Machine learning sparse reaction-diffusion models from stochastic dynamics and spatiotemporal patterns

Bedri Abubaker-Sharif^{1,2}, Peter N. Devreotes^{2,3*}, Pablo A. Iglesias^{1,2,4**}

- 1 Department of Biomedical Engineering, School of Medicine, Johns Hopkins University, Baltimore, MD, 21205, USA
- 2 Department of Cell Biology, School of Medicine, Johns Hopkins University, Baltimore, MD, 21205, USA
- **3** Department of Biological Chemistry, School of Medicine, Johns Hopkins University, Baltimore, MD, 21205, USA
- 4 Department of Electrical and Computer Engineering, Whiting School of Engineering, Johns Hopkins University, Baltimore, MD, 21218, USA

Abstract

In recent years, live-cell imaging has generated detailed spatiotemporal datasets of biochemical networks within cells. These networks often exhibit characteristics of spatially-distributed excitable systems, with propagating waves of signaling activity that govern processes such as cell migration, division, and other essential physiological functions. Traditionally, these reaction-diffusion systems have been modeled using stochastic partial differential equations incorporating spatial Langevin-type dynamics. Although these knowledge-based models have provided valuable insights, they are typically not directly inferred from experimental data. In this study, we introduce and apply two data-driven methodologies for learning the structure and parameters of spatial Langevin equation (SLE) models: 1) Kramers-Moyal regression, an adaptation of an established approach to fit microscale stochastic dynamics, and 2) wave features optimization, a novel approach to match macroscale spatiotemporal patterns. Unlike black-box neural network models that predict system behavior but provide no mechanistic insight, this approach estimates nonlinear model equations that directly relate to system structure and dynamics. As a proof-of-concept, we focus on simulation datasets derived from two stochastic reaction-diffusion models: one based on the FitzHugh-Nagumo equations (FHN model) and another on a biochemically adapted version of the FHN equations (FR model). Our results demonstrate that optimizing stochastic dynamics and wave patterns enables the accurate estimation of SLE model structure and parameters directly from 1D and 2D spatial data. With sparsity enforcement, we also identify novel sparse reaction-diffusion models for excitable systems. We show that this approach effectively approximates system behavior even when working with datasets similar to experimental conditions, with low temporal resolution and unobserved molecular components. By leveraging machine learning techniques for robust estimation of excitable reaction-diffusion models from spatiotemporal data, this work enhances our ability to model and understand the complex systems that regulate cell behavior.

Introduction

Since Hodgkin and Huxley's groundbreaking discovery of the mechanisms underlying nerve cell activity, the theory of excitability, describing dynamical systems that display common fundamental characteristics such as nonlinearity, all-or-none responses, and refractory periods, has been applied

^{*}pnd@jhmi.edu, **pi@jhu.edu

across various scientific disciplines, including physics, chemistry, and biology [1–11]. When these systems are both stochastic and spatially-distributed, they can spontaneously generate a diverse range of patterns, waves, and oscillations that play a crucial role in regulating physiological processes and behavior in various cell types. Mathematical models have greatly aided the understanding of these systems, however, a significant challenge remains in developing data-driven models capable of accurately describing stochastic, spatially-distributed excitable systems. Such models would provide powerful tools for the study of dynamic biological processes at the cellular level as well as other real-world excitable stochastic systems.

Modeling the biochemical network in cells has traditionally focused on replicating the general patterns observed in experiments. In many cell types, extensive imaging data indicates that biomolecules can react and diffuse as a coordinated excitable system, self-organizing into dynamic domains in the cell cortex that propagate as waves [12–21]. For example, in migrating cells, "front" network components, such as Ras and PIP3, generally form the leading edge of the wave, while "refractory" components, such as PKBA potentially, accumulate to varying degrees behind the wave front; other "back" components appear in the inactive regions of the cell cortex both ahead of and behind waves [22–27]. Interestingly, these zones approximate the general patterns observed in activator-inhibitor models such as the FitzHugh-Nagumo (FHN) model with a single positive feedback loop and a single delayed negative feedback loop [11]. Previous studies employing activator-inhibitor models have been used extensively to model a wide array of wave dynamics [28–36]. These conceptual models are simulated as stochastic partial differential equations, with spatial Langevin-type dynamics. Tuning the structure and parameters can yield simulations that replicate waves and patterns observed in various wild-type and experimental perturbations of cells. Model structure and parameters are adjusted manually based on experience and observation. Nonetheless, this iterative process is often time-intensive and does not facilitate the automated discovery of new models with potentially more insightful structures that better capture the dynamics and patterns observed experimentally in cells.

To address the challenge of learning nonlinear models from data, several approaches have been developed in the machine learning literature [37,38]. Unlike most neural-network approaches, these system identification approaches can provide key quantitative insights regarding network structure and dynamics. However, many of these techniques focus on deterministic models [39–44], which have limited utility for complex stochastic systems. On the other hand, approaches for stochastic data usually rely on strict conditions that are often unmet in experimental datasets, such as rapid sampling rates and observations being available for all modeled variables [45,46]. Additionally, there has been limited testing on spatially-distributed models. In recent years, there has also been an advancement of Fokker-Planck (FP)-based approaches for learning the structure and parameters of stochastic differential equations (SDEs) [46–51], yet these do not consider stochastic partial differential equations. In cell migration research, some studies have adapted these FP-based approaches to learn macroscale models, such as models of cell trajectories in chemical gradients [52]. However, few studies address learning stochastic partial differential equation models of excitable systems.

Here, we combine two distinct but complementary approaches: 1) optimizing stochastic dynamics to match microscale network structure, and 2) optimizing spatiotemporal features to match macroscale wave patterns. For the first approach, we use what we refer to as Kramers-Moyal regression, based on previous FP-based algorithms [46,47]. In this algorithm, we learn stochastic models by fitting the deterministic ("drift") and stochastic ("diffusivity") terms of SDEs to corresponding data-derived conditional expectations, also known as Kramers-Moyal (KM) averages, using linear and nonlinear regression. To learn model structure together with model parameters, we enforce sparsity over a library of functional terms using a reverse greedy search algorithm [46,47], and estimate optimal sparsity using Bayesian information criterion (BIC) instead of the cross-validation techniques used in previous work. Additionally, unlike past studies which did not consider spatially-distributed models nor excitable models with rational functions, we adapt these algorithms to learn stochastic reaction-diffusion models with polynomial and rational reaction terms.

11

12

13

14

15

16

17

19

20

21

22

23

24

25

26

27

28

29

30

31

32

37

39

41

43

45

47

49

50

51

52

53

54

In the second approach, we implement a novel Monte Carlo strategy to tune model parameters by optimizing for spatiotemporal wave features. For initialization, we use initial parameters obtained from either KM regression when available, or from previous knowledge. At each iteration, we generate a reconstruction dataset consisting of multiple simulations, and then compute various spatiotemporal metrics such as wave activity, number of firings, and autocorrelation functions. We then optimize a custom objective function based on the relative error in spatiotemporal patterns between the training dataset and the reconstruction dataset in order to estimate optimal reaction-diffusion model parameters.

Our dual strategy here provides a data-driven approach to learning spatially-distributed, stochastic excitable network models. For proof-of-concept, we focus on two-component activator-inhibitor based reaction-diffusion models simulated in 1D and 2D space. These models mimic the type of waves observed in spatiotemporal data acquired from confocal and total internal reflection fluorescence (TIRF) microscopy of cells. To explore the effectiveness in the context of some of the complexities in experimental data, we test cases where model structure is held fixed (parameter estimation/model identification) or variable (model learning), as well as datasets with lower sampling rates and partial observation of system species. This study provides an important, focused application of machine learning approaches for estimating stochastic reaction-diffusion models of excitable systems directly from spatiotemporal data.

Methods

Stochastic reaction-diffusion models

The overall strategy employed for learning stochastic reaction-diffusion models from data is illustrated in Figure 1. Stochastic simulations were first generated based on two distinct models of excitable systems: the classical FitzHugh-Nagumo (FHN) model [53,54], and a modified version with "front" (F) and "refractory" (R) species for biochemical processes, which we refer to as the FR model [28,55]. These activator-inhibitor models (activator x_A , inhibitor x_I) were implemented as spatial Langevin equations in the following Itô differential form (Fig. 1a):

$$dx_A = (D_A \nabla^2 x_A + s_A(x_A, x_I, \theta_A)) dt + g_A dQ_A$$

$$dx_I = (D_I \nabla^2 x_I + s_I(x_A, x_I, \theta_I)) dt + g_I dQ_I$$
(1)

For clarity, we denote the activator and inhibitor species as u and v in the FHN model and as f and r in the FR model. As in previous studies [11,56], the model structures for the FHN and FR models are distinguished by the reaction terms such that,

FHN model reaction terms
$$\begin{cases} s_u(u, v, \theta_u) = a_1 u + a_2 v + a_3 u^2 + a_4 u^3 \\ s_v(u, v, \theta_v) = b_1 u + b_2 v \end{cases}$$
 (2)

FR model reaction terms
$$\begin{cases} s_f(f, r, \theta_f) = a_1 f + a_2 f r + \frac{a_3 f^2}{f^2 + a_4} + a_5 \\ s_r(f, r, \theta_r) = b_1 f + b_2 r \end{cases}$$
 (3)

where reaction parameters $\theta_A = \{a_i\}$ and $\theta_I = \{b_i\}$. Note that while both models are nonlinear in the state variables, the FHN model is linear in the parameters, while the FR model is nonlinear in

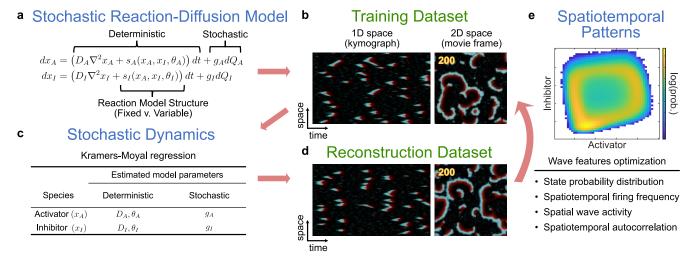


Fig 1. General approach for learning stochastic reaction-diffusion models from data a. A stochastic reaction-diffusion system with two species: an activator (x_A) and an inhibitor (x_I) . The system was modeled with spatial Langevin dynamics and a set of known, constant reaction parameter vectors (θ_A, θ_I) and spatial diffusion parameters (D_A, D_I) . b. Examples of spatiotemporal simulations based on the equations in a. Multiple runs were performed to generate a training dataset. The kymograph of the 1D simulation denotes 400 s of simulation; the yellow label marks the frame time in the 2D simulation. c. Data-driven learning using Kramers-Moyal (KM) averages. Regression was used to fit model functions to KM averages computed for each simulation, providing a set of estimated model parameters. d. Example spatiotemporal simulations obtained using optimal parameters from c. Multiple runs were performed to generate a reconstruction dataset. e. Sample metrics used during wave features optimization to assess the similarity of the training vs. reconstruction datasets. Example of the joint probability distribution over x_A and x_I from a single FHN simulation is shown. These metrics were also used to validate the accuracy of reconstructions from KM regression-based parameters.

the parameters because of the more biochemically relevant Michaelis-Menten-like term. Additionally, while the FHN model variables are based on model membrane potential and take on negative values, the FR model variables describe the concentrations of biochemical species and generally remain non-negative during simulation.

