PROKARYOTES



Draft Genome Sequence of *Bacillus licheniformis* VSD4, a Diesel Fuel–Degrading and Plant Growth–Promoting Phyllospheric Bacterium

Vincent Stevens,^a Sofie Thijs,^a Breanne McAmmond,^b Tori Langill,^a Jonathan Van Hamme,^b Nele Weyens,^a Jaco Vangronsveld^a

Centre for Environmental Sciences, Hasselt University, Diepenbeek, Belgium^a; Department of Biological Sciences, Thompson Rivers University, Kamloops, British Columbia, Canada^b

ABSTRACT We report here the 4.19-Mb draft genome sequence of *Bacillus licheniformis* VSD4, a Gram-positive bacterium of the *Bacillaceae* family, isolated from leaves of *Hedera helix* growing at a high-traffic city center in Belgium. Knowledge about its genome will help to evaluate its potential as an inoculant in phylloremediation applications.

Various bacteria within the genus *Bacillus*, including members of *Bacillus licheniformis*, have previously been associated with diesel fuel degradation (1, 2) and plant growth promotion (3, 4). *B. licheniformis* VSD4 was isolated from the leaves of *Hedera helix* plants growing at a high-traffic city center in Belgium. *In vitro* analyses indicated that this bacterium is capable of utilizing diesel fuel as a carbon source and producing compounds related to plant growth promotion. Partial 16S rRNA gene sequence data showed that VSD4's closest relative is *B. licheniformis* ATCC 14580 (GenBank accession no. CP000002).

RNA-free DNA was extracted from stationary-phase cells grown in LB medium using a PureLink genomic DNA minikit (Thermo Fisher Scientific, Waltham, MA, USA), prior to digesting and ligating sequencing adaptors/barcodes using an Ion Xpress Plus fragment library kit (Thermo Fisher Scientific). Processed DNA was size-selected (480 bp) on a 2% E-Gel SizeSelect agarose gel and purified using Agencourt AMPure XP beads (Beckman Coulter, Inc., Brea, CA, USA). The library dilution factor was determined using an Ion Universal library quantitation kit prior to amplification and enrichment with an Ion PGM Hi-Q Template OT2 400 kit on an Ion OneTouch 2 system. The enriched Ion Sphere Particles were quantified using an Ion Sphere quality control kit. Sequencing was performed on an Ion 316 Chip version 2 (Ion PGM system) with an Ion PGM Hi-Q View sequencing kit (Thermo Fisher Scientific).

In total, 428,739 reads (mean length, 289 bases) generated 124 Mb (114 Mb with \geq Q20) of data. Reads were assembled using MIRA version 4.0rc4 (5), trimmed into 270 contigs \geq 500 bp, giving a consensus length of 4,188,588 bp at 39.4× coverage (largest contig, 471,073 bp; N_{50} , 139,640 bp). The genome sequence of *B. licheniformis* ATCC 14580 was used as a reference to order the VSD4 contigs in Mauve (6, 7). Genome annotation was completed using RAST (8, 9) and NCBI's PGAP (10). The genome of *B. licheniformis* VSD4 has a G+C content of 46.2% and includes 3,674 coding genes, 719 pseudogenes, 34 rRNAs (55, 165, 23S), 83 tRNAs, and five ncRNAs.

Enzymes related to the *Pseudomonas putida* GPo1 *n*-alkane degradation pathway (11) are encoded in *B. licheniformis* VSD4's genome, including homologues of AlkH (aldehyde dehydrogenase), AlkK (acyl-CoA synthetase), AlkT (rubredoxin-NAD⁺ reductase), and AlkN (methyl-accepting chemotaxis protein). Further, homologous enzymes of *P.*

Received 9 January 2017 Accepted 12 January 2017 Published 16 March 2017

Citation Stevens V, Thijs S, McAmmond B, Langill T, Van Hamme J, Weyens N, Vangronsveld J. 2017. Draft genome sequence of *Bacillus licheniformis* VSD4, a diesel fuel– degrading and plant growth–promoting phyllospheric bacterium. Genome Announc 5:e00027-17. https://doi.org/10.1128/ genomeA.00027-17.

