RESEARCH ARTICLE



A new species of Cyrtodactylus (Squamata, Gekkonidae) from Cambodia's Prey Lang Wildlife Sanctuary

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Abstract

Cyrtodactylus phnomchiensis **sp. nov.** is described from Phnom Chi, an isolated mountain in Prey Lang Wildlife Sanctuary, Kampong Thom Province, Cambodia. The new species is recognized by having a unique combination of morphological characters, including snout-vent length 76.1–80.7 mm; paravertebral tubercles 31–36; ventral scales 45–54; enlarged femoral scales 0–8, without pores; enlarged precloacal scales 7–10, bearing pores 4–5 in males, pits 1–7 in females; the posterior border of nuchal loop unbroken and pointed, bordered anteriorly and posteriorly by a broad yellow or yellowish white band; and yellow spots on top of head. The new species also represents a divergent mitochondrial DNA lineage within the *C. irregularis* complex that is closely related to *C. ziegleri*, but the phylogenetic relationships among the new species and two divergent mitochondrial subclades within *C. ziegleri* are not resolved based on available sequence data. *Cyrtodactylus phnomchiensis* **sp. nov.** is the only member of the *C. irregularis* complex known to occur west of the Mekong River. The new species may be endemic to Phnom Chi, and likely faces imminent conservation threats.

Keywords

Cyrtodactylus irregularis, C. ziegleri, Mekong River, Phnom Chi, Sphenomorphus preylangensis

Introduction

Bent-toed Geckos of the genus *Cyrtodactylus* Gray are one of the most species-diverse genera of gekkonid lizards, with 292 recognized species (Uetz et al. 2020). Much of the diversity within *Cyrtodactylus* has been described only during the past decade and from mainland Southeast Asia (Brennan et al. 2017; Uetz et al. 2020), and many of these newly-recognized species are thought to be highly localized with extremely narrow geographic ranges (e.g., Nazarov et al. 2012; Luu et al. 2016; Grismer et al. 2017; Murdoch et al. 2019).

Cyrtodactylus irregularis (Smith, 1921) was originally described from the Langbian Plateau near Da Lat, southern Vietnam. For nearly a century, *C. irregularis* was treated as a single, geographically widespread, but morphologically variable species. Recent taxonomic studies on variation in morphology and, usually, the mitochondrial cytochrome *c* oxidase subunit I (COI) gene (Brennan et al. 2017) have revealed that *C. irregularis* actually represents a complex of at least 19 species distributed in southern and central Vietnam, eastern Cambodia, and southern Laos (Nguyen et al. 2013, 2017; Pauwels et al. 2018). These include 18 named species from Vietnam recognized by Pauwels et al. (2018), as well as *C. buchardi* David, Teynie & Ohler, 2004 from southern Laos, a species that has been hypothesized to be a member of this complex (Ngo and Chan 2010; Nguyen et al. 2013, 2017) but that remains phylogenetically untested owing to lack of molecular data. The monophyly of the *C. irregularis* group has been demonstrated by phylogenetic analysis of the COI gene from most of the species in the complex (Nazarov et al. 2012; Nguyen et al. 2013, 2017; Luu et al. 2017; Schneider et al. 2014).

During field surveys by Wild Earth Allies in June–July 2019, five specimens of the *C. irregularis* complex were collected in Cambodia on the western side of the Mekong River at Phnom Chi (Mountain) in Prey Lang Wildlife Sanctuary, Kampong Thom Province. Herein, we investigate the taxonomic status of the Phnom Chi specimens through comparisons of morphological and mitochondrial DNA data with other members of the *C. irregularis* complex.

Materials and methods

Sampling

Field work was conducted both day and night to search microhabitats for amphibians and reptiles at Phnom Chi. Specimens were collected by hand and kept overnight in individual plastic or cloth bags for photographing the following day. Specimens were euthanized by cardiac injection of high concentration of tricaine methanesulfonate (MS-222) and fixed in 10% formalin after preserving liver tissue in 20% DMSO-salt saturated storage buffer. After a minimum of three days of formalin-fixation, the specimens were soaked in water for six hours to remove formalin, and transferred to 70% ethanol for permanent storage. Specimens were deposited in the herpetological collection at the Centre for Biodiversity Conservation, Royal University of Phnom Penh, Cambodia (**CBC**). Comparative data were taken from original species descriptions and the expanded descriptions of *C. irregularis* by Nazarov et al. (2008) and *C. buchardi* by Teyníe and David (2010).

Morphological analyses

Morphometric and meristic characters were measured and counted using a Nikon SMZ 645 dissecting microscope. Measurements were taken by hand with digital calipers to the nearest 0.1 mm (ratios calculated to 0.001). Measured characters were:

AG	Axilla-groin distance, measured from the posterior margin of forelimb at
	its insertion point on the body to the anterior margin of hind limb at its
	insertion point on the body;
CrusL	Crus length, measured from the knee to the base of the heel;
EarDH	Ear diameter in horizontal distance, measured as the horizontal distance
	between anterior and posterior margins of the ear opening;
EarDV	Ear diameter in vertical distance, measured as the vertical distance be-
	tween dorsal and ventral margins of the ear opening;
END	Eye-nostril distance, measured from the anterior margin of eye to the
	posterior margin of nostril;
ESD	Eye-snout distance, measured from the anterior margin of eye to the tip
	of snout;
EyeD	Eye diameter, measured as the horizontal distance from the anterior to
	the posterior margins of the eyeball;
Eye-EarD	Eye-ear distance, measured from the posterior margin of eye to the ante-
	rior margin of ear opening;
ForeL	Forearm length, measured from the posterior margin of elbow while
	flexed 90° to the wrist inflection;
HeadD	Head depth, measured as the maximum depth of head from the occiput
	to the throat;
HeadL	Head length, measured from the tip of snout to the posterior margin of
	the retroarticular process of the lower jaw;
HeadW	Head width, measured as the maximum head width at the corners of the
	jaws;
IOD	Interorbital distance, measured as the shortest distance between the ante-
	rior corners of the eyes;
IND	Internarial distance, measured as the shortest distance between the
	nostrils;
SVL	Snout to vent length, measured from the tip of the snout to the vent;
TaL	Tail length, measured from the vent to the tip of the tail;
TaW	Tail width, measured at the base of the tail immediately posterior to the
	post-cloacal swelling.

