

Comprehensive evaluation of *SPATS2* expression and its prognostic potential in liver cancer

Jin Xing, PhD^a, Yijun Tian, PhD^b, Wu Ji, PhD^{a,*}, Xinying Wang, PhD^{a,*}

Abstract

Spermatogenesis associated serine rich 2 (*SPATS2*) has been reported to be dysregulated in few types of cancer; however, no reports have investigated *SPATS2* in liver cancer. The aim of the present study was to investigate *SPATS2* expression in liver cancer and to analyze its association with the prognosis of liver cancer patients.

We examined the differential expression of *SPATS2* in liver cancer by exploring The Cancer Genome Atlas (TCGA) database. The diagnostic efficiency of *SPATS2* was obtained by Receiver Operating Characteristic (ROC) curve. The Chi-Squared test was used to assess clinical relevance. Survival analysis and Cox regression model were used to detect the effect of *SPATS2* on the survival of liver cancer patients. Gene Set Enrichment Analysis (GSEA) was used to identify signaling pathways related to *SPATS2* expression.

SPATS2 is highly expressed in liver cancer ($P < 2.2 \times 10^{-16}$) and has the high diagnostic ability (AUC = 0.964). Survival analysis showed that patients with high *SPATS2* expression have an apparently shorter overall survival (OS, $P < .0001$) and relapse-free survival (RFS, $P < .0001$). Cox regression analysis showed that high *SPATS2* expression might be an independent risk factor for liver cancer (OS, HR = 2.41, $P = .000$; RFS, HR = 1.90, $P < .001$). GSEA analysis identified 3 signaling pathways (Mitotic spindle, G2M checkpoint, E2F targets) that were enriched in the presence of high *SPATS2* expression.

SPATS2 expression could be a novel diagnostic and prognostic biomarker in liver cancer.

Abbreviations: AUC = area under curve, FDR = false discovery rate, GSEA = Gene Set Enrichment Analysis, NES = normalized enrichment score, OS = overall survival, RFS = relapse-free survival, ROC = Receiver Operating Characteristic, *SPATS2* = Spermatogenesis associated serine rich 2, TCGA = The Cancer Genome Atlas.

Keywords: diagnosis, liver cancer, prognosis, *SPATS2*

1. Introduction

Liver cancer is the sixth most common malignant tumor in the world.^[1] Epidemiological data show that there are more than 780,000 new cases of liver cancer worldwide every year, which makes liver cancer become the second most common-seen cancerous death.^[2,3] Although great progress has been made in the treatment of liver cancer in recent years, the prognosis remain

poor.^[4,5] Therefore, a reliable prognostic marker is needed to improve the liver cancer patients' prognosis.

Spermatogenesis associated serine rich 2 (*SPATS2*) most expressed in adult testis and slightly expressed in liver and other tissues. Previous studies have found that *SPATS2* is involved in sperm development, and subsequently found that it promotes the progression of prostate cancer.^[6] However, the role of *SPATS2* in liver cancer is unknown, it is necessary to explore the role of *SPATS2* in liver cancer.

In this study, we tested the *SPATS2* mRNA expression difference in liver cancer by exploring TCGA database. ROC curve was drawn to evaluate the diagnostic value. The Chi-Squared test was used to evaluate the clinical correlation. Survival analysis and Cox regression model were executed to identify the effect of *SPATS2* on liver cancer patients' survival rate. GSEA was used to identify signaling pathways related to *SPATS2* expression

2. Materials and methods

2.1. Data source

We obtained the currently available clinical as well as the RNAseq data of 50 normal and 373 liver cancer tissues from TCGA databases (<https://cancergenome.nih.gov/>). The gene-level was estimated as $\log_2(x+1)$ transformed RSEM normalized count. The HCCDB dataset (<http://lifeome.net/database/hccdb/download.html>) was used for validation. No ethical conflict is needed because all the data in this study are from public database and available for research.

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2.2. Statistical analysis

We used R (version 3.5.3)^[7] for statistical analysis. Ggplot2 package was used to plot the boxplot.^[8] The ROC curve drawn by pROC package evaluated the capability of diagnosis as well as setting the optimal cutoff value to separate high *SPATS2* expression group from the low *SPATS2* expression group accordingly.^[9] Besides, the Chi-Squared test was used to evaluate the possible correlation between clinical features and the *SPATS2* expression. Then we used a survival package to plot survival curves and performed a logarithmic rank test.^[10] The univariate Cox model and multivariate Cox model were used to clarify the prognostic role of *SPATS2* independently from other clinical features.^[11]

2.3. Gene set enrichment analysis (GSEA)

GSEA determines whether an a priori defined set of genes has statistically significant differences in expression under 2 different biological conditions.^[12,13] This analysis, performed using GSEA software 3.0 from the Broad Institute, was used for analysis of RNAseq data from TCGA-LIHC. The gene set of “h.all.v6.2.symbols.gmt”, which summarizes and represents specific, well-defined biological states or processes, was downloaded from the Molecular Signatures Database (<http://software.broadinstitute.org/gsea/msigdb/index.jsp>). The normalized enrichment score (NES) was determined by analysis of 1000 permutations. A gene set was considered significantly enriched when the *P* value was less than .05 and the false discovery rate (FDR) was less than .25.

2.4. Ethical approval

Ethics committee approval was not necessary because all clinical data used in this study were obtained from a public database and are available for research.

3. Results

3.1. Patient characteristic

The *SPATS2* expression and clinical features including age, stage, new type, histologic grade, longest dimension, subdivision, sample type, lymphatic invasion, hepatitis virus, vital status, and *SPATS2* expression were shown in Table 1.

