

Letter to the Editor

Epidermal Paracrine Signals May Regulate Dupuytren Contracture Myofibroblasts



Dupuytren contracture (DC), frequently treated by hand surgeons, is a common, benign fibrotic condition driven by myofibroblasts causing contractures and stimulating excess collagen production within DC nodules.¹ Within DC cords, myofibroblasts are present in reduced numbers and appear inactive.² We hypothesized that keratinocytes differentially express local paracrine signals overlying the DC nodules and cords that induce changes in myofibroblasts within DC lesions.

An Albany Medical Center institutional review board-approved study (#2330) was conducted using tissue obtained from 6 patients undergoing fasciectomy for DC. The samples consisted of adherent skin and underlying diseased fascia; these

were analyzed using trichrome staining and standard immunohistochemistry to detect the presence of α -smooth muscle actin and cyclo-oxygenase-2 (COX-2). We observed that within the DC cords, there was abundant fibrous tissue, with reduced numbers of α -smooth muscle actin-positive myofibroblasts that expressed COX-2. Taken together, these data suggested a potentially waning inflammatory local microenvironment because COX-2 induction has been demonstrated to engage in a negative, autocrine feedback loop that restrains the myofibroblast phenotype (Fig.).^{3,4}

Our findings, although preliminary, are consistent with a potential role of epidermal keratinocytes in tempering the profibrotic

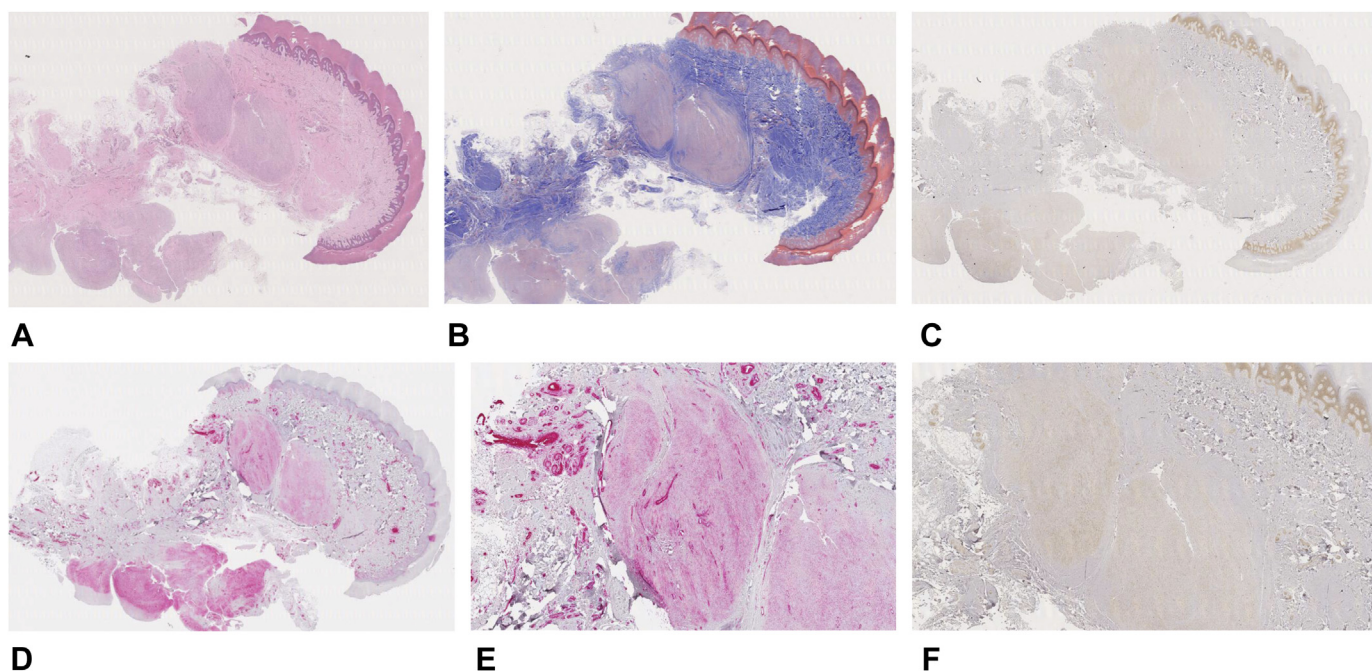


Figure. The adherent skin and underlying diseased fascia excised from a patient undergoing a limited palmdigital fasciectomy. **A** (Hematoxylin-eosin stain; magnification $\times 13.4$). The excised tissue demonstrates nodules of fibroblasts with bland nuclei and surrounding collagen depositions. **B** (Trichrome stain; magnification $\times 13.6$). Collagenous deposition is highlighted within the diseased fascia. **C** (COX-2 immunostain; magnification $\times 13.4$). Cytoplasmic staining of fibroblasts is demonstrated. **D** (SMA- α immunostain; magnification $\times 10$). The deposition of actin within collagen bundles is demonstrated. **E** (SMA- α immunostain; magnification $\times 30$). The deposition of actin within collagen bundles is demonstrated. **F** (COX-2 immunostain; magnification $\times 30$). The cytoplasmic staining of fibroblasts is demonstrated. SMA- α , α -smooth muscle actin.

Declaration of interests: No benefits in any form have been received or will be received related directly or indirectly to the subject of this article.

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myofibroblast phenotype in DC. Paracrine signaling from the epidermis to DC myofibroblasts is currently an understudied area that may provide new therapeutic approaches in which factors inducing fibroblast COX-2 or resulting prostanoids in DC lesions may be beneficial.

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References

1. Tomasek JJ, Schultz RJ, Haaksma CJ. Extracellular matrix-cytoskeletal connections at the surface of the specialized contractile fibroblast (myofibroblast) in Dupuytren disease. *J Bone Joint Surg Am.* 1987;69(9):1400–1407.
2. Verjee LS, Midwood K, Davidson D, Essex D, Sandison A, Nanchahal J. Myofibroblast distribution in Dupuytren's cords: correlation with digital contracture. *J Hand Surg Am.* 2009;34(10):1785–1794.
3. Zheng R, Varney SD, Wu L, DiPersio CM, Van De Water L. Integrin $\alpha 4\beta 1$ is required for IL-1 α - and Nrf2-dependent, Cox-2 induction in fibroblasts, supporting a mechanism that suppresses α -SMA expression. *Wound Repair Regen.* 2021;29(4):597–601.
4. Zheng R, Longmate WM, DeFrest L, et al. Keratinocyte integrin $\alpha 3\beta 1$ promotes secretion of IL-1 α to effect paracrine regulation of fibroblast gene expression and differentiation. *J Invest Dermatol.* 2019;139(9):2029–2038.e3.