



Utilizing regulatory T cells against rheumatoid arthritis

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Regulatory T (Treg) cells are essential for normal immune surveillance systems, and their dysfunction leads to development of diseases, such as autoimmune disorders. CD4⁺CD25⁺ Treg cells are well-known suppressive cells, which express the transcription factor Foxp3, are indispensable for the maintenance of immune self-tolerance and homeostasis by suppressing aberrant or excessive immune response. Other Foxp3⁻ Treg cells include Tr1, Th3, CD8⁺CD28^{-/-}, and Qa1-restricted T cells; however, the contribution of these Treg cells to self-tolerance, immune homeostasis as well as preventing autoimmunity is not well defined. Here, we discuss the phenotypes and function of Foxp3⁺ Treg cells and the potential use of such Treg cells against rheumatoid arthritis (RA). Of note, even though most expanded populations of Foxp3⁺ Treg cells exhibit suppressive activity, tissue-associated or antigen-specific Treg cells appear superior in suppressing local autoimmune disorders such as RA. In addition, utilizing tissue-associated Foxp3⁺ Treg cells from stem cells may stable Foxp3 expression and avoid induction of a potentially detrimental systemic immunosuppression.

Keywords: regulatory T cells, autoimmunity, rheumatoid arthritis, cell-based therapies, suppressive cytokines, stem cells

INTRODUCTION

Rheumatoid arthritis (RA) is a chronic autoimmune disorder, which has approximately 1% prevalence in the industrialized world, and is described by pain and stiffness of the joints, and inflammation of the synovial membrane. This systematic autoimmune disorder results in the accumulation of inflammatory cells such as T cells, B cells, and macrophages in the inflamed joints, which can lead to persistent synovitis and tissue destruction, typically articular cartilage destruction. Being a disabling and painful condition, RA often causes loss of mobility and function, and is commonly accompanied by substantial comorbid conditions in the cardiovascular, neurologic, and metabolic systems (1). The autoantigens are still unidentified in spite that RA is considered as autoimmune disease. The main cause of autoimmune disorder like RA is the failure in the maintenance of immunological self-tolerance. There are multiple mechanisms for maintaining the self-tolerance within the immune system. It is considered that autoreactive T cells and B cells are vital for the pathogenesis of RA. Autoreactive T cells are mainly deleted in the thymus; however, this process is not strict. Nevertheless, autoreactive T cells can escape into the peripheral immune system, and subsequent activation will cause autoimmune pathological disorders (2).

The etiological factor for the development of autoimmune disorder remains unclear; however, the progression of the RA disease is associated with synovial inflammation, which can develop to a Pannus (thickening synovial tissue) and damage the surrounding cartilage and bone. Plasmatic cells infiltrate into the joint and produce antibodies that form aggregates of IgG. In order, the immune system recognizes these IgG aggregates as foreign antigens. Within the synovial membrane, plasmatic cells, T cells, B cells, dendritic cells (DCs), and macrophages develop lymphoid follicle-like constructions, and start to create antibodies against

these constructions recognized as rheumatoid factor RF (3). The IgM class is the most significant type of RF in regards to RA that emerges in 70–80% of patients. During the development of the disease, a wide variety of cells, including B cells, macrophages, DCs, neutrophils, fibroblasts, and granulocytes profoundly infiltrate into the normal and relatively avascular synovium. However, the concept for the pathogenic events that are critical for the triggers for the onset of diseases remains undetermined. In general, RA is induced by interactions of multiple factors, including genetic, biomechanical, and environmental factors, neuro-immune interactions, and impaired articular microvascular function (4). Up to now, by using the genome-wide association studies, a number of genetic loci have been suggested to be associated with RA susceptibility and severity (3, 5, 6).

