

ORIGINAL RESEARCH

Anti-Acid Biomimetic Dentine Remineralization Using Inorganic Silica Stabilized Nanoparticles Distributed Electronspun Nanofibrous Mats

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Methods: The Si/Ca(Sr)-NCF mats were fabricated by dipping the electrospun nano cellulose fiber (NCF) into silica and calcium (strontium) carbonate liquid. Physicochemical characterizations and ion release were confirmed by scanning electron microscopy (SEM), Fourier transform infrared spectroscopy (FTIR), ion release assays and transmission electron microscopy (TEM). Sixty dentine discs were randomly divided into five groups: 1, blank NCF; 2, Si/Ca(Sr 0)-NCF; 3, Si/Ca(Sr 0.01)-NCF; 4, Si/Ca(Sr 0.05)-NCF; 5, Si/Ca(Sr 0.1)-NCF. Dentine discs were mineralized by the mats and observed with SEM immediately, after acid challenge and remineralized in artificial saliva. The releasing liquid was investigated by TEM and type I collagen model. Then, antibacterial property and cytocompatibility were evaluated.

Results: SEM and TEM results confirmed that the experiment mats continuously released amorphous Si/Ca(Sr) NPs and consequently realized anti-acid dentine biomimetic remineralization. Homogeneous surface coverage and collagen intrafibrillar mineralization in strontium adding groups illustrated the mineralization effect was not only by in site precipitation, but also collagen heterogeneous nucleation. Additionally, acceptable antibacterial and cytocompatibility properties were illustrated in low and middle Sr²⁺ containing mats.

Conclusion: In vitro studies on human dentine discs and type I collagen demonstrated that Si/Ca(Sr)–NCF system was a multifunction system inducing anti-acid, biomimetic, antibacterial and cytocompatible dentine remineralization. This multifunction mat would be a promising DH treatment candidate for complicated exposed dentine surfaces.

Keywords: mineralization, silica, dentine hypersensitivity, strontium carbonates, electrospinning

Introduction

Dentine hypersensitivity (DH) is a sharp tooth pain condition in dentistry.¹ Based on available studies, DH prevalence rate ranges from 10–30% of the general population and most of the sufferers are in their 20s to 40s.^{2,3} Irritated by daily activities such as air/water contact, acid food or toothbrushing, this frequent DH pain can be disruptive to basic life.^{2,3} For DH to occur, the outer enamel or

Correspondence: Wei Teng Email tengwei@mail.sysu.edu.cn cementum covering the dentine usually are destroyed with exposed surfaces which are susceptible to external chemical and mechanical stimulus. These irritations can trigger the dentine tubular fluid flow and induce the tooth pain via the excitation of pulp nerve fibers according to the widely accepted hydrodynamic theory.^{4,5}

Nowadays there are two treatment strategies, namely, reducing the pulp nerve response and occluding the exposed dentine tubulars. 6-8 Nerve paralysis agents, for example, potassium salts can interfere with the signaling in nerve receptors and relieve the sharp pain. However, the evidence based research failed to find obvious support for the subjective pain relief for the potassium salts.⁹ Alternatively, various materials are utilized to occluding the exposed dentine surfaces. 6-8 Among the large number of strategies, biomimetic materials are in greater demand compared to the classic ion-based minerals such as calcium salts and phosphates in recent decades.⁸ Biomimetic materials, based on non-classical particle-based crystallization concept, are usually amorphous mineral precursor nanoparticles stabilized with non-collagenous protein analogs. 10,11 With prolonged crystallization process and suitable aggregation sizes, the biomimetic mineralization controlled by organics presents obvious advantages over the violent crystallization by ion based process.^{8,12} Nevertheless, these organic-inorganic strategies still have drawbacks such as the lack of anti-acid property. Acid resistance is an important criterion for the stable existence of the occluding layers.^{2,6} Remineralization structure may be easily lost because of the daily acid food or tooth surface attrition during complicated oral activities. With anti-acid property, another important remineralization material is silicon based materials such as bioactive glass (BG). However, the BG induces rapid ion release with drastically increased pH value. 13 Many of the prior materials are limited with their single function and complicated operations.

Inspired by silica/carbonate co-precipitation nanoparticles originated in natural mineral sedimentation, polymer silica is found to be a special inorganic precursor stabilizer recently. ¹³ In the silica/carbonate co-precipitation nanoparticle, polymer silica shells and amorphous carbonate cores self-assemble with each other by electrostatic chemical compelling under silica rich and weak alkaline environment. ^{15,16} Remarkably, this polymer silica skin can simultaneously be an anti-acid component and kinetically stabilize the inner amorphous carbonates. ^{15,17} This

simple silica shell would be a promising new silicon structure conducting anti-acid biomimetic mineralization.

In addition to the bifunctional silica shell, the inner cores of co-precipitation nanoparticles are alkaline cation carbonates such as calcium (Ca) or barium (Ba) carbonates which are commonly used mineralization components. ^{14–16} With the aim to utilize an anti-acid biomimetic co-precipitation nano particle, the inner carbonates were further selected as mixed strontium (Sr) and calcium carbonates as the addition of Sr²⁺ is reported to enhance crystal anisotropy. ¹⁸ Our previous research also suggested promoting osteogenic differentiation by adding SrCO₃. ¹⁹

To release the silica/calcium (strontium) carbonate coprecipitation nanoparticles (Si/Ca(Sr) NPs) onto dentine surfaces, the present study developed an all-in-one strategy fabricating the mineralized nano cellulose fiber (NCF) mats which can release functional Si/Ca(Sr) NPs. NCF mats is a widely sourced and low-cost fiber and can be hydrolyzed from the electrospun cellulose acetate (CA) mats in alkaline solution.²⁰ Benefit from the rich hydroxyl groups, NCF would provide quantity connecting positions and hydrophilicity would also make the membrane convenient to use. Based on the successful loading onto the negative charged membranes in previous research, dipping the hydrolyzed anionic NCF mats into the silica/carbonate co-precipitation liquid, may facilitate a NCF mat wrapping silica/carbonate particles as well.²¹ Herein, the present study coated the Si/Ca(Sr)-NCFs onto the exposed dentine surfaces with the hypothesis that the Si/Ca(Sr)-NCF mats can conduct anti-acid biomimetic dentine remineralization. Additionally, the antibacterial effects and response of human dental pulp cells (hDPCs) were investigated.

Materials and Methods

Materials

Dentine Sample Preparation

The present study was conducted in accordance with the Declaration of Helsinki. After being authorized by the hospital of stomatology, Wuhan University Ethics Committee (2019-B10), 100 human extracted molars and premolars were collected after the informed consents signed by patients or their legal guardians according to approved protocols for the use of hard tissue research and human dental pulp cell (hDPCs) culture. The dentine discs were cut perpendicularly along the occlusal-apical axis with a low-speed saw (Isomet, Buehler, Lake Bluff, USA) with the thickness of $1.0 \pm 0.1 \,$ mm and be sawed off between the pulp cavity and

superficial enamel. The samples were polished with 600-grit silicon carbide paper to 1.0 mm thickness. In order to remove the smear layer and simulate the exposed hypersensitive dentine surface, the polished samples were soaked into 0.5 M EDTA liquid in an ultrasonic bath for 2 min.^{22,23}

Preparation of Mineralization Liquid

0.1 M sodium metasilicate, 0.1 M calcium chloride and 0.1 M sodium carbonate solution of 10 mL aqueous solution were mixed in centrifuge tubes. 0, 0.01, 0.05 and 0.1 M strontium chloride were respectively added into four carbonate composition groups. The pH value was then adjusted to 10.5 with the HCl solution.

