

MECHANISMS IN ENDOCRINOLOGY

Human brown adipose tissue as a therapeutic target: warming up or cooling down?

Ben T McNeill, Karla J Suchacki and Roland H Stimson

University/BHF Centre for Cardiovascular Science, University of Edinburgh, Queen's Medical Research Institute, Edinburgh, UK

Correspondence
should be addressed
to R H Stimson**Email**
roland.stimson@ed.ac.uk**Abstract**

Excessive accumulation of white adipose tissue leads to obesity and its associated metabolic health consequences such as type 2 diabetes and cardiovascular disease. Several approaches to treat or prevent obesity including public health interventions, surgical weight loss, and pharmacological approaches to reduce caloric intake have failed to substantially modify the increasing prevalence of obesity. The (re-)discovery of active brown adipose tissue (BAT) in adult humans approximately 15 years ago led to a resurgence in research into whether BAT activation could be a novel therapy for the treatment of obesity. Upon cold stimulus, BAT activates and generates heat to maintain body temperature, thus increasing energy expenditure. Activation of BAT may provide a unique opportunity to increase energy expenditure without the need for exercise. However, much of the underlying mechanisms surrounding BAT activation are still being elucidated and the effectiveness of BAT as a therapeutic target has not been realised. Research is ongoing to determine how best to expand BAT mass and activate existing BAT; approaches include cold exposure, pharmacological stimulation using sympathomimetics, browning agents that induce formation of thermogenic beige adipocytes in white adipose depots, and the identification of factors secreted by BAT with therapeutic potential. In this review, we discuss the caloric capacity and other metabolic benefits from BAT activation in humans and the role of metabolic tissues such as skeletal muscle in increasing energy expenditure. We discuss the potential of current approaches and the challenges of BAT activation as a novel strategy to treat obesity and metabolic disorders.

*European Journal of
Endocrinology*
(2021) **184**, R243–R259**Invited Author's profile**

Roland H Stimson is Professor of Endocrinology and a Scottish Senior Clinical Fellow at the University of Edinburgh and an honorary consultant physician at the Royal Infirmary of Edinburgh, UK. His research background is in human experimental medicine and integrative physiology. Prof Stimson's overarching research interest is in obesity and its associated metabolic disease. His laboratory's current focus is on investigating the role of human brown and white adipose tissue and skeletal muscle in the regulation of energy balance with a goal to identify novel pathways amenable to therapeutic manipulation to develop new treatments for obesity and type 2 diabetes mellitus.



Introduction

The prevalence of obesity has increased dramatically over the past 40 years, with over a quarter of adults in the UK now classed as obese (1). During the same period of time, global obesity has increased more than three-fold and this is likely to continue (2, 3). Obesity (defined as a BMI ≥ 30 kg/m²) occurs when energy intake chronically exceeds energy expenditure (EE) with deposition of this excess energy primarily as triglycerides in white adipose tissue (WAT). Obesity substantially increases the risk of developing other diseases such as type 2 diabetes mellitus (T2DM), hypertension, dyslipidaemia and cardiovascular disease and decreases both life expectancy and quality (4, 5). Public health interventions to promote weight loss through dietary caloric restriction and increased physical activity have failed to curb the rise in obesity due to lack of adherence (5). Pharmacological anti-obesity agents have focused on decreasing energy intake/appetite but

have had limited success, in part due to serious adverse side effects leading to their withdrawal (most recently Lorcaserin) (6). Orlistat (a pancreatic lipase inhibitor) and naltrexone-bupropion are the only licensed medications in the UK for obesity and substantial side effects limit patient compliance; other agents are licensed in the USA but are not approved in Europe (Fig. 1) (7). Bariatric surgery can successfully treat obesity (8); however, these procedures are invasive, can cause significant complications and are not suitable for everyone.

There has been less effort on the development of pharmacotherapy to specifically increase EE (energy balance equation; Fig. 1). However, the use of ¹⁸F-fluorodeoxyglucose (¹⁸F-FDG) PET coupled with CT (PET/CT) to diagnose certain malignancies led to the incidental (re-)discovery of brown adipose tissue (BAT) in adult humans approximately 15 years ago (9, 10). This finding has re-ignited interest in this approach to treat obesity (11), as BAT activation in rodents increases EE and improves

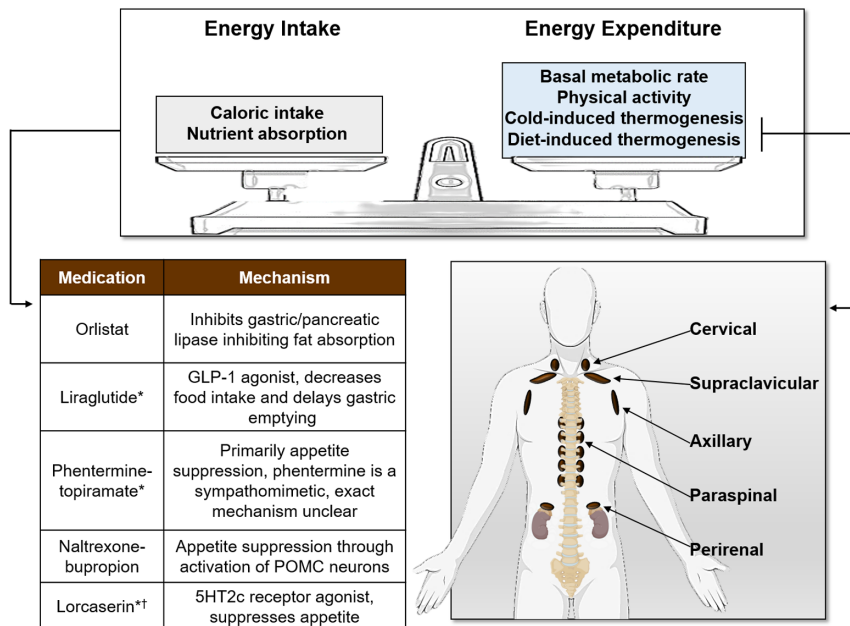


Figure 1

The energy balance equation and current pharmacotherapy to achieve weight loss. Energy balance is governed by the relationship between energy input (calories consumed) and energy output (energy expended). Obesity results from a chronic imbalance of energy intake exceeding energy expenditure with storage of this excess energy as triglycerides mainly in white adipose tissue. All licensed anti-obesity medications primarily cause weight loss by reducing appetite/energy intake (*indicates licensed to treat obesity in US only, †indicates currently withdrawn). Basal metabolic rate, physical activity, diet-induced thermogenesis (DIT) and cold-induced thermogenesis (CIT) all contribute to total energy expenditure. Brown adipose tissue (BAT) is located in adult humans primarily in the cervical, supraclavicular, axillary, paravertebral and peri-renal regions. BAT activation is a key component of both CIT and DIT and is an attractive target to increase energy expenditure to treat obesity. 5HT, 5-hydroxytryptamine; GLP-1, glucagon-like peptide 1; POMC, pro-opiomelanocortin.

insulin sensitivity (12) and dyslipidaemia (13). Thereafter, a number of elegant studies have been undertaken to determine the role, importance and regulation of BAT in humans. This review will discuss the recent advances in our understanding of the physiology and pathophysiology of human BAT, potential approaches to activate BAT and discuss whether this tissue represents a viable therapeutic target for obesity and its associated metabolic disease.

The distribution and function of brown and beige adipose tissue

Predominantly an organ for energy storage, WAT is widely distributed throughout the body and divided into s.c. and visceral depots. Conversely, the primary function of BAT is to generate heat to maintain the body temperature through non-shivering thermogenesis (NST) during cold exposure (14) and is located in the cervical, supraclavicular, axillary, paraspinal, and perirenal regions (Fig. 1) (15, 16, 17). Unlike WAT, BAT contains multilocular

lipid droplets and a high number of mitochondria expressing the thermogenic protein mitochondrial brown fat uncoupling protein 1 (UCP1) (18). When activated, UCP1 dissipates the proton electrochemical gradient across the inner mitochondrial matrix with the energy released as heat in a process termed 'uncoupling' (Fig. 2) (14). Cold exposure stimulates the sympathetic neurones innervating BAT to release noradrenaline which activates β -adrenergic receptors (AR) (classically β_3 -AR but also β_1 - and β_2 -AR (19, 20, 21, 22)). β -AR activation triggers a signalling cascade which results in the hydrolysis of local triglycerides, releasing fatty acids (FA) that activate UCP1 (Fig. 2). In addition to triglyceride stores, BAT sequesters and utilises several circulating substrates such as glucose, fatty acids and some amino acids during thermogenesis (reviewed in (23)). Therefore, BAT activation may improve other metabolic health parameters such as hyperglycaemia and dyslipidaemia in addition to increasing energy expenditure.

In rodents, two distinct types of thermogenic adipose tissue have been identified, classical BAT and beige or

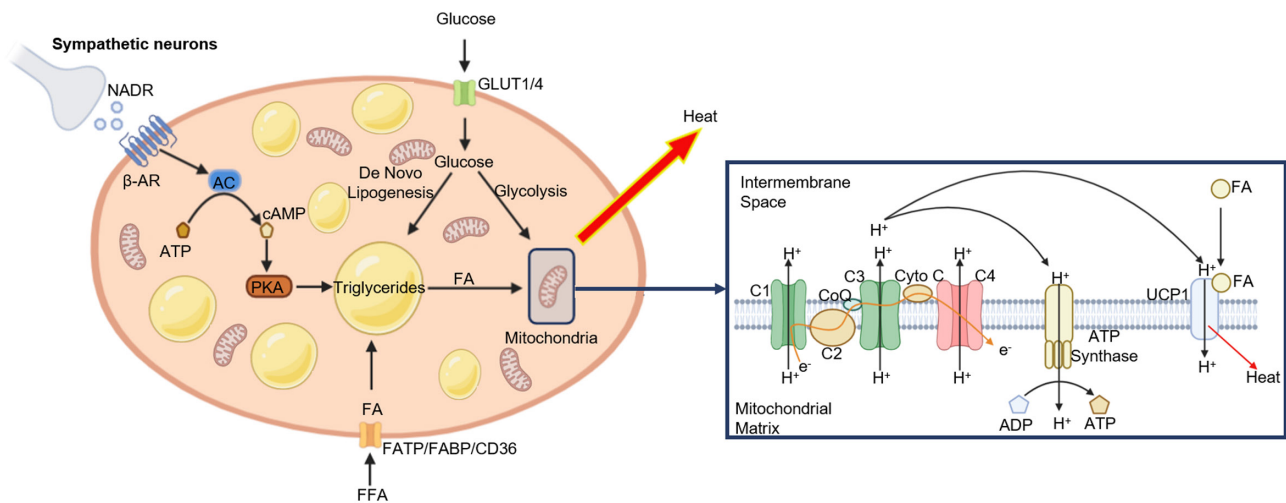


Figure 2

Brown adipocyte activation and molecular mechanism of UCP1 function. Upon cold stimulus, sympathetic neurons innervating BAT release noradrenaline (NADR) from the synapse. NADR binds to various β -adrenergic receptors (β -AR) on the brown adipocyte which activates adenylyl cyclase (AC), converting ATP to cyclic adenosine monophosphate (cAMP). cAMP activates protein kinase A (PKA) which stimulates the lipolysis of triglyceride stores and release of fatty acids (FA). FAs are the primary substrate for thermogenesis but also bind and activate uncoupling protein 1 (UCP1) located in the mitochondria. UCP1 generates heat via transport of protons (H^+) across the inner mitochondrial member using the electrochemical proton gradient generated by the electron transport train, uncoupling respiration from ATP synthase. Uptake of circulating free fatty acids (FFA) and glucose contribute to the regeneration of intracellular triglyceride stores, additionally glucose can be oxidised and enter the tricarboxylic acid (TCA) cycle. FFAs are transported into the cell by fatty acid transport protein (FATP, fatty acid binding protein (FABP), and cluster of differentiation 36 (CD36). Glucose is transported into the cell via the glucose transporters GLUT1 and 4. C1–4, complex 1–4; CoQ, co-enzyme Q; Cyto C, cytochrome C; e^- , electron.

brite. Classical BAT is found mainly in the interscapular region and is derived from myogenic factor 5-positive precursors (24), whilst beige fat is found primarily in the inguinal depot and is derived from different progenitor cells (reviewed in (25)). Beige adipocytes are recruited in response to cold or β -adrenergic stimulation (26), express UCP1 (although levels are substantially lower than in BAT) (26) and contribute to thermogenesis (27, 28). Also thermogenic in nature, human BAT has comparable UCP1 function to rodent BAT (18) and interestingly, the molecular signature of human BAT shares similarities with both rodent classical BAT and beige adipose tissue (29).

The importance of brown adipose tissue in adult humans

Following the discovery of BAT in adults via clinical ^{18}F -FDG-PET/CT scans, healthy volunteer studies confirmed that cold exposure substantially increased ^{18}F -FDG uptake by BAT (17, 30, 31) and ^{18}F -FDG-PET/CT has become the most commonly used technique to quantify the BAT mass and activity (32). During warm conditions, BAT remains metabolically active, with greater glucose and fatty acid uptake compared to that of WAT (33, 34). Clinical ^{18}F -FDG-PET/CT scans performed at room temperature demonstrate that ~5–10% of individuals have detectable ^{18}F -FDG uptake by BAT (35); however, in dedicated studies when subjects are exposed to cold the prevalence is as high as 95% in young healthy men (17). BAT mass is substantially lower than WAT mass (~16–22 kg) even in normal weight adults (36, 37). The quantity of detectable BAT in humans ranges from ~10–300 g (30, 38, 39, 40), although this may be an underestimate as ^{18}F -FDG-PET may not identify all BAT depots and total BAT mass may be as high as ~2550 g (15). However, ^{11}C -acetate PET (used to measure BAT oxidative activity) has not revealed novel BAT depots without substantial glucose uptake (41) indicating that ^{18}F -FDG-PET/CT estimates may be accurate.

