Research Article

Genomic Analysis of Abnormal DNAM Methylation in Parathyroid Tumors

Qing Li[®], Yonghao Li[®], Ximei Sun[®], Xinlei Zhang[®], and Mei Zhang[®]

Department of General Surgery,

The First Affiliated Hospital of Shandong First Medical University & Shandong Provincial Qianfoshan Hospital, No 16766 Jingshi Road, Jinan, Shandong, China

Correspondence should be addressed to Mei Zhang; baichuanldd@163.com

Received 9 December 2021; Revised 20 May 2022; Accepted 17 June 2022; Published 16 July 2022

Academic Editor: Dario de Biase

Copyright © 2022 Qing Li et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Background. Parathyroid tumors are common endocrine neoplasias associated with primary hyperparathyroidism. Although numerous studies have studied the subject, the predictive value of gene biomarkers nevertheless remains low. *Methods.* In this study, we performed genomic analysis of abnormal DNA methylation in parathyroid tumors. After data preprocessing, differentially methylated genes were extracted from patients with parathyroid tumors by using *t*-tests. *Results.* After refinement of the basic differential methylation, 28241 unique CpGs (634 genes) were identified to be methylated. The methylated genes were primarily involved in 7 GO terms, and the top 3 terms were associated with cyst morphogenesis, ion transport, and GTPase signal. Following pathway enrichment analyses, a total of 10 significant pathways were enriched; notably, the top 3 pathways were cholinergic synapses, glutamatergic synapses, and oxytocin signaling pathways. Based on PPIN and ego-net analysis, 67 ego genes were found which could completely separate the diseased group from the normal group. The 10 most prominent genes included POLA1, FAM155 B, AMMECR1, THOC2, CCND1, CLDN11, IDS, TST, RBPJ, and GNA11. SVM analysis confirmed that this grouping approach was precise. *Conclusions.* This research provides useful data to further explore novel genes and pathways as therapeutic targets for parathyroid tumors.

1. Introduction

Parathyroid cancer is a common endocrine disease characterized by excessive secretion of the parathyroid hormone [1]. The molecular pathogenesis of parathyroid tumors has already been partly elucidated; Heppner and Carling have reported that inactivating somatic mutations of tumor suppressor genes multiple endocrine neoplasia type 1 (MEN1), RET, and HRPT2/CDC73 have been identified in parathyroid tumors [2, 3]. Loss of menin, the protein encoded by the oncosuppressor gene MEN1, is characterized by a genetic background of parathyroid tumors [4]. This is called MEN1 syndrome [4].

It has been demonstrated that epigenetic modifications are not only involved in embryogenesis but also in cell fate reprogramming [5–7]. As epigenetic modifications are present in all human cancers, they cooperate with genetic alterations to drive given cancer phenotypes [8]. Furthermore, the abnormal methylation of cytosine phosphate guanine (CpG) in gene-promoter islands has been investigated in MEN1 syndrome cancer [9]. In benign and malignant parathyroid tumors, despite the clear usefulness in performing large-scale DNA methylation analysis, few studies have been published that describe abnormal global methylation on CpG gene-promoter islands. Global promoter methylation by LINE-1 does not differ from levels detected in normal glands. This differs from most other cancers, which display global DNA hypomethylation [10, 11].

Different CpG sites located near promoter regions of more than 14,000 genes have been screened by Starker and Collaborators, using a model composed of normal, benign, and malignant parathyroid tissues (3 normal parathyroid tissues, 14 adenomas, and 7 carcinomas) [12]. A gradient of CpG hypermethylation levels, ranging from normal tissues to adenomas and carcinomas, was identified in a subset of the genes [12]. These genes were involved in key pathways linked to parathyroid tumors. Specific genes such as RIZ1, APC, RASSF1A, CDKN2A/p16 and CDKN2B/p15, RB1, WT1, GATA4, PYCARD, SFRP1, SFRP2, and SFRP4 were found to be hypermethylated; this was in line with the hypothesis that cell cycle, transcription, and WNT pathways would represent biological processes that were deranged [12]. They would, in turn, be found in the onset of parathyroid neoplasms [12].

It is important to note that gene markers based solely on expression are still not reliable [13]. However, using cutting-edge computational methods and genomics data, significant markers can be identified. This can be done by integrating gene expression profiles with protein-protein interaction maps. Thus, in this study, we developed a workflow to identify significant methylation of genes that were functionally associated with diseases. We did this so that feature selection could maximize prediction performance [14].

2. Methods

2.1. Collection of DNA Methylation Data. DNA methylation data for parathyroid tumors (accession no. GSE64412) [15] were accessed from the Gene Expression Omnibus database (GEO, http://www.ncbi.nlm.nih.gov/gds), in the National Center for Biotechnology Information. The microarray data used the GPL11154 platform (Illumina HiSeq 2000 *Homo sapiens*; Illumina Inc., San Diego, CA, USA). GSE64412 included 38 samples: 13 sporadic (non-MEN1) parathyroid adenomas, 12 MEN1-parathyroid tumors, 4 parathyroid carcinomas, and 9 normal parathyroid tissues.

