiological role of Angio II and AT1, could prevent postnatal cardiac remodeling and the early programming of cardiomyopathy induced by neonatal high O₂ exposure.

Here, we aim to describe the effects of neonatal treatments with orally active compounds of Angio-(1–7) or alamandine included in β-cyclodextrins on neonatal heart's shape, remodeling, and function associated with molecular changes in cardiac RAS profile, and on the long-term programming of ventricular dysfunction induced by neonatal high-O₂ exposure. To reach this goal, in addition to the functional and histomorphological cardiac assessments, we developed a noninvasive method of ultrasound imaging of neonatal rats to quantitatively capture in vivo 3-dimensional (3D) geometric ventricular changes caused by neonatal high-O₂ stress and treatments.

METHODS

The authors declare that all supporting data are available within the article and its [Supplemental Material](#).

Animal Model

Experiments were performed on 36 Sprague-Dawley litters each culled to n=12 (50% male) pups. Pups were kept in an oxy-cycler ProOx P110 (Biosherix, Lacona, NY) at 80% O₂ concentration (high O₂) or room air (control) from P3 to P10, as previously described.^{18,19} To avoid maternal morbidity associated with O₂-toxicity, mothers from the room air and high O₂-exposed groups were interchanged every 12 hours, which was previously shown to not affect pup growth or blood pressure levels in this model.¹⁹ Each litter was divided into 3 experimental treatment groups: vehicle cyclodextrin (30 μg/kg per day), cyclodextrin–Angio-(1–7) or cyclodextrin–alamandine oral formulation (inclusion compounds: 74 μg/kg per day [30 μg/kg per day of cyclodextrin+44 μg/kg per day of included peptides])²⁵ from P3 to P10, administered daily by gavage. Different litters were used to assess or carry the assessments at P10 and P28. No more than 2 pups per litter were used per treatment group in each experiment. P28 was chosen to study the effect of therapies on cardiac changes as previously reported by us and before blood pressure changes and overload in this model.^{18,26} Only male rats were used because our data showed no significant cardiovascular alterations in female offspring at P28 before blood pressure changes in this model (Yzodyrczyk et al¹⁹ and [Figure S1](#)). All experimental procedures were approved by the Animal Ethics Committee of the Sainte-Justine University Hospital Research Center and followed the

guidelines of the Canadian Council on Animal Care and the National Institutes of Health Guide for the Care and Use of Laboratory Animals.

Comprehensive Analysis of the Neonatal LV Anatomy by Echocardiography and 3D Computational Models

The method to characterize in vivo LV remodeling was developed by combining echocardiography and computational techniques. It was performed on 24 neonatal rats at P10, immediately after O₂ exposure and treatments ([Figure S2A](#)). To obtain 3D images, VEVO3100 ultrasound system (VisualSonics) and a 30 MHz probe were used. 3D ventricular volumes were recorded using the 3D feature of VEVO3100 system and automated mechanical arm ([Figure S2B](#)) by recording ≈100 sequential LV images in the short-axis view, with a distance of ≈0.1 mm between each image. Segmentation of the LV myocardium was then performed semiautomatically using the Medical Imaging Interaction Toolkit software (<http://www.mitk.org/wiki/MITK>). 3D meshes using cubic interpolation basis functions were then fitted to the segmentation masks using image registration and an idealized LV template, adopting the methods described by Lamata et al.²⁷ Meshes were then aligned by their center of mass and orientation, and an average anatomy was obtained, as illustrated in the atlas presented in [Figure S2C through S2D](#). A principal component analysis (PCA) was then used to analyze the differences of meshes to the average, identifying the 23 linear PCA modes of geometric variation. The amount of variance explained by the 2 main modes and the cumulative variance explained by groups of modes are shown in [Figure S2E](#). In addition, [Figure S2F](#) illustrates the 2 first modes of variation that explained 33.64% of the variance observed in all cases. As a result of this analysis, the LV anatomy of each of the 24 cases is described by 2 coefficients corresponding to the first 2 PCA modes. Each mode was compared between experimental groups to detect shape changes caused by O₂-exposure and drug treatments. In addition, conventional LV bidimensional short-axis M-mode and mitral valve flow Doppler imaging, myocardium strain, and Tissue Doppler analyses were performed. LV mass was calculated by $LV\ mass = 1.053 \times ([LV\ internal\ diameter\ in\ diastole + LV\ posterior\ wall\ thickness\ in\ diastole + intraventricular\ septum\ thickness\ in\ diastole]^3 - LV\ internal\ diameter\ in\ diastole^3)$.

Additional Experimental and Molecular Procedures

We additionally performed echocardiography in juvenile rats at P28 (see [Supplemental Material](#)). Hearts and kidneys were collected after euthanasia immediately following echocardiography imaging. Hearts were washed in 100 mmol/L potassium chloride solution to induce diastolic arrest and then weighed. They were further sectioned and the base of the hearts (including the valves) were immersion-fixed in 4% paraformaldehyde for paraffin embedding and histomorphometry analysis while the apex was snap-frozen in liquid nitrogen and later processed for molecular biology (Western blotting and reverse transcription quantitative polymerase chain reaction of RAS components). Kidneys were collected at P10 and immersion-fixed in 4% paraformaldehyde, cut in half (long-axis sagittal cut), and

embedded in paraffin for histomorphometry. More details about the histological and molecular biology methods used in this study are provided in the [Supplemental Material](#).

Statistical Analysis

We used a total number of 36 litters, 18 per study age (P10 and P28). A sample size of $n=6$ rats per group was used for all statistical analyses based on previous works from our laboratory using this rat model.^{18,20} Experimental groups were compared by 2-way ANOVA followed by Bonferroni posthoc test, having oxygen exposure (high O_2 versus room air) and treatments (vehicle cyclodextrin, cyclodextrin–Angio-(1–7), and cyclodextrin–alamandine) as factors. Growth curve was analyzed using 2-way ANOVA for repeated measures. In the 3D computational anatomy study, PCA were used to identify the anatomic models of variation, and differences in shape descriptors (ie, PCA modes) were studied with an unpaired t test. The software GraphPad Prism version 5.00 for Windows (GraphPad Software, San Diego, CA) was used for all tests performed. The significance level was established at $P<0.05$.

RESULTS

Assessment of Potential Adverse Effects of Neonatal Treatments on Body and Organ Weights and Renal Morphology

Treatments with cyclodextrin–Angio-(1–7) or cyclodextrin–alamandine did not significantly change the weight of the pups in high O_2 -exposed or room air groups throughout time in comparison to vehicle cyclodextrin (Figure S3). We also assessed potential adverse effects of treatments on the renal morphology of neonatal rats. Renal cortex width and glomerular diameter were not significantly changed by treatments in rats kept in room air (Figure S4). The only effect observed was a significant increase in renal cortex width in high O_2 -exposed rats treated with cyclodextrin–Angio-(1–7) versus cyclodextrin–alamandine without reaching statistical significance when compared to vehicle cyclodextrin controls, indicating a possible positive side effect of this drug in exposed rats.

Ventricular Geometric Changes in Neonatal Rats

We first examined the effect of high O_2 exposure and treatments with cyclodextrin–Angio-(1–7) or cyclodextrin–alamandine on ventricular geometric characteristics (shape and size) of neonatal rats at P10, that is, immediately after high- O_2 exposure and treatments. High O_2 exposure was linked to smaller and shorter ventricles with a globular pattern; the characteristics encoded in mode 1 (significant differences between room air and high- O_2 conditions, see Figure 1A). However, the analyses of all rats (room air and high O_2 -exposed) treated either with vehicle cyclodextrin or any inclusion compounds indicated

that treatments triggered significant differences in mode 2 (Figure 1B), leading to smaller and shorter ventricles with a subtle shift of the apex towards the septum (opposite to the free lateral wall).

The average ventricular anatomy of the six experimental groups identified structural alterations compatible with mode 2 induced by cyclodextrin–Angio-(1–7) and cyclodextrin–alamandine treatments in both groups exposed to room air and high O_2 , although they were more pronounced in rats kept at room air than in rats exposed to high O_2 (Figure 1C). In high O_2 -exposed rats, cyclodextrin–Angio-(1–7) treatment induced a more significant difference in mode 2 compared to cyclodextrin–alamandine and vehicle cyclodextrin, indicating an even smaller but slightly less globular LV shape, with reduced ventricular chamber diameter in high O_2 -exposed rats treated with cyclodextrin–Angio-(1–7).

Ventricular Functional Changes in Neonatal Rats

LV functional changes were assessed in neonatal rats at P10 by echocardiography using bidimensional M-mode short-axis and tissue Doppler images. Neonatal echocardiographic findings are shown in Figure 2 and detailed in Table S1. No significant difference was observed between groups on heart rate, intraventricular septum thickness in diastole (intraventricular septum thickness in diastole), LV blood volume in systole, or LV stroke volume. High O_2 exposure reduced LV posterior wall thickness in diastole and the LV mass in neonatal rats (Table S1). However, LV mass when indexed to body weight was no longer statistically different between groups.