Generating simulations

To generate a training dataset, 50 simulations were obtained for each model in 1D and 2D space (Fig. 1b). Each simulation was carried out by integrating the two-component (activator and inhibitor) spatial Langevin equation (SLE) from 1 over a uniform grid of points in either 1D or 2D space, using the Euler-Maruyama method. The simulations were performed with an integration time step of $\Delta t = 0.01\,\mathrm{s}$ and ran for a total duration of 400 seconds. Data was saved for all analyses at a "recording" time-step of $\tau_0 = 0.1\,\mathrm{s}$ for FHN simulations and $\tau_0 = 0.05\,\mathrm{s}$ for FR simulations. These simulations were visualized as kymographs for 1D simulations and videos for 2D simulations, as represented in Fig. 1b. For all 1D simulations, 500×1 grid points were used with $\Delta x = 1$, and for all 2D simulations, a 100×100 grid was used with $\Delta x = \Delta y = 1$. The ground truth SLE parameters used for all training simulations were obtained from iterative tuning and previous studies and are listed in Table 1 and Table 2. Additional details are described in S1 Methods.

Identifying and learning models using Kramers-Moyal (KM) regression with sparsity enforcement

For model identification, we assume a fixed model structure and estimate the parameters of the deterministic and stochastic components of the spatial Langevin equation (SLE) in 1 using data-derived stochastic dynamics (Fig. 1c). For stationary processes with Gaussian white noise, the analytic components of SDEs (the deterministic, or "drift", term and the stochastic, or "diffusivity", term) are directly related to data-based conditional moments known as Kramers-Moyal (KM) coefficients, or KM averages [46,51] (S1 Methods). There are two KM averages that are computed for each species (activator and inhibitor), and these are linked to the deterministic and stochastic components, respectively, of the SLE for each species. To estimate the deterministic parameters for the FHN model, we applied linear regression, while for the FR model, we applied nonlinear regression using the Levenberg-Marquardt algorithm. For both the FHN and FR models, we estimated the stochastic parameter of the activator and inhibitor components in the SLE model directly using the second KM averages (S1 Methods).

Model learning involved estimating the reaction model structure as well as all other SLE parameters using sparse KM regression. A library of up to third-order polynomial reaction terms were used to define an initial set of variable model structures (Fig. 1a), from which a sparse model was learned. Specifically, we considered the following reaction terms for both the activator and inhibitor equations:

Polynomial reaction terms
$$\begin{cases} s_n(x_A, x_I, \theta_n) = a_1 + a_2 x_A + a_3 x_I + a_4 x_A^2 + a_5 x_A x_I + a_6 x_I^2 \\ + a_7 x_A^3 + a_8 x_A^2 x_I + a_9 x_A x_I^2 + a_{10} x_I^3 \end{cases}$$
(4)

where $\theta_n = \{a_1, a_2, ..., a_{10}\}$ denotes the reaction parameters.

In order to learn a sparse polynomial reaction-diffusion model, we used a reverse-greedy search algorithm from [46] with modifications (S1 Methods). Linear regression was performed iteratively, with elimination of the reaction parameter smallest in absolute value at each step. The spatial diffusion term was retained and only removed at the final iteration. Optimal reaction model sparsity was estimated by minimizing the Bayesian information criterion (BIC), which balances complexity and accuracy [38,57]. The optimal model structure was determined using the top model parameters retained over all simulations at a given level of sparsity. We also identified candidate sparse model structures by considering models with fewer terms than the estimated optimal sparsity.

To assess variability and compare with ground truth parameters, mean and standard deviations of all estimated parameters were calculated over 50 simulations. The estimated parameters were then used to generate an additional 20-50 simulations, constituting a reconstruction dataset (Fig. 1d).

Quantification and optimization of wave features (WF) for validating and estimating models

A comprehensive, quantitative comparison was performed to evaluate the quality of the reconstructed simulations against the original training dataset. This assessment involved measuring the joint probability distribution of model species, spatiotemporal firing frequencies, and spatiotemporal autocorrelations. Together, these metrics of the patterns observed in the simulations provided a rigorous validation of estimated models (Fig. 1e, S1 Methods). Wilcoxon rank-sum tests were conducted as a nonparametric test to compare the distribution of wave metrics of reconstruction simulations to that of training simulations.

Using these metrics, we also developed a Monte Carlo optimization scheme to estimate model parameters that generate reconstructions with wave features that best match the wave features in the training data. A Nelder-Mead simplex search was performed to minimize a custom objective function involving the sum of squared relative errors in each wave feature (S1 Methods). This WF optimization approach was applied in three cases of model estimation: 1) estimation after sparse KM regression for FHN-1D and FR-1D models; 2) estimation for FHN-2D models from temporally subsampled data; and 3) estimation for FHN-2D models temporally subsampled data only observing the activator or the inhibitor. After optimization, to assess the quality of parameter estimates in each case, 20 reconstructions were generated, and wave features were analyzed.

Results

Visualization of spatiotemporal data from excitable systems

Figure 2 presents visualizations of the data observed in 1D and 2D simulations of the FHN and FR models. Three different visualizations are shown: the spatiotemporal trajectories, the temporal dynamics at a single spatial grid point, and phase portraits of system trajectories at a single grid point. The spatiotemporal trajectories of 1D simulations were visually represented fully in kymographs, where the x-axis is time, and the y-axis represents spatial points, while for 2D simulations, selected frame sequences of a movie were shown.

Dynamic features spontaneously emerged in both the FHN-1D and FR-1D kymographs (Fig. 2a,c) with the formation of 1D waves originating at random initiation points and expanding in V-shaped patches in space-time with characteristic length scales in both time and space. The activator and the inhibitor in these two models demonstrated similar spatial profiles as visible by the significant overlap in the vertical propagation distance of both the activator and the inhibitor for any given wave (Fig. 2a,c). Temporally, FHN-1D wave patterns for the activator and the inhibitor showed behavior characteristic of excitability and exhibited similar duration as indicated by the strong horizontal overlap in the patches. The temporal signal at a single grid point in the left plot in Figure 2b shows this more clearly, with activator and inhibitor spikes lasting about 30 and 40 seconds, respectively, and separated by a delay of \sim 20 seconds. In contrast, in the FR-1D model, the wave patterns for the activator lasted significantly shorter (\sim 20 s) compared to the inhibitor (\sim 60 s), and with shorter delay (\sim 5 s) between the two species (Fig. 2d, left).

Examining trajectories in phase space, the FHN-1D and FR-1D models demonstrated similar excursions during a firing when the system crossed an activation threshold as a result of noise or spatial diffusion (Fig. 2b,d). The nullclines of the dynamics of the reaction terms (without spatial diffusion) are displayed together with the trajectories at a single grid point as a wave passed through. The behaviors in the phase plane are indicative of a system with a stable equilibrium point near a Hopf bifurcation point. As two waves passed through the single grid point shown in orange in

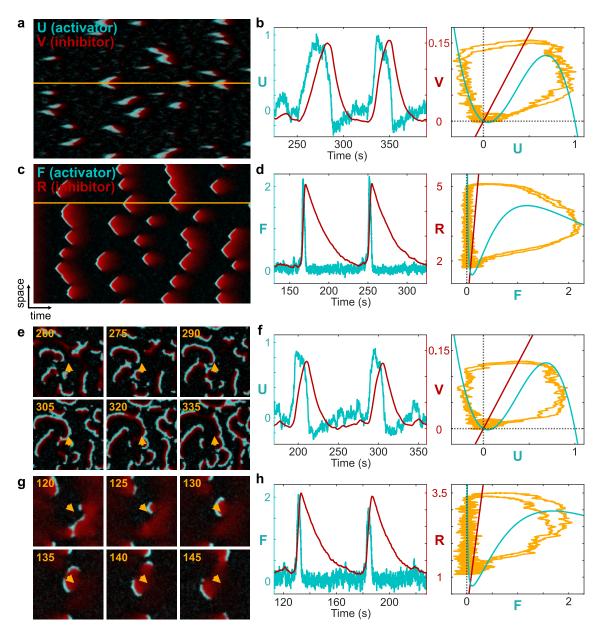


Fig 2. Visualization of training simulations of excitable reaction-diffusion models simulated in 1D and 2D space.

a. Kymograph of a 400 s stochastic reaction-diffusion simulation based on the classical FitzHugh-Nagumo (FHN) model over a 1D spatial domain (FHN-1D). Activator (U) and inhibitor (V) signals are shown in cyan and red, respectively. b. Signal values of U and V plotted together over time (left), and trajectories of U and V plotted in the phase plane (right). Both correspond to signals at the grid point indicated in a by the yellow line. c-d. Similar visualizations of the FR-1D model. F and R denote the activator and inhibitor, respectively. e-h. Frames from FHN (e) and FR (g) models simulated over a 2D spatial domain for 400 s. Time stamps indicate simulation time (seconds). Yellow arrows indicate spatial points at which trajectories were plotted over time (f,h, left) and in the phase plane (f,h, right).

Figure 2a and were plotted in time (Fig. 2b, left), the FHN-1D activator and inhibitor traced a stereotypical trajectory in the phase plane (Fig. 2b, right). The excursion began from the equilibrium point (intersection of the nullclines) and moved to the right as the activator increased while the inhibitor remained low, then moved upward as the delayed inhibitor accumulated, then moved left as the activator decreased by the negative feedback from the inhibitor, before returning to the equilibrium. In the FR-1D simulations, for two waves passing through a grid point shown in Figure 2c, we displayed the time series and corresponding trajectories in the phase plane (Fig. 2d). Compared to the FHN model, the FR model parameters yielded faster activator dynamics and slower inhibitor decay.

Excitability in 2D spatial simulations formed traveling waves with rich features. FHN-2D showed circular waves that spontaneously initiated and propagated radially (Fig. 2e,S1 Video). Over time, as a result of annihilations when inhibitor signals from two waves merged, large waves were split into patches propagating through the spatial domain (Fig. 2e). FR-2D simulations showed similar traveling waves but with limited propagation range (Fig. 2g, S2 Video). The inhibitor R appeared to accumulate rapidly and with long duration, hindering propagation and extinguishing the waves (Fig. 2g). The temporal signals at a single grid point in Figure 2f,h show this distinction more clearly between the FHN-2D and FR-2D models. In FHN-2D, the activator and inhibitor lasted $\sim 30-50$ seconds and were separated by a slight delay of ~ 10 seconds (Fig. 2f, left). In contrast, in the FR-2D model, the wave patterns for the activator were significantly shorter (\sim 5–10 s) compared to the inhibitor ($\sim 40 \,\mathrm{s}$), with a time delay under 5 seconds (Fig. 2h, left). The trajectories in the phase plane at the same grid point plotted in Figure 2e,g for both FHN-2D and FR-2D, were remarkably similar to those found in 1D spatial simulations (Fig. 2f,h, right). The FR-2D simulation showed some slight differences compared to FR-1D in phase plane analysis with the trajectories at a single grid point not settling to the equilibrium point of the spatially-homogeneous reaction equation nullclines (Fig. 2h, right).

