

The Natural Killer T (NKT) Cell Ligand α -Galactosylceramide Demonstrates Its Immunopotentiating Effect by Inducing Interleukin (IL)-12 Production by Dendritic Cells and IL-12 Receptor Expression on NKT Cells

By Hidemitsu Kitamura,^{*§} Kenji Iwakabe,[‡] Takashi Yahata,[‡] Shin-ichiro Nishimura,[§] Akio Ohta,^{*} Yasushi Ohmi,^{*} Marimo Sato,^{*} Kazuyoshi Takeda,^{||} Ko Okumura,^{||} Luc Van Kaer,[¶] Tetsu Kawano,^{**} Masaru Taniguchi,^{**} and Takashi Nishimura^{*‡}

From the ^{*}Section of Genetic Engineering, Research Center for Genetic Engineering and Cell Transplantation, and [‡]Department of Immunology, Tokai University School of Medicine, Isehara 259-1193, Japan; the [§]Division of Biological Sciences, Graduate School of Science, Hokkaido University, Sapporo 060-0810, Japan; the ^{||}Department of Immunology, Juntendo University School of Medicine, Tokyo 113-0033, Japan; the [¶]Howard Hughes Medical Institute, Department of Microbiology and Immunology, Vanderbilt University School of Medicine, Nashville, Tennessee 37232; and ^{**}Core Research for Evolutional Science and Technology (CREST) Project and Department of Molecular Immunology, Graduate School of Medicine, Chiba University, Chiba 260-8670, Japan

Summary

The natural killer T (NKT) cell ligand α -galactosylceramide (α -GalCer) exhibits profound antitumor activities in vivo that resemble interleukin (IL)-12-mediated antitumor activities. Because of these similarities between the activities of α -GalCer and IL-12, we investigated the involvement of IL-12 in the activation of NKT cells by α -GalCer. We first established, using purified subsets of various lymphocyte populations, that α -GalCer selectively activates NKT cells for production of interferon (IFN)- γ . Production of IFN- γ by NKT cells in response to α -GalCer required IL-12 produced by dendritic cells (DCs) and direct contact between NKT cells and DCs through CD40/CD40 ligand interactions. Moreover, α -GalCer strongly induced the expression of IL-12 receptor on NKT cells from wild-type but not CD1^{-/-} or V α 14^{-/-} mice. This effect of α -GalCer required the production of IFN- γ by NKT cells and production of IL-12 by DCs. Finally, we showed that treatment of mice with suboptimal doses of α -GalCer together with suboptimal doses of IL-12 resulted in strongly enhanced natural killing activity and IFN- γ production. Collectively, these findings indicate an important role for DC-produced IL-12 in the activation of NKT cells by α -GalCer and suggest that NKT cells may be able to condition DCs for subsequent immune responses. Our results also suggest a novel approach for immunotherapy of cancer.

Key words: natural killer T cells • dendritic cells • α -galactosylceramide • interleukin 12 • interleukin 12 receptor

Natural killer T (NKT)¹ cells represent a novel lymphoid lineage distinct from mainstream T cells, B cells, and NK cells. NKT cells are characterized by the expression of an invariant TCR encoded by V α 14 and J α 281 gene segments and V β 8, 7, or 2 gene segments (1, 2). It was demonstrated recently that NKT cells are strongly stimulated by the glycolipid α -galactosylceramide (α -Gal-

Cer), a potent inducer of antitumor immunity in mice (3–5). Recognition of α -GalCer by NKT cells appeared to depend on the interaction of the invariant TCR of these cells with α -GalCer presented by the nonclassical MHC molecule CD1d on APCs (6). Stimulation of NKT cells by α -GalCer resulted in the production of large amounts of IFN- γ and some IL-4, and the development of a cytotoxic phenotype (7).