Regulation and dysregulation of human BAT activity

Clinical PET/CT studies identified that increased outdoor temperature and male sex were associated with reduced ^{18}F -FDG uptake by BAT (40, 42). However, dedicated cold exposure studies have not revealed substantial differences in BAT activity between sexes (43), potentially indicating that females activate their BAT at higher room

temperature than males. Ethnicity may also alter BAT mass/activity, which has been implicated in the greater risk of metabolic disease in individuals of South Asian origin (44). Perhaps, the most interesting observation was that reduced ^{18}F -FDG uptake by BAT was observed with increasing age, fasting glucose and body weight, implicating dysregulation of BAT activity in metabolic disease (17, 31, 42, 45). In addition, obese subjects have reduced fatty acid (using the PET tracer ^{18}F -fluoro-6-thiaheptadecanoic acid) uptake by BAT during both warm and cold exposure in keeping with decreased BAT mass and activity (34), although greater insulin resistance may also contribute to the reduced glucose/fatty acid uptake by BAT in obesity (46). Dysfunctional BAT in obese subjects could reduce EE and contribute to weight gain, as observed in mice with selective disruption of *Ucp1* that develop obesity when housed at thermoneutral conditions (47). However, *Ucp1*^{-/-} mice housed below thermoneutrality have resistance to diet-induced obesity due to decreased metabolic efficiency, highlighting that dysfunctional BAT does not necessarily cause weight gain and is dependent on the environmental conditions (48). BAT mass and ^{18}F -FDG uptake by BAT are also substantially reduced in older subjects (49) and in those with T2DM (49, 50), although interestingly oxidative metabolism is maintained, indicating that functional BAT is preserved in these cohorts (49). These data highlight a critical issue in the therapeutic potential of activating BAT, as the target patient groups require enough BAT mass and function to benefit from activation. Therefore, effective expansion of BAT mass will likely be required to obtain improvements in metabolic health.

There are substantial data that BAT mass can expand or regress in response to different stimuli. For example, in colder climates, greater BAT mass is found in individuals who work outdoors compared with indoor workers (51). Furthermore, repeated intermittent cold exposure for ~7–10 days increased BAT mass and glucose uptake (using ^{18}F -FDG-PET/CT), BAT oxidative metabolism, NST and wider cold-induced thermogenesis (CIT) (38, 52). Rare diseases also highlight the plasticity of BAT, as previously mentioned BAT activation is under sympathetic control and patients with catecholamine-secreting tumours (called pheochromocytomas) often have substantial BAT mass and function which regress upon surgical removal of the tumour (53, 54). Importantly, BAT function can be increased in obese subjects both with and without T2DM (the target patient group) by weight loss, which increased ^{18}F -FDG uptake by BAT, BAT volume and non-shivering thermogenesis in some subjects (50, 55, 56). These data

suggest that brown adipocyte precursors are present in individuals without detectable BAT and can differentiate into functional brown adipocytes upon appropriate stimulation. Therefore, treatments to increase BAT mass may be successful in patients with metabolic disease. These data also indicate that reduced BAT mass may be a consequence of obesity and it is to be determined whether activating BAT can cause weight loss in obese individuals. However, there are substantial differences in capacity and function of BAT even in healthy individuals, and further research is needed to determine the causes of this variability and whether reduced or absent BAT mass and function can be rescued in all subjects. In addition, it is important to consider other factors regulating BAT mass and function in individuals. For example, BAT activity demonstrates a circadian rhythm in both rodents and humans (57, 58) while dietary composition and timing of feeding/fasting have powerful effects on BAT activity and browning at least in rodents (59, 60, 61). Exercise may also regulate BAT activity, as seen in endurance-trained athletes who have reduced ^{18}F -FDG uptake by BAT during cold exposure compared with sedentary adults (62, 63).

Quantification of cold-induced thermogenesis and energy expenditure by BAT

Determining the maximal capacity of BAT is key to understanding its therapeutic potential. Early research estimated that 50 g of activated human BAT could increase EE by 20% above basal metabolic rate (64). In addition, EE increases by ~250–300 kcal/24 h during mild cold exposure (Fig. 3) and CIT is higher in subjects with greater BAT mass in some (65, 66) but not in all studies (67). However, the use of $^{15}\text{O}_2$ -PET suggested that BAT only accounts for a very small contribution to CIT, <20 kcal/24 h even in subjects with substantial BAT mass (67, 68). Thus, unlike in rodents, non-shivering thermogenesis accounts for a small proportion (~1%) of CIT in humans, indicating approaches to activate BAT alone will not significantly increase whole body EE. However, it is interesting to note that the deep muscles (particularly in the neck) located adjacent to BAT are responsible for the majority of CIT (67, 69). It is possible that greater sympathetic activation in subjects with BAT also increases skeletal muscle thermogenesis, or that BAT secretes factors that enhance EE in skeletal muscle in a paracrine fashion, as seen in rodents (70). In addition, interventions that increase BAT mass also increase wider CIT, highlighting the potential benefits of this approach (38). For example, repeated cold exposure at 17°C for 2 h/day for 6 weeks

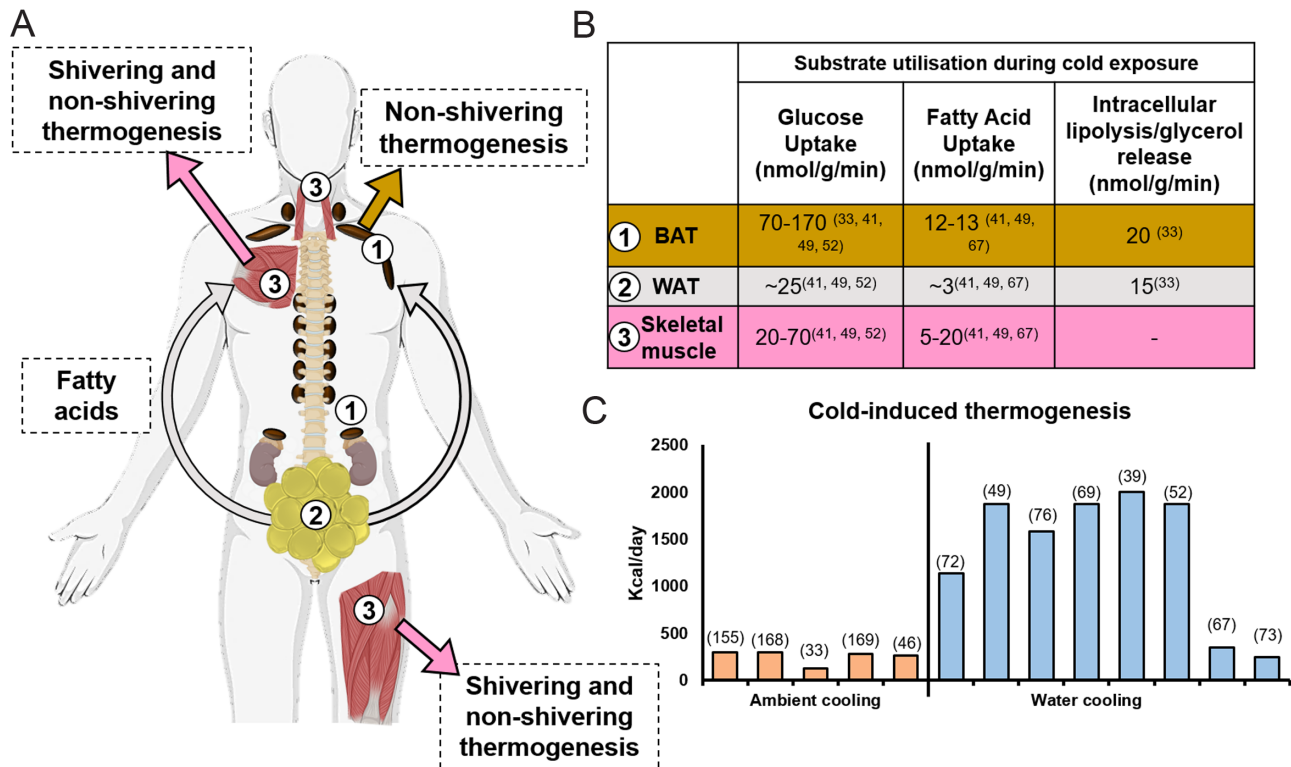
increased cold-induced ^{18}F -FDG uptake by BAT, CIT by ~200 kcal/day and reduced fat mass (66). This substantial increase in EE during acute cold exposure highlights the potential benefits of activating this pathway, to put this into context increasing EE by 50–60 kcal/day for 1 year would result in weight loss of ~2.5 kg (71) if there was no compensatory increase in food intake. Further research to dissect the pathways regulating CIT may identify novel targets for manipulation to increase EE.

Metabolic effects of BAT activation and cold-induced thermogenesis

The metabolic benefits of BAT activation and wider CIT extend beyond burning calories. During warm exposure when thermogenesis is not required, BAT sequesters and utilises circulating metabolic substrates such as glucose and FAs to a greater extent than WAT (33, 34). However, uptake of these substrates and others such as glutamate by BAT increases substantially following cold activation, in addition to hydrolysis and oxidation of its own triglyceride stores (33, 72). CIT also leads to increased glucose and FA uptake by skeletal muscle and lipolysis in WAT (41, 49, 52, 67, 73). Highlighting the substantial metabolic activity of BAT, glucose and potentially FA uptake are greater per gram of tissue in BAT than in either skeletal muscle or WAT (Fig. 3). However, skeletal muscle mass (~30 kg) is substantially greater than BAT mass and accounts for ~50% of whole body glucose uptake during cold, compared with ~1% for BAT (72).

As mentioned previously, in line with increased substrate utilisation BAT activation improves glucose homeostasis. For example, acute cold exposure in overweight/obese men increased glucose disposal and oxidation in addition to insulin sensitivity in subjects with detectable BAT, these changes were not observed in the 'BAT negative' group (74). Furthermore, 4 weeks of mild cold exposure (19°C for 10 h/night) in lean healthy men improved postprandial insulin sensitivity, reduced leptin levels and increased adiponectin concentrations in addition to increasing ^{18}F -FDG uptake by BAT (39). In T2DM subjects, cold exposure (~15°C for 2–6 h/day for 10 days) improved whole body insulin sensitivity (primarily through increased glucose disposal) and increased ^{18}F -FDG uptake by BAT and skeletal muscle but not WAT (50).

BAT activation is also associated with changes in circulating lipids. For example, in overweight/obese men those with greater BAT activation had increased cold-induced lipolysis, FA oxidation and adipose tissue insulin sensitivity (75). In addition, acute cold exposure

**Figure 3**

Whole body cold-induced thermogenesis and substrate utilisation. (A) Cold exposure stimulates WAT lipolysis to provide FAs for utilisation by both BAT and skeletal muscle (grey arrows). BAT uses FAs released from intracellular triglyceride stores to fuel non-shivering thermogenesis (orange arrow) but also sequesters circulating FAs and glucose. Skeletal muscle shivering accounts for the largest proportion of whole body heat production, glucose and FA uptake during cold-induced thermogenesis (CIT) (pink arrows). Muscles that contribute substantially to shivering thermogenesis include the longus colli, sternocleidomastoid, pectoralis major, and the rectus femoris. (B) During cold exposure, glucose uptake per gram of tissue is greater in BAT than skeletal muscle but with similar fatty acid uptake. However, whole body FA and glucose uptake by BAT is comparatively low due to substantially greater skeletal muscle mass. (C) Quantification of CIT varies greatly depending on the cooling method used and temperature, ambient air cooling protocols (orange columns) typically elicit a lower increase in energy expenditure compared to water cooling blanket/suit protocols (blue columns), but substantial CIT is induced by both methods. Additional references used for data in panel C (168,169).

decreased triglyceride and very low-density lipoproteins (VLDL)-cholesterol concentrations in those subjects the following day, suggesting BAT activation may have prolonged beneficial effects. This may be in part due to sequestration of fatty acids derived from triglyceride-rich lipoproteins (TRLs) (76). In addition, BAT thermogenesis is activated to a similar extent by a meal as by cold, this postprandial thermogenesis utilises TRL-derived FAs and glucose which may improve systemic FA oxidation in addition to glucose disposal (73). These data suggest that BAT activation and wider CIT may improve insulin resistance and dyslipidaemia in addition to increasing EE, making activation of this pathway an attractive prospect to treat metabolic disease. BAT activation may also have

additional beneficial effects, for example short-term cold exposure reduced local inflammation within fat depots (77) and the wider benefits of BAT activation and CIT remain relatively unexplored.

Approaches to activating BAT

As described above, there are clear metabolic benefits from acute activation of BAT and wider CIT, questions remain as to whether these improvements will be maintained during chronic activation. In addition, a major challenge for the field is how to safely achieve long-term expansion and activation of BAT.

