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Case report: Optical coherence tomography for monitoring biologic therapy in psoriasis and atopic dermatitis

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Biologic therapies are increasingly used to treat chronic inflammatory skin diseases such as psoriasis and atopic dermatitis. In clinical practice, scores based on evaluation of objective and subjective symptoms are used to assess disease severity, leading to evaluation of treatment goals with clinical decisions on treatment initiation, switch to another treatment modality or to discontinue current treatment. However, this visual-based scoring is relatively subjective and inaccurate due to inter- and intraobserver reliability. Optical coherence tomography (OCT) is a fast, high-resolution, in vivo imaging modality that enables the visualization of skin structure and vasculature. We evaluated the use of OCT for guantification and monitoring of skin inflammation to improve objective assessment of disease activity in patients with psoriasis and atopic dermatitis. We assessed the following imaging parameters including epidermal thickness, vascular density, plexus depth, vessel diameter, and vessel count. A total of four patients with psoriasis or atopic dermatitis were treated with biologic agents according to current treatment guidelines. OCT was used to monitor their individual treatment response in a target lesion representing disease activity for 52 weeks. Psoriatic and eczema lesions exhibited higher epidermal thickness, increased vascular density, and higher vessel count compared to uninvolved skin. An upward shift of the superficial vascular plexus accompanied by smaller vessel diameters was seen in psoriasis in contrast to atopic dermatitis, where larger vessels were observed. A response to biologic therapy was characterized by normalization of the imaging parameters in the target lesions in comparison to uninvolved skin during the observation period of 52 weeks. Optical coherence tomography potentially serves as an instrument to monitor biologic therapy in inflammatory skin diseases. Imaging parameters may enable objective quantification of inflammation in psoriasis or atopic dermatitis in selected representative skin areas. OCT may reveal persistent subclinical inflammation in atopic dermatitis beyond clinical remission.

KEYWORDS

optical coherence tomography (OCT), skin inflammation, psoriasis, atopic dermatitis, biologic therapy, *in vivo* imaging

Introduction

Psoriasis and atopic dermatitis are both common inflammatory skin diseases. About 30% of the patients with psoriasis (1) and about 10% with atopic dermatitis (2) require a systemic therapy due to their disease severity. Due to their safety profile and efficacy, biologic agents (3–5), and small molecules (6) are being increasingly used. A main drawback of these innovative treatments is that they may lose efficacy over time and that not all patients respond to a selected therapy. Also, in some cases the current dosing is not needed and potentially exposes the individuals to adverse events that may be avoided by adjusted dosing that fits their individual needs. Currently, however, indication and continuation of treatment are mainly dependent on clinical scores. These clinical scores are based on visual examination that might be biased by the experience of the clinician resulting in inter- and intraobserver variabilities (7, 8).

Facing these limitations, we aimed to develop an objective and more reliable evaluation method using image-based scoring to enable objective guidance of clinical decision making to avoid delayed and inconsistent therapy decisions.

Optical coherence tomography (OCT) is a fast, highresolution, in vivo imaging method with growing influence in the dermatological practice, especially in non-melanoma skin cancer (9-12). The imaging technique of OCT is based on Michelson interferometry (13). In the assessment of inflammatory skin diseases, additional information on vascular network is of special interest. OCT angiography based on speckle variance detection allows the visualization of cutaneous vasculature (14). It was reported that different vascular patterns and shapes (dots, blobs, coils, lines, curves, and serpiginous vessels) could distinguish healthy skin from lesional inflammatory skin (15, 16). In psoriatic skin, structural changes and alterations in vessel density and size were shown (17-19). Previous studies demonstrated the correlation between histopathological findings and structural features of psoriasis and chronic inflammation apparent in OCT (20, 21). In atopic dermatitis, epidermal hypertrophy and vascular depth were discussed as important parameters for disease severity (22).

In the past 20 years, OCT was used for single timepoint observations and short-term monitoring of psoriasis and atopic dermatitis (23–25). Based on these observations, here, we performed an interim analysis of a large ongoing prospective, long-term, observational study using a clinically approved OCT system for monitoring patients with psoriasis and atopic dermatitis undergoing biologic treatment over 52 weeks.

Materials and methods

Subjects and biologic therapies

The study was conducted according to the guidelines of the Declaration of Helsinki and ethical approval was obtained from the Ethics Committee of the University of Lübeck. Adult subjects eligible for inclusion were diagnosed with moderate to severe plaque psoriasis or atopic dermatitis and received treatment according to current treatment guidelines. Written informed consent was obtained from all patients. OCT data of three patients with psoriasis and one patient with atopic dermatitis was analyzed. They underwent biologic treatment with anti-IL-17 ixekizumab (n = 1), anti-IL-23 risankizumab (n = 1), anti-TNF- α certolizumab (n = 1) or anti-IL-4R α dupilumab (n = 1). At baseline, week 2, 4, 16, 28, 40, and 52 of treatment, clinical scores were determined and OCT scans were performed at one lesional site and one perilesional control site for each patient (**Figure 1**).

Clinical scoring

Visual-based scores such as psoriasis area and severity index (PASI) (7), eczema area and severity index (EASI) (8), and body surface area (BSA) (7) were determined by an experienced physician. Additional digital dermoscopy images were captured with DermoGenius ultra polarized (DermoScan GmbH, Regensburg, Germany).