The immune system can defend against pathogenic attack, and is responsible for maintaining effective immune response as well as promoting an adequate inflammatory response. Regulatory T (Treg) cells, formerly known as suppressor T cells, are a developmentally and functionally distinct T cell subpopulation that modulates the immune system, retains tolerance to self-antigens, and eliminates autoimmunity. CD4⁺CD25⁺ suppressor T cells are well-known Treg cells that express the transcription factor forkhead box P3 (Foxp3), are indispensable for the maintenance of immune self-tolerance and homeostasis by suppressing aberrant or excessive immune response. Other Foxp3⁻ suppressor T cells include Tr1, Th3, CD8⁺CD28^{-/-}, and Qa1-restricted T cells; however, the contribution of these Treg cells to self-tolerance, immune homeostasis as well as preventing autoimmunity is not well defined. The main functions of Foxp3⁺ Treg cells are to migrate into inflammation sites and suppress various effector lymphocytes, especially the subsets of CD4⁺ helper T (Th) cells: Th1, Th2, Th17, and follicular Th cells. Over the past decade, a number

of studies have addressed that the majority of the Foxp3⁺ Treg cells are produced in the thymus as an antigen-primed and functionally mature T cell subpopulation specialized for immune suppression. However, some of Foxp3⁺ Treg cells differentiate from naive conventional T cells in the periphery under certain condition (7). The characteristics of different Treg cell subsets are briefly described in **Table 1**.

CD4⁺CD25⁺ Treg CELL FUNCTION IN RHEUMATIC DISEASE

Thymic selection results in the appearance of T cells with two types of T cell receptor (TCR). The majority express $\alpha\beta$ chains in the TCR, forming unique structure on each T cell. $\alpha\beta$ T cells represent mature T cells that circulate through the secondary lymphoid organs and develop adaptive immune response. Some other fraction of T cells express $\gamma\delta$ chains in TCR, reside in skin and mucosal surfaces that play a role in the initial response to microbial invasion. On the basis of lineage marker and functional activities, $\alpha\beta$ T cells are subdivided into several groups, including CD4⁺ and CD8⁺, or naive, effector, and memory T cells. Naive T cells are the most homogenous representative of CD4⁺ and CD8⁺ subsets. The activated CD4⁺ T cells can be subdivided into Th1, Th2, Th17, and Treg cell subsets based on production of signature cytokines.

There are two main classes of CD4⁺CD25⁺ Treg cells: naturally occurring Treg (nTreg) cells and induced or adaptive Treg (iTreg) cells. nTreg cells are CD4⁺CD25⁺Foxp3⁺ Treg cells that develop in the thymus. iTreg cells are induced Foxp3⁺ Treg cells converted from Foxp3⁻ T effectors (Teffs) (8). There are multiple signaling pathways that are involved in the activation and control of T cell responses. There are no unique surface markers for Treg cells; however, Treg cells are usually distinguished by the expression of CD25 (the alpha chain of the IL-2 receptor), CTLA-4 (CD152, cytotoxic T-lymphocyte antigen 4), and glucocorticoid-induced tumor-necrosis-factor-receptor-related protein (GITR).

These proteins are expressed not only on Treg cells but also on activated T cells that make complications for their use as markers of Treg cells. It remains unclear how the Treg cells are generated and the mechanisms that relate to their function. Nevertheless, CTLA-4 and TGF- β are critical in activity of Treg cells. CTLA-4^{-/-} or TGF- β ^{-/-} mice have a more extensive pathology than those lacking CD4⁺CD25⁺ Treg cells, although such Treg cells can function in the absence of TGF- β production or signaling. In general, the signaling mechanisms by which Treg cells are developed and exert their suppressive function still need to be determined.

The transcription factor Foxp3 is highly expressed in lymphoid tissues such as CD4⁺ T cells; however, approximately undetectable in B cells and CD8⁺ T cells. Foxp3 is similar to other genes such as CTLA-4, OX40, and GITR, is a gene associated with cell activation and the Treg lineage. Foxp3 is not expressed in CD4⁺CD25⁻ Th cells upon stimulation. Foxp3 is the most specific gene for the lineage of Treg cells as compared with other associated genes like GITR or CTLA-4. Being an X chromosome-encoded transcription factor, Foxp3 is indispensable for both development and function of Treg cells. Mice mutated in Foxp3 as well as patients with immune dysregulation polyendocrinopathy, enteropathy, and X chromosome-linked syndrome (IPEX) result in the development of complex autoimmune diseases due to the deficiency of Treg cells. When manipulated to ectopically express Foxp3, T cells acquire the phenotype of Treg cells. Furthermore, approximately 90% decrease of Foxp3 protein expression due to destabilizing alterations in the 3' UTR of the Foxp3 messenger RNA (mRNA), thereby destabilizing mRNA, results in significantly impaired Treg cell-mediated suppression, demonstrating that the amount of Foxp3 protein directly correlates to the function of Treg cells (9). Constitutive expression of Foxp3 is fundamental for the maintenance of the suppressive function of Treg cells (10).