2.2. Data Preprocessing and Identification of Differentially Methylated Genes. A methylation-identification algorithm found in Genelibs (http://www.genelibs.com) was utilized. Raw microarray data containing 1954997 CpGs sites were filtered down in order to better focus our study. Sites were eliminated when they met the following criteria: (i) distance from CpG to single-nucleotide polymorphism (SNP) was less than or equal to 2; (ii) minor allele frequency (MAF) was less than 0.05; (iii) cross-hybridized probes were found, or on sex chromosomes. 1925786 CpGs were kept for further study. The DNA methylation microarray data were then processed using the Lumi package (bioconductor.org/ packages/release/bioc/html/lumi) [16, 17]. Data were normalized via the β -mixture quantile normalization method [18].

In this study, β values have been represented graphically, both the diseased group and the normal group. The percentage of methylation used the following formula: methylated/(methylated + unmethylated); results range between 0 and 1, where zero represented fully unmethylated genes and one represented fully methylated. Subsequently, the absolute value of the difference in mean β values was calculated, termed A. *T*-tests were then employed to identify the differentially methylated CpGs and identified at the threshold of P < 0.05 and A > 0.01.

In order to decrease the number of nonvariable sites, further filtering steps were performed. All sites with methylation scores were >50; the differential expression p value was <0.001, and the absolute value of >3 remained. Only CpGs termed A with a value of \geq 17 (and corrected p value <0.05) were applied. This was done in order to detect substantial methylation differences.

2.3. Hierarchical Clustering Analysis. Hierarchical clustering is an analytical tool applied to discover the closest associations that exist between gene profiles and specimens [19]. Generally, cancers have similar methylation profiles that tend to cluster together. To analyze whether these differentially methylated CpGs would segregate into two distinct clusters, unsupervised hierarchical clustering was conducted using Euclidian distance and average linkage criteria [20]. The matrix of mean β -value levels was then formed between the parathyroidoma and normal samples.

2.4. Gene Ontology (GO) Analysis of Differentially Methylated Genes. Gene ontology (GO) was applied to analyze the main function of differentially expressed genes (DEGs), the key functional classification used by the National Center of Biotechnology Information (NCBI). In the present study, Fisher's exact test was used to ascertain the GO category; furthermore, P values were corrected using the false discovery rate (FDR) along with the Benjamini & Hochberg method [21]. Functional terms with an FDR <0.01 and gene count >10 were considered to be statistically significant.

2.5. Pathway Enrichment Analysis of Differentially Methylated Genes. Pathway analysis was used to find significant pathways of the DEGs according to the Kyoto Encyclopedia of Genes and Genomes (KEGG) [22]. In the present study, Fisher's exact test was used to extract significant pathways, and the threshold of significance was defined by FDR. Significant pathways were selected according to the thresholds of FDR <0.01 and gene count >10.

2.6. Identification of the Ego Genes in the Methylation Correlation Network. The Search Tool for the Retrieval of Interacting Genes (STRING) database (http://www.bork. embl-heidelberg.de/STRING) is a global resource used for analyzing gene-gene interaction [23]. In the present study, STRING was utilized to identify the correlations between the differentially methylated genes.

First, a network of all human gene interactions was obtained from the STRING database; a special subnetwork that contained differentially methylated genes and parathyroid adenoma genes (CDC73, MEN1, CCND1, RET) was then used for further analysis. The absolute value of the Pearson coefficient was calculated: interaction pairs with a p value of 0.05 defined the difference expression network [24]. Finally, the weight value of interaction in the network was calculated using the following formula (EgoNet algorithm) [25]:

$$\omega_{ij} = \frac{\sqrt{\log p_i + \log p_j}}{\sqrt{2^* \max_{i \in V} |\log p_i|}} (\operatorname{ifcor}(i, j) \ge \delta) \omega_{ij},$$

$$= 0 (if \operatorname{cor}(i, j) < \delta),$$
(1)

where V represents the node set in the network.

The topological analysis of the differential expression network was measured by four indicators: degree, closeness, betweenness, and transitivity (a measure of the clustering coefficient). In order to identify the methylation markers, the context likelihood of relatedness (CLR) was applied to the algorithm to square weight and then to form an adjacency matrix. Genes were sorted in descending order based on the importance of genes in the netweb (g (i) = z-score).

2.7. Support Vector Machine Analysis. The support vector machine (SVM) is a machine learning method commonly used in pattern recognition for binary classification [26]. In this study, SVM was used to test whether ego genes could completely separate the diseased group from the normal group and to further verify the feasibility of our analysis methods.

The normal group and the diseased group were together termed the total population. In accordance with the ratio of 6:4, the total population was randomly divided into an experimental group and a testing group. SVM C-classification was then performed using linear kernel and 5-fold cross-validation. The experimental group served as the basis for classification, and the testing group as the basis for regression [27].

3. Results

3.1. Identification of Differentially Methylated Genes. Following quality control and normalization to (i) remove probes with an SNP-CpG distance of ≤ 2 , (ii) remove probes with a minimum allelic frequency of <0.05, and (iii) demonstrate cross-hybridization, a total of 1925786 methylated CpGs remained. A volcano plot exhibiting the distribution of the 1925786 methylated CpGs is presented in Figure 1. Among these 1925786 methylated CpGs, 192613 of them (representing 26926 genes) were differentially methylated, where the absolute value of the mean β -value was >0.05, and the *P* value <0.05. A total of 192340 of the CpGs were hypermethylated, and 273 CpGs were hypomethylated in the disease group.