The in vivo antitumor activity of α -GalCer strongly resembles the antitumor activity mediated by the cytokine IL-12 (8, 9). Moreover, both α -GalCer and IL-12 are

¹Abbreviations used in this paper: α -GalCer, α -galactosylceramide; DC, dendritic cell; GAPDH, glyceraldehyde 3-phosphate dehydrogenase; NKT, natural killer T; RT, reverse transcription.

strong inducers of NKT cell activity and exert their antitumor activities through activation of these cells (8, 9). Because of these striking similarities between α -GalCer and IL-12 for activation of NKT cells, we decided to investigate whether α -GalCer activation of NKT cells involves regulation by IL-12. First, we demonstrated that NKT cells are the main, if not the only, target for activation by α -GalCer in spleen cell populations of mice. Second, we showed that endogenous IL-12 produced by dendritic cells (DCs) is critically important for the activation of NKT cells by α -GalCer and that the interaction between DCs and NKT cells involves CD40 and its ligand. Third, α -GalCer induced the expression of IL-12R on NKT cells, which required the production of IFN- γ by NKT cells. Fourth, α -GalCer acted synergistically with IL-12 in the activation of natural killing activity and IFN- γ production in vivo. Collectively, these findings indicate that α -GalCer exerts its function through IL-12 and suggest a novel approach for therapeutic intervention in cancer and other disease processes.

Materials and Methods

Mice. C57BL/6 mice were purchased from Charles River Japan. V α 14 NKT cell-deficient (J α 281^{-/-}) and CD1d^{-/-} mice were established by specific deletion of the J α 281 and CD1d gene segment, respectively (3, 10). All mice used in this study were at 5–8 wk of age and were maintained in specific pathogen-free conditions.

α -GalCer. α -GalCer [(2S,3S,4R)-1-O-(α -D-galactopyranosyl)-2-(N-hexacosanoylamino)-1,3,4-octadecanetriol] used for this study was provided by Dr. Y. Koezuka (Kirin Brewery Co., Ltd., Gunma, Japan [4, 5]). The stock solution of α -GalCer (220 μ g/ml) was diluted in 0.5% polysorbate 20 (Nikko Chemical) in 0.9% NaCl solution. This stock solution was further diluted into an appropriate concentration with saline and used for the experiments. A vehicle control solution was prepared from a solution of 0.5% polysorbate 20 in 0.9% NaCl solution. The vehicle control was used in all experiments.

Isolation of Lymphoid Cell Subsets by FACS[®]. Spleen cells were incubated on nylon wool columns for 45 min, and the nonadherent cells were used for the isolation of NKT cells, NK cells, CD4⁺ T cells, and CD8⁺ T cells by cell sorting using a FACS Vantage[™] instrument (Becton Dickinson). All mAbs used in these experiments (mAbs against NK1.1, CD4, CD8, and TCR- α/β) were purchased from PharMingen. Unless noted otherwise, NK1.1⁺TCR- α/β ⁺ cells were used as purified NKT cells. The stained cells were isolated using the FACS Vantage[™]. The purity of the sorted cells was >98%. The details of the staining and sorting have been described previously (11).

Coculture of DCs and NKT Cells. DCs were prepared according to the method of Steinman et al. (12) with some modifications. In brief, spleen cells were incubated in 10-cm plastic dishes (Falcon; Becton Dickinson) for 2 h, and the nonadherent cells were removed from the culture. The adherent cells were further incubated overnight and the nonadherent cells were harvested. Then, CD11c⁺B220⁻CD4⁻CD8⁻ cells were isolated from the nonadherent populations by cell sorting and used as the source of DCs. Generally, DCs (10⁵) were cocultured with purified NK1.1⁺TCR- α/β ⁺ NKT cells (2 \times 10⁵) in the presence of 50 ng/ml of α -GalCer in 96-well U-bottomed plates (Costar

Corp.). After incubation for 36 h, the culture supernatants were harvested to detect cytokine levels.

Detection of Cytokine Activity. IL-4 or IFN- γ activity in culture supernatants was determined using the Biotrac[™] mouse IL-4 or Biotrac[™] mouse IFN- γ ELISA system (Nycomed Amersham plc). Serum samples were obtained from C57BL/6 mice 24 h after injection of α -GalCer (200 ng/mouse) and/or IL-12 (200 U/mouse; donated by Genetics Institute, Inc., Cambridge, MA), and cytokine levels were measured using ELISA kits (Nycomed Amersham plc). IL-12 p70 activity in culture supernatants was measured using Intertest-12X[™] ELISA kits (Genzyme Corp.).

Cytotoxicity Assay. The natural killing activity of spleen cells was determined by 4-h ⁵¹Cr-release assays using YAC-1 cells as target. 1 lytic unit (LU) was defined as the number of effector cells required to cause 25% lysis of 2,500 target cells as described previously (13).