Cold exposure

To date, the most common method to activate BAT in humans is either to reduce ambient room temperature to $\sim 16\text{--}19^\circ\text{C}$ (30, 33) or use a cold water-infused suit/jacket (41, 49). Both methods elicit similar levels of BAT activation at least as measured by glucose uptake (33, 41, 46, 52). Repeated cold exposure for several hours per day for up to 6 weeks increased BAT mass/activity (as measured using ^{18}F -FDG), CIT and decreased fat mass (38, 50, 66). Although it is possible that the increased ^{18}F -FDG uptake demonstrates the activation of previously dormant BAT, in rodents repeated cold exposure leads to differentiation of new thermogenic beige adipocytes (78) and the same is most likely true in humans. Importantly, these studies reveal that short-term cold exposure improves cardiometabolic markers and potentially decreases fat mass in humans without the need for pharmacotherapy. However, this technique is time-consuming and may be uncomfortable for patients. It is also unclear whether these benefits are maintained over time and ongoing studies will determine whether chronic cold exposure (or repeated short-term cold exposure for several months) improves metabolic health (79, 80).

Increased ambient temperature is associated with the prevalence of obesity in some (81, 82) but not in all studies (83). Indoor housing temperatures in the UK have increased since the 1970s, potentially due to greater use of central heating including in more energy-efficient homes (84, 85, 86). The reduced requirement for CIT due to warmer ambient temperatures could lower EE (Fig. 3) and contribute to the increased prevalence of obesity. Therefore, a concerted effort to reduce room temperature through the reduced usage of central heating to increase EE may have metabolic benefits in addition to being the most cost-effective 'therapeutic' option in cold climates such as the UK. However, in rodents, intermittent cold exposure causes a compensatory increase in food intake to meet the increased thermogenic demands (12), while in humans increased EE during cold exposure results in a parallel increase in *ad libitum* food intake (87). Therefore, while cold exposure may improve metabolic health this may not necessarily result in weight loss.

Pharmacotherapy for BAT activation

Pharmacotherapy to activate BAT (and potentially other tissues involved in CIT) is an attractive option as this would be a more comfortable method of activation. However, a subject must have enough BAT to respond

to a 'BAT activator' particularly as the target patient group (typically obese subjects with T2DM who may be older) generally have very little BAT. Therefore, the ideal drug would expand BAT mass in addition to activating BAT. As BAT expansion and activation are both under sympathetic regulation (53, 88), the majority of research in this area has focused on the effect of sympathetic agonists.

Sympathomimetics

Activation of the β_3 -AR induces browning and BAT thermogenesis, while administration of β_3 -agonists induces weight loss and improves hyperglycaemia in rodents (89). Consequently, there was significant interest in β_3 -AR agonists in humans even prior to the recent identification of BAT in adult humans (90, 91). β_3 -agonist administration for 4–8 weeks in humans improved lipids and insulin sensitivity although there was no effect on body weight (92, 93). More recently, a single high dose (200 mg) of the β_3 -AR agonist mirabegron (licensed for urinary frequency/incontinence) in humans housed at 23°C increased ^{18}F -FDG uptake by BAT and increased EE by ~ 200 kcal/24 h (19). In addition, an administration of 100 mg mirabegron daily for 4 weeks in healthy women increased BAT mass and volume (using ^{18}F -FDG) and increased EE but did not alter body weight (94). In accordance with the earlier studies, mirabegron improved insulin sensitivity and increased high-density lipoprotein cholesterol in these subjects. However, a lower dose of mirabegron (50 mg) did not activate BAT thermogenesis (22) and the effects at higher doses may be due to off target activation of particularly the β_2 -AR and also β_1 -AR that are more highly expressed in human BAT than β_3 -AR (21, 22). These data highlight the difficulties with developing selective adrenergic receptor agonists to activate BAT.

The mixed adrenoceptor agonist ephedrine (which also inhibits noradrenaline re-uptake in post-synaptic neurons (95, 96)) also increased ^{18}F -FDG uptake by BAT in lean (but not obese) adults at room temperature (97). However, ephedrine also increased heart rate and blood pressure, side effects also induced by mirabegron that limit the potential of this approach (19, 94, 97). In addition, lower dose ephedrine (1 mg/kg) did not acutely activate BAT in healthy humans (98) while chronic administration may in fact reduce BAT activity (99). These data suggest that chronic sympathetic stimulation may result in desensitisation in BAT.

Thyroid hormones

Thyroid hormone receptor activation is crucial for BAT thermogenesis and adrenergic responsiveness in mice (100). Similarly, individuals with thyrotoxicosis have increased BAT glucose uptake, lipid oxidation, EE, and possibly improved insulin sensitivity; these changes are reversed once euthyroidism is restored (101, 102). A recent trial also demonstrated that administration of thyrotropin-releasing hormone in healthy subjects increased glucose uptake by BAT but only during cold exposure (103). While these studies highlight the importance of the thyroid in BAT activation, the long-term effects of thyroid hormone administration on BAT function are unknown.

Other drugs known to activate BAT

Capsaicin and capsinoids are substances naturally present in chilli peppers that are agonists of the transient receptor potential vanilloid type 1 (TRPV1) receptor (104). In rodents, capsinoids stimulate sympathetic activation of BAT and increase UCP1 expression in both BAT and WAT (104, 105). In healthy humans, acute ingestion of capsinoids (9 mg) significantly increased whole body EE only in subjects with detectable BAT (106). Chronic capsinoid supplementation (9 mg daily for 6 weeks) increased CIT in healthy subjects (66) and potentially increased resting EE in overweight individuals (107), although whether these effects are mediated by BAT is unclear. These data provide proof-of-concept that dietary supplementation could be a relatively safe method to increase EE and BAT activity.

Produced by the liver and modified by gut microbiota, bile acids (BA) are released into the intestinal lumen and circulation in the postprandial period and regulate metabolism (reviewed in (108)). In rodents, BAs increase BAT thermogenesis and induce browning of WAT through the G-protein-coupled bile acid receptor TGR5 and the cyclic-AMP-dependent thyroid hormone activating enzyme type 2 iodothyronine deiodinase (109, 110, 111). In humans, administration of the BA chenodeoxycholic acid (CDCA) for 2 days increased BAT activation and whole body EE *in vivo* (112) and CDCA increased mitochondrial uncoupling in human brown (but not white) adipocytes through TGR5 (112).

In rodents, acute and chronic glucocorticoid excess decreases UCP1 expression and reduces BAT thermogenesis, conversely in humans acute glucocorticoid excess increases UCP1 and oxygen consumption *in vitro* and increases ¹⁸F-FDG uptake and heat production by BAT and CIT *in vivo* (113, 114, 115). However, chronic glucocorticoid excess

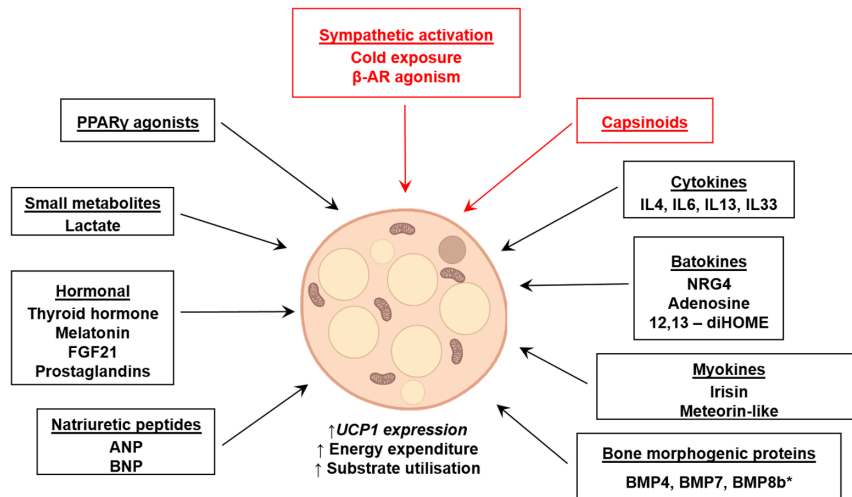
reduces BAT function so would not be an appropriate therapeutic agent (114, 116), but importantly, these data highlight the species-specific regulation of BAT activation.

Browning agents to enhance thermogenesis

Due to the relatively small quantity of BAT in adult humans, upon activation, the systemic clearance of glucose and caloric capacity is substantially lower than skeletal muscle (69, 72). Therefore, therapeutic strategy is to expand the thermogenic adipose tissue mass in a process termed 'browning' (Fig. 4). Increasing WAT thermogenesis could have profound metabolic effects as, in obese subjects, WAT accounts for over one-quarter of total body weight (117). Cold exposure is a powerful inducer of browning in rodents (78) and increases supraclavicular BAT mass in humans; however, studies have not demonstrated increased glucose uptake (118) or oxidative capacity (52) (measured by ¹¹C-acetate) in classical WAT depots *in vivo* following repeated cold exposure. These data suggest either that repeated cold exposure is not a sufficient stimulus to increase WAT thermogenesis or that classical WAT depots have a low browning capacity. Pharmacotherapy may hold greater promise, browning agents such as β 3-agonists can induce formation of UCP1-positive 'beige' thermogenic adipocytes in WAT depots in both rodents and humans, although as discussed above their adverse effects limit their potential (26, 119, 120).

Numerous browning agents have been identified in rodents, some of which have been investigated in humans. For example, fibroblast growth factor-21 (FGF21) release is induced by cold and increases UCP1 expression in murine WAT and BAT, and in human adipocytes (121, 122, 123). FGF21 is expressed in BAT (123, 124), although hepatic FGF21 primarily accounts for circulating levels and the beneficial metabolic effects (125). FGF21 analogue administration to obese humans with T2DM improved circulating lipids and reduced body weight (potentially by reducing food intake although this was not measured), although it did not improve glucose levels (126, 127). However, BAT activity and WAT browning were not measured in these studies so it is unclear if adipose thermogenesis contributed to the observed metabolic improvements, further research is needed to determine if FGF21 administration *in vivo* induces browning in humans. FGF21 also induces bone loss, which lessens the therapeutic potential of this approach (128).

Several bone morphogenic proteins (BMPs, members of the transforming growth factor superfamily) drive

**Figure 4**

Factors that induce browning of typical WAT depots. An illustration of factors that induce thermogenic beige adipocyte formation *in vivo* and *in vitro* with greater UCP1 expression and uncoupled respiration compared to white adipocytes. The small number of factors that induce browning of white adipose tissue *in vivo* in humans are highlighted in red and underlined. *BMP8b is classed additionally as a BATokine. 12,13-dihOME, 12,13-dihydroxy-9Z-octadecenoic acid; β -AR, β -adrenoreceptor; BMP, bone morphogenic protein; BNP, brain natriuretic peptide; FGF21, fibroblast growth factor-21; IL, interleukin; NRG4, neuregulin-4; PPAR γ , peroxisome proliferator-activated receptor- γ .

brown adipogenesis, notably BMP7. Transgenic disruption of *Bmp7* in mice substantially reduced UCP1 expression and BAT mass, while its overexpression increased UCP1 and EE and reduced body mass (129). In addition, BMP7 (and BMP4) induces browning in human adipocyte cell models (130, 131). However, BMP7 may only work as a browning agent below thermoneutrality, which is an important consideration for any therapeutic agent (132). The effect of BMP7 on metabolic health has not been studied *in vivo* in humans, although BMP7 has been FDA approved for bone fracture treatment in clinical trials (133).

Thiazolidinediones are peroxisome proliferator-activated receptor- γ (PPAR γ) agonists used as insulin-sensitising drugs for the treatment of T2DM, although adverse side effects have reduced their use substantially (134). In rodents, PPAR γ -agonists are powerful browning agents, both *in vivo* and *in vitro* (135, 136). In humans, PPAR γ -agonists also induce browning in adipocytes but *in vivo* in fact reduce cold-induced ^{18}F -FDG uptake by BAT (137). Another anti-diabetic drug, the dipeptidyl peptidase-4 (DPP-IV) inhibitor sitagliptin decreased body weight, increased energy expenditure and increased UCP1 protein expression in BAT in obese mice (138). However, DPP-IV inhibition using sitagliptin in overweight pre-diabetic subjects for 12 weeks increased ^{18}F -FDG uptake slightly by subcutaneous WAT but not by BAT during cold exposure (139). Therefore, it is unlikely that DPP-IV inhibitors induce substantial browning in humans.

Irisin is another browning agent that has received substantial attention. Irisin is secreted from skeletal muscle during exercise and substantially increases UCP1 expression in inguinal WAT, increases EE and protects against weight gain in mice (140). In humans, irisin induced browning in white adipocytes *in vitro* while circulating irisin concentrations are increased by exercise and cold exposure (122, 140). However, irisin had no effect on brown adipocytes and may also induce osteogenesis (141). Data in humans on the effects of exercise programmes on circulating irisin levels and induction of browning are inconsistent, calling into question whether irisin mediates any of the beneficial effects of exercise, although methodologies often vary widely between studies (142, 143). Despite differences in methodology to quantify irisin concentrations, systemic irisin levels are increased in obesity which may reduce the potential of using irisin as a therapeutic agent, although levels are likely decreased in T2DM (144). The effect of irisin administration *in vivo* in humans is yet to be tested. While numerous factors have been identified as browning agents in rodents (145), the above data highlight the importance of assessing the effect of browning agents *in vivo* in humans and at present there are very limited data on the majority of these factors in humans. β -agonists have successfully demonstrated proof-of-concept but further work is necessary to dissect the mechanisms regulating adipose tissue browning in humans and the thermogenic capacity of various WAT

depots to determine the therapeutic potential of this approach.