Estimating FHN reaction-diffusion model parameters from stochastic dynamics

Kramers-Moyal regression, a parameter estimation algorithm based on stochastic dynamics, was initially applied to FHN-1D and FHN-2D simulations, assuming a known reaction-diffusion model structure for each. A training dataset of 50 1D and 2D simulations recorded every $\tau_0=0.1\,\mathrm{s}$, was used for analysis (Methods). From the kymographs or movies, Kramers-Moyal (KM) averages were computed over a binned Laplacian-extended state space (S1 Methods). The first KM average, $H_n^{(1)}$, relates to the deterministic component of the FHN spatial Langevin equation (SLE), as a function of the binned values of u, v, and the Laplacian of the corresponding n-th species. For visualization, projected KM averages were displayed as a heatmap over the activator-inhibitor phase plane. As seen in S1 Fig, the heatmap for $H_u^{(1)}$ and $H_v^{(1)}$ matched the stochastic dynamics observed in the time series at single grid points (Fig. 2b). The $H_u^{(2)}$ heatmap corresponding to the constant stochastic term in the activator SLE was relatively constant, while the $H_v^{(2)}$ heatmap for the inhibitor was approximately zero, owing to the absence of stochastic noise in the inhibitor dynamics.

Linear regression over a library of basis functions, corresponding precisely to the terms in the FHN model, was performed on these KM averages to identify an optimal set of parameters that minimized the mean squared error (MSE) between empirical KM averages and the corresponding deterministic/stochastic model terms (theoretical KM averages). The parameters, computed separately for each simulation, demonstrated remarkable accuracy and precision when compared to the ground truth values used in simulation generation, even across a wide range of magnitudes (Table 1).

To validate these parameters further, a set of 50 FHN-1D and FHN-2D simulations were generated based on the estimated parameters, forming a reconstruction dataset (Fig. 3). Comparisons between representative kymographs from the training and reconstruction datasets (Fig. 3a,e) revealed the inherent stochastic nature of the system, with no two simulations being

Table 1. True and estimated spatial Langevin equation (SLE) parameters for FHN-1D and FHN-2D models using Kramers-Moyal regression on training simulations (N=50).

Activator SLE: $du = D_u \nabla^2 u + a_1 u + a_2 v + a_3 u^2 + a_4 u^3 + g_u dQ_u$.

Inhibitor SLE: $dv = D_v \nabla^2 v + b_1 u + b_2 v + g_v dQ_v$.

	D_u	a_1	a_2	a_3	a_4	g_u
FHN-1D						_
Ground truth	1.4	-0.2	-2	2.2	-2	0.1
Mean	1.382	-0.193	-1.984	2.163	-1.969	0.103
SD	0.035	0.025	0.070	0.089	0.091	0.001
FHN-2D						
Ground truth	0.13	-0.2	-2	2.2	-2	0.0725
Mean	0.129	-0.189	-1.986	2.140	-1.942	0.0735
SD	0.003	0.014	0.031	0.048	0.048	0.001
	D_v	b_1	b_2	g_v		
FHN-1D						_
Ground truth	1.2	0.02	-0.06	0		
Mean	1.194	0.0200	-0.0599	0.0006		
SD	0.005	2.2×10^{-5}	1.3×10^{-4}	2.5×10^{-5}		
FHN-2D						
Ground truth	0.00625	0.0150	-0.0450	0		
Mean	0.00626	0.0150	-0.0449	0.0006		
SD	1.00×10^{-4}	9.16×10^{-6}	6.08×10^{-5}	9.20×10^{-6}		

identical. Feature analysis was employed for a more nuanced comparison of the two datasets. Characterization of these spatiotemporal features involved computing metrics for temporal firing frequency, spatial wave activity, and spatiotemporal autocorrelations (Fig. 3b–d,f–h).

For both 1D and 2D simulations, the FHN training and reconstruction data displayed similar distributions for the number of firing events at a grid point, with the peak centered around 1 or 3 firings for 1D or 2D simulations, respectively, and a slight rightward skew for 1D data due to a small proportion of grid points with greater than 2 firings (Fig. 3b,f). The spatial wave activity, as indicated by the proportion of spatial points above the threshold at any given time, was remarkably consistent, with variability centered around 5% for 1D models and 20% for 2D models (Fig. 3c,g). Wave activity was particularly consistent in the 2D model in which oscillations continued through the last 300 s of training and reconstruction simulations. Temporal and spatial autocorrelations were also strikingly similar, indicating comparable pattern length scales in time (1D: \sim 40-50 s; 2D: \sim 50-70 s) and space (1D/2D: \sim 5-10 px.) for both the training and reconstruction FHN datasets (Fig. 3d,h).

Estimating FR reaction-diffusion model parameters from stochastic dynamics

The process of identifying model parameters for the FR model, under the assumption of a known reaction-diffusion model structure, involved the application of the KM regression algorithm with modifications. Given the faster dynamics, the FR training dataset was analyzed using simulations recorded at $\tau_0 = 0.05 \, \text{s}$. KM averages were computed from the kymographs or movies, and nonlinear regression (using the Levenberg-Marquardt algorithm) was subsequently performed to determine the optimal set of parameters minimizing the MSE between the empirical KM averages and the corresponding deterministic/stochastic terms in the FR spatial Langevin equation (Methods). The comparison of estimated parameters to the ground truth values of training data revealed their remarkable accuracy and precision across both 1D and 2D spatial FR simulations (Table 2). The

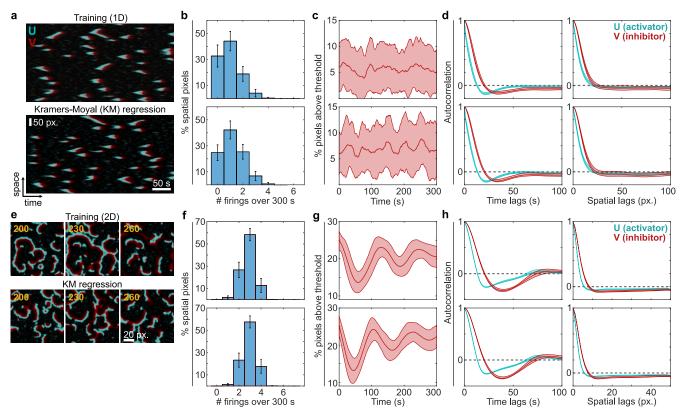


Fig 3. Reconstruction and validation with Kramers-Moyal (KM) based estimates of FHN reaction-diffusion model parameters.

a. Representative kymographs from a 400 s simulation in the FHN-1D training dataset (top) and from a reconstruction dataset generated by FHN reaction-diffusion model parameters estimated using KM regression (bottom). (N=50 each; 50 spatial pixels (px.) and 50 s fiducial markers apply to both kymographs).
b. Temporal firing frequency, computed as the distribution of the number of firings in a 300 s interval at spatial points over all simulations in the FHN-1D training data (top) and KM-based reconstructions (bottom). (N=50 each; error bars indicate SD).
c. Spatial wave activity, computed as the percentage of spatial points (pixels) above threshold in a 300 s interval over all simulations in the FHN-1D training data (top) and KM-based reconstructions (bottom). (N=50 each; mean ± SD is plotted).
d. Spatiotemporal patterns as quantified on each training kymograph (top row) and KM-based reconstruction (bottom row) using a temporal autocorrelation function averaged over spatial grid points (left column) and a spatial autocorrelation function averaged over time points (right column). (N=50 each; mean ± SD is plotted).
e. Frames from representative training and KM-based reconstruction simulations for the FHN-2D reaction-diffusion model. (N=50 each).
f-h. Similar to b-d, for FHN-2D reaction-diffusion simulations.

Table 2. True and estimated spatial Langevin equation (SLE) parameters for FR-1D and FR-2D models using Kramers-Moyal regression on training simulations (N=50).

Activator SLE: $df = D_f \nabla^2 f + a_1 f + a_2 f r + a_3 f^2 / (f^2 + a_4) + a_5 + g_f dQ_f$ Inhibitor SLE: $dr = D_r \nabla^2 r + b_1 f + b_2 r + g_r dQ_r$

	D_f	a_1	a_2	a_3	a_4	a_5	g_f
FR-1D							
Ground truth	2.6	-0.00835	-0.8335	8.35	1.44	0.0635	0.15
Mean	2.583	0.00643	-0.8305	8.280	1.439	0.0620	0.161
SD	0.025	0.0484	0.0078	0.126	0.016	0.0038	0.001
FR-2D							
Ground truth	1.14	-0.0120	-1.200	12.024	2.88	0.0482	0.31
Mean	1.135	0.0177	-1.199	11.898	2.873	0.0435	0.324
SD	0.013	0.0546	0.014	0.157	0.058	0.0054	0.002
	D_r	b_1	b_2	g_r			
FR-1D							
Ground truth	5.2	0.5875	-0.025	0			
Mean	5.197	0.5872	-0.0250	0.036			
SD	0.004	0.0002	3.80×10^{-5}	0.0007			
FR-2D					_		
Ground truth	1	0.5229	-0.045	0			
Mean	0.999	0.5226	-0.0449	0.0384			
SD	0.001	0.0003	0.0001	0.0007			

estimated parameters for the activator in the FR model displayed larger variability compared to those for the inhibitor, which lacked a stochastic term in its equation. Additionally, the variability of parameters estimated for FR was slightly less than that observed for FHN, likely due to the smaller recording time step used for FR data analysis.

To assess the robustness of these parameters, an additional 50 1D and 2D FR simulations were generated based on the estimated parameters, forming reconstruction datasets. A comparative analysis of representative kymographs and movies from the 1D or 2D reconstruction dataset (Fig. 4b,d) and the corresponding training dataset (Fig. 4a,e) underscored the inherent stochastic nature of the system, highlighting the unique nature of each simulation. As above, wave feature analysis was subsequently employed for a nuanced comparison of the two datasets.

A high degree of similarity was observed between training and reconstruction datasets. The number of firings at any grid point exhibited similar distributions in both FR training and reconstruction simulations (Fig. 4b,f), with a peak centered around 3 firings for 1D simulations, and around 4 firings for 2D simulations. The spatial wave activity, quantified by the proportion of grid points above threshold at any given time, exhibited high similarity between training and reconstruction simulations, with variability centered around 10% for 1D simulations and 20% for 2D simulations (Fig. 4c,g). Strikingly, even the slight oscillations in spatial wave activity over the last 300 s of training for 1D and 2D simulations were replicated in the reconstructions. The temporal and spatial autocorrelations also demonstrated a high degree of similarity between training and reconstruction data (Fig. 4d,h), indicating comparable wavelength scales in time (1D: \sim 5–20 s; 2D: \sim 5-15 s) and space (1D: \sim 25-50 px.; 2D: \sim 10-50 px.).