Measurement of the Synergistic Effect of α -GalCer and IL-12 In Vivo. Wild-type C57BL/6 mice received an injection of α -GalCer (200 ng/mouse i.v.), and 6 h later mice were injected with recombinant IL-12 (200 U/mouse i.p.) or saline. 1 d after treatment with IL-12, IFN- γ production in the serum and NK activity of spleen cells were determined. Control mice were treated with vehicle only.

Quantitative Reverse Transcription PCR Assay for IL-12R mRNA Measurement. C57BL/6, CD1d^{-/-}, and V α 14 NKT cell-deficient mice were injected with α -GalCer (2 μ g/mouse i.v.) or vehicle. At different time points (0–6 h) after treatment, mice were killed and spleen cells were isolated. TaqMan[™] real-time quantitative reverse transcription (RT)-PCR assay was carried out for the detection of IL-12R mRNA expression by these cells according to published methods (14). In brief, total RNA extracts from the cells were added to the master mixture. To detect the amount of the IL-12R mRNA RT-PCR amplicon, target (IL-12R β 1, IL-12R β 2) and control (glyceraldehyde 3-phosphate dehydrogenase [GAPDH]) hybridization probes were mixed with target and control PCR primers, respectively. This mixture was transferred to a set of thermocycler tubes and transcribed at 42°C for 30 min, followed by 40 cycles of amplification at 95°C for 15 s and 60°C for 1.5 min, and analyzed using an ABI PRISM 7700 sequence detector (Applied Biosystems). IL-12R β 1 and IL-12R β 2 mRNA expression were estimated from the ratio of fluorescence intensity to GAPDH. IL-12R expression induced by α -GalCer is indicated in the figures as induction index, calculated as follows: induction index = IL-12R expression of α -GalCer-stimulated sample/IL-12R expression of unstimulated sample.

TaqMan[™] probes used for these analyses are as follows: IL-12R β 1 mRNA-605T, 5'-CGGATGCCCAACGAAT-TGGA-3'; IL-12R β 2 mRNA-551T, 5'-AGCCACCTCAAAA-CATATCATGTGTCCAGG-3'; GAPDH-542T, 5'-CCTGGC-CAAGTCATCCATGACAACCTTT-3'.

PCR primers used for these analyses are as follows: IL-12 β 1 mRNA, forward primer (-563F) 5'-AATGTGTCTGAAGAGGC-CGGT-3' and reverse primer (-657R) 5'-GAGTTAACCTGAG-GTCCGCAGT-3'; IL-12R β 2 mRNA, forward primer (-529F) 5'-ATCTCAGTTGGTGTGCTCCA-3' and reverse primer (-602R) 5'-GCCACAGTTCATTTTCTCCT-3'; GAPDH, forward primer (-368F) 5'-CTTACCACCATGGAGAAGGC-3' and reverse primer (-605R) 5'-GGCATGGACTGTGGTCAT-GAG-3'.

Blocking of IL-12R Induction by Anti-IFN- γ mAb. Wild-type C57BL/6 mice were injected with 500 μ g i.p. anti-IFN- γ mAb (R4-6A2; PharMingen) or IL-12 (C15.1 and C15.6, donated by Dr. G. Trinchieri, Wistar Institute of Anatomy and Biology, Philadelphia, PA) at 0 and 1 d before priming with α -GalCer. As a

control, the same amount of rat IgG1 (PharMingen) was injected intraperitoneally into control mice before injection of α -GalCer.

Results

α -GalCer Selectively Activates $NK1.1^+TCR-\alpha/\beta^+$ NKT Cells. To provide direct evidence that NKT cells are the only target cells for activation by α -GalCer, various lymphoid subsets were isolated from mouse spleen cell suspensions by flow cytometry and cocultured with DCs in the presence of α -GalCer. After 36 h of culture, the supernatants were harvested and their IL-4 and IFN- γ contents were measured by ELISA. Fig. 1 shows that purified $NK1.1^+$ T cells produce higher levels of IL-4 and IFN- γ than unfractionated spleen cells. The IFN- γ produced in these cultures was not derived from classical NK cells, because enrichment of $NK1.1^+TCR-\alpha/\beta^-$ NK cells showed no significant cytokine production. In contrast, $NK1.1^+TCR-\alpha/\beta^+$ cells, which represent the NKT cell population, revealed markedly high levels of IL-4 and IFN- γ production. Although $CD4^+$ T cells produced higher levels of cytokines compared with unfractionated spleen cells, this appeared to be due to the presence of $CD4^+NK1.1^+$ NKT cells, because $CD4^+NK1.1^-$ cells produced neither IL-4 nor IFN- γ in response to α -GalCer. Culture of $NK1.1^+TCR-\alpha/\beta^+$ NKT cells alone or with DCs in the absence of α -GalCer caused no significant production of IFN- γ or IL-4, indicating that DCs are essential for the stimulation of cytokine production by NKT cells.

