SCIENTIFIC REPORTS

Received: 17 July 2015 Accepted: 08 January 2016 Published: 10 February 2016

OPEN Taxonomic status and molecular phylogeography of two sibling species of Polytremis (Lepidoptera: Hesperiidae)

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The skipper Polytremis theca species complex is widely distributed in the south of the Qinling Mountains in China. A recent study of the Polytremis genus suggested that this species might encompass two differentiated lineages. We tested this hypothesis, by carrying out a phylogenetic study of this agricultural pest based on nationwide sampling and the evaluation of mitochondrial and nuclear DNA markers. We show that this species is actually an amalgamation of two sibling taxa (P. t. theca and P. t. fukia), which displayed levels of genetic divergence as great as those generally found between sister species in the *Polytremis* genus, suggesting that they actually correspond to two distinct species. The Divergence time estimates suggest that an active period of speciation within Polytremis occurred within the Pleistocene eras. Based on its distinct phylogenetic placement and geographical isolation, we suggest that the subspecies should be elevated to full species status under the phylogenetic species concept, which has significant management implications.

Evans¹ described a lot of skipper species from Europe, Asia and Australia. He often described varieties, now considered subspecies under the current Code of Zoological Nomenclature², when specimens exhibited morphology similar to known taxa. It appears to follow the classical typological species concept where small differences represented regional variation in the species. Close inspection of such intraspecific variations has sometimes revealed that the species' actually contain several closely related multiple species, so-called 'sibling species' or 'cryptic species', that are difficult to distinguish morphologically from each other, at least without skilful eyes³⁻⁵. With the development of molecular techniques and the collection of greater number of specimens, some of his subspecies were argued, revised and elevated to species rank. Chiba & Eliot⁶, Devyatkin & Monastyrskii⁷, Guo et al.⁸ reevaluated Parnara guttatus batta Evans, 1949 by morphological features, molecular data and biogeographical implications and treated it as an independent species named Parnara batta Evans, 1949. Chiba et al.9 and Hsu¹⁰ regarded Polytremis menica kirazana Evans, 1949 as valid species-level named Polytremis kiraizana Sonan, 1938 by further morphological and phylogenetic analysis.

The skipper Polytremis theca species complex is widely distributed in the south of the Qinling Mountains in China, except Hainan Province and the southern tropical regions of Yunnan Province^{1,11,12}. Evans¹ described P. t. theca Evans, 1937 from Sichuan Province. He then described the P. t. fukia Evans, 1940 from Fujian Province that was lighter in coloration than the typical P. theca and existed 1or 2 white spots in space Cu₂ on the upperside of forewing. P. t. theca and P. t. fukia are nearly separated geographically.

If genetic differentiation has occurred, the taxonomic status of P. theca species complex can be reevaluation by molecular techniques. In a preliminary study of molecular phylogeny of the genus Polytremis Mabille 1904 using mitochondrial cytochrome c oxidase I (COI), we found the inter-subspecific distance between P. t. theca and P. t. fukia ranged up to 4.2%, which is higher than some inter-specific genetic distances in Polytremis¹³. Additionally, P. theca is also the only species whose intra-specific distance is more than 3%, a thresholds of species identification have been proposed in Lepidoptera for COI, in genus Polytremis¹⁴. Thus, we suspected the possible existence of a sibling species paired or the cryptic diversity in the species. However, this assertion was based on

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a very small number of samples and required confirmation in a larger sample. To answer the question, we conducted molecular phylogenetic analyses of *P. theca* species complex collected from 13 geographic localities in China using a region of mitochondrial genes (COI) and a region of nuclear genes (*wingless*).

Materials and methods

Samples collection. We collected specimens of *P. t. theca* (8) from five local regions and *P. t. fukia* (25) from eight different localities in China from 2008 to 2014 (Fig. 1; Table 1), and preliminarily identified them based on traditional wing and genitalia morphology characters recommended by Evans¹, Eliot¹⁵ and Chou¹⁶. Our sampling of 33 specimens covered the major range of the *P. theca* (Fig. 1). The outgroups included eight specimens of *P. nascens* Leech, 1893, five specimens of *P. mencia* Moore, 1877 (Table 1). All specimens were caught in the field, preserved by dehydration in small envelopes and dried with silica desiccant for further processing.