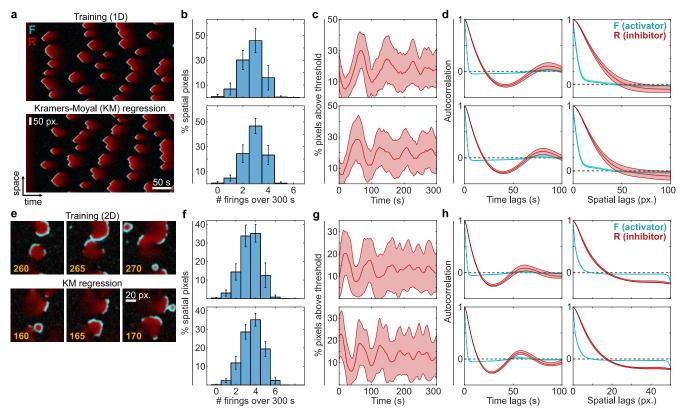


Fig 4. Reconstruction and validation with Kramers-Moyal (KM) based estimates of FR reaction-diffusion model parameters

a. Representative kymographs from a 400 s simulation in the FR-1D training dataset (top) and from a reconstruction dataset generated by FR reaction-diffusion model parameters estimated using KM regression (bottom). (N=50 each; 50 spatial pixels (px.) and 50 s fiducial markers apply to both kymographs).
b. Temporal firing frequency, computed as the distribution of the number of firings in a 300 s interval at spatial points over all simulations in the FR-1D training data (top) and KM-based reconstructions (bottom). (N=50 each; error bars indicate SD).
c. Spatial wave activity, computed as the percentage of spatial points (pixels) above threshold in a 300 s interval over all simulations in the FR-1D training data (top) and KM-based reconstructions (bottom). (N=50 each; mean ± SD is plotted).
d. Spatiotemporal patterns as quantified on each training kymograph (top row) and KM-based reconstruction (bottom row) using a temporal autocorrelation function averaged over spatial grid points (left column) and a spatial autocorrelation function averaged over time points (right column). (N=50 each; mean ± SD is plotted).
e. Frames from representative training and KM-based reconstruction simulations for the FR-2D reaction-diffusion model. (N=50 each).
f-h. Similar to b-d, for FR-2D reaction-diffusion simulations.

Learning sparse polynomial reaction-diffusion models from FHN dynamics and patterns

Building on the above work estimating parameters for models with a fixed structure for the deterministic and stochastic components of the spatial Langevin equation (SLE), we next addressed the challenge of learning model structure and parameters together from FHN-1D training simulations. Specifically, we considered a library of polynomial reaction-diffusion models based on equations 1, 2, and 4. Note that the library varies only in the polynomial reaction terms in the deterministic components of the SLE, and that the forms of the spatial diffusion and noise terms were held fixed for both the activator and inhibitor equations. Importantly, the true FHN reaction-diffusion model structure is a subset of this library.

To learn sparse models initially from stochastic dynamics, we used sparse KM regression (Methods). This involved a series of linear regressions to minimize the MSE between the empirical and parameterized theoretical KM averages for the deterministic components of the activator and inhibitor SLE separately. Starting with 11 deterministic parameters (one spatial diffusion and ten polynomial reaction model terms) for the initial regression, we sequentially eliminated the smallest reaction parameter in absolute value at each step. The stochastic components of the activator and inhibitor SLE were estimated assuming constant noise coefficients. Figure 5a illustrates the specific model terms eliminated at each iteration. Sparse KM regression was applied separately to each of the 50 training simulations, and the proportion of simulations in which a specific model term in the polynomial reaction-diffusion library remained selected was plotted for each column (iteration). Owing to the inherent stochasticity in the training dataset, some variability in the eliminated terms was observed resulting in shaded regions, especially in early steps where few model terms had been eliminated (higher model complexity).

To determine the optimal number of parameters, we employed the Bayesian Information Criterion (BIC), which balances accuracy and sparsity (Methods, Fig. 5b). Analysis of the mode of the distribution of optimal sparsity (BIC minimum) from all simulations (S2 Figa) indicated that the optimal number of reaction-diffusion parameters was five for the activator and three for the inhibitor (denoted "5u3v"). The reaction-diffusion terms corresponding to these parameters were identified from the model terms at the iteration highlighted in white in Figure 5a, by selecting the top five activator terms and the top three inhibitor terms remaining over 50 training simulations (Fig. 5a). Consequently, this method successfully recovered the original FHN reaction-diffusion model structure.

Figure 5c presents representative reconstructions using parameters from sparse KM regression for the optimal model structure, 5u3v, alongside reconstructions using three sparser models: 5u2v, 4u3v, and 4u2v (S1 Table). The 5u2v model displayed similar wave patterns to those of the 5u3v model, albeit with reduced activity, while the 4u3v and 4u2v models predominantly exhibited oscillatory behavior, differing in period and synchronization across the spatial dimension. The insets reveal that these differences arise because the inhibitor nullclines intersect the activator nullcline at the local minimum, positioning the system at a Hopf bifurcation.

To quantify the differences in the wave patterns clearly observed among the models tested, we considered several normalized wave metrics (Fig. 5d). These included the following averaged spatiotemporal features: the joint probability distribution of U and V, spatiotemporal wave frequency, spatial wave activity, and spatiotemporal patterns analyzed using autocorrelation functions. Analysis of these metrics revealed that reconstructions of the first two models (5u3v and 5u2v) showed reasonably close alignment with the training data, while the last two models (4u3v and 4u2v) exhibited significant discrepancies.

Motivated by these findings, we explored whether optimizing based on these wave features could identify model parameters that more closely matched the training data. Using a Nelder-Mead simplex search, we optimized a custom objective function involving the sum of squared relative errors in wave features of a reconstruction dataset relative to that of training FHN-1D data (Methods, S2 Figb). Figure 5e shows kymographs for the optimal parameters based on wave features optimization for the four model structures considered earlier. The 5u3v and 5u2v models displayed

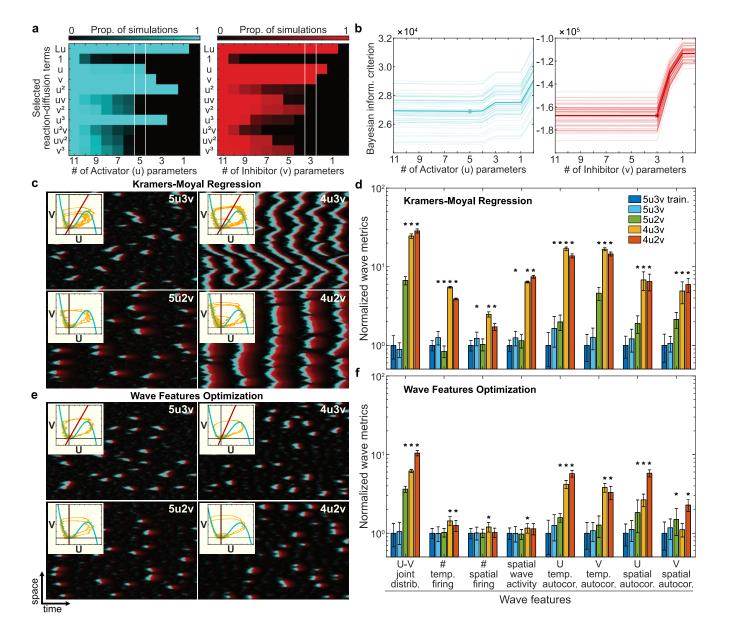


Fig 5. Learning sparse models from FHN-1D data by optimizing Kramers-Moyal (KM) averages and wave features

a. Reverse-greedy search approach estimating parameters using KM-based regression at each iteration, and sequentially eliminating terms from a library of polynomial reaction-diffusion models. This sparse KM regression was performed separately for the activator equation (left) and inhibitor equation (right), and repeated for each kymograph in the FHN-1D training dataset (N=50). Colors indicate the proportion of simulations for which each model term remained selected as model complexity decreased from 11 to zero parameters over the range of iterations. White lines indicate the optimal sparsity as estimated from the next panel. b. Bayesian information criterion (BIC) estimated at each iteration of sparse KM regression for the activator and inhibitor equations and plotted as separate lines for each simulation (average is in bold). The star indicates the most common minimum of the BIC (optimal sparsity) over all simulations. c. Reconstructions using KM-based parameters for four sparse models labeled by the number of terms in the activator (U) and inhibitor (V) equations: 5u3v (optimal sparsity; five terms for U equation, three terms for V equation), 5u2v, 4u3v, and 4u2v. Insets show representative nullclines and phase-plane trajectories. d. Spatiotemporal wave metrics computed for FHN-1D training data (N=50) and reconstruction datasets of the four sparse model structures (N=20 each). Metrics were normalized relative to training data and plotted on a logarithmic scale. Stars indicate statistically significant differences relative to training data for each wave feature (Wilcoxon rank-sum test, p < 0.01). e. Similar to c, for reconstructions based on parameters estimated for the same sparse model structures and optimizing for wave features. f. Similar to d, a quantitative comparison of each normalized wave metric in the reconstruction dataset for each sparse model structure (N=20) to the normalized wave metrics of the original FHN-1D training data.

similar behavior, whereas the 4u3v simulation showed no oscillations, and the 4u2v model exhibited broken patches with some oscillatory behavior. Quantification of individual raw wave features (S3 Fig, S4 Fig), normalized wave metrics (Fig. 5d), and overall wave metrics error (Fig. 5f) all confirmed the improved characteristics of simulations after wave features optimization.

Learning sparse polynomial reaction-diffusion models from FR dynamics and patterns

Extending this approach to real systems requires fitting data-derived quantities to a library of generalized models that may not include the exact system. To explore the feasibility of this, we used training data generated by the FR-1D model which includes a rational term in the activator dynamics. As before, we began with initially fitting KM quantities to a polynomial reaction-diffusion library, and then fine-tuned parameters using wave features optimization. Unlike the FHN dataset, however, the true FR reaction-diffusion model structure is a subset of the library for the inhibitor (R) dynamics, but not for the activator (F).

In Figure 6a, we started with 11 deterministic parameters for both the activator and inhibitor and sequentially eliminated terms using sparse KM regression, as previously. In contrast to FHN data, here there was decreased variability (fewer shaded regions) in the selected reaction diffusion terms for both the activator and the inhibitor even at early iterations with higher model complexity, likely due to the smaller recording time step $\tau_0 = 0.05\,\mathrm{s}$ used for analysis. Once again, we computed the BIC over all iterations for each simulation (Fig. 6b); then we used the mode of the distribution of optimal sparsity (BIC minimum) from all simulations to obtain the final estimate of optimal sparsity (S5 Figa). The optimal model structure corresponded to a model denoted "9f3r" with 9 parameters for the activator (one spatial diffusion and eight reaction) and three parameters for the inhibitor (one spatial diffusion and two reaction). The model terms at the corresponding iterations highlighted in Figure 6a matched the exact structure of the deterministic component of the inhibitor equation; however, for the activator, the estimated 9-term polynomial reaction-diffusion model was quite distinct from the training FR model. Additionally, despite 9 parameters being optimal, we observed minimal differences in BIC from models with 9 parameters to 5 parameters, as seen in Fig. 6b.