Cyclodextrin–Angio-(1–7) and cyclodextrin–alamandine treatments did not induce significant changes in LV function in neonatal rats kept in room air. However, in high O_2 -exposed rats, cyclodextrin–Angio-(1–7) treatment triggered more pronounced positive LV functional changes in comparison to cyclodextrin–alamandine. High O_2 -exposed rats treated with cyclodextrin–Angio-(1–7) had reduced LV internal chamber diameters (LV internal diameter in diastole, Figure 2A) and blood volume in diastole (Figure 2B), which are features compatible with the smaller and less globular shape identified in the 3D geometric analyses. Furthermore, cyclodextrin–Angio-(1–7) prevented reduced LV posterior wall thickness in diastole in high O_2 -exposed rats (Figure 2C), although it significantly reduced LV mass in these rats in comparison to rats kept in room air treated with vehicle cyclodextrin (Table S1). However, LV mass when indexed to body weight was no longer different between groups.

Cyclodextrin–Angio-(1–7) treatment markedly improved LV systolic function in neonatal high O_2 -exposed rats. This improvement was indicated by an average 10% higher fraction of shortening (Figure 2D) and a significant increment of radial endocardial strain

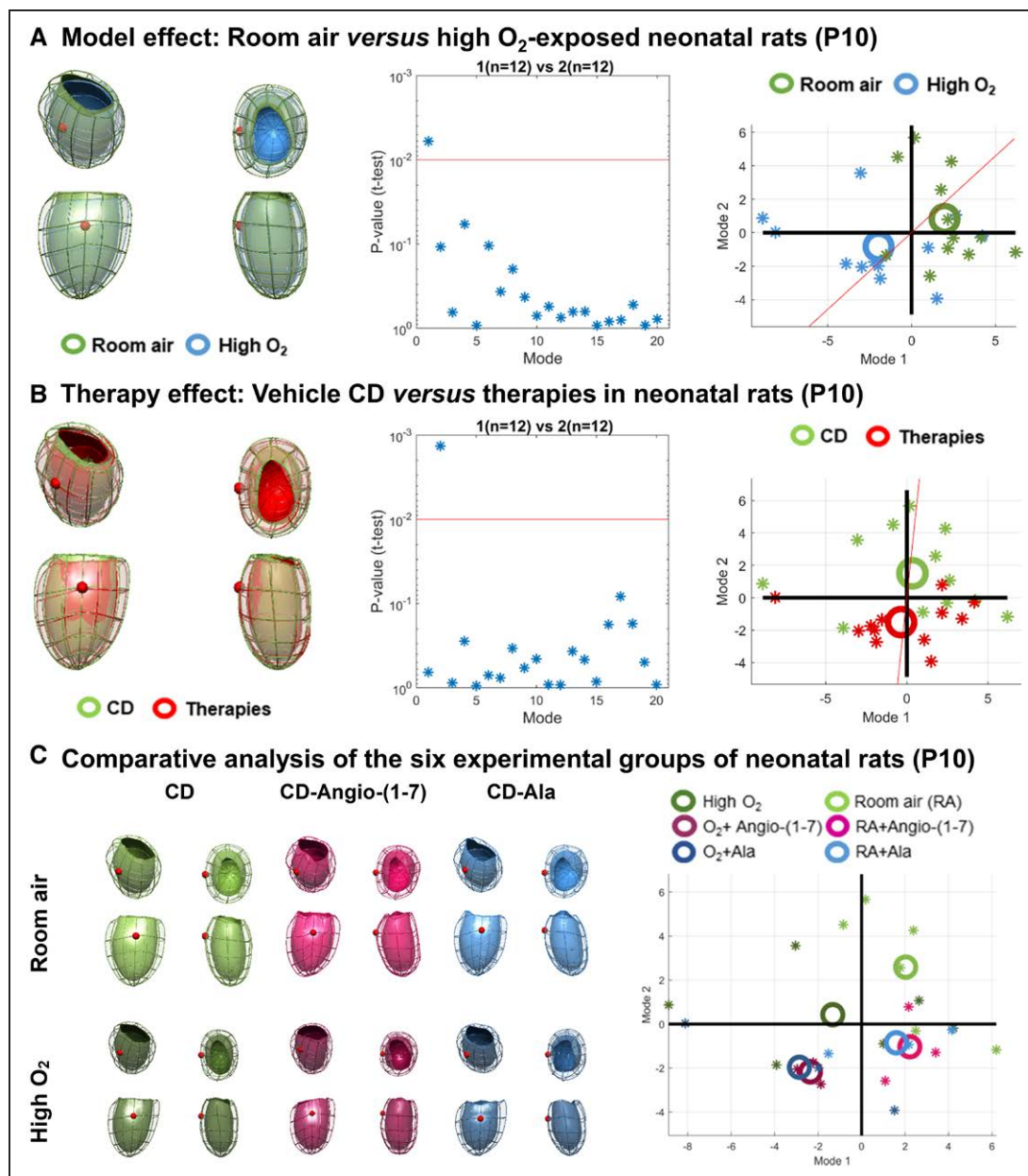
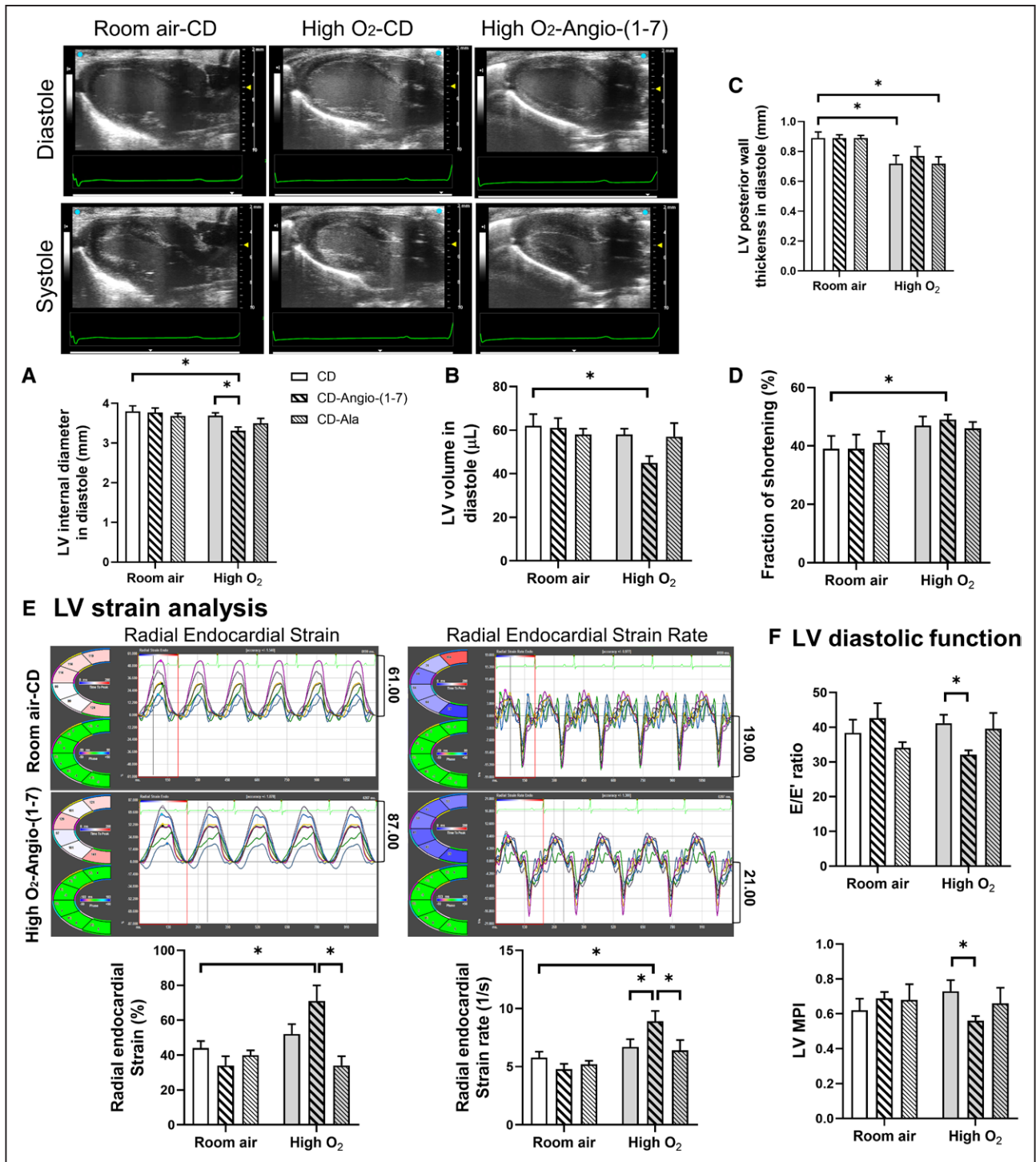


Figure 1. Computational geometric ventricular analyses of neonatal rats (P10).