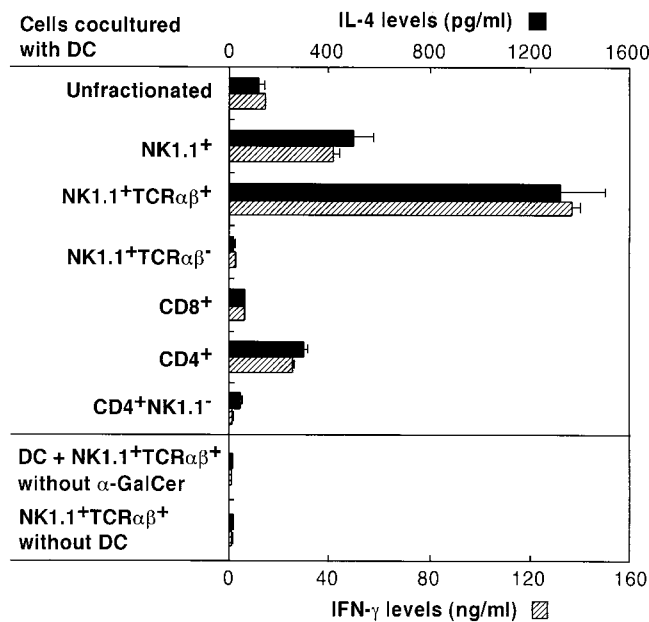


Figure 1. α -GalCer selectively activates $NK1.1^+TCR-\alpha/\beta^+$ NKT cells. Spleen cells from C57BL/6 mice were separated into a variety of lymphoid cell subsets by cell sorting as described in Materials and Methods. Their responsiveness to α -GalCer in the presence of DCs was then determined by measuring IL-4 (■) and IFN- γ (▨) levels in the culture supernatants using ELISA. As a control, $NK1.1^+TCR-\alpha/\beta^+$ NKT cells were cultured alone or with DCs in the absence of α -GalCer. The bars represent mean \pm SE of triplicate samples.

Endogenous IL-12 Production by DCs Is Essential for the Triggering of NKT Cells. Fig. 2 A shows that coculture of DCs and NKT cells in the presence of α -GalCer results in high levels of IFN- γ production. However, addition of anti-IL-12 mAb into these cultures caused a marked inhibition of IFN- γ production. Such inhibition was not observed when control anti-CD8 rat IgG mAb was added. Therefore, these results indicated that endogenously produced IL-12 by DCs was essential for the early activation of NKT cells by α -GalCer. The effect of mAbs against CD40 and CD40L on the activation of NKT cells by α -GalCer was also investigated (Fig. 2 B). Both anti-CD40 mAb and anti-CD40L mAb greatly inhibited the production of IFN- γ by NKT cells in response to α -GalCer. These findings suggested that direct contact between DCs and NKT cells through CD40/CD40L interactions is critically important