To examine the behavior of these different stochastic models, we generated a set of 20 reconstructions using the estimated parameters from sparse KM regression for the 9f3r model and

334

335

336

337

338

339

340

341

342

343

345

347

348

349

351

353

354

355

356

357

358

359

360

361

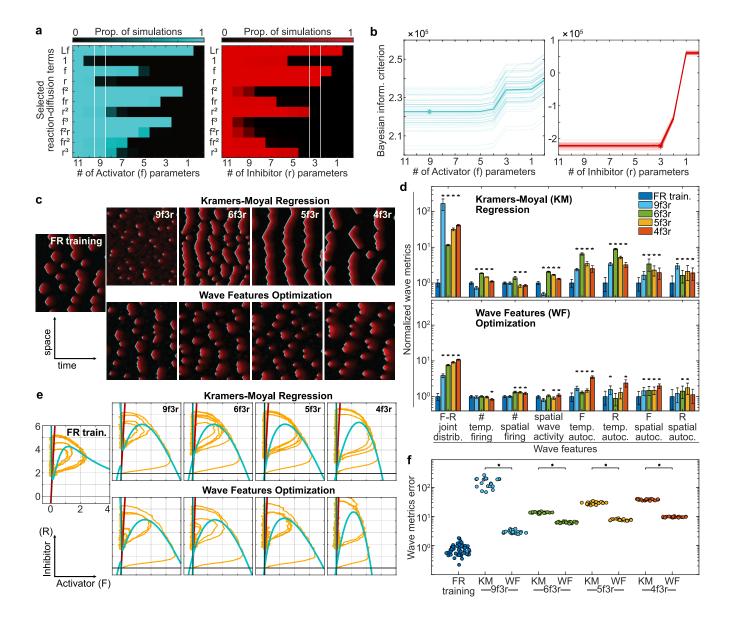


Fig 6. Learning sparse models from FR-1D data by optimizing Kramers-Moyal averages and wave features

a. Reverse-greedy search approach estimating parameters using KM regression at each iteration, and sequentially eliminating terms from a library of polynomial reaction-diffusion models. This sparse KM regression was performed separately for the activator equation (left) and inhibitor equation (right), and repeated for each kymograph in the FR-1D training dataset (N=50). Colors indicate the proportion of simulations for which each model term remained selected as model complexity decreased from 9 to zero parameters over the range of iterations. White lines indicate the optimal sparsity as estimated from the next panel. b. Bayesian information criterion (BIC) estimated at each iteration for the activator and inhibitor equations and plotted as separate lines for each simulation (average is in bold). The star indicates the most common minimum of the BIC (optimal sparsity) over all simulations. c. Reconstructions using KM-based parameters (top row) for four sparse models labeled by the number of terms in the activator (F) and inhibitor (R) equations: 9f3r (optimal sparsity; nine terms for U equation, three terms for V equation), 6f3r, 5f3r, and 4f3r. Bottom row shows representative reconstructions based on parameters estimated for the same sparse model structures and optimizing for wave features. A representative kymograph from the FR-1D training data is plotted on the far left for comparison. d. Spatiotemporal wave metrics computed for FR-1D training data (N=50) and both KM-based and wave features (WF) optimization-based reconstruction datasets of the four sparse model structures (N=20 each). Metrics were normalized relative to training data and plotted on a logarithmic scale. Stars indicate statistically significant differences relative to training data for each wave metric (Wilcoxon rank-sum test, p < 0.01). e. Representative nullclines and phase-plane trajectories for KM-based reconstructions (top row) and wave features based reconstructions (bottom row), as well as the original FR-1D training data (far left). f. Scalar wave metrics error computed for each simulation in the FR-1D training data, and both reconstruction datasets (KM-based and WF-based). Each dot represents the square root of the sum of the squared relative error between wave metrics in a given simulation and the mean wave metrics in the FR-1D training data. Stars indicate statistically significant differences (Wilcoxon rank-sum test, p < 0.01).

three sparser models: 6f3r, 5f3r, and 4f3r (S2 Table). Fig. 6c shows representative reconstructions with SLE parameters estimated using KM regression. By visual inspection, the general wave patterns observed in simulations of three of the models deviated from the FR training data: 9f3r had considerably smaller wave firings compared to the training data; 6f3r, 5f3r, and 4f3r showed oscillatory behavior with increasing wave period. Spatiotemporal wave metrics computed from the reconstruction sets quantified the differences in the patterns observed for each model structure (Fig. 6d).

We then performed wave features optimization to identify model parameters to match the patterns observed in FR training data. The coarse-grained approach of sparse KM regression provided candidate sparse model structures with a wide range of spatiotemporal patterns significantly differing from the training data. Fine-tuning as before, we used initial parameters from sparse KM regression, and optimized a wave metrics error objective function over multiple rounds for the reaction, spatial diffusion, and noise parameters (S5 Figb). Compared to patterns in FR training data, wave features (WF) optimization achieved improved matching of wave metrics for all model structures, with the most improvements visible in the 9f3r model (Fig. 6c). Quantification confirmed these trends across both individual raw wave features (S6 Fig, S7 Fig), normalized wave metrics (Fig. 6d), and overall wave metrics error (Fig. 6f). The improvement in wave metrics for each model tested demonstrated the success of this combined approach in learning sparse polynomial reaction-diffusion models even when the true FR model structure is not fully captured by a polynomial model.

Exploring possible reasons for the improvement, we plotted the spatially homogeneous reaction equation nullclines and phase plane trajectories for each of the models learned from KM regression and wave features optimization (Fig. 6e). In general, for both KM regression and wave features (WF) optimization, all the models had an approximate inverted "N" shape for the F-nullcline. However, for the sparser models, 663r, 563r, and 463r, the 463r model, this inverted shape consisted two curves for the F-nullcline: one vertical line at F=0 and a second concave down curve. The intersecting curves resulted in an approximate inverted "N" shape in the region of the phase plane

363

364

365

366

367

368

369

371

372

373

374

375

377

378

379

380

381

382

383

384

386

387

388

accessed by stochastic trajectories. During WF optimization, the local shape of the curves around the equilibrium point, which relates to system threshold, appeared to become tuned for matching excitability. For example, for the 9f3r model, compared to the reduced wave activity in the KM regression model, the WF optimization model exhibited a decrease in the F-nullcline minimum similar to the training model, resulting in a lower threshold and increased wave activity comparable to training data (Fig. 6e). Interestingly, for the sparsest 4f3r model, WF optimization resulted in more complex changes to the nullclines, but locally around the equilibrium point of the system, the curves moved closer to that of the training model, resulting in improved firing patterns (S5 Figc).

Estimating 2D-spatial reaction-diffusion models from datasets with limited observations

We further challenged our approach by testing scenarios of low-resolution data arising in real-world contexts. Signals acquired from live-cell microscopy are often propagating in 2D space and are measured every few seconds, with only 1-2 species observed at a time. To evaluate estimating stochastic reaction-diffusion models from limited data, in Figure 7 we applied our approach to estimating parameters for the FHN-2D model from datasets with various rates of temporal sampling and observations limited to only one of the species (activator (U) or inhibitor (V)).

In Fig. 7a, we show the effects of different rates of temporal subsampling on data quality obtained at a particular spatial grid point. Compared to full signals acquired at the simulation time step of dt = 0.01 s, decreasing sampling rates with acquisition every $\tau = 0.1, 1, 4$ s resulted in smoothing of high frequency components in the stochastic signals. However, subsampling at even $\tau = 4$ s, with a 99.75% reduction in temporal data, still retained much of the general profile of a propagating wave. To study the effects of subsampling on parameter estimation, we first computed finite-time KM averages (S8 Fig) for each of the four sampling rates. We then applied KM regression to estimate parameters for the deterministic and stochastic components of the SDE that best fit the finite-time KM averages and generated reconstruction datasets of 20 simulations for each case. Maximum intensity frames from representative 2D-spatial simulations based on parameters from finite-time KM regression showed the qualitative deterioration of patterns as sampling rate decreased, with barely any visible patterns at $\tau = 4$ s (Fig. 7b, top panel; S3 Video, S4 Video).

Next, we checked whether wave features optimization could yield improved estimates from subsampled data. Similar to before, we first computed wave metrics from FHN-2D training data based on both the activator and inhibitor signals sampled every $\tau=1$ or $\tau=4$ s (Methods). Using the previous objective function with subsampled wave features, we then optimized over the set of reaction, diffusion, and noise parameters of a fixed FHN-2D model structure (S3 Table). We tested three cases, each with an initial set of parameters obtained from finite-time KM regression (Fig. 7b, top panel): 1) $\tau=0.1\,\mathrm{s}$, 2) $\tau=1\,\mathrm{s}$, and 3) $\tau=4\,\mathrm{s}$. For cases 1 and 2, wave features were optimized to training data wave features at $\tau=1\,\mathrm{s}$; for case 3, wave features were optimized to training data wave features at $\tau=4\,\mathrm{s}$. A set of 20 reconstructions were generated for each case, and maximum intensity frames from representative simulations demonstrated marked improvement in wave patterns for cases 1 and 2, and moderate improvement for case 3 (Fig. 7b, bottom panel; S3 Video, S4 Video). Significant improvements in individual raw wave features (S9 Fig, S10 Fig), normalized wave metrics (Fig. 7c), and wave metrics error (Fig. 7d) were observed over the range of cases, demonstrating the effectiveness of wave features optimization for parameter estimation of 2D-spatial reaction-diffusion models from subsampled datasets.

To further examine the robustness of our approach, we tested the feasibility of parameter estimation from FHN-2D training datasets that were subsampled and only partially observed. Here, we used subsampled data at $\tau=4\,\mathrm{s}$ and considered two cases: 1) only observing the activator (U), and 2) only observing the inhibitor (V). Since KM averages cannot be estimated from partially-observed systems, we only focused on wave features optimization. For initialization, we used a set of parameters designed to give near-oscillatory behavior with large propagating waves that qualitatively differed from training data as visible in the frames from 2D-simulations (Fig. 7e, top