Scale counts are reported in right and left (R, L) order. The presence, absence and/ or numbers of the following characters were recorded:

EFS	Enlarged femoral scales;
EPrecS	Enlarged precloacal scales;
FP	Femoral pores;
InL	Infralabials, counted as the number of scales from the first lower labial scale immediately posterior to mental to the last scale below posterior edge of the eyeball;
LDRT	Longitudinal dorsal rows of enlarged tubercles, counted as the number of tubercles transversely across the dorsum between ventrolateral folds;
PrecG	Precloacal groove;
PrecP	Precloacal pores;
PostPSR	Post precloacal scale rows;
PostSP	Post cloacal spur;
PVT	Paravertebral tubercles, counted as the number of enlarged tubercles in a straight line between limb insertions left of the vertebral column;
SDLF4	Subdigital lamellae beneath fourth finger, counted as the number of both expanded proximal subdigital lamellae from the base to the largest scale on the digital inflection, and unmodified distal lamellae beneath fourth finger to the claw sheath;
SDLT4	Subdigital lamellae on fourth toe, counted as the number of expanded proximal subdigital lamellae from the base to the largest scale on digital inflection and unmodified distal subdigital lamellae beneath fourth toe to the claw sheath;
SL	Supralabials, counted as the number of scales from the first upper labial scale immediately posterior to rostral to the last scale below posterior edge of the eveball;
VS	Ventral scales, counted as the number of scales transversely across the ventral surface at midbody between ventrolateral folds.

Molecular analyses

Total genomic DNA was extracted from preserved liver tissue of two Phnom Chi specimens (CBC 03003–04) using the DNeasy Blood and Tissue Kit (Qiagen). A 658 bp fragment of mitochondrial (mt) DNA that encodes part of the COI gene was amplified in a 25 ul reaction by the polymerase chain reaction (PCR; 35 cycles of 95° 30s, 53 °C 40s, 72° 90s) and sequenced using the primers VF1d and VR1d (Ivanova et al. 2006). PCR products were cleaned using ExoSAP-IT (Applied Biosystems) and sequenced in both directions by direct double strand cycle sequencing using the Big-Dye Terminator version 3.1 Cycle Sequencing Kit on a 3130 DNA Analyzer (Applied Biosystems). Sequences were edited with Sequencher version 5.4.6 (Gene Codes) and deposited in GenBank under accession numbers MT066405–MT066406. All available *Cyrtodactylus* COI sequences (*n* = 453), and the outgroup *Hemidactylus frenatus* (GenBank accession GQ245970), were downloaded from GenBank on 1 October 2019. The downloaded sequences were aligned and visually inspected in Sequencher to ensure that insertion-deletions did not disrupt the coding region. Preliminary phylogenetic analysis (not shown) was performed on the alignment under the parsimony criterion using a heuristic search with equal weighting of nucleotide substitutions in PAUP* version 4.0a165 (Swofford 2003). Those *Cyrtodactylus* sequences that clustered in the clades around the Phnom Chi samples (= *C. irregularis* group) in a strict consensus of the equally most parsimonious trees were retained in the alignment. Exemplar sequences of other major clades were also retained to represent known phylogenetic diversity within *Cyrtodactylus*, including *C. auribalteatus* (GenBank accession AP018116), *C. badenensis* (KF929505), *C. chanhomeae* (MF169908), *C. condorensis* (MF169910), *C. interdigitalis* (MF169919), *C. intermedius* (MF169920), *C. jellesmae* (MF169923), *C. peguensis* (AP018114), *C. russelli* (MF169938), and *C. thirakhupti* (AP018115).

The resulting pruned COI alignment contained 270 taxa and 717 characters, with no insertion-deletions. The alignment was partitioned by codon position, and the best-fit partitioning scheme and models of sequence evolution were selected using PartitionFinder 2 (Lanfear et al. 2017). Two partitions were selected, with the first and second codon positions merged into a single partition under the model TVM+I+G, and the third codon position under the model GTR+G. Four independent partitioned Bayesian analyses were performed using MrBayes 3.2.7a (Ronquist et al. 2012) on the Cyberinfrastructure for Phylogenetic Research (CIPRES) Science Gateway version 3.3 (Miller et al. 2010). In each analysis, four chains were run for 20 million generations using the default priors, the chain temperature was set to 0.1, trees were sampled every 4,000 generations, and the first 25% of trees were discarded as 'burn-in'. The resulting trace plots were viewed using Tracer v.1.7 (Rambaut et al. 2018). A 50% majority-rule consensus of the post burn-in trees was constructed to calculate the posterior probabilities of nodes. Nodes with posterior probabilities ≥ 0.95 were considered to be statistically supported. Uncorrected pairwise distances were calculated using PAUP* version 4.0a165 (Swofford 2003).

Results

Morphological analyses

The Phnom Chi specimens could not be referred to any other named members of the *C. irregularis* complex owing to having a unique combination of morphological characters. These characters included body size such as having a relatively long body and tibia; scalation, such as the number of subdigital lamellae under the fourth finger and fourth toe, number of longitudinal dorsal and paravertebral rows of tubercles, number of ventral scales, number of enlarged precloacal scales and associated pores (in males) and pits (in females), absence of pores in their enlarged femoral scales, and size of the median subcaudal scale rows from other species in the complex; and pattern and col-



Figure 1. Map illustrating (1) the type locality of *Cyrtodactylus phnomchiensis* sp. nov. at Prey Lang Wildlife Sanctuary, Kampong Thom Province, Cambodia; (2) the type locality of *C. ziegleri* at Chu Yang Sin National Park, Dak Lak Province, Vietnam (Nazarov et al. 2008); (3) the second known locality of *C. ziegleri* at Nam Nung Nature Reserve, Dak Nong Province, Vietnam (Nguyen et al. 2013); and (4) the type locality of *C. bugiamapensis* at Bu Gia Map National Park, Binh Phuoc Province, Vietnam (Nazarov et al. 2012).

oration, including an unbroken nuchal loop bordered anteriorly and posteriorly by a broad yellow or yellowish white band, three or four dark brown body bands, and two or three yellowish white or light brown body bands, about half the width of the brown body bands, and yellow spots on top of the head.

