**GENOME SEQUENCES**





## **Complete Genome Sequence and Annotation of Campylobacter jejuni YH003, Isolated from Retail Chicken**

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**ABSTRACT** The complete genome sequence of Campylobacter jejuni YH003, isolated from retail chicken, was determined using PacBio and Illumina technologies. The assembled genome is 1,743,985 bp (G-C content of 30.3%). Genome annotation revealed several genes encoding virulence and antibiotic resistance factors, including a type VI secretion system, cytolethal distending toxins, and a multidrug efflux system.

Campylobacteriosis caused by *Campylobacter jejuni* is the most prevalent foodborne<br>illness in the United States and worldwide [\(1,](#page-1-0) [2\)](#page-1-1). Wild and domestic birds, especially poultry, are the primary reservoirs, which leads to exposure for humans through contact with raw or undercooked poultry [\(3](#page-1-2)[–](#page-1-3)[5\)](#page-1-4). While campylobacteriosis is usually self-limiting, treatment with antibiotics, mainly fluoroquinolones, may be necessary in some cases [\(6\)](#page-1-5). Use of antibiotics in poultry to curtail infections has led to an increase in antibiotic resistance in *Campylobacter* spp., which can have an impact on antibiotic treatment in humans  $(7, 8)$  $(7, 8)$  $(7, 8)$ . The whole-genome sequence and annotation of C. jejuni YH003, isolated from retail chicken, can reveal genetic information related to virulence and antibiotic resistance and thus assist in the identification of pathogenicity and antibiotic susceptibilities of the pathogen.

C. jejuni YH003 was recently isolated from chicken drumsticks purchased from a local supermarket. The strain was isolated using a passive filtration method [\(9,](#page-1-8) [10\)](#page-1-9). Confirmation of the genus and species was done by 16S rRNA gene sequencing and multiplex real-time PCR targeting the hipO and cdtA genes [\(11,](#page-1-10) [12\)](#page-1-11). The strain was grown in Brucella broth (Difco; Becton, Dickinson, Franklin Lakes, NJ) for approximately 40 h under microaerophilic conditions and pelleted by centrifugation, and genomic DNA was extracted using the Genomic tip 100/G (Qiagen, Valencia, CA) and sequenced using both single-molecule real-time (SMRT) sequencing (Pacific Bioscience, Menlo Park, CA) and the MiSeq platform (Illumina, San Diego, CA). The Nextera XT library preparation kit (Illumina) was used to prepare the library for Illumina sequencing. SMRTbell library preparation and PacBio sequencing were performed by the University of Delaware DNA Sequencing and Genotyping Center (Newark, DE). Coverages for the PacBio and Illumina reads were  $125\times$  and  $506\times$ , respectively. Illumina reads (a total of 1,759,375 reads, with an average length of 251 nucleotides) were quality controlled by FastQC and assembled using SPAdes v.3.7.1 [\(13\)](#page-1-12), while a total of 17,686 PacBio reads, with an average length of 12,592 nucleotides, were assembled using Canu v.1.3 [\(14\)](#page-1-13). The two de novo assemblies were joined and corrected using Pilon v.1.22 to obtain a single contig. The draft genome was manually edited and oriented. CLC Workbench (Qiagen Bioinformatics, Redwood City, CA) was used to confirm the assembly by mapping the reads back. Default parameter settings were used for all software. Annotation of the final assembly was performed using Rapid Annotation using Subsystem Technology (RAST) [\(http://rast.nmpdr.org\)](http://rast.nmpdr.org). The NCBI Prokaryotic Genome Annotation Pipeline (PGAP) was also used for genome annotation, and the results are publicly available.

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C. jejuni YH003 has a circular chromosome of 1,743,985 bp (G+C content of 30.3%), encoding 1,785 proteins. Based on a BLAST search against all of the available Campylobacter genomes in the NCBI database and comparative genome analysis with Mauve alignment using default settings, this strain is most closely related to C. jejuni NCTC 11168. Distinctly, C. jejuni YH003 contains 16 annotated proteins of the type VI secretion system (T6SS), an important virulence factor exporting cytotoxic molecules to host cells. Strain YH003 also contains the ATP-dependent chaperone ClpB as part of the T6SS. In addition to 86 motility and chemotaxis genes, this strain possesses 65 virulence genes, including cadF, jlpA, ciaB, pebAC, omp, htrA, and the cytolethal distending toxin genes cdtCBA. Interestingly, RAST annotation revealed a type II CRISPR system containing the cas1, cas2, and cas9/csn1 genes, a type I restriction-modification system with the hsdM, hsdS, rloC, and hsdR genes, a Campylobacter multidrug efflux pump consisting of cmeABCR, and a 36-kb complete Mu-like phage in the chromosome of C. jejuni YH003.

**Data availability.** This complete genome sequence has been deposited in GenBank under accession number [CP041584.](https://www.ncbi.nlm.nih.gov/nuccore/CP041584) The raw reads from PacBio RS II and Illumina MiSeq sequencing are available in the NCBI Sequence Read Archive (SRA) under accession numbers [SRX7021025](https://www.ncbi.nlm.nih.gov/sra/SRX7021025) and [SRX7021026,](https://www.ncbi.nlm.nih.gov/sra/SRX7021026) respectively. The BioProject accession number is [PRJNA553752,](https://www.ncbi.nlm.nih.gov/bioproject/PRJNA553752) and the BioSample accession number is [SAMN12249490.](https://www.ncbi.nlm.nih.gov/biosample/SAMN12249490)

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## <span id="page-1-0"></span>**REFERENCES**

