

# In Reply

In their letter to the editor [1], Raj et al. suggested that extreme caution should be taken prior to infusing mesenchymal stem cells (MSCs) as a cell-based drug delivery vector into patients with cancer due to the potential risk for promoting disease progression. Although we tested unmodified MSCs with no therapeutic potential [2], the approach used in this study was considered safe given that infusing allogeneic MSCs 4–6 days preprostatectomy is unlikely to alter the natural course of an individual's prostate cancer. It cannot be emphasized enough that caution should always be paramount when translating a new therapeutic platform into the clinic, and we acknowledge that tumor promotion is a theoretical concern. However, it is important to consider that this is not strongly supported by available clinical evidence that includes hundreds of clinical trials and thousands of patients, although it is also not clear that rigorous human studies in cancer patients have been conducted to assess this specific risk. We agree that MSCs have well-documented pro-tumorigenic properties, including proangiogenic and immunosuppressive potential as published by ourselves and many others in multiple preclinical animal models [3-8]; however, this does not necessarily preclude their use as a cell-based drug delivery vector in future studies for men with metastatic prostate cancer, assuming tumor homing efficiency can be improved upon and cells can be engineered to deliver a cytotoxic payload [2]. In addition, there is sparse evidence of long-term engraftment and the most consistent result from prior studies testing MSCs is that they are safe and well-tolerated in patients across a broad disease spectrum [9]. This includes no evidence of cellular transformation or tumor promotion in patients receiving large numbers of infused MSCs (i.e.,  $2 \times 10^6$  cells per kilogram up to  $2 \times 10^8$  total); the first of whom were treated more than two decades ago [10-13]. For prostate cancer, the sheer incidence would argue that many of the men enrolled in these studies had diagnosed or occult disease, not to mention the thousands of patients that receive bone marrow transplants annually that also contain MSCs if an unselected product is used.

We would like to highlight that there is a distinct difference between the recruitment and/or expansion of endogenous (i.e., autologous) MSCs in tumors and the infusion of allogeneic MSCs as a cell-based drug delivery vector; the vast majority of which are entrapped in the lung and do not home to tumor tissue in robust numbers [14]. As stated above, there is no evidence of long-term MSC engraftment in any clinical study to date, at least in an allogeneic nonbone marrow transplant immunocompetent setting [11]. This is true of preclinical rodent and advanced primate models as well with limited persistence observed in vivo due to alloreactivity and death of the majority of cells <48 hours postinfusion [15-20]. Therefore, the direct effects of allogeneic MSCs are acute in nature; a point further documented by the weekly infusions required for the only clinical indication in which MSCs have demonstrated sufficient clinical efficacy to receive approval thus far-acute graft versus host

# LETTERS TO THE EDITOR

disease. This observation does not argue that acute effects cannot promote long-term beneficial outcomes in damaged tissue for example, by limiting acute fibrotic responses. Furthermore, if one believes that MSC recruitment is an ongoing process during malignant progression, as we have argued [6], and one assumes that infused allogeneic MSCs can supplant endogenously recruited ones that can promote disease progression, it would seem preferable to have those recruited cells carry with them a cytotoxic agent to counterbalance the pro-tumorigenic properties with a therapeutic effect [21].

In summary, although preclinical models have demonstrated that MSCs have pro-tumorigenic properties, primarily in syngeneic or immunocompromised coinoculation settings, clinical evidence does not support this being a significant risk in the context of allogeneic infusion for cell-based drug delivery. This is based on the collective experience of thousands of patients that have received MSCs in clinical trials worldwide for a variety of clinical indications with a well-tolerated safety profile and no evidence of ectopic tissue formation or tumor promotion [10-13]. It is also worth noting that there are three additional ongoing or completed phase I or I/II trials at independent institutions using genetically modified MSCs administered for therapeutic benefit in ovarian, lung, and gastrointestinal stromal tumor patients with no adverse events reported [14, 20, 22]. It is understood that these clinical observations are not definitive evidence of no risk and that more rigorous human studies with longer follow-up are required to be conclusive. However, they do support proceeding with clinical evaluation of these engineering cell platforms for tumors in organ sites that show high levels of entrapment or clearance postinfusion (e.g., lung, liver) or local delivery is an option (e.g., ovarian, glioblastoma), and potentially other tumor types if methods to enhance tumor tropism are developed or efficacy can be demonstrated at low levels of homing with transient engraftment. Future strategies that successfully enhance tumor tropism could also incorporate "suicide genes" or other approaches to eliminate the infused cells if necessary to further mitigate the potential for adverse outcomes. Despite the low perceived risk based on available evidence, continued development of these platforms should be predicated on careful monitoring of continued safety in treatment cohorts for unexpected sequelae. This is essential for further clinical translation and hopefully implementation of these innovative next-generation cell-based drug delivery strategies in a safe and effective manner.

