



Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information website.

Elsevier hereby grants permission to make all its COVID-19-related research that is available on the COVID-19 resource centre - including this research content - immediately available in PubMed Central and other publicly funded repositories, such as the WHO COVID database with rights for unrestricted research re-use and analyses in any form or by any means with acknowledgement of the original source. These permissions are granted for free by Elsevier for as long as the COVID-19 resource centre remains active.

The physiology and pharmacology of singlet oxygen

Thomas W. Stief

University Hospital, Marburg, Germany

Summary Reactive oxygen species (ROS) are generated by many different cells. Singlet oxygen ($^1\text{O}_2$) and a reaction product of it, excited carbonyls ($\text{C}=\text{O}^*$), are important ROS. $^1\text{O}_2$ and $\text{C}=\text{O}^*$ are nonradical and emit light (one photon/molecule) when returning to ground state oxygen. Especially activated polymorphonuclear neutrophil granulocytes (PMN) produce large amounts of $^1\text{O}_2$. Via activation of the respiratory burst (NADPH oxidase and myeloperoxidase) they synthesize hypochlorite (NaOCl) and chloramines (in particular *N*-chlorotaurine). Chloramines are selective and stable chemical generators of $^1\text{O}_2$. In the human organism, $^1\text{O}_2$ is both a signal and a weapon with therapeutic potency against very different pathogens, such as microbes, virus, cancer cells and thrombi. Chloramines at blood concentrations between 1 and 2 mmol/L inactivate lipid enveloped virus and chloramines at blood concentrations below 0.5 mmol/L, i.e. at oxidant concentrations that do not affect thrombocytes or hemostasis factors, act antithrombotically by activation of the physiologic PMN mediated fibrinolysis; this thrombolysis is of selective nature, i.e. it does not impair the hemostasis system of the patient allowing the antithrombotic treatment in patients where the current risky thrombolytic treatment is contraindicated. The action of $^1\text{O}_2$ might be compared to the signaling and destroying gunfire of soldiers directed against bandits at night, resulting in an autorecruitment of the physiological inflammatory response. Chloramines (such as the mild and un toxic oxidant chloramine T® (*N*-chloro-*p*-toluene-sulfonamide)) and their signaling and destroying reaction product $^1\text{O}_2$ might be promising new therapeutic agents against a multitude of up to now refractory diseases.

© 2003 Elsevier Science Ltd. All rights reserved.

INTRODUCTION

The human redox state is a balanced system of pro- and anti-oxidants. The main cellular reactive oxygen species (ROS) are hydrogen peroxide (H_2O_2), superoxide anion (O_2^-), hydroxyl radical (HO^\bullet), and singlet oxygen ($^1\text{O}_2$). Singlet oxygen – in contrast to the other oxidants – is nonradical and excited, i.e. $^1\text{O}_2$ or the reaction product of $^1\text{O}_2$ with a $\text{C}=\text{C}$ group, i.e. an excited carbonyl, emits 1 photon when returning to ground state oxygen (1). Whereas the radicalic oxygen species are harmful for the

organism, nonradicalic $^1\text{O}_2$ is rather mild and un toxic for mammalian tissue. This mild oxidative character has been used for diagnostic purposes, such as the radio-halogenation of proteins (2–4).

GENERATION OF $^1\text{O}_2$

ROS are generated by pro-oxidative enzyme systems or by redox-cycling of pro-oxidative compounds. Pro-oxidative enzymes are the NADPH-oxidase (5), myeloperoxidase (6), NO-synthase (7,8), or the cytochrome P-450 chain (9–11). Physiologic activation of these pro-oxidative enzymes results into the normal oxidative state. NADPH-oxidase is mainly found in polymorphonuclear leukocytes (PMN). The membranous NADPH-oxidase generates superoxide anions that dismutate to hydrogen peroxide. H_2O_2 can react with superoxide anions or with HOCl or chloramines to form the nonradicalic $^1\text{O}_2$ (10,11). Since NADPH-oxidase is present in many

Received 20 June 2002

Accepted 11 November 2002

Correspondence to: Thomas W. Stief, MD, Priv.-Doz. Department of Clinical Chemistry and Molecular Diagnostics, University Hospital, D-35033 Marburg, Germany. Phone: +49-6421-64471; Fax: +49-6421-65594; E-mail: thstief@post.med.uni-marburg.de

different cells (5), diverse cells seem to generate the signal/messenger $^1\text{O}_2$ for inter- or intra-cellular signaling.