Synthesis of Electrospinning NCF Mats

NCF mats were fabricated by electrospinning of a 15 wt% CA (Mn 30,000) and solution prepared in a 2/1 (wt/wt) acetone/N, N-dimethylacetamide mixture according to the previous studies.²⁰ Hydrolysis of the CA mats was implemented in a 0.05 M sodium hydrate aqueous solution at ambient for 7 days and the CA mats were then transformed into NCF mats.

Preparation of Si/Ca(Sr)-NCF Mats

Five groups of NCF mats were dipped into different mineral liquid for 24 hours as following protocols:

group 1: blank NCF mats;

group 2: Si/Ca(Sr 0)-NCF mats, Sr²⁺ free;

group 3: Si/Ca(Sr 0.01)-NCF mats, low Sr²⁺;

group 4: Si/Ca(Sr 0.05)-NCF mats, middle Sr²⁺;

group 5: Si/Ca(Sr 0.1)-NCF mats, high Sr²⁺.

The mats were dried in room temperature and applied onto dentine discs.

Preparation of the Remineralized Dentine Discs

After thorough inspection, teeth with cavity and defects were excluded and sixty discs divided into five groups were applied in the following experiments. The dentine discs with the mineralization mats were placed in petri dishes and were immersed in shallow purified water for 24 hours. Then, discs were picked out, and the covering mats were peeled off. There were twelve samples for each group for remineralization morphology, acid challenge and AS remineralization observation.

Characterization of the Si/Ca(Sr)-NCF Remineralization System

The Characterization of Si/Ca(Sr)–NCF Mats

The morphology and physical and chemical properties of Si/Ca(Sr)-NCF mats were investigated with scanning

electron microscopy (SEM; S-4800, Hitachi, Japan), Fourier transform infrared spectroscopy (FTIR; Thermo Nicollet 5700, USA) Raman spectra (i-Raman Portble Raman Spectrometer, B&W TEK Inc, USA), drop shape analysis system CASTTR3.0 (USA KINO Industry Co., USA) and tensile test system Tex-ture Analyzer (TA.XT. plus, Stable Micro Systems, UK).

Element Entrapment and Release Assay

Newly synthesized Si/Ca(Sr)-NCF mats were collected and treated with hydrogen nitrite. After diluting, the solution samples were analyzed for concentration of Ca, Sr and Si ions using an atomic absorption spectrophotometer (AAS; ContrAA 700, Analytik Jena AG, Germany). At time point day 0–3, 4–6, 7–9 and 10–12, the non-accumulative and total concentration of ions released were investigated.

Then, the suspension released from the Si/Ca(Sr)-NCF mats instantly and after aging for 7 days were examined by transmission electron microscopy (TEM; JEOL JEM 2100, Japan) and the selected area electron diffraction (SAED). The releasing liquid for 7 days was dehydrated in ambient environment and exterminated by X-ray diffraction (XRD; X'Pert PRO, PANalytical, The Netherlands).

The Characteristic of Remineralized Dentine Discs and Type I Collagen

SEM observation was performed on Si/Ca(Sr)–NCF remineralized dentine discs, after acid leaching by 6 wt% citric acid for 1 min and after crystallization in artificial saliva (AS) for 7 days. The AS was prepared with 1.5 mM calcium chloride, 50 mM potassium chloride, 0.9 mM monobasic potassium phosphate and 20 mM Tris.²¹

The type I collagen fibers constructed on nickel mesh was stained by 0.1% uranyl acetate as the previous study.²⁴ After immersing in the selected Si/Ca(Sr)-NCF mat releasing liquid for 6, 12 and 24 hours, the collagen fibers were observed by the TEM.

In vitro Anti-Bacterial Ability

To analyse the inner tubular antibacterial ability, *Enterococcus faecalis* (*E. faecalis*; ATCC 29212, ATCC, Manassas, VA, USA) was selected. Both sides of the mats (6 mm × 6 mm) were disinfected for 1 hour under ultraviolet. The suspended bacteria colonies counting was conducted by coating the suspension evenly on agar plates after co-culturing for 6, 12 and 24 hours. Plates with representative colony numbers were recorded in photos.

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The live bacteria colonies were quantified, and the anti-bacterial ratio (%) was calculated.

In vitro Cellular Studies

Cell Proliferation

Human dental pulp cells (hDPCs) were isolated from young patients' pulp tissue via improved explant culture. In-vitro cytotoxicity and morphology observation had been proceeded with 3–5 passage cells. Cell proliferation on the mats was determined using a cell counting kit-8 (CCK-8; Dojindo Laboratories, Japan) at 4 hours, 1, 4 and 7 days.

Cell Adhesion and Morphology with Si/Ca(Sr)-NCF Mats

At the time point of 24 h hours, the adhesion of cocultured cells with the material was observed under fluorescence microscopy (Leica DM4000B, Germany). Filamentous actins (F-actins) were stained with rhodamine phalloidin (R-415 kit, Invitrogen, USA) and the nuclei were stained with 2-(4-amidinophenyl)-6-indolecarbamidine dihydrochloride (DAPI; Invitrogen, Basel, Switzerland). The cell viability was evaluated by Live/ Dead assay with calcein-AM (Beyotime, China) and propidium iodide (PI; Beyotime, China).

Alkaline Phosphatase (ALP) Activity and RT-qPCR for Osteogenesis-Related Gene Expression

To evaluate the ALP activity, cells were cultured with hDPCs osteogenic factors which contains 100 nM dexamethasone, 10 mM-glycerol phosphate and 50 g/mL ascorbic acid (Sigma, USA). The total cellular lysate harvested on day 7 and day 14 was measured in alkaline buffer solution (A9226; Sigma Chemicals) with p-nitrophenyl phosphate (pNPP; Sigma, USA). Intracellular total protein content was determined via the BCA protein assay kit (Thermo Fisher Scientific, Waltham, MA). Then the ALP activity was normalized to the total protein content.

The gene expressions of osteogenesis related genes including ALP, osteocalcin (OCN), runt-related transcription factor 2 (Runx2), osteopontin (OPN) and glyceraldehyde 3-phosphate dehydrogenase (GAPDH) in hDPCs after osteogenic induction for 7 days were further evaluated by RT-qPCR according to the method in previous research.

Statistical Analysis

Statistical analysis was performed by SPSS 19.0 (SPSS Inc, USA). Results in ion release assay, CFU counting, CCK-8 and ALP experiments were analyzed using one-

way ANOVA with post hoc Tukey's test, respectively. The significance level was set at $\alpha = 0.05$.