- 1. Kaakoush NO, Castano-Rodriguez N, Mitchell HM, Man SM. 2015. Global epidemiology of Campylobacter infection. Clin Microbiol Rev 28: 687–720. [https://doi.org/10.1128/CMR.00006-15.](https://doi.org/10.1128/CMR.00006-15)
- <span id="page-1-1"></span>2. Platts-Mills JA, Kosek M. 2014. Update on the burden of Campylobacter in developing countries. Curr Opin Infect Dis 27:444 – 450. [https://doi](https://doi.org/10.1097/QCO.0000000000000091) [.org/10.1097/QCO.0000000000000091.](https://doi.org/10.1097/QCO.0000000000000091)
- <span id="page-1-2"></span>3. Wagenaar JA, Mevius DJ, Havelaar AH. 2006. Campylobacter in primary animal production and control strategies to reduce the burden of human campylobacteriosis. Rev Sci Tech 25:581–594. [https://doi.org/10](https://doi.org/10.20506/rst.25.2.1680) [.20506/rst.25.2.1680.](https://doi.org/10.20506/rst.25.2.1680)
- <span id="page-1-3"></span>4. Janssen R, Krogfelt KA, Cawthraw SA, van Pelt W, Wagenaar JA, Owen RJ. 2008. Host-pathogen interactions in Campylobacter infections: the host perspective. Clin Microbiol Rev 21:505–518. [https://doi.org/10.1128/CMR](https://doi.org/10.1128/CMR.00055-07) [.00055-07.](https://doi.org/10.1128/CMR.00055-07)
- <span id="page-1-5"></span><span id="page-1-4"></span>5. Skarp CPA, Hanninen ML, Rautelin H. 2016. Campylobacteriosis: the role of poultry meat. Clin Microbiol Infect 22:103–109. [https://doi.org/10](https://doi.org/10.1016/j.cmi.2015.11.019) [.1016/j.cmi.2015.11.019.](https://doi.org/10.1016/j.cmi.2015.11.019)
- <span id="page-1-6"></span>6. Konkel ME, Monteville MR, Rivera-Amill V, Joens LA. 2001. The pathogenesis of Campylobacter jejuni-mediated enteritis. Curr Issues Intest Microbiol 2:55–71.
- 7. Epps SV, Harvey RB, Hume ME, Phillips TD, Anderson RC, Nisbet DJ. 2013. Foodborne Campylobacter: infections, metabolism, pathogenesis and reservoirs. Int J Environ Res Public Health 10:6292-6304. [https://doi.org/](https://doi.org/10.3390/ijerph10126292) [10.3390/ijerph10126292.](https://doi.org/10.3390/ijerph10126292)
- <span id="page-1-7"></span>8. Luangtongkum T, Jeon B, Han J, Plummer P, Logue CM, Zhang Q. 2009. Antibiotic resistance in Campylobacter: emergence, transmission and persistence. Future Microbiol 4:189 –200. [https://doi.org/10.2217/17460913.4.2](https://doi.org/10.2217/17460913.4.2.189) [.189.](https://doi.org/10.2217/17460913.4.2.189)
- <span id="page-1-8"></span>9. Jokinen CC, Koot JM, Carrillo CD, Gannon VP, Jardine CM, Mutschall SK, Topp E, Taboada EN. 2012. An enhanced technique combining preenrichment and passive filtration increases the isolation efficiency of Campylobacter jejuni and Campylobacter coli from water and animal fecal samples. J Microbiol Methods 91:506 –513. [https://doi.org/10.1016/j](https://doi.org/10.1016/j.mimet.2012.09.005) [.mimet.2012.09.005.](https://doi.org/10.1016/j.mimet.2012.09.005)
- <span id="page-1-9"></span>10. Speegle L, Miller ME, Backert S, Oyarzabal OA. 2009. Use of cellulose filters to isolate Campylobacter spp. from naturally contaminated retail broiler meat. J Food Prot 72:2592–2596. [https://doi.org/10.4315/0362](https://doi.org/10.4315/0362-028x-72.12.2592) [-028x-72.12.2592.](https://doi.org/10.4315/0362-028x-72.12.2592)
- <span id="page-1-10"></span>11. He Y, Yao X, Gunther NW, Xie Y, Tu S-I, Shi X. 2010. Simultaneous detection and differentiation of Campylobacter jejuni, C. coli, and C. lari in chickens using a multiplex real-time PCR assay. Food Anal Methods 3:321–329. [https://doi.org/10.1007/s12161-010-9136-6.](https://doi.org/10.1007/s12161-010-9136-6)
- <span id="page-1-11"></span>12. Irwin P, Nguyen TL, Chen CY. 2008. Binding of nontarget microorganisms from food washes to anti-Salmonella and anti-E. coli O157 immunomagnetic beads: minimizing the errors of random sampling in extreme dilute systems. Anal Bioanal Chem 391:515–524. [https://doi.org/10.1007/](https://doi.org/10.1007/s00216-008-1961-8) [s00216-008-1961-8.](https://doi.org/10.1007/s00216-008-1961-8)
- <span id="page-1-12"></span>13. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455– 477. [https://doi.org/10.1089/cmb.2012.0021.](https://doi.org/10.1089/cmb.2012.0021)
- <span id="page-1-13"></span>14. Koren S, Walenz BP, Berlin K, Miller JR, Bergman NH, Phillippy AM. 2017. Canu: scalable and accurate long-read assembly via adaptive k-mer weighting and repeat separation. Genome Res 27:722–736. [https://doi](https://doi.org/10.1101/gr.215087.116) [.org/10.1101/gr.215087.116.](https://doi.org/10.1101/gr.215087.116)