Molecular analyses

The standard deviation of split frequencies among the four Bayesian runs was 0.006260 and the Estimated Sample Sizes (ESS) of parameters were \geq 1,606, indicating that the four runs were sufficiently sampled and had converged. The Phnom Chi specimens represented a distinct mitochondrial lineage that did not match any other named species (Fig. 2). The Phnom Chi lineage was recovered with strong support (Bayesian posterior probability 1.00) to be phylogenetically nested within a clade containing two mitochondrial subclades of *C. ziegleri* (subclades Z1 and Z2; Fig. 2), but the relationships among the Phnom Chi lineage and the two subclades of *C. ziegleri* were unresolved, rendering *C. ziegleri* non-monophyletic (Fig. 2). The clade containing the Phnom Chi lineage and the two subclades of *C. ziegleri* was recovered with strong support (Bayesian posterior probability 1.00) to be sister to *C. bugiamapensis* (Fig. 2).

The Phnom Chi samples had uncorrected *p*-distances in COI of 4.3–6.2% from *C. ziegleri* (all samples) and 7.0–8.6% from *C. bugiamapensis. Cyrtodactylus ziegleri* (all samples) had uncorrected *p*-distances of 6.7–8.5% from *C. bugiamapensis. Cyrtodactylus ziegleri* subclade Z1 had uncorrected *p*-distances of 4.7–5.2% from *C. ziegleri* subclade Z2.

Species description

On the basis of their distinctiveness in morphology and mitochondrial DNA, including from *C. ziegleri* to which they are phylogenetically related (but exact relationship unresolved; Fig. 2), and further corroborated by their geographic distance to any other named members in the complex (and the only member known from west of the Mekong River), the Phnom Chi specimens are hypothesized to represent a distinct species, described herein as:

Cyrtodactylus phnomchiensis sp. nov. http://zoobank.org/103B12F4-6D6F-4928-85A5-A927A81225FE

Figures 3-6

Holotype. CBC 03012, adult male (Fig. 3), Cambodia, Kampong Thom Province, Sandan District, Phnom Chi, Prey Lang Wildlife Sanctuary, 12°56'11.6"N, 105°39'17.1"E, 237 m elevation, collected on 18 July 2019 by Thy Neang and En E.

Paratypes. All from Cambodia, Kampong Thom Province, Sandan District, Phnom Chi, Prey Lang Wildlife Sanctuary: CBC 03003, adult male, 12°56'09.2"N, 105°39'12.7"E, 269 m elevation, coll. 13 June 2019 by Thy Neang; CBC 03004, adult female, 12°56'09.7"N, 105°39'14.4"E, 271 m elevation, coll. 13 June 2019 by Thy Neang; CBC 03013, adult female, same data as holotype; CBC 03014, adult female, same data as holotype except 12°56'08.7"N, 105°39'12.6"E, 284 m elevation.



Figure 2. Upper (**A**) and lower (**B**) portions of a fifty percent majority-rule consensus phylogram resulting from partitioned Bayesian analysis of 717 aligned characters of the mitochondrial cytochrome *c* oxidase subunit I (COI) gene from geckos in the *Cyrtodactylus irregularis* group. The outgroup *Hemidactylus frenatus* (GenBank accession GQ245970) and exemplars of other *Cyrtodactylus* clades including *C. auribalteatus* (GenBank accession AP018116), *C. badenensis* (KF929505), *C. chanhomeae* (MF169908), *C. interdigitalis* (MF169919), *C. intermedius* (MF169920), *C. jellesmae* (MF169923), *C. peguensis* (AP018114), *C. russelli* (MF169938), and *C. thirakhupti* (AP018115) were also included in the analysis (not shown). Black circles at nodes indicate Bayesian posterior probabilities \geq 0.99, and open circles at nodes indicate Bayesian posterior probabilities \geq 0.95. Numbers at terminal tips are GenBank accession numbers.



Figure 2. Continued.



Figure 3. *Cyrtodactylus phnomchiensis* sp. nov. in life. **A** Male holotype CBC 03012 and **B** female paratype CBC 03013.

Etymology. The specific epithet is taken from the type locality of Phnom Chi and the Latin suffix *-ensis* meaning "originating from." The specific epithet is masculine in agreement with the gender of *Cyrtodactylus*.

Diagnosis. *Cyrtodactylus phnomchiensis* sp. nov. is distinguished from the 19 other named species in the *C. irregularis* group (Ngo and Chan 2010; Nguyen et al. 2013, 2017; Pauwels et al. 2018) by having the combination of SVL 76.1–80.7 mm; relatively long body, AG/SVL 0.451–0.481; relatively long tibia, CrusL/SVL 0.172–0.200; subdigital lamellae on fourth finger 18–20; subdigital lamellae on fourth toe 20–23; longitudinal dorsal rows of tubercles 18–20; paravertebral rows of tubercles 31–36; ventral scales 45–54; enlarged femoral scales 0–8, without pores; enlarged precloacal scales 7–10, bearing pores 4 or 5 in males, pits 1–7 in females; precloacal groove absent; median row of transverse subcaudal scales only slightly enlarged; posterior border of nuchal loop unbroken and pointed, bordered anteriorly and posteriorly by broad yellow or yellowish white band; dark brown body bands 3 or 4, the first intact, the second, third and fourth more irregular, alternating with two or three yellowish white or light brown body bands, about half the width of dark brown body bands; and yellow spots on top of head.