3.2. Differential expression of *SPATS2* in liver cancer

Boxplots showed that *SPATS2* expression was high in liver cancer compared with which in normal liver tissues ($P < 2.2e-16$, Fig. 1), which was also validated by HCCDB dataset (Fig. 1). In addition, *SPATS2* was also expressed differently in different groups of stage ($P = .00051$), vital status ($P = .013$), age ($P = .049$), gender ($P = .021$), T classification ($P = .00029$), and histologic grade ($P = 4.5e-10$).

3.3. The diagnostic capability of *SPATS2*

According to the performance of the ROC curve, we found the area under curve (AUC) was 0.964, which represents the high diagnostic ability. In addition, we reached the same results by analyzing the subgroups of different stages (AUC: stage I was 0.957, stage II was 0.976, stage III was 0.979, stage IV was 0.872; Fig. 2).

Table 1

Clinical characteristics of the liver cancer patients.

| characteristics | Number of patients (%) |
|----------------------------------|------------------------|
| Age | |
| <55 | 117 (31.45) |
| >=55 | 255 (68.55) |
| Gender | |
| FEMALE | 121 (32.44) |
| MALE | 252 (67.56) |
| Histological type | |
| Fibrolamellar Carcinoma | 3 (0.8) |
| Hepatocellular Carcinoma | 363 (97.32) |
| Hepatocholangiocarcinoma (Mixed) | 7 (1.88) |
| Histologic grade | |
| NA | 5 (1.34) |
| G1 | 55 (14.75) |
| G2 | 178 (47.72) |
| G3 | 123 (32.98) |
| G4 | 12 (3.22) |
| Stage | |
| NA | 24 (6.43) |
| I | 172 (46.11) |
| II | 87 (23.32) |
| III | 85 (22.79) |
| IV | 5 (1.34) |
| T classification | |
| NA | 2 (0.54) |
| T1 | 182 (48.79) |
| T2 | 95 (25.47) |
| T3 | 80 (21.45) |
| T4 | 13 (3.49) |
| TX | 1 (0.27) |
| N classification | |
| NA | 1 (0.27) |
| N0 | 253 (67.83) |
| N1 | 4 (1.07) |
| NX | 115 (30.83) |
| M classification | |
| M0 | 267 (71.58) |
| M1 | 4 (1.07) |
| MX | 102 (27.35) |
| Hepatitis virus | |
| FALSE | 219 (58.71) |
| TRUE | 154 (41.29) |
| Radiation therapy | |
| NA | 25 (6.7) |
| NO | 340 (91.15) |
| YES | 8 (2.14) |
| Residual tumor | |
| NA | 7 (1.88) |
| R0 | 326 (87.4) |
| R1 | 17 (4.56) |
| R2 | 1 (0.27) |
| RX | 22 (5.9) |
| Vital status | |
| DECEASED | 130 (34.85) |
| LIVING | 243 (65.15) |
| relapse | |
| NO | 179 (55.94) |
| YES | 141 (44.06) |
| SPATS2 | |
| High | 139 (37.27) |
| Low | 234 (62.73) |