Subsequently, these 192613 methylated CpGs were subjected to further filtering. 31207 unique CpGs (covering 13798 genes) met the following conditions: [1] a methylation score >50; [2] differential expression *P* value < 0.001, and [3] absolute value > 3. The only CpGs with the absolute value of the mean β -value difference were \geq 17, with the corrected *P* value <0.05.28241. CpGs (covering 634 genes) were selected for further analysis. According to various studies, CDC73, MEN1, CCND1, and RET were shown to be parathyroid adenoma susceptible genes [28–31]. Their *P* values are demonstrated in Table 1.



FIGURE 1: Volcano plot exhibiting the methylation data of 29 parathyroid tumors and 9 normal samples. X-axis represents the mean methylation difference between parathyroid tumor and normal samples. Y-axis represents the log-transformed *P* values. A total of 192613 CpG sites were considered to be significantly differentially methylated, indicated in blue.

TABLE 1: P values of susceptible parathyroid adenoma genes.

| | | - |
|---------|-------------|--------|
| ID | p value | Symbol |
| 1173651 | 0.003161659 | RET |
| 1173667 | 0.003246324 | RET |
| 1173680 | 7.16E-05 | RET |
| 1173684 | 0.000342777 | RET |
| 1173722 | 0.002885583 | RET |
| 1173736 | 0.002602326 | RET |
| 1173749 | 0.001025146 | RET |
| 1173756 | 0.000703971 | RET |
| 1173773 | 0.000353432 | RET |
| 1301532 | 0.001475949 | MEN1 |
| 1312429 | 0.004835513 | CCND1 |
| 1312477 | 0.003523037 | CCND1 |
| 150982 | 0.001799107 | CDC73 |
| | | |

3.2. Hierarchical Clustering Analysis. To further explore the changes in the methylation levels, cluster analysis was conducted. The cluster heat map is demonstrated in Figure 2. In this figure, it is observed that there were distinctive methylation patterns in the parathyroidoma and normal samples, further segregating the samples into two distinct groups.

3.3. GO Enrichment. To determine the primary functions of the differentially methylated genes, a comprehensive gene ontology (GO) analysis was performed. As shown in Figure 3, 7 signaling pathways were significantly enriched by the differentially methylated genes, including regulation of small

International Journal of Endocrinology



FIGURE 3: Differentially methylated genes identified between parathyroidoma and normal samples were functionally classified via gene ontology analysis. The most enriched terms, which satisfied the criteria of FDR <0.01 and gene count >10, are represented.

GTPase mediated signal transduction, regulation of cell morphogenesis, calcium ion transmembrane transport, glutamate receptor signaling pathway, regulation of cell morphogenesis involved in differentiation, regulation of ion transmembrane transport, and ERBB signaling pathway. These GO terms were sorted in ascending order based on FDR value. Overall, the GO terms were mainly involved in cell morphogenesis, ion transport, and GTPase signal.

3.4. KEGG Pathway Analysis. Pathway enrichment analysis of the differentially methylated genes was conducted given the KEGG pathway database. This analysis yielded 48 pathways. Table 2 lists the top 10 pathways. These pathways were sorted in ascending order based on FDR value. The top 3 significantly enriched pathways were cholinergic synapses, glutamatergic synapses, and the oxytocin signaling pathway. The oxytocin signaling pathway was associated with CCND1.

3.5. Ego Analysis. Given the goal of analyzing the association between the differentially methylated genes, STRING software was used to establish the PPI network (PPIN). A network of all human gene interactions containing 787896 interaction pairs (16730 genes) was obtained from STRING. A total of the 793 interaction pairs (the target-PPI) were extracted from the PPIN. The target-PPI included differentially methylated genes as well as parathyroid adenoma susceptible genes (CDC73, MEN1, CCND1, RET). This covered 299 genes. The weight value of the interaction in the network was calculated, and genes were sorted in descending order based on topological degree. The top 30% were selected as ego genes. A total of 67 ego genes were obtained (Figure 4). The differential expression of the 67 genes ensures complete separation between the normal and parathyroid tumor populations. Genes with higher z-scores were given greater importance. The top 10 of the 67 ego genes are shown in Table 3.



FIGURE 2: Hierarchical clustering analysis of significantly differentially methylated CpGs between parathyroidoma and normal samples. DNA methylation across the 28241 sites in each of the samples was analyzed by this method. Each row is an individual CpG site and each column is a different sample. The color gradation from green to red denotes low to high DNA methylation, with β -values ranging from 0 (no methylation; green) to 1 (complete methylation; red).

| TABLE 2: The to | p 10 key pathway | ys according to | KEGG analysis |
|-----------------|------------------|-----------------|---------------|
|-----------------|------------------|-----------------|---------------|

| Pathway name | P value |
|---|-------------|
| 4725 Cholinergic synapse [PATH:hsa04725] | 3.27E-07 |
| 4724 Glutamatergic synapse [PATH:hsa04724] | 4.63E-07 |
| 4921 Oxytocin signaling pathway [PATH:hsa04921] | 5.67E-06 |
| 4261 Adrenergic signaling in cardiomyocytes [PATH:hsa04261] | 4.82E-05 |
| 4911 Insulin secretion [PATH:hsa04911] | 0.000101911 |
| 4713 Circadian entrainment [PATH:hsa04713] | 0.0002795 |
| 4020 Calcium signaling pathway [PATH:hsa04020] | 0.000363009 |
| 4912 GnRH signaling pathway [PATH:hsa04912] | 0.000829401 |
| 4730 Long-term depression [PATH:hsa04730] | 0.00111826 |
| 5205 Proteoglycans in cancer [PATH:hsa05205] | 0.001260031 |