Copyright © 2017 Stevens et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Jaco Vangronsveld, jaco.vangronsveld@uhasselt.be. *putida* G7's naphthalene degradation pathway (12) were located, including all four subunits of naphthalene dioxygenase (NahA). Genes associated with plant growth–promoting characteristics were also present: indole-3-acetic acid, acetoin, and siderophore production. *B. licheniformis* VSD4 is being further evaluated as an inoculant to enhance phylloremediation of environments contaminated with diesel fuel–associated air pollutants.

Accession number(s). This whole-genome sequencing project has been deposited in GenBank under the accession number MLKN00000000. The version described in this paper is the second version, MLKN00000000.2.

ACKNOWLEDGMENT

This work was supported by the Hasselt University Methusalem project 08M03VGRJ.

REFERENCES

- Azmatunnisa M, Rahul K, Subhash Y, Sasikala Ch, Ramana ChV. 2015. Bacillus oleivorans sp. nov., a diesel oil-degrading and solvent-tolerant bacterium. Int J Syst Evol Microbiol 65:1310–1315. https://doi.org/ 10.1099/ijs.0.000103.
- Purwanti IF, Abdullah SRS, Hamzah A, Idris M, Basri H, Mukhlisin M, Latif MT. 2015. Biodegradation of diesel by bacteria isolated from *Scirpus mucronatus* rhizosphere in diesel-contaminated sand. Adv Sci Lett 21: 140–143. https://doi.org/10.1166/asl.2015.5843.
- Asari S, Matzén S, Petersen MA, Bejai S, Meijer J. 2016. Multiple effects of Bacillus amyloliquefaciens volatile compounds: plant growth promotion and growth inhibition of phytopathogens. FEMS Microbiol Ecol 92: fiw070. https://doi.org/10.1093/femsec/fiw070.
- Goswami D, Dhandhukia P, Patel P, Thakker JN. 2014. Screening of PGPR from saline desert of Kutch: growth promotion in *Arachis hypogea* by *Bacillus licheniformis* A2. Microbiol Res 169:66–75. https://doi.org/ 10.1016/j.micres.2013.07.004.
- Chevreux B, Pfisterer T, Drescher B, Driesel AJ, Müller WE, Wetter T, Suhai S. 2004. Using the miraEST assembler for reliable and automated mRNA transcript assembly and SNP detection in sequenced ESTs. Genome Res 14:1147–1159. https://doi.org/10.1101/gr.1917404.
- Darling AE, Mau B, Perna NT. 2010. progressiveMauve: multiple genome alignment with gene gain, loss and rearrangement. PLoS One 5:e11147. https://doi.org/10.1371/journal.pone.0011147.
- 7. Rissman AI, Mau B, Biehl BS, Darling AE, Glasner JD, Perna NT. 2009.

Reordering contigs of draft genomes using the Mauve aligner. Bioinformatics 25:2071–2073. https://doi.org/10.1093/bioinformatics/btp356.

- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: rapid annotations using subsystems technology. BMC Genomics 9:75. https://doi.org/10.1186/1471-2164-9-75.
- Overbeek R, Olson R, Pusch GD, Olsen GJ, Davis JJ, Disz T, Edwards RA, Gerdes S, Parrello B, Shukla M, Vonstein V, Wattam AR, Xia F, Stevens R. 2014. The SEED and the rapid annotation of microbial genomes using subsystems technology (RAST). Nucleic Acids Res 42:D206–D214. https://doi.org/10.1093/nar/gkt1226.
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI prokaryotic genome annotation pipeline. Nucleic Acids Res 44: 6614–6624. https://doi.org/10.1093/nar/gkw569.
- van Beilen JB, Panke S, Lucchini S, Franchini AG, Röthlisberger M, Witholt B. 2001. Analysis of *Pseudomonas putida* alkane-degradation gene clusters and flanking insertion sequences: evolution and regulation of the *alk* genes. Microbiology 147:1621–1630. https://doi.org/10.1099/00221287-147-6-1621.
- Suenaga H, Koyama Y, Miyakoshi M, Miyazaki R, Yano H, Sota M, Ohtsubo Y, Tsuda M, Miyazaki K. 2009. Novel organization of aromatic degradation pathway genes in a microbial community as revealed by metagenomic analysis. ISME J 3:1335–1348. https://doi.org/10.1038/ismej.2009.76.