DNA extraction and sequencing. The DNA was isolated from legs of adult butterfly using a QIAamp DNA Mini kit (Qiagen, Hilden, Germany) essentially following the manufacturer's instructions but with some modification. Briefly, after adding proteinase K and buffer AL (QIAGEN®), the mixed homogeneous solution was incubated at 70 °C for 2 h. Subsequently, 200 µL of 100% ethanol was added and the mixture transferred to a QIAamp spin column. The mixture in the spin column was subjected to 3cycles of centrifugation at full speed (14,000g) for 1 min and the filtrate was returned to the spin column to increase the amount of DNA obtained. Partial sequences of mitochondrial gene COI (487 bp) was amplified and sequenced with primers HCO2198: 5'-TAA ACT TCA GGG TGA CCA AAA AAT CA-3' and LCO1490: 5'-GGT CAA CAA ATC ATA AAG ATA TTG G-3'17. The nuclear DNA fragments of wingless (389) were amplified and sequenced with primers WG1z: 5'-GAR TGY AAR TGY CAY GG-3' and WG2a: 5'-ACT ICG CAR CAC CAR TGG AAT GTR CA-3'¹⁸. The PCR for all amplicons were conducted in $20 \,\mu$ L final volume reactions containing $1 \,\mu$ L template DNA, $0.4 \,\mu$ M each of the amplification primers, 200 µM each dNTP, 1.5 mM MgCl₂, 50 mM KCl, 10 mM Tris-HCl (pH 8.3) and 2 U Taq polymerase (Takara, Otsu, Shiga, Japan). The thermal profile used was as follows. COI: 95 °C denaturation for 5 min followed by 35 cycles of 94 °C for 45 s, 42 °C for 1 min, 90 °C for 90 s and a final elongation step at 72 °C for 10 min. Wingless: 95 °C denaturation for 5 min followed by 30 cycles of 94 °C for 60 s, annealing at 55 °C for 60 s and extension at 72 °C for 2 min, with a final extension of 72 °C for 10 min.

Extraction blanks were run in all reactions to control for contamination during the extraction and PCR processes. The amplification products were subjected to electrophoresis in a 2% (w/v) agarose gel in TAE buffer (0.04 M Tris–acetate, 0.001 M EDTA) with a DL1000 ladder size marker (Takara, Otsu, Shiga, Japan) to determine whether the amplification reactions were successful. In addition, after electrophoresis in agarose gels, the amplification products were extracted using the Wizard SV Gel and PCR Clean-up System (Promega, Madison, WI, USA) for sequencing. Finally, all the haplotypes obtained were deposited in GenBank (Table 1).

Statistical analyses. The sequence data of the mitochondrial COI and nuclear *wingless* were aligned with published homologous sequence from genus *Polytremis* (e. g., *P. eltola* Hewitson, 1869, *P. discreta* Elwes & Edwards, 1897, etc.) as well as *Borbo cinnara* Wallace, 1866 and *Pseudoborbo bevani* Moore, 1878 (presented in Table 1) respectively, translated to amino acid sequences to check for nuclear mitochondrial pseudogenes (numts) and pruned to remove redundant sequences with Bioedit v.7.0¹⁹. The haplotype sequence matrix was used for all subsequent phylogenetic analyses (Table 1). Phylogenetic trees were constructed by the ML (maximum-likelihood) methods with PhyML²⁰. Modeltest 3.7²¹ was used to select the optimal nucleotide substitution models following the Akaike Information Criterion (AIC). In the ML analysis, a heuristic search was

