

# USP2a alters chemotherapeutic response by modulating redox

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Cancer cells are characterized by altered ubiquitination of many proteins. The ubiquitin-specific protease 2a (USP2a) is a deubiquitinating enzyme overexpressed in prostate adenocarcinomas, where it exhibits oncogenic behavior in a variety of ways including targeting c-Myc via the miR-34b/c cluster. Here we demonstrate that USP2a induces drug resistance in both immortalized and transformed prostate cells. Specifically, it confers resistance to typically pro-oxidant agents, such as cisplatin (CDDP) and doxorubicin (Doxo), and to taxanes. USP2a overexpression protects from drug-induced oxidative stress by reducing reactive oxygen species (ROS) production and stabilizing the mitochondrial membrane potential ( $\Delta\Psi$ ), thus impairing downstream p38 activation and triggering of apoptosis. The molecular mediator of the USP2a protective function is the glutathione (GSH). Through miR-34b/c-driven c-Myc regulation, USP2a increases intracellular GSH content, thus interfering with the oxidative cascade triggered by chemotherapeutic agents. In light of these findings, targeting Myc and/or miR-34b/c might revert chemo-resistance.

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**Subject Category:** Cancer

Deubiquitinating enzymes (DUBs) represent one of the largest families of proteins responsible for regulating the ubiquitin–proteasome system by opposing ubiquitination.<sup>1</sup> Although DUBs have been first identified for their role in protein stabilization, in recent years several other mechanisms have emerged by which the balance ubiquitination/deubiquitination can regulate protein level, activity and subcellular location, thus driving different cellular processes from gene transcription and DNA repair to cell cycle and apoptosis.<sup>2</sup>

Alteration in the ubiquitination pattern is observed in most cancer cells, where it is manifested by destabilization of tumor suppressors and activation of oncogenes. This justifies the efforts for developing proteasome inhibitors in cancer therapy, like Bortezomib used successfully in myeloma, neuroblastoma and mesothelioma therapy.<sup>3,4</sup> DUB enzymes and proteasome activity also affect cancer chemo- and radio-resistance. In fact, proteasome inhibitors have been proposed to be used in combination with conventional antineoplastic agents. Therapeutic strategies are currently being investigated to overcome chemo-resistance based on the mechanisms of ubiquitination/deubiquitination.<sup>5–9</sup> However, despite both the *in vitro* and *in vivo* (clinical) evidence of beneficial antineoplastic effects, the reason why blocking proteasomal degradation in a ‘non-specific’ manner results in differential killing of tumor cell remains elusive.

In prostate, USP2a is a deubiquitinating enzyme specifically overexpressed in about 40% of prostate adenocarcinomas.<sup>10,11</sup> Its oncogenic behavior is ascribed to its interaction with and prevention of proteasomal degradation of specific protein targets involved in different cellular pathways, such as fatty acid synthase (FASN), Mdm2, MdmX, AIF, Cyclin D, Aurora-A and EGFR.<sup>12–18</sup> Moreover, we recently reported that USP2a is a master regulator of microRNA (miRNA or miR) expression in both immortalized and transformed prostate epithelial cells, where its overexpression specifically regulates the miR-34b/c cluster to target c-Myc.<sup>19</sup>

miRNAs have been shown to be differently expressed in benign compared with malignant prostate tissue and in different stages of prostatic carcinogenesis.<sup>20–22</sup> In addition, certain miRs have been found to predict and affect sensitivity to anticancer treatment.<sup>23</sup> For instance, the tumor suppressor miR-148a attenuates paclitaxel resistance in PC3 cells,<sup>24</sup> while overexpression of miR-143 suppresses prostate proliferation and migration, and increases sensitivity to docetaxel (DTX) by targeting the EGFR/RAS/MAPK pathway.<sup>25</sup> miR-34 family members in particular have been often associated to cancer resistance. In prostate cancer, ectopic miR-34 expression triggers cell cycle arrest and growth inhibition and attenuates chemo-resistance to camptothecin (CPT), Doxo and paclitaxel exposure.<sup>26–28</sup>

