Short Communication

BUDR INHIBITION OF POST-DMSO-INDUCED ERYTHROLEUKAEMIA CELL DIFFERENTIATION IN VITRO

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THE ADDITION of dimethylsulfoxide (DMSO) to suspension cultures of Friend leukaemia cells results in the erythroid differentiation of a substantial proportion of cells in the culture (Friend et al., 1971). The continuous presence of DMSO in the culture medium is not necessary for differentiation to occur since exposure of the cells to DMSO for 2 days results in the differentiation of a substantial number of cells after subsequent culture in DMSO-free medium (Preisler and Giladi, 1975). Whilst cells continue to differentiate after removal of DMSO from the culture medium, it is not known whether the continued differentiation is a result of the utilization of mRNAs synthesized during growth in the presence of DMSO, or whether the process of differentiation including the synthesis of globin mRNA continues despite the removal of DMSO from the culture medium. The studies reported here strongly suggest the latter alternative.

Friend leukaemia cells (line 745A) were cultured as previously described (Preisler, Scher and Friend, 1973). Cells were grown in the presence or absence of DMSO (2% v/v) for 2 days and the amount of haem present in 10⁷ cells determined spectrophotometrically (Preisler and Giladi, 1975). The cells were washed twice with cold phosphate-buffered saline and pelleted at 900 g and then placed into fresh room-temperature DMSO-free medium at a concentration of 3×10^5 cells/ml (secondary culture). 5-bromo-2'-deoxyuridine (BUdR) was added to half of the secondary cultures to a final concentration of $3 \mu g/ml$. After 3 additional days of secondary culture the cells were harvested, counted and the amount of haem present in 10^7 cells determined. In experiments I and II slides were made and benzidine-positive cells were determined by a single observer who scored the cells in a single-blind fashion.

As we have previously reported there is a slight but detectable increase in the amount of haem present in cultures after 2 days' growth in the presence of DMSO. When these cells were washed and subsequently grown in DMSO-free medium the amount of haem substantially increased during secondary culture (Table The total amount of haem present I). in secondary cultures of control cells also increased, but the increase was smaller and could be accounted for by an increase in cell number during secondary culture with a small proportion of spontaneous differentiation (Scher, Holland and Friend, 1971). The addition of BUdR to secondary cultures of cells previously exposed to DMSO resulted in a 50% decrease in the total amount of haem accumulated by the secondary cultures (Table I). The addition of BUdR to secondary cultures of control cells did not detectably inhibit erythroid spontaneous differentiation. This finding has been previously reported

TABLE I.—Effect of BUdR on the Accumulation of Haem During Secondary Culture

		Experiment		
		Í	II	Ш
*Start of secondary culture	Control DMSO	$16 \\ 24$	$\begin{array}{c} 27\\ 50 \end{array}$	$\frac{19}{34}$
†After 3 days secondary culture	$egin{array}{l} { m Control} \ { m Control} \ + \ { m BUdR} \ { m DMSO} \ { m DMSO} \ + \ { m BUdR} \ { m DMSO} \ + \ { m BUdR} \end{array}$	56 71 367 180	$209 \\ 266 \\ 2030 \\ 1235$	$212 \\ 280 \\ 1843 \\ 972$

 $O.D._{415}/10$ ml of culture of Friend leukaemia cells at the start of secondary culture.

 $\pm 0.0_{415}/10$ ml of culture of Friend leukaemia cells at the end of secondary culture.

The $O.D._{415}^{\prime}/10$ ml of culture was calculated as follows: the $O.D._{415}^{\prime}/10^7$ cells was determined, and multiplied by: Cells/10 ml culture $\times 10^{-7}$

TABLE II.—Effect of BUdR on Haem Synthesis After Exposure to and Removal from DMSO