						Accession number		
Locality	Specimen ID	Sex	Date	Altitude	HaolotypeAltitude(COI/wingless)		wingless	
Polytremis theca theca	[1	1			
Lianglu, Tianquan County, Sichuan Province (N	Ptt_LL 1	Ŷ	6-Jul-11	not recorded	Ptt I/Ptt I	KC684410 [°]	KR911938	
29°55′; E 102°23′)	Ptt_LL 2	ð	11-Aug-11	not recorded	Ptt I/Ptt II	KC684410*	KR911939	
Wolong, Wenchuan	Ptt_WL 1	ð	6-Aug-11	not recorded	Ptt II/Ptt II	KC684411*	KR911939	
(N 31°29′; E 103°35′)	Ptt_WL 2	ð	6-Aug-11	not recorded	Ptt II/Ptt II	KC684411*	KR911939	
Luding, Sichuan Province (N 29°55′; E 102°15′)	Ptt_LD 1	ð	19-Jul-11	not recorded	Ptt II / Ptt II	KC684411*	KR911939	
Feng County, Shanxi	Ptt_FC 1	Ŷ	19-Jul-11	not recorded	Ptt III/Ptt I	KR911924	KR911938	
Province (N 33°54'; E 106°31')	Ptt_FC 2	ð	19-Jul-11	not recorded	Ptt III/Ptt I	KR911924	KR911938	
Qinling, Shanxi Province 34°14′N 103°54′E	Ptt_QL 1	ð	16-Jul-10	not recorded	Ptt III/Ptt I	KR911924	KR911938	
Polytremis theca fukia				-				
Qinliangfeng, Lin'an County, Zhejiang Province (N 30°59'; E 118°52')	Ptf_QLF 1	ð	11-Apr-13	800 m	Ptf II/Ptf I	KC684406*	KR911935	
	Ptf_TMS 1	Ŷ	8-May-09	not recorded	Ptf I/Ptf I	KC684409*	KR911935	
Tianmushan, Lin'an	Ptf_TMS 2	ð	20-Sep-08	not recorded	Ptf I/Ptf I	KC684409*	KR911935	
(N 30°19'; E 119°25')	Ptf_TMS 3	ð	17-Aug-09	not recorded	Ptf I/Ptf I	KC684409*	KR911935	
	Ptf_TMS 4	ð	20-Aug-13	300 m	Ptf II/Ptf I	KC684406*	KR911935	
Wuyishan, Fujian Province	Ptf_WYS 1	ð	19-Sep-08	not recorded	Ptf III/Ptf II	KC684408*	KR911936	
(N 27°44'; E 118°01')	Ptf_WYS 2	ð	19-Sep-08	not recorded	Ptf III/Ptf II	KC684408*	KR911936	
lingangshan Ji'an county	Ptf_JGS 1	Ŷ	19-Oct-10	not recorded	Ptf I/Ptf I	KC684409*	KR911935	
Jiangxi Province (N 26°45';	Ptf_JGS 2	Ŷ	19-May-10	not recorded	Ptf II/Ptf I	KC684406*	KR911935	
E 114°17′)	Ptf_JGS 3	Ŷ	20-May-10	not recorded	Ptf II/Ptf I	KC684406*	KR911935	
Yueshan, Fengxin County,	Ptf_YS 1	Ŷ	13-Jul-13	400-600 m	Ptf II/Ptf I	KC684406*	KR911935	
Jiangxi Province (N 28°79'; E 115°18')	Ptf_YS 2	Ŷ	13-Jul-13	400-600 m	Ptf IV/Ptf I	KR911919	KR911935	
	Ptf MES 1	Ŷ	8-Jul-11	400 m	Ptf V/Ptf II	KC684407*	KR911936	
Maoershan , Xin'an County, Guangxi Province (N 25°49' - E 110°00')	Ptf MES 2	ð	7-Jul-11	400 m	Ptf II/Ptf III	KC684406*	KR911937	
	Ptf_MES 3	ð	7-Jul-11	400 m	Ptf VI/Ptf II	KR911920	KR911936	
	Ptf_MES 4	Ŷ	7-Jul-11	400 m	Ptf VII / Ptf III	KR911921	KR911937	
	Ptf_MES 5	ð	8-Jul-11	400 m	Ptf II/Ptf III	KC684406*	KR911937	
(1(20 1), 2110 00)	Ptf_MES 6	ð	8-Jul-11	400 m	Ptf II/Ptf III	KC684406*	KR911937	
	Ptf_MES 7	ð	8-Jul-11	400 m	Ptf VIII/Ptf I	KR911922	KR911935	
	Ptf_MES 8	ð	9-Jul-11	2000 m	Ptf VIII/Ptf I	KR911922	KR911935	
	Ptf_AJP 1	ð	13-Jul-11	1200 m	Ptf I/Ptf III	KC684409*	KR911937	
Anjiangpin ,Lingui County,	Ptf_AJP 2	ð	13-Jul-11	1200 m	Ptf II/Ptf II	KC684406*	KR911936	
Guangxi Province (N 25°14'; E 110°12')	Ptf_AJP 3	ð	13-Jul-11	1200 m	Ptf IX/Ptf III	KR911923	KR911937	
	Ptf_AJP 4	ð	14-Jul-11	1350 m	Ptf VIII/Ptf II	KR911922	KR911936	
Bifengxia, Yaan County, Sichuan Province (N 30°03'; E 102°62')	Ptf_BFX 1	Ŷ	2-Aug-14	not recorded	Ptf II/Ptf I	KC684409*	KR911935	
Outgroups								
Polytremis nascens								
Zhanghe, Langao County, Shaanxi Province (N	Pn_ZH 1	ð	30-Jul-12	1600 m	Pn I/Pn I	KJ574014*	KR911931	
32°32′; E 108°14′)	Pn_ZH 2	ð	30-Jul-12	1600m	Pn II/Pn I	KC684397*	KR911931	
Hailuogou, Tianquan	Pn_HLG 1	ð	27-Jul-06	2300 m	Pn I/Pn II	KJ574014*	KR911932	
(N 29°34'; E 102°04')	Pn_HLG 2	ð	27-Jul-06	2300 m	Pn II/Pn II	KC684397*	KR911932	
Lianglu, Tianquan County, Sichuan Province (N 29°55'; E 102°23')	Pn_LL 1	ð	3-Sep-10	1450 m	Pn I/Pn II	KJ574014*	KR911932	
Fenghuangshan, Hanyin	Pn_FHS 1	Ŷ	3-Aug-12	1400 m	Pn II/Pn I	KC684397*	KR911931	
County, Shaanxi Province (N 32°90'; E 108°50')	Pn_FHS 2	Ŷ	3-Aug-12	1600 m	Pn III/Pn I	KJ574015 [*]	KR911931	
Houhe, Wufeng County, HuBei Province (N 30°22'; E 110°68')	Pn_HH 1	ð	9-Jul-13	1200 m	Pn II/Pn III	KC684397*	KR911933	
Continued								