A, Analysis of the effect of high-oxygen (O₂) stress exposure on the anatomy of neonatal rats kept at room air (RA, green) vs high O₂-exposed (blue) at P10: illustration of the average left ventricular anatomy of each group, statistical comparison of principal component analysis (PCA) modes and a graph indicating individual values (stars) and averages (large circles) per group comparing PCA mode 1 and 2 (n=12 per exposure group). **B**, Analyses of the effect of the treatments, comparing neonatal rats at P10 kept at room air and high O₂-exposed that were either treated with vehicle cyclodextrin (CD) or with any inclusion compounds (n=12 per exposure group). **C**, Average ventricular anatomy of the six experimental groups and graph with individual values (stars) and averages per group (circles) comparing PCA mode 1 and 2 (n=3 per treatment per exposure group). Red sphere in 3-dimensional meshes points in the direction of the left ventricle lateral wall. Ala indicates alamandine; and Angio, angiotensin.

in this group in comparison to neonatal rats kept in room air treated with vehicle cyclodextrin (Figure 2E). Similarly, the radial endocardial strain rate was also markedly increased in high O₂-exposed rats treated with cyclodextrin–Angio-(1–7) versus room air and high O₂-exposed rats treated with vehicle cyclodextrin (Figure 2E), supporting enhanced myocardial contractility in cyclodextrin–Angio-(1–7) treated group. We also observed

improvements in LV diastolic function (Figure 2F) in high O₂-exposed rats treated with cyclodextrin–Angio-(1–7). The significant reduction in LV mitral valve Doppler E wave/ tissue Doppler E' wave ratio (E/E') ratio in this group indicates a better balance between ventricular relaxation and filling pressures (Figure 2F). This is corroborated by a significantly lower LV myocardial performance index (MPI) in this group (Figure 2F), indicating



enhanced global systolic and diastolic functions in high O_2 -exposed rats treated with cyclodextrin–Angio-(1–7) versus vehicle cyclodextrin.

Cyclodextrin–alamandine treatment did not induce any significant LV functional changes in neonatal rats with features very similar to rats treated with vehicle cyclodextrin in both exposure groups (Table S1).

Histomorphological Ventricular Changes in Neonatal Rats

At P10, high O_2 exposure and drug treatments did not induce significant changes in body weight in comparison to rats kept at room air and treated with vehicle cyclodextrin (Figure 3A). The only statistically significant difference observed between groups was a lower weight in pups exposed to high O_2 treated with cyclodextrin–Angio-(1–7) versus pups kept in room air receiving the same treatment. Cardiac index, calculated by the ratio of the LV weight (extracted postmortem) to body weight, was not significantly different between groups (Figure 3B).

Histological analyses of P10 hearts showed increased average LV cardiomyocyte surface area as an indication of hypertrophy in high O_2 -exposed rats versus room air rats treated with vehicle cyclodextrin (Figure 3C and 3D). However, this effect was removed by treating high O_2 -exposed rats with cyclodextrin–Angio-(1–7) or cyclodextrin–alamandine. In rats kept at room air, both cyclodextrin–Angio-(1–7) treatment and cyclodextrin–alamandine did not significantly change cardiomyocyte surface area in comparison to rats treated with vehicle cyclodextrin.

Long-Term Effects of Neonatal Treatments on Ventricular Function and Histomorphologic Changes

We examined whether neonatal treatment with cyclodextrin–Angio-(1–7) and cyclodextrin–alamandine could induce long-term positive effects on LV function and remodeling assessed in juvenile rats at P28. LV functional changes were assessed in juvenile rats at P28 by echocardiography using a similar protocol as for P10 with bidimensional M-mode short-axis and tissue Doppler images to obtain wall thickness and chamber diameter measurements and to calculate the fraction of shortening (Table S2 and Figure 4). We have previously shown that high O_2 -exposed rats can develop impaired systolic function at this age, mainly characterized by reduced fraction of shortening with increased myocardium hypertrophy and fibrosis.^{18,20} No significant difference was observed for body weight (Figure 4A) or LV posterior wall thickness in diastole (Figure 4B) between groups. Furthermore, no significant difference was observed between juvenile rats previously exposed to room air and high O_2 on left ventricle internal diameter in systole (Figure 4C). Cyclodextrin–alamandine

treatment, however, significantly reduced this parameter in room air rats and cyclodextrin–Angio-(1–7) in high O_2 -exposed rats in comparison to rats previously kept at room air treated with vehicle cyclodextrin. Fraction of shortening was significantly reduced in juvenile rats previously exposed to high O_2 versus room air, which was prevented by neonatal treatments with cyclodextrin–Angio-(1–7) and cyclodextrin–alamandine treatments (Figure 4D).

LV cardiomyocyte surface area was significantly higher in juvenile rats previously exposed to high O_2 versus room air, indicative of cardiomyocyte hypertrophy (Figure 4E). However, this effect was completely prevented by neonatal treatment with cyclodextrin–Angio-(1–7). In juvenile rats previously kept at room air, only alamandine treatment significantly reduced cardiomyocyte surface area. Both neonatal treatments had also markedly reduced LV fibrosis in juvenile rats previously exposed to high O_2 without significant effects in rats kept in room air (Figure 4F).

Cardiac RAS Changes in Neonatal Rats

We assessed changes in cardiac RAS profile at P10 in neonatal rats treated with cyclodextrin–Angio-(1–7) or cyclodextrin–alamandine (Figure 5). Figure 5A describes differences in *ACE*, *AT1a*, *AT1b*, and *AT2* mRNA expression in ventricles. *ACE* mRNA expression was significantly higher in high O_2 -exposed rats treated with vehicle cyclodextrin at P10, which was prevented by both treatments (Figure 5A). *AT1a* mRNA expression was similarly higher in high O_2 -exposed neonatal rats treated with vehicle cyclodextrin (Figure 5A). This was prevented only by cyclodextrin–alamandine treatment. However, in rats kept in room air, cyclodextrin–alamandine treatment increased cardiac *AT1a* mRNA expression (Figure 5A). This paradoxical response of cyclodextrin–alamandine between rats kept at room air and high O_2 suggests that this peptide may indirectly regulate the transcription of *AT1a* gene in ventricles. High O_2 exposure also increased the mRNA expression of *AT1b* (Figure 5A) and *AT2* (Figure 5A), and no treatment significantly affected their transcription in high O_2 -exposed rats. However, cyclodextrin–Angio-(1–7) and cyclodextrin–alamandine treatments increased *AT1b* mRNA expression in ventricles of neonatal rats kept in room air (Figure 5A), also suggesting an indirect effect of these peptides on *AT1b* gene regulation.

Figure 5B shows results of *ACE2* and Mas receptor protein and mRNA expressions. High O_2 exposure did not change cardiac *ACE2* mRNA expression in neonatal rats treated with vehicle cyclodextrin but significantly reduced *ACE2* protein expression (Figure 5B). Cyclodextrin–Angio-(1–7) and cyclodextrin–alamandine treatments markedly increased *ACE2* mRNA expression but not the protein expression in ventricles of neonatal rats exposed to high O_2 (Figures 5B). The mRNA expression of Mas receptor was significantly increased in neonatal rats

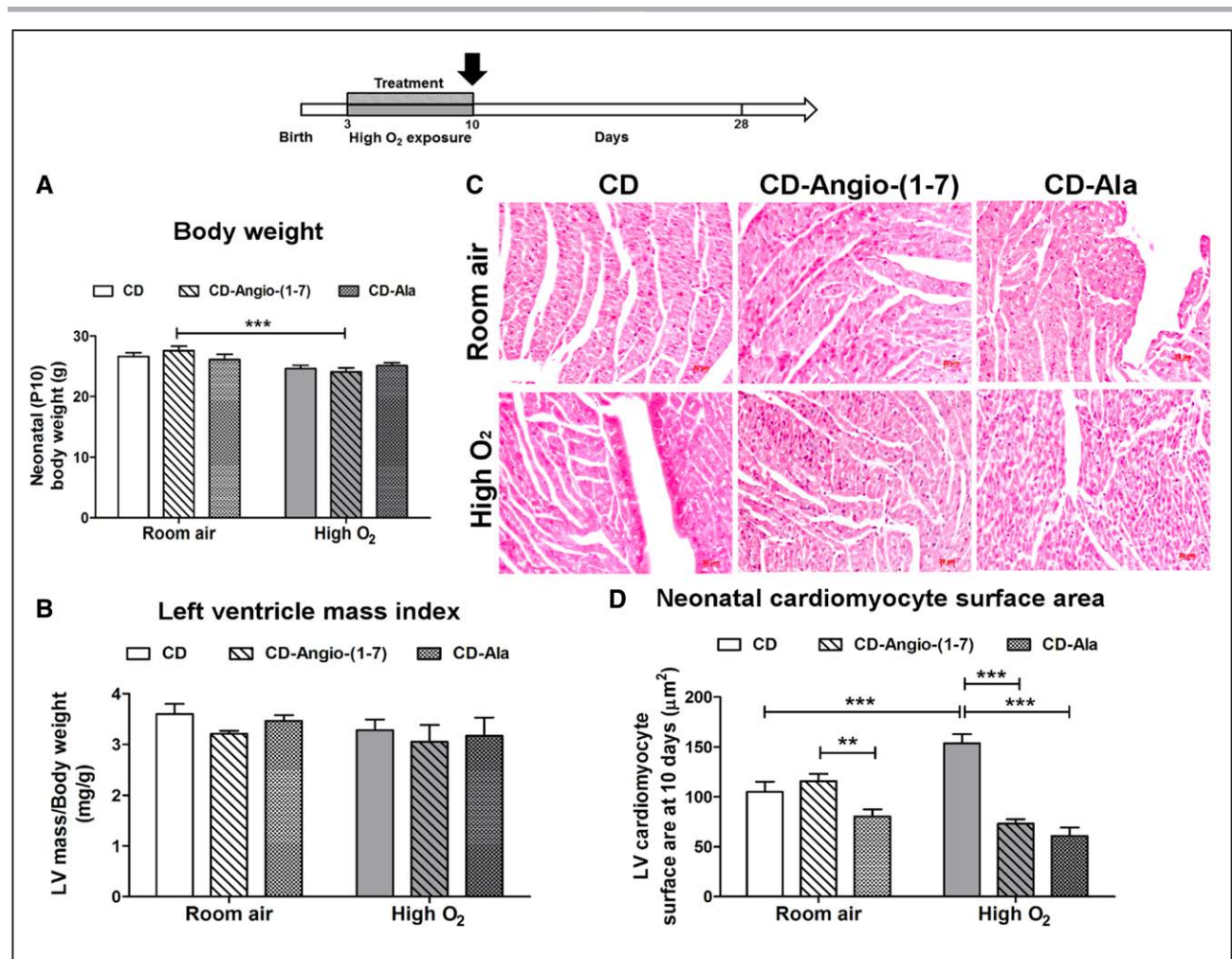


Figure 3. Histomorphometry analyses of neonatal hearts (P10).