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**Abbreviations:** CDDP, cisplatin; CPT, camptothecin;  $\Delta\Psi$ , mitochondria membrane potential; Doxo, doxorubicin; DUB, deubiquitinating enzyme; DTX, docetaxel; EV, empty vector;  $\gamma$ GCS, the  $\gamma$ -glutamyl-cysteine synthetase; GSH, glutathione; iPrEC, human immortalized non-transformed prostate epithelial cells; miRNAs or miRs, microRNAs; NAC, N-acetyl-cysteine; PI, propidium iodide; ROS, reactive oxygen species; USP2a, ubiquitin-specific protease 2a; USP2aWT, wild-type ubiquitin-specific protease 2a; USP2aMUT, mutant (C276A and H549R) ubiquitin-specific protease 2a

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Here we demonstrate that USP2a modulates the susceptibility to antineoplastic agents in prostate cells. In particular, when overexpressed, USP2a<sup>WT</sup> is able to induce chemo-resistance to typically pro-oxidant agents, such as CDDP and Doxo, and also to the DTX taxane. The latter is in accordance to that previously reported by our group, showing a marked resistance to paclitaxel displayed by the USP2a<sup>WT</sup> prostate cells if compared with EV (empty vector) and USP2a<sup>MUT</sup> transfectants.<sup>11</sup> In this regard, although taxanes are known to be mainly mitotic inhibitors through disruption of microtubule function, recent data on mitochondrial ROS formation showed that they can trigger a direct mitochondrial damage, induce mitochondrial permeability transition and ROS formation.<sup>29–31</sup>

Cisplatin in particular has been widely used in prostate chemotherapy, along with mitoxantrone and DTX.<sup>32</sup> At molecular level, it triggers apoptosis through oxidative stress, p38 kinase activation and mitochondria impairment in different cancer cell types, including prostate cells.<sup>33–36</sup> In our experimental model, we demonstrate for the first time that a specific prostatic DUB, USP2a, protects from drug-induced oxidative stress. The molecular mediator of the USP2a protective function is the glutathione. Through miR-driven c-Myc regulation, USP2a implements the intracellular glutathione content by stimulating its synthesis, thus interfering with the oxidative cascade triggered by the chemotherapeutic agents.

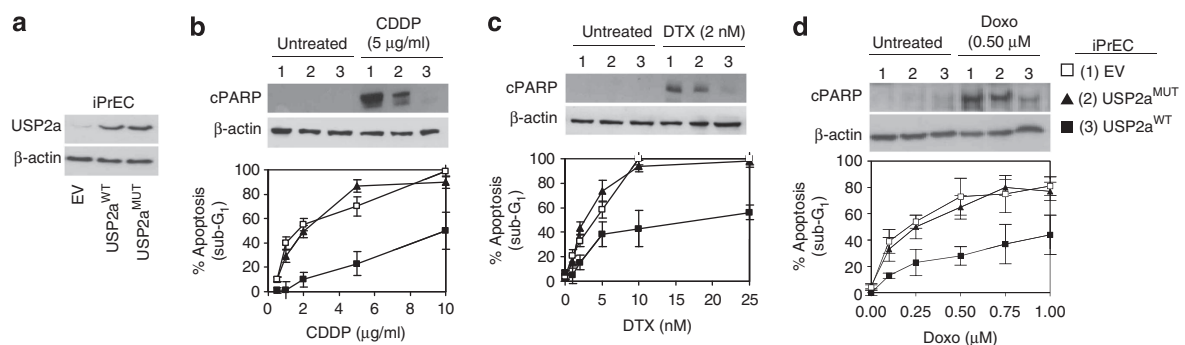
## Results

**USP2a overexpression confers resistance to pro-oxidant agents to prostate cells.** In a previously characterized experimental model of human immortalized non-transformed prostate epithelial cells (iPrEC), overexpressing either wild-type (USP2a<sup>WT</sup>) or catalytically mutant USP2a (USP2a<sup>MUT</sup>),<sup>11,19</sup> we evaluated the response to antineoplastic agents in terms of both subG<sub>1</sub> percentage and apoptotic PARP cleavage.