Result

Physicochemical Characterization of the Si/Ca(Sr)-NCF Mats

Surface Topography and Microstructural Characterization

The electrospun NCF mats showed in loose woven form with smooth surfaces (Figure 1a1–a3). In Si/Ca(Sr 0)-NCF group, accrose aggregations erupted from the granular aggregation attaching to the fibers and 10–20 µm diameter sphere particles were dispersed among the fibrous structure (Figure 1b1–b3). While in Sr²⁺ containing groups, no large bulk was detected. Denser accrose aggregations were presented in Si/Ca(Sr 0.01)-NCF group (Figure 1c1–c3). In Si/Ca(Sr 0.05)-NCF group, there were obvious segmented nano granular attachments onto the fibers (Figure 1d1–d3). As for the images of Si/Ca(Sr 0.1)-NCF group, the aggregation was more evenly attached onto the fibers (Figure 1e1–e3).

The undistinguished peak at 1750 cm⁻¹ in NCF FTIR results implied the successful transformation from CA to NCF mats (Figure 2A).²⁰ Peaks at 850 cm⁻¹ (v₂), and 1449 cm⁻¹ (v₃) representing the witherite or calcium carbonates further showed the existence of carbonates. Specially, the absence of 694 cm⁻¹ (v₁) peaks, with the peak splitting of 1449 cm⁻¹ (splitting into 1420 cm⁻¹ and 1450 cm⁻¹) in all mineral groups further indicated that there are amorphous carbonates entrapping in the polymerized silica and NCF networks (Figure 2A). 14,15 In the Raman spectrum, the peaks at 701 cm⁻¹ and 1080 cm⁻¹ also indicated the existence of alkali metal carbonates (Figure 2B). 25 Additionally, the Si/Ca(Sr)-NCF hydrophilicity decreased as higher Sr²⁺ containing (Figure 2C) and the elastic modulus decreased with the combination of minerals as the result illustrated in Figure 2D, namely, 4.38 MPa in NCF, 1.82 MPa in Si/Ca(Sr 0)-NCF, 1.06 MPa in Si/Ca(Sr 0.01)-NCF, 0.98 MPa in Si/Ca(Sr 0.05)-NCF and 0.29 MPa in Si/Ca(Sr 0.1)-NCF mats.

Element Entrapment and Ion Release Assay

The Sr entrapment varied in different groups (Figure 3Aa-c), and the noncumulative ion concentrations of Sr, Ca and Si showed no evident burst release in the long-term observation (within 12 days) (Figure 3Ad-f). Within 12 days, the total Sr²⁺ release was tested as 0.48 mM, 2.59 mM, and 4.75 mM in the three Sr²⁺ groups.

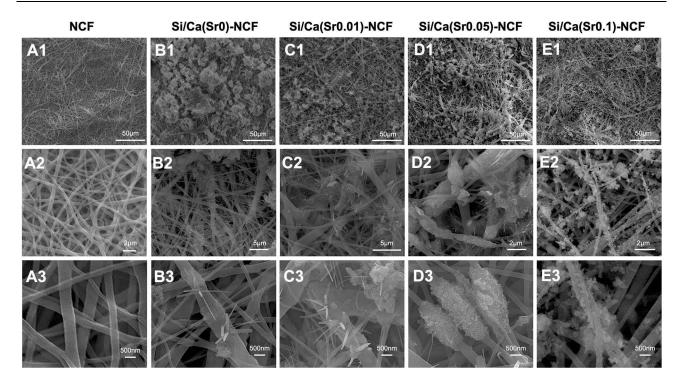


Figure 1 The SEM observation of Si/Ca(Sr)-NCF mats. SEM images of (A1-A3) control NCF mats, (B1-B3) Si/Ca(Sr 0)-NCF mats, (C1-C3) Si/Ca(Sr 0.01)-NCF mats, (D1-D3) Si/Ca(Sr 0.05)-NCF mats and (E1-E3) Si/Ca(Sr 0.1)-NCF mats.

Abbreviations: SEM, scanning electron microscopy; NCF, nano cellulose fiber; Si/Ca(Sr)-NCF, the nano cellulose fiber mineralized with silica and carbonates.

Liquid released immediately from the films was observed by TEM in Figure 3B. Si/Ca(Sr)-NCF mats with or without Sr²⁺ addition all presented nanoparticles with a diameter of approximately 50 nm (Figure 3Ba–d) with amorphous SAED patterns (Figure 3Be–h). The EDS results illustrated the Ca/Sr containing in the releasing liquid showing the increasing Sr²⁺ containing (Figure 3Bi–l).

Liquid released from the representative Sr²⁺ containing group: Si/Ca(Sr 0.05)-NCF group for 7 days was observed with TEM. After 7 days' coagulation, long columnar and small needle shaped crystals in the diluent were showed by TEM (Figure 3Cb and c) and SEM (Figure 3Ch and i). The 50 nm sized packed particles attaching to the crystal edge implied the existing of nanoparticles, which proposed the stabilization of amorphous carbonates for long-term (Figure 3Ca, c and d). Characteristic shell shedding phenomenon of Si/Ca(Sr) NPs was also detected around the crystal edges (Figure 3Cc and d; red arrows) and SEM images further illustrated the accumulated shedding silica membrane wrapping the crystals and the aggregated nanoparticles (Figure 3Ci; red colored). This silica shell and membrane in aged liquid evidently demonstrated the coprecipitation growth behavior of the silica and the carbonates.

Next, in the lattice spacing analyses, (221) of calcite (PDF# 51-1524) and the (021) of strontianite (PDF# 05-0418) in Si/Ca(Sr 0.05)-NCF group were illustrated (Figure 3Ce and f). Corresponding XRD peaks at 31.82° and 25.80° were confirmed (Figure 3Cg). Besides, the aragonite pattern (PDF# 41-1475) at 27.35°, 45.85°, 53.94° and 66.19° demonstrated in all groups implied the co-existence of aragonite and calcite.

The Mineralization of in vitro Human Dentine Discs and Type I Collagen SEM Observation of Mineralized in vitro Human Dentine Disc

In the dentine SEM images after remineralization by the Si/Ca(Sr)-NCF mats' coverage for 1 day, full surface occluding was presented in all Sr²⁺ adding groups (Figure 4A). While in Sr²⁺ free Si/Ca(Sr 0)-NCF group, there were precipitations in peritubular dentine with partially patent tubular (Figure 4Ab1 and b2). In Si/Ca(Sr 0.01)–NCF and Si/Ca(Sr 0.05)–NCF groups, 0.5–2.0 µm sized particles scattered on the homogeneous basal precipitation layer (Figure 4Ac1, c2, d1 and d2), while the large blade shaped crystals densely covered the full dentine surface in Si/Ca(Sr 0.1)-NCF group (Figure 4Ac1 and e2). The longitudinal sections showed corresponding