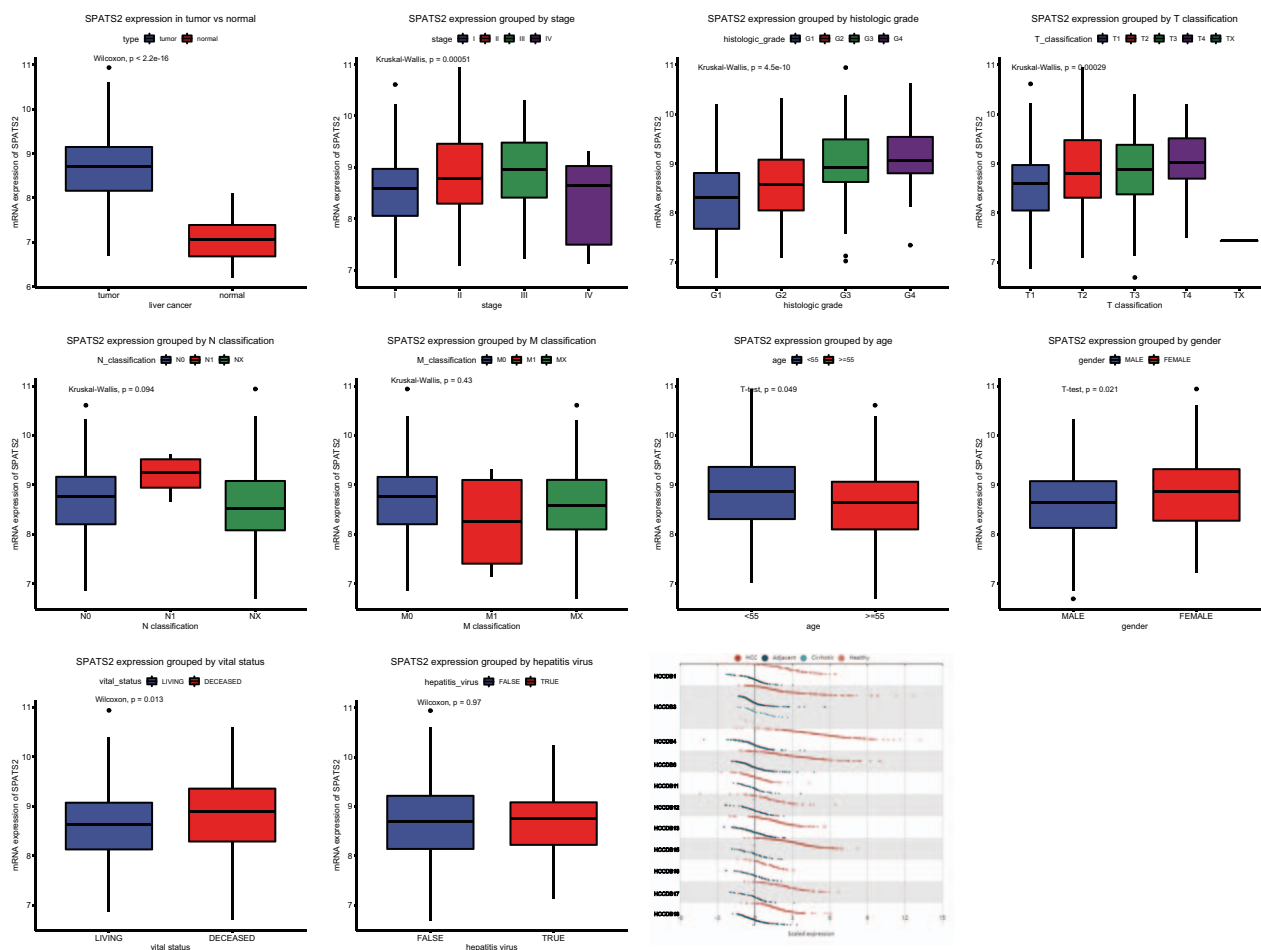


Figure 1. SPATS2 expression in liver cancer. SPATS2 expression between normal tissues and liver cancer, stages, histologic grade, TNM classification, age, gender, vital status. SPATS2 expression in liver cancer validated by HCCDB datasets.

3.4. Relationship between SPATS2 expression and clinical features of liver cancer

As show in Table 2, the expression of *SPATS2* was strongly associated with age ($P=.024$), vital status ($P=.001$), stage ($P=.002$), T classification ($P=.002$) and histologic grade ($P=.000$) of liver cancer patients.

3.5. High SPATS2 expression is related to liver cancer patients' poor overall survival

As shown in Figure 3, patients with higher *SPATS2* expression had particularly shorter OS ($P<.0001$), which was validated by HCCDB datasets (Fig. 3) and consistent with results of subgroup analysis, especially in stage I/II ($P=.03$), stage III/IV ($P<.001$), stage G1/G2 ($P<.0001$), stage G3/G4 ($P=.0054$), male ($P<.0001$), female ($P=.0047$), younger ($P<.0001$), older ($P=1e-04$), hepatitis virus positive ($P=.013$), and hepatitis virus negative ($P<.0001$). Cox model and Multivariate Cox model suggested that high *SPATS2* expression was an independent risk factor for the OS of liver cancer (HR=2.47, $P<.001$, Table 3).

3.6. High SPATS2 expression is related to liver cancer patients poor relapse-free survival

As shown in Figure 4, patients with higher *SPATS2* expression had particularly shorter RFS ($P<.0001$), which was consistent with results of subgroup analysis, especially in stage I/II ($P=.016$), stage III/IV ($P=.037$), stage G1/G2 ($P=.00014$), stage G3/G4 ($P=.016$), male ($P=.00014$), female ($P=.048$), younger ($P<.0036$), older ($P=.0028$), and hepatitis virus negative ($P<.0001$). Univariate Cox model and Multivariate Cox model suggested that high *SPATS2* expression was an independent risk factor for the RFS of liver cancer (HR=1.92, $P<.001$, Table 4).

3.7. GSEA identifies SPATS2-related signaling pathway

We compared the data sets for low and high *SPATS2* expression using GSEA to identify signaling pathways activated during liver cancer. The results indicated significant differences (FDR<0.25, NOM P value<.05) in the enrichment of the MSigDB collection (h.all.v6.2.symbols.gmt; Table 5). We selected the most significantly enriched signaling pathways based on normalized enrichment score (NES) (Fig. 5, Table 5).