Secreted factors from BAT

As previously discussed, the presence of BAT is associated with favourable metabolic profiles (74) and there is recent evidence BAT secretes factors (often termed 'BATokines', although most if not all of these factors are also secreted from other tissues) with beneficial paracrine and endocrine functions. Consequently, there is substantial interest in identifying BATokines with therapeutic potential. Many factors have been identified, the vast majority in rodents. BMP8b has gained interest as a BATokine as its expression is induced in BAT by cold exposure and BMP8b directly enhances sympathetic-stimulated BAT thermogenesis (146). In addition, secreted BMP8b increased adipose tissue browning through enhanced sympathetic innervation and vascularisation even at thermoneutrality in mice through secretion of another BATokine neuregulin-4 (NRG4) (147). NRG4 also exerts beneficial metabolic effects in other tissues such as the liver where it inhibits *de novo* lipogenesis (148) and increases FA oxidation, while NRG4 also increases glucose uptake in skeletal muscle (149). In humans, NRG4 expression in WAT is reduced in obese subjects and circulating levels are lower in non-alcoholic fatty liver disease (150). However, the contribution of BAT to total serum NRG4 is unknown and therapeutic administration has not been tested in humans.

In both rodents and humans, circulating concentrations of the BATokine 12,13-dihydroxy-9Z-octadecenoic acid (12,13-diHOME) are increased following cold exposure (151) and exercise (152). In mice, 12,13-diHOME administration increased fatty acid uptake by brown/beige adipocytes and skeletal muscle (151, 152). In addition, plasma 12,13-diHOME concentrations in humans were inversely associated with fat mass, fasting insulin and triglyceride concentrations (153); however, the effect of 12,13-diHOME administration to humans is currently unknown. Adenosine is another BATokine that activates BAT and induces browning *in vivo* in mice and *in vitro* in humans, via activation of the A_{2A} receptor (154). Intravenous adenosine administration in lean healthy men *in vivo* increased BAT but not WAT perfusion in keeping with increased BAT oxidative metabolism (155). Further work is required to determine whether prolonged activation of A_{2A} receptors induces browning in humans *in vivo*. Recent work has focused on identifying BATokines in humans. For example, comparative analysis of the

human brown and white adipocyte secretome recently identified ~100 human BATokines (156), and through this ependymin-related protein 1 was identified as an important regulator of both thermogenic differentiation and noradrenaline-stimulated thermogenesis in human brown adipocytes (156). Further work investigating the role of the BAT secretome in humans is needed to determine if any BATokines offer a therapeutic potential.

Potential concerns with BAT activation as a therapeutic strategy

The metabolic benefits of BAT activation and cold exposure make increasing EE an attractive target for obesity and metabolic disease; however, there are concerns with this approach that must be taken into consideration. For example, selective activation of BAT may prove difficult to achieve or even be desired, as evidenced by selective β_3 -AR agonists that may in fact require activation of other β -receptor subtypes in BAT, and part of their beneficial effects may be mediated by other tissues (19, 21, 22, 94). Chronic sympathetic activation (e.g. from β_3 -AR agonists (19, 94), thyrotoxicosis (101) or from supraphysiological thyroid hormone replacement (157)) causing tachycardia and hypertension may result in unacceptable cardiovascular side effects such as myocardial infarction or stroke (158). It is also possible that elevated heart rate is essential for increased EE and additional research is required to identify whether there is a safe threshold of heart rate that does not increase cardiovascular risk (159). Further research to identify pharmacological mechanisms to safely activate BAT and/or wider cold-induced thermogenesis is urgently required.

Another potential issue is dissipation of the heat generated by pharmacological BAT activation. BAT activation prevents reduced body temperature during cold exposure, however, when activated chronically at room temperature or above thermoneutrality there is a risk of inducing hyperthermia. For example, 2,4-Dinitrophenol (DNP) was used as a weight loss medication as early as the 1930s (160). DNP caused generalised uncoupling of oxidative phosphorylation, leading to hyperthermia particularly during overdose that could be fatal (161, 162). Whilst selective BAT activation is unlikely to cause such thermal stress due to its low quantity, any pharmacological approach to activate BAT will have to be specific and avoid off target effects.

Finally, an approach to increase EE may cause a compensatory hyperphagic response, as observed in mice

and potentially humans during cold exposure, to meet the increased metabolic demands (12, 87, 163). However, not all pharmacotherapy that activates BAT in rodents causes hyperphagia (164), so it is unclear whether weight loss from increased EE by BAT may be neutralised by increased caloric consumption. Synergistic combination therapy with appetite suppressants may be required to maintain the benefits of BAT activation (165).

Perspective and conclusion

Since the identification of BAT in adult humans, there has been a resurgence in investigation of BAT activation and wider thermogenesis as a therapeutic strategy for obesity and metabolic disease. Thanks largely to PET imaging and to other novel *in vivo* techniques for measuring human BAT activity, significant progress has been made in understanding the role and regulation of human BAT, although to date most of the metabolic benefits from BAT activation have occurred in the context of acute cold exposure. In addition, pharmacological activation of BAT has been demonstrated at room temperature in important proof-of-concept studies but more research is required to fully understand the pathways regulating adipose tissue thermogenesis in order to develop treatments to safely activate BAT. Recent innovative approaches in rodents have identified the therapeutic potential of increasing BAT mass to treat metabolic disease, as evidenced by transplantation of either BAT (166) or beige adipocytes (167) which improves weight loss, glucose homeostasis and insulin sensitivity. While this approach remains untested in humans, it provides a clear proof of principle that increasing BAT mass improves metabolic health, which justifies further research to increase BAT mass and activity in humans. Pharmacological browning of white adipose tissue offers a larger adipose depot to increase EE and act as a glucose and lipid sink, although it remains unknown how much browning of these depots is possible. These current data suggest that selective BAT activation without significant expansion of BAT mass would not increase EE sufficiently to induce weight loss in humans and the most effective use of BAT activators may be as treatments for the comorbidities associated with obesity such as hyperglycaemia and dyslipidaemia rather than obesity itself. Further research into chronic BAT activation, potentially in combination with other approved weight loss therapies such as appetite suppressants, will determine whether BAT activation can complement current treatment options.

Declaration of interest

The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of this review.

Funding

This work was supported by the Medical Research Council (MR/K010271/1, MR/S035761/1), the Chief Scientist Office (SCAF/17/02), and the British Heart Foundation.

Author contribution statement

B T M and R H S wrote the manuscript and K J S critically revised the manuscript. All authors approved the submitted version.

References

- 1 Who.int. Obesity and overweight [Web Page]. Internet (www.who.int): World Health Organization, 2020 [updated 1 April 2020]. (available from: <https://www.who.int/news-room/fact-sheets/detail/obesity-and-overweight>)
- 2 NCD Risk Factor Collaboration (NCD-RisC). Worldwide trends in body-mass index, underweight, overweight, and obesity from 1975 to 2016: a pooled analysis of 2416 population-based measurement studies in 128.9 million children, adolescents, and adults. *Lancet* 2017 **390** 2627–2642. ([https://doi.org/10.1016/S0140-6736\(17\)32129-3](https://doi.org/10.1016/S0140-6736(17)32129-3))
- 3 Ng M, Fleming T, Robinson M, Thomson B, Graetz N, Margono C, Mullany EC, Biryukov S, Abbafati C, Abera SF et al. Global, regional, and national prevalence of overweight and obesity in children and adults during 1980–2013: a systematic analysis for the Global Burden of Disease Study 2013. *Lancet* 2014 **384** 766–781. ([https://doi.org/10.1016/S0140-6736\(14\)60460-8](https://doi.org/10.1016/S0140-6736(14)60460-8))
- 4 Bellou V, Belbasis L, Tzoulaki I & Evangelou E. Risk factors for type 2 diabetes mellitus: an exposure-wide umbrella review of meta-analyses. *PLoS ONE* 2018 **13** e0194127. (<https://doi.org/10.1371/journal.pone.0194127>)
- 5 Blüher M. Obesity: global epidemiology and pathogenesis. *Nature Reviews: Endocrinology* 2019 **15** 288–298. (<https://doi.org/10.1038/s41574-019-0176-8>)
- 6 Cataldi M, Cignarelli A, Giallauria F, Muscogiuri G, Barrea L, Savastano S, Colao A & Obesity Programs of Nutrition, Education, Research and Assessment (OPERA) Group. Cardiovascular effects of antiobesity drugs: are the new medicines all the same? *International Journal of Obesity Supplements* 2020 **10** 14–26. (<https://doi.org/10.1038/s41367-020-0015-3>)
- 7 Douglas IJ, Bhaskaran K, Batterham RL & Smeeth L. The effectiveness of pharmaceutical interventions for obesity: weight loss with orlistat and sibutramine in a United Kingdom population-based cohort. *British Journal of Clinical Pharmacology* 2015 **79** 1020–1027. (<https://doi.org/10.1111/bcp.12578>)
- 8 Sjostrom L, Lindroos AK, Peltonen M, Torgerson J, Bouchard C, Carlsson B, Dahlgren S, Larsson B, Narbro K, Sjostrom CD et al. Lifestyle, diabetes, and cardiovascular risk factors 10 years after bariatric surgery. *New England Journal of Medicine* 2004 **351** 2683–2693. (<https://doi.org/10.1056/NEJMoa035622>)
- 9 Hany TF, Gharehpapagh E, Kamel EM, Buck A, Himms-Hagen J & von Schulthess GK. Brown adipose tissue: a factor to consider in symmetrical tracer uptake in the neck and upper chest region. *European Journal of Nuclear Medicine and Molecular Imaging* 2002 **29** 1393–1398. (<https://doi.org/10.1007/s00259-002-0902-6>)
- 10 Yeung HWD, Grewal RK, Gonen M, Schöder H & Larson SM. Patterns of 18F-FDG uptake in adipose tissue and muscle: a potential source