Description of holotype. Adult male with SVL 76.1 mm; head slightly elongate, HeadL 22.1 mm, about 30% of SVL, moderately widened, HeadW 14.1 mm, HeadW/ HeadL 0.64, slightly depressed, HeadD 9.4 mm, HeadD/HeadL 0.43, distinct from neck, triangular in dorsal profile; snout rather elongated, rounded in rostral region, ESD 9.0 mm, slightly less than HeadD, ESD/HeadL 0.41, frontonasal region flattened, prefrontal region slightly concave, forming elongated medial rostral groove, canthus rostralis flattened, slightly angled between loreal region and rostral groove; lores posterior to nostrals depressed, anterior to orbit flattened; eye large, eyeball rounded, slightly protruding, EyeD 5.1 mm, shorter than the distance between eye and ear, Eve-EarD 5.7 mm, pupil vertical, covered by crenellate supraciliaries; ear opening oval, deeply sunk, rather small, elongated in oblique position, EarDV 1.2 mm, almost twice longer than its diameter in horizontal position, EarDH 0.7 mm; rostral large, subrectangular, height 1.9 mm, shorter than its width 3.6 mm, medially divided dorsally by a suture, reaching to about half way of rostral height, in contact with 1st SL and nostrils laterally, supranasals and internasal dorsally (Fig. 4); nostrils pieced at anterior angle of snout, directed lateroposteriorly, surrounded by rostral anteriorly, 1st SL ventrally, supranasals dorsally, and three small postnasal scales; internarial distance narrow, IND 2.9 mm; supranasals subrectangular, separated by intersupranasal, slightly smaller in size, in contact with rostral anteriorly, nostrals laterally, four small scales posteriorly; intersupranasal single, subpentagonal, slightly protruding rostral, in contact with two small scales posteriorly; interorbital rather narrow, IOD 5.5 mm, longer than EyeD, slightly shorter than Eye-EarD 5.7 mm; supralabials (12R, 13L), subrectangular anteriorly, circular shape posteriorly, anterior SL separated from small scales on loreal region by row of slightly enlarged scales; infralabials (9R, 9L), larger than SL, first InL bordered by mental anteriorly, first postmental ventrally, second InL bordered by second enlarged postmental, enlarged chin shield scale ventrally, 3-7th InL bordered by a row of slightly enlarged chin shield scales ventrally; mental large, triangular, width 3.3 mm in width, 2.3 mm in length, in contact with first InL laterally, two pairs postmentals posteriorly; the first pair largest, subrectangular, in broad contact medially, second pair enlarged, half the size of the first pair, separated by four smaller gular scales medially, in contact with smaller scales posteriorly (Fig. 4). Scales on frontonasal, prefrontal, loreal



Figure 4. Head of male holotype CBC 03012 of *Cyrtodactylus phnomchiensis* sp. nov. in preservative. **A** Dorsal view illustrating the rostral, supranasal and internsupranasal scales **B** ventral view illustrating the mental and postmental scales.

regions small, almost homogenous, slightly larger than those on top of head; scales on occiput intermixed with scattered larger, more rounded, conical tubercles, more prominent tubercles on region between orbit and area above ear opening, a noticeably larger tubercle (compared to those surrounding) at the corner of jaw.

Body slightly slender, AG 36.6 mm, nearly half SVL, AG/SVL 0.481 with welldefined narrow vertebral furrow posteriorly; scales on dorsum small, mostly homogenous, granular, interspersed with larger, low, weakly keeled, irregularly arranged, tubercles; longitudinal dorsal rows of enlarged tubercles approximately 18; paravertebral tubercles 32; tubercles on nape within dark brown nuchal loop, anterior dorsal surface at level above shoulder smaller, more rounded, sparser than those on mid-dorsum and posterior dorsal surface, more prominent, being denser, weekly keeled, more regularly arranged on sacral and tail base region; tubercles on lateral body sparsely; ventral scales small, not imbricate, those near midline larger than lateral and dorsal scales; scales on throat and gular region the smallest; faint ventrolateral folds with few emerged tubercles; ventral scales at midbody between ventrolateral folds 47; precloacal region moderately enlarged, a few rows of enlarged precloacal non-pore bearing scales anterior to pore bearing precloacal scales; enlarged precloacal scales 7, in angular series, bearing 5 pores, terminal scale on each side poreless; post precloacal scale rows 3, the first row immediately posterior to enlarged precloacal pore-bearing scales with six scales in angular series, the second row with four scales in angular series, the third row with three scale in straight line, the medial scale largest; femoral scales slightly enlarged (8R, 8L), distal scales more than twice the size of proximal scales, all smaller than those of pore-bearing precloacal scales, separated from precloacal scales by diastema; precloacal groove absent; fully everted hemipenes thick, 5.9 mm in length, two penes at each sheath, two sockets posterior to hemipenal bases (Fig. 5).

Limbs rather slender; digits with strongly inflected interphalangeal joints; forelimbs bearing five relatively slender fingers, moderately bowed, ending with curved claws, ForeL/SVL 0.162; expanded proximal subdigital lamellae on fourth finger 6, unmodified distal subdigital lamellae on fourth finger 12, total subdigital lamellae on fourth finger 18; hind limbs bearing five relatively slender toes, strongly bent, ending with curved claws, CrusL/SVL 0.176; expanded proximal subdigital lamellae on fourth toe 7, unmodified distal subdigital lamellae on fourth toe 15, total subdigital lamellae on fourth toe 22; all digits lacking scansorial setae on ventral surface; scales on limbs small, interspersed with larger, low, conical, weakly keeled tubercles; scales on palmar and plantar surfaces small.

Tail moderately wide anteriorly, TaW 5.5 mm, segmented, cylindrical, becoming slender toward tip, regenerated posteriorly; dorsal caudal longitudinal tubercle rows at base of tail 8; 2 transverse rows of dorsal caudal tubercles at posterior margin of third band on tail, 22.7 mm from tail base; vertebral caudal surface with scattered bump at approximate intervals of 3 mm; subcaudal scale rows smooth, small, differing in size and irregular in shape, usually alternating between a single slightly enlarged and two smaller scales, 2 or 3 times larger than neighboring lateral caudal scales (Fig. 5).



Figure 5. Cloacal region of male holotype CBC 03012 of *Cyrtodactylus phnomchiensis* sp. nov. in preservative illustrating the enlarged precloacal scales, enlarged femoral scales, and not enlarged subcaudal scales.