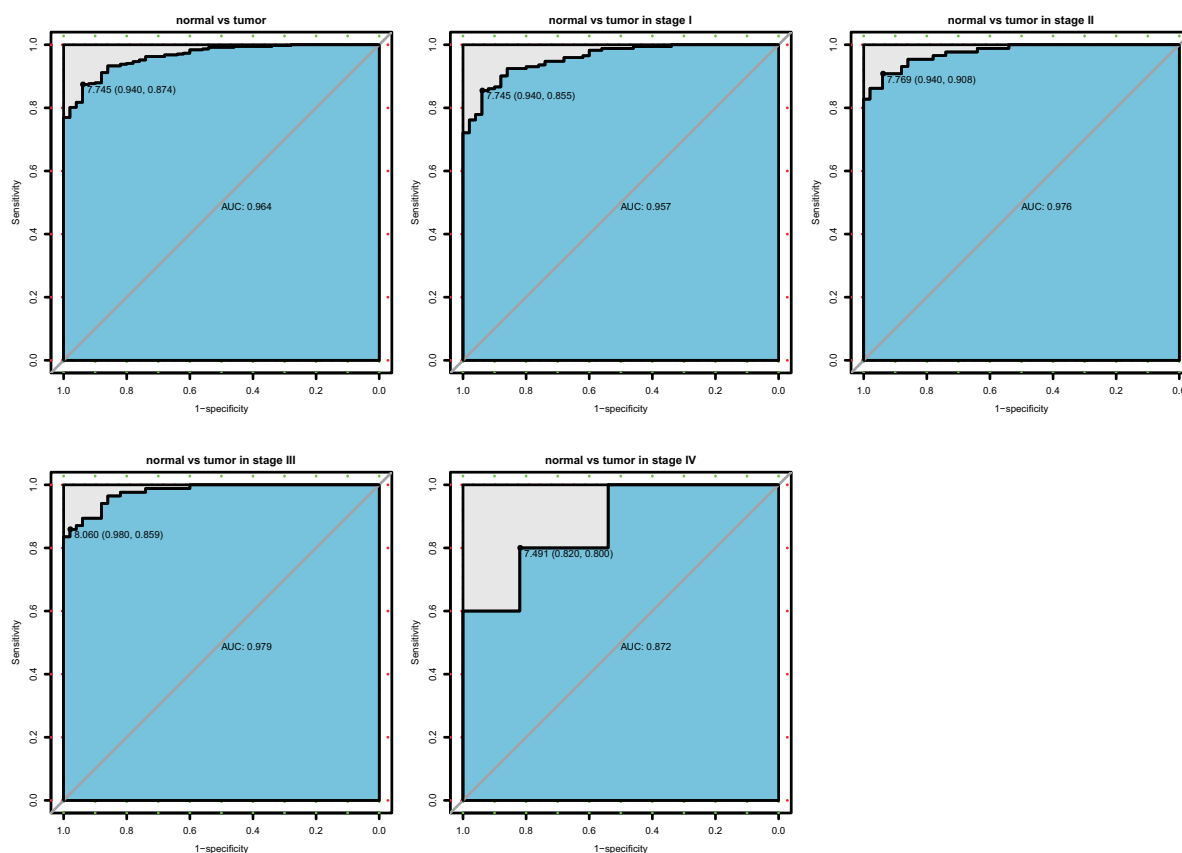


Figure 2. Diagnosis value of SPATS2. The ROC curve of normal tissues and liver cancer, and subgroup analysis of stage I, II, III, IV.

Table 2

Relationship between the clinical features and SPATS2 expression in liver cancer patients.

| Clinical characteristics | Variable | No. of patients | SPATS2 expression | | χ ² | P value | | |
|--------------------------|----------------------------------|-----------------|-------------------|---------|----------------|---------|---------|------|
| | | | High | Low | | | | |
| Age | <55 | 117 | 54 | -38.85 | 5.0981 | .024 | | |
| | ≥55 | 255 | 85 | -61.15 | | | | |
| Gender | FEMALE | 121 | 54 | -38.85 | 3.6998 | .054 | | |
| | MALE | 252 | 85 | -61.15 | | | | |
| Histological type | Fibrolamellar Carcinoma | 3 | 0 | 3 | -1.28 | 5.2958 | .096 | |
| | Hepatocellular Carcinoma | 363 | 134 | 229 | -97.86 | | | |
| | Hepatocholangiocarcinoma (Mixed) | 7 | 5 | 2 | -0.85 | | | |
| Histologic grade | G1 | 55 | 12 | -8.7 | 43 | -18.7 | 21.6385 | 0 |
| | G2 | 178 | 56 | -40.58 | 122 | -53.04 | | |
| | G3 | 123 | 62 | -44.93 | 61 | -26.52 | | |
| | G4 | 12 | 8 | -5.8 | 4 | -1.74 | | |
| Stage | I | 172 | 48 | -37.21 | 124 | -56.36 | 14.4648 | .002 |
| | II | 87 | 35 | -27.13 | 52 | -23.64 | | |
| | III | 85 | 44 | -34.11 | 41 | -18.64 | | |
| | IV | 5 | 2 | -1.55 | 3 | -1.36 | | |
| T classification | T1 | 182 | 51 | -36.69 | 131 | -56.47 | 16.0251 | .002 |
| | T2 | 95 | 42 | -30.22 | 53 | -22.84 | | |
| | T3 | 80 | 38 | -27.34 | 42 | -18.1 | | |
| | T4 | 13 | 8 | -5.76 | 5 | -2.16 | | |
| | TX | 1 | 0 | 0 | 1 | -0.43 | | |
| N classification | N0 | 253 | 97 | -70.29 | 156 | -66.67 | 3.4399 | .166 |
| | N1 | 4 | 3 | -2.17 | 1 | -0.43 | | |
| | NX | 115 | 38 | -27.54 | 77 | -32.91 | | |
| M classification | M0 | 267 | 102 | -73.38 | 165 | -70.51 | 0.7578 | .629 |
| | M1 | 4 | 2 | -1.44 | 2 | -0.85 | | |
| Hepatitis virus | FALSE | 102 | 35 | -25.18 | 67 | -28.63 | 0 | 1 |
| | TRUE | 219 | 82 | (58.99) | 137 | (58.55) | | |
| Radiation therapy | NO | 154 | 57 | (41.01) | 97 | (41.45) | 1.2796 | .258 |
| | YES | 340 | 131 | -99.24 | 209 | -96.76 | | |
| Residual tumor | R0 | 8 | 1 | -0.76 | 7 | -3.24 | 4.305 | .19 |
| | R1 | 326 | 116 | -85.29 | 210 | -91.3 | | |
| | R2 | 17 | 9 | -6.62 | 8 | -3.48 | | |
| | RX | 1 | 0 | 0 | 1 | -0.43 | | |
| Vital status | DECEASED | 22 | 11 | -8.09 | 11 | -4.78 | 11.4474 | .001 |
| | LIVING | 130 | 64 | -46.04 | 66 | -28.21 | | |
| | | 243 | 75 | -53.96 | 168 | -71.79 | | |