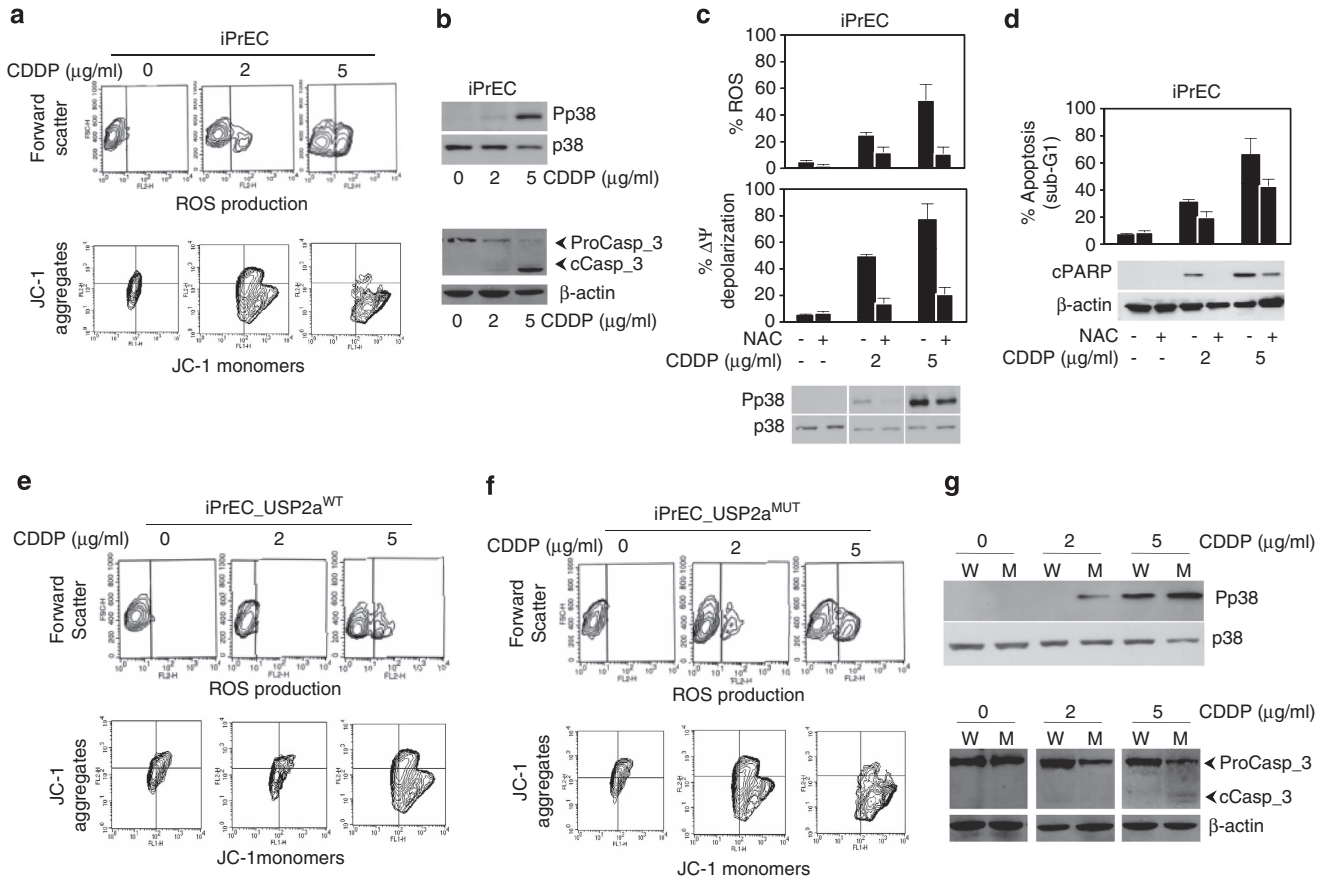
Figure 1 shows that the ectopic USP2a<sup>WT</sup> expression (Figure 1a) significantly affects the response to different drugs. Specifically, USP2a<sup>WT</sup> confers resistance to typically pro-oxidant agents, such as cisplatin (Figure 1b) and doxorubicin (Figure 1d), and to taxanes (docetaxel, Figure 1c, and Priolo *et al.*<sup>11</sup>). The USP2a antiapoptotic function seems to be related to its enzymatic activity, as the dose-dependent effects reported for USP2a<sup>MUT</sup> resemble those elicited by the empty vector clone. Also in a model of