Study design showing hearts collected at day 10 of life (P10) after 7 d of oxygen exposure and treatment interventions in neonatal rats. **A**, Bodyweight and **(B)** left ventricular (LV) mass per body weight index of neonatal rats at P10. **C**, Representative histological images and **(D)** quantification of LV cardiomyocytes surface area of the hearts of neonatal rats ($n=6$ /group) exposed to high-O₂ or kept at room air and treated with vehicle cyclodextrin (CD) or included CD-Angio (angiotensin)-(1-7) or CD-alamandine (Ala). Data presented as mean \pm SEM. Two-way ANOVA followed by Bonferroni posthoc test. * $P<0.05$, ** $P<0.01$ and *** $P<0.001$.

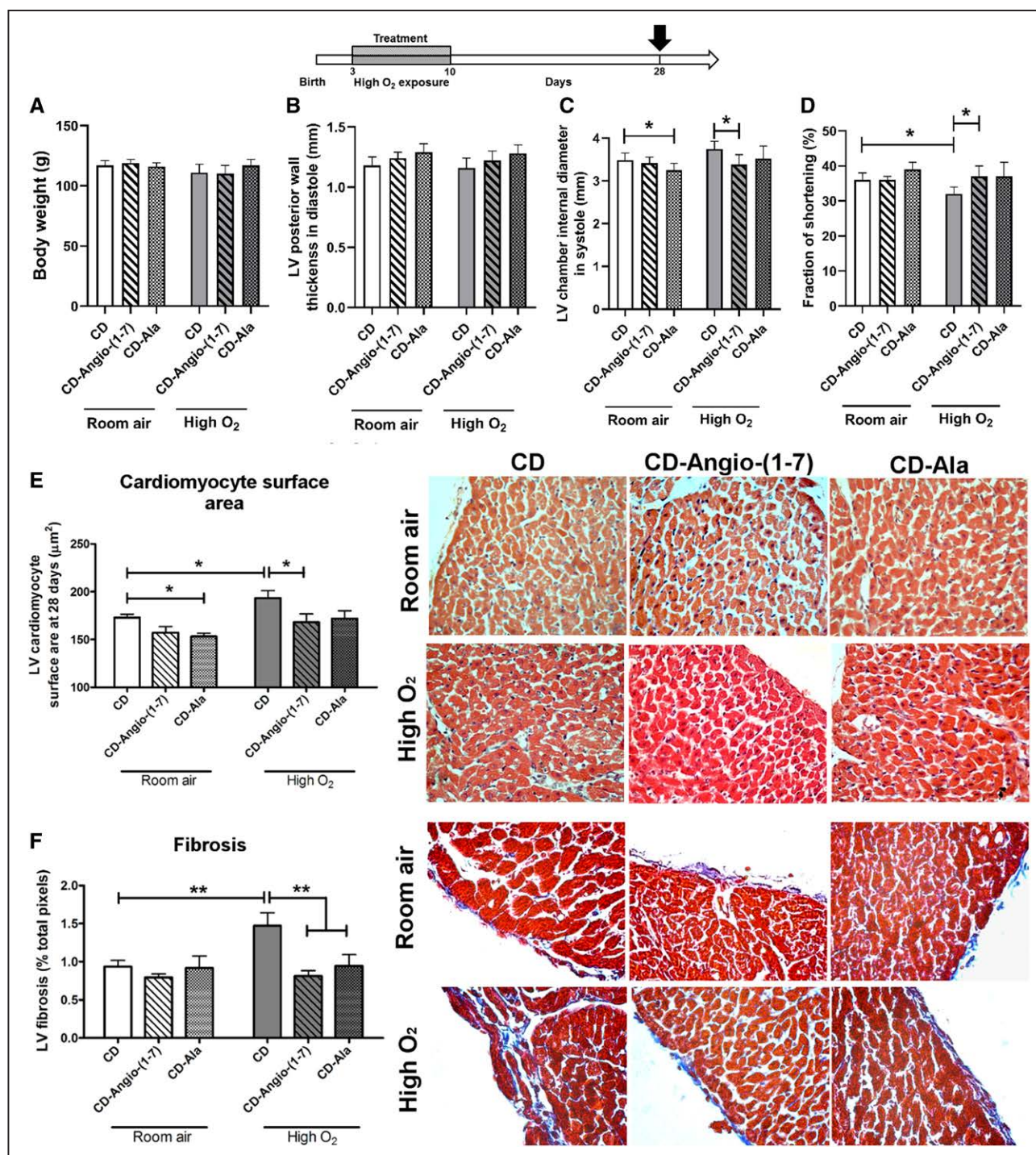
exposed to high O₂ compared to room air, while its protein expression was not different (Figure 5B). Only cyclodextrin–Angio-(1-7) treatment significantly increased the protein expression of Mas receptor in high O₂-exposed rats (Figure 5B). A summary of our findings in ventricles of neonatal rats is shown in Figure 5C and 5D. Briefly, they indicate an inverse regulation of ACE and ACE2, prevailing the upregulation of ACE2, by cyclodextrin–Angio-(1-7) and cyclodextrin–alamandine treatments in neonatal rats immediately after high O₂ exposure without significant downregulation of AT1 receptor genes.

Long-Term Changes in Cardiac RAS Induced by Neonatal Treatments

We examined the same RAS components in the LV of juvenile rats at P28, previously exposed to high O₂ or kept at room air and treated with cyclodextrin–Angio-(1-7) or cyclodextrin–alamandine from P3 to P10 (Figure 6). No

difference was observed in ACE mRNA expression between groups (Figure 6A). Cyclodextrin–Angio-(1-7) and cyclodextrin–alamandine treatments significantly increased AT1a mRNA expression compared to vehicle cyclodextrin in rats kept at room air, whereas only alamandine treatment increased it in the LV of rats previously exposed to high O₂ (Figure 6A). AT1b mRNA expression was increased in LV of juvenile rats exposed to high O₂ but prevented only by cyclodextrin–Angio-(1-7) treatment (Figure 6A). AT2 mRNA expression was significantly lower in LV of juvenile rats previously exposed to high O₂ and both treatments did not change this low transcription in juvenile rats (Figure 6A). However, in rats previously kept at room air, only early cyclodextrin–Angio-(1-7) treatment significantly increased long-term cardiac AT2 mRNA expression in comparison to rats treated with vehicle cyclodextrin (Figure 6A).

Juvenile rats previously exposed to high O₂ did not have ACE2 mRNA expression changes in LV but had lower protein expression of ACE2 (Figure 6B). Both



cyclodextrin-Angio-(1-7) and cyclodextrin-alamandine treatments significantly increased *ACE2* mRNA and protein expressions in LV in juvenile rats previously exposed to high O₂. No significant change was observed in Mas

receptor mRNA and protein expressions in ventricles of juvenile rats previously exposed to high O₂ (Figure 6B). However, cyclodextrin-Angio-(1-7) treatment markedly increased long-term mRNA and protein expressions of