To date, whole body OCT scanners do not exist and largearea scanning cannot be performed within a reasonable time. Thus, we must select target lesions for small-area imaging. The total sign score (TSS) was used to evaluate selected skin areas. For psoriasis, three clinical signs including erythema, induration, and desquamation were assessed using a 5-point scale (0 absent, 1 mild, 2 moderate, 3 severe, and 4 very severe)

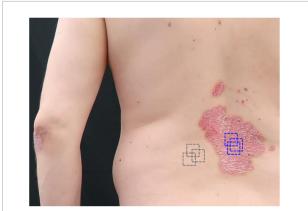


FIGURE 1

Selection of target lesions and control sites for optical coherence tomography (OCT). Patient 3 was a 45-year-old male patient with plaque psoriasis showing PASI = 10.8 and BSA = 10% at baseline. A representative target lesion (blue) on his trunk was selected for 52 weeks monitoring. A perilesional location was selected as the control site (gray). The scans were performed repetitively at the same lesional and non-lesional location.

with a total range of 0-12 (26). For atopic dermatitis, six clinical signs including erythema, edema/papulation, oozing/crusting, excoriation, lichenification, and dryness were graded using a 4-point scale (0 absent, 1 mild, 2 moderate, and 3 severe) with a total range of 0-18 (27).

Optical coherence tomography

We utilized a clinically approved VivoSight Dx OCT scanner (Michelson Diagnostics, Maidstone, Kent, UK). The emission wavelength is 1,305 nm. The lateral and axial resolution is <7.5 μm and <5 μm , respectively. The scan area is 6 mm \times 6 mm. The penetration depth is about 1.2 mm. A three-dimensional image stack with 6 mm width records a sequence of 180 B-scans with an interslice spacing of $33.3 \,\mu\text{m}$. Images were acquired as vertical B-scans and en-face scans. The acquisition time of an image stack is about 30 s generating both structural and vascular images. The en-face scans are reconstructed images to show skin layers at a constant depth from the skin surface. For an image stack with 1.2 mm depth, a sequence of 120 en-face scans with an interslice spacing of 10 µm can be obtained. Patients were at rest prior to scanning. Measurement conditions (location, patient position, and room temperature) were kept constant. Three regions of the same target lesion and three regions of its perilesional, clinically uninvolved skin area as reference were repetitively imaged. The target lesion was located either in upper extremities or trunk. Comparable to conventional computer-assisted dermoscopy, we used system-integrated overview images of the body and skin areas to make sure that measurements are done in consistent areas. The laser scanning handpiece uses a plastic tip as a distance holder between optical component and skin surface. The distance holder was gently placed onto the skin to avoid pressure- or shear-induced effects on the vessel diameter (28). Terminal hair was carefully trimmed. Scales were not removed. According to the patients' verbal feedback, the OCT scans do not lead to any patient discomfort.

Imaging parameters

Clinical signs such as erythema and skin thickening can result from microvascular changes and epidermal hypertrophy, respectively. Thus, we defined epidermal thickness, vascular density, depth, diameter, and vessel count as imaging parameters for quantification of skin inflammation (Figure 2). The calculation of vascular density, depth, and diameter was assisted by the proprietary vessel analysis software (Michelson Diagnostics, Maidstone, Kent, UK). All measures were performed by the same operator.

Epidermal thickness ET

We used the integrated on-screen ruler tool to measure the maximum epidermal thickness (*ET*) per image stack, then a mean ET of three image stacks was calculated with standard deviation. ET_L is the epidermal thickness of the target lesion and ET_C of the control at baseline. The same indices were used analogously for the other imaging parameters.

Vascular density ϱ

The vascular density ϱ was calculated at the depth of the superficial plexus. It is the density of the top of the superficial plexus calculated over the depth range, which is the plexus depth \pm 60 $\mu m.$

Plexus depth θ

The plexus depth θ is the depth of the top of the superficial plexus, where the density reaches 50% of the maximum vessel density.

Vessel diameter δ

The vessel diameter δ is a modal value calculated at the depth of the superficial plexus. It is the diameter of the majority of the vessels.

Vessel count N

The vessel count N was calculated using ImageJ 1.53k (U.S. National Institutes of Health, Bethesda, MD, USA). We anticipated that the cross sections of the elongated capillaries could be most accurately measured at about 200 μ m depth, as the last few *en-face* scans capture undesired projection artifacts from the superficial vessels. The *en-face* images were thresholded and converted into binary images. Adjacent or overlapping vessels were divided by watershed segmentation.

The vessels were counted by automatic particle analyzer using Sobel edge detection.

Response rate

We obtained fit coefficients λ using MATLAB R2021b (The MathWorks, Natick, MA, USA) to fit *ET* to an exponential equation $ET = A \times \exp(-\lambda \times t) + ET_R$ as a function of time *t* based on least squares algorithm, where ET_R is the estimated minimum *ET* during 52 weeks of therapy and *A* is an adjusting parameter. The number of weeks of therapy t_R required for *ET* reduction achieving $1.25 \times ET_R$ is then given by $t_R = -\lambda^{-1} \times ln[(k \times ET_R - ET_R)/A]$ for k = 1.25. If physiologically achievable, instead of $k \times ET_R$, t_R can also be calculated for $0.5 \times ET_L$ (50% reduction of ET_L from baseline).