						Accession number	
Locality	Specimen ID	Sex	Date	Altitude	Haolotype (COI/wingless)	COI	wingless
Polytremis mencia							
	Pm_TMS 1	ð	5-Jun-11	not recorded	Pm I/Pm I	KC684415*	KR911940
Tianmushan, Lin'an County, Zheijang Province	Pm_TMS 2	ð	15-Jun-10	not recorded	Pm I/Pm I	KC684415*	KR911940
(N 30°19′; E 119°25′)	Pm_TMS 3	ð	30-May-10	not recorded	Pm II/Pm I	KC684414*	KR911940
	Pm_TMS 4	Ŷ	20-Sep-08	not recorded	Pm III/Pm I	KC684416 [*]	KR911940
Ningbo, Zhejiang Province (N 29°52'; E 121°32')	Pm_NB 1	ð	23-Aug-11	not recorded	PmIV/Pm I	KC684417 [*]	KR911940
Polytremis pellucid							
Tianmushan, Lin'an County, Zhejiang Province (N 30°19′; E 119°25′)	121112009	ę	19-Sep-08	not recorded	P. pellucid	KC684393*	KR911941
Polytremis zina		-					
Tianmushan, Lin'an County, Zhejiang Province (N 30°19'; E 119°25')	121116021	ð	18-Aug-09	not recorded	P. zina	KC684395*	KR911942
Polytremis discreta							
Baoxing County, Sichuan Province (N 30°22'; E 102°48')	121116024	ð	2-Jul-08	not recorded	P. discreta	KC684392*	KR911943
Polytremis lubricans	1				1		1
Baoxing County, Sichuan Province (N 30°22'; E 102°48')	121116027	ð	2-Jul-08	not recorded	P. lubricans	KC684391*	KR911944
Polytremis eltola							
Jingxiu, Guangxi Province (N 24°07'; E 110°11')	121116030	ð	28-Jul-11	not recorded	P. eltola	KC684389*	KR911945
Polytremis gigantean							
Qingchengshan, Sichuan Province (N 30°53'; E 103°34')	121119033	Ŷ	28-Aug-11	not recorded	P. gigantean	KC684403*	KR911929
Polytremis matsuii							
Hongya, Sichuan Province (N 29°54'; E 103°22')	121119036	ð	4-Jun-11	not recorded	P. matsuii	KC684400*	KR911927
Polytremis caerulescens							
Lianglu, Tianquan County, Sichuan Province (N 29°55'; E 102°23')	121119039	ð	21-Jul-11	not recorded	P. caerulescens	KC684399*	KR911925
Polytremis jigongi							
Tianmushan, Lin'an County, Zhejiang Province (N 30°20'; E 119°23')	121119042	ð	11-Jul-09	not recorded	P. jigongi	KC684404*	KR911934
Polytremis kiraizana							
Qilai, Taiwan Province (N 24°01'; E 121°22')	121124051	ð	11-Jul-91	not recorded	P. kiraizana	KC684401*	KR911926
Polytremis suprema							
Jingxiu, Guangxi Province (N 24°07′; E 110°11′)	121124052	ð	31-Jul-11	not recorded	P. suprema	KC684402*	KR911928
Polytremis gotama							
Lijiang, Yunnan Province (N 26°51'; E 100°13')	130101067	ð	28-Jul-06	not recorded	P. gotama	KC684396*	KR911930
Borbo cinnara							
Taidong, Taiwan Province (N 22°59'; E 120°59')	121124069	ð	17-Sep-07	not recorded	B. cinnara	KC684418*	KR911947
Pseudoborbo bevani			1				
Nantou, Taiwan Province (N 23°55'; E 120°41')	121124067	Ŷ	14-Sep-03	not recorded	P. bevani	KC684419*	KR911946

Table 1. List of specimen information used for this study. *indicates the sequences retrieved from GenBank.

conducted. The starting tree for branch-swapping was from stepwise addition. Nodal support of the ML tree was estimated by 1000 bootstraps. Network profile of the haplotypes identified in *P. t. theca*, *P. t. fukia*, *P. nascens* and *P. mencia* was constructed with Network4.5 using the median-joining method²². The haplotype diversity (Hd) and nucleotide diversity (π) for *P. t. theca* and *P. t. fukia* were estimated by DnaSP4.90²³.