Table 1 | Characteristics of different Treg cell subsets.

Cell type	Phenotype	Mechanism	Origin
nTreg cells	Foxp3 ⁺ CTLA4 ⁺ TNFR SF18 ⁺ IL7R ^{low} CD25 ^{hi} (mouse) CD25 ^{very hi} (human)	Contact dependent, CTLA4, IL-10, and TGF- β 1	Thymus
nTreg cells (activated)	Foxp3 ⁺ HLA ⁻ DR ⁺ CD69 ⁺ Granzyme B ⁺ (mouse) Granzyme A ⁺ (human)	Contact dependent	Expansion of natural Treg cells
iTreg cells	Foxp3 ⁺ CTLA4 ⁺ TNFR SF18 ⁺	Contact dependent and in some cases TGF- β 1 dependent	Conversion and/or expansion of naive CD4 ⁺ T cells
TH3 cells	Foxp3 ⁻	TGF- β and or/IL-10	Periphery
Tr1 cells	Foxp3 ⁻ IFN γ ⁻ (mouse) IFN γ ^{low} (human)	IL-10	Periphery
CD8 ⁺ T cells	CD28 ^{+/-}	Cell contact ⁻ , LILRB4 ⁻ , and LILRB2 dependent	Periphery

Although the precise signaling mechanisms regulating Foxp3 expression are not fully understood, TGF- β , IL-2, or TCR stimulation of T cells results in increased Foxp3 expression. This is most likely modulated by the demethylation of the Foxp3 promoter or conserved non-coding regions in the Foxp3 locus (11). In addition, multiple transcription factors, including cAMP response element modulator (CREB)/activating transcription factors (ATF), Ets-1 (protein C-ets-1), forkhead box protein O1 (Foxo1), forkhead box protein O3 (Foxo3), and signal transducer and activator of transcription 5 (STAT5), regulate Foxp3 transcription (12, 13). It is not fully clear whether Foxp3⁺ Treg cells can lose Foxp3 expression and suppressive function, as well as whether Foxp3⁺ Treg cells exhibit characteristics of other Th cell subsets. A number of studies in which Foxp3⁺ Treg cells were adoptively transferred into lymphopenic mice demonstrated that approximately 10–50% of the transferred Treg cells lost Foxp3 expression (14–16). Furthermore, Treg cells from both the periphery and the thymus were converted into Th17 cells upon stimulation with anti-CD3, anti-CD28, and IL-6, demonstrating a degree of plasticity (17). In addition, Foxp3⁺ Treg cells can be converted to a Foxp3 Th1 cell phenotype upon *Toxoplasma* infection (18). In fact, Foxp3 can be polyubiquitinated; however, the regulation of this process and the modulators remain elusive (19–21). Deubiquitinating enzyme (DUB) ubiquitin-specific-processing protease 7 (USP7, also known as HAUSP, ubiquitin carboxyl-terminal hydrolase 7 or herpesvirus-associated ubiquitin-specific protease) is active in primary Treg cells and associates with Foxp3. Ectopic expression of USP7 specifically decreased Foxp3 polyubiquitination, resulting in increased Foxp3 expression. Conversely, knockdown of USP7 resulted in decreased Foxp3 expression. Furthermore, the function of Treg cells was noticeably decreased when USP7 was knocked down or when DUB activity was inhibited both *in vitro* and *in vivo* (22). The manipulation of Foxp3 ubiquitination provides a method for temporally controlling the expression of Foxp3 in T cells, thereby regulating the numbers and function of Treg cells.

CD4⁺CD25⁺ Treg cells comprise approximately 5–10% of the mature CD4⁺ T cells in mice and humans, and approximately 1–2% of CD4⁺ Treg cells are detectable in peripheral blood. Murine and human nTreg cells are phenotypically similar on the basis of surface markers, expressing MHC-class II molecules, CD25, CD122, CD132, GITR, CTLA-4, PD-L1, CD62, CD38, CD45RO, and Foxp3. Although both nTreg cells can use the cell–cell contact mechanism to mediate their suppressive function, murine nTreg cells are via a granzyme-B-dependent and perforin-independent pathway, while human nTreg cells are via a granzyme-A and perforin-dependent pathway. Number of CD4⁺CD25⁺ Treg cells in peripheral blood is differed; however, the frequency of Treg cells is constantly higher in the synovial fluid than in the peripheral blood. Furthermore, Treg cells from RA patients still keep their suppressive ability; however, these Treg cells have no capacity to stop the production of inflammatory cytokines such as TNF- α from monocytes or activated T cells (23). This functional defect of Treg cells in RA was related with a high expression of TNF- α that reduced expression of Foxp3, or resulted in defective expression of CTLA-4 (24, 25). Cell number is critical during arthritis development. In many autoimmune disorders, e.g., juvenile idiopathic arthritis, psoriatic arthritis, multiple sclerosis, systemic