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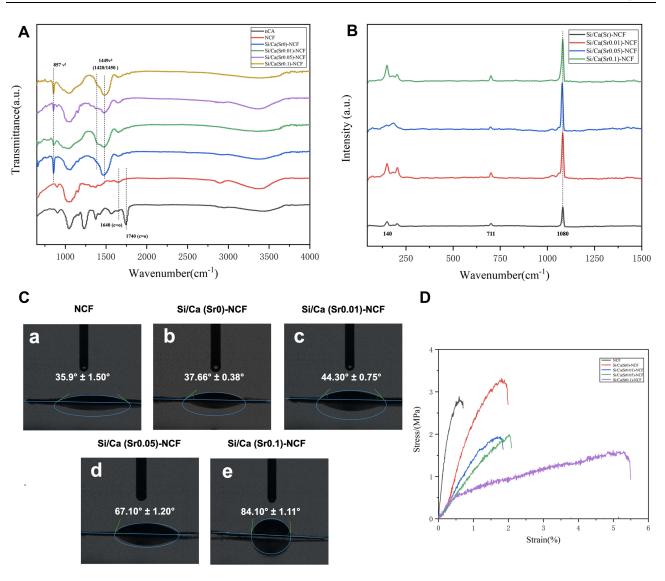


Figure 2 The FTIR, Raman spectrum, water contact angles and elastic tests of Si/Ca(Sr)-NCF mats. (A) FTIR spectrum of Si/Ca(Sr)-NCF mats. (B) FTIR spectrum of Si/Ca(Sr)-NCF mats. (C) water contact angles of Si/Ca(Sr)-NCF mats, (D) elastic tests for Si/Ca(Sr)-NCF mats.

Abbreviations: FTIR, Fourier transform infrared spectroscopy; NCF, nano cellulose fiber; Si/Ca(Sr)-NCF, the nano cellulose fiber mineralized with silica and carbonates.

preferable results in low and middle Sr^{2+} containing groups: no longitudinal infiltration (Figure 4a3), 8 μ m in Sr^{2+} free group (Figure 4b3), 12 μ m in low Sr^{2+} group (Figure 4c3); 20 μ m in middle Sr^{2+} containing group (Figure 4d3) and 6 μ m in high Sr^{2+} containing group (Figure 4e3).

Then, SEM images of acid challenge showed Sr²⁺ containing groups all presented better tubular obstruction than NCF and Sr²⁺ free groups after acid challenge (Figure 4B). The granular particles in Sr²⁺ containing groups were evident (Figure 4Bc1-c3, d1-d3, e1-d3) while the Sr²⁺ free group showed patent tubular openings (Figure 4Bb1-b3).

Immersing in the AS, the SEM images indicating denser full surface coverage in Sr²⁺ containing groups compared with residual groups (Figure 4C).

TEM Observation of Mineralized in Type I Collagen Fibers

Further exploration of type I collagen infiltration ability was conducted with the mineral liquid released from the representative Si/Ca(Sr 0.05)-NCF mats. The amorphous SAED image changed at time point 1 hour (Figure 5A) changed into poly crystal rings after 6 and 12 hours (Figure 5B and C) implying the successful collagen infiltration mineralization.

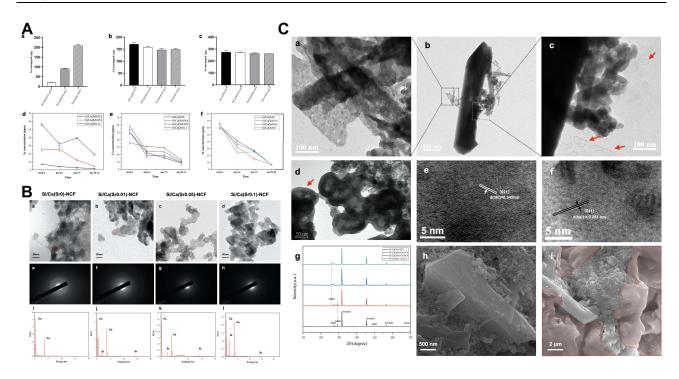


Figure 3 The ion release assay of Si/Ca(Sr)-NCF mats and the long-term crystallization. (A) (a–c) Sr, Ca, and Si ion entrapments and (d–f) nun-cumulative release of ions at day 0–3, day 4–6 and day 7–9. (B) (a–d) TEM images of the released liquid, (e–h) SAED patterns of the particles in different groups (red circles indicating the 50 nm sized particles) and (i–l) EDS results of the nanoparticles of different groups. (C) Crystallization after aging for 7 days of the liquid released from Si/Ca(Sr)-NCF mats. (a–d) TEM images of the crystals and the residual co-precipitation particles. (c and d) The shedding silica skin during the crystallization procedure (red arrows). (e and f) Lattice spacing analyses of the crystals of the aged releasing liquid. (g) XRD patterns of the Si/Ca(Sr)-NCF mats' leaching liquid. (h and i) SEM images of the crystals. The red membranes indicated the sheded silica wrapping the crystals and NPs.

Abbreviations: NCF, nano cellulose fiber; Si/Ca(Sr)-NCF, the nano cellulose fiber mineralized with silica and carbonates; TEM, transmission electron microscopy; SAED, selected area electron diffraction; EDS, energy-dispersive X-ray spectrometry; XRD, X-Ray Diffraction; SEM, scanning electron microscopy; NPs, nano particles.

In vitro Anti-Bacterial Ability

The results of bacteria colonies counting illustrated that all the testing groups have significant antibacterial effects than compare groups at 3 time points in the representative photos and anti-bacterial ratios (Figure 6). The antibacterial ratios of the four mineralized groups were 17.33%, 30.18%, 36.13%, 46.17% for 6 hours; 35.35%, 45.27%, 54.31%, 68.97% for 12 hours, 69.43%, 68.66%, 71.97%, 70.47% for 24 hours, respectively (Figure 6B).

In vitro Cellular Studies with Si/ Ca(Sr)-NCF Mats Cell Proliferation

In the results of CCK-8 results, the OD value of Si/Ca (Sr0)-NCF and Si/Ca(Sr 0.01)-NCF were significantly higher than the control group at day 1, 7 and 14 (Figure 7B). Significant decrease was present in Si/Ca(Sr 0.1)-NCF group compared to control groups and no significant difference was found between Si/Ca(Sr0)-NCF and Si/Ca(Sr 0.01)-NCF groups at all time points (Figure 7B).

Cell Adhesion and Morphology with Si/Ca(Sr)-NCF Mats

Verified by the Live/Dead (Figure 7A) and the cellular skeleton staining (Figure 7H), the proliferation and adhesion of the hDPCs on the Si/Ca(Sr)-NCF mats had been demonstrated. Dead cells were obvious in Si/Ca(Sr 0.1)-NCF group (Figure 7Ae1–e3) which was in accordance with the CCK-8 results (Figure 7B). For cell adhesion, the cell nucleus stained by PI grew into the fibrous structure (Figure 7H) showing decreased nucleus density in the Si/Ca(Sr 0.1)-NCF (Figure 7He1–e4) in accordance with CCK-8 and Live/Dead staining. Dense microfilaments and nucleus were presented in Si/Ca(Sr 0.01)-NCF and Si/Ca(Sr 0.05)-NCF groups indicating the preferable proliferation and adhesion activity.