- of false-positives for PET. *Journal of Nuclear Medicine* 2003 **44** 1789–1796.
- 11 Nedergaard J, Bengtsson T & Cannon B. Unexpected evidence for active brown adipose tissue in adult humans. *American Journal of Physiology: Endocrinology and Metabolism* 2007 **293** E444–E452. (<https://doi.org/10.1152/ajpendo.00691.2006>)
 - 12 Ravussin Y, Xiao C, Gavrillova O & Reitman ML. Effect of intermittent cold exposure on brown fat activation, obesity, and energy homeostasis in mice. *PLoS ONE* 2014 **9** e85876. (<https://doi.org/10.1371/journal.pone.0085876>)
 - 13 Bartelt A, Bruns OT, Reimer R, Hohenberg H, Ittrich H, Peldschus K, Kaul MG, Tromsdorf UI, Weller H, Waurisch C *et al.* Brown adipose tissue activity controls triglyceride clearance. *Nature Medicine* 2011 **17** 200–205. (<https://doi.org/10.1038/nm.2297>)
 - 14 Nedergaard J, Matthias A, Golozoubova V, Jacobsson A & Cannon B. UCP1: the original uncoupling protein – and perhaps the only one? New perspectives on UCP1, UCP2, and UCP3 in the light of the bioenergetics of the UCP1-ablated mice. *Journal of Bioenergetics and Biomembranes* 1999 **31** 475–491. (<https://doi.org/10.1023/a:1005400507802>)
 - 15 Leitner BP, Huang S, Brychta RJ, Duckworth CJ, Baskin AS, McGehee S, Tal I, Dieckmann W, Gupta G, Kolodny GM *et al.* Mapping of human brown adipose tissue in lean and obese young men. *PNAS* 2017 **114** 8649–8654. (<https://doi.org/10.1073/pnas.1705287114>)
 - 16 Jespersen NZ, Feizi A, Andersen ES, Heywood S, Hattel HB, Daugaard S, Peijs L, Bagi P, Feldt-Rasmussen B, Schultz HS *et al.* Heterogeneity in the perirenal region of humans suggests presence of dormant brown adipose tissue that contains brown fat precursor cells. *Molecular Metabolism* 2019 **24** 30–43. (<https://doi.org/10.1016/j.molmet.2019.03.005>)
 - 17 van Marken Lichtenbelt WD, Vanhomerig JW, Smulders NM, Drossaerts JM, Kemerink GJ, Bouvy ND, Schrauwen P & Teule GJ. Cold-activated brown adipose tissue in healthy men. *New England Journal of Medicine* 2009 **360** 1500–1508. (<https://doi.org/10.1056/NEJMoa0808718>)
 - 18 Porter C, Herndon DN, Chondronikola M, Chao T, Annamalai P, Bhattarai N, Saraf MK, Capek KD, Reidy PT, Daquinag AC *et al.* Human and mouse brown adipose tissue mitochondria have comparable UCP1 function. *Cell Metabolism* 2016 **24** 246–255. (<https://doi.org/10.1016/j.cmet.2016.07.004>)
 - 19 Cypess AM, Weiner LS, Roberts-Toler C, Franquet Elia E, Kessler SH, Kahn PA, English J, Chatman K, Trauger SA, Doria A *et al.* Activation of human brown adipose tissue by a beta3-adrenergic receptor agonist. *Cell Metabolism* 2015 **21** 33–38. (<https://doi.org/10.1016/j.cmet.2014.12.009>)
 - 20 Cypess AM, White AP, Vernochet C, Schulz TJ, Xue R, Sass CA, Huang TL, Roberts-Toler C, Weiner LS, Sze C *et al.* Anatomical localization, gene expression profiling and functional characterization of adult human neck brown fat. *Nature Medicine* 2013 **19** 635–639. (<https://doi.org/10.1038/nm.3112>)
 - 21 Riis-Vestergaard MJ, Richelsen B, Bruun JM, Li W, Hansen JB & Pedersen SB. Beta-1 and not beta-3 adrenergic receptors may be the primary regulator of human brown adipocyte metabolism. *Journal of Clinical Endocrinology and Metabolism* 2020 **105** e994–e1005. (<https://doi.org/10.1210/clinem/dgz298>)
 - 22 Blondin DP, Nielsen S, Kuipers EN, Severinsen MC, Jensen VH, Miard S, Jespersen NZ, Kooijman S, Boon MR, Fortin M *et al.* Human brown adipocyte thermogenesis is driven by beta2-AR stimulation. *Cell Metabolism* 2020 **32** 287.e7–300.e7. (<https://doi.org/10.1016/j.cmet.2020.07.005>)
 - 23 McNeill BT, Morton NM & Stimson RH. Substrate utilization by brown adipose tissue: what's hot and what's not? *Frontiers in Endocrinology* 2020 **11** 571659. (<https://doi.org/10.3389/fendo.2020.571659>)
 - 24 Seale P, Bjork B, Yang W, Kajimura S, Chin S, Kuang S, Scime A, Devarakonda S, Conroe HM, Erdjument-Bromage H *et al.* PRDM16 controls a brown fat/skeletal muscle switch. *Nature* 2008 **454** 961–967. (<https://doi.org/10.1038/nature07182>)
 - 25 Ikeda K, Maretich P & Kajimura S. The common and distinct features of brown and beige adipocytes. *Trends in Endocrinology and Metabolism* 2018 **29** 191–200. (<https://doi.org/10.1016/j.tem.2018.01.001>)
 - 26 Wu J, Bostrom P, Sparks LM, Ye L, Choi JH, Giang AH, Khandekar M, Virtanen KA, Nuutila P, Schaart G *et al.* Beige adipocytes are a distinct type of thermogenic fat cell in mouse and human. *Cell* 2012 **150** 366–376. (<https://doi.org/10.1016/j.cell.2012.05.016>)
 - 27 Shabalina IG, Petrovic N, de Jong JM, Kalinovich AV, Cannon B & Nedergaard J. UCP1 in brite/beige adipose tissue mitochondria is functionally thermogenic. *Cell Reports* 2013 **5** 1196–1203. (<https://doi.org/10.1016/j.celrep.2013.10.044>)
 - 28 Kazak L, Chouchani ET, Jedrychowski MP, Erickson BK, Shinoda K, Cohen P, Vetrivelan R, Lu GZ, Laznik-Bogoslavski D, Hasenfuss SC *et al.* A creatine-driven substrate cycle enhances energy expenditure and thermogenesis in beige fat. *Cell* 2015 **163** 643–655. (<https://doi.org/10.1016/j.cell.2015.09.035>)
 - 29 Jespersen NZ, Larsen TJ, Peijs L, Daugaard S, Homoe P, Loft A, de Jong J, Mathur N, Cannon B, Nedergaard J *et al.* A classical brown adipose tissue mRNA signature partly overlaps with brite in the supraclavicular region of adult humans. *Cell Metabolism* 2013 **17** 798–805. (<https://doi.org/10.1016/j.cmet.2013.04.011>)
 - 30 Virtanen KA, Lidell ME, Orava J, Heglind M, Westergren R, Niemi T, Taittonen M, Laine J, Savisto NJ, Enerback S *et al.* Functional brown adipose tissue in healthy adults. *New England Journal of Medicine* 2009 **360** 1518–1525. (<https://doi.org/10.1056/NEJMoa0808949>)
 - 31 Cypess AM, Lehman S, Williams G, Tal I, Rodman D, Goldfine AB, Kuo FC, Palmer EL, Tseng YH, Doria A *et al.* Identification and importance of brown adipose tissue in adult humans. *New England Journal of Medicine* 2009 **360** 1509–1517. (<https://doi.org/10.1056/NEJMoa0810780>)
 - 32 Chen KY, Cypess AM, Laughlin MR, Haft CR, Hu HH, Bredella MA, Enerback S, Kinahan PE, Lichtenbelt Wv, Lin FI *et al.* Brown adipose reporting criteria in imaging studies (BARCIST 1.0): recommendations for standardized FDG-PET/CT experiments in humans. *Cell Metabolism* 2016 **24** 210–222. (<https://doi.org/10.1016/j.cmet.2016.07.014>)
 - 33 Weir G, Ramage LE, Akyol M, Rhodes JK, Kyle CJ, Fletcher AM, Craven TH, Wakelin SJ, Drake AJ, Gregoriades ML *et al.* Substantial metabolic activity of human brown adipose tissue during warm conditions and cold-induced lipolysis of local triglycerides. *Cell Metabolism* 2018 **27** 1348.e4–1355.e4. (<https://doi.org/10.1016/j.cmet.2018.04.020>)
 - 34 Saari TJ, Raiko J, U-Din M, Niemi T, Taittonen M, Laine J, Savisto N, Haaoranta-Solin M, Nuutila P & Virtanen KA. Basal and cold-induced fatty acid uptake of human brown adipose tissue is impaired in obesity. *Scientific Reports* 2020 **10** 14373. (<https://doi.org/10.1038/s41598-020-71197-2>)
 - 35 Worku MG, Seretew WS, Angaw DA & Tesema GA. Prevalence and associated factor of brown adipose tissue: systematic review and meta-analysis. *BioMed Research International* 2020 **2020** 9106976. (<https://doi.org/10.1155/2020/9106976>)
 - 36 He X, Li Z, Tang X, Zhang L, Wang L, He Y, Jin T & Yuan D. Age- and sex-related differences in body composition in healthy subjects aged 18 to 82 years. *Medicine* 2018 **97** e11152. (<https://doi.org/10.1097/MD.00000000000011152>)
 - 37 Bazzocchi A, Diano D, Ponti F, Andreone A, Sassi C, Albisinni U, Marchesini G & Battista G. Health and ageing: a cross-sectional study of body composition. *Clinical Nutrition* 2013 **32** 569–578. (<https://doi.org/10.1016/j.clnu.2012.10.004>)
 - 38 van der Lans AA, Hoeks J, Brans B, Vijgen GH, Visser MG, Vosselman MJ, Hansen J, Jorgensen JA, Wu J, Mottaghy FM *et al.* Cold acclimation recruits human brown fat and increases

- nonshivering thermogenesis. *Journal of Clinical Investigation* 2013 **123** 3395–3403. (<https://doi.org/10.1172/JCI68993>)
- 39 Lee P, Smith S, Linderman J, Courville AB, Brychta RJ, Dieckmann W, Werner CD, Chen KY & Celi FS. Temperature-acclimated brown adipose tissue modulates insulin sensitivity in humans. *Diabetes* 2014 **63** 3686–3698. (<https://doi.org/10.2337/db14-0513>)
- 40 Persichetti A, Sciuto R, Rea S, Basciani S, Lubrano C, Mariani S, Ulisse S, Nofroni I, Maini CL & Gnessi L. Prevalence, mass, and glucose-uptake activity of ¹⁸F-FDG-detected brown adipose tissue in humans living in a temperate zone of Italy. *PLoS ONE* 2013 **8** e63391. (<https://doi.org/10.1371/journal.pone.0063391>)
- 41 Ouellet V, Labbe SM, Blondin DP, Phoenix S, Guerin B, Haman F, Turcotte EE, Richard D & Carpentier AC. Brown adipose tissue oxidative metabolism contributes to energy expenditure during acute cold exposure in humans. *Journal of Clinical Investigation* 2012 **122** 545–552. (<https://doi.org/10.1172/JCI60433>)
- 42 Ouellet V, Routhier-Labadie A, Bellemare W, Lakhal-Chaieb L, Turcotte E, Carpentier AC & Richard D. Outdoor temperature, age, sex, body mass index, and diabetic status determine the prevalence, mass, and glucose-uptake activity of ¹⁸F-FDG-detected BAT in humans. *Journal of Clinical Endocrinology and Metabolism* 2011 **96** 192–199. (<https://doi.org/10.1210/jc.2010-0989>)
- 43 Mengel LA, Seidl H, Brandl B, Skurk T, Holzapfel C, Stecher L, Clausnitzer M & Hauner H. Gender differences in the response to short-term cold exposure in young adults. *Journal of Clinical Endocrinology and Metabolism* 2020 **105** e1938–e1948. (<https://doi.org/10.1210/clinem/daaa110>)
- 44 Bakker LE, Boon MR, van der Linden RA, Arias-Bouda LP, van Klinken JB, Smit F, Verberne HJ, Jukema JW, Tamsma JT, Havekes LM *et al.* Brown adipose tissue volume in healthy lean South Asian adults compared with white Caucasians: a prospective, case-controlled observational study. *Lancet: Diabetes and Endocrinology* 2014 **2** 210–217. ([https://doi.org/10.1016/S2213-8587\(13\)70156-6](https://doi.org/10.1016/S2213-8587(13)70156-6))
- 45 Saito M, Okamatsu-Ogura Y, Matsushita M, Watanabe K, Yoneshiro T, Nio-Kobayashi J, Iwanaga T, Miyagawa M, Kameya T, Nakada K *et al.* High incidence of metabolically active brown adipose tissue in healthy adult humans: effects of cold exposure and adiposity. *Diabetes* 2009 **58** 1526–1531. (<https://doi.org/10.2337/db09-0530>)
- 46 Orava J, Nuutila P, Noponen T, Parkkola R, Viljanen T, Enerback S, Rissanen A, Pietilainen KH & Virtanen KA. Blunted metabolic responses to cold and insulin stimulation in brown adipose tissue of obese humans. *Obesity* 2013 **21** 2279–2287. (<https://doi.org/10.1002/oby.20456>)
- 47 Feldmann HM, Golozoubova V, Cannon B & Nedergaard J. UCP1 ablation induces obesity and abolishes diet-induced thermogenesis in mice exempt from thermal stress by living at thermoneutrality. *Cell Metabolism* 2009 **9** 203–209. (<https://doi.org/10.1016/j.cmet.2008.12.014>)
- 48 Jaroslawska J, Chabowska-Kita A, Kaczmarek MM & Kozak LP. Npvh: hypothalamic biomarker of ambient temperature independent of nutritional status. *PLoS Genetics* 2015 **11** e1005287. (<https://doi.org/10.1371/journal.pgen.1005287>)
- 49 Blondin DP, Labbe SM, Noll C, Kunach M, Phoenix S, Guerin B, Turcotte EE, Haman F, Richard D & Carpentier AC. Selective impairment of glucose but not fatty acid or oxidative metabolism in brown adipose tissue of subjects with type 2 diabetes. *Diabetes* 2015 **64** 2388–2397. (<https://doi.org/10.2337/db14-1651>)
- 50 Hanssen MJ, Hoeks J, Brans B, van der Lans AA, Schaart G, van den Driessche JJ, Jorgensen JA, Boekschoten MV, Hesselink MK, Havekes B *et al.* Short-term cold acclimation improves insulin sensitivity in patients with type 2 diabetes mellitus. *Nature Medicine* 2015 **21** 863–865. (<https://doi.org/10.1038/nm.3891>)
- 51 Huttunen P, Hirvonen J & Kinnula V. The occurrence of brown adipose tissue in outdoor workers. *European Journal of Applied Physiology and Occupational Physiology* 1981 **46** 339–345. (<https://doi.org/10.1007/BF00422121>)
- 52 Blondin DP, Labbe SM, Tingelstad HC, Noll C, Kunach M, Phoenix S, Guerin B, Turcotte EE, Carpentier AC, Richard D *et al.* Increased brown adipose tissue oxidative capacity in cold-acclimated humans. *Journal of Clinical Endocrinology and Metabolism* 2014 **99** E438–E446. (<https://doi.org/10.1210/jc.2013-3901>)
- 53 Geslot A, Bennet A, Hitzel A, Thoulouzan M, Mouly C, Savagner F, Quintyn-Ranty ML, Caron P & Vezzosi D. Weight-loss with activation of brown fat: suspect pheochromocytoma. *Annales d'Endocrinologie* 2019 **80** 314–318. (<https://doi.org/10.1016/j.ando.2019.06.004>)
- 54 Kuji I, Imabayashi E, Minagawa A, Matsuda H & Miyauchi T. Brown adipose tissue demonstrating intense FDG uptake in a patient with mediastinal pheochromocytoma. *Annals of Nuclear Medicine* 2008 **22** 231–235. (<https://doi.org/10.1007/s12149-007-0096-x>)
- 55 Dadson P, Hannukainen JC, Din MU, Lahesmaa M, Kalliokoski KK, Iozzo P, Pihlajamaki J, Karlsson HK, Parkkola R, Salminen P *et al.* Brown adipose tissue lipid metabolism in morbid obesity: effect of bariatric surgery-induced weight loss. *Diabetes, Obesity and Metabolism* 2018 **20** 1280–1288. (<https://doi.org/10.1111/dom.13233>)
- 56 Vijgen GH, Bouvy ND, Teule GJ, Brans B, Hoeks J, Schrauwen P & van Marken Lichtenbelt WD. Increase in brown adipose tissue activity after weight loss in morbidly obese subjects. *Journal of Clinical Endocrinology and Metabolism* 2012 **97** E1229–E1233. (<https://doi.org/10.1210/jc.2012-1289>)
- 57 van den Berg R, Kooijman S, Noordam R, Ramkisoensing A, Abreu-Vieira G, Tambyrajah LL, Dijk W, Ruppert P, Mol IM, Kramar B *et al.* A diurnal rhythm in brown adipose tissue causes rapid clearance and combustion of plasma lipids at waking. *Cell Reports* 2018 **22** 3521–3533. (<https://doi.org/10.1016/j.celrep.2018.03.004>)
- 58 Lee P, Bova R, Schofield L, Bryant W, Dieckmann W, Slattery A, Govendir MA, Emmett L & Greenfield JR. Brown adipose tissue exhibits a glucose-responsive thermogenic biorhythm in humans. *Cell Metabolism* 2016 **23** 602–609. (<https://doi.org/10.1016/j.cmet.2016.02.007>)
- 59 de Goede P, Sen S, Oosterman JE, Foppen E, Jansen R, la Fleur SE, Challet E & Kalsbeek A. Differential effects of diet composition and timing of feeding behavior on rat brown adipose tissue and skeletal muscle peripheral clocks. *Neurobiology of Sleep and Circadian Rhythms* 2018 **4** 24–33. (<https://doi.org/10.1016/j.nbscr.2017.09.002>)
- 60 Kim KH, Kim YH, Son JE, Lee JH, Kim S, Choe MS, Moon JH, Zhong J, Fu K, Lenglin F *et al.* Intermittent fasting promotes adipose thermogenesis and metabolic homeostasis via VEGF-mediated alternative activation of macrophage. *Cell Research* 2017 **27** 1309–1326. (<https://doi.org/10.1038/cr.2017.126>)
- 61 Li G, Xie C, Lu S, Nichols RG, Tian Y, Li L, Patel D, Ma Y, Brocker CN, Yan T *et al.* Intermittent fasting promotes white adipose browning and decreases obesity by shaping the gut microbiota. *Cell Metabolism* 2017 **26** 672.e4–685.e4. (<https://doi.org/10.1016/j.cmet.2017.08.019>)
- 62 Singhal V, Maffazioli GD, Ackerman KE, Lee H, Elia EF, Woolley R, Kolodny G, Cypess AM & Misra M. Effect of chronic athletic activity on brown fat in young women. *PLoS ONE* 2016 **11** e0156353. (<https://doi.org/10.1371/journal.pone.0156353>)
- 63 Vosselman MJ, Hoeks J, Brans B, Pallubinsky H, Nascimento EBM, van der Lans AAJJ, Broeders EPM, Mottaghy FM, Schrauwen P & van Marken Lichtenbelt WD. Low brown adipose tissue activity in endurance-trained compared with lean sedentary men. *International Journal of Obesity* 2015 **39** 1696–1702. (<https://doi.org/10.1038/ijo.2015.130>)
- 64 Garrow JS. Luxuskonsumption, brown fat, and human obesity. *BMJ* 1983 **286** 1684–1686. (<https://doi.org/10.1136/bmj.286.6379.1684-a>)
- 65 Yoneshiro T, Aita S, Matsushita M, Kameya T, Nakada K, Kawai Y & Saito M. Brown adipose tissue, whole-body energy expenditure, and thermogenesis in healthy adult men. *Obesity* 2011 **19** 13–16. (<https://doi.org/10.1038/oby.2010.105>)
- 66 Yoneshiro T, Aita S, Matsushita M, Kayahara T, Kameya T, Kawai Y, Iwanaga T & Saito M. Recruited brown adipose tissue as an