Statistical analysis

We used Pearson correlation coefficient r to show the correlation between *ET* or *N* and the clinical scores.

Results

Description of patients at baseline

Patient 1 was a 52-year-old male with psoriasis (PASI = 19.5, BSA = 37%) who was treated with ixekizumab. He had no previous systemic therapy.

Patient 2 was a 30-year-old male with psoriasis (PASI = 8.7, BSA = 11%) who received risankizumab. Prior therapies were acitretin, dimethyl fumarate, methotrexate, certolizumab, ustekinumab, and secukinumab.

Patient 3 was a 45-year-old male with psoriasis (PASI = 10.8, BSA = 10%) who underwent therapy with certolizumab. He was previously treated with methotrexate.

Patient 4 was a 50-year-old female with atopic dermatitis (EASI = 17, BSA = 25%) who was treated with dupilumab. She had no previous systemic therapy.

In patient 1 and 2, the target lesions were located on their right arms. In patient 3 and 4, the target lesions were located on their trunk. The control area was their perilesional skin.

Structural imaging parameters

Epidermal thickness

In line with clinical and histopathological findings, at baseline we measured a thicker ET_L in psoriasis due to hyperkeratosis (**Figure 3**, top). A similar observation was made in atopic dermatitis where the increased ET_L was due

to spongiosis and/or lichenification (**Figure 3**, bottom). At baseline, the mean ET_L was 405.8 μ m (±20.8 μ m) in psoriatic skin and 270 μ m (±25.5 μ m) in atopic dermatitis. ET_C ranged from 90 to 110 μ m and was similar in uninvolved skin of patients with psoriasis and atopic dermatitis. The thickened ET_L decreased during the observation period under therapy (**Figure 4**, row 1). According to t_R , 0.5 × ET_L was achieved as follows: ixekizumab (after 3.95 weeks), certolizumab (after 4.23 weeks), risankizumab (after 6.08 weeks), and dupilumab (after 22.13 weeks). 1.25 × ET_R was achieved as follows: ixekizumab (after 4.31 weeks), certolizumab (after 6.52 weeks), risankizumab (after 11.23 weeks), and dupilumab (after 33.89 weeks).

In good agreement with the OCT measurements, the clinical severity was reduced in the target lesion (TSS) and in the global assessment (PASI or EASI) (**Supplementary Figure 1**). A positive correlation was observed between ET_L and clinical scores as follows: ixekizumab ($r_{TSS} = 0.83$, p = 0.0208; $r_{PASI} = 0.9564$, p = 0.0007), certolizumab ($r_{TSS} = 0.8857$, p = 0.008; $r_{PASI} = 0.9265$, p = 0.0027), risankizumab ($r_{TSS} = 0.8968$, p = 0.0062; $r_{PASI} = 0.8017$, p = 0.0301), and dupilumab ($r_{TSS} = 0.9778$, p = 0.0007; $r_{EASI} = 0.9184$, p = 0.0097).

Decay rate

The decrease of ET_L under therapy yielded the following decay rates: ixekizumab ($\lambda = 0.4418$, $R^2 = 0.96$), certolizumab ($\lambda = 0.3224$, $R^2 = 0.95$), risankizumab ($\lambda = 0.2117$, $R^2 = 0.96$), and dupilumab ($\lambda = 0.05817$, $R^2 = 0.93$). In this comparison, the highest rate was observed for ixekizumab followed by certolizumab and risankizumab. The decay rate under dupilumab was a factor of 3.6–7.6 slower than the biologic therapies of psoriasis.

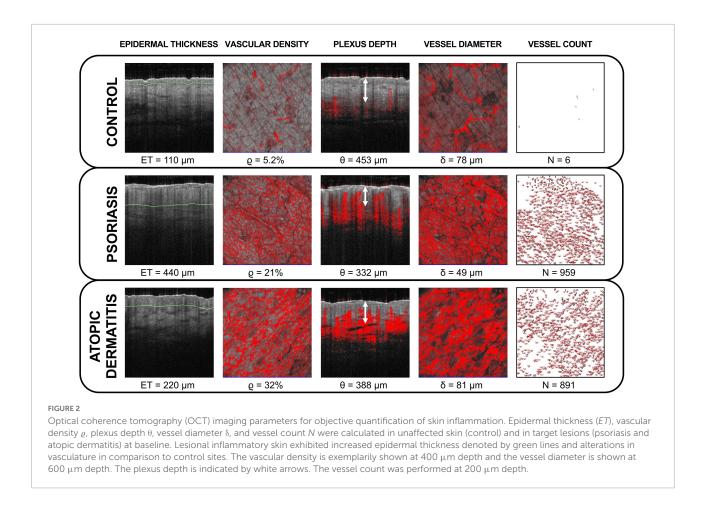
Vascular imaging parameters

Vascular patterns and shapes

We used the terminology as proposed by Ulrich et al. (15). Characteristic vascular patterns and shapes were visible in the *en-face* mode at about 200 μ m depth (Supplementary Figure 2). Psoriatic skin exhibited "dotted" or "pinpoint-like" vessels resulting from vessel elongation within extended dermal papillae. Lichenification is often observed in chronic eczema. Coarsening and wrinkling of the lichenified skin may change the direction of the capillary loops causing a striped pattern. Thus, "comma-like" vessels were observed in atopic dermatitis. Uninvolved skin typically has "linear" vessels generating a reticular pattern.