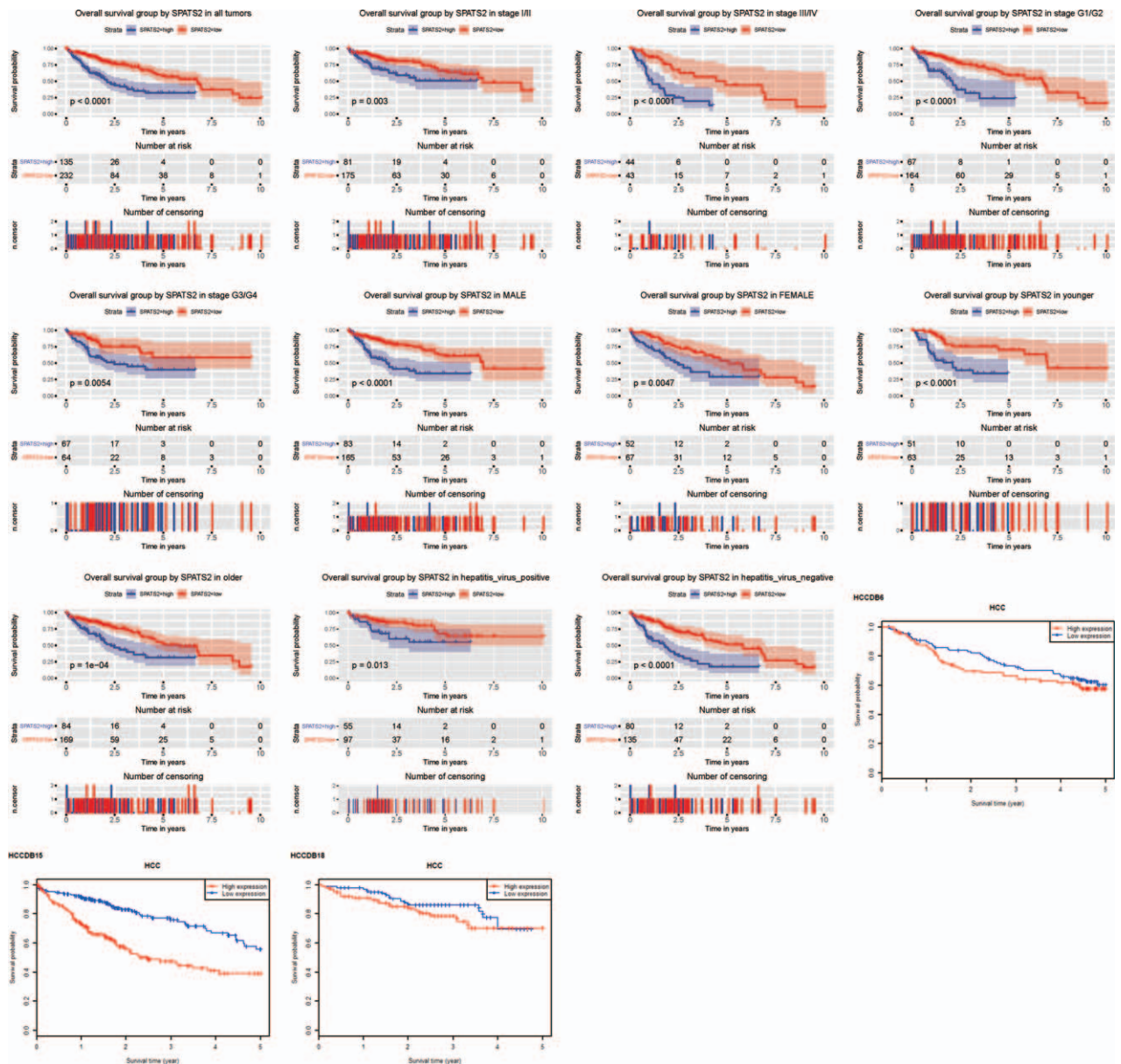


Figure 3. Survival curves of OS in liver cancer. Kaplan–Meier curves of liver cancer patients OS in all tumors, histologic grade G1/G2, G3/G4, clinical stage I/II, III/IV, male, female, younger, and older. Survival curves of OS grouped by SPATS2 in liver cancer validated by HCCDB datasets.

Table 3

Univariate analysis and multivariate analysis of liver cancer patients overall survival.

| Parameters | Univariate analysis | | | Multivariate analysis | | |
|-------------------|---------------------|---------------------|---------|-----------------------|---------------------|---------|
| | Hazard ratio | 95%CI (lower-upper) | P value | Hazard ratio | 95%CI (lower-upper) | P value |
| Age | 1 | 0.69–1.45 | .997 | | | |
| Gender | 0.8 | 0.56–1.14 | .22 | | | |
| Histological type | 0.99 | 0.27–3.66 | .986 | | | |
| Histologic grade | 1.04 | 0.84–1.3 | .698 | | | |
| Stage | 1.38 | 1.15–1.66 | .001 | 0.89 | 0.71–1.11 | .29 |
| T classification | 1.66 | 1.39–1.99 | 0 | 1.61 | 1.26–2.06 | 0 |
| N classification | 0.73 | 0.51–1.05 | .086 | | | |
| M classification | 0.72 | 0.49–1.04 | .077 | | | |
| Hepatitis virus | 0.51 | 0.35–0.74 | .001 | 0.64 | 0.43–0.95 | .028 |
| Radiation therapy | 0.51 | 0.26–1.03 | .06 | | | |
| Residual tumor | 1.42 | 1.13–1.8 | .003 | 1.35 | 1.05–1.74 | .018 |
| SPATS2 | 2.72 | 1.9–3.89 | 0 | 2.47 | 1.71–3.56 | 0 |