Alkaline Phosphatase (ALP) Activity and RT-qPCR for Osteogenesis-Related Gene Expression

The ALP activity representing the matrix remineralization ability had been explored on day 7 and day 14. Adding of low Sr²⁺ dose realized the statistically highest ALP activity compared with Sr²⁺ free and NCF groups on day 7 and

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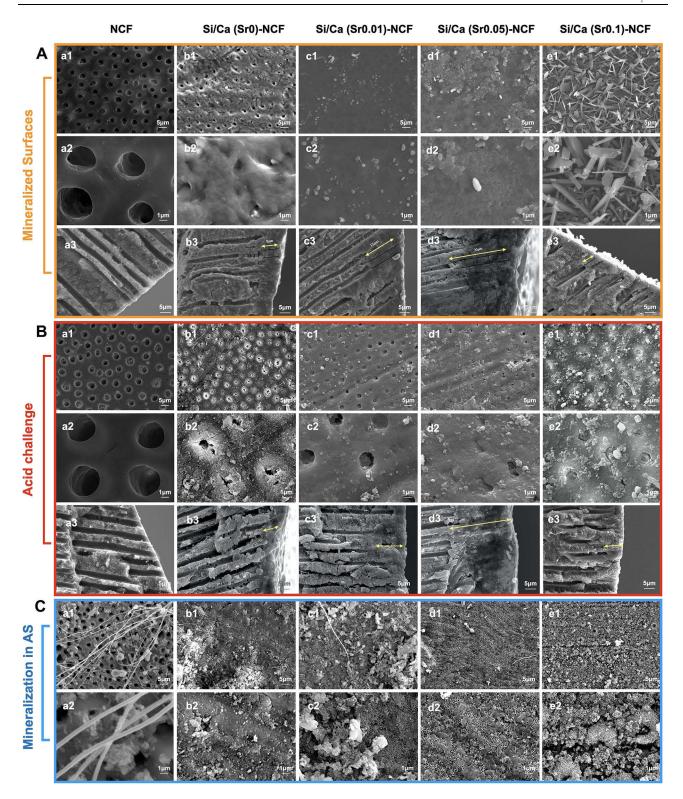


Figure 4 The SEM images of the remineralized dentine surfaces and longitudinal sections after applying the Si/Ca(Sr)-NCF mats. (A) SEM images presenting surface occluding and longitudinal infiltration of dentine discs (a1-a3) in distilled water, (b1-b3) in Si/Ca(Sr 0)-NCF group, (c1-c3) in Si/Ca(Sr 0.01)-NCF group, (d1-d3) in Si/Ca(Sr 0.01)-NCF group, (d1 0.05)-NCF group and (e1-e3) in Si/Ca(Sr 0.1)-NCF group. (B) The SEM images presenting surface occluding and longitudinal infiltration after 6 wt% citric acid leaching. Images of dentine discs (a1-a3) in distilled water, (b1-b3) in Si/Ca(Sr 0)-NCF group, (c1-c3) in Si/Ca(Sr 0.01)-NCF group, (d1-d3) in Si/Ca(Sr 0.05)-NCF group, and (e1-e3) in Si/Ca(Sr 0.1)-NCF group. (C) The SEM images after immersing in artificial saliva for 7 days. Images presenting surface occluding (a1-a2) in distilled water, (b1-b2) in Si/Ca (Sr 0)-NCF group, (cI-c2) in Si/Ca(Sr 0.01)-NCF group, (dI-d2) in Si/Ca(Sr 0.05)-NCF group, and (eI and e2) in Si/Ca(Sr 0.1)-NCF group. Abbreviations: SEM, scanning electron microscopy; NCF, nano cellulose fiber; Si/Ca(Sr)-NCF, the nano cellulose fiber mineralized with silica and carbonates.

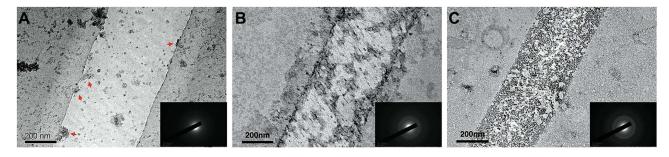


Figure 5 The TEM images of the Si/Ca(Sr0.05)-NCF releasing liquid mineralized type I collagen for I h, 12 h and 24 h with uranium acetate dyeing. The image revealed dispersed nanospheres (red arrows) attaching to the collagen and the SAED results indicated (**A**) the amorphous SAED phase at time point I h, (**B**) SAED results indicated polycrystals at 12 h, (**C**) SAED results indicated polycrystals at 24 h.

Abbreviations: Si/Ca(Sr)-NCF, the nano cellulose fiber mineralized with silica and carbonates; TEM, transmission electron microscopy; SAED, selected area electron diffraction.

14 (Figure 7C). While the OD value in middle and high Sr²⁺ groups showed no significant difference with compare group (Figure 7C).

Osteogenesis and odontoblastic-related genes in hDPCs, including ALP (early marker for osteogenic differentiation and odontoblast differentiation), OCN (late marker for osteogenic differentiation and odontoblastic differentiation), Runx2 (vital transcription maker for bone formation), and OPN (crucial marker in apatite crystals) were significantly upregulated in Si/Ca(Sr 0.01)-NCF group at day 7(Figure 7D–G).^{25,26}

Discussion

In present research, Si/Ca(Sr)-NCF mats were successfully fabricated and Si/Ca(Sr) NPs releasing from the mats conducted desirable anti-acid biomimetic dentine remineralization, antibacterial activity with *E. faecalis* and biocompatibility with hDPCs (Scheme 1). Moreover, the applying of Sr²⁺ yielded several benefits: (1) middle Sr²⁺ adding in the system conducted continuous mechanical tubular occluding, preferable acid resistance and remineralization in AS; (2) ALP activity was improved by limited Sr²⁺ adding.

Remarkably, continuous Si/Ca(Sr) NPs release from Si/Ca(Sr) – NCF mats was verified by the nanoparticles in leaching liquid (Figure 3B), characteristic shell shedding images in TEM results (Figure 3Bc, Bd) and the ion release assay (Figure 3A). In present study, the mineralized NCF mats provided silica rich and alkalescence environment which is essential for the Si/Ca(Sr) NPs self-assembling. ¹⁴ The stabilizing effect of this inorganic silica stabilized amorphous precursor was confirmed in the TEM images (Figure 3B). Even in the images after aging for a week, there were still isolated nanoparticles in the

releasing dilution showing the long term carbonates stabilization (Figure 3C). As a carrier and storage, the mineralized NCF mats with disordered nano fibers and partial crystallization (Figure 1) also showed low elastic modulus (Figure 2D) and high surface hydrophilicity (Figure 2C) in low Sr²⁺ group making it easy to apply as lining and medical dressing. Base films combining with silica and carbonate components was reported before.²⁷ The explanation might be the negative charged NCF surface interacted with the positive charged cations synergistically with the silica and from a whole NCF (-OH)–Ca²⁺/Sr²⁺–silanol (Si-OH) network.