Vascular density, plexus depth, and vessel diameter

Vascular parameters decreased or normalized during therapy course (Figure 4, rows 2–4). Before treatment the mean



 ρ_L in psoriatic skin was 4.6% (±1.9%), which was higher than ρ_C with 2.1% (±1.23%). ρ_L in atopic dermatitis was 24.5% (±8.4%), while ρ_C was 6.3% (±1.24%). The higher ρ_L from baseline decreased after therapy start as a result of reduction in vessel diameter and/or vessel count (Supplementary Figure 3).

We defined the location of the origin (0,0) in the upper left of the image according to the general convention in image processing. This means that plexus depth and optical axis *z* have the same direction. For plots, the origin is located in the lower left. We observed an upward shift of the plexus $\Delta \theta = \theta_L \cdot \theta_C$ by $-83.4 \,\mu\text{m} (\pm 42.3 \,\mu\text{m})$ in psoriatic skin θ_L when compared to θ_C . At baseline, the mean δ_L in psoriatic skin was $35.2 \,\mu\text{m} (\pm 6.9 \,\mu\text{m})$, which was smaller than δ_C with $58.6 \,\mu\text{m}$ ($\pm 26.8 \,\mu\text{m}$). We did not observe an upward shift in the patient with atopic dermatitis. In contrast to the psoriatic lesions, in atopic dermatitis δ_L was $81 \,\mu\text{m} (\pm 3.1 \,\mu\text{m})$, which was larger than δ_C with 57.7 $\mu\text{m} (\pm 6.9 \,\mu\text{m})$.

Vessel count

The mean N_L in psoriasis and atopic dermatitis was 531.4 (±285.9) and 987 (±37.5), respectively. The mean N_C was 35.08 (±14.27). After 52 weeks, we observed a full normalization of the vessel count (**Figure 4**, row 5).

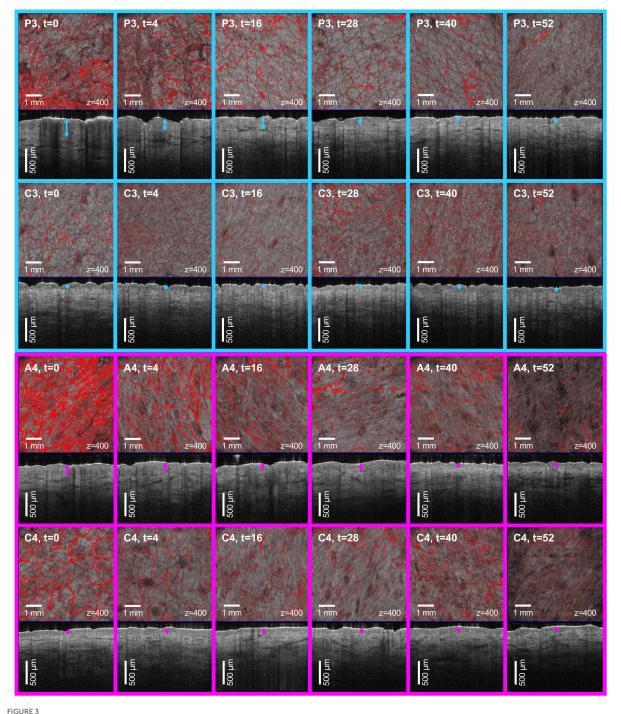
Also, a positive correlation was observed between *N* and clinical scores as follows: ixekizumab ($r_{TSS} = 0.85$, p = 0.0154; $r_{PASI} = 0.9589$, p = 0.0006), certolizumab ($r_{TSS} = 0.8978$, p = 0.0061; $r_{PASI} = 0.9327$, p = 0.0022), risankizumab ($r_{TSS} = 0.6055$, p = 0.1497; $r_{PASI} = 0.6792$, p = 0.0933), and dupilumab ($r_{TSS} = 0.9378$, p = 0.0057; $r_{EASI} = 0.9272$, p = 0.0078).

Clinical response

The results of this case series indicated a good correlation with the clinical scores. A 75% reduction from baseline in the TSS (TSS-75) was achieved under ixekizumab after 3 weeks and under certolizumab after 3.5 weeks. In this observation period, TSS-75 under risankizumab was not achieved. After 3 weeks, TSS-75 was achieved under dupilumab.