FIGURE 4: Protein-protein interaction network constructed from differentially methylated genes. Nodes colored in orange are gene-associated ego genes. Yellow nodes represent parathyroid adenoma susceptibility genes. It is paramount to explicate that CCDN 1 and RET are parathyroid adenoma susceptibility genes as well ego genes.

| Node | z-Score |
|---------|-------------|
| POLA1 | 219.1896777 |
| FAM155B | 202.6303015 |
| AMMECR1 | 161.7614615 |
| THOC2 | 135.7028647 |
| CCND1 | 132.8239023 |
| CLDN11 | 128.873499 |
| IDS | 126.8873715 |
| TST | 125.1721484 |
| RBPJ | 117.4724126 |
| GNA11 | 115.4442218 |

3.6. Support Vector Machine Analysis. The use of machine learning in medical diagnosis is gaining currency. This is mainly because the effectiveness of classification and recognition systems has significantly improved, helping medical experts in diagnosing diseases [32, 33]. In this paper, C-classification was performed using linear kernel and 5-fold cross-validation. Assessment indicators were AUC (the area under receiver operating characteristic curve), accuracy (classification accuracy), MCC (Mathews' correlation coefficient), specificity (TNR, true-negative rates), and sensitivity (TPR, true-positive rates). The results of the analysis are as follows: AUC = 0.9123, accuracy = 99.29%, MCC = 0.8549, specificity = 83%, and sensitivity = 92%. Thus, the methods used in this study could completely separate the diseased group from the normal one.

4. Discussion

Analysis of DNA methylation data has been widely used to identify abnormally methylated genes associated with tumors. It has also enabled the extraction of targets for therapeutic strategies. In this study, the pathogenesis of parathyroid tumors was analyzed by means of bioinformatics. This included the detection of differentially methylated genes, gene ontology (GO), pathway enrichment, PPI construction, and ego genes identification. Potential mechanisms of parathyroid tumors have been thus revealed, providing novel insights into parathyroid diagnosis and therapy.

In general, the genetic background of parathyroid tumors is characterized by the loss of two oncosuppressor genes. First, a loss of menin, the protein encoded by the MEN1 gene, takes place. This occurs in the parathyroid lesions found in multiple endocrine neoplasia type 1 (MEN1), [34]. Second is a loss of parafibromin, encoded by the CDC73/HRPT2 gene. This occurs in hyperparathyroidism-jaw tumors (HPT-JT) [35].

The heterogeneity of parathyroid tumors' biological and clinical presentation may be due to epigenetic deregulation [36, 37]. As mentioned, epigenetic modifications at DNA and chromatin levels are present in all human cancers [38–40]. Epigenetic regulation is achieved through multiple regulatory pathways unifying sequence-specific, DNAbinding transcription factors, ATP-dependent, nucleosome remodeling, and long, noncoding RNA and DNA methylation [41-44].

Our model aimed at finding the diseased genes by means of the following underlying assumption: if the majority of neighbors of a central disease gene are also disease genes, then its other neighbors are likely to be involved in the disease pathway. This model's benefit is that it can find hidden genes that would otherwise show no significance by themselves but are nevertheless clustered in a subnetwork module. It is in this subnetwork where collectively the genes are highly predictive of disease status. Furthermore, machine-learning techniques can be used to assess the association between ego-networks and clinical outcomes.

Based on pathway analyses, pathways of cholinergic synapses, glutamatergic synapses, and oxytocin signaling were considered the most important. The oxytocin signaling pathway in particular was associated with CCND1. On the other hand, cholinergic and glutamatergic synapses were closely related to the regulation of nerve excitement and inhibition. In GO analysis, the differentially methylated genes were enriched in 7 GO terms, which were mainly involved in cell morphogenesis, ion transport, and GTPase signal. Recent studies suggest that there is a cross-regulation between morphogenesis and EMT processes. Aberrant activation of these coregulators can drive different stages of cancer progression, including tumor invasion, cell spread, and metastatic colonization and growth [45] The ion (such as $\mathrm{Ca}^{2+},\,\mathrm{K}^{+},\,\mathrm{Fe}^{3+},\,\mathrm{and}\,\,\mathrm{Cl}^{-})$ and channel proteins play important roles in tumor progression. These membrane ion transport systems have great potential as diagnostic biomarkers and therapeutic targets in tumor treatment [46-48]. GTPase-related signaling pathways are widely abnormally activated in tumors and are involved in the regulation of tumor cell growth, metastasis, and drug resistance [49]. However, there are few studies on the relationship between these three cell functions and the tumorigenesis of parathyroid tumors, suggesting that further research in this area should be conducted in the future.