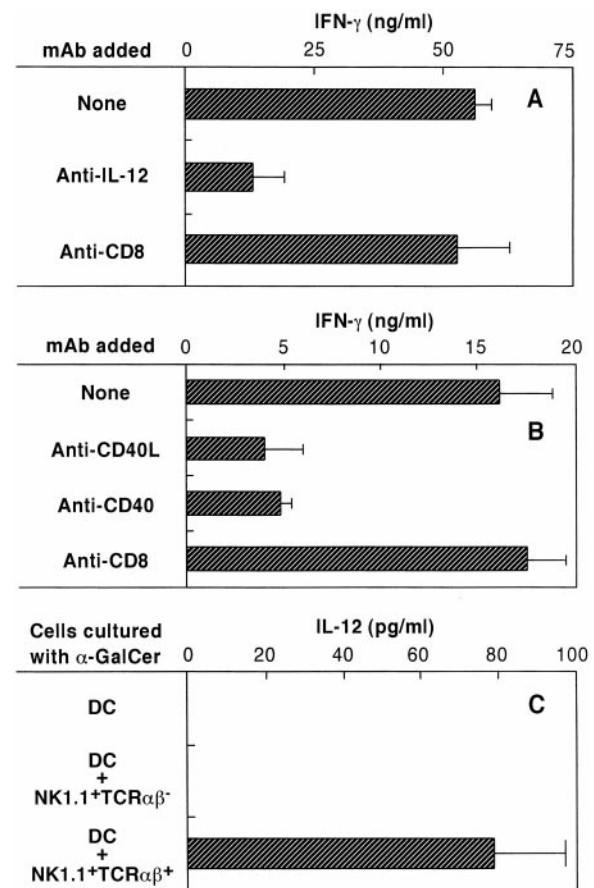


Figure 2. Endogenously produced IL-12 and CD40/CD40L interaction during coculture of DCs and NKT cells is essential for NKT cell activation by α -GalCer. Purified NKT cells were cocultured with DCs in the presence of α -GalCer for 36 h. The IFN- γ levels in culture supernatants were then determined by ELISA. (A) The ability of anti-IL-12 mAb to block NKT cell activation by α -GalCer. Anti-CD8 mAb was used as control rat IgG Ab. (B) The ability of anti-CD40 mAb and anti-CD40L mAb to block NKT cell activation by α -GalCer. As a control, rat anti-CD8 IgG mAb was added to the culture. (C) IL-12 production by DCs cultured with α -GalCer and NKT cells. DCs (5×10^5) were activated with 50 ng/ml of α -GalCer for 8 h in the presence or absence of $NK1.1^+TCR-\alpha/\beta^-$ NK cells (10^5) or $NK1.1^+TCR-\alpha/\beta^+$ NKT cells (10^5). The bars represent mean \pm SE of triplicate samples.

for the activation of NKT cells by α -GalCer. To study the requirements for IL-12 production by DCs in these cultures in further detail, IL-12 p70 activity in culture supernatants was measured by ELISA. As shown in Fig. 2 C, DCs produced IL-12 p70 when cultured with NKT cells and α -GalCer. However, DCs did not produce IL-12 p70 when cultured with α -GalCer alone or when cultured with α -GalCer and NK (NK1.1⁺TCR- α/β ⁻) cells.

α -GalCer Induces IL-12R Expression on NKT Cells. The effect of α -GalCer on the induction of IL-12R mRNA expression in spleen cells was examined by RT-PCR. As shown in Fig. 3, intravenous injection of α -GalCer into C57BL/6 mice caused the induction of mRNA for both IL-12R β 1 and IL-12R β 2 in spleen cells within 4 h. This upregulation of IL-12R was strongly blocked by administration of anti-IL-12 mAb or anti-IFN- γ mAb before injection of α -GalCer (Fig. 4). Moreover, the IL-12R induction by α -GalCer was almost completely abolished in both CD1d^{-/-} and V α 14 NKT cell-deficient mice (Fig. 5, A and B). Thus, these results suggested that CD1d-dependent α -GalCer-induced IFN- γ production by NKT cells may be critically important for the upregulation of IL-12R on NKT cells. To provide direct evidence for this hypothesis, we measured the expression of IL-12R on purified NKT cells that were previously activated in the presence of DCs and α -GalCer, either in vitro or in vivo. Fig. 5 C shows that in vitro activation of spleen cells by DCs plus α -GalCer strongly induced the expression of IL-12R on NKT cells. Similar findings were made when mice were injected in vivo with α -GalCer (Fig. 5 D).

α -GalCer Synergistically Acts with Exogenously Administered IL-12 in the Activation of Natural Killing Activity and IFN- γ Production In Vivo. C57BL/6 mice were injected intravenously with α -GalCer, and their splenic natural killing activity against YAC-1 cells was determined 24 h later. As