$^1\text{O}_2$ AS A CELL SIGNAL/MESSENGER

$^1\text{O}_2$ is a cell signal and messenger (12–14): redox active agents regulate ion channel activity in animals and plants (15). $^1\text{O}_2$ activates large-conductance, Ca^{2+} -activated (maxi) K^+ channels (16): monochloramine (NH_2Cl) – in contrast to Tau-Cl – is membrane permeating and at 3–30 $\mu\text{mol/L}$ it increases outward currents more than 8-fold (17,18). $^1\text{O}_2$, generated by chloramine-T[®] (*N*-chloro-*p*-toluene-sulfonamide), also inactivates the Na^+ currents from skeletal or heart muscle fibers, presumably by oxidation of methionine residues (19–21). Chloramine-T[®] has also been shown to modulate dose dependently outward currents in rabbit atrial cells (22,23) or potassium channels (24–27). Chloramine-T[®] is known to abolish inactivation of Na^+ and K^+ channels (28–33). Potential receptors for excited oxygen species/light are cryptochromes (34), that consist of flavin- and pteridine- prosthetic groups. Pteridines seem to interact with excited oxygen (35–37).

$^1\text{O}_2$ AS A WEAPON

Important armatory functions of $^1\text{O}_2$ are:

- (a) antiinfectious (antibacterial, antiviral);
- (b) cytostatic (anticancer);
- (c) antiatherothrombotic (selective thrombolysis).

Ad (a)

Chloramine-T[®] is bactericidal (38,39). *N*-chloramines exhibit low toxicity and skin irritation and are superior to chlorhexidine in preventing the expansion of the normal skin flora in vivo (40). Chloramine-T[®] is better than HOCl in inactivation of *Staphylococcus aureus* (41) and monochloramine is superior to *N*-chlorotaurine in inactivation of *Mycobacterium terrae* (42). NaOCl shows higher activity than chloramine-T[®] against *Bacillus subtilis* spores, coat and cortex material was degraded by chloramine-T[®] (43).

Because of their unotoxicity and antimicrobial power (44), chloramines – especially chloramine-T[®] – is used for disinfection of drinking water, dialysate, or ice cream machines (45–48). Chloramine T[®] is also a therapeutic drug for treating bacterial gill disease, a predominant disease of a variety of fish species (49). However, chloramine-T[®] at 10 g/L (35 mM) has been shown to be ineffective as fungicide (50).

Chloramines are virucidal, too (51–56). Even such dangerous viruses as the Marburg virus (57), or the Ebola virus (58,59) are inactivated by chloramines.

Bhanja virus (60), lymphocytic choriomeningitis virus (61), simian rotavirus (62), or poliovirus (63–65) are sensible to NaOCl/chloramines. Even replicating agents of the Creutzfeldt–Jakob disease show some sensibility to NaOCl (74,75).

Poliovirus on whole hands is inactivated (reduction factor >100) by 35 mM chloramine T[®] (63,67). Coxsackievirus B3, adenovirus type 5, parainfluenza virus type 3 and coronavirus 229E are inactivated (reduction factor >1000) by a 100 mM chloramine-T[®] solution (68). NaOCl inactivates HIV-1 (66,69–72). The 1.5 mM NaOCl inactivated more than 10 000 fold HIV in serum and 7.5 mM more than 10 fold in blood (73). Own experiments show that chloramine-T[®] at blood concentrations that are tolerable for normal hemostasis function inactivate the lipid enveloped model virus VSV (vesicular stomatitis virus): 1 mmol/L chloramine-T[®] inactivates 90% of added VSV, 2 mmol/L chloramine-T[®] inactivate 99% of added VSV, i.e. there seems to exist a narrow therapeutical window for $^1\text{O}_2$ treatment of human infections by enveloped viruses. Intravenous infusions of 1–1.5 mmol/L (blood concentration) chloramine (chloramine-T[®] or the physiologic *N*-chlorotaurine) once a week for several weeks might be a potent treatment modality for infections with lipid enveloped viruses, such as human immunodeficiency virus (HIV) (74).

Ad (b)

Singlet oxygen is tumoricidal (75). In photodynamic therapy (PDT) high concentrations of singlet oxygen are generated by illumination of a photosensitizer, resulting in a cytostatic action of PDT (76,77). However, excessive oxidant concentrations are carcinogenic (78–82).

Ad (c)

$^1\text{O}_2$ mediates PMN adherence to the endothelium (12,83,84) and subsequently selective thrombolysis (10,11). $^1\text{O}_2$ activates the complement cascade, transforming C5 into a C5b-like molecule (85); activation of the complement cascade results in increased PMN adhesion to endothelial cells (86,87). Since cholesterol is an inhibitor of $^1\text{O}_2$, the atherogenic action of cholesterol might be explained by insufficient thrombolytic capacity of a hypercholesterolemic individuuum (10,11,88).