For the COI data set, pairwise F_{ST} was also calculated with Arlequin v3.0²⁴, which accurately reflects patterns of genetic variation. Correspondingly, gene flow was estimated in Arlequin v3.0²⁴. Two level hierarchical analyses

A. COI

B.

COI	11111111111111122222222333333333333344444444	
	1224444555566778011233445556899000026667823345678888901112234578	
	8124569013989479139519690284519246914792548970840289613692819874	
Ptf I	ACCTCATATATTCATTTAATTCTTCTTCCTGTTAACTCGTTTTGAACCTTCCTCGACATTTTGC	5
Ptf II		ğ
D+f III		0
FUL III DAE IV		1
PTI IV	A	1
Ptf V		1
Ptf VI	. GG C	1
Ptf VII	. GG C	1
Ptf VIII	CGT	3
Ptf IX	G	1
Ptt I	CTT C GC GCC ATTC G. TT ATT CA.	2
Ptt II	TT C GC GCC ATTC C G. TT ATT CA.	3
Ptt III	TT C GC GCC AATTC G. TTCAT TATT CA.	3
Pn I	TT CT. AT CCT CCT. ATTC. A. C. T. TA C. A TT TCT C A.	3
Pn II	TT CT. AT CCT CC ATTC C. T. TA C. A TT T. T C A.	4
Pn III	TT CT. AT CCT CC ATTCA CTT. TA C. A TT T CA. A.	1
Pm I	TTTCATTCATTT.T.TAC.TT.TT.TA.TT.	2
Pm II	ТТТСАТТСАТТТ.Т.ТАС.ТТ.ТТ.ТА.ТТ.А.А.	1
Pm III	TT. T. CAT. T. C. ATT. T. T. TA. C. TT. T. A. TTC. A.	1
Pm IV	TT. T. CAT. T. C. ATT. T. T. TA. A. TT. T. A. TTC. CA.	î
		-
wingless	12222222333	
0	2374014445688234	

	2374014445688234	
	12071572459289138	
Ptf I	CTGGGGCCAAATCCCCC	12
Ptf II	A	6
Ptf III	G T.	6
Ptt I	G. A. A T T	4
Ptt II	G. A. AA. T T	4
Pn I	. C GG. C	4
Pn II	. C A. T. GG. C	3
Pn III	. C GG. C T	1
Pm I	G. A. A T TTT	Ę

Figure 2. Sequence nucleotide substitutions of (**A**) the 19 haplotypes identified in 45 COI gene sequences and (**B**) the 9 haplotypes identified in 45 *wingless* gene sequences of *P. t. theca* (Ptt), *P. t. fukia* (Ptf), *P. nascens* (Pn) and *P. mencia* (Pm). The numbers on top of sequences indicates the nucleotide position which is relative to beginning of the fragment investigated in this study. The last column shows the number of individuals shared each haplotype. Dots (·) denote the identity with the reference sequence (Ptf I).

of molecular variance (AMOVA) were conducted to evaluate possible population genetic structure of *P. t. theca* and *P. t. fukia* using Arlequin v3.0 with 1,000 permutations. We calculated Tajima's D^{25} and Fu's F statistic²⁶ and ran 10,000 coalescent simulations for each statistic to create 95% confidence intervals investigate the historical population demographics and testing whether the sequences conformed to the expectations of neutrality. Pairwise mismatch distribution analyses were performed for all *P. theca* specimens and two subspeices specimens separately to find the evidence of past demographic expansions using DnaSP4.90²³. The times to the most recent common ancestor of the major lineages and the whole population were estimated using relax-clock molecular dating estimation implemented in the BEAST $1.5.2^{27}$. Analyses using the HKY model of nucleotide substitution with gamma distributed rate variation among sites were performed. The Yule speciation method was assumed and the nucleotide substitution rate of 3.54% per million years that has generally been calibrated for COI in insect²⁸ was used. Chains were run for 50 million generations, with the first 20% discarded as burn-in. The results were summarized through TRACER 1.5^{29} .