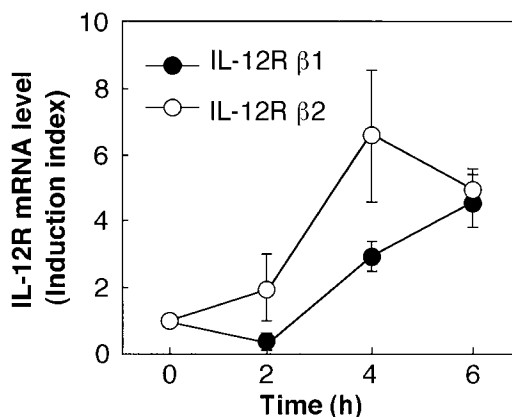


Figure 3. Upregulation of IL-12R expression in the spleen upon in vivo administration of α -GalCer. C57BL/6 mice were injected intravenously with α -GalCer. Various times (0, 2, 4, and 6 h) after the treatment, spleen cells were prepared and their expression of IL-12R β 1 (●) and IL-12R β 2 (○) mRNA was measured by quantitative RT-PCR. The IL-12R mRNA levels are represented as an induction index, as described in Materials and Methods. The bars represent mean \pm SE of triplicate samples.

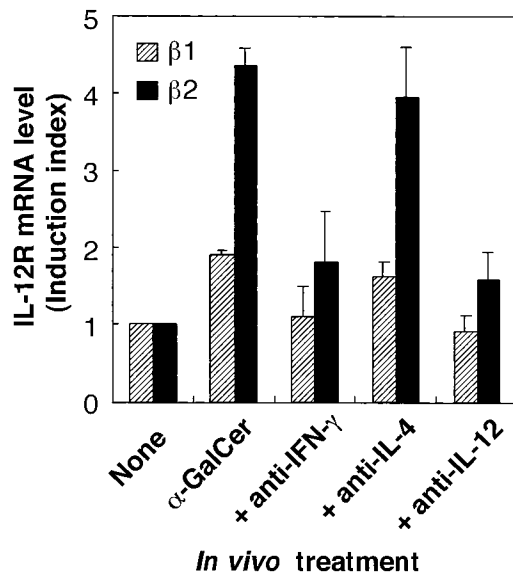


Figure 4. Role of IL-12 and IFN- γ in the induction of IL-12R expression in the spleen. C57BL/6 mice were injected intravenously with α -GalCer. 4 h after the injection, spleen cells were prepared and their expression of IL-12R β 1 (▨) and IL-12R β 2 (■) mRNA was measured by quantitative RT-PCR. The IL-12R mRNA levels are presented as an induction index, as described in Materials and Methods. The blocking effect of anti-IL-12 mAb and anti-IFN- γ mAb before injection of α -GalCer was determined by injection of these mAbs (500 μ g/mouse i.p.) into the mice 1 and 0 d before the treatment with α -GalCer. The bars represent mean \pm SE of triplicate samples.

shown in Fig. 6 A, a suboptimal dose of neither α -GalCer nor IL-12 was able to activate natural killing activity in vivo. However, combined administration of α -GalCer and IL-12 at a suboptimal dose caused a marked augmentation of natural killing.

A similar synergistic effect of α -GalCer and IL-12 was demonstrated for the elevation of serum IFN- γ production. As shown in Fig. 6 B, the administration of α -GalCer plus IL-12 resulted in a strong enhancement of serum IFN- γ levels in C57BL/6 mice compared with mice treated with α -GalCer or IL-12 only.

Discussion

The finding that NKT cells recognize α -GalCer presented by DCs in a CD1d-dependent manner represents a novel recognition mechanism in the immune system (15). NKT cells, which can produce both IFN- γ and IL-4 (16, 17), play an important role in immunoregulation and have been considered to play a central role as innate effector cells involved in both the protection and the onset of immune diseases (18). The NKT cell ligand α -GalCer has a strong immunopotentiating effect in vivo, and this chemical mediates strong antitumor activity (3–5, 9). Therefore, it is important to dissect the mechanism by which α -GalCer activates NKT cells.

The previous finding (3) that NKT-deficient mice did not respond to α -GalCer strongly suggested that NKT cells may be the primary target cells to α -GalCer. However, it