TOXICOLOGY OF $^1\text{O}_2$

However, and according to Paracelsus (dosis sola venenum facit (only the dosage makes the poison)), high concentrations of chloramines can act toxic to normal tissue (89). 3 mM monochloramine induced DNA breakage (90). PMN are the main cells that use singlet oxygen as

a weapon. They also dispose of an enzyme that reverses methionine oxidation – the methionine sulfoxide-peptide reductase (91). Taurine-chloramine is the major chloramine generated in activated PMN as a result of the reaction between HOCl (92) and taurine, an abundant free amino acid in their cytosol (93–96). Also other plasma proteins react with hypochlorite to chloramines (97). HOCl (25 μ M) or NH_2Cl (10 μ M) – but not Tau-Cl (100 μ M) – increase endothelial permeability (98) or epithelial cell injury (99). NH_2Cl , the reaction product of hypochlorite with ammonia (NH_3), seems to be more toxic than Tau-Cl (100,101). The 60 mM NH_2Cl (about 10 times the concentration generated by activated PMN!) is ulcerogenic in rat stomachs, taurine application (1 ml 200 mM) attenuates the deleterious action of NH_2Cl (102), NH_2Cl induces apoptosis in gastric mucosa (103). Tau-Cl selectively modulates the ability of dendritic cells to induce the release of IL-2 and IL-10 from T cells (104). Tau-Cl inhibits monocyte chemoattractant protein-1 and macrophage inflammatory protein-2 production in glioma cells (105). Tau-Cl inhibits the production of NO and superoxide anions (106–109), prostaglandin E2 (110, 111), interleukin 6, and tumor necrosis factor- α and it has been suggested that Tau-Cl may regulate the balance between protective, microbicidal and toxic effect of PMN, Tau-Cl at 0.1–0.3 mM inhibits interleukin-2 release of purified T cells (112).

Chloramines – in contrast to sodium chlorite – do not induce detectable hematologic (\rightarrow methemoglobin) or hepatic (\rightarrow elevation of serum alanine-amino-transferase) in African Green monkeys (113). However, a chloramine-induced haemolysis and erythropoietin resistance occurred when the dialysate chloramine levels rose from <0.1 to 0.3 p.p.m. (about 1 mM) resulting in an increase in mean methaemoglobin of 23% and a 21% fall in mean haptoglobin during haemodialysis; only one patient with glucose-6-phosphate-dehydrogenase deficiency had Heinz bodies (114,115). Dogs treated with 1 mmol/L blood concentration of chloramine T[®] 3 times a week for several months did not show toxic side effects (116).

CONCLUSION

Singlet oxygen is a major agent generated by many different cell types, especially by neutrophil granulocytes. $^1\text{O}_2$ is nonradical and emits light when returning to ground state oxygen. Like the gunfire of soldiers directed against bandits, $^1\text{O}_2$ is both a signal and a weapon, directed against multiple pathogens – including microbes, virus, cancer cells, thrombi – and resulting in an autorecruitment of the physiological inflammatory response. Chloramines are stable chemical generators of $^1\text{O}_2$. *N*-chlorotaurine is an important physiological chloramine, for therapeutic purposes chloramine-T[®]

seems to be a promising new therapeutic agent against a multitude of up to now refractory diseases.

REFERENCES

1. Olszowski S., Olszowska E., Stelmaszynska T., Krawczyk A. Chemiluminescence of ABEI-labelled IgG triggered by the *N*-chloramine- H_2O_2 -*p*-iodophenol system. *Luminescence* 1999; **14**: 139–145.
2. Tejedor F., Ballesta J. P. Iodination of biological samples without loss of functional activity. *Anal Biochem* 1982; **127**: 143–149.
3. Silberring J., Golda W., Szybinski Z. A universal and simple chloramine T version for hormone iodination. *Int J Appl Radiat Isot* 1982; **33**: 117–119.
4. Lindgren S., Skarnemark G., Jacobsson L., Karlsson B. Chloramine-T in high-specific-activity radioiodination of antibodies using *N*-succinimidyl-3-(trimethylstannyl) benzoate as an intermediate. *Nucl Med Biol* 1998; **25**: 659–665.
5. Babior B. M. NADPH-oxidase: an update. *Blood* 1999; **93**: 1464–1476.
6. Tatsuzawa H., Maruyama T., Hori K., Sano Y., Nakano M. Singlet oxygen ($(^1\Delta(g)\text{O}(2))$) as the principal oxidant in myeloperoxidase-mediated bacterial killing in neutrophil phagosome. *Biochem Biophys Res Commun* 1999; **262**: 647–650.
7. Furchgott R. Endothelium-dependent relaxation, endothelium-derived relaxing factor and photorelaxation of blood vessels. *Semin Perinatol* 1991; **15**: 11–15.
8. Stuehr D. J., Kwon N. S., Nathan C. F. FAD and GSH participate in macrophage synthesis of nitric oxide. *Biochem Biophys Res Commun* 1990; **168**: 558–565.
9. Stuehr D. J., Ikeda-Saito M. Spectral characterization of brain and macrophage nitric oxide synthases. Cytochrome P-450 like heme proteins that contain a flavin semiquinone radical. *JBC* 1992; **267**: 20547–20550.
10. Stief T. W., Fareed J. The antithrombotic factor singlet oxygen/light ($^1\text{O}_2/h\nu$). *Clin Appl Thromb/Hemostasis* 2000; **6**: 22–30.
11. Stief T. W. The blood fibrinolysis/deep-sea analogy: a hypothesis on the cell signals singlet oxygen/photons as natural antithrombotics. *Thromb Res* 2000; **99**: 1–20.
12. Grether-Beck S., Olaizola-Horn S., Schmitt H., Grewe M., Jahnke A., Johnson J. P., Briviba K., Sies H., Krutmann J. Activation of transcription factor AP-2 mediates UVA radiation- and singlet oxygen induced expression of the human intercellular adhesion molecule1 gene. *Proc Natl Acad Sci USA* 1996; **93**: 14586–14591.
13. Gorman A. A., Rodgers M. A. Current perspectives of singlet oxygen detection in biological environments. *J Photochem Photobiol B* 1992; **14**: 159–176.
14. Briviba K., Klotz L. O., Sies H. Toxic and signaling effects of photochemically or chemically generated singlet oxygen in biological systems. *Biol Chem* 1997; **378**: 1259–1265.
15. Carpaneto A., Cantu A. M., Gambale F. Redox agents regulate ion channel activity in vacuoles from higher plant cells. *FEBS Lett* 1999; **442**: 129–132.
16. Schoonbroodt S., Legrand-Poels S., Best-Belpomme M., Piette J. Activation of the NF- κ B transcription factor in a T-lymphocytic cell line by hypochlorous acid. *Biochem J* 1997; **321**: 777–785.
17. Prasad M., Matthews J. B., He X. D., Akbarali H. I. Monochloramine directly modulates $\text{Ca}(2+)$ -activated $\text{K}(+)$