Color of holotype in life (Fig. 3). Dorsal surface, nape, and tail yellowish white to light brown; top of head with yellowish spots; interorbital region, rostral and loreal regions lighter brown with scattered yellowish scales; eye ring yellowish; rostral, mental lighter brown; supralabials, corner of jaw, and region extending through dorsal margin of ear opening to shoulder yellowish; nuchal loop with large dark brown band, pointed, extending between posterior margins of eyes, bordered anteriorly by broad yellow band

along upper edge of dark brown nuchal loop, posteriorly by yellowish white band; three dorsal dark brown bands on body, the first more regular, the second and third bands irregular, interrupted by white irregular blotches; all dark brown bands ending near to mid-flank region, bordered below by lighter brown extending to lateral folds; dark brown body bands bordered by yellowish white or light brown bands about half the width of dark brown body bands, the last light brown band ending on tail base; anterior and posterior margins of body bands with darker brown coloration; dark brown bands 6 on regenerated tail, margins at tail base darker; white bands on tail 5, nearly encircling the tail except subcaudal scale row; subcaudal scales lighter brown than dorsal caudal scales; limbs lighter brown with orangish or yellowish on enlarged tubercles; ventral surfaces between ventrolateral folds, chin, throat, and limbs white with tiny black dots on tip of scales; ventral surfaces of fingers and toes dark brown. In preservative, all yellowish, yellowish white or orange coloration faded to white, cream, or light brown (Fig. 6).

Variations. Morphometric and meristic characters of the type series are presented in Table 1. The paratypes generally resemble the holotype (Fig. 6), except as follows. CBC 03003 has four enlarged post precloacal scale rows in an angular series, the last row with only a single enlarged scale. CBC 03004 and CBC 03014 have darker brown body bands. CBC 03003–03004 have a more pointed nuchal loop. CBC 03003 has more dense dark dots causing ventral surfaces to be darker brown. CBC 03003 has more slender, fully everted hemipenes. Females have enlarged precloacal scales with pits rather than pores.

Distribution and natural history. The new species is known only from the type locality at Phnom Chi in Prey Lang Wildlife Sanctuary, Kampong Thom Province, Sandan District, Cambodia. All individuals were found at night between 2001–2147 hr in evergreen-large dipterocarp dominated forest associated with rocky terrain (Fig. 7). The holotype CBC 03012 was found on a rock face following evening rain, paratypes CBC 03013–14 were on boulders following evening rain, paratype CBC 03003 was on leaf litter along a forest trail, and paratype CBC 03004 was on a rock wall at the entrance to a cave. Only five individuals were found during five-survey nights, suggesting the species is relatively uncommon. None were encountered during a brief survey by NT in the wet season of 2014 (Hayes et al. 2015). The new species is the only member of the C. *irregularis* complex known to occur west of the Mekong River (Nguyen et al. 2017; Pauwels et al. 2018).

Comparisons. *Cyrtodactylus phnomchiensis* sp. nov. is distinguishable from all 19 other members of the *C. irregularis* group by a unique combination of morphological characters (and in mitochondrial DNA; Fig. 2).

Cyrtodactylus phnomchiensis sp. nov. differs from *C. bidoupimontis* Nazarov, Poyarkov, Orlov, Phung, Nguyen, Hoang & Ziegler, 2012 by having ventral scales 45–54 (vs. 38–43 in *bidoupimontis*), precloacal pits in females 1–7 (vs. absent in *bidoupimontis*), dark brown body bands larger than yellowish white or light brown dorsal bands (vs. dark brown bands, when present, narrower than light yellow dorsal bands in *bidoupimontis*), and distinct large yellow band on anterior margin of dark brown nuchal loop (vs. narrow light margin in *bidoupimontis*), and yellow spots on top of head (vs. dark spots in *bidoupimontis*).

Voucher specimen	CBC 03012	CBC 03003	CBC 03004	CBC 03013	CBC 03014	Range
Type status	Holotype	Paratype	Paratype	Paratype	Paratype	
Sex	Male	Male	Female	Female	Female	
SVL	76.1	79.0	77.3	80.7	76.7	76.1-80.7
TaL*	75.1	56.9	66.6	79.1	64.6	56.9-79.1
TaW	5.5	4.4	5.3	5.6	5.7	4.4–5.7
TaW/SVL	0.072	0.056	0.069	0.069	0.074	0.056-0.74
ForeL	12.3	13.2	12.1	13.7	11.7	11.7–13.7
ForeL/SVL	0.162	0.167	0.157	0.170	0.153	0.153-0.170
CrusL	13.4	14.9	13.8	16.1	13.2	14.2–16.6
Crus/SVL	0.176	0.189	0.179	0.200	0.172	0.172-0.200
AG	36.6	36.1	35.3	36.4	35.4	35.3–36.6
AG/SVL	0.481	0.457	0.457	0.451	0.462	0.451-0.481
HeadL	22.1	23.5	22.2	23.6	23.4	22.1-23.6
HeadL/SVL	0.290	0.297	0.287	0.292	0.305	0.287-0.305
HeadW	14.1	14.5	13.7	15.2	13.7	13.7–15.2
HeadD	9.4	9.2	8.6	9.8	8.6	8.6–9.8
EyeD (eye diameter)	5.1	5.1	4.8	4.8	4.5	4.5-5.1
EyeD/SVL	0.067	0.065	0.062	0.059	0.059	0.059-0.067
Ear-EyeD (eye-ear	5.7	6.1	6.0	6.5	5.7	5.7-6.5
distance)						
ESD (eye-snout distance)	9.0	9.5	9.0	9.9	9.3	9.0–9.9
ESD/SVL	0.118	0.120	0.116	0.123	0.121	0.116-0.123
END (eye-nostrial	6.6	6.9	6.4	7.0	7.0	6.4–7.0
distance)						
IO (interorbital distance)	5.5	4.8	5.0	5.7	5.2	4.8–5.7
IND (internarial distance)	2.9	2.8	2.6	2.9	2.7	2.6-2.9
EarDV (vertical)	1.2	1.3	1.2	1.3	1.3	1.2–1.3
EarDH (horizontal)	0.7	1.1	0.7	0.7	0.7	0.6-1.1
Intersupranasal scales	1	1	1	1	1	1
Supralabials (SL)	12R/13L	11R/11L	11R/11L	12R/12L	12R/13L	11–13
Infralabials (InL)	9R/9L	10R/9L	10R/10L	10R/9L	10R/10L	8-10
PVT	32	31	36	34	32	31–36
LDRT	18	20	20	20	19	18-20
VS	47	47	52	45	54	45–54
Median subcaudal scales	yes	yes	yes	yes	yes	yes
slightly enlarged						
SDLF4	18	20	18	19	19	18-20
SDE14	22	23	20	21	21	20-23
EFS	8R8L	3R/3L	0	7R/6L	0	0-8
FP	0	0	0	0	0	0
EPrecS	7	9	7	10	9	7–10
PrecP	5	4	4	1	7	1–7
PrecG	0	0	0	0	0	0
PostPSR	3	4	3	3	3	3-4
PostSP	4	3	4	4	3	3-4
Number of dark brown	3	3	4	3	3	3-4
body bands						
Femoral and precloacal	no	no	no	no	no	no
scales continuous						
reliowish spots on top	yes	yes	yes	yes	yes	yes
or nead						

Table 1. Mensural, meristic and color pattern characters of *Cyrtodactylus phnomchiensis* sp. nov. Abbreviations defined in the text. All specimens have regenerated portions of tails (*).