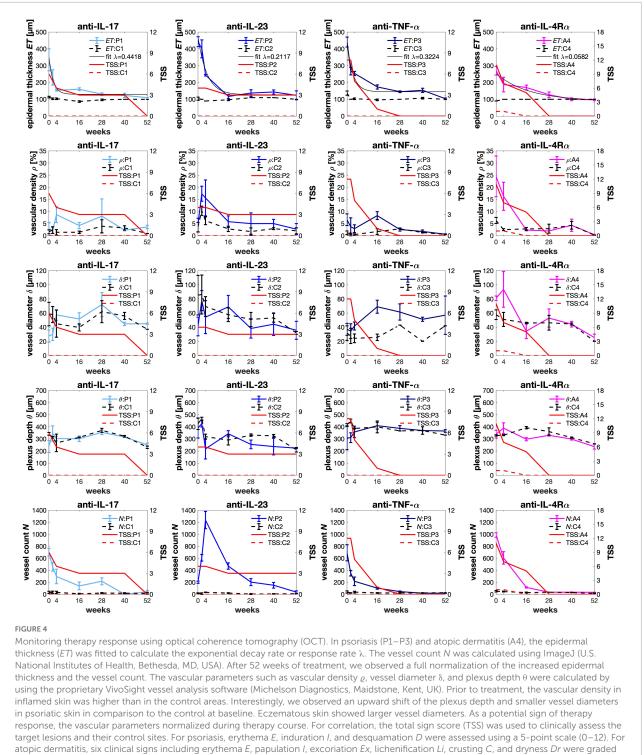
Discussion

Optical coherence tomography is a suitable imaging tool for the investigation and monitoring of inflammatory skin diseases. The VivoSight OCT offers user-friendly handling and fast scanning, which is important for its application as



Optical coherence tomography (OCT) guided monitoring of biologic therapy in psoriasis and atopic dermatitis. Patient 3 with psoriasis (P3) was treated with certolizumab (top) and patient 4 with atopic dermatitis (A4) was treated with dupilumab (bottom) for t = 52 weeks. The vertical B-scans and the corresponding en-face scans at 400 µm depth were collected in the course of therapy. Hyperkeratotic psoriatic plaques led to a signal reduction at baseline. Under therapy a decrease of the epidermal thickening (arrows) and a normalization of the vascularization (red signal) were observed in psoriasis and atopic dermatitis. No significant changes were detected in the control sites (C3 and C4).

a bedside monitoring device. But the system applied in this study has a limited resolution capacity that does not allow the detection of single cells. In comparison, the investigational line-field confocal OCT provides superior axial resolution of 1.1 µm and lateral resolution of 1.3 µm (29). This would allow a characterization of individual cells and microstructures



using a 4-point scale (0–18).

in inflammatory skin diseases (30). Furthermore, OCT with dynamic contrast allows color-coding of different epithelial cell layers based on micromotions of cellular structures (31, 32). So far, these technical upgrades have not yet been incorporated

into a clinically approved OCT system for dermatological applications. The future goal is to obtain contactless, whole body scans *via* rapid scanning of large body sites as commercial OCT systems only provide small fields of view (33, 34). To date, we

are dependent on the selection of target lesions for imaging. While psoriatic lesions tend to recur at the same skin sites that were affected previously, in atopic dermatitis the eczema lesions tend to shift location. Under this aspect, monitoring of target lesions could potentially miss an eczema flare or even psoriasis that exacerbates on a different body part.

Psoriasis and atopic dermatitis are associated with lower skin hydration (35, 36). Consequently, effects of laser-tissue interactions such as scattering (e.g., scaling, hyperkeratosis, and lichenification) and shielding (e.g., shadow artifacts caused by hair or crusts) could lower optical penetration and might masquerade as a loss of perfusion.

In an attempt to define imaging biomarkers for inflammatory skin diseases, we demonstrated the application of OCT for monitoring biologic therapy in psoriasis and atopic dermatitis based on four case studies. Biomarkers are objective, quantifiable, and reproducible (37). We should differentiate between robust and weak imaging biomarkers. Robust biomarkers might be ET and N, as these parameters are less influenced by internal and external factors. In this study, manual measurements of the maximum ET still remain vulnerable to intraobserver variability, which was minimized by repeated measurements. Hence, further studies should incorporate computer-assisted analysis of the ET. Weaker biomarkers such as ρ and δ might be highly influenced by internal stress factors of the subject (comparable to the "white coat effect") and outdoor temperatures (hot temperatures lead to vasodilation and cold temperatures lead to vasoconstriction) as we observed intraindividual variations in control sites.

In this work, we provided a detailed description on vascular alterations in psoriasis and atopic dermatitis compared to non-lesional skin. Our findings were consistent with previous observations reported on higher vascularization and characteristic vascular pattern in psoriasis (16-18). While "dotted" vessels were previously described in psoriasis (38), we firstly described the appearance of "comma-like" vessels in atopic dermatitis with lichenification. We detected an upward shift of the plexus in psoriasis. We anticipated that a plexus shift results from an elongation of the capillary loops, which is more distinctive in severe psoriasis. At baseline, no shift of the plexus depth was observed in our patient with atopic dermatitis. Byers et al. (22) described deeper vascular layers in atopic dermatitis so that we need to make further investigations before coming to a conclusion. We observed large vessels and high vascular densities in atopic dermatitis that were consistent with former studies (38, 39). Manfredini et al. (23) reported on a reduction of dermal edema and vascularization under dupilumab. In our analysis, we also observed a normalization of the vasculature under therapy. Further, we observed that vessel elongation in psoriasis appeared with smaller capillary loop diameters. Evidently, a larger case number is required to confirm our preliminary observations.