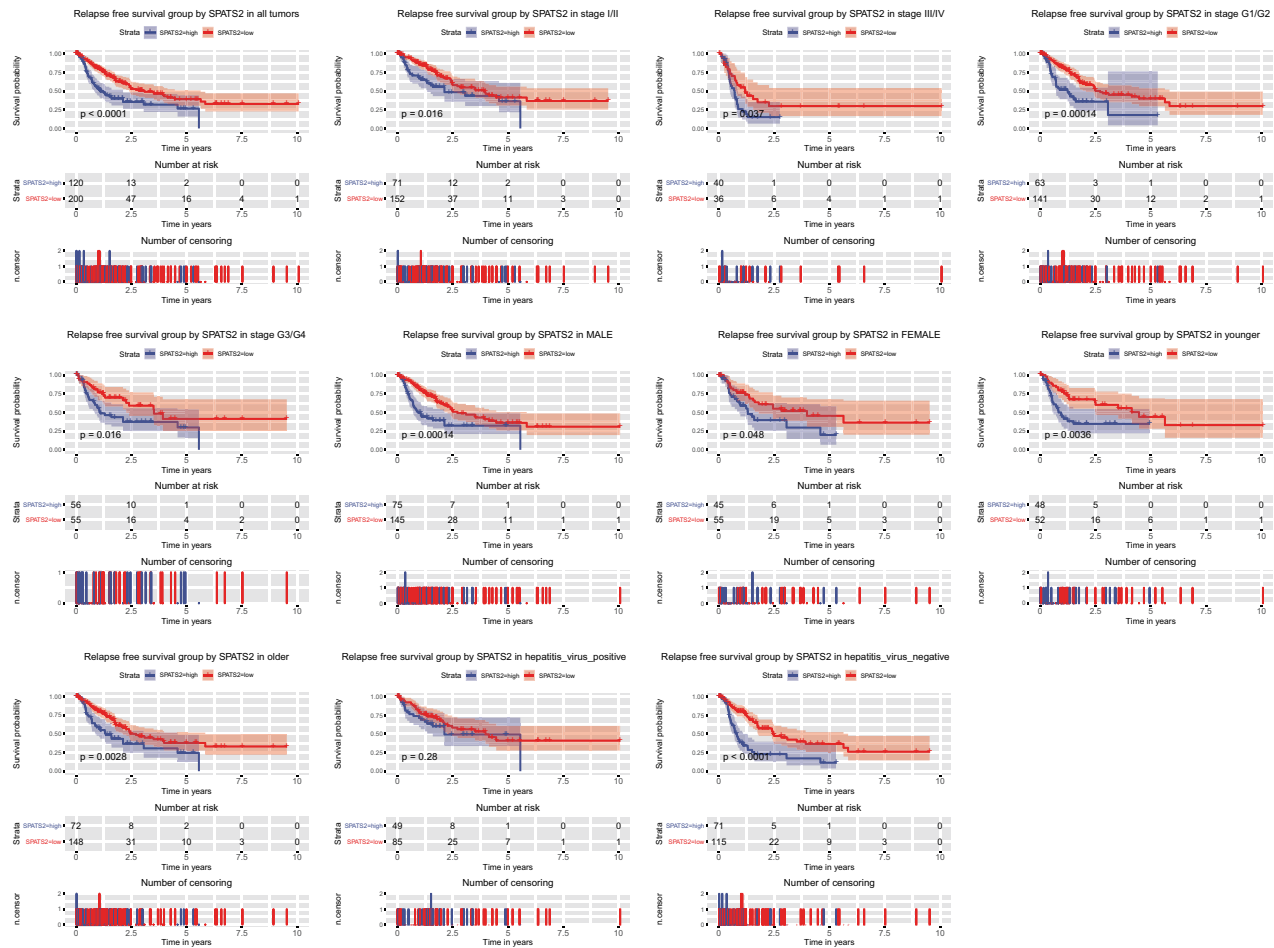


Figure 4. Survival curves of RFS in liver cancer. Kaplan–Meier curves of liver cancer patients RFS in all tumors, histologic grade G1/G2, G3/G4, clinical stage I/II, III/IV, male, female, younger, and older.

The results indicated the data set with high *SPATS2* expression was enriched in Mitotic spindle, G2M checkpoint, E2F targets.

4. Discussion

At present, liver cancer can be treated in various ways, such as hepatectomy, liver transplantation, ablation, interventional

therapy, radiotherapy, drug therapy, and biological immunity. However, the prognosis of patients is still poor. Therefore, it is urgent to find novel markers to predict the prognosis of liver cancer.^[14] Many researches have been working to find out the prognostic maker of liver cancer, and aim to guide clinicians to evaluate the prognosis of liver cancer patients recently.^[15–22] In this research, we found that *SPATS2*

Table 4
Univariate analysis and multivariate analysis of liver cancer patients' recurrence-free survival.