- channels in rabbit colonic muscularis mucosae. *Gastroenterology* 1999; **117**: 906–917.
18. Nakamura T. Y., Yamamoto I., Nishitani H., Matozaki T., Suzuki T., Wakabayashi S., Shigekawa M., Goshima K. Detachment of cultured cells from the substratum induced by the neutrophil-derived oxidant NH₂Cl: synergistic role of phosphotyrosine and intracellular Ca²⁺ concentration. *J Cell Biol* 1995; **131**: 509–524.
 19. Vogt W. Oxidation of methionyl residues in proteins: tools, targets, and reversal. *Free Radic Biol Med* 1995; **18**: 93–105.
 20. Quinonez M., DiFranco M., Gonzalez F. Involvement of methionine residues in the fast inactivation mechanism of the sodium current from toad skeletal muscle fibers. *J Membr Biol* 1999; **169**: 83–90.
 21. Koumi S., Sato R., Hayakawa H. The activation gate of cardiac Na⁺ channel modulates voltage- and pH-dependent unbinding of disopyramide. *Eur J Pharmacol* 1995; **277**: 165–172.
 22. Tanaka H., Habuchi Y., Nishio M., Suto F., Yoshimura M. Modulation by chloramine-T of 4-aminopyridine-sensitive transient outward current in rabbit atrial cells. *Eur J Pharmacol* 1998; **358**: 85–92.
 23. Ulbricht W. The inactivation of sodium channels in the node of Ranvier and its chemical modification. *Ion Channels* 1990; **2**: 123–168.
 24. Bauer C. K., Falk T., Schwarz J. R. An endogenous inactivating inward-rectifying potassium current in oocytes of *Xenopus laevis*. *Pflugers Arch* 1996; **432**: 812–820.
 25. Schlieff T., Schonherr R., Heinemann S. H. Modification of C-type inactivating Shaker potassium channels by chloramine-T. *Pflugers Arch* 1996; **431**: 483–493.
 26. Stephens G. J., Robertson B. Inactivation of the cloned potassium channel mouse Kv1.1 by the human Kv3.4 'ball' peptide and its chemical modification. *J Physiol* 1995; **484**: 1–13.
 27. Koumi S., Sato R., Hayakawa H. Modulation of voltage-dependent inactivation of the inwardly rectifying K⁺ channel by chloramine-T. *Eur J Pharmacol* 1994; **258**: 281–284.
 28. Wu J. R., Zhou Z., Bittar E. E. Abolition with chloramine-T of inactivation in barnacle muscle fibers results in stimulation of the ouabain-insensitive sodium efflux. *Biochim Biophys Acta* 1992; **1112**: 99–104.
 29. Niemann P., Schmidtmayer J., Ulbricht W. Chloramine-T effect on sodium conductance of neuroblastoma cells as studied by whole-cell clamp and single-channel analysis. *Pflugers Arch* 1991; **418**: 129–136.
 30. Rouzair-Dubois B., Dubois J. M. Modification of electrophysiological and pharmacological properties of K channels in neuroblastoma cells induced by the oxidant chloramine-T. *Pflugers Arch* 1990; **416**: 393–397.
 31. Mozhayeva G. N., Naumov A. P., Kuryshv Yu. A., Nosyeva E. D. Some properties of sodium channels in neuroblastoma cells modified with scorpion toxin and chloramine-T. Single channel measurements. *Gen Physiol Biophys* 1990; **9**: 3–17.
 32. Wang G. K. Irreversible modification of sodium channel inactivation in toad myelinated nerve fibres by the oxidant chloramine-T. *J Physiol* 1984; **346**: 127–141.
 33. Wang G. K., Brodwick M. S., Eaton D. C. Removal of sodium channel inactivation in squid axon by the oxidant chloramine-T. *J Gen Physiol* 1985; **86**: 289–302.
 34. Cashmore A. R., Jarillo J. A., Wu Y.-L., Liu D. Cryptochromes: blue light receptors for plants and animals. *Science* 1999; **284**: 760–765.