## W. NATHANIEL BRENNEN

Department of Oncology, Johns Hopkins University School of Medicine, Baltimore, Maryland, USA

## MICHAEL T. SCHWEIZER

Department of Medicine, University of Washington, Seattle, Washington, USA; Clinical Research Division, Fred Hutchinson Cancer Research Center, Seattle, Washington, USA

This is an open access article under the terms of the Creative Commons Attribution-NonCommercial-NoDerivs License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made.

STEM CELLS TRANSLATIONAL MEDICINE 2019;8:739–740 www.StemCellsTM.com © 2019 The Authors. STEM CELLS TRANSLATIONAL MEDICINE published by Wiley Periodicals, Inc. on behalf of AlphaMed Press

#### HAO WANG

Department of Oncology, Johns Hopkins University School of Medicine, Baltimore, Maryland, USA

## TRINITY J. BIVALACQUA and ALAN W. PARTIN

Department of Urology, Johns Hopkins University School of Medicine, Baltimore, Maryland, USA

## SU JIN LIM, CAROLYN CHAPMAN, and REHAB ABDALLAH

Department of Oncology, Johns Hopkins University School of Medicine, Baltimore, Maryland, USA

## **OREN LEVY**

Center for Nanomedicine and Division of Engineering in Medicine, Department of Medicine, Brigham and Women's Hospital, Harvard Medical School, Boston, Massachusetts, USA Harvard Stem Cell Institute, Cambridge, Massachusetts, USA Division of Health Sciences and Technology, Harvard-MIT, Cambridge, Massachusetts, USA

### NEIL A. BHOWMICK

Department of Medicine, Cedars-Sinai Medical Center, Los Angeles, California, USA

#### JEFFREY M. KARP

Center for Nanomedicine and Division of Engineering in Medicine, Department of Medicine, Brigham and Women's Hospital, Harvard Medical School, Boston, Massachusetts, USA Harvard Stem Cell Institute, Cambridge, Massachusetts, USA Division of Health Sciences and Technology, Harvard-MIT, Cambridge, Massachusetts, USA

#### ANGELO DE MARZO

Department of Pathology, Johns Hopkins University School of Medicine, Baltimore, Maryland, USA

## JOHN T. ISAACS and SAMUEL R. DENMEADE 回

Department of Oncology, Johns Hopkins University School of Medicine, Baltimore, Maryland, USA

### **DISCLOSURE OF POTENTIAL CONFLICTS OF INTEREST**

The authors indicated no potential conflicts of interest.

#### REFERENCES

**1** Raj AJ, Kheur S, Bhonde R et al. Use of Bone Marrow-Derived Mesenchymal Stem Cells in Prostate Cancer Could Increase the Risk of Cancer Progression. STEM CELLS TRANSLATIONAL MEDICINE 2019;8:737–738.

**2** Schweizer MT, Wang H, Bivalacqua TJ et al. A phase I study to assess the safety and cancer-homing ability of allogeneic bone marrow-derived mesenchymal stem cells in men with localized prostate cancer. STEM CELLS TRANSLATIONAL MEDICINE 2019;8:441–449.

**3** Klopp AH, Gupta A, Spaeth E et al. Concise review: Dissecting a discrepancy in the literature: Do mesenchymal stem cells support or suppress tumor growth? STEM CELLS 2011;29:11–19.