Exp. I		Exp. II		Exp. III
$egin{array}{c} 6\cdot5 imes10^5\ 2\cdot7 imes10^5 \end{array}$		$11 \cdot 0 imes 10^5 \\ 8 \cdot 1 imes 10^5$		$8.5 imes 10^{5} \ 3.0 imes 10^{5}$
$5 \cdot 22 imes 10^{-3} \ 8 \cdot 42 imes 10^{-3}$		$8 \cdot 84 \times 10^{-3}$ $16 \cdot 7 \times 10^{-3}$		$6 \cdot 23 imes 10^{-3} \ 11 \cdot 2 imes 10^{-3}$
$egin{array}{llllllllllllllllllllllllllllllllllll$		$3 \cdot 87 imes 10^{6} \\ 4 \cdot 07 imes 10^{6} \\ 2 \cdot 82 imes 10^{6} \\ 2 \cdot 52 imes 10^{6}$		$3 \cdot 12 imes 10^{6} \ 3 \cdot 08 imes 10^{6} \ 2 \cdot 88 imes 10^{6} \ 2 \cdot 7 imes 10^{6}$
	% B+' cells	**	m % B + cells	
$3\cdot 55 imes 10^{-3}\ 3\cdot 44 imes 10^{-3}\ 13\cdot 6 imes 10^{-3}\ 8\cdot 2 imes 10^{-3}$	$\begin{array}{c} 0\\ 0\\ 13\\ 5\end{array}$	$5 \cdot 4 \times 10^{-3}$ $6 \cdot 54 \times 10^{-3}$ $72 \cdot 0 \times 10^{-3}$ $49 \cdot 0 \times 10^{-3}$	$0 \\ 1 \\ 24 \\ 11$	$6 \cdot 9 \times 10^{-3}$ $9 \cdot 1 \times 10^{-3}$ $64 \cdot 0 \times 10^{-3}$ $36 \cdot 0 \times 10^{-3}$
	$\begin{array}{c} \cdot\\ 6\cdot5\times10^5\\ 2\cdot7\times10^5\\ 5\cdot22\times10^{-3}\\ 8\cdot42\times10^{-3}\\ 1\cdot57\times10^6\\ 2\cdot7\times10^6\\ 2\cdot7\times10^6\\ 2\cdot7\times10^6\\ 3\cdot55\times10^{-3}\\ 3\cdot44\times10^{-3}\\ \end{array}$	$\begin{array}{c} \cdot \\ 6 \cdot 5 \times 10^{5} \\ 2 \cdot 7 \times 10^{5} \\ \hline \\ 5 \cdot 22 \times 10^{-3} \\ 8 \cdot 42 \times 10^{-3} \\ \hline \\ 1 \cdot 57 \times 10^{6} \\ 2 \cdot 07 \times 10^{6} \\ 2 \cdot 7 \times 10^{6} \\ 2 \cdot 7 \times 10^{6} \\ \hline \\ \end{array}$	$\begin{array}{ccccccc} 6\cdot5\times10^5 & 11\cdot0\times10^5 \\ 2\cdot7\times10^5 & 8\cdot1\times10^5 \\ \hline\\ 5\cdot22\times10^{-3} & 8\cdot84\times10^{-3} \\ 8\cdot42\times10^{-3} & 16\cdot7\times10^{-3} \\ \hline\\ 1\cdot57\times10^6 & 3\cdot87\times10^6 \\ 2\cdot07\times10^6 & 4\cdot07\times10^6 \\ 2\cdot7\times10^6 & 2\cdot82\times10^6 \\ 2\cdot7\times10^6 & 2\cdot52\times10^6 \\ 2\cdot52\times10^6 & 2\cdot52\times10^6 \\ \hline\\ & & & & \\ & & & & \\ & & &$	$\begin{array}{cccccccccccccccccccccccccccccccccccc$

Note the difference in cell growth and haem synthesis between experiments I, II, and III. Different batches of foetal calf serum were used. Despite the variability in the absolute amount of haem synthesized, in all 3 experiments BUdR inhibited post-DMSO differentiation by approximately 50%.

* Control cells placed in secondary culture in the presence of BUdR.

† Cells which were exposed to DMSO during primary culture and placed in secondary culture in fresh DMSO-free medium.

 \ddagger Same cells as above (†) but BUdR was present during secondary culture. ** Benzidine + ve.

using ⁵⁹Fe incorporation into haem as an index of haem synthesis (Scher, Preisler and Friend, 1973).

The addition of BUdR to secondary cultures of either control or DMSOtreated cells had no effect on the cell growth during the 3-day period of secondary culture (Table II). The amount of haem synthesized/cell in the culture and the proportion of benzidine-positive cells was decreased in each instance by approximately 50%. At first glance it seems surprising that the increment in the amount of haem/10⁶ cells during secondary culture (i.e. the difference between the amount of haem present at the start and at the end of the secondary culture of cells exposed to DMSO during primary culture) appeared to be less than the increment in haem/culture. This observation arises because the rate of increase in differentiated cells during

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secondary culture was less than the rate of increase in undifferentiated cells in the same culture (Preisler and Giladi, 1975) and hence the proportion of differentiated cells in the secondary culture actually declined.

It has previously been demonstrated that BUdR interferes with the DMSOinduced differentiation of Friend leukaemia cells (Ostertag et al., 1973; Scher et al., 1973) and that this inhibition results from interference with the accumulation of globin mRNA (Conkie et al., 1974; Preisler *et al.*, 1973). These observations, taken together with those reported here, strongly suggest that these leukaemic cells continue to synthesize and accumulate globin mRNA after removal of DMSO from the culture medium and that interference with this process inhibits differentiation during secondary culture in DMSO-free medium.

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