transformed prostate cells (LNCaP cancer cell line), cells acquire resistance to CDDP, DTX and Doxo upon ectopic expression of USP2a<sup>WT</sup> (Supplementary Figure 1a) as the percentage of both apoptotic cells and PARP cleavage levels is significantly reduced in USP2a<sup>WT</sup> when compared with control cells (Supplementary Figure 1b). Accordingly, USP2a silencing (Supplementary Figure 1c) triggers a massive increase of apoptosis in response to CDDP, DTX and Doxo treatment, as demonstrated by the augmented subG<sub>1</sub> percentage and downstream PARP cleavage (Supplementary Figure 1d). Still, the chemo-resistance reported in the LNCaP\_USP2a<sup>WT</sup> cells is specifically dependent on the catalytic activity of the protein, as the drug response observed in the USP2a<sup>MUT</sup> cells resembles what reported in the LNCaP\_EV cells (Supplementary Figure 1e).

**USP2a modulates the cell redox state.** Cisplatin is known to trigger apoptosis through oxidative stress, p38 kinase activation and mitochondria impairment in different cancer cell types, including prostate.<sup>34–36</sup> We thus evaluated the generation of ROS and the depolarization of mitochondria membrane potential in iPrEC cells, following CDDP administration. Cytofluorimetric analysis revealed that the susceptibility of epithelial prostate cells to CDDP proceeds via a dose-dependent ROS generation and  $\Delta\Psi$  depolarization (Figure 2a), accompanied by activation of p38 through phosphorylation and caspase-3 cleavage (Figure 2b). CDDP-induced oxidative stress appears to be an upstream molecular event, as specific inhibition of ROS generation by NAC (*N*-acetyl-cysteine) pre-treatment (Figure 2c) reduces p38 phosphorylation, mitochondria depolarization and downstream apoptosis, as evidenced by decreased PARP cleavage and subG<sub>1</sub> percentage (Figure 2d). However, as NAC pre-loading does not completely abolish p38 activation and PARP cleavage at the CDDP concentration of 5  $\mu$ g/ml (Figures 2c and d), it suggests that the drug-induced apoptosis can involve alternative pathways at the higher doses. As detailed in Supplementary Figure 2, only when administered in combination with the specific p38 inhibitor (SB202190), NAC interferes with the caspase-3 and PARP activation (Supplementary Figure 2a) and reduces cell death (Supplementary Figure 2b), suggesting the occurrence of a double lane of cellular mediators (both ROS and p38) to death (Supplementary Figure 2c).



**Figure 1** USP2a triggers chemo-resistance in prostate epithelial cells. Evaluation of (a) USP2a expression and (b–d) of the apoptotic response carried out by western blot of cleaved PARP (cPARP) and cytofluorimetric assay of the subG<sub>1</sub> percentage in the indicated iPrEC cells, following treatment with CDDP, DTX and Doxo



**Figure 2** USP2a has an antioxidant effect in response to CDDP. Evaluation of (a) ROS production and  $\Delta\Psi$  depolarization, and (b) p38 kinase phosphorylation (Pp38) and caspase-3 activation, carried out in iPrEC cells, following CDDP exposure. Upon NAC pre-treatment, iPrEC cells have been characterized in response to CDDP in terms of (c) ROS production,  $\Delta\Psi$  depolarization and p38 activation, and of (d) cPARP cleavage and subG<sub>1</sub> apoptotic percentage. Evaluation of (e, f) ROS and  $\Delta\Psi$  depolarization, and (g) p38 and caspase-3 activation has been further carried out in iPrEC\_USP2a<sup>WT</sup> (W) and iPrEC\_USP2a<sup>MUT</sup> (M) cells, respectively, following CDDP exposure