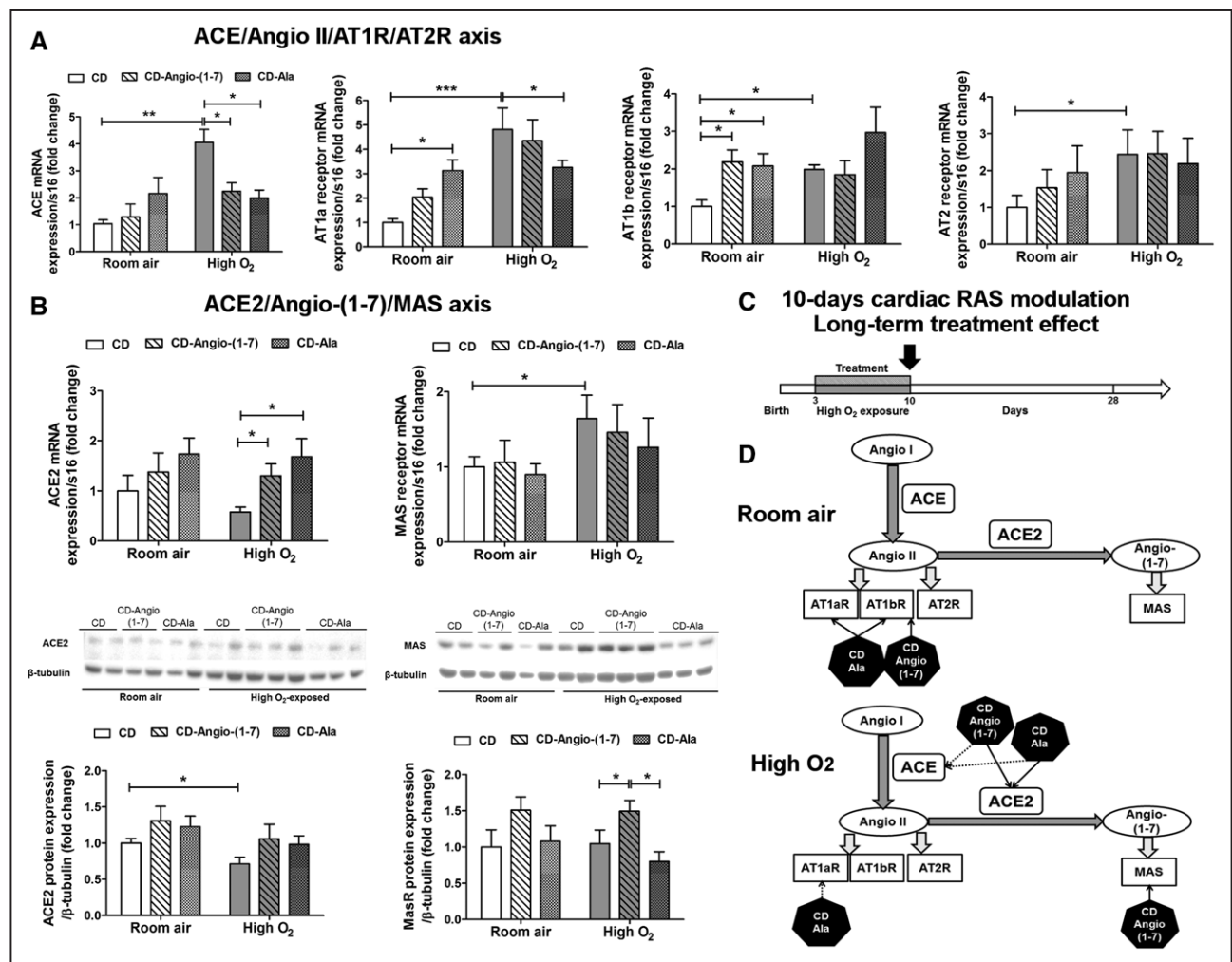


Figure 5. Cardiac renin-angiotensin system (RAS) changes in neonatal rats (P10).

A, mRNA expression of components of the ACE (angiotensin-converting enzyme)/Angio II (angiotensin II)/AT1a (angiotensin receptor type 1a), AT1b and AT2 (angiotensin receptor type 2) receptors axis, and **(B)** mRNA and protein expressions of ACE2 and Mas receptor in left ventricle (LV) of neonatal rats at P10 ($n=6/\text{group}$) exposed to high O₂ or at room air and treated with vehicle cyclodextrin (CD) or included CD-Angio-(1-7) or CD-alamandine (Ala). Data presented as mean \pm SEM. Two-way ANOVA followed by Bonferroni posthoc test. * $P<0.05$, ** $P<0.01$ and *** $P<0.001$. **C**, Study design indicating that hearts were collected at P10. **D**, Summary of cardiac RAS findings indicating which treatment (black hexagons) has stimulated (arrow with full line) or inhibited (arrow with dotted line) RAS components in neonatal rats kept at room air or high O₂. AT1R indicates angiotensin II receptor type 1; AT2R, angiotensin II receptor type 2; AT1aR, angiotensin II type 1 receptor a; AT1bR, angiotensin II type 1 receptor b; MAS, Mas receptor protein; and MasR, Mas receptor gene.

Mas receptor in ventricles of juvenile rats kept at room air but not in high O₂-exposed rats, whereas cyclodextrin-alamandine had no significant effect (Figures 6B). A summary of the main findings in ventricles of juvenile rats at P28 is shown in Figure 6C and 6D. Briefly, they indicate a long-term upregulation of ACE2 in ventricles of juvenile rats previously exposed to high O₂ and treated with either cyclodextrin-Angio-(1-7) or cyclodextrin-alamandine.

DISCUSSION

In this study, we showed that treatment with cyclodextrin-Angio-(1-7) and cyclodextrin-alamandine were effective at preventing pathological cardiac remodeling in juvenile rats exposed to high-O₂ during the neonatal period. Of

the 2 treatments, cyclodextrin-Angio-(1-7) presented the most significant positive effects on LV remodeling and function in neonatal rats exposed to high O₂. In addition, although cyclodextrin-Angio-(1-7) induced significant changes in ventricular geometry, remodeling, and function, cyclodextrin-alamandine effectiveness was limited to the reduction of LV myocardium hypertrophy in neonatal rats and in overall remodeling in juvenile rats exposed to high O₂. Importantly, no treatment significantly downregulated or blocked Angio II AT1 receptor expression, whereas both cyclodextrin-Angio-(1-7) and cyclodextrin-alamandine reduced ACE gene expression to values similar to controls. In high-O₂ animals, ACE2 was similarly stimulated by both treatments through gene upregulation in neonatal and juvenile rats.

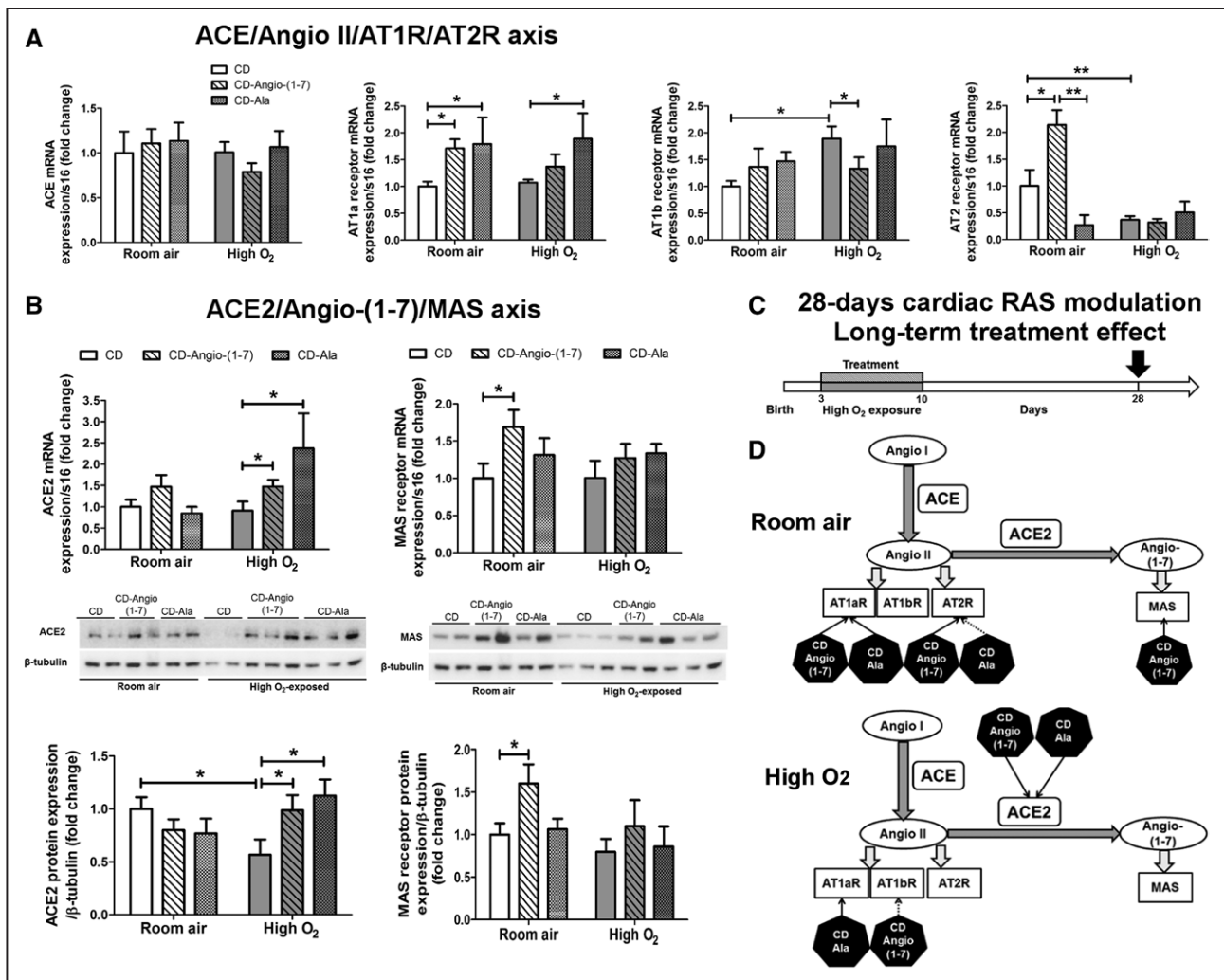


Figure 6. Cardiac renin-angiotensin system (RAS) changes in juvenile rats (P28).