Result

Genetic Divergences and haplotype networks. All 46 samples yielded high-quality of DNA and were successfully sequenced for the mitochondrial COI and nuclear *wingless* (accession numbers KR911919-KR911947, Table 1, Fig. 2). The data set of COI alignment contains 487 nucleotide positions, of which 64 positions are variable and 47 are parsimony informative. The mean base composition of the fragment shows a strong bias of A + T (T 39.4%, C 18.2%, A 28.7% and G 13.7%), as found commonly in insect mitochondrial genomes³⁰. Nineteen haplotypes were identified in all 46 samples (three in *P. t. theca*, nine in *P. t. fukia*, three in *P. nascens* and four in *P. mencia*) and the haplotype network was constructed and presented in Fig. 3. There was no shared haplotype among the four taxa. Haplotypes of the same taxon differed from each other by no more than five mutation distance. The five mutation distance existed between the haplotype Ptt I and Ptt III of *P. t. theca*. The potential ancestral haplotype of *P. t. fukia*, defined by its central position in the network, was designated as Ptf I, which was found in three samples from West Tianmu Mountains, one from Jinggang Mountains, and one from Lingui. Ptf II was the most common haplotype in *P. t. fukia* and shared with ten samples. Haplotype Ptf III was found in two samples from Wuyishan. Haplotype Ptf XIII was identified in two sampled from Maoershan and one sampled from Anjiangpin. The remaining haplotypes of *P. t. fukia* occurred in only one individual. At least 18 nucleotide substitutions were observed between the potential ancestral haplotypes of *P. t. theca* (Ptt I) and that



Figure 3. Network profile of (**A**) COI and (**B**) *wingless* gene haplotypes based on the nucleotide sequences of *P. t. theca, P. t. fukia, P. nascens* and *P. mencia*. Each haplotype is represented by a circle. The area of the circles is proportional to the number of individuals that shared the haplotype. The circles labelled 1, 2, 5, 10 and 15 on the right hand side represent the number of individuals. The number of unique and shared haplotypes identified in different locations is shown in Table 1. The order of mutations (which were scored relative to haplotype Ptt I) on a branch is arbitrary.

of *P. t. fukia* (Ptf I). In addition, the haplotype (Pn I) of *P. nascens* differed from Ptt I and Ptf I by 29 and 25 nucleotide substitutions while the haplotype (Pm I) of *P. mencia* differed from Ptt I and Ptf I by 26 and 23 nucleotide substitutions, respectively (Figs 2 and 3).

The data set of nuclear *wingless* contains 390 nucleotide positions without gaps or stop codons, of which 18 positions are variable and 9 are parsimony informative. In total, ten haplotypes were found in all samples, in which two haplotypes were found in *P. t. theca*, four in *P. t. fukia*, three in *P. nascens* and one in *P. mencia*. All the haplotypes were used for network construction with the software Network 4.5 using the median-joining method (Fig. 3). Haplotypes of the same taxon differed from each other by no more than two mutation distance, in particular, the two haplotypes in 13 samples of *P. t. theca* differed by only one-mutation distance. Five nucleotide substitutions were observed between the potential ancestral haplotypes of *P. t. theca* (Ptt I) and that of *P. t. fukia* (Ptf I). In addition, the haplotype (Pn I) of *P. mencia* differed from Ptt I and Ptf I by nine and five nucleotide substitutions, respectively (Figs 2 and 3).

	Ns	Nh	Hd	Nv	π	SD (π)	D	F
All P. theca samples (COI)	33	12	0.875	38	0.0207	0.0019	0.281	2.368
P. t. theca samples (COI)	8	3	0.750	6	0.0064	0.0048	1.598	2.631
P. t. fukia samples (COI)	25	9	0.803	13	0.0050	0.0070	-1.014	-1.886*
All P. theca samples (wingless)	33	5	0.773	9	0.0078	0.0057	1.122	1.509
P. t. theca samples (wingless)	8	2	0.571	1	0.0015	0.0010	1.444	1.100
P. t. fukia samples (wingless)	25	3	0.640	3	0.0030	0.0020	1.080	1.159

Table 2. Genetic diversity and neutrality tests calculated for *P. t. theca* and *P. t. fukia*. Ns-number of samples, Nh–Number of haplotypes, Hd–Haplotype diversity, Nv–Number of variable sites, π –nucleotide diversity, SD–standard deviation, D–Tajima's D statistic, F–Fu's F statistic, *–significant difference.





Source of variation	df	Sum of squares	Variance components	Percentage variation	Φ Statistic
Among populations	1	169.650	9.98266 Va	88.53	_
Within populations	11	45.269	1.29341 Vb	11.47	0.896 (p < 0.01)
Total	12	214.919	11.27607		
Fixation index	0.8853				

Table 3. Analysis of molecular variance (AMOVA) for the COI sequences of P. t. theca and P. t. fukia.

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Overall, *P. t. theca* had a lower diversity than *P. t. fukia* according to the result of analysis of both mitochondrial COI and nuclear *wingless*. The haplotype diversity (Hd) and nucleotide diversity (π) for *P. t. theca* and *P. t. fukia* are given in Table 2. Additionally, they differed from each other by $5.07 \pm 0.49\%$ (4.3–5.9% divergence) for the COI sequences and by $1.70 \pm 0.27\%$ (1.3–2.1% divergence) for the *wingless* sequences.



Figure 5. Mismatch distributions of pairwise nucleotide differences for (**A**) *P. t. theca*, (**B**) *P. t. fukia* and (**C**) *P. t. theca* and *P. t. fukia*. X axis: Pairwise Differences. Y axis: Frequency. The circles show the observed distribution of pairwise difference. The solid lines represent the expected equilibruim distributions.