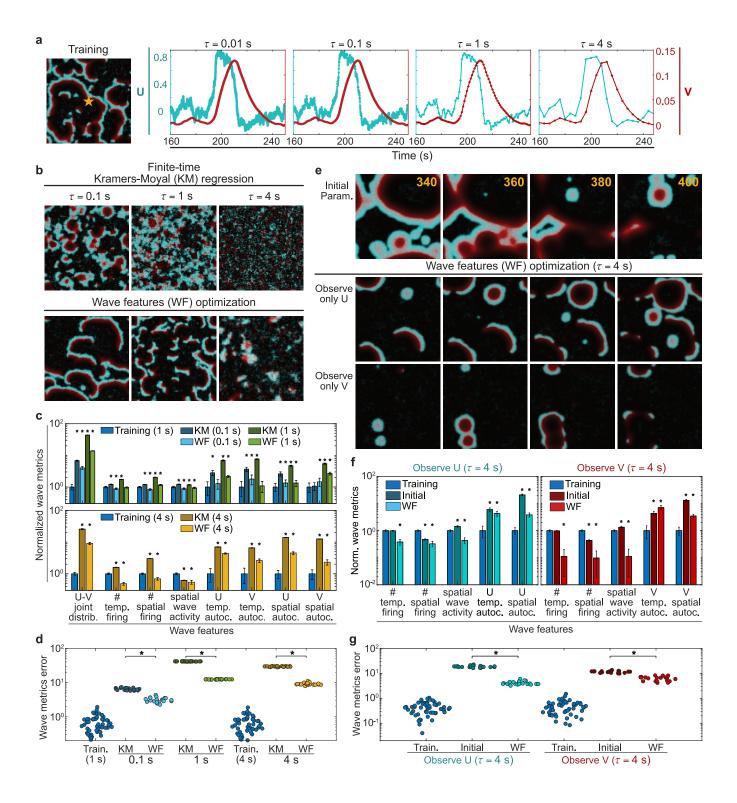


Fig 7. Estimating stochastic reaction-diffusion models from limited observations

a. Maximum intensity frame (226s) from a 2-D spatial simulation in the FHN-2D training dataset. Time series for the activator (U, cyan) and inhibitor (V, red) were extracted at a single spatial grid point (orange star), and plotted with decreasing temporal sampling rates, from $\tau = 0.01\,\mathrm{s}$ (integration time step) to $\tau = 4\,\mathrm{s}$. b. Maximum intensity frames from reconstructions based on FHN-2D model parameters estimated using finite-time Kramers-Moyal (KM) regression (top row) or wave features (WF) optimization (bottom row). Finite-time KM regression parameters were estimated from data sampled at $\tau = 0.01, 0.1, \text{ or } 4\,\text{s}$ and then used as initial parameters for wave features optimization. c. Wave metrics corresponding to 8 wave features computed for reconstruction datasets (N=20 simulations) generated with parameters from finite-time KM regression or WF optimization and compared to that of training data (N=50 simulations). For parameters estimated from data sampled at $\tau = 0.01$ and 1 s, wave metrics from corresponding reconstructions were computed at $\tau = 1$ s and compared with metrics from training data sampled at $\tau = 1$ s (top panel). For parameters estimated from data sampled at $\tau = 4$ s, wave metrics from the corresponding reconstructions were computed at $\tau = 4$ s and compared with metrics from training data sampled at $\tau = 4$ s (bottom panel). d. Wave metrics errror based on the wave features objective function used during optimization and computed for FHN-2D training data sampled at 1s and 4s, as well as reconstruction datasets from finite-time KM regression and WF optimization. Each dot represents the error between wave metrics in each simulation and the mean wave metrics of the training data. e. Frames from simulations generated with an initial parameter set (top row) and after wave features optimization when only observing either activator (middle panel) or the inhibitor (bottom panel) signals sampled at $\tau = 4$ s. The time stamps (seconds) in the top panel indicate corresponding simulation time points for the frames in each column. f. Normalized wave metrics corresponding to 5 single-species wave features computed for reconstruction datasets (N=20 simulations) generated with parameters from WF optimization and compared to that of training data (N=50 simulations) when only observing U (left panel), or V (right panel). g. As in panel d., wave metrics error based on the single-species wave features objective function used during optimization and computed for FHN-2D training data as well as reconstruction datasets from WF optimization. Stars indicate statistically significant differences between compared groups, or differences relative to training data, as in \mathbf{c} or \mathbf{f} , respectively (Wilcoxon rank-sum test, p < 0.01).

row). Because only one species was observed, we computed a set of 5 wave features from FHN-2D training data for each case, and optimized the objective function based on this subset of wave features to estimate FHN-2D reaction-diffusion model parameters (S4 Table). Representative frames from reconstructions based on parameters after wave features optimization demonstrated the improvements in wave patterns (Fig. 7e, S5 Video, S6 Video). Remarkably, in both cases, the wave propagation range and wave profile thicknesses of both species more closely matched that of training data (Fig. 7e). Optimization when only U signal was observed appeared to show more similar wave activity and frequency to that of training data, while optimization when only V signal was observed resulted in markedly reduced wave activity and frequency. Quantitative analysis of each of the 5 raw wave features (S11 Fig), normalized wave metrics (Fig. 7f), and the overall wave metrics objective function each confirmed these trends (Fig. 7g). These results indicate the effectiveness of wave features optimization to generate parameter estimates for 2D-spatial reaction-diffusion models even from datasets with limited observations.

Discussion

The development of mathematical models to capture the intricate behaviors of complex systems has a rich history. Traditionally, these models have been knowledge-based, with parameters adjusted through an iterative exploration of parameter space guided by experience and intuition. This method, however, is limited to handcrafted models of low-dimensional systems with few variables and becomes computationally intensive when applied to spatially-distributed systems. Recent advances in machine learning have offered new possibilities for nonlinear system identification and computational biology, yet, to our knowledge, these techniques have not been extensively applied to stochastic reaction-diffusion systems. Such systems are crucial in biological contexts, where signaling

440

442

444

445

446

447

448

449

450

451

452

454

456

457

458

459

460

networks exhibit noise-driven, dynamic patterns that propagate across cellular boundaries.

Overall, this study introduces a proof-of-concept approach for using spatiotemporal data to learn sparse model equations that govern an excitable, stochastic system's emergent properties and patterns. To this end, we first extracted key features characterizing the datasets: microscale stochastic dynamics and macroscale spatiotemporal patterns. We then estimated model structure and parameters using a two-step machine-learning algorithm that first matches stochastic dynamics using sparse Kramers-Moyal (KM) regression, and second, matches spatiotemporal patterns using wave features (WF) optimization. Using a set of only 8 wave metrics and a computationally efficient approach for estimating KM averages in a Laplacian-extended state space, our methods demonstrate efficacy over a range of model complexity and data paucity. We validated the method for multiple nonlinear activator-inhibitor models, 1D and 2D spatially-distributed systems, temporally subsampled data, and data with unobserved model species.

Our approach presents new avenues for understanding not only biochemical waves and pattern formation but also spatially-distributed dynamical systems in general. We learned novel, sparse polynomial reaction-diffusion models for both the FHN and FR excitable systems. From a theoretical perspective, the identification of alternative equations for excitable systems that are simpler than the classical model equations opens the door for further research questions. From an applications perspective, this is essential for understanding real-world systems where the correct model structure is not predetermined, and sparse models can yield more robust predictions and insightful system information. Compared to other methods which focus on learning system trajectories, our approach learns sparse stochastic equations, in which the parameters can provide quantitative, mechanistic insights regarding the excitable system, e.g. reaction model structure, feedback loop strengths, temporal and spatial scales, process noise, and threshold. This presents a significant advantage when modeling dynamic biological systems where learning simplified system structure is just as important as approximating system behavior.

Despite these advances, KM approaches have inherent limitations. Fundamentally, KM averages estimate stochastic dynamics in the limit of infinitesimal time steps and under the assumption of a stationary Markov process. Hence, for systems with rapid dynamics, fitting KM averages works best for datasets that are highly sampled in time. Visualizing the projected finite-time KM averages computed over larger time steps (S8 Figb-d), we observed decreasing accuracy compared to the standard KM averages, even while keeping data quantity consistent. This effect was less pronounced for the inhibitor due to the absence of process noise in the inhibitor equation here. Optimizing sampling rate for KM-based analysis is also complicated in real-world settings, by the fact that sampling too rapidly can lead to non-Markovian behavior due to physical properties of the system; in these cases, sampling at time steps around the Einstein-Markov time scale is recommended for stochastic modeling [47,58]. Additionally, because KM-based methods estimate parameters separately for each species and their SLE components, the coupling between equations is not accounted for, limiting effectiveness when learning sparse models. There are also often a range of model structures with similar errors in KM quantities that produce dramatically different wave patterns (Fig. 5b, Fig. 6b), highlighting that KM-based stochastic dynamics offer a limited view of system properties. To address these challenges, we developed wave features (WF) optimization.

This Monte Carlo, features-based optimization approach has several advantages. Through the use of a customized objective function, it allows for optimization of a set of key macroscale quantities characterizing system behavior. When KM averages are available as initial parameters for WF optimization, these initial parameters, which approximate stochastic dynamics but deviate significantly in reconstruction patterns, can be effectively tuned to replicate general system features. A key part of the effectiveness lies in the fact that, unlike KM regression where deterministic and stochastic parameters are estimated separately for each species, features-based optimization adjusts the deterministic and stochastic parameters for both activator and inhibitor equations simultaneously.

The strengths of WF optimization are particularly evident when learning sparse models. By jointly tuning parameters of the reaction equation, spatial diffusion coefficients, and noise terms, the

algorithm effectively matches wave features (S1 Table, S2 Table). Even for the sparsest models tested in the FHN and FR models, the 4u2v model and the 5f3r model, respectively, WF optimization recovered wave patterns from initial KM-based reconstructions that were oscillatory (Fig. 5c,e, Fig. 6c). Interestingly, the KM-based and WF-based 4u2v and 5f3r models were all at the Hopf bifurcation point when examining the spatially-homogeneous reaction equation nullclines (insets in Fig. 5c,e, Fig. 6e). This highlights an important distinction from spatially-homogeneous deterministic systems where system behavior can only be tuned by reaction-parameters. WF optimization based models did have slight reaction parameter adjustments for the activator nullcline to approach the training nullclines locally near the system equilibrium (S2 Figc, S5 Figc). However, there were also consistent changes in other parameters not visible in phase-plane analysis, such as the spatial diffusion coefficient and state noise parameters, that contributed to the recovery of wave patterns by possibly increasing effective threshold in the system despite being at the Hopf bifurcation (S1 Table, S2 Table, S2 Figc, S5 Figc).

Additionally, using WF optimization to fine-tune parameters for candidate models from sparse KM regression allowed for the identification of novel sparse models for excitable systems. We identified multiple sparse models for both the FHN model and the FR model. For the classical FHN model, which already was a simplified 5u3v model, we found sparser model structures with as few as three activator reaction terms and only one inhibitor term (4u2v, since spatial diffusion adds an extra term for both species). The 4u2v model, after tuning parameters with WF optimization, demonstrated similar stochastic dynamics and wave features to the original model. Similarly, we approximated a biologically-relevant FR model using as few as 4 polynomial terms. Notably, although the FR model contained a rational Hill function, its activator nullcline can be written as a fourth-order polynomial equation. Hence, our library of up to third-order polynomial terms could be used to approximate the nullcline. The sparse models we identified all generally approximated the spatially homogeneous nullclines; however the 4f3r model after WF optimization demonstrated more complicated adjustments (S5 Figc).

Examining the parameters for these alternative model structures of excitable systems, we observed that the effective positive and negative feedback loops were maintained, indicating that despite the mathematical structure being sparser, the general network structure remained consistent (S1 Table, S2 Table. Splitting the reaction terms into the signage of the terms that depend on only U, only V, or mixed terms, the reaction network can be inferred. For example, in the 4u2v model structure for the U reaction equation, the V term has a negative coefficient (negative feedback), and the terms that depend on U only (u^2, u^3) yield a positive value in the right quadrant where trajectories lie (S1 Table). Our combined approach with approximation of stochastic dynamics and spatiotemporal patterns is thus of high value in modeling biological systems where insights reside in both microscale network structure defined by the stochastic equations as well as the emergent macroscale wave features.

WF optimization also improves estimation from signals with low temporal resolution and unobserved species. In these limited data cases, KM regression yielded poor model estimates (in the case of subsampled data) or was not well-defined (in the case of partially-observed data). Using WF optimization on the other hand, we can tune initial KM-based parameters to obtain approximate models from data with $\tau=4\,\mathrm{s}$, where temporal resolution is reduced by 99.75% compared to full simulations (Fig. 7b–d). When learning from datasets that resemble experimental data with only one of the two modeled species being observed and a temporal resolution of $\tau=4\,\mathrm{s}$, WF optimization still provides reasonable parameter estimates, given a set of initial parameters (Fig. 7e). Interestingly, although the activator and inhibitor signals differed significantly in that only the former was modeled with system noise, the optimization procedure converged to similar results. In fact, estimation using the filtered inhibitor signal was slightly poorer based on the wave features in the reconstructions than estimation using the highly noisy activator signal. Additional interesting questions regarding observability of the system arise given that the parameters after optimization using partially-observed data differed significantly from the ground-truth parameters.

The development and validation of these algorithms represent a significant contribution to

data-driven learning of spatially-distributed stochastic models of excitable systems. These computational approaches are of interest in our field of cell biology where high spatial resolution, time-lapse imaging of various pattern-forming systems are readily available but often underutilized when constructing models. Future efforts should focus on integrating experimental biology with machine learning and dynamical systems to gain deeper insights into cellular signaling pathways in both health and disease.

Supporting information

S1 Methods Supplementary Methods.