Apart from the stabilization effect of the silica shell, the Sr²⁺ addition further promoted particle dispersion on dentine surfaces. Interestingly, uniform segmental particles connected onto fibrous mat (Figure 1d1), homogeneous dentine surface covering (Figure 4Ad1 and d2), deep longitudinal sedimentation (Figure 4Ad3), preferable results of anti-acid (Figure 4Bd1-d3) and remineralization in AS (Figure 4Cd1 and d2) were all showed in the low and middle Sr2+ containing groups. As the tendency of polycrystal ball forming in Sr²⁺ free group, the micrometer sized polycrystal particles were too large to obstruct the 1-2 µm diameter tubular openings (Figure 1b1). This polycrystal sphere was previously reported and explained as the much stronger interaction between Ca²⁺ and silicate ions than the homologous such as Sr2+ and Be2+.28,29 While, in the high Sr²⁺ concentration group, large blade shaped crystals also showed shorter infiltration (Figure 4Ae1-e3). Based on previous research indicating the increased anisotropic after Sr²⁺ addition, the possible explanation for the homogeneous particle dispersion may be that the appropriate amount of Sr²⁺ may competitively inhibit the coagulation of CaCO₃, inducing a less violent

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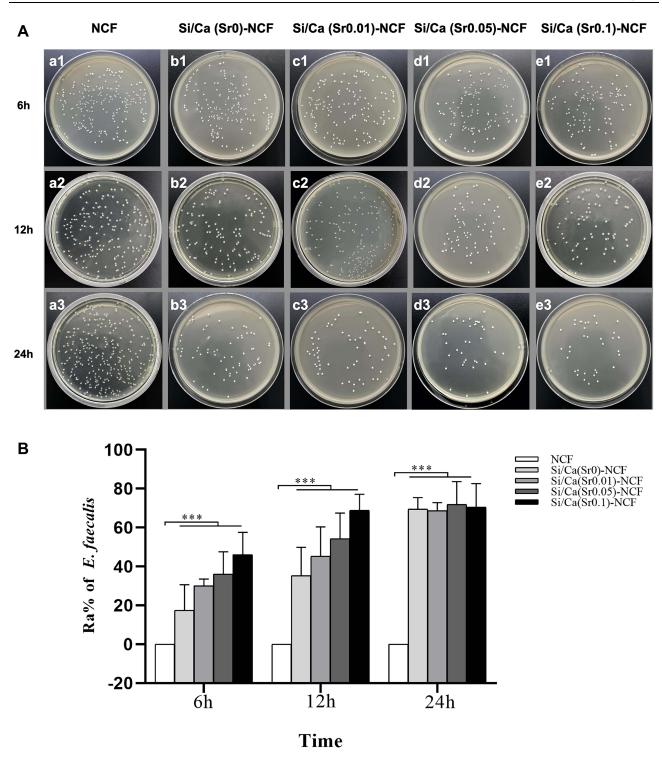


Figure 6 The anti-bacteria tests for Enterococcus faecalis. (A) (a I -e I) The representative photographs of recultivated colonies of on agar plates at time points of 4 h, (a2-e2) 12 h and (a3–e3) 24 h, and (B) the corresponding anti-bacteria ratios. ***p < 0.001. Abbreviations: NCF, nano cellulose fiber; Si/Ca(Sr)-NCF, the NCF mineralized with silica and carbonates.

mineralization process and more uniform particle distribution.

In addition to the mechanical obstruction by Si/Ca(Sr) NPs in site precipitation, the precursor NPs further conducted type I collagen biomimetic remineralization with the leaching liquid of middle Sr²⁺ group. Generally, small stabilizers (<65k Da) were demanded for type I collagen intrafibrillar mineralization. 30,31 Oligomeric

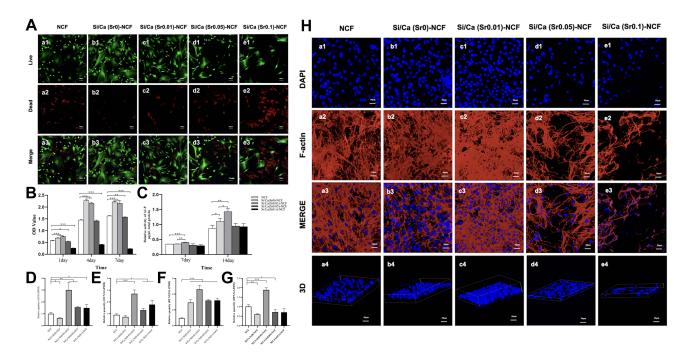


Figure 7 The hDPCs response to the Si/Ca(Sr)-NCF mats. (A) The Live/Dead fluorescent dyeing of hDPCs after cultured with Si/Ca(Sr)-NCF mats for 1 day. (B) The CCK-8 results revealed the cell proliferation at time points of day 1, 7 and 14. (C) The ALP activity of the hDPCs at day 7 and 14. (D-G) Gene expression levels of osteogenesis-related markers: (D) ALP, (E) OCN, (F) Runx2, (E) OPN. (H) The cell adhesion with the Si/Ca(Sr)-NCF mats represented by the florescence images of the cell skeletons. *p < 0.05, **p < 0.01, ***p < 0.001.

Abbreviations: hDPCs, human dental pulp cells; NCF, nano cellulose fiber; Si/Ca(Sr)-NCF, the nano cellulose fiber mineralized with silica and carbonates; ALP, alkaline phosphatase. OCN, osteocalcin; Runx2, runt-related transcription factor; OPN, osteopontin.

silica was small enough and early collagen mineralization with polycrystal SAED rings confirmed that Si/Ca(Sr) NPs as an inorganic silica stabilized precursor can induce dentine biomimetic remineralization (Figure 5). Moreover, the remineralized dentine tubular structure showing no regular parallel arrangement, but a continuous fused peritubular and tubular pattern, also indicated the prominent biomimetic dentine remineralization (Figure 4A and B).

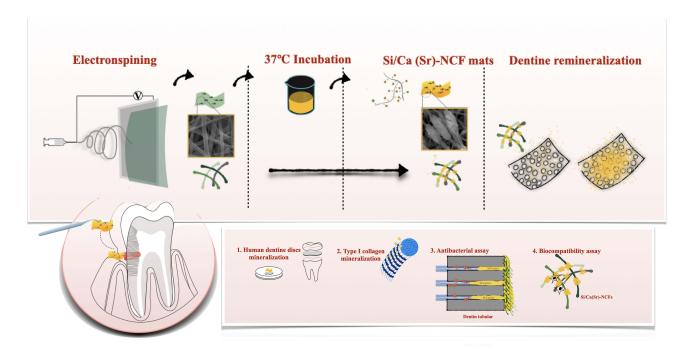
According to the previous studies of silica-carbonate co-precipitation structure, the silica shells remain after the acid challenge. ^{14,32} In the SEM images after acid leaching, although the precipitation of different groups showed corrosions in varying degrees, the anti-acid effects were still reliable as the residual particle fusion was evident in Sr²⁺ containing groups (Figure 4B) and longitudinal depth illustrated no significant reduction (Figure 4Aa3–e3, Ba3–e3).

Based on the dentine occluding evaluation above, the all-in-one Si/Ca(Sr)-NCF mats realized 6–20 µm infiltration within 24 hours and conducted anti-acid biomimetic remineralization not only by mechanical in site precipitation, but also via dentine collagen infiltration. Preferable occluding effects were presented in low and middle Sr²⁺ groups. The tubular obstructing by hydrophilic mat

covering in present study was convenient and timeefficient, as many strategies nowadays still requiring several steps for days with undefined obstructing depth and acid resistance.