Voucher specimen	CBC 03012	CBC 03003	CBC 03004	CBC 03013	CBC 03014	Range
Posterior border of nuchal	yes	yes	yes	yes	yes	yes
loop pointed						
First body band complete	yes	yes	yes	yes	yes	yes
Second to fourth body	yes	yes	yes	yes	yes	yes
bands more irregular						
Yellowish white or light	yes	yes	yes	yes	yes	yes
brown bands about half						
the width of dark brown						
body bands						
Number of yellowish	2	2	3	2	2	2-3
white or light brown body						
bands						
Yellowish spot above ear	yes	yes	yes	yes	yes	yes
opening						
Enlarged tubercle at	yes	Yes	yes	yes	yes	yes
corner of jaw						

Cyrtodactylus phnomchiensis sp. nov. differs from *C. buchardi* by having SVL 76.1–80.7 mm (vs. 60–65 mm in *buchardi*), SDLF4 18–20 (vs. 14 in *buchardi*), SDLT4 20–23 (vs. 12–14 in *buchardi*), ventral scales 45–54 (vs. 30 in *buchardi*), LDRT 18–20 (vs. 25 in *buchardi*), precloacal pores in males 4–5 (vs. 9 in *buchardi*), and irregular dorsal body bands (vs. blotches in *buchardi*).

Cyrtodactylus phnomchiensis sp. nov. differs from *C. bugiamapensis* Nazarov, Poyarkov, Orlov, Phung, Nguyen, Hoang & Ziegler, 2012 by having LDRT 18–20 (vs. 20–24 in *bugiamapensis*), ventral scales 45–54 (vs. 36–46 in *bugiamapensis*), precloacal pores in males 4–5 (vs. 7–11 in *bugiamapensis*), SDLF4 18–20 (vs. 15–17 in *bugiamapensis*), SDLT4 20–23 (vs. 17–20 in *bugiamapensis*), CrusL/SVL in adult specimens 0.172–0.200 (vs. 0.144–0.157 in *bugiamapensis*), large nuchal loop bordered anteriorly and posteriorly by broad yellow bands (vs. narrow nuchal loop bordered by distinct narrow white lines in *bugiamapensis*), dark brown body bands 3–4 (vs. seven highly irregular dark blotches with light margins in *bugiamapensis*), and top of head with yellowish spots (vs. distinct dark brown spots in *bugiamapensis*).

Cyrtodactylus phnomchiensis sp. nov. differs from *C. caovansungi* Orlov, Nguyen, Nazarov, Ananjeva & Nguyen, 2007 by having SVL 76.1–80.7 mm (vs. 90.4–94.0 mm in *caovansungi*), ventral scales 45–54 (vs. 38–44 in *caovansungi*), femoral pores absent (vs. 6 in *caovansungi*), precloacal pores in males 4–5 (vs. 9 in *caovansungi*), SDLF4 18–20 (vs. 22 in *caovansungi*), and enlarged subcaudals absent (vs. present in *caovansungi*).

Cyrtodactylus phnomchiensis sp. nov. differs from *C. cattienensis* Geissler, Nazarov, Orlov, Böhme, Phung, Nguyen & Ziegler, 2009 by having SVL 76.1–80.7 mm (vs. 69.0 mm maximum in *cattienensis*), ventral scales 45–54 (vs. 28–42 in *cattienensis*), SDLF4 18–20 (vs. 12–16 in *cattienensis*), and SDLT4 20–23 (vs. 14–19 in *cattienensis*).

Cyrtodactylus phnomchiensis sp. nov. differs from *C. cucdongensis* Schneider, Phung, Le, Nguyen & Ziegler, 2014 by having SVL 76.1–80.7 mm (vs. 55.8–65.9 mm in *cucdongensis*), ventral scales 45–54 (vs. 35–44 in *cucdongensis*), SDLF4 18–20 (vs. 13–18 in *cucdongensis*), and SDLT4 20–23 (vs. 15–20 in *cucdongensis*).



Figure 6. Dorsal view of the type series of *Cyrtodactylus phnomchiensis* sp. nov. in preservative.

Cyrtodactylus phnomchiensis sp. nov. differs from *C. cryptus* Heidrich, Rösler, Vu, Böhme & Ziegler, 2007 by having precloacal pores in males 4–5 (vs. 9–11 in *cryptus*), and precloacal pits in females 1–7 (vs. absent in *cryptus*).

Cyrtodactylus phnomchiensis sp. nov. differs from *C. dati* Ngo, 2013 by having SVL 76.1–80.7 mm (vs. 70.1 mm maximum in *dati*), regenerated TaL 56.9–79.1 mm vs. (vs. 50.3 mm maximum, non-regenerated TaL in *dati*), femoral pores in both sexes absent (vs. present in *dati*), nuchal loop continuous (vs. broken in *dati*), and dark brown body bands (vs. irregular dark brown blotches on body in *dati*).

Cyrtodactylus phnomchiensis sp. nov. differs from *C. gialaiensis* Luu, Dung, Nguyen, Le & Ziegler, 2017 by having SVL 76.1–80.7 mm (vs. 62.8 mm maximum in *gialaiensis*), precloacal pores in males 4–5 (vs. 9–10 in *gialaiensis*), SDLF4 18–20 (vs. 14–15 in *gialaiensis*), and SDLT4 20–23 (vs. 15–17 in *gialaiensis*).