- S1 Fig. Visualizing KM averages for FHN and FR datasets. KM averages computed over a Laplacian-extended state space from FHN-1D/2D and FR-1D/2D training data, and then averaged over Laplacian values and plotted as a heatmap in the activator-inhibitor phase plane. Computations from a single simulation in each of the four training sets are shown.
- S2 Fig. FHN system identification using sparse KM regression and wave features optimization. a. Distribution of the minimum of the Bayesian information criterion (BIC) computed for each simulation. b. Wave metrics objective function values tracked over the first 100 iterations of wave features optimization, with model structure labeled. c. Spatially-homogeneous nullclines plotted in the phase-plane for three models: FHN training model parameters (solid lines); 4u2v sparse-KM regression based parameters (short dashes); and 4u2v WF optimization based parameters (long dashes). The right panel is a zoomed in plot of the region depicted by the blue box in the left panel.
- S3 Fig. Full spatiotemporal analysis of reconstructions of sparse FHN-1D models estimated by KM regression. Eight wave features computed for the FHN training set (5u3v, N=50 simulations), and four polynomial reaction-diffusion model reconstruction sets using KM-based parameters (5u3v, 5u2v, 4u3v, and 4u2v; N=20 each). a. Joint probability distribution of the activator and inhibitor. b. Temporal firing frequency. c. Spatial firing frequency. d. Spatial wave activity. e. Temporal autocorrelation of the activator (cyan) and inhibitor (red). f. Spatial autocorrelation of the activator (cyan) and inhibitor (red).
- S4 Fig. Full spatiotemporal analysis of reconstructions of sparse FHN-1D models estimated by wave features (WF) optimization. Eight wave features computed for the FHN training set (5u3v, N=50 simulations), and four polynomial reaction-diffusion model reconstruction sets using WF optimization based parameters (5u3v, 5u2v, 4u3v, and 4u2v; N=20 each). a. Joint probability distribution of the activator and inhibitor. b. Temporal firing frequency. c. Spatial firing frequency. d. Spatial wave activity. e. Temporal autocorrelation of the activator (cyan) and inhibitor (red). f. Spatial autocorrelation of the activator (cyan) and inhibitor (red).
- S5 Fig. FR system identification using sparse KM regression and wave features optimization. a. Distribution of the minimum of the Bayesian information criterion (BIC) computed for each simulation. b. Wave metrics objective function values tracked over the first 100 iterations of wave features optimization, with model structure labeled. c. Spatially-homogeneous nullclines plotted in the phase-plane for three models: FR training model parameters (solid lines); 4f3r sparse-KM regression based parameters (short dashes); and 4f3r WF optimization based parameters (long dashes). The right panel is a zoomed in plot of the region depicted by the blue box in the left panel.

S6 Fig. Spatiotemporal analysis of reconstructions of sparse FR-1D models estimated by KM regression. Eight wave features computed for the FR training set (N=50 simulations), and four polynomial reaction-diffusion model reconstruction sets using KM-based parameters (9f3r, 6f3r, 5f3r, and 4f3r; N=20 each). a. Joint probability distribution of the activator and inhibitor. b. Temporal firing frequency. c. Spatial firing frequency. d. Spatial wave activity. e. Temporal autocorrelation of the activator (cyan) and inhibitor (red). f. Spatial autocorrelation of the activator (cyan) and inhibitor (red).

- S7 Fig. Spatiotemporal analysis of reconstructions of sparse FR-1D models estimated by wave features (WF) optimization. Eight wave features computed for the FR training set (N=50 simulations), and four polynomial reaction-diffusion model reconstruction sets using WF optimization based parameters (9f3r, 6f3r, 5f3r, and 4f3r; N=20 each). a. Joint probability distribution of the activator and inhibitor. b. Temporal firing frequency. c. Spatial firing frequency. d. Spatial wave activity. e. Temporal autocorrelation of the activator (cyan) and inhibitor (red). f. Spatial autocorrelation of the activator (cyan) and inhibitor (red).
- S8 Fig. Visualizing finite-time KM averages computed from FHN-2D data with various rates of temporal subsampling ($\tau=0.1,1,4\,\mathrm{s}$). First and second KM averages for the activator and inhibitor computed over a Laplacian-extended state space, averaged over Laplacian values, and plotted as a heatmap in the activator-inhibitor phase plane. **a**. KM averages computed in the limit of $\tau\to0.01$ from one FHN-2D training simulation recorded at $\tau=0.1\,\mathrm{s}$. **b**. Finite-time KM averages computed for $\tau=0.1\,\mathrm{s}$ from one FHN-2D training simulation. **c**. Finite-time KM averages computed for $\tau=1\,\mathrm{s}$ from 10 FHN-2D training simulations. **d**. Finite-time KM averages computed for $\tau=4\,\mathrm{s}$ from 40 FHN-2D training simulations.
- S9 Fig. Spatiotemporal analysis of FHN-2D reconstructions based on parameters from finite-time KM regression and wave features (WF) optimization on subsampled data ($\tau = 0.1, 1\,\mathrm{s}$). Eight wave features computed for the FHN-2D training model (N=50 simulations) and reconstruction sets using finite-time KM and WF based parameters (N=20 each), all subsampled at the indicated time step, τ . a. Joint probability distribution of the activator and inhibitor. b. Temporal firing frequency. c. Spatial firing frequency. d. Spatial wave activity. e. Temporal autocorrelation of the activator (cyan) and inhibitor (red). f. Spatial autocorrelation of the activator (cyan) and inhibitor (red).
- S10 Fig. Spatiotemporal analysis of FHN-2D reconstructions based on parameters from finite-time KM regression and wave features (WF) optimization on subsampled data ($\tau = 4\,\mathrm{s}$). Eight wave features computed for the FHN-2D training model (N=50 simulations) and reconstruction sets using finite-time KM and WF based parameters (N=20 each), all subsampled at the indicated time step, τ . a. Joint probability distribution of the activator and inhibitor. b. Temporal firing frequency. c. Spatial firing frequency. d. Spatial wave activity. e. Temporal autocorrelation of the activator (cyan) and inhibitor (red). f. Spatial autocorrelation of the activator (cyan) and inhibitor (red).
- S11 Fig. Spatiotemporal analysis of reconstructions based on parameters from wave features (WF) optimization on subsampled ($\tau = 4\,\mathrm{s}$), partially-observed data. Five wave features computed for the FHN-2D training model (N=50 simulations) and reconstruction sets using initial parameters and WF optimization based parameters when only observing U or only observing V (N=20 each). a. Temporal firing frequency. b. Spatial firing frequency. c. Spatial wave activity. d. Temporal autocorrelation of either only the activator (cyan), or only the inhibitor (red). e. Spatial autocorrelation of either only the activator (cyan) and or only the inhibitor (red).

S1 Video. Representative FHN-2D spatial simulations of training and reconstruction

data from KM regression. S2 Video. Representative FR-2D spatial simulations of training and reconstruction 655 data from KM regression. 656 S3 Video. Representative FHN-2D spatial reconstructions based on parameters from finite-time KM regression and wave features optimization on subsampled, 658 fully-observed training data ($\tau = 1 \, \mathrm{s}$). S4 Video. Representative FHN-2D spatial reconstructions based on parameters from 660 finite-time KM regression and wave features optimization on subsampled, 661 fully-observed training data ($\tau = 4 \, \mathrm{s}$). 662 S5 Video. Representative FHN-2D spatial reconstructions based on parameters from 663 wave features optimization on subsampled training data with only the activator (U) 664 observed ($\tau = 4 \, \mathrm{s}$). S6 Video. Representative FHN-2D spatial reconstructions based on parameters from 666 wave features optimization on subsampled training data with only the inhibitor (V) observed ($\tau = 4 \, \mathrm{s}$). 668 S1 Table. Estimated FHN-1D model structures and parameters from sparse KM 669 regression and wave features optimization. S2 Table. Estimated FR-1D model structures and parameters from sparse KM 671 regression and wave features optimization. 672 S3 Table. Estimated FHN-2D model parameters from finite-time KM regression and 673 wave features optimization on subsampled, fully-observed data ($\tau = 0.1, 1, 4 \text{ s}$). 674 S4 Table. Estimated FHN-2D model parameters from wave features optimization on 675 subsampled, partially-observed data ($\tau = 4 \, \mathrm{s}$). Contributions 677 B.A.-S. and P.A.I. conceptualized the overall study. B.A.-S. developed the code, conducted the 678 simulations and analysis, and wrote the manuscript. ChatGPT-40 (OpenAI) was used only to edit 679 some of the initial writing and to vectorize a MATLAB script for computing Kramers-Moyal 680 averages. P.A.I. and P.N.D. provided guidance throughout the study and edited the manuscript. 681 Acknowledgments 682 We thank Y. Kevrekidis (School of Engineering, JHU) and M.I. Miller (School of Engineering, JHU) for their input and advice as thesis committee members. We thank T. Banerjee (Devreotes lab) and 684 A. Titi (School of Engineering, JHU) for stimulating discussions. We thank all members of the Devreotes, Iglesias, Kevrekidis, and D.N. Robinson (School of Medicine, JHU) labs for their helpful 686 feedback. This work was supported by the following NIH grants: R35 GM118177 (P.N.D) with 687 Diversity Supplement (B.A.-S.) and T32 GM136577 Medical Scientist Training Program (JHU). 688

References

- 1. Volpert V, Petrovskii S. Reaction-diffusion waves in biology. Phys Life Rev. 2009;6(4):267–310. doi:10.1016/j.plrev.2009.10.002.
- 2. Deneke VE, Di Talia S. Chemical waves in cell and developmental biology. J Cell Biol. 2018;doi:10.1083/jcb.201701158.
- 3. Krinsky V, Swinney H. Wave and patterns in biological and chemical excitable media. Noth-Holland, Amsterdam. 1991;.
- 4. Gray P, Scott SK. Sustained oscillations and other exotic patterns of behavior in isothermal reactions. J Phys Chem. 1985;89(1):22–32. doi:10.1021/j100247a009.
- Jahnke W, Skaggs WE, Winfree AT. Chemical vortex dynamics in the Belousov-Zhabotinskii reaction and in the two-variable oregonator model. J Phys Chem. 1989;93(2):740–749. doi:10.1021/j100339a047.
- 6. Falcke M. Reading the patterns in living cells —the physics of ca2+ signaling. Adv Phys. 2004;53(3):255-440. doi:10.1080/00018730410001703159.
- 7. Winfree AT. The Geometry of Biological Time. Springer New York; 2001.
- Kapral R, Showalter K. Chemical Waves and Patterns. Springer Science & Business Media; 2012.
- 9. Izhikevich EM. Dynamical Systems in Neuroscience. MIT Press; 2007.
- 10. Keener J, Sneyd J, editors. Mathematical Physiology. vol. 1: Cellular Physiology of Interdisciplinary Applied Mathematics. Springer New York; 2009.
- 11. Iglesias PA. Excitable systems in cell motility. In: 52nd IEEE Conference on Decision and Control; 2013. p. 757–762.
- Bretschneider T, Anderson K, Ecke M, Müller-Taubenberger A, Schroth-Diez B, Ishikawa-Ankerhold HC, et al. The three-dimensional dynamics of actin waves, a model of cytoskeletal self-organization. Biophys J. 2009;96(7):2888–2900. doi:10.1016/j.bpj.2008.12.3942.
- Arai Y, Shibata T, Matsuoka S, Sato MJ, Yanagida T, Ueda M. Self-organization of the phosphatidylinositol lipids signaling system for random cell migration. Proceedings of the National Academy of Sciences. 2010;107(27):12399–12404. doi:10.1073/pnas.0908278107.
- 14. Bement WM, Leda M, Moe AM, Kita AM, Larson ME, Golding AE, et al. Activator-inhibitor coupling between Rho signalling and actin assembly makes the cell cortex an excitable medium. Nat Cell Biol. 2015;17(11):1471–1483. doi:10.1038/ncb3251.
- Weiner OD, Marganski WA, Wu LF, Altschuler SJ, Kirschner MW. An actin-based wave generator organizes cell motility. PLoS Biol. 2007;5(9):2053–2063. doi:10.1371/journal.pbio.0050221.
- 16. Case LB, Waterman CM. Adhesive F-actin waves: a novel integrin-mediated adhesion complex coupled to ventral actin polymerization. PLoS One. 2011;6(11):e26631. doi:10.1371/journal.pone.0026631.
- 17. Vicker MG. Eukaryotic cell locomotion depends on the propagation of self-organized reaction-diffusion waves and oscillations of actin filament assembly. Exp Cell Res. 2002;275(1):54–66. doi:10.1006/excr.2001.5466.