lupus erythematous, autoimmune hepatitis, and type-1 diabetes, the numbers and suppressive activity of circulating CD4⁺CD25⁺ Treg cells dramatically reduced (26). Despite presenting in the joints of patients with RA, Treg cells did not possess normal immune suppressive activity. Treg cells in the synovial fluid from RA patients are exposed to a numbers of inflammatory cytokines; high amounts of TNF- α secreted by the inflamed synovium into the joint fluid likely cause the abnormal phosphorylation of Foxp3, resulting in abnormal suppressive function of Treg cells.

The function and frequency of Treg cells can be measured in peripheral blood as well as at the site of inflammation in arthritic patients. Circulating Treg cells in RA patients holding mutable functional activity, particularly with regard to the suppressive function (27); however, at the inflamed joints, the suppressive activity of enriched Treg cells is high and consistent (28). The general agreement is that these are highly reactive Treg cells, which have an increased suppressive activity (29). Similarly in synovium of RA patients, Foxp3 DNA methylation resulted in a higher dedication toward Treg cell lineage (30). In the inflamed synovium of RA patients, local tissue and different immune cells interrelate through cytokines and/or cell–cell contact. Pro-inflammatory cytokines, e.g., TNF- α , IL-6, and antigen presenting cells (APCs) also affect the function of Treg cells. Despite presenting in large amounts and suppressive function *in vitro*, Treg cells likely display different suppressive capacity *in vivo*, countered by the inflammatory environment, or hampered by the resistance from Teffs (31).

MODULATION OF CD4⁺CD25⁺ REGULATORY T CELLS

A number of therapeutic strategies have been implemented in the clinical and many others are being developed for the treatments of RA including: biological therapy, stem cell therapy, mesenchyme stem cell (MSC)-based therapy, hematopoietic stem cell (HSC)-based therapy, and Treg cell-based therapy. In the field of RA, the biological therapy is a fairly new type of therapeutics, and is usually managed at specific biological targets, which have a function in the inflammatory cascade. In general, these targets are TNF- α , IL-6, the IL-1 receptor antagonist, and soluble CTLA-4. There are three biological agents targeting TNF- α . These are infliximab, adalimumab, and etanercept. Currently, these biological agents are used to treating RA. Despite targeting TNF- α is a successful therapeutic strategy in the treatment of RA, methotrexate (MTX), not targeting TNF- α , is also an effective drug (28). MTX is an antifolate drug that suppresses purine and pyrimidine synthesis and inhibits DNA replication. Early treatment with TNF- α inhibitors combined with MTX has been shown to significantly improve treatments of RA (30, 32). In addition to TNF- α and MTX, there are some other biologic agents for treating moderate to severe RA, including abatacept, rituximab, and tocilizumab (31, 33).

Another effective biological target of RA is IL-6. IL-6 is significantly increased in synovial fluid of RA patients, and functions as a pro-inflammatory cytokine, which directly reduced the suppressive function of Treg cells (23). In addition, IL-6 dramatically influenced the conversion of Foxp3⁺ CD4 T cells into Th17 cells (34). Th17 cells play a crucial role in RA pathology through secreting IL-17. IL-17 has ability to activate a number of cells, such as synovial fibroblasts and monocytes that were involved in causing

joint damage (35). Neutralization of IL-17 during reactivation of antigen-induced arthritis in animals prevented joint inflammation and bone erosion (36). Another drug that targets IL-6 receptor is tocilizumab, a humanized antibody that has been proven as a successful therapy for the treatments of RA (37). Tocilizumab can ameliorate the symptom of RA by reducing the Th17 cells, and increasing the number of CD4⁺CD25⁺ Treg cells at the site of inflammation. As a matter of fact, tocilizumab corrected the balance of Th17/Treg cells in RA patients (38).