Cyrtodactylus phnomchiensis sp. nov. differs from *C. huynhi* Ngo & Bauer, 2008 by having SDLF4 18–20 (vs. 14–17 in *huynhi*), AGL/SVL 0.451–0.481 (vs. 0.370–0.428 in *huynhi*), ventral scales 45–54 (vs. 43–46 in *huynhi*), precloacal pores in males 4–5 (vs. 7–9 in *huynhi*), dark brown body bands 3–4 (vs. 5–6 in *huynhi*); femoral pores in both sexes absent (vs. 3–8 in *huynhi*), and nuchal loop bordered anteriorly and posteriorly by broad yellow bands (vs. narrow cream margin in *huynhi*).

Cyrtodactylus phnomchiensis sp. nov. differs from *C. irregularis* by lacking enlarged triangular tubercles at base of tail (vs. present in *irregularis*), CrusL/SVL 0.172–0.200 (vs. 0.138–0.156 in *irregularis*), LDRT 18–20 (vs. 22–24 in *irregularis*), paravertebral tubercles 31–36 (vs. 38–48 in *irregularis*); ventral scales 45–54 (vs. 38–45 in *irregularis*), SDLF4 18–20 (vs. 15–16 in *irregularis*), SDLT4 20–23 (vs. 18–20 in *irregularis*), dark brown body bands 3–4 (vs. 5–7, mostly as irregular blotches in *irregularis*), and yellowish spots on top of head (vs. distinct dark brown spots in *irregularis*).

Cyrtodactylus phnomchiensis sp. nov. differs from *C. kingsadai* Ziegler, Phung, Le & Nguyen, 2013 by having SVL 76.1–80.7 mm (vs. 83.0–94.0 mm in *kingsadai*), enlarged femoral scales 0–8 (vs. 9–12 in *kingsadai*), precloacal pore in males 4–5 (vs. 7–9 in *kingsadai*), and subcaudal scales not enlarged (vs. enlarged in *kingsadai*).

Cyrtodactylus phnomchiensis sp. nov. differs from *C. phuocbinhensis* Nguyen, Le, Tran, Orlov, Lathrop, MacCulloch, Le, Jin, Nguyen, Nguyen, Hoang, Che, Murphy & Zhang, 2013 by having SVL 76.1–80.7 mm (vs. 46.0–60.4 in *phuocbinhensis*); precloacal pits in females 1–7 (vs. absent in *phuocbinhensis*), top of head with yellow spots (vs. dark brown spots in *phuocbinhensis*), and dark brown body bands (vs. two dark brown longitudinal stripes or blotches in *phuocbinhensis*).

Cyrtodactylus phnomchiensis sp. nov. differs from *C. pseudoquadrivirgatus* Rösler, Vu, Nguyen, Ngo & Ziegler, 2008 by having yellow spots on top of head (vs. dark blotches on top of head in *pseudoquadrivirgatus*) and dark brown body bands (vs. highly irregular body blotches in *pseudoquadrivirgatus*).

Cyrtodactylus phnomchiensis sp. nov. differs from *C. sangi* Pauwels, Nazarov, Bobrov & Poyarkov, 2018 by having SVL 76.1–80.7 mm (vs. 56.3 mm maximum in *sangi*), paravertebral tubercles 31–36 (vs. 27–29 in *sangi*), ventral scales 45–54 (vs. 37 in *sangi*), precloacal pores in males 4–5 (vs. 7 in *sangi*), and first dark brown body band complete, second, third, and fourth more irregular (vs. highly irregular bands in *sangi*).



Figure 7. Habitat at Phnom Chi, the type locality of Cyrtodactylus phnomchiensis sp. nov.

Cyrtodactylus phnomchiensis sp. nov. differs from *C. takouensis* Ngo & Bauer, 2008 by having LDRT 18–20 (vs. 9–10 smooth tubercles in *takouensis*); ventral scales 45–54 (vs. 39–40 in *takouensis*), SDLF4 18–20 (vs. 16–17 in *takouensis*), SDLT4 20–23 (vs. 18–20 in *takouensis*), and dark brown canthal stripe absent (vs. present in *takouensis*).

Cyrtodactylus phnomchiensis sp. nov. differs from *C. taynguyenensis* Nguyen, Le, Tran, Orlov, Lathrop, MacCulloch, Le, Jin, Nguyen, Nguyen, Hoang, Che, Murphy & Zhang, 2013 by having supralabials 11–13 (vs. 8–9 in *taynguyenensis*), precloacal pits in females present (vs. absent in *taynguyenensis*), SDLF 18–20 (vs. 13–18 in *taynguyenensis*), top of head with yellow spots (vs. dark brown blotches in *taynguyenensis*), and dark brown body bands (vs. black irregular blotches margined by light brown in *taynguyenensis*).

Cyrtodactylus phnomchiensis sp. nov. differs from *C. yangbayensis* Ngo & Chan, 2010 by having SDLT4 20–23 (vs. 15–17 in *yangbayensis*) and lacking enlarged subcaudal scales (vs. present in *yangbayensis*).

Cyrtodactylus phnomchiensis sp. nov. is most closely related in mitochondrial DNA to *C. ziegleri* Nazarov, Orlov, Nguyen & Ho, 2008 (Fig. 2), but differs in morphology from *C. ziegleri* by having SVL 76.1–80.7 mm (vs. 84.6–93.0 mm in *ziegleri*), paravertebral tubercles 31–36 (vs. 38–46 in *ziegleri*), ventral scales 45–54 (vs. 33–45 in *ziegleri*), HeadL/SVL 0.287–0.305 (vs. 0.263–0.284 in *ziegleri*), ESD/SVL 0.116–0.123 (vs. 0.103–0.113 in *ziegleri*), CrusL/SVL 0.172–0.200 (vs. 0.140–0.168 in *ziegleri*), AG/SVL 0.451–0.481 (vs. 0.390–0.444 in *ziegleri*), eyeD/SVL 0.059–0.067 (vs. 0.053–0.057 in *ziegleri*), top of head with yellow spots (vs. dark brown spots in

ziegleri), large dark brown nuchal loop (vs. narrow in *ziegleri*), distinct, broad yellow band on anterior margin of dark brown nuchal loop (vs. absent in *ziegleri*), and dark brown body bands bordered by yellowish white or light brown bands about half the width of dark brown bands (vs. light yellow or light brown body bands about same width as dark brown body bands in *ziegleri*).