The aim of biologic treatment especially in atopic dermatitis is to improve the skin barrier. We observed a good correlation

between *ET* and TSS. Our results on the decrease of *ET* under therapy were in good agreement with previous studies using topical (25, 40) and non-biologic systemic therapies (19). In contrast to conventional therapies of psoriasis, we observed an accelerated decrease of *ET*. Similar results on *ET* in atopic dermatitis were reported under dupilumab (23). We also anticipated that *ET* may be a robust imaging biomarker as already stated by Byers et al. (22). The rapid decrease in *ET* can be regarded as an exponential decay. We interpreted the decay rate λ as a response rate. In theory, $\lambda = 0$ means steady state. Values of $\lambda > 0$ implicate therapy response meaning the larger λ the higher the response. $\lambda < 0$ refers to therapy failure.

Patient 2 was bio-experienced with a long record of pretreatments, therefore, a slower therapy response in TSS was seen under risankizumab. Interestingly, in patient 4 with atopic dermatitis higher vessel density and diameter were observed in the control area at baseline indicating subclinical inflammation in clinically healthy-appearing skin. In addition, although a rapid improvement of patient 4 was clinically observed shown as TSS-75 after 3 weeks, using OCT we were able to detect a prolonged epidermal thickening for $t_R(1.25 \times ET_R) = 33.89$ weeks as a sign of persistent disease activity. Comparable observations were reported by Byers et al. (22). The understanding of subclinical inflammation highlights the importance of therapy continuation to avoid the risk of relapse (41).

Conclusion

Preliminary observations of this work showed that OCT may be suitable for objective quantification of structural (epidermal thickness) and vascular parameters (vascular density, depth, diameter, and count). These parameters may serve as objective imaging biomarkers for monitoring therapeutic effects in psoriasis and atopic dermatitis. The relatively short acquisition time of OCT is an important demand for medical imaging to minimize time burden for patients.

Therapy response may be characterized by reduction of epidermal thickness and normalization of vascular network. Potential imaging biomarkers such as epidermal thickness and vessel count exhibited rapid changes. The calculated response rate λ may serve as a useful parameter in the assessment of therapeutic effects. Additional diagnostic value of OCT angiography could be seen in the detection of subclinical inflammation that implicates the need for therapy continuation beyond clinical remission.

In this preliminary evaluation, all patients have responded well to their systemic treatment. In the next step, we will also evaluate insufficient therapy responses or therapy failures in a large prospective, long-term, observational study. Additional imaging biomarkers revealing desquamation, excoriation, and lichenification should be considered in upcoming studies. The vision of the future is to provide "OCT-guided therapy" enhancing the current dermatological assessment and contributing to personalized medicine.

Data availability statement

The original contributions presented in this study are included in the article/Supplementary material, further inquiries can be directed to the corresponding author.

Ethics statement

The studies involving human participants were reviewed and approved by Ethics Committee of the University of Lübeck, Germany. The patients/participants provided their written informed consent to participate in this study.

Author contributions

LH-W and HY contributed to data acquisition. LH-W performed the data analysis and wrote the main manuscript text. All authors contributed to manuscript revision, read, and approved the submitted version.

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References

1. Martin G, Young M, Aldredge L. Recommendations for initiating systemic therapy in patients with psoriasis. *J Clin Aesthet Dermatol.* (2019) 12:13–26.

2. Werfel T, Schwerk N, Hansen G, Kapp A. The diagnosis and graded therapy of atopic dermatitis. *Dtsch Arztebl Int.* (2014) 111:509–20. doi: 10.3238/arztebl.2014. 0509

3. Armstrong AW, Puig L, Joshi A, Skup M, Williams D, Li J, et al. Comparison of biologics and oral treatments for plaque psoriasis: a metaanalysis. *JAMA Dermatol.* (2020) 156:258–69. doi: 10.1001/jamadermatol.2019. 4029

4. Guttman-Yassky E, Krueger JG, Lebwohl MG. Systemic immune mechanisms in atopic dermatitis and psoriasis with implications for treatment. *Exp Dermatol.* (2018) 27:409–17. doi: 10.1111/exd. 13336

5. Thaçi D, Simpson EL, Deleuran M, Kataoka Y, Chen Z, Gadkari A, et al. Efficacy and safety of dupilumab monotherapy in adults with moderate-to-severe atopic dermatilis: a pooled analysis of two phase 3 randomized trials (LIBERTY AD SOLO 1 and LIBERTY AD SOLO 2). *J Dermatol Sci.* (2019) 94:266–75. doi: 10.1016/j.jdermsci.2019. 02.002

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Conflict of interest

Author DT declares being a consultant, investigator, speaker, and participating in scientific advisory boards for AbbVie, Almirall, Amgen, Biogen Idec, Eli Lilly, Galapagos, Janssen-Cilag, LEO Pharma, MSD, Novartis, Pfizer, Regeneron, Samsung, Sanofi, and UCB; and research/educational grants from AbbVie, LEO Pharma, and Novartis.