| Parameters | Univariate analysis | | | Multivariate analysis | | |
|-------------------|---------------------|---------------------|---------|-----------------------|---------------------|---------|
| | Hazard ratio | 95%CI (lower-upper) | P value | Hazard ratio | 95%CI (lower-upper) | P value |
| Age | 0.9 | 0.63–1.28 | .55 | | | |
| Gender | 0.99 | 0.7–1.41 | .966 | | | |
| Histological type | 2.02 | 0.66–6.24 | .22 | | | |
| Histologic grade | 0.98 | 0.8–1.21 | .883 | | | |
| stage | 1.66 | 1.38–1.99 | 0 | 1.12 | 0.87–1.44 | .363 |
| T classification | 1.78 | 1.49–2.12 | 0 | 1.55 | 1.18–2.02 | .002 |
| N classification | 0.97 | 0.67–1.4 | .874 | | | |
| M classification | 1.17 | 0.79–1.74 | .432 | | | |
| Hepatitis virus | 0.63 | 0.44–0.89 | .008 | 0.83 | 0.57–1.2 | .318 |
| Radiation therapy | 0.74 | 0.26–2.16 | .584 | | | |
| Residual tumor | 1.28 | 1.01–1.61 | .042 | 1.34 | 1.05–1.71 | .018 |
| SPATS2 | 2.06 | 1.47–2.89 | 0 | 1.92 | 1.36–2.71 | 0 |

Table 5**Gene set enrichment analysis (GSEA) of signaling pathways activated during liver cancer.**

| Gene set | ES | NES | NOM p-val | FDR q-val |
|--------------------------|--------|--------|-----------|-----------|
| HALLMARK_MITOTIC_SPINDLE | -0.601 | -1.922 | 0.000 | 0.050 |
| HALLMARK_G2M_CHECKPOINT | -0.724 | -1.819 | 0.002 | 0.064 |
| HALLMARK_E2F_TARGETS | -0.714 | -1.748 | 0.004 | 0.077 |

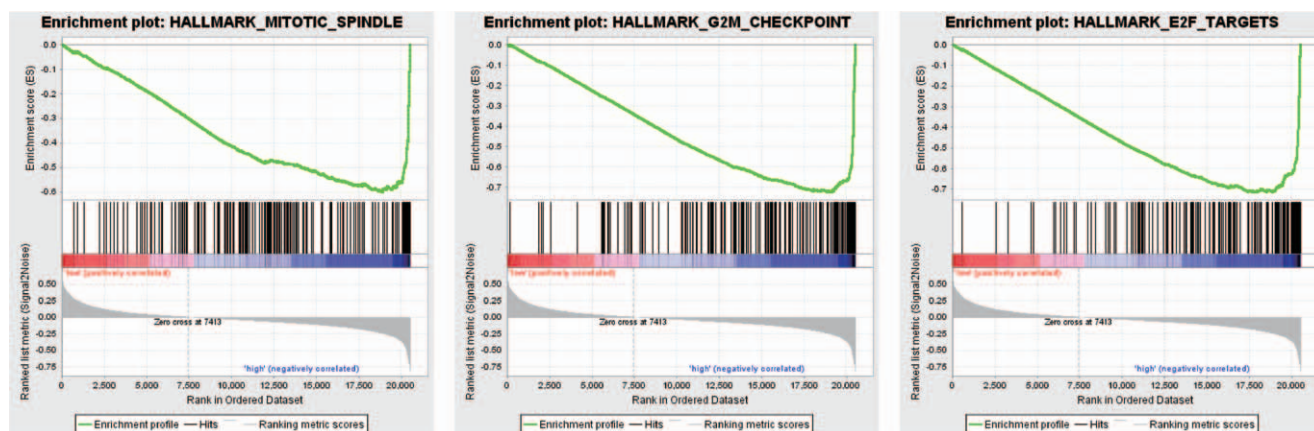


Figure 5. Gene set enrichment analysis (GSEA) of signaling pathways activated during liver cancer. The high *SPATS2* expression phenotype had enrichment of Mitotic spindle, G2M checkpoint, E2F targets.

expression could be a novel diagnostic and prognostic biomarker in liver cancer.

The gene *SPATS2* was firstly found to encode a polypeptide containing 545 amino acid residues in mouse testis, which is involved in sperm growth and development.^[23] Subsequently, the researchers found that *SPATS2* was also expressed in 25 human tissues.^[24] Recent studies have found that *SPATS2* is highly expressed in squamous cell carcinoma but lowly expressed in non-lepidic AD.^[25] In this study, we found that *SPATS2* was highly expressed in liver cancer compared with normal liver tissue. This may be due to the organization specificity. In addition, *SPATS2* can induce the transcription of *SNHG5*, promoting the survival of colon cancer cells.^[26] Meanwhile, we found that the *SPATS2* expression gradually increased in the stage, histologic stage and T classification, indicating that *SPATS2* might promote the progress of liver cancer through helping cancer cell survival. Importantly, *SPATS2* was highly expressed in the deceased than it in the survivors, so further survival analysis is necessary.

The survival of cancer cells is the main cause of cancer recurrence. Previous studies have found that *SPATS2* removes H3K27me3 histone markers and promotes the survival of prostate cancer.^[6] Interestingly, patients with high *SPATS2* expression have a significantly shorter OS and RFS in liver cancer. Subgroup analysis also found the prognostic significance in the stage I/II, stage III/IV, G1/G2, G3/G4, male, female, younger, and older. This suggests that *SPATS2* plays an important prognostic role in the whole process of liver cancer development. Consistent with these findings, we found that high *SPATS2* could predict poor prognosis in liver cancer, which may involve in Mitotic spindle, G2 M checkpoint, E2F targets.

To our knowledge, this is the first study to examine the prognostic value of *SPATS2* expression in liver cancer. Together

with other studies about the functions of *SPATS2*, we have contributed to a better understanding of the role of *SPATS2*, as well as great possibility of accurately predicting the prognosis of liver cancer. However, the sample size is limited in this study. In the future, more samples need to be involved to validate the prognostic role of *SPATS2* in liver cancer.