Following the implementation of the PPIN and ego-net, 67 ego genes were found, which completely separates normal from parathyroid tumor populations. The top 10 of the 67 ego genes were POLA1, FAM155 B, AMMECR1, THOC2, CCND1, CLDN11, IDS, TST, RBPJ, and GNA11. At present, there have been studies conducted on CCND1, but little research on the other genes. It is reported that AMMECR1 is associated with growth, bone, and heart alterations; CLDN11 is an epigenetic biomarker for malignancy, and THOC2 mutations are related to X-linked intellectual disability [50-52]. POLA1 is the catalytic subunit of DNA polymerase and plays an essential role in the initiation of DNA replication [53]. FAM155B is predicted to be a multipass membrane protein with unknown function. IDS is involved in the lysosomal degradation of heparan sulfate and dermatan sulfate [54]. TST is mainly localized to mitochondria and catalyzes the conversion of thiosulfate and cyanide to thiocyanate and sulfite [55]. As molecular switches in regulating the notch response, transcription factor RBPJ involves in the regulation of the progression of a variety of tumors, such as colorectal cancer [56], lung cancer [57], hepatocellular carcinoma [58], glioblastoma [59, 60], and prostate cancer [61]. As a member of the G protein family, *GNA11* plays an important role in tumor progression [62]. Research shows that CCND1 and POLA1 are potential targets of miR-206 and may share a common regulatory network [63]. The interrelationship between these genes has not been studied.

Global promoter methylation by CCND1 often displays differentially expressed DNA hypomethylation, especially when compared to normal tissue [64]. Different CpG sites located near promoter regions of more than 14,000 predicted genes were screened. Both CCND1 and CCND2 have been previously reported as deregulated in many cancers [65, 66]. Furthermore, they represent an alternative marker to study epigenetic modifications [66]. The reversibility of these changes makes them prime targets for therapeutic manipulations. A number of small molecules targeting chromatin-based mechanisms are currently being tested in clinical trials [67]. However, the results of this study were based on a limited clinical sample, which limited the accuracy of the data. The role of the above biomarkers still needed to be verified with more samples. In addition, we did not analyze the difference of MEN1 in adenomas, carcinomas, or sporadic tumors due to the limitation of sample size. Importantly, our conclusions still require experimental confirmation, not just bioinformatics data analysis. The expression of MEN1 in subgroups of parathyroid tumors is still worthy of further analysis.

In conclusion, we have evaluated the performance of EgoNet in human protein-protein interaction networks. This method has not only successfully identified CCND1 from significant ego-networks but has also detected several novel targets, such as POLA1, FAM155 B, AMMECR1, and THOC2. We expect that EgoNet can be widely used to infer novel biomarkers for phenotypic prediction of many human diseases.

Abbreviations

| MEN1: | Multiple endocrine neoplasia type 1 |
|-----------|--|
| CpG: | Cytosine phosphate guanine |
| GEO: | Gene expression omnibus |
| SNP: | Single-nucleotide polymorphism |
| MAF: | Minor allele frequency |
| GO: | Gene ontology |
| DEGs: | Differentially expressed genes |
| NCBI: | National Center of Biotechnology Information |
| FDR: | False discovery rate |
| KEGG: | Kyoto Encyclopedia of Genes and Genomes |
| STRING: | Search Tool for the Retrieval of Interacting Genes |
| CLR: | Context likelihood of relatedness |
| SVM: | Support vector machine |
| GO: | Gene ontology |
| PPIN: | PPI network |
| AUC: | The area under receiver operating characteristic |
| | curve |
| Accuracy: | Classification accuracy |
| MCC: | Mathews' correlation coefficient |
| TND. | Truce acaptize actor |

TNR: True-negative rates

| TPR: | True-positive rates |
|---------|---------------------------------|
| HPT-JT: | Hyperparathyroidism-jaw tumors. |

Data Availability

The data supporting the conclusions of this paper are included within the manuscript.

Ethical Approval

This article does not contain any studies with human participants or animals performed by any of the authors.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

Authors' Contributions

Qing Li and Yonghao Li contributed equally to this research.

Acknowledgments

This study was supported by the Natural Science Funds of Shandong Province (Project: Mutation and Expression of Parathyroid Carcinoma Susceptibility Gene HRPT2, MEN1, CyclinD1, and RET) (grant no. ZR2012HL04) and the National Natural Science Foundation for Cultivation in Shandong Provincial Qianfoshan Hospital, The First Hospital Affiliated with Shandong First Medical University (grant no.QYPY2019NSFC0809), 2021 Shandong Medical Association Clinical Research Fund-Qilu Special Project (grant no. YXH2022ZX02167), and General Program of Natural Science Foundation of Shandong Province (grant no. ZR2021MH325).

References

- H. Siilin, E. Lundgren, H. Mallmin et al., "Prevalence of primary hyperparathyroidism and impact on bone mineral density in elderly men: MrOs Sweden," *World Journal of Surgery*, vol. 35, no. 6, pp. 1266–1272, 2011.
- [2] C. Heppner, M. B. Kester, S. K. Agarwal et al., "Somatic mutation of the MEN1 gene in parathyroid tumours," *Nature Genetics*, vol. 16, pp. 375–378, 1997.
- [3] T. Carling, J. Rastad, G. Akerström, and G. Westin, "Vitamin D receptor (VDR) and parathyroid hormone messenger ribonucleic acid levels correspond to polymorphic VDR alleles in human parathyroid tumors," *Journal of Clinical Endocrinology and Metabolism*, vol. 83, no. 7, pp. 2255–2259, 1998.
- [4] Y. Jiao, C. Shi, B. H. Edil et al., "DAXX/ATRX, MEN1, and mTOR pathway genes are frequently altered in pancreatic neuroendocrine tumors," *Science*, vol. 331, no. 6021, pp. 1199–1203, 2011.
- [5] J. L. R. Lorenzo, R. Hobza, and B. Vyskot, "Epigenetic control of reproductive development," *Plant Systematics and Evolution*, vol. 303, no. 8, pp. 991–1000, 2017.
- [6] K. Takahashi, "Cellular reprogramming lowering gravity on Waddington's epigenetic landscape," *Journal of Cell Science*, vol. 125, no. Pt 11, pp. 2553–2560, 2012.