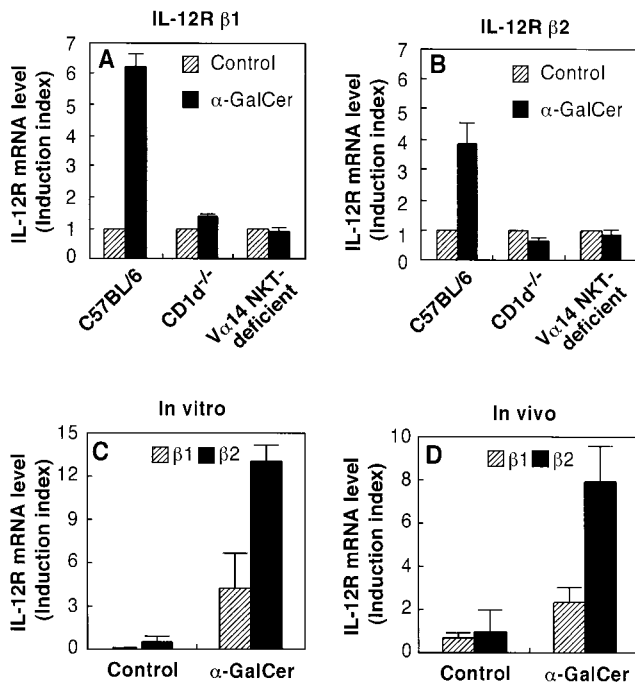


Figure 5. α -GalCer induces IL-12R expression on NKT cells. C57BL/6 wild-type mice, CD1d^{-/-} mice, and NKT-deficient mice were injected intravenously with vehicle (▨) or α -GalCer (■). 4 h after the injection, spleen cells were prepared and their expression of IL-12Rβ1 (A) and IL-12Rβ2 (B) mRNA was measured by quantitative RT-PCR. (C) Spleen cells from wild-type mice were cultured with DCs plus α -GalCer or vehicle for 8 h, NK1.1⁺TCR- α /β⁺ NKT cells were purified by cell sorting, and the expression of IL-12Rβ1 (▨) and IL-12Rβ2 (■) was examined by quantitative RT-PCR. (D) Wild-type mice were injected intravenously with α -GalCer or vehicle; after 6 h the spleens from these mice were isolated, and NK1.1⁺TCR- α /β⁺ NKT cells were purified by cell sorting, and expression of IL-12Rβ1 (▨) and IL-12Rβ2 (■) was examined by quantitative RT-PCR. The IL-12R mRNA levels are presented as an induction index, as described in Materials and Methods. The bars represent mean \pm SE of triplicate samples.

still remained unclear whether only NKT cells responded to α -GalCer. To answer this question, we used highly purified splenic NK cells, NKT cells, CD4⁺ T cells, and CD8⁺ T cells and determined their responsiveness to α -GalCer in the presence of DCs. The data illustrated in Fig. 1 clearly demonstrate that NKT cells are the only cells that respond to α -GalCer (3). It is surprising that neither classical NK cells nor mainstream CD4⁺ T cells or CD8⁺ T cells revealed a significant response to α -GalCer even in the presence of DCs. Together with previous findings (3), the present data indicate that α -GalCer selectively stimulates NKT cells in the presence of DCs.

Recently, the mechanisms of activation of naive CD4⁺ T cells through interaction with DCs have been examined (12, 19–22). Cell–cell adhesion between CD4⁺ T cells and DCs through CD40/CD40L and B7.1/CD28 resulted in the activation of both DCs and T cells, which triggered the production of IL-12 by DCs and IFN- γ by Th1 cells (12, 19, 20, 23–25). Such conditioned DCs were able to prime cytotoxic T cells (22, 26, 27). This recognition system has resemblance to that discussed here. As shown in Fig. 2,

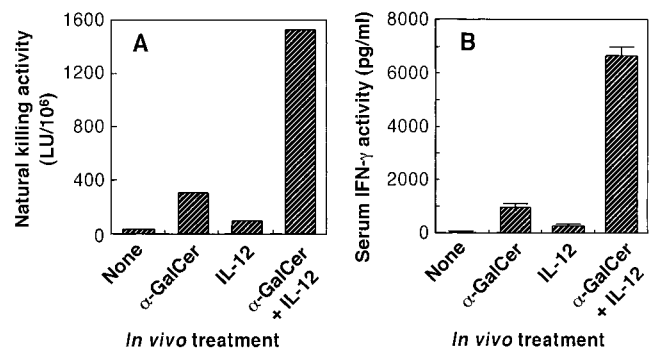


Figure 6. Synergistic effect of α -GalCer and IL-12 in vivo. C57BL/6 mice were injected with a suboptimal dose of α -GalCer (200 ng/mouse i.v.) and 6 h later, mice were injected with a suboptimal dose of IL-12 (200 U/mouse i.p.). 1 d after the treatment with IL-12, the mice were killed and splenic natural killing activity (A) and serum IFN- γ levels (B) were determined as described in Materials and Methods. The bars represent mean \pm SE of triplicate samples.