A, mRNA expression of components of the ACE (angiotensin-converting enzyme)/Angio II (angiotensin II)/AT1a (angiotensin receptor type 1a), AT1b (angiotensin receptor type 1b), and AT2 (angiotensin receptor type 2) receptors axis, and **(B)** mRNA and protein expressions of ACE2 and Mas receptor in left ventricle (LV) of juvenile rats at P28 ($n=6/\text{group}$) exposed to high O₂ or at room air and treated with vehicle cyclodextrin (CD) or included CD-Angio-(1-7) or CD-alamandine (Ala) from P3 to P10. Data presented as mean \pm SEM. Two-way ANOVA followed by Bonferroni posthoc test. * $P<0.05$ and ** $P<0.01$. **C**, Study design indicating that hearts were collected at P28. **D**, Summary of cardiac RAS findings indicating which treatment (black hexagons) has stimulated (arrow with full line) or inhibited (arrow with dotted line) RAS components in juvenile rats kept at room air or high O₂ from P3 to P10. AT1R indicates angiotensin II receptor type 1; AT2R, angiotensin II receptor type 2; AT1aR, angiotensin II type 1 receptor a; AT1bR, angiotensin II type 1 receptor b; and MAS, Mas receptor protein.

The 3D computational cardiac images revealed altered ventricular shape and remodeling at neonatal age (P10) in rats exposed to high-O₂, mainly characterized by small and short ventricles with a more globular pattern. Histomorphometry further indicated that this pattern was associated with cardiomyocyte hypertrophy in high O₂-exposed rats, which may represent a myocardial compensatory adaptation to maintain the cardiac output in such small hearts. Still, LV function was preserved at neonatal age in exposed rats. These changes in ventricular shape and remodeling were also shown in our study to predispose the developmental programming of cardiomyopathy in O₂-exposed rats when reaching juvenile age. Interestingly, both treatments failed to rescue the smaller and shorter pattern of ventricles in O₂-exposed

rats; however, cyclodextrin-Angio-(1-7) has more effectively reshaped it to a slightly smaller aspect with a consequent reduction of ventricular diastolic volume. Compensatory improvements for systolic and diastolic ventricular function and myocardium contractility were observed in neonatal rats exposed to high O₂ treated with vehicle cyclodextrin and the inclusion compounds, probably due to their lower ventricular volumes. However, LV functional improvements were more remarkable and not associated with cardiomyocyte hypertrophy only in neonatal rats treated with cyclodextrin-Angio-(1-7). Cyclodextrin-alamandine, however, did not induce changes in ventricular shape and function as observed with cyclodextrin-Angio-(1-7) treatment, maintaining instead a pattern and functional parameters that were very close to

O₂-exposed rats treated with vehicle. However, cyclodextrin-alamandine still effectively reduced cardiomyocyte surface area and hypertrophy in high O₂-exposed neonatal rats. Although we observed subtle geometric changes in control groups treated with the inclusion compounds, they have not associated with significant LV remodeling and dysfunction.

Our ventricular geometric results are similar to clinical findings in preterm-born infants. Previously, a large study performing echocardiographic imaging in >390 preterm and full-term born newborns described smaller hearts in preterm newborns with disproportionate catch-up growth in the first 3 months of life.⁸ In this study, a similar method of computational ventricular analysis was performed from bidimensional ultrasound imaging, revealing in preterm babies a ventricular pattern consistent with a globular shape. In line with these findings, Cox et al⁵ have used cardiac magnetic resonance scans of preterm-born neonates and computational atlases to also demonstrate a ventricular globular shape with spherical blood pool in preterm-born neonates. Similar to our findings, early in life cardiac remodeling was described to persist into adulthood in preterm-born individuals. This was first reported by Lewandowski et al,²⁸ where they showed short-patterned LV associated with dysfunction and higher LV mass in young adults born prematurely compared to those born full-term. LV dysfunction was further linked to greater diffuse myocardial fibrosis in the LV of young adults born preterm, which relates to the degree of prematurity.²⁹

Hence, our rat model successfully reproduced the main cardiac features observed in preterm-born young adults that may be predisposing them to an accelerated progression towards dysfunction and disease.

Crispi et al³⁰ previously described the impact of intrauterine ventricular remodeling on perinatal and postnatal growth outcomes in small for gestational age newborns.³¹ Their findings also revealed that globular and hypertrophied ventricular patterns in fetal hearts can associate with sustained hemodynamic and ventricular diastolic dysfunction into postnatal life³² and up to preadolescence.³³ However, another fetal ventricular pattern described by them that is consistent with an elongated shape has been associated with better outcomes.³¹ These findings indicate that early compensatory ventricular patterns could lead to better outcomes in this population.

In our rat model, all O₂-exposed groups developed shorter ventricles in comparison to room air controls, indicating an activation of O₂-sensitive mechanisms capable of regulating cardiac development that was not rescued by any treatment. However, our findings indicate that the main positive responses triggered by both treatments in O₂-exposed rats were related to the inhibition of cardiomyocyte hypertrophy and ventricular fibrosis rather than rescuing the smaller and shorter

LV shape observed in high O₂-exposed rats. Therefore, our treatments targeting the cardioprotective RAS axis revealed an important effect of this system on heart plasticity during neonatal life.

The RAS is a well-recognized central mechanism regulating blood pressure through the actions of vasoactive peptides that can also control blood volume. In fetal life, RAS components such as Angio II and AT1 receptors are essential for kidney and heart formation and maturation, notably in the last gestational trimester.^{24,34–36} However, in prematurity, this system can be overactivated, which has been shown to trigger a paradoxical negative effect by stimulating an accelerated organ maturation and remodeling.²¹ Our results showed that restoring the balance of this system during development can be beneficial to prevent heart diseases later in life in our animal model. More importantly, we have not observed any adverse effects of Angio-(1–7) or alamandine treatments in neonatal rats. We have only reported a significant increase in renal cortex width in neonatal rats exposed to high O₂ treated with Angio-(1–7), which is a positive effect. A previous study from our laboratory has shown that hyperoxia exposure resulted in a significant reduction in both nephrogenic zone width and glomerular diameter at P5 in this model, and a significantly increased apoptotic cell count, without affecting nephron number at P10. We also observed that the expression of the HIF (hypoxic-inducible factor)-1 α was significantly reduced in the developing kidney following hyperoxia exposure.³⁷ In this study, a systemic administration of the HIF-1 α stabilizer dimethyloxalylglycine resulted in enhanced expression of HIF-1 α and improved nephrogenesis in hyperoxia-exposed pups mainly by increasing nephrogenic zone width and glomerular diameter. However, interactions between the RAS and particularly of Angio-(1–7), and HIF-1 α are still unclear, especially in the kidney.

In our study, both treatments were effective at preventing programming of cardiomyopathy in juvenile rats previously exposed to high O₂. Over the years, the cardioprotective effect of Angio-(1–7) has emerged as a promising therapeutic avenue for the management of several pathological outcomes.²² Angio-(1–7)-mediated antifibrogenic and antihypertensive effects have been systematically reported in different experimental models of hypertension and heart diseases and also when using a similar orally active formula to deliver this peptide.^{25,38,39} In addition, the recently discovered alamandine peptide is shown to stimulate cardioprotective responses comparable to those of Angio-(1–7) through mechanisms that also involve the reduction of cardiac fibrosis and blood pressure in a heart failure model.²² This was further confirmed by Oliveira et al⁴⁰ describing a severe programming of dilated cardiomyopathy in a mice deficient in alamandine's receptor MrgD (Mas-related G protein-coupled receptor member D).

A common positive effect of both treatments observed in our study was the upregulation of cardiac ACE2 in O₂-exposed rats. Although our rats were only treated during the neonatal period, this effect was shown as long-lasting and associated with better ventricular remodeling and function in rats into adulthood. ACE2 is a well-known mechanism shifting the RAS deleterious actions towards its cardioprotective axis, prevailing Angio-(1–7) actions rather than Angio II.^{41,42} ACE2 has also been considered a potential target molecule for therapies because of its extensive distribution within the cardiovascular system. By cleaving Angio II and forming Angio-(1–7), ACE2 further bypasses the activation of AT1 receptors.^{41,42} This was previously shown by others when treating ACE2 knock-out mice infused with Angio II with an exogenous human recombinant ACE2.⁴³ It stimulated a dramatic decrease on plasmatic and myocardial Angio II levels while increasing Angio-(1–7) in treated mice. In our study, both treatments have sustained a balance in favor of ACE2. Acutely, they have mainly stimulated ACE2 gene expression, whereas in the long-term, both therapies caused a sustained gene and protein ACE2 overexpression in O₂-exposed rats, indicating potential epigenetic modifications targeting this gene induced by early treatment with both peptides. However, the molecular or genetic mechanisms up-regulating ACE2 triggered by these treatments are still unclear.

Novel downstream players involved in the biological differences and similarities between Angio-(1–7) and alamandine are not fully understood. This is a fast-advancing field of research that faces additional challenges in newborns when RAS plays an essential but also dueling role under stress. The positive effects of cyclodextrin–Angio-(1–7) and cyclodextrin–alamandine treatments observed in our animal model of neonatal O₂-stress represent an important and novel therapeutic avenue to prevent the programming of diseases in prematurity.