- antiobesity agent in humans. *Journal of Clinical Investigation* 2013 **123** 3404–3408. (<https://doi.org/10.1172/JCI67803>)
- 67 U Din M, Raiko J, Saari T, Kudomi N, Tolvanen T, Oikonen V, Teuho J, Sipilä HT, Savisto N, Parkkola R *et al*. Human brown adipose tissue [(15)O]O₂ PET imaging in the presence and absence of cold stimulus. *European Journal of Nuclear Medicine and Molecular Imaging* 2016 **43** 1878–1886. (<https://doi.org/10.1007/s00259-016-3364-y>)
- 68 Muzik O, Mangner TJ, Leonard WR, Kumar A, Janisse J & Granneman JG. 15O PET measurement of blood flow and oxygen consumption in cold-activated human brown fat. *Journal of Nuclear Medicine* 2013 **54** 523–531. (<https://doi.org/10.2967/jnumed.112.111336>)
- 69 Blondin DP, Labbe SM, Phoenix S, Guerin B, Turcotte ÉE, Richard D, Carpentier AC & Haman F. Contributions of white and brown adipose tissues and skeletal muscles to acute cold-induced metabolic responses in healthy men. *Journal of Physiology* 2015 **593** 701–714. (<https://doi.org/10.1113/jphysiol.2014.283598>)
- 70 Kong X, Yao T, Zhou P, Kazak L, Tenen D, Lyubetskaya A, Dawes BA, Tsai L, Kahn BB, Spiegelman BM *et al*. Brown adipose tissue controls skeletal muscle function via the secretion of myostatin. *Cell Metabolism* 2018 **28** 631.e3–643.e3. (<https://doi.org/10.1016/j.cmet.2018.07.004>)
- 71 Saeidifard F, Medina-Inojosa JR, Supervia M, Olson TP, Somers VK, Erwin PJ & Lopez-Jimenez F. Differences of energy expenditure while sitting versus standing: a systematic review and meta-analysis. *European Journal of Preventive Cardiology* 2018 **25** 522–538. (<https://doi.org/10.1177/2047487317752186>)
- 72 Blondin DP, Frisch F, Phoenix S, Guerin B, Turcotte ÉE, Haman F, Richard D & Carpentier AC. Inhibition of intracellular triglyceride lipolysis suppresses cold-induced brown adipose tissue metabolism and increases shivering in humans. *Cell Metabolism* 2017 **25** 438–447. (<https://doi.org/10.1016/j.cmet.2016.12.005>)
- 73 U Din M, Saari T, Raiko J, Kudomi N, Maurer SF, Lahesmaa M, Fromme T, Amri EZ, Klingenspor M, Solin O *et al*. Postprandial oxidative metabolism of human brown fat indicates thermogenesis. *Cell Metabolism* 2018 **28** 207.e3–216.e3. (<https://doi.org/10.1016/j.cmet.2018.05.020>)
- 74 Chondronikola M, Volpi E, Borsheim E, Porter C, Annamalai P, Enerback S, Lidell ME, Saraf MK, Labbe SM, Hurren NM *et al*. Brown adipose tissue improves whole-body glucose homeostasis and insulin sensitivity in humans. *Diabetes* 2014 **63** 4089–4099. (<https://doi.org/10.2337/db14-0746>)
- 75 Chondronikola M, Volpi E, Borsheim E, Porter C, Saraf MK, Annamalai P, Yfanti C, Chao T, Wong D, Shinoda K *et al*. Brown adipose tissue activation is linked to distinct systemic effects on lipid metabolism in humans. *Cell Metabolism* 2016 **23** 1200–1206. (<https://doi.org/10.1016/j.cmet.2016.04.029>)
- 76 Blondin DP, Tingelstad HC, Noll C, Frisch F, Phoenix S, Guerin B, Turcotte ÉE, Richard D, Haman F & Carpentier AC. Dietary fatty acid metabolism of brown adipose tissue in cold-acclimated men. *Nature Communications* 2017 **8** 14146. (<https://doi.org/10.1038/ncomms14146>)
- 77 Becker M, Serr I, Salb VK, Ott VB, Mengel L, Blüher M, Weigmann B, Hauner H, Tschöp MH & Daniel C. Short-term cold exposure supports human Treg induction in vivo. *Molecular Metabolism* 2019 **28** 73–82. (<https://doi.org/10.1016/j.molmet.2019.08.002>)
- 78 Wang QA, Tao C, Gupta RK & Scherer PE. Tracking adipogenesis during white adipose tissue development, expansion and regeneration. *Nature Medicine* 2013 **19** 1338–1344. (<https://doi.org/10.1038/nm.3324>)
- 79 Søberg S. Winter-swimming and brown adipose tissue activity in middle-aged obese subjects (WinterBAT). NCT03541096 <https://clinicaltrials.gov>. United States National Library of Medicine, 2018. (available at: <https://clinicaltrials.gov/ct2/show/NCT03541096>)
- 80 (NIDDK) NIOHCCCNIODaDaKD. Chronic cold exposure and energy metabolism in humans <https://clinicaltrials.gov>. United States National Library of Medicine, 2018 [updated 14.05.2018]. (available at: <https://clinicaltrials.gov/ct2/show/NCT01730105>)
- 81 Yang HK, Han K, Cho JH, Yoon KH, Cha BY & Lee SH. Ambient temperature and prevalence of obesity: a nationwide population-based study in Korea. *PLoS ONE* 2015 **10** e0141724. (<https://doi.org/10.1371/journal.pone.0141724>)
- 82 Valdes S, Maldonado-Araque C, Garcia-Torres F, Goday A, Bosch-Comas A, Bordiu E, Calle-Pascual A, Carmena R, Casamitjana R, Castano L *et al*. Ambient temperature and prevalence of obesity in the Spanish population: the Di@bet.es study. *Obesity* 2014 **22** 2328–2332. (<https://doi.org/10.1002/oby.20866>)
- 83 Daly M. Association of ambient indoor temperature with body mass index in England. *Obesity* 2014 **22** 626–629. (<https://doi.org/10.1002/oby.20546>)
- 84 Hunt DRG & Gidman MI. A national field survey of house temperatures. *Building and Environment* 1982 **17** 107–124. ([https://doi.org/10.1016/0360-1323\(82\)90048-8](https://doi.org/10.1016/0360-1323(82)90048-8))
- 85 Shipworth M, Firth SK, Gentry MI, Wright AJ, Shipworth DT & Lomas KJ. Central heating thermostat settings and timing: building demographics. *Building Research and Information* 2010 **38** 50–69. (<https://doi.org/10.1080/09613210903263007>)
- 86 Mavrogianni A, Johnson F, Ucci M, Marmot A, Wardle J, Oreszczyn T & Summerfield A. Historic variations in winter indoor domestic temperatures and potential implications for body weight gain. *Indoor and Built Environment* 2013 **22** 360–375. (<https://doi.org/10.1177/1420326X11425966>)
- 87 Westerterp-Plantenga MS, van Marken Lichtenbelt WD, Strobbe H & Schrauwen P. Energy metabolism in humans at a lowered ambient temperature. *European Journal of Clinical Nutrition* 2002 **56** 288–296. (<https://doi.org/10.1038/sj.ejcn.1601308>)
- 88 Perkins MN, Rothwell NJ, Stock MJ & Stone TW. Activation of brown adipose tissue thermogenesis by the ventromedial hypothalamus. *Nature* 1981 **289** 401–402. (<https://doi.org/10.1038/289401a0>)
- 89 Arbeeny CM, Meyers DS, Hillyer DE & Bergquist KE. Metabolic alterations associated with the antidiabetic effect of beta 3-adrenergic receptor agonists in obese mice. *American Journal of Physiology* 1995 **268** E678–E684. (<https://doi.org/10.1152/ajpendo.1995.268.4.E678>)
- 90 Cawthorne MA, Sennitt MV, Arch JR & Smith SA. BRL 35135, a potent and selective atypical beta-adrenoceptor agonist. *American Journal of Clinical Nutrition* 1992 **55** (Supplement) 252S–257S. (<https://doi.org/10.1093/ajcn/55.1.252s>)
- 91 Mitchell TH, Ellis RD, Smith SA, Robb G & Cawthorne MA. Effects of BRL 35135, a beta-adrenoceptor agonist with novel selectivity, on glucose tolerance and insulin sensitivity in obese subjects. *International Journal of Obesity* 1989 **13** 757–766.
- 92 Larsen TM, Toubro S, van Baak MA, Gottesdiener KM, Larson P, Saris WH & Astrup A. Effect of a 28-d treatment with L-796568, a novel beta(3)-adrenergic receptor agonist, on energy expenditure and body composition in obese men. *American Journal of Clinical Nutrition* 2002 **76** 780–788. (<https://doi.org/10.1093/ajcn/76.4.780>)
- 93 Weyer C, Tataranni PA, Snitker S, Danforth E, Jr & Ravussin E. Increase in insulin action and fat oxidation after treatment with CL 316,243, a highly selective beta3-adrenoceptor agonist in humans. *Diabetes* 1998 **47** 1555–1561. (<https://doi.org/10.2337/diabetes.47.10.1555>)
- 94 O'Mara AE, Johnson JW, Linderman JD, Brychta RJ, McGehee S, Fletcher LA, Fink YA, Kapuria D, Cassimatis TM, Kelsey N *et al*. Chronic mirabegron treatment increases human brown fat, HDL cholesterol, and insulin sensitivity. *Journal of Clinical Investigation* 2020 **130** 2209–2219. (<https://doi.org/10.1172/JCI131126>)
- 95 Dulloo AG, Seydoux J & Girardier L. Peripheral mechanisms of thermogenesis induced by ephedrine and caffeine in brown adipose tissue. *International Journal of Obesity* 1991 **15** 317–326.
- 96 Becker DE. Basic and clinical pharmacology of autonomic drugs. *Anesthesia Progress* 2012 **59** 159–168; quiz 169. (<https://doi.org/10.2344/0003-3006-59.4.159>)