- [7] B. Nashun, P. W. S. Hill, and P. Hajkova, "Reprogramming of cell fate: epigenetic memory and the erasure of memories past," *The EMBO Journal*, vol. 34, no. 10, pp. 1296–1308, 2015.
- [8] S. B. Baylin and P. A. Jones, "Epigenetic determinants of cancer," *Cold Spring Harbor Perspectives in Biology*, vol. 8, no. 9, Article ID a019505, 2016.
- [9] D. Lindberg, G. Akerström, and G. Westin, "Evaluation of CDKN2C/p18, CDKN1B/p27 and CDKN2B/p15 mRNA expression, and CpG methylation status in sporadic and MEN1-associated pancreatic endocrine tumours," *Clinical Endocrinology*, vol. 68, no. 2, pp. 271–277, 2008.
- [10] N. Kitkumthorn and A. Mutirangura, "Long interspersed nuclear element-1 hypomethylation in cancer: biology and clinical applications," *Clinical Epigenetics*, vol. 2, no. 2, pp. 315–330, 2011.
- [11] N. Kitkumthorn, T. Tuangsintanakul, P. Rattanatanyong, D. Tiwawech, and A. Mutirangura, "LINE-1 methylation in the peripheral blood mononuclear cells of cancer patients," *Clinica Chimica Acta*, vol. 413, no. 9-10, pp. 869–874, 2012.
- [12] L. F. Starker, J. Svedlund, R. Udelsman et al., "The DNA methylome of benign and malignant parathyroid tumors," *Genes, Chromosomes and Cancer*, vol. 50, no. 9, pp. 735–745, 2011.
- [13] D. Venet, J. E. Dumont, and V. Detours, "Most random gene expression signatures are significantly associated with breast cancer outcome," *PLoS Computational Biology*, vol. 7, no. 10, Article ID e1002240, 2011.
- [14] X. Wang, N. Gulbahce, and H. Yu, "Network-based methods for human disease gene prediction," *Briefings in functional* genomics, vol. 10, no. 5, pp. 280–293, 2011.
- [15] Z. Yuan, C. S. Claros, M. Suzuki et al., "Loss of MEN1 activates DNMT1 implicating DNA hypermethylation as a driver of MEN1 tumorigenesis," *Oncotarget*, vol. 7, no. 11, pp. 12633–12650, 2016.
- [16] P. Du, W. A. Kibbe, and S. M. Lin, "lumi: a pipeline for processing Illumina microarray," *Bioinformatics*, vol. 24, no. 13, pp. 1547-1548, 2008.
- [17] P. Du, X. Zhang, C. C. Huang et al., "Comparison of Betavalue and M-value methods for quantifying methylation levels by microarray analysis," *BMC Bioinformatics*, vol. 11, no. 1, p. 587, 2010.
- [18] A. E. Teschendorff, F. Marabita, M. Lechner et al., "A betamixture quantile normalization method for correcting probe design bias in Illumina Infinium 450 k DNA methylation data," *Bioinformatics*, vol. 29, no. 2, pp. 189–196, 2013.
- [19] H. A. Boateng, "Mesh-free hierarchical clustering methods for fast evaluation of electrostatic interactions of point multipoles," *The Journal of Chemical Physics*, vol. 147, no. 16, Article ID 164104, 2017.
- [20] A. Sturn, J. Quackenbush, and Z. Trajanoski, "Genesis: cluster analysis of microarray data," *Bioinformatics*, vol. 18, no. 1, pp. 207-208, 2002.
- [21] Y. Benjamini, D. Drai, G. Elmer, N. Kafkafi, and I. Golani, "Controlling the false discovery rate in behavior genetics research," *Behavioural Brain Research*, vol. 125, no. 1-2, pp. 279–284, 2001.
- [22] J. Zhu and X. Yao, "Use of DNA methylation for cancer detection: promises and challenges," *The International Journal* of Biochemistry & Cell Biology, vol. 41, no. 1, pp. 147–154, 2009.
- [23] C. von Mering, M. Huynen, D. Jaeggi, S. Schmidt, P. Bork, and B. Snel, "STRING: a database of predicted functional associations between proteins," *Nucleic Acids Research*, vol. 31, no. 1, pp. 258–261, 2003.