Strengths and Limitations

This is the first study describing the direct impact of environmental stress such as high-O₂ exposure and treatments on neonatal heart remodeling, at a time when heart development is still ongoing in rats. The description of different ventricular shape patterns was possible due to the creation of a 3D computational method using high-frequency ultrasound imaging of neonatal rats at P10. The 3D model was further complemented by detailed ventricular histomorphometry and functional assessments. Previous limitations to assess neonatal ventricular shape and function in rodents were probably due to their small ventricle size (≈8 mm) and accelerated heart rate, which can quickly and significantly vary in response to environmental factors such as anesthesia and temperature changes, for example. Therefore, image acquisition requires a fast recording and experienced personnel. When using a high-frequency ultrasound probe in neonatal rats, these problems are compensated

by anatomic characteristics such as thin skin and lower thoracic bone density, which significantly facilitated the 3D screening of neonatal ventricles. These anatomic characteristics were also crucial in our study to warranty the quality of our images and contours during the segmentation process. Still, the quality of some images obtained from the ventricular base (area close to the valves) were compromised by the sternum superposition. This issue also made recording of 3D ventricular images in juvenile rats in our study unfeasible, mainly due to their higher bone density in the thoracic cage. Nevertheless, our main goal in this study was to record neonatal images and to associate ventricular patterns with later-in-life adaptive or nonadaptive cardiac function into adulthood. Therefore, in juvenile rats, we performed the same echocardiographic method using the same equipment as in our previous studies, in which we described for the first time the programming of cardiomyopathy induced by neonatal exposure to high O₂.^{18,20}

Another important strength of our study was the use of an orally active formulation of cyclodextrin–Angio-(1–7) and cyclodextrin–alamandine included in β-cyclodextrins to treat neonatal rats. The structure of the inclusion compound protects the internal peptide against proteolytic enzymes present in the stomach and intestine.⁴⁴ In addition, cyclodextrins are enzymatically degraded by bacterial enzymes at the distal portions of the digestive tract, releasing the peptide that is mainly absorbed in the colon. This therapeutic strategy could therefore more closely mimic the clinical scenario for nasogastric tube feeding and treatment of preterm newborns.

An important limitation of this study was the use of only male rats in our analyses. This was justified by the fact that female rats exposed to neonatal high O₂ stress do not present significant cardiac remodeling and functional changes before the increment of blood pressure (data provided in [Supplemental Material](#)), which happens only after 7 weeks of age in this model.¹⁹ Blood pressure changes may, therefore, be an important factor triggering decompensation with heart changes and dysfunction in female rats in this model. Further studies from our group are needed to elucidate sex-based differences in the developmental programming of cardiac diseases using this model.

Perspectives

Our findings have used a newly developed method for 3D geometric ventricular analyses to investigate ventricular shape differences and cardiac remodeling induced by neonatal high O₂ stress and treatments targeting the cardioprotective RAS axis in rats. The observed different ventricular remodeling patterns may indicate a higher predisposition of hyperoxia-exposed neonatal rats to develop cardiomyopathy into adulthood, which could be modified or prevented by cyclodextrin–alamandine and more significantly by cyclodextrin–Angio-(1–7) treatments. In both treatments, the upregulation of cardiac ACE2 could also

be key to balancing RAS and preventing the deleterious effects of Angio II/AT1 on heart remodeling and programming of cardiomyopathy in this model of neonatal stress.

Taken together, our findings describe an important cardiac plasticity in neonatal rats in response to stressful conditions and therapies. Furthermore, we have also shown that the neonatal period may be an important intervention window to effectively prevent ventricular remodeling and the programming of cardiomyopathy in preterm-born subjects.

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Disclosures

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REFERENCES

- Blencowe H, Cousens S, Oestergaard MZ, Chou D, Moller AB, Narwal R, Adler A, Vera Garcia C, Rohde S, Say L, et al. National, regional, and worldwide estimates of preterm birth rates in the year 2010 with time trends since 1990 for selected countries: a systematic analysis and implications. *Lancet*. 2012;379:2162–2172. doi: 10.1016/S0140-6736(12)60820-4
- Carr H, Cnattingius S, Granath F, Ludvigsson JF, Edstedt Bonamy AK. Preterm birth and risk of heart failure up to early adulthood. *J Am Coll Cardiol*. 2017;69:2634–2642. doi: 10.1016/j.jacc.2017.03.572
- Hovi P, Vohr B, Ment LR, Doyle LW, McGarvey L, Morrison KM, Evensen KA, van der Pal S, Grunau RE, Brubakk AM, et al; APIC Adults Born Preterm International Collaboration. Blood pressure in young adults born at very low birth weight: adults born preterm international collaboration. *Hypertension*. 2016;68:880–887. doi: 10.1161/HYPERTENSIONAHA.116.08167
- Crump C, Howell EA, Stroustrup A, McLaughlin MA, Sundquist J and Sundquist K. Association of preterm birth with risk of ischemic heart disease in adulthood. *JAMA Pediatr*. 2019;173:736–743. doi: 10.1001/jamapediatrics.2019.1327
- Cox DJ, Bai W, Price AN, Edwards AD, Rueckert D, Groves AM. Ventricular remodeling in preterm infants: computational cardiac magnetic resonance atlas shows significant early remodeling of the left ventricle. *Pediatr Res*. 2019;85:807–815. doi: 10.1038/s41390-018-0171-0
- Edstedt Bonamy AK, Mohlkert LA, Hallberg J, Liuba P, Fellman V, Domellöf M, Norman M. Blood pressure in 6-year-old children born extremely preterm. *J Am Heart Assoc*. 2017;6:e005858. doi: 10.1161/JAHA.117.005858
- Mohlkert LA, Hallberg J, Broberg O, Rydberg A, Halvorsen CP, Liuba P, Fellman V, Domellöf M, Sjöberg G, Norman M. The preterm heart in childhood: left ventricular structure, geometry, and function assessed by echocardiography in 6-year-old survivors of periviable births. *J Am Heart Assoc*. 2018;7:e007742. doi: 10.1161/JAHA.117.007742
- Aye CYL, Lewandowski AJ, Lamata P, Upton R, Davis E, Ohuma EO, Kenworthy Y, Boardman H, Wopperer S, Packham A, et al. Disproportionate cardiac hypertrophy during early postnatal development in infants born preterm. *Pediatr Res*. 2017;82:36–46. doi: 10.1038/pr.2017.96
- Denney PA. Effects of oxidative stress on embryonic development. *Birth Defects Res C Embryo Today*. 2007;81:155–162. doi: 10.1002/bdrc.20098
- Kamlin CO, O'Donnell CP, Davis PG, Morley CJ. Oxygen saturation in healthy infants immediately after birth. *J Pediatr*. 2006;148:585–589. doi: 10.1016/j.jpeds.2005.12.050
- Saugstad OD. Optimal oxygenation at birth and in the neonatal period. *Neonatology*. 2007;91:319–322. doi: 10.1159/000101349
- Huckstep OJ, Williamson W, Telles F, Burchert H, Bertagnolli M, Herdman C, Arnold L, Smillie R, Mohamed A, Boardman H, et al. Physiological stress elicits impaired left ventricular function in preterm-born adults. *J Am Coll Cardiol*. 2018;71:1347–1356. doi: 10.1016/j.jacc.2018.01.046
- Alphonse RS, Vadivel A, Fung M, Shelley WC, Critser RJ, Ionescu L, O'Reilly M, Ohls RK, McConaghy S, Eaton F, et al. Existence, functional impairment, and lung repair potential of endothelial colony-forming cells in oxygen-induced arrested alveolar growth. *Circulation*. 2014;129:2144–2157. doi: 10.1161/CIRCULATIONAHA.114.009124
- O'Reilly M, Thébaud B. Animal models of bronchopulmonary dysplasia. The term rat models. *Am J Physiol Lung Cell Mol Physiol*. 2014;307:L948–L958. doi: 10.1152/ajplung.00160.2014
- Dorfman AL, Polosa A, Joly S, Chemtob S, Lachapelle P. Functional and structural changes resulting from strain differences in the rat model of oxygen-induced retinopathy. *Invest Ophthalmol Vis Sci*. 2009;50:2436–2450. doi: 10.1167/iovs.08-2297
- Rivera JC, Sitaras N, Noueihed B, Hamel D, Madaan A, Zhou T, Honoré JC, Quiniou C, Joyal JS, Hardy P, et al. Microglia and interleukin-1 β in ischemic retinopathy elicit microvascular degeneration through neuronal semaphorin-3A. *Arterioscler Thromb Vasc Biol*. 2013;33:1881–1891. doi: 10.1161/ATVBAHA.113.301331
- Black MJ, Siebel AL, Gezmish O, Moritz KM, Wlodek ME. Normal lactational environment restores cardiomyocyte number after uteroplacental insufficiency: implications for the preterm neonate. *Am J Physiol Regul Integr Comp Physiol*. 2012;302:R1101–R1110. doi: 10.1152/ajpregu.00030.2012
- Bertagnolli M, Huyard F, Cloutier A, Anstey Z, Huot-Marchand JÉ, Fallaha C, Paradis P, Schiffrin EL, Deblois D, Nuyt AM. Transient neonatal high oxygen exposure leads to early adult cardiac dysfunction, remodeling, and activation of the renin-angiotensin system. *Hypertension*. 2014;63:143–150. doi: 10.1161/HYPERTENSIONAHA.113.01760
- Zydorczyk C, Comte B, Cambonie G, Lavoie JC, Germain N, Ting Shun Y, Wolff J, Deschepper C, Touyz RM, Lelièvre-Pegorier M, et al. Neonatal oxygen exposure in rats leads to cardiovascular and renal alterations in adulthood. *Hypertension*. 2008;52:889–895. doi: 10.1161/HYPERTENSIONAHA.108.116251
- Bertagnolli M, Dios A, Béland-Bonenfant S, Gascon G, Sutherland M, Lukaszewski MA, Cloutier A, Paradis P, Schiffrin EL, Nuyt AM. Activation of the cardiac renin-angiotensin system in high oxygen-exposed newborn rats: angiotensin receptor blockade prevents the developmental programming of cardiac dysfunction. *Hypertension*. 2016;67:774–782. doi: 10.1161/HYPERTENSIONAHA.115.06745
- Bertagnolli M. Preterm birth and renin-angiotensin-aldosterone system: evidences of activation and impact on chronic cardiovascular