With respect to biological properties, antibacterial and biocompatibility activity were evaluated (Scheme 1). *E. faecalis* is a dominant bacterium inside the root canal. Evading in tubular system and the remaining viable are recognized to be the reasons causing periapical diseases. ^{33,34} In consideration of the infiltration potential, the antibacterial activity of the Si/Ca(Sr)-NCF mats was evaluated by co-cultured with the *E. faecalis*. In BG antibacterial study based on large dataset, researchers believe the antibacterial effect has correlations with the unfavorable interface shaped by immediate alkaline species release. ^{35,36} Higher solubility and denser cation ion release would be the reasons why the high Sr²⁺ containing group showed better antibacterial effect on *E. faecalis*.

In view of the penetration activity, cytocompatibility of Si/Ca(Sr)-NCF remineralization mats was another issue to be explored. After co-culturing with the hDPCs, the adhesion, proliferation and ALP activities were evaluated (Figure 7). Based on the research, Sr²⁺ promotes osteogenesis in mature



Scheme I Schematic illustration of Si/Ca(Sr)-NCF mats application to conduct multifunctional remineralization in dentine hypersensitivity model. Si/Ca(Sr)-NCF mats were assembled by electrospinning NCF mats and mineral loading. The dentine tubular obstruction effects were evaluated with (1) in vitro dentine discs, (2) the type I collagen model, (3) the antibacterial activity and (4) cytocompatibility with human dental pulp cells. Si/Ca(Sr) NPs were continuously released from the minerals (Ca^{2+}/Sr^{2+}) and silica rich mat surfaces facilitating the dentine remineralization not only through mechanical occluding but also homogeneous nucleation of the type I collagen. The antibacterial activity and cytocompatibility further indicated the multifunctional all-in-one Si/Ca(Sr)-NCF mats is a promising candidate to treat the dentine hypersensitivity. **Abbreviations**: NCF, nano cellulose fiber; NPs, nano particles; Si/Ca(Sr)-NCF, the nano cellulose fiber mineralized with silica and carbonates.

osteocytes in a dose dependent manner within the concentration from 10–1000 uM. ^{37,38} Similar Sr²⁺ dose dependent osteo-induction and cell viability is also reported with ALP activity promotion in concentration of 0.1–2.5 mM in hDPCs. ³⁹ Based on the Sr ion release within 12 days, the high Sr²⁺ containing group (4.75 mM, Figure 3Ad–f) showing cytotoxicity, and low Sr²⁺ containing group (0.48 mM, Figure 3Ad–f) showing ALP promotion and cytocompatibility with hDPCs in present study could be explained.

As high calcified tissue, the development of hard tissue is closely related to the bone formation. The Si/Ca(Sr0.01)-NCF releasing liquid with low Sr²⁺ concentration significantly improved the gene expression for the early and late odontoblastic differentiation. While in the high Sr²⁺ group, the proliferation and odontoblastic differentiation were inhibited. These results were in accordance with the dose dependence results.³⁷ In addition, the potential mechanism for Sr²⁺ may be that the Sr²⁺ promote osteoblast differentiation via the CaSR pathway, which is similar to the Ca ions.²⁶ While in present study, the low Sr²⁺ group significantly improved the ALP activities and the related gene expression compared with the pure calcium group (Figure 7D–G).The

underlying cell differentiation promoting mechanism of Sr²⁺ needs to be further explored.

Fundamentally, this in vitro study is still limited as the oral activities are accompanied by more complicated physico-chemical interactions. The ion release in distilled water in present study may also be quite different with the release in saliva liquid. Therefore, further studies such as acid challenge for longer time, wear resistance tests, and the in vivo experiments are indispensable.

Conclusion

Present study confirmed that the Si/Ca(Sr)-NCF system with low and middle Sr²⁺ containing (0.48–2.59 mM Sr²⁺ release) successfully realized acid-resistant, bacteriostatic, and low cytotoxic dentine biomimetic remineralization. This multifunction all-in-one Si/Ca(Sr)-NCF system was convenient and effective to utilize as the Si/Ca(Sr) NPs continuously releasing from the carrier mats. Moreover, dentine tubular bacterial *E. faecalis* was inhibited, and hDPCs proliferation and ALP activity representing matrix mineralization was promoted. The multifunction Si/Ca(Sr)-NCF system, with soft NCF basement, which may be easily

applied onto complicated dentine surfaces such as the areas adjacent to pulp cavity and with potential bacterial infections, could be a promising candidate managing the DH in the future.

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Disclosure

The authors declare that they have no conflict of interests.

References

- Favaro Zeola L, Soares PV, Cunha-Cruz J. Prevalence of dentin hypersensitivity: systematic review and meta-analysis. *J Dent.* 2019;81:1–6. doi:10.1016/j.jdent.2018.12.015
- West N, Seong J, Davies M. Dentine hypersensitivity. Monogr Oral Sci. 2014;25:108–122. doi:10.1159/000360749
- 3. Thomas MS. Dentin hypersensitivity. *J Evid Base Dent Pract*. 2011;142:220–228. doi:10.1016/S1532-3382(12)70043-X
- Brannstrom M. Dentin sensitivity and aspiration of odontoblasts. J Am Dent Assoc. 1963;66:366–370. doi:10.14219/jada.archive. 1963.0104
- Walters PA. Dentinal hypersensitivity: a review. J Contemp Dent Pract. 2005;6:107–117. doi:10.5005/jcdp-6-2-107
- Yu J, Yang H, Li K, et al. A novel application of nanohydroxyapatite/ mesoporous silica biocomposite on treating dentin hypersensitivity: an in vitro study. *J Dent.* 2016;50:21–29. doi:10.1016/j. jdent.2016.04.005
- Liang K, Xiao S, Liu H, et al. 8DSS peptide induced effective dentinal tubule occlusion in vitro. *Dent Mater.* 2018;34(4):629–640. doi:10.1016/j.dental.2018.01.006
- Niu LN, Zhang W, Pashley DH, et al. Biomimetic remineralization of dentin. Dent Mater. 2014;30:77–96. doi:10.1016/j.dental.2013.07.013
- Poulsen S, Errboe M, Lescay Y, et al. Potassium containing toothpastes for dentine hypersensitivity. *Cochrane Database Syst Rev.* 2006;(3):CD001476. doi:10.1002/14651858.CD001476.pub2
- Zhou YZ, Cao Y, Liu W, et al. Polydopamine-induced tooth remineralization. Acs Appl Mater Interfaces. 2012;4:6901–6910. doi:10.1021/am302041b
- Liu Z, Shao C, Jin B, et al. Crosslinking ionic oligomers as conformable precursors to calcium carbonate. *Nature*. 2019;574 (7778):394–398. doi:10.1038/s41586-019-1645-x
- 12. Gower LB. Biomimetic model systems for investigating the amorphous precursor pathway and its role in biomineralization. *Cheminform*. 2009;40(5):4551–4627. doi:10.1021/cr800443h
- Xu Z, Long J, Zhang N, et al. Strong mineralization ability of strontium zinc silicate: formation of a continuous biomorphic mineralized layer with enhanced osteogenic activity. *Colloids Surf B Biointerfaces*. 2019;176:420–430. doi:10.1016/j.colsurfb.2019. 01.026
- Hyde ST, Carnerup AM, Larsson AK, et al. Self-assembly of carbonate-silica colloids: between living and non-living form. *Phys A*. 2004;339:24–33. doi:10.1016/j.physa.2004.03.045
- Eiblmeier J, Kellermeier M, Deng M, et al. Bottom-up self-assembly of amorphous core–shell–shell nanoparticles and biomimetic crystal forms in inorganic silica–carbonate systems. *Chem Mater*. 2013;25:1842–1851. doi:10.1021/cm4003959
- Kellermeier M, Melero-García E, Glaab F, et al. Stabilization of amorphous calcium carbonate in inorganic silica-rich environments. *J Am Chem Soc.* 2010;132(50):17859–17866. doi:10.1021/ja106959p