 35. Horejsi R., Estelberger W., Mlekusch W., Moller R., Ottl K., Vrecko K., Reibnegger G. Effects of pteridines on chloramine-T-induced growth inhibition in *E. coli* strains: correlations with molecular structure. *Free Radic Biol Med* 1996; **21**: 133–138.
 36. Reibnegger G., Fuchs D., Murr C., Dierich M. P., Pfeleiderer W., Wachter H. Effects of pteridines on luminol-dependent chemiluminescence induced by chloramine-T. *Free Radic Biol Med* 1995; **18**: 515–523.
 37. Zgliczynski J. M., Olszowska E., Olszowski S., Stelmaszynska T., Kwasnowska E. A possible origin of chemiluminescence in phagocytosing neutrophils. Reaction between chloramines and H₂O₂. *Int J Biochem* 1985; **17**: 515–519.
 38. Fuursted K., Hjort A., Knudsen L. Evaluation of bactericidal activity and lag of regrowth (postantibiotic effect) of five antiseptics on nine bacterial pathogens. *J Antimicrob Chemother* 1997; **40**: 221–226.
 39. Gutierrez C. B., Rodriguez Barbosa J. I., Suarez J., Gonzalez O. R., Tascon R. I., Rodriguez Ferri E. F. Efficacy of a variety of disinfectants against *Actinobacillus pleuropneumoniae* serotype 1. *Am J Vet Res* 1995; **56**: 1025–1029.
 40. Selk S. H., Pogany S. A., Higuchi T. Comparative antimicrobial activity, in vitro and in vivo, of soft N-chloramine systems and chlorhexidine. *Appl Environ Microbiol* 1982; **43**: 899–904.
 41. Peters J., Spicher G. Model tests for the efficacy of disinfectants on surfaces. IV. communication: dependence of test results on the amount of contamination and the kind of active substance. *Zentralbl Hyg Umweltmed* 1998; **201**: 311–323.
 42. Nagl M., Gottardi W. Rapid killing of *Mycobacterium terrae* by N-chlorotaurine in the presence of ammonium is caused by the reaction product monochloramine. *J Pharm Pharmacol* 1998; **50**: 1317–1320.
 43. Bloomfield S. F., Arthur M. Interaction of *Bacillus subtilis* spores with sodium hypochlorite, sodium dichloroisocyanurate and chloramine-T. *J Appl Bacteriol* 1992; **72**: 166–172.
 44. Wede I., Widner B., Fuchs D. Neopterin derivatives modulate toxicity of reactive species on *Escherichia coli*. *Free Radic Res* 1999; **31**: 381–388.
 45. Kool J. L., Carpenter J. C., Fields B. S. Effect of monochloramine disinfection of municipal drinking water on risk of nosocomial Legionnaires' disease. *Lancet* 1999; **353**: 272–277.
 46. Richardson D., Bartlett C., Goutcher E., Jones C. H., Davison A. M., Will E. J. Erythropoietin resistance due to dialysate chloramine: the two-way traffic of solutes in haemodialysis. *Nephrol Dial Transplant* 1999; **14**: 2625–2627.
 47. Perez-Garcia R., Rodriguez-Benitez P. Chloramine, a sneaky contaminant of dialysate. *Nephrol Dial Transplant* 1999; **14**: 2579–2582.
 48. Beljaars P. R., Rondags T. M. Spectrodensitometric determination of chloramine-T in ice cream. *J Assoc Off Anal Chem* 1978; **61**: 1415–1418.
 49. Meinertz J. R., Schmidt L. J., Stehly G. R., Gingerich W. H. Liquid chromatographic determination of para-toluenesulfonamide in edible fillet tissues from three species of fish. *J AOAC Int* 1999; **82**: 1064–1070.
 50. Bundgaard-Nielsen K., Nielsen P. V. Fungicidal effect of 15 disinfectants against 25 fungal contaminants commonly found in bread and cheese manufacturing. *J Food Prot* 1996; **59**: 268–275.