- [24] T. Ahn, E. Lee, N. Huh, and T. Park, "Personalized identification of altered pathways in cancer using accumulated normal tissue data," *Bioinformatics*, vol. 30, no. 17, pp. i422–9, 2014.
- [25] R. Yang, Y. Bai, Z. Qin, and T. Yu, "EgoNet: identification of human disease ego-network modules," *BMC Genomics*, vol. 15, no. 1, p. 314, 2014.
- [26] W. S. Noble, "What is a support vector machine?" Nature Biotechnology, vol. 24, no. 12, pp. 1565–1567, 2006.
- [27] B. Zhou, C. Yang, H. Guo, and J. Hu, A Quasi-Linear SVM Combined with Assembled SMOTE for Imbalanced Data Classification, International Joint Conference on Neural Networks, Dallas, USA, 2013.
- [28] J. M. Sharretts and W. F. Simonds, "Clinical and molecular genetics of parathyroid neoplasms," *Best Practice & Research Clinical Endocrinology & Metabolism*, vol. 24, no. 3, pp. 491–502, 2010.
- [29] J. Costaguda and A. Arnold, "Genetic and epigenetic changes in sporadic endocrine tumors: parathyroid tumors," *Molecular and Cellular Endocrinology*, vol. 386, no. 1-2, pp. 46–54, 2014.
- [30] G. Masi, L. Barzon, M. Iacobone et al., "Clinical, genetic, and histopathologic investigation of CDC73-related familial hyperparathyroidism," *Endocrine-Related Cancer*, vol. 15, no. 4, pp. 1115–1126, 2008.
- [31] M. A. Hahn, V. M. Howell, A. J. Gill et al., "CDC73/HRPT2 CpG island hypermethylation and mutation of 5'-untranslated sequence are uncommon mechanisms of silencing parafibromin in parathyroid tumors," *Endocrine-Related Cancer*, vol. 17, no. 1, pp. 273–282, 2010.
- [32] K. Polat and S. Güneş, "Breast cancer diagnosis using least square support vector machine," *Digital Signal Processing*, vol. 17, no. 4, pp. 694–701, 2007.
- [33] Y. Lee and C. K. Lee, "Classification of multiple cancer types by multicategory support vector machines using gene expression data," *Bioinformatics*, vol. 19, no. 9, pp. 1132–1139, 2003.
- [34] F. Marini, A. Falchetti, E. Luzi, F. Tonelli, and B. Maria Luisa, Multiple Endocrine Neoplasia Type 1 (MEN1) Syndrome, 2009.
- [35] O. Gimm, P. Nguyen Thanh, M. Bloching et al., "Hyperparathyroidism-jaw tumor (HPT-JT)-syndrome: three case reports," *Experimental and Clinical Endocrinology & Diabetes*, vol. 113, no. S 1, 2005.
- [36] C. Verdelli, I. Forno, V. Vaira, and S. Corbetta, "Epigenetic alterations in human parathyroid tumors," *Endocrine*, vol. 49, no. 2, pp. 324–332, 2015.
- [37] J. Costa-Guda and A. Arnold, "Genetic and epigenetic changes in sporadic endocrine tumors: parathyroid tumors," *Molecular and Cellular Endocrinology*, vol. 386, no. 1-2, pp. 46–54, 2014.
- [38] M. Esteller and J. G. Herman, "Cancer as an epigenetic disease: DNA methylation and chromatin alterations in human tumours," *The Journal of Pathology*, vol. 196, no. 1, pp. 1–7, 2002.
- [39] L. Ellis, P. W. Atadja, and R. W. Johnstone, "Epigenetics in cancer: targeting chromatin modifications," *Molecular Cancer Therapeutics*, vol. 8, no. 6, pp. 1409–1420, 2009.
- [40] S. B. Baylin, M. Esteller, M. R. Rountree, K. E. Bachman, K. Schuebel, and J. G. Herman, "Aberrant patterns of DNA methylation, chromatin formation and gene expression in cancer," *Human Molecular Genetics*, vol. 10, no. 7, pp. 687–692, 2001.