Phylogenetic and population structure analysis. Phylogenetic analyses were performed using ML method. For the COI gene sequence data, the submodel GTR + I + G was selected. For the *wingless* sequence data, the submodel GTR + G was selected. Figure 4A shows the ML tree based on the data set of COI. On the phylogeny, the haplotypes of *P. theca* were split into two discrete clades. One clade with three haplotypes consisted exclusively of the *P. t. theca* (strongly supported by the bootstrap probability of 99%), whereas the other clade with nine haplotypes exclusively consisted of the *P. t. fukia* (strongly supported with bootstrap probability of 99%). Figure 4B shows the ML tree based on the data set of *wingless*. Three haplotypes, constituted a monophyletic group, exclusively containing *P. t. fukia* butterflies whereas two haplotypes of the *P. t. theca*, *P. pellucida* Murray, 1875, *P. zina* Evans, 1932 and *P. mencia* Moore, 1877 constituted a strongly supported clade (86%). On the other hand, two *P. t. theca* haplotypes formed a strongly supported clade (86%). Mitochondrial COI and nuclear *wingless* strongly supports the distinction between *P. t. theca* and *P. t. fukia*.

The AMOVA for the COI sequences of *P. t. theca* and *P. t. fukia* revealed that 88.53% of the genetic variation was among populations and 11.47% was within populations (Table 3). The average Φ ST value as 0.896 (p < 0.01), suggesting significant genetic variation among the populations. Pairwise estimates of F_{ST} (0.885) and gene flow (Nm = 0.065) between *P. t. theca* and *P. t. fukia* suggests that the subspecies in this species are highly differentiated.

Demographic inference and estimation of divergence times. Demographic history changes were analyzed for *P. t. theca* and *P. t. fukia* populations through neutrality tests and mismatch distribution. The neutrality tests reveal that the Mitochondrial COI appear to be not evolving neutrally as Fu's F values in *P. t. fukia* group are negative significantly (Table 2). The Tajima's D and Fu's F values were non-significantly positive in





	wing								
	Color of cilia of wings	Color of Underside ground	Number of spots in space Cu2 of the forewing	Color of scales scattered in costa and subapical area of forewing	Color of scales scattered in discal area and dorsum of hindwing	Ductus bursae			
P. t. theca	brown	yellowish brown	0	greenish ochreous	greenish ochreous	thin			
P. t. fukia	greyish white	greenish ochreous	1 or 2	greyish white	greyish white	thick			

Table 4. Different morphological features of genitals and wings between P. t. theca and P. t. fukia.

P. t. theca group and all *P. theca* samples group. The mismatch analysis yielded a unimodal distribution of pairwise differences for *P. t. fukia* (Fig. 5B) compared to the multimodal distribution of *P. t. theca* samples (Fig. 5A) and the pooled samples (Fig. 5C). According to Rogers and Harpending³¹, the observed curves with unimodal representing population expansion and the observed curves with many peaks or resemblance to expected curves mean equilibrium population, which further elucidating the demographic history of *P. theca*. The results suggest that population expansion in *P. t. fukia* and population equilibrium in *P. t. theca*. Divergence time analysis with an uncorrelated lognormal relaxed clock run in BEAST produced a tree with a topology similar to ML tree (Fig. 6). *P. t. theca* diverged from *P. t. fukia* around 0.81 (HPD = 0.53–1.28) million years ago (Mya) during the Pleistocene (node a in Fig. 6). *P. theca* diverged from other congeners included in the analysis about 1.36 (HPD = 1.02–1.53) Mya during the Pleistocene eras (node b in Fig. 6).

Discussion

There is a small region of overlap in central Sichuan province in the distribution of *P. t. theca* and *P. t. fukia*, but otherwise they are not sympatric (Fig. 1). *P. t. theca* inhabits the higher elevations of west-central Sichuang Province and the Qinling Mountains¹⁶. *P. t. fukia* occurs in the whole southeastern of China, from northern Zhejiang Province, Jiangxi Province to east-central Yunnan Province^{9,32,33}. According to the description on morphological variation between *P. t. theca* and *P. t. fukia*¹², we found a different morphological feature existing in female genitalia except for the different color and spot number in some part of wings (Table 4). The ductus bursae of female *P. t. theca* is thinner than that of female *P. t. fukia*. We suspected thetaxonomic status of the subspecies from their geographic separation and the morphological variation.