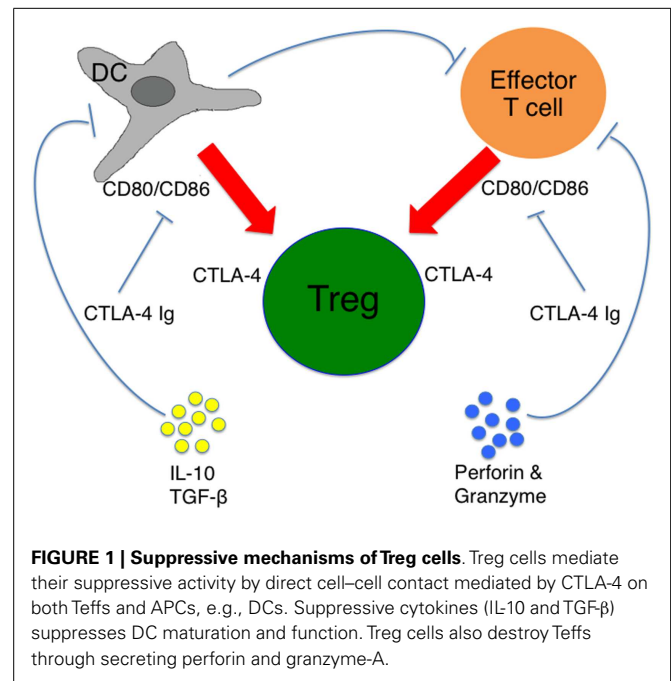
IL-1 is a central pro-inflammatory cytokine that impacts on a number of cell types and subsequently induces bone and cartilage destruction. IL-1 is another important biological that modulates RA pathology. Human recombinant IL-1 receptor antagonist (IL-1ra) can block the IL-1 mediated effects, and restore the balance of Th17/Treg cells. Even though the exact mechanisms remain largely unknown, the use of anakinra, an IL-1ra, as a therapy in RA was effective and safe (39). Treg cells highly express CTLA-4, which controls the suppressive function of Treg cells (40). CTLA-4 is also expressed on activated T cells, however, binding to its ligands induces opposite effects in effector and Treg cells. For example, ligation of CTLA-4 with CD80 (B7-1) and CD86 (B7-2) inhibits the function of Teffs but augments the suppressive capacity of Treg cells. CTLA-4-Ig (also known as abatacept) is a human fusion protein that consists of the extracellular domain of CTLA-4 and the Fc portion of IgG1, which is used for the treatments of RA patients.

In addition, nTregs and iTreg cells can directly kill autologous target cells through a CD18 and perforin-dependent manner (41–43). Upon activation, nTreg cells predominantly express granzyme-A while iTreg cells express granzyme B. Both subtypes of Treg cells exhibit perforin-dependent cytotoxicity against a variety of autologous target cells, including CD4⁺ and CD8⁺ T cells, CD14⁺ monocytes, and DCs. The mechanism by which the Treg cell subsets recognize their targets is unclear; however, several lines of evidence suggest that it is an MHC/TCR independent process. The potential suppressive mechanisms of Treg cells are briefly described in **Figure 1**.

Treg CELL-BASED THERAPIES IN RA

Regulatory T cells play a central role in controlling autoimmunity. Deficiencies in the functions of Treg cells have been identified in a number of autoimmune diseases including RA. A number of studies suggested that stem cell therapy or biological therapy induced a potent population of CD4⁺CD25⁺ Treg cells in patients with RA (44); however, the deficit of nTreg cells persisted in the patients even after the treatment with anti-TNF- α (45). CD4⁺CD25⁺ Treg cells isolated from active RA patients were capable of suppressing the proliferation of conventional T cells; however, these Treg cells could not suppress the cytokine production. It is suggested that reduced expression and functional abnormalities in Treg cells associated with CTLA-4 might account for the defect of Treg cells in patients with RA (25).

After the success of the biological therapies in RA, new therapeutic adventures with the goal to ultimately reach complete clinical remission become more attractive. For reestablish the tolerance by utilizing Treg cells, numerous studies have been performed on Treg cell-based therapies. This approach can be reached



by expansion of CD4⁺CD25⁺ Treg cells *ex vivo* or induction of Treg cells *in vivo*. Because of the inherent resistance of Treg cells to exogenous expansion, and large numbers of Treg cells essentially required to perform the cell-based therapies, it is critical to isolate a high number of Treg cells for exogenous expansion, and the following adoptive transfer. However, caution is needed when translating *in vitro* induction of Treg cells to *in vivo* application. To make sure that Treg cells suppress Teffs at the site of inflammation, a number of strategies have been suggested to regulate the numbers and functionality of Treg cells such as ectopic expression or the acetylation modulation of Foxp3 (46).