51. Quiberone A., Suarez V. B., Reinheimer J. A. Inactivation of *Lactobacillus helveticus* bacteriophages by thermal and chemical treatments. *J Food Prot* 1999; **62**: 894–898.
52. Doultree J. C., Druce J. D., Birch C. J., Bowden D. S., Marshall J. A. Inactivation of feline calicivirus, a Norwalk virus surrogate. *J Hosp Infect* 1999; **41**: 51–57.
53. Krilov L. R., Harkness S. H. Inactivation of respiratory syncytial virus by detergents and disinfectants. *Pediatr Infect Dis J* 1993; **12**: 582–584.
54. Tyler R., Ayliffe G. A. A surface test for virucidal activity of disinfectants: preliminary study with herpes virus. *J Hosp Infect* 1987; **9**: 22–29.
55. Hunter D. T. Sodium hypochlorite in the treatment of herpes simplex virus infections. *Cutis* 1983; **31**: 328–332.
56. Jensen H., Thomas K., Sharp D. G. Inactivation of coxsackieviruses B3 and B5 in water by chlorine. *Appl Environ Microbiol* 1980; **40**: 633–640.
57. Muntianov V. P., Kriuk V. D., Belanov E. F. Disinfecting action of chloramine B on Marburg virus. *Vopr Virusol* 1996; **41**: 42–43.
58. Chepurnov A. A., Chuev Iu. P., P'iankov O. V., Efimova I. V. The effect of some physical and chemical factors on inactivation of the Ebola virus. *Vopr Virusol* 1995; **40**: 74–76.
59. Georges A. J., Baize S., Leroy E. M., Georges-Courbot M. C. Ebola virus: what the practitioner needs to know. *Med Trop* 1998; **58**: 177–186.
60. Hubalek Z. Some physical and chemical properties of Bhanja virus. *Acta Virol* 1986; **30**: 440–442.
61. Podoplekina L. E., Shutova N. A., Fyodorov Yu. V. Influence of several chemical reagents on lymphocytic choriomeningitis and Tacaribe viruses. *Virologie* 1986; **37**: 43–48.
62. Berman D., Hoff J. C. Inactivation of simian rotavirus SA11 by chlorine, chlorine dioxide, and monochloramine. *Appl Environ Microbiol* 1984; **48**: 317–323.
63. Gowda N. M., Trieff N. M., Stanton G. J. Inactivation of poliovirus by chloramine-T. *Appl Environ Microbiol* 1981; **42**: 469–476.
64. Tyler R., Ayliffe G. A., Bradley C. Virucidal activity of disinfectants: studies with the poliovirus. *J Hosp Infect* 1990; **15**: 339–345.
65. Sharp D. G., Leong J. Inactivation of poliovirus I (Brunhilde) single particles by chlorine in water. *Appl Environ Microbiol* 1980; **40**: 381–385.
66. Sagripanti J. L., Lightfoote M. M. Cupric and ferric ions inactivate HIV. *AIDS Res Hum Retroviruses* 1996; **12**: 333–337.
67. Steinmann J., Nehr Korn R., Meyer A., Becker K. Two in-vivo protocols for testing virucidal efficacy of handwashing and hand disinfection. *Zentralbl Hyg Umweltmed* 1995; **196**: 425–436.
68. Sattar S. A., Springthorpe V. S., Karim Y., Loro P. Chemical disinfection of non-porous inanimate surfaces experimentally contaminated with four human pathogenic viruses. *Epidemiol Infect* 1989; **102**: 493–505.
69. Shapshak P., McCoy C. B., Shah S. M., Page J. B., Rivers J. E., Weatherby N. L., Chitwood D. D., Mash D. C. Preliminary laboratory studies of inactivation of HIV-1 in needles and syringes containing infected blood using undiluted household bleach. *J Acquir Immune Defic Syndr* 1994; **7**: 754–759.
70. Bloomfield S. F., Smith-Burchnell C. A., Dalgleish A. G. Evaluation of hypochlorite-releasing disinfectants against the human immunodeficiency virus (HIV). *J Hosp Infect* 1990; **15**: 273–278.