IL-12 production by DCs appears to be essential for NKT cell activation by α -GalCer, because neutralization of endogenously produced IL-12 by anti-IL-12 mAb caused a strong inhibition of IFN- γ production by NKT cells. The important role of CD40/CD40L for the production of IFN- γ in the cocultures of DCs and NKT cells with α -GalCer is also apparent from these experiments (Fig. 2 B). As demonstrated in Fig. 2 C, DCs produce IL-12 only when they are cultured with α -GalCer in the presence of NKT cells, indicating that direct contact between α -GalCer-bound DCs and NKT cells may be essential for IL-12 production by DCs. This interaction may be required for the production of IFN- γ by IL-12-activated NKT cells, because mAbs directed against CD40/CD40L greatly inhibited IFN- γ production by NKT cells (Fig. 2). These findings indicate that the interaction of NKT cells with DCs may be very similar to the interaction of helper T cells with DCs (22, 26, 27). Since the interactions between DCs and NKT cells occur very quickly after administration of α -GalCer, NKT cells may be able to condition DCs very early in an immune response, and affect subsequent adaptive responses.

In this paper, we also demonstrate that α -GalCer upregulates IL-12R expression in vivo (Fig. 3). IL-12R upregulation is blocked by mAbs against IL-12 or IFN- γ and is absent in CD1d^{-/-} and NKT-deficient mice (Figs. 4 and 5). Moreover, activation of NKT cells in vitro and in vivo results in a strong induction of IL-12Rβ1 and IL-12Rβ2 on these cells (Fig. 5, C and D). Therefore, we speculate that the following series of events is induced upon culture of α -GalCer with DCs and NKT cells: (a) α -GalCer first binds to CD1d molecules on DCs; (b) NKT cells recognize α -GalCer-bound DCs via their TCRs and also interact with DCs via CD40/CD40L; (c) during this interaction, DCs produce IL-12; (d) the endogenously produced IL-12 stimulates IFN- γ production by NKT cells; and (e) IFN- γ produced by NKT cells upregulates IL-12R on NKT cells in an autocrine manner. The dramatic synergistic effect of

suboptimal α -GalCer and exogenously administered IL-12 indicates that expression of IL-12R β 1 and β 2, detected by quantitative RT-PCR, is functionally upregulated in vivo. Moreover, since this synergistic effect of α -GalCer and IL-12 was not demonstrated in NKT-deficient mice, we conclude that in wild-type mice coadministration of α -GalCer and IL-12 leads to upregulation of IL-12R on CD1-dependent NKT cells.

Both α -GalCer and IL-12 have been demonstrated to exhibit potent antitumor activity in vivo. IL-12 has multiple effects on the immune system that are beneficial for the induction of antitumor immunity in vivo (28–30). However, the unexpected severe side effects of IL-12 have made it difficult to use this cytokine in clinical trials (31). We demonstrated that α -GalCer synergistically acts with small doses of IL-12 in vivo to activate NKT cells and to induce IFN- γ production (Fig. 6). These findings suggest that

coadministration of α -GalCer with IL-12 could be used as a new approach for tumor immunotherapy.

Recent studies have demonstrated that Th1 immunity regulated by IL-12 and IFN- γ plays a critical role in the induction of protective immunity against tumors and infectious agents (32, 33). Although NKT cells are involved in both Th1 and Th2 immunity through IFN- γ or IL-4 production, the immunomodulating protocol using α -GalCer and IL-12 preferentially induces NKT cells that produce large amounts of IFN- γ (34). These NKT cells may facilitate the development of Th1-dominant cellular immunity essential for the induction of protective immunity against tumors and some infectious agents. Recently, it was demonstrated that α -GalCer can stimulate human NKT cells in a CD1d-dependent manner (35, 36), indicating that our proposed immunotherapy protocol using α -GalCer and IL-12 will be useful for the application to human immune diseases, including cancer.

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Address correspondence to Takashi Nishimura, Section of Genetic Engineering, Research Center for Genetic Engineering and Cell Transplantation, Department of Immunology, Tokai University School of Medicine, Bohseidai, Isehara 259-1193, Japan. Phone: 81-463-93-1121; Fax: 81-463-96-5438; E-mail: tak24@is.icc.u-tokai.ac.jp

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