Embryonic stem cells (ESCs) or induced pluripotent stem cells (iPSCs) have the strong potential to differentiate into a number of different cell types in the body. Different types of blood cells including myeloid and lymphoid lineages can arise from ESCs or iPSCs. The approach to obtain ESCs from patients is not feasible. HSCs have a higher potential to pass the bone marrow barrier and travel in the blood, which allows HSCs to be easily and safely harvested from the patient blood following mobilization with hematopoietic growth factors. Thus, the use of HSC for therapeutic purposes has been widely applied in clinic, especially in HSC transplantations. In addition, HSCs also can be generated from differentiated cultures from ESCs and fetal-derived embryonic germ stem cells (47). However, compared to pluripotent ESCs or iPSCs, HSCs or MSCs have less potential for self-renewal, and the differentiation declines in response to differentiation signals with each cycle (48). In fact, the ability to expand numbers *in vivo* or *in vitro* would be a huge boost to all current and future medical uses of HSCs and MSCs. In contrast, iPSCs can be easily generated from patients' somatic cells by transduction of various transcription factors and exhibit characteristics identical to those of ESCs (49, 50). Many genetic methods as well as the protein-based approach have been developed to produce iPSC with potentially reduced

risks, including that of immunogenicity and tumorigenicity (51–53). This approach provides an opportunity to generate patient- or disease-specific iPSCs (54–56). Because of the plasticity and the potential for an unlimited capacity for self-renewal, iPSCs have high potential for advancing the field of cell-based therapies (57, 58). Recent results have shown that programming of functional CD4⁺ Treg cells or CD8⁺ cytotoxic T lymphocytes from iPSCs can be used for adoptive immunotherapy of cancers and autoimmune arthritis (59–62).

Embryonic stem cells and iPSCs are exposed to a number of signals responsible for their progression. Although the exact signals are not fully understood, part of the mechanism known to be critical for directing T-cell fate occurs via the Notch signaling pathway (63, 64). Notch is evolutionarily conserved regulating cell fate decisions in a number of cell and tissue types. Ligand binding by members of the Jagged or Delta-like families results in the proteolytic cleavage and release of the intracellular fragment of the Notch heterodimer. Translocation to the nucleus then allows for its regulation of gene expression. Notch-1, specifically, has been shown to be critical for the establishment of T-cell fate. Loss of function results in the blockade of T cell development and enhanced B cell production, while over-expression results in the blockade of B cell lymphopoiesis and leads to the generation of T cells (64). However, the intracellular signaling pathways by which Notch signaling regulates Ag-specific Treg differentiation remain unknown. Genetic modification of iPSCs with the Foxp3 gene and an antigen-specific TCR as well as stimulation with an *in vitro* Notch ligand may direct iPSC differentiation into antigen-specific Treg cells, which produce suppressive cytokines and inhibit other immune cell activities. Adoptive transfer of iPSC-derived Treg cells has been shown to suppress the development of arthritis in a murine model (62).

IMMUNOSUPPRESSION BY Treg CELL THERAPY

The transfer of *ex vivo* expanded Treg cells into the patients is presently the focus of intense research to treat autoimmune disorders and to suppress the occurrence of graft-versus-host disease (GVHD) after bone marrow transplantation (65, 66). *Ex vivo* expanded Treg cells were dramatically useful in the prevention of autoimmune diseases, and in many cases, in the treatments of ongoing inflammatory and autoimmune disorders in mice (4). A number of mechanisms have been suggested for Treg cell-mediated suppression, including secretion of immunosuppressive cytokines from the Treg cells, cell contact dependent suppression, and functional modification or killing of APCs. However, it is not clear whether Treg cells *in vivo* use same or different mechanisms. Nevertheless, a number of key issues need to be considered prior to their clinical use. First, because of the heterogeneity of Foxp3⁺ Treg cells, it is important to determine the optimal Treg cells that should be isolated for expansion. Expansion of Foxp3-expressing T cells is based on isolation of CD25 and CD127; this method usually results in contamination of non-regulatory CD45RO⁺Foxp3^{low} T cells. CD45RO⁺FoxP3^{low} cells are capable of producing pro-inflammatory cytokines, and likely constitute 30–50% of the Foxp3⁺CD4⁺ T cells (67). Because FoxP3 expression is crucial for the suppressive activity of Treg cells, and CD45RA⁺Foxp3^{low} naive Treg cells have the highest capacity

of maintaining the expression of Foxp3 after expansion, the subset of CD45RA⁺Foxp3^{low} naive Treg cells is the best population of choice for isolation (68).