- disease risks. *Protein Pept Lett* 2017;24:793–798. doi: 10.2174/0929866524666170728160243
22. Lautner RQ, Vilella DC, Fraga-Silva RA, Silva N, Verano-Braga T, Costa-Fraga F, Jankowski J, Jankowski V, Sousa F, Alzamora A, et al. Discovery and characterization of alamandine: a novel component of the renin-angiotensin system. *Circ Res*. 2013;112:1104–1111. doi: 10.1161/CIRCRESAHA.113.301077
 23. Price RL, Carver W, Simpson DG, Fu L, Zhao J, Borg TK, Terracio L. The effects of angiotensin II and specific angiotensin receptor blockers on embryonic cardiac development and looping patterns. *Dev Biol*. 1997;192:572–584. doi: 10.1006/dbio.1997.8764
 24. Quan A. Fetopathy associated with exposure to angiotensin converting enzyme inhibitors and angiotensin receptor antagonists. *Early Hum Dev*. 2006;82:23–28. doi: 10.1016/j.earlhumdev.2005.11.001
 25. Bertagnolli M, Casali KR, De Sousa FB, Rigatto K, Becker L, Santos SH, Dias LD, Pinto G, Dartora DR, Schaan BD, et al. An orally active angiotensin-(1-7) inclusion compound and exercise training produce similar cardiovascular effects in spontaneously hypertensive rats. *Peptides*. 2014;51:65–73. doi: 10.1016/j.peptides.2013.11.006
 26. Zydorczyk C, Comte B, Huyard F, Cloutier A, Germain N, Bertagnolli M, Nuyt AM. Developmental programming of eNOS uncoupling and enhanced vascular oxidative stress in adult rats after transient neonatal oxygen exposure. *J Cardiovasc Pharmacol*. 2013;61:8–16. doi: 10.1097/FJC.0b013e318274d1c4
 27. Lamata P, Niederer S, Nordsletten D, Barber DC, Roy I, Hose DR, Smith N. An accurate, fast and robust method to generate patient-specific cubic Hermite meshes. *Med Image Anal*. 2011;15:801–813. doi: 10.1016/j.media.2011.06.010
 28. Lewandowski AJ, Augustine D, Lamata P, Davis EF, Lazdam M, Francis J, McCormick K, Wilkinson AR, Singhal A, Lucas A, et al. Preterm heart in adult life: cardiovascular magnetic resonance reveals distinct differences in left ventricular mass, geometry, and function. *Circulation*. 2013;127:197–206. doi: 10.1161/CIRCULATIONAHA.112.126920
 29. Lewandowski AJ, Raman B, Bertagnolli M, Mohamed A, Williamson W, Pelado JL, McCance A, Lapidair W, Neubauer S, Leeson P. Association of preterm birth with myocardial fibrosis and diastolic dysfunction in young adulthood. *J Am Coll Cardiol*. 2021;78:683–692. doi: 10.1016/j.jacc.2021.05.053
 30. Crispi F, Bijmens B, Figueras F, Bartrons J, Eixarch E, Le Noble F, Ahmed A, Gratacós E. Fetal growth restriction results in remodeled and less efficient hearts in children. *Circulation*. 2010;121:2427–2436. doi: 10.1161/CIRCULATIONAHA.110.937995
 31. Rodríguez-López M, Cruz-Lemini M, Valenzuela-Alcaraz B, Garcia-Otero L, Sitges M, Bijmens B, Gratacós E, Crispi F. Descriptive analysis of different phenotypes of cardiac remodeling in fetal growth restriction. *Ultrasound Obstet Gynecol*. 2017;50:207–214. doi: 10.1002/uog.17365
 32. Cruz-Lemini M, Crispi F, Valenzuela-Alcaraz B, Figueras F, Sitges M, Bijmens B, Gratacós E. Fetal cardiovascular remodeling persists at 6 months in infants with intrauterine growth restriction. *Ultrasound Obstet Gynecol*. 2016;48:349–356. doi: 10.1002/uog.15767
 33. Sarvari SI, Rodríguez-Lopez M, Nuñez-García M, Sitges M, Sepulveda-Martinez A, Camara O, Butakoff C, Gratacos E, Bijmens B, Crispi F. Persistence of cardiac remodeling in preadolescents with fetal growth restriction. *Circ Cardiovasc Imaging*. 2017;10:e005270. doi: 10.1161/CIRCIMAGING.116.005270
 34. Gomez RA, Norwood VF. Developmental consequences of the renin-angiotensin system. *Am J Kidney Dis*. 1995;26:409–431. doi: 10.1016/0272-6386(95)90487-5
 35. Schütz S, Le Moullec JM, Corvol P, Gasc JM. Early expression of all the components of the renin-angiotensin-system in human development. *Am J Pathol*. 1996;149:2067–2079.
 36. Vinturache AE, Smith FG. Angiotensin type 1 and type 2 receptors during ontogeny: cardiovascular and renal effects. *Vascul Pharmacol*. 2014;63:145–154. doi: 10.1016/j.vph.2014.11.001
 37. Popescu CR, Sutherland MR, Cloutier A, Benoit G, Bertagnolli M, Zydorczyk C, Germain N, Phan V, Lelièvre-Pegorier M, Sartelet H, et al. Hyperoxia exposure impairs nephrogenesis in the neonatal rat: role of HIF-1 α . *PLoS One*. 2013;8:e82421. doi: 10.1371/journal.pone.0082421
 38. Marques FD, Ferreira AJ, Sinisterra RD, Jacoby BA, Sousa FB, Caliari MV, Silva GA, Melo MB, Nadu AP, Souza LE, et al. An oral formulation of angiotensin-(1-7) produces cardioprotective effects in infarcted and isoproterenol-treated rats. *Hypertension*. 2011;57:477–483. doi: 10.1161/HYPERTENSIONAHA.110.167346
 39. Marques FD, Melo MB, Souza LE, Irigoyen MC, Sinisterra RD, de Sousa FB, Savergnini SQ, Braga VB, Ferreira AJ, Santos RA. Beneficial effects of long-term administration of an oral formulation of Angiotensin-(1-7) in infarcted rats. *Int J Hypertens*. 2012;2012:795452. doi: 10.1155/2012/795452
 40. Oliveira AC, Melo MB, Motta-Santos D, Peluso AA, Souza-Neto F, da Silva RF, Almeida JFQ, Canta G, Reis AM, Goncalves G, et al. Genetic deletion of the alamandine receptor MRGD leads to dilated cardiomyopathy in mice. *Am J Physiol Heart Circ Physiol*. 2019;316:H123–H133. doi: 10.1152/ajpheart.00075.2018
 41. Santos RAS, Sampaio WO, Alzamora AC, Motta-Santos D, Alenina N, Bader M, Campagnole-Santos MJ. The ACE2/Angiotensin-(1-7)/MAS axis of the renin-angiotensin system: focus on Angiotensin-(1-7). *Physiol Rev*. 2018;98:505–553. doi: 10.1152/physrev.00023.2016
 42. Santos RA, Ferreira AJ, Verano-Braga T, Bader M. Angiotensin-converting enzyme 2, angiotensin-(1-7) and Mas: new players of the renin-angiotensin system. *J Endocrinol*. 2013;216:R1–R17. doi: 10.1530/JOE-12-0341
 43. Zhong J, Basu R, Guo D, Chow FL, Byrns S, Schuster M, Loibner H, Wang XH, Penninger JM, Kassiri Z, et al. Angiotensin-converting enzyme 2 suppresses pathological hypertrophy, myocardial fibrosis, and cardiac dysfunction. *Circulation*. 2010;122:717–728. doi: 10.1161/CIRCULATIONAHA.110.955369
 44. Lula I, Denadai AL, Resende JM, de Sousa FB, de Lima GF, Pilo-Veloso D, Heine T, Duarte HA, Santos RA, Sinisterra RD. Study of angiotensin-(1-7) vasoactive peptide and its beta-cyclodextrin inclusion complexes: complete sequence-specific NMR assignments and structural studies. *Peptides*. 2007;28:2199–2210. doi: 10.1016/j.peptides.2007.08.011