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/ fmed.2022.995883/full#supplementary-material

6. Solimani F, Meier K, Ghoreschi K. Emerging topical and systemic JAK inhibitors in dermatology. *Front Immunol.* (2019) 3:2847. doi: 10.3389/fimmu. 2019.02847

7. Bożek A, Reich A. The reliability of three psoriasis assessment tools: psoriasis area and severity index, body surface area and physician global assessment. *Adv Clin Exp Med.* (2017) 26:851–6. doi: 10.17219/acem/69804

8. Bożek A, Reich A. Assessment of intra- and inter-rater reliability of three methods for measuring atopic dermatitis severity: EASI, objective SCORAD, and IGA. *Dermatology*. (2017) 233:16–22. doi: 10.1159/000472711

9. Wan B, Ganier C, Du-Harpur X, Harun N, Watt FM, Patalay R, et al. Applications and future directions for optical coherence tomography in dermatology. *Br J Dermatol.* (2021) 184:1014–22. doi: 10.1111/bjd.19553

10. Olsen J, Holmes J, Jemec GB. Advances in optical coherence tomography in dermatology-a review. *J Biomed Opt.* (2018) 23:1–10. doi: 10.1117/1.JBO.23.4. 040901

11. Ulrich M, Themstrup L, de Carvalho N, Manfredi M, Grana C, Ciardo S, et al. Dynamic optical coherence tomography in dermatology. *Dermatology*. (2016) 232:298–311. doi: 10.1159/000444706

12. Batz S, Wahrlich C, Alawi A, Ulrich M, Lademann J. Differentiation of different nonmelanoma skin cancer types using OCT. *Skin Pharmacol Physiol.* (2018) 31:238–45. doi: 10.1159/000489269

13. Huang D, Swanson EA, Lin CP, Schuman JS, Stinson WG, Chang W, et al. Optical coherence tomography. *Science*. (1991) 254:1178–81. doi: 10.1126/science. 1957169

14. Casper M, Schulz-Hildebrandt H, Evers M, Birngruber R, Manstein D, Hüttmann G. Optimization-based vessel segmentation pipeline for robust quantification of capillary networks in skin with optical coherence tomography angiography. *J Biomed Opt.* (2019) 24:1–11. doi: 10.1117/1.JBO.24.4.046005

15. Ulrich M, Themstrup L, de Carvalho N, Ciardo S, Holmes J, Whitehead R, et al. Dynamic optical coherence tomography of skin blood vessels - proposed terminology and practical guidelines. *J Eur Acad Dermatol Venereol.* (2018) 32:152–5. doi: 10.1111/jdv.14508

16. Schuh S, Holmes J, Ulrich M, Themstrup L, Jemec GBE, de Carvalho N, et al. Imaging blood vessel morphology in skin: dynamic optical coherence tomography as a novel potential diagnostic tool in dermatology. *Dermatol Ther.* (2017) 7:187– 202. doi: 10.1007/s13555-017-0175-4

17. Deegan AJ, Talebi-Liasi F, Song S, Li Y, Xu J, Men S, et al. Optical coherence tomography angiography of normal skin and inflammatory dermatologic conditions. *Lasers Surg Med.* (2018) 50:183–93. doi: 10.1002/lsm.22788

18. Zafar H, Enfield J, O'Connell ML, Ramsay B, Lynch M, Leahy MJ. Assessment of psoriatic plaque *in vivo* with correlation mapping optical coherence tomography. *Skin Res Technol.* (2014) 20:141–6. doi: 10.1111/srt.12096

19. Aldahan AS, Chen LL, Fertig RM, Holmes J, Shah VV, Mlacker S, et al. Vascular features of nail psoriasis using dynamic optical coherence tomography. *Skin Appendage Disord*. (2017) 2:102–8. doi: 10.1159/000449230

20. Morsy H, Kamp S, Thrane L, Behrendt N, Saunder B, Zayan H, et al. Optical coherence tomography imaging of psoriasis vulgaris: correlation with histology and disease severity. *Arch Dermatol Res.* (2010) 302:105–11. doi: 10.1007/s00403-009-1000-4

21. Silver R, Helms A, Fu W, Wang H, Diaconu D, Loyd CM, et al. Using optical coherence tomography for the longitudinal non-invasive evaluation of epidermal thickness in a murine model of chronic skin inflammation. *Skin Res Technol.* (2012) 18:225–31. doi: 10.1111/j.1600-0846.2011.00558.x

22. Byers RA, Maiti R, Danby SG, Pang EJ, Mitchell B, Carré MJ, et al. Sub-clinical assessment of atopic dermatitis severity using angiographic optical coherence tomography. *Biomed Opt Express.* (2018) 9:2001–17. doi: 10.1364/BOE.9.002001

23. Manfredini M, Liberati S, Ciardo S, Bonzano L, Guanti M, Chester J, et al. Microscopic and functional changes observed with dynamic optical coherence tomography for severe refractory atopic dermatitis treated with dupilumab. *Skin Res Technol.* (2020) 26:779–87. doi: 10.1111/srt.12868

24. Odorici G, Losi A, Ciardo S, Pellacani G, Conti A. Non-invasive evaluation of Secukinumab efficacy in severe plaque psoriasis with confocal microscopy and optical coherence tomography: a case report. *Skin Res Technol.* (2018) 24:160–2. doi: 10.1111/srt.12409