Discussion

Mitochondrial DNA serves as a useful but imperfect tool for delimiting species within the C. irregularis complex (Nguyen et al. 2017; Pauwels et al. 2018). Two divergent mitochondrial lineages occur within C. ziegleri, with both lineages found at its type locality of Chu Yang Sin National Park, Dak Lak Province, and one lineage at Nam Nung Nature Reserve, Dak Nong Province, central Vietnam (Figs 1, 2; Nguyen et al. 2013, 2017; Ziegler et al. 2013; Schneider et al. 2014; Pauwels et al. 2018). Thus, from a matrilineal perspective, some individuals of C. ziegleri at Chu Yang Sin are more closely related to those at Nam Nung than to other individuals at Chu Yang Sin. Likewise, two divergent mitochondrial lineages occur within C. cattienensis (Fig. 2; Nazarov et al. 2012; Nguyen et al. 2013, 2014, 2017; Schneider et al. 2014; Pauwels et al. 2018), and both lineages of C. cattienensis can be found in sympatry at Ta Kou Mountain, Binh Thuan Province, southern Vietnam (Nguyen et al. 2017). The mitochondrial divergences within C. ziegleri and C. cattienensis are uncorroborated by divergences in morphology (or in one or two nuclear markers; Nguyen et al. 2017; Pauwels et al. 2018), and therefore these each continue to each be treated as single species that harbor considerable intraspecific mitochondrial DNA variation (Nguyen et al. 2017; Pauwels et al. 2018). The processes that resulted in the formation of these divergent mitochondrial lineages within C. ziegleri and C. cattienensis are unknown, but might be a consequence of a period of past separation of populations by geological or climatic events, the subsequent accumulation of mutations in the mitochondrial genome during isolation, and eventually recontact of these separated populations that today exhibit homogeneous morphology but persistent, divergent mitochondrial genomes (i.e., ancestral polymorphism). Similarly, Nguyen et al. (2017) preferred an explanation of allopatric divergence and subsequent migration into sympatry, rather than sympatric divergence, to explain the co-occurrence of the two mitochondrial lineages of C. cattienensis at Ta Kou Mountain. Importantly for this study, the divergent and unique mitochondrial lineage of C. phnomchiensis sp. nov. is corroborated by a divergence in morphology from C. ziegleri and all other members of the C. irregularis complex, and so we posit that *C. phnomchiensis* sp. nov. should be recognized as a distinct species.

Unfortunately, our phylogenetic analysis of the COI gene does not resolve the relationships among *C. phnomchiensis* sp. nov. and the two subclades of *C. ziegleri* (subclades Z1 and Z2; Fig. 2). This means that the resulting polytomy renders *C. ziegleri* as non-monophyletic in our analysis (Fig. 2). This polytomy could be a consequence of a near-simultaneous divergence among these three lineages (i.e., short internodes), but is more likely to be a result of insufficient molecular data (Brennan et al. 2017). Unfortunately, most members of the *C. irregularis* complex have been represented by mitochondrial DNA in previous studies only with <700 bp of the COI gene (Brennan et al. 2017), and hence we were limited in our analyses by available comparative sequence data. Additional sequence data for members of this complex are needed. Specifically, additional mitochondrial data may resolve the polytomy found here among the two subclades of *C. ziegleri* and *C. phnomchiensis* sp. nov., and additional nuclear data can be used to test species boundaries that have been hypothesized from morphological and mitochondrial data (one or two nuclear markers have provided some, but limited, phylogenetic utility; Nguyen et al. 2017; Pauwels et al. 2018).

Phnom Chi consists of an isolated small rocky mountain (peak of 652 m elevation) and a few associated smaller hills, altogether encompassing an area of approximately 4,464 ha within the Prey Lang Wildlife Sanctuary in Kampong Thom and Kratie provinces, Cambodia. The base and lower elevations of Phnom Chi have dry and mixed deciduous forest, whereas upper elevations have large dipterocarp-dominated evergreen and semi-evergreen forest. The current habitat remains in relatively good condition, but this long-overlooked site needs urgent conservation attention. Local communities utilize Phnom Chi for resource extraction, notably the tapping of liquid resin from large dipterocarp trees on the mountain, and small-scale, illegal gold extraction around the base, in addition to forest burning during the dry season (possibly by resin tappers). A small pagoda at the base of the mountain and the scenic beauty of the area (Fig. 7) attracts local and domestic tourists, and will likely attract international tourists in the near future. A second species of lizard, the scincid Sphenomorphus preylangensis Grismer, Wood, Quah, Anuar, Poyarkov, Neang, Orlov, Thammachoti & Hun, 2019, was also recently described from Phnom Chi. Phnom Chi is the only feature with any significant topographic relief in Prey Lang Wildlife Sanctuary, other than some isolated limestone karst blocks in the northern section that have not yielded Cyrtodactylus during field surveys (TN, unpublished data). As such, C. phnomchiensis sp. nov. may be endemic to the immediate vicinity of Phnom Chi, and together with S. preylangensis, underscores the importance of the area for biodiversity conservation. Due to having a small area of occupancy, being relatively uncommon, and experiencing ongoing conservation threats, an assessment of C. phnomchiensis sp. nov. by the IUCN Red List of Threatened Species (IUCN 2020) is urgently warranted.

Species diversity of *Cyrtodactylus* in Cambodia is likely to be significantly underestimated. Recently, five species were described within the *C. intermedius* complex from the Cardamom Mountains of southwestern Cambodia (Murdoch et al. 2019), and an additional undescribed species in this complex has been reported from northern Cambodia near the Thai border (Geissler et al. 2019). The species diversity of the *C. irregularis* complex in Cambodia is even less known. Stuart et al. (2006) and Nazarov et al. (2008, 2012) referred to unstudied specimens in the *C. irregularis* complex from Mondolkiri and Ratanakiri Provinces in hilly eastern Cambodia, and those from Ratanakiri Province were later referred by Stuart et al. (2010) to *C. pseudoquadrivirgatus*. The rapid rate of taxonomic partitioning within the *C. irregularis* complex during the past decade, underscored by the realization that many of these newly-recognized species have very narrow geographic ranges, suggests that re-evaluation of the species identities of the Mondolkiri and Ratanakiri specimens is needed.

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