71. Aranda-Anzaldo A., Viza D., Busnel R. G. Chemical inactivation of human immunodeficiency virus in vitro. *J Virol Methods* 1992; **37**: 71–81.
72. Resnick L., Veren K., Salahuddin S. Z., Tondreau S., Markham P. D. Stability and inactivation of HTLV-III/LAV under clinical and laboratory environments. *JAMA* 1986; **255**: 1887–1891.
73. Van Bueren J., Simpson R. A., Salman H., Farrelly H. D., Cookson B. D. Inactivation of HIV-1 by chemical disinfectants: sodium hypochlorite. *Epidemiol Infect* 1995; **115**: 567–579.
74. Stief TW, Slenczka W, Renz H, Klenk HD. Singlet oxygen (1O_2)-generating chloramines at concentrations that are tolerable for normal hemostasis function inactivate the lipid enveloped vesicular stomatitis virus in human blood. In: 3rd Symposium on the Biology of Endothelial Cells; Pathophysiology of the Endothelium: Vascular and Infectious Diseases, May 24–26, 2001, Giessen, Germany, Abstr. D10.
75. Docherty J. G., McGregor J. R., Purdie C. A., Galloway D. J., O'Dwyer P. J. Efficacy of tumoricidal agents in vitro and in vivo. *Br J Surg* 1995; **82**: 1050–1052.
76. McCaughan J. S., Jr Photodynamic therapy: a review. *Drugs Aging* 1999; **15**: 49–68.
77. de Vree W. J., Essers M. C., de Bruijn H. S., Star W. M., Koster J. F., Sluiter W. Evidence for an important role of neutrophils in the efficacy of photodynamic therapy in vivo. *Cancer Res* 1996; **56**: 2908–2911.
78. Iseki K., Tatsuta M., Iishi H., Baba M., Mikuni T., Hirasawa R., Yano H., Uehara H., Nakaizumi A. Attenuation by methionine of monochloramine-enhanced gastric carcinogenesis induced by *N*-methyl-*N'*-nitro-*N*-nitrosoguanidine in Wistar rats. *Int J Cancer* 1998; **76**: 73–76.
79. Soffritti M., Belpoggi F., Lenzi A., Maltoni C. Results of long-term carcinogenicity studies of chlorine in rats. *Ann N Y Acad Sci* 1997; **837**: 189–208.
80. Dunnick J. K., Melnick R. L. Assessment of the carcinogenic potential of chlorinated water: experimental studies of chlorine, chloramine, and trihalomethanes. *J Natl Cancer Inst* 1993; **85**: 817–822.
81. Suzuki H., Seto K., Mori M., Suzuki M., Miura S., Ishii H. Monochloramine induced DNA fragmentation in gastric cell line MKN45. *Am J Physiol* 1998; **275**: G712–G726.
82. Khan A. U., Kasha M. Singlet molecular oxygen evolution upon simple acidification of aqueous hypochlorite: application to studies on the deleterious health effects of chlorinated drinking water. *Proc Natl Acad Sci USA* 1994; **91**: 12362–12364.
83. Suzuki M., Asako H., Kubes P., Jennings S., Grisham M. B., Granger D. N. Neutrophil-derived oxidants promote leukocyte adherence in postcapillary venules. *Microvasc Res* 1991; **42**: 125–138.
84. Stapleton P. P., Redmond H. P., Bouchier-Hayes D. J. Myeloperoxidase (MPO) may mediate neutrophil adherence to the endothelium through upregulation of CD 11b expression – an effect downregulated by taurine. *Adv Exp Med Biol* 1998; **442**: 183–192.
85. Vogt W., Hesse D. Oxidants generated by the myeloperoxidase-halide system activate the fifth component of human complement, C5. *Immunobiology* 1994; **192**: 1–9.
86. Kilgore K. S., Ward P. A., Warren J. S. Neutrophil adhesion to human endothelial cells is induced by the membrane attack complex: the roles for P-selectin and platelet activating activating factor. *Inflammation* 1998; **22**: 583–598.