- [41] P. A. Jones and D. Takai, "The role of DNA methylation in mammalian epigenetics," *Science*, vol. 293, no. 5532, pp. 1068–1070, 2001.
- [42] P. B. Becker and W. Hörz, "ATP-dependent nucleosome remodeling," *Annual Review of Biochemistry*, vol. 71, no. 1, pp. 247–273, 2002.
- [43] T. R. Mercer, M. E. Dinger, and J. S. Mattick, "Long noncoding RNAs: insights into functions," *Nature Reviews Genetics*, vol. 10, no. 3, pp. 155–159, 2009.
- [44] S. B. Baylin, "DNA methylation and gene silencing in cancer," *ChemInform*, vol. 37, p. S4, 2006.
- [45] C. C. Wang, "Metabolic stress adaptations underlie mammary gland morphogenesis and breast cancer progression," *Cells*, vol. 10, no. 10, p. 2641, 2021.
- [46] E. Bulk, L. M. Todesca, and A. Schwab, "Ion channels in lung cancer," *Reviews of Physiology, Biochemistry & Pharmacology*, vol. 181, pp. 57–79, 2021.
- [47] K. J. Anderson, R. T. Cormier, and P. M. Scott, "Role of ion channels in gastrointestinal cancer," *World Journal of Gastroenterology*, vol. 25, no. 38, pp. 5732–5772, 2019.
- [48] N. Prevarskaya, R. Skryma, and Y. Shuba, "Ion channels in cancer: are cancer hallmarks oncochannelopathies?" *Physiological Reviews*, vol. 98, no. 2, pp. 559–621, 2018.
- [49] E. Crosas-Molist, R. Samain, L. Kohlhammer et al., "Rho GTPase signaling in cancer progression and dissemination," *Physiological Reviews*, vol. 102, no. 1, pp. 455–510, 2022.
- [50] L. Gao, K. van den Hurk, P. T. M. Moerkerk et al., "Promoter CpG island hypermethylation in dysplastic nevus and melanoma: CLDN11 as an epigenetic biomarker for malignancy," *Journal of Investigative Dermatology*, vol. 134, no. 12, pp. 2957–2966, 2014.
- [51] M. Moysésoliveira, G. Giannuzzi, R. J. Fish, J. A. Rosenfeld, F. Petit, and M. F. Soares, *Inactivation of AMMECR1 Is Associated with Growth, Bone, and Heart Alterations*, Human Mutation, Wiley-Liss Inc., Hoboken, NJ, USA, 2017.
- [52] R. Kumar, M. A. Corbett, B. W. van Bon et al., "THOC2 mutations implicate mRNA-export pathway in X-linked intellectual disability," *The American Journal of Human Genetics*, vol. 97, no. 2, pp. 302–310, 2015.
- [53] C. Toukoki and I. Gryllos, "PolA1, a putative DNA polymerase I, is coexpressed with PerR and contributes to peroxide stress defenses of group A Streptococcus," *Journal of Bacteriology*, vol. 195, no. 4, pp. 717–725, 2013.
- [54] K. Matsuhisa and K. Imaizumi, "Loss of function of mutant IDS due to endoplasmic reticulum-associated degradation: new therapeutic opportunities for mucopolysaccharidosis type II," *International Journal of Molecular Sciences*, vol. 22, no. 22, Article ID 12227, 2021.
- [55] P. Singh, P. Rao, and R. Bhattacharya, "Dose and time-dependent effects of cyanide on thiosulfate sulfurtransferase, 3mercaptopyruvate sulfurtransferase, and cystathionine λ -lyase activities," *Journal of Biochemical and Molecular Toxicology*, vol. 27, no. 12, pp. 499–507, 2013.
- [56] G. Q. Chen, Z. M. Liao, J. Liu, F. Li, D. Huang, and Y. D. Zhou, "LncRNA FTX promotes colorectal cancer cells migration and invasion by miRNA-590-5p/RBPJ Axis," *Biochemical Genetics*, vol. 59, no. 2, pp. 560–573, 2021.
- [57] H. Onishi, S. Ichimiya, K. Yanai et al., "RBPJ and MAML3: potential therapeutic targets for small cell lung cancer," *Anticancer Research*, vol. 38, no. 8, pp. 4543–4547, 2018.
- [58] Y. C. Ye, J. L. Zhao, Y. T. Lu et al., "NOTCH signaling via WNT regulates the proliferation of alternative, CCR2-independent tumor-associated macrophages in hepatocellular

carcinoma," Cancer Research, vol. 79, no. 16, pp. 4160-4172, 2019.

- [59] B. D. Giaimo, E. K. Gagliani, R. A. Kovall, and T. Borggrefe, "Transcription factor RBPJ as a molecular switch in regulating the notch response," *Advances in Experimental Medicine and Biology*, vol. 1287, pp. 9–30, 2021.
- [60] G. Zhang, S. Tanaka, S. Jiapaer et al., "RBPJ contributes to the malignancy of glioblastoma and induction of proneuralmesenchymal transition via IL-6-STAT3 pathway," *Cancer Science*, vol. 111, no. 11, pp. 4166–4176, 2020.
- [61] L. Xue, H. Li, Q. Chen et al., "Inhibition of recombining binding protein suppressor of hairless (RBPJ) impairs the growth of prostate cancer," *Cellular Physiology and Biochemistry*, vol. 36, no. 5, pp. 1982–1990, 2015.
- [62] L. Larribère and J. Utikal, "Update on GNA alterations in cancer: implications for uveal melanoma treatment," *Cancers*, vol. 12, no. 6, p. 1524, 2020.
- [63] D. Mihály, G. Papp, Z. Mervai et al., "The oncomir face of microRNA-206: a permanent miR-206 transfection study," *Proceedings of The Society for Experimental Biology and Medicine*, vol. 243, no. 12, pp. 1014–1023, 2018.
- [64] S. Gomez, G. Castellano, G. Mayol et al., "DNA methylation fingerprint of neuroblastoma reveals new biological and clinical insights," *Epigenomics*, vol. 7, no. 7, pp. 1137–1153, 2015.
- [65] A. K. Eisfeld, J. Kohlschmidt, S. Schwind et al., "Mutations in the CCND1 and CCND2 genes are frequent events in adult patients with t(8;21)(q22;q22) acute myeloid leukemia," *Leukemia*, vol. 31, no. 6, pp. 1278–1285, 2017.
- [66] R. Buschges, R. G. Weber, B. Actor, P. Lichter, V. P. Collins, and G. Reifenberger, "Amplification and expression of cyclin D genes (CCND1, CCND2 and CCND3) in human malignant gliomas," *Brain Pathology*, vol. 9, no. 3, pp. 435–442, 1999.
- [67] M. T. Mccabe, J. C. Brandes, and P. M. Vertino, "Cancer DNA methylation: molecular mechanisms and clinical implications," *Clinical Cancer Research*, vol. 15, no. 12, pp. 3927–3937, 2009.