- 97 Carey AL, Formosa MF, Van Every B, Bertovic D, Eikelis N, Lambert GW, Kalff V, Duffy SJ, Cherk MH & Kingwell BA. Ephedrine activates brown adipose tissue in lean but not obese humans. *Diabetologia* 2013 **56** 147–155. (<https://doi.org/10.1007/s00125-012-2748-1>)
- 98 Cypess AM, Chen YC, Sze C, Wang K, English J, Chan O, Holman AR, Tal I, Palmer MR, Kolodny GM *et al.* Cold but not sympathomimetics activates human brown adipose tissue in vivo. *PNAS* 2012 **109** 10001–10005. (<https://doi.org/10.1073/pnas.1207911109>)
- 99 Carey AL, Pajtak R, Formosa MF, Van Every B, Bertovic DA, Anderson MJ, Eikelis N, Lambert GW, Kalff V, Duffy SJ *et al.* Chronic ephedrine administration decreases brown adipose tissue activity in a randomised controlled human trial: implications for obesity. *Diabetologia* 2015 **58** 1045–1054. (<https://doi.org/10.1007/s00125-015-3543-6>)
- 100 Ribeiro MO, Carvalho SD, Schultz JJ, Chiellini G, Scanlan TS, Bianco AC & Brent GA. Thyroid hormone – sympathetic interaction and adaptive thermogenesis are thyroid hormone receptor isoform – specific. *Journal of Clinical Investigation* 2001 **108** 97–105. (<https://doi.org/10.1172/JCI12584>)
- 101 Lahesmaa M, Orava J, Schalin-Jantti C, Soinio M, Hannukainen JC, Noponen T, Kirjavainen A, Iida H, Kudomi N, Enerback S *et al.* Hyperthyroidism increases brown fat metabolism in humans. *Journal of Clinical Endocrinology and Metabolism* 2014 **99** E28–E35. (<https://doi.org/10.1210/jc.2013-2312>)
- 102 Skarulis MC, Celi FS, Mueller E, Zemska M, Malek R, Hugendubler L, Cochran C, Solomon J, Chen C & Gorden P. Thyroid hormone induced brown adipose tissue and amelioration of diabetes in a patient with extreme insulin resistance. *Journal of Clinical Endocrinology and Metabolism* 2010 **95** 256–262. (<https://doi.org/10.1210/jc.2009-0543>)
- 103 Heinen CA, Zhang Z, Kliverik LP, de Wit TC, Poel E, Yaqub M, Boelen A, Kalsbeek A, Bisschop PH, van Trotsenburg ASP *et al.* Effects of intravenous thyrotropin-releasing hormone on (18) F-fluorodeoxyglucose uptake in human brown adipose tissue: a randomized controlled trial. *European Journal of Endocrinology* 2018 **179** 31–38. (<https://doi.org/10.1530/EJE-17-0966>)
- 104 Ono K, Tsukamoto-Yasui M, Hara-Kimura Y, Inoue N, Nogusa Y, Okabe Y, Nagashima K & Kato F. Intragastric administration of capsiate, a transient receptor potential channel agonist, triggers thermogenic sympathetic responses. *Journal of Applied Physiology* 2011 **110** 789–798. (<https://doi.org/10.1152/jappphysiol.00128.2010>)
- 105 Masuda Y, Haramizu S, Oki K, Ohnuki K, Watanabe T, Yazawa S, Kawada T, Hashizume S & Fushiki T. Upregulation of uncoupling proteins by oral administration of capsiate, a nonpungent capsaicin analog. *Journal of Applied Physiology* 2003 **95** 2408–2415. (<https://doi.org/10.1152/jappphysiol.00828.2002>)
- 106 Yoneshiro T, Aita S, Kawai Y, Iwanaga T & Saito M. Nonpungent capsaicin analogs (capsinoids) increase energy expenditure through the activation of brown adipose tissue in humans. *American Journal of Clinical Nutrition* 2012 **95** 845–850. (<https://doi.org/10.3945/ajcn.111.018606>)
- 107 Fuse S, Endo T, Tanaka R, Kuroiwa M, Ando A, Kume A, Yamamoto A, Kuribayashi K, Somekawa S, Takeshita M *et al.* Effects of capsinoid intake on brown adipose tissue vascular density and resting energy expenditure in healthy, middle-aged adults: a randomized, double-blind, placebo-controlled study. *Nutrients* 2020 **12** 2676. (<https://doi.org/10.3390/nu12092676>)
- 108 Lefebvre P, Cariou B, Lien F, Kuipers F & Staels B. Role of bile acids and bile acid receptors in metabolic regulation. *Physiological Reviews* 2009 **89** 147–191. (<https://doi.org/10.1152/physrev.00010.2008>)
- 109 Fang S, Suh JM, Reilly SM, Yu E, Osborn O, Lackey D, Yoshihara E, Perino A, Jacinto S, Lukasheva Y *et al.* Intestinal FXR agonism promotes adipose tissue browning and reduces obesity and insulin resistance. *Nature Medicine* 2015 **21** 159–165. (<https://doi.org/10.1038/nm.3760>)
- 110 Velazquez-Villegas LA, Perino A, Lemos V, Zietak M, Nomura M, Pols TWH & Schoonjans K. TGR5 signalling promotes mitochondrial fission and beige remodelling of white adipose tissue. *Nature Communications* 2018 **9** 245. (<https://doi.org/10.1038/s41467-017-02068-0>)
- 111 Watanabe M, Houten SM, Matakai C, Christoffolete MA, Kim BW, Sato H, Messaddeq N, Harney JW, Ezaki O, Kodama T *et al.* Bile acids induce energy expenditure by promoting intracellular thyroid hormone activation. *Nature* 2006 **439** 484–489. (<https://doi.org/10.1038/nature04330>)
- 112 Broeders EP, Nascimento EB, Havekes B, Brans B, Roumans KH, Tailleux A, Schaart G, Kouach M, Charton J, Deprez B *et al.* The bile acid chenodeoxycholic acid increases human brown adipose tissue activity. *Cell Metabolism* 2015 **22** 418–426. (<https://doi.org/10.1016/j.cmet.2015.07.002>)
- 113 Scotney H, Symonds ME, Law J, Budge H, Sharkey D & Manolopoulos KN. Glucocorticoids modulate human brown adipose tissue thermogenesis in vivo. *Metabolism: Clinical and Experimental* 2017 **70** 125–132. (<https://doi.org/10.1016/j.metabol.2017.01.024>)
- 114 Ramage LE, Akyol M, Fletcher AM, Forsythe J, Nixon M, Carter RN, van Beek EJ, Morton NM, Walker BR & Stimson RH. Glucocorticoids acutely increase brown adipose tissue activity in humans, revealing species-specific differences in UCP-1 regulation. *Cell Metabolism* 2016 **24** 130–141. (<https://doi.org/10.1016/j.cmet.2016.06.011>)
- 115 Strack AM, Bradbury MJ & Dallman MF. Corticosterone decreases nonshivering thermogenesis and increases lipid storage in brown adipose tissue. *American Journal of Physiology* 1995 **268** R183–R191. (<https://doi.org/10.1152/ajpregu.1995.268.1.R183>)
- 116 Thuzar M, Law WP, Ratnasingam J, Jang C, Dimeski G & Ho KKY. Glucocorticoids suppress brown adipose tissue function in humans: a double-blind placebo-controlled study. *Diabetes, Obesity and Metabolism* 2018 **20** 840–848. (<https://doi.org/10.1111/dom.13157>)
- 117 Zhang Q, Zhou S, Yan X, Nan L & Yang L. Optimum body fat percentage cut-off in evaluation of overweight and obesity among adult people. *Wei Sheng Yan Jiu* 2019 **48** 573–576.
- 118 Hanssen MJ, van der Lans AA, Brans B, Hoeks J, Jardon KM, Schaart G, Mottaghy FM, Schrauwen P & van Marken Lichtenbelt WD. Short-term cold acclimation recruits brown adipose tissue in obese humans. *Diabetes* 2016 **65** 1179–1189. (<https://doi.org/10.2337/db15-1372>)
- 119 Okamatsu-Ogura Y, Fukano K, Tsubota A, Uozumi A, Terao A, Kimura K & Saito M. Thermogenic ability of uncoupling protein 1 in beige adipocytes in mice. *PLoS ONE* 2013 **8** e84229. (<https://doi.org/10.1371/journal.pone.0084229>)
- 120 Finlin BS, Memetimin H, Confides AL, Kasza I, Zhu B, Vekaria HJ, Harfmann B, Jones KA, Johnson ZR, Westgate PM *et al.* Human adipose beigeing in response to cold and mirabegron. *JCI Insight* 2018 **3** e121510.
- 121 Fisher FM, Kleiner S, Douris N, Fox EC, Mepani RJ, Verdeguer F, Wu J, Kharitononkov A, Flier JS, Maratos-Flier E *et al.* FGF21 regulates PGC-1alpha and browning of white adipose tissues in adaptive thermogenesis. *Genes and Development* 2012 **26** 271–281. (<https://doi.org/10.1101/gad.177857.111>)
- 122 Lee P, Linderman JD, Smith S, Brychta RJ, Wang J, Idelson C, Perron RM, Werner CD, Phan GQ, Kammula US *et al.* Irisin and FGF21 are cold-induced endocrine activators of brown fat function in humans. *Cell Metabolism* 2014 **19** 302–309. (<https://doi.org/10.1016/j.cmet.2013.12.017>)
- 123 Hondares E, Iglesias R, Giral A, Gonzalez FJ, Giral M, Mampel T & Villarroya F. Thermogenic activation induces FGF21 expression and release in brown adipose tissue. *Journal of Biological Chemistry* 2011 **286** 12983–12990. (<https://doi.org/10.1074/jbc.M110.215889>)
- 124 Hondares E, Gallego-Escuredo JM, Flachs P, Frontini A, Cereijo R, Goday A, Perugini J, Kopecky P, Giral M, Cinti S *et al.* Fibroblast