87. Bhakdi S. Complement and atherogenesis: the unknown connection. *Ann Med* 1998; **30**: 503–507.
88. Hazen S. L., Hsu F. F., Duffin K., Heinecke J. W. Molecular chlorine generated by the myeloperoxidase–hydrogen peroxide–chloride system of phagocytes converts low density lipoprotein cholesterol into a family of chlorinated sterols. *J Biol Chem* 1996; **271**: 23080–23088.
89. Tanen D. A., Graeme K. A., Raschke R. Severe lung injury after exposure to chloramine gas from household cleaners. *N Engl J Med* 1999; **341**: 848–849.
90. Shibata H., Sakamoto Y., Oka M., Kono Y. Natural antioxidant, chlorogenic acid, protects against DNA breakage caused by monochloramine. *Biosci Biotechnol Biochem* 1999; **63**: 1295–1297.
91. Carp H., Janoff A., Abrams W., Weinbaum G., Drews R. T., Weissbach H., Brot N. Human methionine sulfoxide-peptide reductase, an enzyme capable of reactivating oxidized α -1-proteinase inhibitor in vitro. *Am Rev Respir Dis* 1983; **127**: 301–305.
92. Schraufstatter I. U., Browne K., Harris A., Hyslop P. A., Jackso J. H., Quehenberger O., Cochrane C. G. Mechanisms of hypochlorite injury of target cells. *J Clin Invest* 1990; **85**: 554–562.
93. Thomas E. L. Myeloperoxidase, hydrogen peroxide, chloride antimicrobial system: nitrogen–chlorine derivatives of bacterial components in bactericidal action against *Escherichia coli*. *Infect Immun* 1979; **23**: 522–531.
94. Weiss S. J. Tissue destruction by neutrophils. *N Engl J Med* 1989; **320**: 365–376.
95. Ossana P. J., Test S. T., Matheson N. R., Regiani S., Weiss S. J. Oxidative regulation of neutrophil elastase- α -1-proteinase inhibitor interactions. *J Clin Invest* 1986; **77**: 1939–1951.
96. McKenzie S. J., Baker M. S., Buffinton G. D., Doe W. F. Evidence of oxidant-induced injury to epithelial cells during inflammatory bowel disease. *J Clin Invest* 1996; **98**: 136–141.
97. Hawkins C. L., Davies M. J. Hypochlorite-induced oxidation of proteins in plasma: formation of chloramines and nitrogen-centred radicals and their role in protein fragmentation. *Biochem J* 1999; **340**: 539–548.
98. Tatsumi T., Fliss H. Hypochlorous acid and chloramines increase endothelial permeability: possible involvement of cellular zinc. *Am J Physiol* 1994; **267**: H1597–H1607.
99. Cantin A. M. Taurine modulation of hypochlorous acid-induced lung epithelial cell injury in vitro. Role of anion transport. *J Clin Invest* 1994; **93**: 606–614.
100. Yajima N., Hiraishi H., Yamaguchi N., Ishida M., Shimada T., Terano A. Monochloramine-induced cytolysis to cultured rat gastric mucosal cells: role of glutathione and iron in protection and injury. *J Lab Clin Med* 1999; **134**: 372–377.
101. Nishiwaki H., Umeda M., Araki H., Fujita A., Furukawa O., Takeuchi K. Effect of monochloramine on recovery of gastric mucosal integrity and blood flow response in rat stomachs – relations to capsaicin-sensitive sensory neurons. *Life Sci* 1999; **65**: 1207–1216.
102. Nishiwaki H., Kato S., Sagumoto S., Umeda M., Morita H., Yoneta T., Takeuchi K. Ulcerogenic and healing impairing actions of monochloramine in rat stomachs: effects of zinc L-carnosine, polaprezinc. *J Physiol Pharmacol* 1999; **50**: 183–195.
103. Naito Y., Yoshikawa T., Fujii T., Boku Y., Yagi N., Dao S., Yoshida N., Kondo M., Matsui H., Ohtani-Fujita N., Sakai T. Monochloramine-induced cell growth inhibition and apoptosis in a rat gastric mucosal cell line. *J Clin Gastroenterol* 1997; **25**(Suppl. 1): S179–185.
104. Marcinkiewicz J., Nowak B., Grabowska A., Bobek M., Petrovska L., Chain B. Regulation of murine dendritic cell functions in vitro by taurine chloramine, a major product of the neutrophil myeloperoxidase-halide system. *Immunology* 1999; **98**: 371–378.
105. Liu Y., Schuller-Levis G., Quinn M. R. Monocyte chemoattractant protein-1 and macrophage inflammatory protein-2 production is inhibited by taurine chloramine in rat C6 glioma cells. *Immunol Lett* 1999; **70**: 9–14.
106. Park E., Schuller-Levis G., Jia J. H., Quinn M. R. Preactivation exposure of RAW 264.7 cells to taurine chloramine attenuates subsequent production of nitric oxide and expression of iNOS mRNA. *J Leukoc Biol* 1997; **61**: 161–166.
107. Park E., Alberti J., Quinn M. R., Schuller-Levis G. Taurine chloramine inhibits the production of superoxide anion, IL-6 and IL-8 in activated human polymorphonuclear leukocytes. *Adv Exp Med Biol* 1998; **442**: 177–182.
108. Ogino T., Kobuchi H., Sen C. K., Roy S., Packer L., Maguire J. J. Monochloramine inhibits phorbol ester-inducible neutrophil respiratory burst activation and T cell interleukin-2 receptor expression by inhibiting inducible protein kinase C activity. *J Biol Chem* 1997; **272**: 26247–26252.
109. Kim C., Park E., Quinn M. R., Schuller-Levis G. The production of superoxide anion and nitric oxide by cultured murine leukocytes and the accumulation of TNF- α in the conditioned media is inhibited by taurine chloramine. *Immunopharmacology* 1996; **34**: 89–95.
110. Liu Y., Tonna-DeMasi M., Park E., Schuller-Levis G., Quinn M. R. Taurine chloramine inhibits production of nitric oxide and prostaglandin E2 in activated C6 glioma cells by suppressing inducible nitric oxide synthase and cyclooxygenase-2 expression. *Brain Res Mol Brain Res* 1998; **59**: 189–195.
111. Quinn M. R., Park E., Schuller-Levis G. Taurine chloramine inhibits prostaglandin E2 production in activated RAW 264.7 cells by post-transcriptional effects on inducible cyclooxygenase expression. *Immunol Lett* 1996; **50**: 185–188.
112. Marcinkiewicz J., Grabowska A., Chain B. M. Modulation of antigen-specific T-cell activation in vitro by taurine chloramine. *Immunology* 1998; **94**: 325–330.
113. Bercz J. P., Jones L., Garner L., Murray D., Ludwig D. A., Boston J. Subchronic toxicity of chlorine dioxide and related compounds in drinking water in the nonhuman primate. *Environ Health Perspect* 1982; **46**: 47–55.
114. Fluck S., McKane W., Cairns T., Fairchild V., Lawrence A., Lee J., Murray D., Polgitiye M., Palmer A., Taube D. Chloramine-induced haemolysis presenting as erythropoietin resistance. *Nephrol Dial Transplant* 1999; **14**: 1687–1691.
115. Lockhart A. C. A hemodialysis patient with chloramine-induced hemolysis. A discussion of the mechanism. *N C Med J* 1998; **59**: 248–250.
116. Abrams W. R., Cohen A. B., Damiano V. V., Eliraz A., Kimbel P., Meranze D. R., Weinbaum G. A model of decreased functional α -1-proteinase inhibitor. Pulmonary pathology of dogs exposed to chloramine T. *J Clin Invest* 1981; **68**: 1132–1139.