Second, the apoptotic properties of Treg cells should be investigated because these cells may be used to modulate the ratio of Treg to Teff cells. Treg cells are prone to die by apoptosis (69) and it is hard to expand in cultures with high dose of IL-2 (67). In addition, naive Treg cells are easier to expand and develop a high frequency of functional Foxp3⁺ Treg cells; the use of CD45RA⁺Foxp3^{low} naive Treg cells is the best choice for the expansion of Treg cells. It has been shown that purified naive Treg cells, when cultured with high-dose IL-2, could give rise to inflammatory cytokine producing T cells (70). Therefore, certain cytokines and chemicals need to be determined to enable the expansion of functional Treg cells, which will not secrete pro-inflammatory cytokines. Rapamycin (also known as Sirolimus/Rapamune) likely be useful as a component as it substantially increases the purity of Treg cells by eliminating non-Treg cells (71).

To onset the development and pathogenesis of arthritis in mice, a number of cytokines are regarded to play a crucial role (72). Among them, IL-17 has an important role; IL-17 is a T cell derived pro-inflammatory cytokine produced by TCR α/β ⁺CD4⁻CD8⁻ thymocytes, as well as activated CD4⁺ and CD4⁺CD45RO⁺ memory T cells (73). IL-17 is involved in the development of RA because of the existence in the synovial fluid of RA patients and the production from T cell clones established from RA patients (74). IL-17 played a role in the early and late stages of collagen induced arthritis (75). Adoptively transfer of engineered Treg cells substantially decreased inflammatory knee swelling and a significant reduction of Th17 cells in the draining lymph nodes (76). Treg cells were capable of limiting the detrimental effects of Th17 cells by reducing accumulation at the site of inflammation and in the draining lymph nodes (77). However, Treg cells might be inefficient suppressors of Th17 cells (25, 26). Taken together, a successful therapy using Foxp3⁺ Treg cells (both nTreg and iTreg cells) depends on how Treg cells are prepared, the purity of the Treg cells, and how the Treg cells are maintained as a functionally stable cells *in vivo* as well as *in vitro* expanded.

For a successful and potential immunotherapy using Treg cells, Ag specificity of Treg cells are highly recommended. The importance of antigen specificity is a strong indication where high doses of polyclonal Treg cells fail to reverse ongoing autoimmunity. Antigen specificity is required for Treg cell to home or retained at the appropriate site and exerts active suppression. A retroviral transduction protocol in which OTII TCR genes were transduced in primary Treg cells and in CD4⁺ T cells allows these cells to redirect the specificity without the needs for lengthy *in vitro* culture. Within 3 days, their redirected Treg cells and converted CD4⁺ T cells were available for adoptive transfer into recipient mice, where they engrafted efficiently, maintained their phenotype, and suppressed arthritic joint inflammation in an antigen dependent fashion (76). Foxp3 transduction of naive CD4⁺ T cells can convert these cells into functional Foxp3⁺ Treg cells that prevent or suppress the development of a number of autoimmune diseases in animals (78–80). Because of the excellent capacity of stem cell-derived Treg cells in suppression of autoimmunity (62), generation of antigen-specific Foxp3⁺ Treg cells from stem cells such as iPSCs

and HSCs may open a new area in Treg cell-based immunotherapy in transplantation and autoimmune disorders.

ADVANTAGES AND DISADVANTAGES FOR THE USE OF Treg CELL-BASED THERAPY

Reestablishing self-tolerance in autoimmunity depends on self-reactive Treg cells. The advantages of Treg cell-based immunotherapy over conventional treatments are numerous. These benefits include: (1) the potential for antigen specificity with the lack of general immunosuppression; (2) the possibility of inducing long-lasting regulation *in vivo*; and (3) custom-made product that can be designed for each patient, with very limited or absent side effects. However, due to the lack of Treg cell specific surface antigens, purification of a more comprehensive Treg cell subset increases the contamination by the non-regulatory T cells. Although the therapeutic potential for Treg cell-based immunotherapy is now well established in animal models (81–87), to date, such therapy has not been directly applied to suppress autoimmunity in humans.