**4** Bergfeld SA, DeClerck YA. Bone marrow-derived mesenchymal stem cells and the tumor microenvironment. Cancer Metastasis Rev 2010;29:249–261.

**5** Brennen WN, Nguyen H, Dalrymple SL et al. Assessing angiogenic responses induced by primary human prostate stromal cells in a three-dimensional fibrin matrix assay. Oncotarget 2016;7: 71298–71308.

**6** Brennen WN, Zhang B, Kulac I et al. Mesenchymal stem cell infiltration during neoplastic transformation of the human prostate. Oncotarget 2017;8:46710–46727.

**7** Krueger TE, Thorek DLJ, Meeker AK et al. Tumor-infiltrating mesenchymal stem cells: Drivers of the immunosuppressive tumor microenvironment in prostate cancer? Prostate 2019;79:320–330.

**8** Ridge SM, Sullivan FJ, Glynn SA. Mesenchymal stem cells: Key players in cancer progression. Mol Cancer 2017;16:31.

**9** Squillaro T, Peluso G, Galderisi U. Clinical trials with mesenchymal stem cells: An update. Cell Transplant 2016;25:829–848.

**10** Wang Y, Han ZB, Song YP et al. Safety of mesenchymal stem cells for clinical application. Stem Cells Int 2012;2012:652034.

**11** von Bahr L, Batsis I, Moll G et al. Analysis of tissues following mesenchymal stromal cell therapy in humans indicates limited long-term engraftment and no ectopic tissue formation. STEM CELLS 2012;30: 1575–1578.

**12** Karantalis V, Hare JM. Use of mesenchymal stem cells for therapy of cardiac disease. Circ Res 2015;116:1413–1430.

**13** Lalu MM, McIntyre L, Pugliese C et al. Safety of cell therapy with mesenchymal stromal cells (SafeCell): A systematic review and meta-analysis of clinical trials. PLoS One 2012;7:e47559.

**14** Krueger TEG, Thorek DLJ, Denmeade SR et al. Concise review: Mesenchymal stem cell-based drug delivery: The good, the bad, the ugly, and the promise. STEM CELLS TRANSLATIONAL MEDICINE 2018;7: 651–663.

**15** Lee RH, Pulin AA, Seo MJ et al. Intravenous hMSCs improve myocardial infarction in mice because cells embolized in lung are activated to secrete the anti-inflammatory protein TSG-6. Cell Stem Cell 2009;5:54–63.

**16** Huang XP, Sun Z, Miyagi Y et al. Differentiation of allogeneic mesenchymal stem cells induces immunogenicity and limits their long-term benefits for myocardial repair. Circulation 2010;122:2419–2429.

17 Isakova IA, Lanclos C, Bruhn J et al. Allo-reactivity of mesenchymal stem cells in rhesus macaques is dose and haplotype dependent and limits durable cell engraftment in vivo. PLoS One 2014;9: e87238.

**18** Sivanathan KN, Gronthos S, Rojas-Canales D et al. Interferongamma modification of mesenchymal stem cells: Implications of autologous and allogeneic mesenchymal stem cell therapy in allotransplantation. Stem Cell Rev 2014;10:351–375.

**19** Ankrum JA, Ong JF, Karp JM. Mesenchymal stem cells: Immune evasive, not immune privileged. Nat Biotechnol 2014;32:252–260.

**20** Mohr A, Zwacka R. The future of mesenchymal stem cell-based therapeutic approaches for cancer—From cells to ghosts. Cancer Lett 2018;414:239–249.

**21** Levy O, Brennen WN, Han E et al. A prodrug-doped cellular Trojan Horse for the potential treatment of prostate cancer. Biomaterials 2016;91:140–150.

22 von Einem JC, Peter S, Gunther C et al. Treatment of advanced gastrointestinal cancer with genetically modified autologous mesenchymal stem cells—TREAT-ME-1—A phase I, first in human, first in class trial. Oncotarget 2017;8:80156–80166.

http://dx.doi.org/10.1002/sctm.19-0068