5. Conclusion

In conclusion, we mainly focused on the diagnostic and prognostic value of *SPATS2* in patients with liver cancer. *SPATS2* expression could be a novel diagnostic and prognostic biomarker in liver cancer

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Author contributions

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Investigation: Yijun Tian, Wu Ji.

Methodology: Jin Xing.

Project administration: Yijun Tian.

Resources: Yijun Tian.

Software: Jin Xing.

Validation: Yijun Tian, Wu Ji.

Visualization: Jin Xing.

Writing – original draft: Jin Xing.

Writing – review & editing: Xinying Wang.

References

- [1] Bray F, Ferlay J, Soerjomataram I, et al. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin* 2018;68:394–424.
- [2] Ferlay J, Soerjomataram I, Dikshit R, et al. Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. *Int J Cancer* 2015;136:E359–386.
- [3] Llovet JM, Zucman-Rossi J, Pikarsky E, et al. Hepatocellular carcinoma. *Nat Rev Dis Primers* 2016;2:16018.
- [4] Tovoli F, Negrini G, Benevento F, et al. Systemic treatments for hepatocellular carcinoma: challenges and future perspectives. *Hepat Oncol* 2018;5:HE01.
- [5] Ikeda K. Recent advances in medical management of hepatocellular carcinoma. *Hepatol Res* 2019;49:14–32.
- [6] Ngollo M, Lebert A, Daures M, et al. Global analysis of H3K27me3 as an epigenetic marker in prostate cancer progression. *BMC cancer* 2017;17:261.
- [7] Team RDCJ. *RA language and environment for statistical computing*. R Foundation for Statistical Computing, Vienna, Austria 2009;14:12–21.
- [8] Wickham H. *Ggplot2: elegant graphics for data analysis*. *J R Stat Soc* 2011;174:245–6.
- [9] Robin X, Turck N, Hainard A, et al. pROC: an open-source package for R and S+ to analyze and compare ROC curves. *BMC Bioinformatics* 2011;12:77.
- [10] Therneau TM, April. *A Package for Survival Analysis in S*. 1994.
- [11] Therneau TM, Grambsch PM. *Modeling Survival Data: Extending the Cox Model*. New York: Springer; 2000.
- [12] Mootha VK, Lindgren CM, Eriksson KF, et al. PGC-1 α -responsive genes involved in oxidative phosphorylation are coordinately down-regulated in human diabetes. *Nat Genet* 2003;34:267–73.
- [13] Subramanian A, Tamayo P, Mootha VK, et al. Gene set enrichment analysis: a knowledge-based approach for interpreting genome-wide expression profiles. *Proc Natl Acad Sci USA* 2005;102:15545–50.
- [14] Guo W, Tan HY, Wang N, et al. Deciphering hepatocellular carcinoma through metabolomics: from biomarker discovery to therapy evaluation. *Cancer Manag Res* 2018;10:715–34.
- [15] Jiao Y, Fu Z, Li Y, et al. High EIF2B5 mRNA expression and its prognostic significance in liver cancer: a study based on the TCGA and GEO database. *Cancer Manag Res* 2018;10:6003–14.
- [16] Jiao Y, Li Y, Lu Z, et al. High trophinin-associated protein expression is an independent predictor of poor survival in liver cancer. *Dig Dis Sci* 2019;64:137–43.
- [17] Jiao Y, Fu Z, Li Y, et al. Aberrant FAM64A mRNA expression is an independent predictor of poor survival in pancreatic cancer. *PLoS One* 2019;14:e0211291.
- [18] Jiao Y, Li Y, Jiang P, et al. PGM5: a novel diagnostic and prognostic biomarker for liver cancer. *PeerJ* 2019;7:e7070.
- [19] Jiao Y, Li Y, Liu S, et al. ITGA3 serves as a diagnostic and prognostic biomarker for pancreatic cancer. *Onco Targets Ther* 2019;12:4141–52.
- [20] Li Y, Jiao Y, Fu Z, et al. High miR-454-3p expression predicts poor prognosis in hepatocellular carcinoma. *Cancer Manag Res* 2019;11:2795–802.
- [21] Jiao Y, Li Y, Fu Z, et al. OGDHL expression as a prognostic Biomarker for liver cancer patients. *Dis Markers* 2019;2019:9.
- [22] Hou L, Zhang X, Jiao Y, et al. ATP binding cassette subfamily B member 9 (ABCB9) is a prognostic indicator of overall survival in ovarian cancer. *Medicine (Baltimore)* 2019;98:e15698.
- [23] Senoo M, Hoshino S, Mochida N, et al. Identification of a novel protein p59(scr), which is expressed at specific stages of mouse spermatogenesis. *Biochem Biophys Res Commun* 2002;292:992–8.
- [24] Seki K, Koshi R, Sugano N, et al. Microarray analysis of bisphenol A-induced changes in gene expression in human oral epithelial cells. *Acta biochimica et biophysica Sinica* 2007;39:879–84.
- [25] Takamochi K, Ohmiya H, Itoh M, et al. Novel biomarkers that assist in accurate discrimination of squamous cell carcinoma from adenocarcinoma of the lung. *BMC Cancer* 2016;16:760.
- [26] Damas ND, Marcatti M, Come C, et al. SNHG5 promotes colorectal cancer cell survival by counteracting STAU1-mediated mRNA destabilization. *Nat Commun* 2016;7:13875.