- growth factor-21 is expressed in neonatal and pheochromocytoma-induced adult human brown adipose tissue. *Metabolism: Clinical and Experimental* 2014 **63** 312–317. (<https://doi.org/10.1016/j.metabol.2013.11.014>)
- 125 Markan KR, Naber MC, Ameka MK, Anderegg MD, Mangelsdorf DJ, Kliewer SA, Mohammadi M & Potthoff MJ. Circulating FGF21 is liver derived and enhances glucose uptake during refeeding and overfeeding. *Diabetes* 2014 **63** 4057–4063. (<https://doi.org/10.2337/db14-0595>)
- 126 Talukdar S, Zhou Y, Li D, Rossulek M, Dong J, Somayaji V, Weng Y, Clark R, Lanba A, Owen BM *et al*. A long-acting FGF21 molecule, PF-05231023, decreases body weight and improves lipid profile in non-human primates and Type 2 diabetic subjects. *Cell Metabolism* 2016 **23** 427–440. (<https://doi.org/10.1016/j.cmet.2016.02.001>)
- 127 Gaich G, Chien JY, Fu H, Glass LC, Deeg MA, Holland WL, Kharitonov A, Bumol T, Schilske HK & Moller DE. The effects of LY2405319, an FGF21 analog, in obese human subjects with type 2 diabetes. *Cell Metabolism* 2013 **18** 333–340. (<https://doi.org/10.1016/j.cmet.2013.08.005>)
- 128 Wei W, Dutchak PA, Wang X, Ding X, Wang X, Bookout AL, Goetz R, Mohammadi M, Gerard RD, Dechow PC *et al*. Fibroblast growth factor 21 promotes bone loss by potentiating the effects of peroxisome proliferator-activated receptor gamma. *PNAS* 2012 **109** 3143–3148. (<https://doi.org/10.1073/pnas.1200797109>)
- 129 Tseng YH, Kokkotou E, Schulz TJ, Huang TL, Winnay JN, Taniguchi CM, Tran TT, Suzuki R, Espinoza DO, Yamamoto Y *et al*. New role of bone morphogenetic protein 7 in brown adipogenesis and energy expenditure. *Nature* 2008 **454** 1000–1004. (<https://doi.org/10.1038/nature07221>)
- 130 Okla M, Ha JH, Temel RE & Chung S. BMP7 drives human adipogenic stem cells into metabolically active beige adipocytes. *Lipids* 2015 **50** 111–120. (<https://doi.org/10.1007/s11745-014-3981-9>)
- 131 Elsen M, Raschke S, Tennagels N, Schwahn U, Jelenik T, Roden M, Romacho T & Eckel J. BMP4 and BMP7 induce the white-to-brown transition of primary human adipose stem cells. *American Journal of Physiology: Cell Physiology* 2014 **306** C431–C440. (<https://doi.org/10.1152/ajpcell.00290.2013>)
- 132 Boon MR, van den Berg SA, Wang Y, van den Bossche J, Karkampouna S, Bauwens M, De Saint-Hubert M, van der Horst G, Vukicevic S, de Winther MP *et al*. BMP7 activates brown adipose tissue and reduces diet-induced obesity only at subthermoneutrality. *PLoS ONE* 2013 **8** e74083. (<https://doi.org/10.1371/journal.pone.0074083>)
- 133 White AP, Vaccaro AR, Hall JA, Whang PG, Friel BC & McKee MD. Clinical applications of BMP-7/OP-1 in fractures, nonunions and spinal fusion. *International Orthopaedics* 2007 **31** 735–741. (<https://doi.org/10.1007/s00264-007-0422-x>)
- 134 Hiatt WR, Kaul S & Smith RJ. The cardiovascular safety of diabetes drugs – insights from the rosiglitazone experience. *New England Journal of Medicine* 2013 **369** 1285–1287. (<https://doi.org/10.1056/NEJMp1309610>)
- 135 Coelho MS, de Lima CL, Royer C, Silva JB, Oliveira FC, Christ CG, Pereira SA, Bao SN, Lima MC, Pitta MG *et al*. GQ-16, a TZD-derived partial PPARgamma agonist, induces the expression of thermogenesis-related genes in brown fat and visceral white fat and decreases visceral adiposity in obese and hyperglycemic mice. *PLoS ONE* 2016 **11** e0154310. (<https://doi.org/10.1371/journal.pone.0154310>)
- 136 Ohno H, Shinoda K, Spiegelman BM & Kajimura S. PPARgamma agonists induce a white-to-brown fat conversion through stabilization of PRDM16 protein. *Cell Metabolism* 2012 **15** 395–404. (<https://doi.org/10.1016/j.cmet.2012.01.019>)
- 137 Loh RKC, Formosa MF, Eikelis N, Bertovic DA, Anderson MJ, Barwood SA, Nanayakkara S, Cohen ND, La Gerche A, Reutens AT *et al*. Pioglitazone reduces cold-induced brown fat glucose uptake despite induction of browning in cultured human adipocytes: a randomised, controlled trial in humans. *Diabetologia* 2018 **61** 220–230. (<https://doi.org/10.1007/s00125-017-4479-9>)
- 138 Shimasaki T, Masaki T, Mitsutomi K, Ueno D, Gotoh K, Chiba S, Kakuma T & Yoshimatsu H. The dipeptidyl peptidase-4 inhibitor des-fluoro-sitagliptin regulates brown adipose tissue uncoupling protein levels in mice with diet-induced obesity. *PLoS ONE* 2013 **8** e63626. (<https://doi.org/10.1371/journal.pone.0063626>)
- 139 Nahon KJ, Doornink F, Straat ME, Botani K, Martinez-Tellez B, Abreu-Vieira G, van Klinken JB, Voortman GJ, Friesema ECH, Ruiz JR *et al*. Effect of sitagliptin on energy metabolism and brown adipose tissue in overweight individuals with prediabetes: a randomised placebo-controlled trial. *Diabetologia* 2018 **61** 2386–2397. (<https://doi.org/10.1007/s00125-018-4716-x>)
- 140 Bostrom P, Wu J, Jedrychowski MP, Korde A, Ye L, Lo JC, Rasbach KA, Bostrom EA, Choi JH, Long JZ *et al*. A PGC1-alpha-dependent myokine that drives brown-fat-like development of white fat and thermogenesis. *Nature* 2012 **481** 463–468. (<https://doi.org/10.1038/nature10777>)
- 141 Zhang Y, Xie C, Wang H, Foss RM, Clare M, George EV, Li S, Katz A, Cheng H, Ding Y *et al*. Irisin exerts dual effects on browning and adipogenesis of human white adipocytes. *American Journal of Physiology: Endocrinology and Metabolism* 2016 **311** E530–E541. (<https://doi.org/10.1152/ajpendo.00094.2016>)
- 142 Otero-Diaz B, Rodriguez-Flores M, Sanchez-Munoz V, Monraz-Preciado F, Ordonez-Ortega S, Becerril-Elias V, Baay-Guzman G, Obando-Monge R, Garcia-Garcia E, Palacios-Gonzalez B *et al*. Exercise induces white adipose tissue browning across the weight spectrum in humans. *Frontiers in Physiology* 2018 **9** 1781. (<https://doi.org/10.3389/fphys.2018.01781>)
- 143 Hecksteden A, Wegmann M, Steffen A, Kraushaar J, Morsch A, Ruppenthal S, Kaestner L & Meyer T. Irisin and exercise training in humans – results from a randomized controlled training trial. *BMC Medicine* 2013 **11** 235. (<https://doi.org/10.1186/1741-7015-11-235>)
- 144 Perakakis N, Triantafyllou GA, Fernandez-Real JM, Huh JY, Park KH, Seufert J & Mantzoros CS. Physiology and role of irisin in glucose homeostasis. *Nature Reviews: Endocrinology* 2017 **13** 324–337. (<https://doi.org/10.1038/nrendo.2016.221>)
- 145 Kaisanlahti A & Glumoff T. Browning of white fat: agents and implications for beige adipose tissue to type 2 diabetes. *Journal of Physiology and Biochemistry* 2019 **75** 1–10. (<https://doi.org/10.1007/s13105-018-0658-5>)
- 146 Whittle AJ, Carobbio S, Martins L, Slawik M, Hondares E, Vazquez MJ, Morgan D, Csikasz RI, Gallego R, Rodriguez-Cuenca S *et al*. BMP8B increases brown adipose tissue thermogenesis through both central and peripheral actions. *Cell* 2012 **149** 871–885. (<https://doi.org/10.1016/j.cell.2012.02.066>)
- 147 Pellegrinelli V, Peirce VJ, Howard L, Virtue S, Turei D, Senzacqua M, Frontini A, Dalley JW, Horton AR, Bidault G *et al*. Adipocyte-secreted BMP8b mediates adrenergic-induced remodeling of the neurovascular network in adipose tissue. *Nature Communications* 2018 **9** 4974. (<https://doi.org/10.1038/s41467-018-07453-x>)
- 148 Wang GX, Zhao XY, Meng ZX, Kern M, Dietrich A, Chen Z, Cozacov Z, Zhou D, Okunade AL, Su X *et al*. The brown fat-enriched secreted factor Nrg4 preserves metabolic homeostasis through attenuation of hepatic lipogenesis. *Nature Medicine* 2014 **20** 1436–1443. (<https://doi.org/10.1038/nm.3713>)
- 149 Chen Z, Wang GX, Ma SL, Jung DY, Ha H, Altamimi T, Zhao XY, Guo L, Zhang P, Hu CR *et al*. Nrg4 promotes fuel oxidation and a healthy adipokine profile to ameliorate diet-induced metabolic disorders. *Molecular Metabolism* 2017 **6** 863–872. (<https://doi.org/10.1016/j.molmet.2017.03.016>)
- 150 Wang R, Yang F, Qing L, Huang R, Liu Q & Li X. Decreased serum neuregulin 4 levels associated with non-alcoholic fatty liver disease in children with obesity. *Clinical Obesity* 2019 **9** e12289. (<https://doi.org/10.1111/cob.12289>)

- 151 Lynes MD, Leiria LO, Lundh M, Bartelt A, Shamsi F, Huang TL, Takahashi H, Hirshman MF, Schlein C, Lee A *et al.* The cold-induced lipokine 12,13-diHOME promotes fatty acid transport into brown adipose tissue. *Nature Medicine* 2017 **23** 631–637. (<https://doi.org/10.1038/nm.4297>)
- 152 Stanford KI, Lynes MD, Takahashi H, Baer LA, Arts PJ, May FJ, Lehnig AC, Middelbeek RJW, Richard JJ, So K *et al.* 12,13-diHOME: an exercise-induced lipokine that increases skeletal muscle fatty acid uptake. *Cell Metabolism* 2018 **27** 1111.e3–1120.e3. (<https://doi.org/10.1016/j.cmet.2018.03.020>)
- 153 Vasan SK, Noordam R, Gowri MS, Neville MJ, Karpe F & Christodoulides C. The proposed systemic thermogenic metabolites succinate and 12,13-diHOME are inversely associated with adiposity and related metabolic traits: evidence from a large human cross-sectional study. *Diabetologia* 2019 **62** 2079–2087. (<https://doi.org/10.1007/s00125-019-4947-5>)
- 154 Gnad T, Scheibler S, von Kügelgen I, Scheele C, Kilić A, Glöde A, Hoffmann LS, Reverte-Salasa L, Horn P, Mutlu S *et al.* Adenosine activates brown adipose tissue and recruits beige adipocytes via A2A receptors. *Nature* 2014 **516** 395–399. (<https://doi.org/10.1038/nature13816>)
- 155 Lahesmaa M, Oikonen V, Helin S, Luoto P, Din MU, Pfeifer A, Nuutila P & Virtanen KA. Regulation of human brown adipose tissue by adenosine and A2A receptors – studies with [(15)O]H2O and [(11)C]TMSX PET/CT. *European Journal of Nuclear Medicine and Molecular Imaging* 2019 **46** 743–750. (<https://doi.org/10.1007/s00259-018-4120-2>)
- 156 Deshmukh AS, Peijs L, Beaudry JL, Jespersen NZ, Nielsen CH, Ma T, Brunner AD, Larsen TJ, Bayarri-Olmos R, Prabhakar BS *et al.* Proteomics-based comparative mapping of the secretomes of human brown and white adipocytes reveals EPDR1 as a novel fatokine. *Cell Metabolism* 2019 **30** 963.e7–975.e7. (<https://doi.org/10.1016/j.cmet.2019.10.001>)
- 157 Osman F, Franklyn JA, Daykin J, Chowdhary S, Holder RL, Sheppard MC & Gammage MD. Heart rate variability and turbulence in hyperthyroidism before, during, and after treatment. *American Journal of Cardiology* 2004 **94** 465–469. (<https://doi.org/10.1016/j.amjcard.2004.04.061>)
- 158 Scheen AJ. Sibutramine on cardiovascular outcome. *Diabetes Care* 2011 **34** (Supplement 2) S114–S119. (<https://doi.org/10.2337/dc11-s205>)
- 159 Ceesay SM, Prentice AM, Day KC, Murgatroyd PR, Goldberg GR, Scott W & Spurr GB. The use of heart rate monitoring in the estimation of energy expenditure: a validation study using indirect whole-body calorimetry. *British Journal of Nutrition* 1989 **61** 175–186. (<https://doi.org/10.1079/bjn19890107>)
- 160 Cutting WC, Mehrtens HG & Tainter ML. Actions and uses of dinitrophenol – promising metabolic applications. *JAMA* 1933 **101** 193–195. (<https://doi.org/10.1001/jama.1933.02740280013006>)
- 161 Hoch FL & Hogan FP. Hyperthermia, muscle rigidity, and uncoupling in skeletal muscle mitochondria in rats treated with halothane and 2,4-dinitrophenol. *Anesthesiology* 1973 **38** 237–243. (<https://doi.org/10.1097/0000542-197303000-00007>)
- 162 Grundlingh J, Dargan PI, El-Zanfaly M & Wood DM. 2,4-Dinitrophenol (DNP): a weight loss agent with significant acute toxicity and risk of death. *Journal of Medical Toxicology* 2011 **7** 205–212. (<https://doi.org/10.1007/s13181-011-0162-6>)
- 163 Langeveld M, Tan CY, Soeters MR, Virtue S, Ambler GK, Watson LP, Murgatroyd PR, Chatterjee VK & Vidal-Puig A. Mild cold effects on hunger, food intake, satiety and skin temperature in humans. *Endocrine Connections* 2016 **5** 65–73. (<https://doi.org/10.1530/EC-16-0004>)
- 164 Caldeira da Silva CC, Cerqueira FM, Barbosa LF, Medeiros MH & Kowaltowski AJ. Mild mitochondrial uncoupling in mice affects energy metabolism, redox balance and longevity. *Aging Cell* 2008 **7** 552–560. (<https://doi.org/10.1111/j.1474-9726.2008.00407.x>)
- 165 Blundell J, Finlayson G, Axelsen M, Flint A, Gibbons C, Kvist T & Hjerstedt JB. Effects of once-weekly semaglutide on appetite, energy intake, control of eating, food preference and body weight in subjects with obesity. *Diabetes, Obesity and Metabolism* 2017 **19** 1242–1251. (<https://doi.org/10.1111/dom.12932>)
- 166 Liu X, Wang S, You Y, Meng M, Zheng Z, Dong M, Lin J, Zhao Q, Zhang C, Yuan X *et al.* Brown adipose tissue transplantation reverses obesity in ob/ob mice. *Endocrinology* 2015 **156** 2461–2469. (<https://doi.org/10.1210/en.2014-1598>)
- 167 Min SY, Kady J, Nam M, Rojas-Rodriguez R, Berkenwald A, Kim JH, Noh HL, Kim JK, Cooper MP, Fitzgibbons T *et al.* Human ‘Brite/beige’ adipocytes develop from capillary networks, and their implantation improves metabolic homeostasis in mice. *Nature Medicine* 2016 **22** 312–318. (<https://doi.org/10.1038/nm.4031>)
- 168 Brychta RJ, Huang S, Wang J, Leitner BP, Hattenbach JD, Bell SL, Fletcher LA, Perron Wood R, Idelson CR, Duckworth CJ *et al.* Quantification of the capacity for cold-induced thermogenesis in young men With and Without obesity. *Journal of Clinical Endocrinology and Metabolism* 2019 **104** 4865–4878. (<https://doi.org/10.1210/jc.2019-00728>)
- 169 Orava J, Nuutila P, Lidell ME, Oikonen V, Noponen T, Viljanen T, Scheinin M, Taittonen M, Niemi T, Enerback S *et al.* Different metabolic responses of human brown adipose tissue to activation by cold and insulin. *Cell Metabolism* 2011 **14** 272–279. (<https://doi.org/10.1016/j.cmet.2011.06.012>)

Received 15 December 2020

Revised version received 4 March 2021

Accepted 10 March 2021