There are a number of problems and limitations in generating a large number of Treg cells, in spite of the growing number of methods for isolating Treg cells. No approach or method has been established to isolate the population of Treg cells with 100% specificity. In addition to the issue of isolation, the survival of Treg cells is another critical factor in Treg cell-based therapies. Treg cells are disposed to apoptosis in the absence of certain cytokines (e.g., IL-2) since Treg cells do not secrete IL-2, and are essentially dependent on other cells (88). Also, Treg cells are prone to apoptosis because of the low expression of the bcl-2 family proteins. Bcl-xL is an anti-apoptotic protein that can sustain the survival of T lymphocytes (88). β -catenin promoted the survival of Treg cells *in vitro* without changing their anergic state or suppressive function by regulating expression of anti-apoptotic bcl-xL (89). Indeed, bcl-xL is involved in the development and function of Treg cells by inducing the expression of Foxp3, CTLA-4, TGF- β , and repressing the programmed death receptor-1 (PD-1) expression (90, 91). Foxp3 and bcl-xL cooperatively promote the survival of Treg cells and prevent arthritis development because of the increased survival of Treg cells (80). These approaches likely induce the differentiation and survival of Treg cells, and result in the persistence of Treg cells.

In addition, there are several unsettled issues regarding the function of Treg cells in rheumatic diseases. These unresolved questions significantly limit the potential use of Treg cell-based therapies in clinic. For instance, it remains unclear whether the existed effects of Treg cell-based therapies redirect the fundamental mechanism that donates to medical progress. In fact, the differences in the compartment of Treg cells may not be directly involved in the improvement of symptoms. Assessment of the function of Treg cells is also an obstacle, because there is no specific surface marker of Treg cells in humans, and highly activated T cells also transiently express Foxp3. Furthermore, the accessibility of the site of inflammation in humans is limited. Of note, most investigations are studies in the periphery, which possibly will not uncover the interactions occurring at the site of inflammation (92).

CONCLUSION

Due to the plasticity and possibly infinite ability for self-renewal, stem cell-derived CD4⁺CD25⁺ Treg cells likely are applicable for

Treg cell-based immunotherapy, such as autoimmune disorders. Several questions remain: (i) how to direct the differentiation of Treg cells, (ii) how to obtain a large number of functional Treg cells, and (iii) which type of stem cells can be feasibly applicable for individual use. Stem cell or their offspring likely is an optimal option for the use of Treg cell-based therapies. iPSCs have great potential as a source for the generation of antigen-specific Treg cells to be utilized in personalized therapies. However, significant challenges remain to bring the use of stem cell-based technologies into the clinic to treat destructive chronic diseases like RA.

More also needs to be known about the effects of standard therapy on Treg cells, because the precise contribution of Treg cells to the pathogenesis of RA remains unclear, even Treg cells play an important role. Regarding MTX, it is still controversial that Treg cells are impacted by the treatments of MTX. With regards to targeting the pro-inflammatory cytokines TNF- α , IL-6, IL-1, and the costimulatory molecule CTLA-4, most investigations strengthen a stimulating effect of blocking activity on Treg cells. This underscores the importance of further studies on the effects of current therapies on the function of Treg cells, which could eventually lead to the therapeutic applications by utilizing Treg cells.

ACKNOWLEDGMENTS

This project was funded, in part, under grants from the National Institute of Health Grant K18CA151798, the Leona M. and Harry B. Helmsley Charitable Trust Grant No. 2014PG-T1D049, and the Pennsylvania Department of Health using Tobacco CURE Funds to Jianxun Song.

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Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Received: 31 May 2014; accepted: 23 July 2014; published online: 08 August 2014.
Citation: Haque M, Fino K, Lei F, Xiong X and Song J (2014) Utilizing regulatory T cells against rheumatoid arthritis. *Front. Oncol.* **4**:209. doi: 10.3389/fonc.2014.00209
This article was submitted to Tumor Immunity, a section of the journal *Frontiers in Oncology*.
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