Mitochondrial haplotypes sampled from *P. theca* form well supported clades that closely correspond with subspecific boundaries delimited primarily on the basis of wing color and pattern (Fig. 4A). The haplotype clades associated with both subspecies are deeply genetically divergent, differing from each other by $5.07 \pm 0.49\%$ (4.3–5.9% divergence). This degree of divergence suggests that evolutionary separation of both subspecies occurred about 0.81 (HPD = 0.53-1.28) Mya, likely sometime during the Pleistocene based on a molecular clock calibration of 3.54% pairwise divergence per million years for a homologous mtDNA fragment in other insect species²⁸. (fossil data and geographical calibration event are not available for this group). It is noteworthy for the nuclear *wingless* sequences that *P. t. theca* and *P. mencia* are considered distinct species with a genetic divergence of $0.65 \pm 0.15\%$ while the *P. t. fukia* is currently considered a subspecies of the *P. t. theca* despite $1.70 \pm 0.27\%$

sequence divergence. The phylogenetic of *wingless* gene indicates that the *P. theca* is paraphyly with three species sister to the clade of *P. t. theca* (Fig. 4B). While 100% of *P. t. fukia* constitutes one separate clade, the clade consisting of *P. t. theca* also includes *P. pellucid*, *P. zina* and *P. mencia*. The clade consisting of *P. t. theca* is not monophyletic, but complex. This suggests that *P. t. theca* and *P. t. fukia* differ from each other, as evident from the COI tree where they form two separate clades (Fig. 4A). Concordance between strongly differentiated mtDNA, nuclear haplotype clades and phenotypic variation supports the hypothesis that both subspecies of *P. theca* deserves recognition at the species-level under the general lineage concept of species^{4,34}.

Two neutrality tests were chosen for the demographic history analysis, and significantly negative values of neutrality statistics can be indicative of background selection, but are also consistent with either population subdivision or expansion. Fu's Fs is significantly negative in P. t. fukia group reveal that the mtDNA of P. t. fukia appear to be not evolving neutrally. Additionally, we analysed the population size change of P. t. theca, P. t. fukia and all *P. theca*, respectively by the software $DnaSP4.90^{23}$ and got no evidence for population expansion in P. t. theca group³¹ (Fig. 5A). However, we got unimodal curves representing population expansion in P. t. fukia³¹ (Fig. 5B). The results suggest that population expansion in P. t. fukia. We still confirm the result of the population size change in haplotype network (Fig. 3). Statistical parsimony network reflects genealogical relationships of the mtDNA haplotypes, that is, single mutation steps separate adjacent haplotypes in the network and older haplotypes are placed at internal branching points whereas younger ones occur toward the tip positions³⁵. The haplotypes network of P. t. fukia displays a star-like pattern (Fig. 3). Haplotype I, the second most common and geographically widespread in central-west of China, is in the star's centre and derivatives are connected to it by short branches. Based on coalescence theory, the star-like topologies for this cluster strongly suggest the effect of a population expansion³⁶. In our study, a higher F_{ST} value indicated a lower level of gene flow (Nm) and higher genetic differentiation among populations. The results of two-level AMOVA show that significant genetic variation exists among the examined populations. These results provide a second line of support to a conclusion that the P. t. fukia is a different species.

In conclusion, our investigations and analyses revealed significant molecular and biogeographical differences between *P. t. theca* and *P. t. fukia*. We propose that *P. t. fukia* should be treated as a distinct species called *Polytremis fukia* Evans 1940 under the Phylogenetic Species Concept. In fact, it has been recently found that other species previously considered subspecies based on morphology are in fact sibling species that passed unnoticed until the advent of molecular techniques^{37–40}. Results from our study strengthen information about the *Polytremis* species complex and help in developing appropriate integrated pest management strategies for these insect pests.

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Acknowledgements

We thank for Huang Hao (Qingdao, China), Chen Zhibin (Shanghai Zoological Park, Shanghai, China), Tang Liang, Hu Jiayao, Yin Ziwei, Li Lizhen and their colleagues (Shanghai Normal University, Shanghai, China) for providing valuable comments and specimens throughout the project. This study was financially supported by grants from National Natural Science Foundation of China (No. 31401997), Innovation Program of Shanghai Municipal Education Commission–China (No. 14YZ068) and Innovation Program for University Student in Shanghai, China (No. A-9103-15-035088 and A-9103-15-035089).

Author Contributions

W.J. and J.Z. initiated the project. W.J. performed bioinformatics analyses. W.J. and W.Y. designed experiments. H.H., Y.L., M.R., Y.M. and L.Z. carried out experiments. W.J. and J.Z. analysed data and interpreted results. W.J. wrote the manuscript.

Additional Information

Competing financial interests: The authors declare no competing financial interests.

How to cite this article: Jiang, W. et al. Taxonomic status and molecular phylogeography of two sibling species of *Polytremis* (Lepidoptera: Hesperiidae). *Sci. Rep.* **6**, 20820; doi: 10.1038/srep20820 (2016).

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