- 18. Winans AM, Collins SR, Meyer T. Waves of actin and microtubule polymerization drive microtubule-based transport and neurite growth before single axon formation. Elife. 2016;5:e12387. doi:10.7554/eLife.12387.
- 19. Marchesin V, Montagnac G, Chavrier P. ARF6 promotes the formation of Rac1 and WAVE-dependent ventral F-actin rosettes in breast cancer cells in response to epidermal growth factor. PLoS One. 2015;10(3):e0121747. doi:10.1371/journal.pone.0121747.
- 20. Wu M, Wu X, De Camilli P. Calcium oscillations-coupled conversion of actin travelling waves to standing oscillations. Proc Natl Acad Sci U S A. 2013;110(4):1339–1344. doi:10.1073/pnas.1221538110.
- 21. Yang JM, Bhattacharya S, West-Foyle H, Hung CF, Wu TC, Iglesias PA, et al. Integrating chemical and mechanical signals through dynamic coupling between cellular protrusions and pulsed ERK activation. Nat Commun. 2018;9(1):4673. doi:10.1038/s41467-018-07150-9.
- 22. Banerjee T, Biswas D, Pal DS, Miao Y, Iglesias PA, Devreotes PN. Spatiotemporal dynamics of membrane surface charge regulates cell polarity and migration. Nat Cell Biol. 2022;doi:10.1038/s41556-022-00997-7.
- 23. Miao Y, Bhattacharya S, Banerjee T, Abubaker-Sharif B, Long Y, Inoue T, et al. Wave patterns organize cellular protrusions and control cortical dynamics. Mol Syst Biol. 2019;15(3):e8585. doi:10.15252/msb.20188585.
- Matsuoka S, Ueda M. Mutual inhibition between PTEN and PIP3 generates bistability for polarity in motile cells. Nat Commun. 2018;9(1):4481. doi:10.1038/s41467-018-06856-0.
- 25. Taniguchi D, Ishihara S, Oonuki T, Honda-Kitahara M, Kaneko K, Sawai S. Phase geometries of two-dimensional excitable waves govern self-organized morphodynamics of amoeboid cells. Proc Natl Acad Sci U S A. 2013;110(13):5016–5021. doi:10.1073/pnas.1218025110.
- Devreotes PN, Bhattacharya S, Edwards M, Iglesias PA, Lampert T, Miao Y. Excitable Signal Transduction Networks in Directed Cell Migration. Annu Rev Cell Dev Biol. 2017;33(1):annurev-cellbio-100616-060739. doi:10.1146/annurev-cellbio-100616-060739.
- 27. Ridley AJ, Schwartz MA, Burridge K, Firtel RA, Ginsberg MH, Borisy G, et al. Cell migration: integrating signals from front to back. Science. 2003;302(5651):1704–1709. doi:10.1126/science.1092053.
- 28. Miao Y, Bhattacharya S, Edwards M, Cai H, Inoue T, Iglesias PA, et al. Altering the threshold of an excitable signal transduction network changes cell migratory modes. Nat Cell Biol. 2017;19(4):329–340. doi:10.1038/ncb3495.
- 29. Zhan H, Bhattacharya S, Cai H, Iglesias PA, Huang CH, Devreotes PN. An Excitable Ras/PI3K/ERK Signaling Network Controls Migration and Oncogenic Transformation in Epithelial Cells. Dev Cell. 2020;54(5):608–623.e5. doi:10.1016/j.devcel.2020.08.001.
- 30. Huang CH, Tang M, Shi C, Iglesias PA, Devreotes PN. An excitable signal integrator couples to an idling cytoskeletal oscillator to drive cell migration. Nat Cell Biol. 2013;15(11):1307–1316. doi:10.1038/ncb2859.
- 31. Fukushima S, Matsuoka S, Ueda M. Excitable dynamics of Ras triggers spontaneous symmetry breaking of PIP3 signaling in motile cells. J Cell Sci. 2019;132(5). doi:10.1242/jcs.224121.
- 32. Flemming S, Font F, Alonso S, Beta C. How cortical waves drive fission of motile cells. Proc Natl Acad Sci U S A. 2020;117(12):6330–6338. doi:10.1073/pnas.1912428117.

- 33. Pal DS, Banerjee T, Lin Y, de Trogoff F, Borleis J, Iglesias PA, et al. Actuation of single downstream nodes in growth factor network steers immune cell migration. Dev Cell. 2023;58(13):1170–1188.e7. doi:10.1016/j.devcel.2023.04.019.
- 34. Lin Y, Pal DS, Banerjee P, Banerjee T, Qin G, Deng Y, et al. Ras suppression potentiates rear actomyosin contractility-driven cell polarization and migration. Nat Cell Biol. 2024;26(7):1062–1076. doi:10.1038/s41556-024-01453-4.
- 35. Shi C, Huang CH, Devreotes PN, Iglesias PA. Interaction of motility, directional sensing, and polarity modules recreates the behaviors of chemotaxing cells. PLoS Comput Biol. 2013;9(7):e1003122. doi:10.1371/journal.pcbi.1003122.
- 36. Banerjee T, Matsuoka S, Biswas D, Miao Y, Pal DS, Kamimura Y, et al. A dynamic partitioning mechanism polarizes membrane protein distribution. Nat Commun. 2023;14(1):7909. doi:10.1038/s41467-023-43615-2.
- 37. Karniadakis GE, Kevrekidis IG, Lu L, Perdikaris P, Wang S, Yang L. Physics-informed machine learning. Nature Reviews Physics. 2021;3(6):422–440. doi:10.1038/s42254-021-00314-5.
- 38. Brunton SL, Kutz JN. Data-driven science and engineering: Machine learning, dynamical systems, and control. Cambridge, England: Cambridge University Press (Virtual Publishing); 2019.
- 39. Brunton SL, Proctor JL, Kutz JN. Discovering governing equations from data by sparse identification of nonlinear dynamical systems. Proc Natl Acad Sci U S A. 2016;113(15):3932–3937. doi:10.1073/pnas.1517384113.
- 40. Rudy SH, Brunton SL, Proctor JL, Kutz JN. Data-driven discovery of partial differential equations. Sci Adv. 2017;3(4):e1602614. doi:10.1126/sciadv.1602614.
- 41. Van Breugel F, Liu Y, Brunton BW, Kutz JN. PyNumDiff: A Python package for numerical differentiation of noisy time-series data. J Open Source Softw. 2022;7(71):4078. doi:10.21105/joss.04078.
- 42. de Silva B, Champion K, Quade M, Loiseau JC, Kutz J, Brunton S. PySINDy: A Python package for the sparse identification of nonlinear dynamical systems from data. J Open Source Softw. 2020;5(49):2104. doi:10.21105/joss.02104.
- 43. Kaheman K, Kutz JN, Brunton SL. SINDy-PI: a robust algorithm for parallel implicit sparse identification of nonlinear dynamics. Proc Math Phys Eng Sci. 2020;476(2242):20200279. doi:10.1098/rspa.2020.0279.
- 44. Yazdani A, Lu L, Raissi M, Karniadakis GE. Systems biology informed deep learning for inferring parameters and hidden dynamics. PLoS Comput Biol. 2020;16(11):e1007575. doi:10.1371/journal.pcbi.1007575.
- 45. Dietrich F, Makeev A, Kevrekidis G, Evangelou N, Bertalan T, Reich S, et al. Learning effective stochastic differential equations from microscopic simulations: Linking stochastic numerics to deep learning. Chaos. 2023;33(2):023121. doi:10.1063/5.0113632.
- 46. Boninsegna L, Nüske F, Clementi C. Sparse learning of stochastic dynamical equations. J Chem Phys. 2018;148(24):241723. doi:10.1063/1.5018409.
- 47. Callaham JL, Loiseau JC, Rigas G, Brunton SL. Nonlinear stochastic modelling with Langevin regression. Proc Math Phys Eng Sci. 2021;477(2250):20210092. doi:10.1098/rspa.2021.0092.

- 48. Lade SJ. Finite sampling interval effects in Kramers–Moyal analysis. Phys Lett A. 2009;373(41):3705–3709. doi:10.1016/j.physleta.2009.08.029.
- 49. Friedrich R, Siegert S, Peinke J, Lück S, Siefert M, Lindemann M, et al. Extracting model equations from experimental data. Phys Lett A. 2000;271(3):217–222. doi:10.1016/S0375-9601(00)00334-0.
- Friedrich R, Renner C, Siefert M, Peinke J. Comment on "indispensable finite time corrections for Fokker-Planck equations from time series data". Phys Rev Lett. 2002;89(14). doi:10.1103/physrevlett.89.149401.
- 51. Siegert S, Friedrich R, Peinke J. Analysis of data sets of stochastic systems. Phys Lett A. 1998;243(5):275–280. doi:10.1016/S0375-9601(98)00283-7.
- 52. Amselem G, Theves M, Bae A, Bodenschatz E, Beta C. A stochastic description of Dictyostelium chemotaxis. PLoS One. 2012;7(5):e37213. doi:10.1371/journal.pone.0037213.
- 53. Fitzhugh R. Impulses and Physiological States in Theoretical Models of Nerve Membrane. Biophys J. 1961;1(6):445–466. doi:10.1016/s0006-3495(61)86902-6.
- 54. Nagumo J, Arimoto S, Yoshizawa S. An Active Pulse Transmission Line Simulating Nerve Axon. Proceedings of the IRE. 1962;50(10):2061–2070. doi:10.1109/JRPROC.1962.288235.
- 55. Bhattacharya S, Iglesias PA. The Regulation of Cell Motility Through an Excitable Network. IFAC-PapersOnLine. 2016;49(26):357–363. doi:10.1016/j.ifacol.2017.03.001.
- 56. Bhattacharya S, Iglesias PA. The threshold of an excitable system serves as a control mechanism for noise filtering during chemotaxis. PLoS One. 2018;13(7):e0201283. doi:10.1371/journal.pone.0201283.
- 57. Hastie T, Tibshirani R, Friedman J. The Elements of Statistical Learning: Data Mining, Inference, and Prediction. 2nd ed. New York, NY: Springer; 2009.
- 58. Friedrich R, Peinke J, Sahimi M, Reza Rahimi Tabar M. Approaching complexity by stochastic methods: From biological systems to turbulence. Phys Rep. 2011;506(5):87–162. doi:10.1016/j.physrep.2011.05.003.