25. Welzel J, Bruhns M, Wolff HH. Optical coherence tomography in contact dermatitis and psoriasis. *Arch Dermatol Res.* (2003) 295:50–5. doi: 10.1007/s00403-003-0390-y

26. Gooderham M, Debarre JM, Keddy-Grant J, Xu Z, Kurvits M, Goodfield M. Safety and efficacy of calcipotriol plus betamethasone dipropionate gel in the treatment of scalp psoriasis in adolescents 12-17 years of age. *Br J Dermatol.* (2014) 171:1470–7. doi: 10.1111/bjd.13235

27. Guttman-Yassky E, Ungar B, Malik K, Dickstein D, Suprun M, Estrada YD, et al. Molecular signatures order the potency of topically applied anti-inflammatory

drugs in patients with atopic dermatitis. J Allergy Clin Immunol. (2017) 140:1032-42.e13. doi: 10.1016/j.jaci.2017.01.027

28. Wang-Evers M, Casper MJ, Glahn J, Luo T, Doyle AE, Karasik D, et al. Assessing the impact of aging and blood pressure on dermal microvasculature by reactive hyperemia optical coherence tomography angiography. *Sci Rep.* (2021) 11:13411. doi: 10.1038/s41598-021-92712-z

29. Ruini C, Schuh S, Sattler E, Welzel J. Line-field confocal optical coherence tomography-Practical applications in dermatology and comparison with established imaging methods. *Skin Res Technol.* (2021) 27:340–52. doi: 10.1111/srt. 12949

30. Boone M, Norrenberg S, Jemec G, Del Marmol V. High-definition optical coherence tomography: adapted algorithmic method for pattern analysis of inflammatory skin diseases: a pilot study. *Arch Dermatol Res.* (2013) 305:283–97. doi: 10.1007/s00403-012-1311-8

31. Münter M, Vom Endt M, Pieper M, Casper M, Ahrens M, Kohlfaerber T, et al. Dynamic contrast in scanning microscopic OCT. *Opt Lett.* (2020) 45:4766–9. doi: 10.1364/OL.396134

32. Scholler J, Groux K, Goureau O, Sahel JA, Fink M, Reichman S, et al. Dynamic full-field optical coherence tomography: 3D live-imaging of retinal organoids. *Light Sci Appl.* (2020) 17:140. doi: 10.1038/s41377-020-00375-8

33. Göb M, Burhan S, Draxinger W, Kolb JP, Huber R. Towards densely sampled ultra-large area multi-MHz-OCT for *in vivo* skin measurements beyond 1 cm²/sec. *European Conferences on Biomedical Optics 2021 (ECBO). OSA Technical Digest.* Washington, DC: Optical Society of America (2021).

34. Huang Y, Li X, Liu J, Qiao Z, Chen J, Hao Q. Robotic-arm-assisted flexible large field-of-view optical coherence tomography. *Biomed Opt Express.* (2021) 12:4596–609. doi: 10.1364/BOE.431318

35. Egawa M, Kunizawa N, Hirao T, Yamamoto T, Sakamoto K, Terui T, et al. *In vivo* characterization of the structure and components of lesional psoriatic skin from the observation with Raman spectroscopy and optical coherence tomography: a pilot study. *J Dermatol Sci.* (2010) 57:66–9. doi: 10.1016/j.jdermsci.2009.10.006

36. Hon KL, Lam PH, Ng WG, Kung JS, Cheng NS, Lin ZX, et al. Age, sex, and disease status as determinants of skin hydration and transepidermal water loss among children with and without eczema. *Hong Kong Med J.* (2020) 26:19–26. doi: 10.12809/hkmj198150

37. Sullivan DC, Obuchowski NA, Kessler LG, Raunig DL, Gatsonis C, Huang EP, et al. Metrology standards for quantitative imaging biomarkers. *Radiology.* (2015) 277:813–25. doi: 10.1148/radiol.201514 2202

38. Rajabi-Estarabadi A, Vasquez-Herrera NE, Martinez-Velasco MA, Tsatalis J, Verne SH, Nouri K, et al. Optical coherence tomography in diagnosis of inflammatory scalp disorders. *J Eur Acad Dermatol Venereol.* (2020) 34:2147–51. doi: 10.1111/jdv.16304

39. Blatter C, Weingast J, Alex A, Grajciar B, Wieser W, Drexler W, et al. In situ structural and microangiographic assessment of human skin lesions with high-speed OCT. Biomed Opt Express. (2012) 3:2636–46. doi: 10.1364/BOE.3. 002636

40. Yélamos O, Alejo B, Ertekin SS, Villa-Crespo L, Zamora-Barquero S, Martinez N, et al. Non-invasive clinical and microscopic evaluation of the response to treatment with clobetasol cream vs. calcipotriol/betamethasone dipropionate foam in mild to moderate plaque psoriasis: an investigator-initiated, phase IV, unicentric, open, randomized clinical trial. *J Eur Acad Dermatol Venereol.* (2021) 35:143–9. doi: 10.1111/jdv.16559

41. Tang TS, Bieber T, Williams HC. Are the concepts of induction of remission and treatment of subclinical inflammation in atopic dermatitis clinically useful?. J Allergy Clin Immunol. (2014) 133:1615–25.e1. doi: 10.1016/j.jaci.2013.12.1079