 Kellermeier M, Melero-García E, Kunz W, et al. Local autocatalytic co-precipitation phenomena in self-assembled silica–carbonate materials. *Colloids Surf B Biointerfaces*. 2012;380:1–7. doi:10.1016/j. jcis.2012.05.009

- Knaus J, Sommer M, Duchstein P, et al. Polar structure formation in solid solution of strontium-substituted fluorapatite-gelatin composites: from structural and morphogenetic aspects to pyroelectric properties. Chem Mater. 2020;32:8619–8632. doi:10.1021/acs.chemmater.0c02993
- Ma KN, Huang D, Cai J, et al. Surface functionalization with strontium-containing nanocomposite coatings via EPD. *Colloids Surf B Biointerfaces*. 2016;146:97–106. doi:10.1016/j.colsurfb.2016.05.036
- Deng H, Zhou X, Wang X, et al. Layer-by-layer structured polysaccharides film-coated cellulose nanofibrous mats for cell culture. *Carbohydr Polym*. 2010;80(2):474–479. doi:10.1016/j.carbpol.2009.12.004
- Nakouzi E, Fares HM, Schlenoff JB, et al. Polyelectrolyte complex films influence the formation of polycrystalline micro-structures. *Soft Matter*. 2018;14(16):3164–3170. doi:10.1039/C7SM02466E
- Wang Z, Sa Y, Sauro S, et al. Effect of desensitising toothpastes on dentinal tubule occlusion: a dentine permeability measurement and SEM in vitro study. J Dent. 2010;38:400–410. doi:10.1016/j.jdent.2010.01.007
- João-Souza SH, Machado C, Lopes M, et al. Effectiveness and acid/tooth brushing resistance of in-office desensitizing treatments-A hydraulic conductance study. Arch Oral Biol. 2018;96:130–136. doi:10.1016/j.archoralbio.2018.09.004
- Nudelman F, Pieterse K, George A. The role of collagen in bone apatite formation in the presence of hydroxyapatite nucleation inhibitors. Nat Mater. 2010;9:1004–1009. doi:10.1038/nmat2875
- Gan X, He KH, Qian BS, et al. The effect of glycine on the growth of calcium carbonate in alkaline silica gel. J Cryst Growth. 2017;458:60–65. doi:10.1016/j.jcrysgro.2016.11.027
- 26. Qin W, Liu P, Zhang R, et al. JNK MAPK is involved in BMP-2-induced odontoblastic differentiation of human dental pulp cells. Connect Tissue Res. 2014;55:217–224. doi:10.3109/ 03008207.2014.882331
- 27. Huang D, Ma K, Cai X, et al. Evaluation of antibacterial, angiogenic, and osteogenic activities of green synthesized gap-bridging copper-doped nanocomposite coatings. *Int J Nanomedicine*. 2017;12:7483–7500. doi:10.2147/IJN.S141272
- Eiblmeier J, Dankesreiter S, Pfitzner A, et al. Crystallization of mixed alkaline-earth carbonates in silica solutions at high Ph. Cryst Growth Des. 2014;14:6177–6188. doi:10.1021/cg5004116
- Nakouzi E, Rendina R, Palui G, et al. Effect of inorganic additives on the growth of silica–carbonate biomorphs. *J Cryst Growth*. 2016;452:166–171. doi:10.1016/j.jcrysgro.2015.12.042
- Niu LN, Jee SE, Jiao K, et al. Collagen intrafibrillar mineralization as a result of the balance between osmotic equilibrium and electroneutrality. Nat Mater. 2017;16:370–378. doi:10.1038/nmat4789
- Liu Y, Luo D, Kou XI, et al. Hierarchical intrafibrillar nanocarbonated apatite assembly improves the nanomechanics and cytocompatibility of mineralized collagen. *Adv Funct Mater*. 2012;23:1404–1411. doi:10.1002/adfm.201201611
- Bittarello E, Massaro FR, Aquilano D. The epitaxial role of silica groups in promoting the formation of silica/carbonate biomorphs:
 a first hypothesis. J Cryst Growth. 2010;312(3):402–412. doi:10.1016/j.jcrysgro.2009.11.004
- George S, Kishen A, Song P. The role of environmental changes on monospecies biofilm formation on root canal wall by enterococcus faecalis. *J Endodontics*. 2005;31:867–872. doi:10.1097/01. doi:0000164855.98346.fc
- 34. Fan W, Li YY, Sun Q, et al. Calcium-silicate mesoporous nanoparticles loaded with chlorhexidine for both anti- Enterococcus faecalis and mineralization properties. *J Nanobiotechnol*. 2016;14(1). doi:10.1186/s12951-016-0224-7
- Echezarreta-López MM, Landin M. Using machine learning for improving knowledge on antibacterial effect of bioactive glass. *Int J Pharm*. 2013;453:641–647. doi:10.1016/j.ijpharm.2013.06.036

- 36. Liu J, Rawlinson SC, Hill RG, et al. Strontium-substituted bioactive glasses in vitro osteogenic and antibacterial effects. Dent Mater. 2016;32:412-422. doi:10.1016/j.dental.2015.12.013
- 37. Aimaiti A, Maimaitiyiming A, Boyong X, et al. Low-dose strontium stimulates osteogenesis but high-dose doses cause apoptosis in human adipose-derived stem cells via regulation of the ERK1/2 signaling pathway. Stem Cell Res Ther. 2017;8:282. doi:10.1186/s13287-017-0726-8
- 38. Mohan BG, Suresh BS, Varma HK, et al. In vitro evaluation of bioactive strontium-based ceramic with rabbit adipose-derived stem cells for bone tissue regeneration. J Mater Sci Mater Med. 2013;24:2831-2844. doi:10.1186/s13287-017-0726-8
- 39. Huang M, Hill RG, Rawlinson SC. Strontium (Sr) elicits odontogenic differentiation of human dental pulp stem cells (hDPSCs): a therapeutic role for Sr in dentine repair? Acta Biomater. 2016;38:201-211. doi:10.1016/j.actbio.2016.04.
- 40. Pokrowiecki R, Wojnarowicz J, Zareba T, et al. Nanoparticles and human saliva: a step towards drug delivery systems for dental and craniofacial biomaterials. Int J Nanomedicine. 2019;14:9235–9257. PMID: 31819427; PMCID: PMC6886554. doi:10.2147/IJN. S221608

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