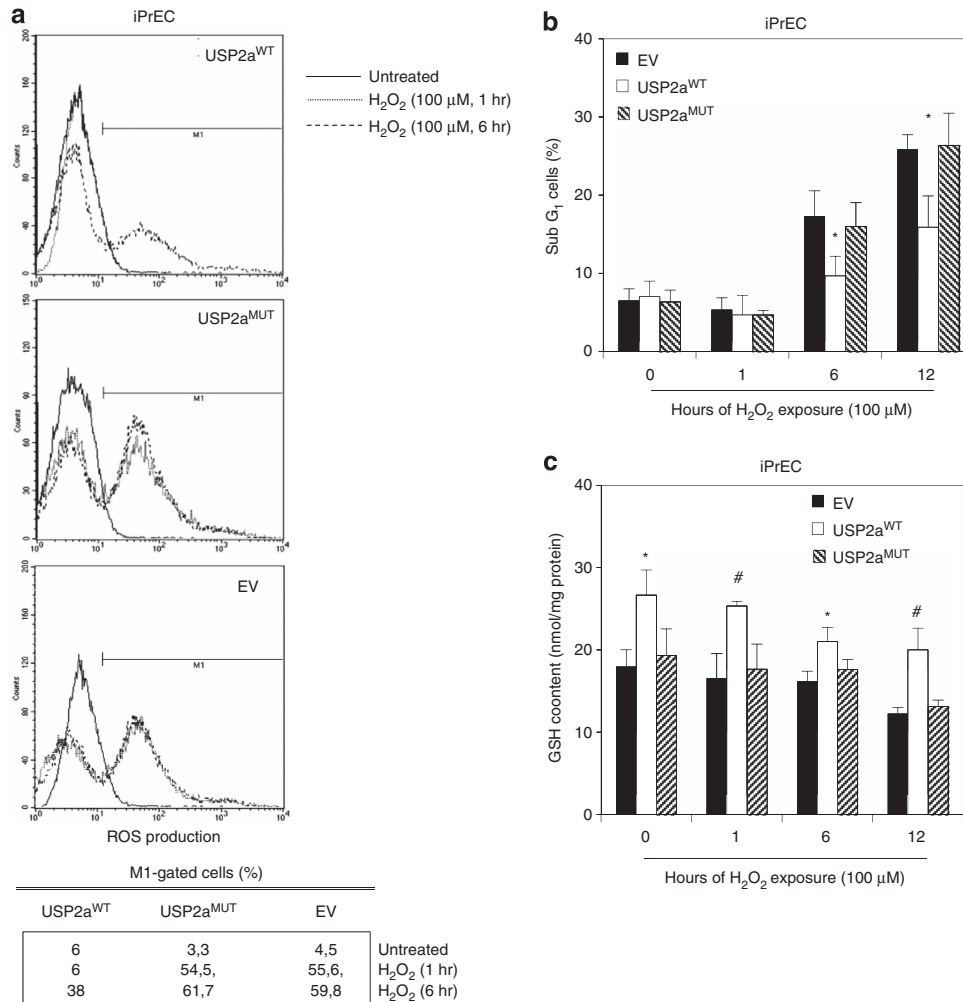
Interestingly, when overexpressed, USP2a<sup>WT</sup> displays an antioxidant effect. Its expression reduces the CDDP-generated ROS and protects prostate cells from drug-induced  $\Delta\Psi$  impairment at both the doses administered (Figure 2e). If catalytically inactive, USP2a does not modify the iPrEC sensitivity to CDDP-triggered oxidative stress and mitochondria dysfunction, the iPrEC-USP2a<sup>MUT</sup> profile being comparable to control cells (Figure 2f). Still, the ROS-scavenging ability driven by USP2a<sup>WT</sup> in response to CDDP is able to inhibit downstream p38 phosphorylation and caspase-3 cleavage if compared with mutant protein (Figure 2g). No difference in p38 phosphorylation has been reported between mutant and empty vector cells in response to drug (data not shown).

Also in response to Doxo and DTX, iPrEC cells display an increased ROS generation, which is actively scavenged by NAC pre-exposure (Supplementary Figure 3a). Interestingly, NAC treatment abolishes Doxo-induced cytotoxicity, whereas it only partially interferes with the apoptotic cascade initiated by DTX (Supplementary Figure 3b). As USP2a<sup>WT</sup> scavenges drug-induced ROS species, as much as NAC pre-treatment (Supplementary Figure 3c), but also significantly impairs apoptotic response to DTX (Figure 1c), it suggests that the USP2a-dependent chemo-resistance to taxanes proceeds via

multiple pathways and that its antioxidant function is just one of the involved mechanism.

To further characterize the antioxidant function, USP2a<sup>WT</sup> overexpression was tested in response to direct H<sub>2</sub>O<sub>2</sub> exposure in iPrEC cells (Figure 3). As hypothesized, it protects from H<sub>2</sub>O<sub>2</sub>-induced ROS in a time-dependent manner (Figure 3a), the M<sub>1</sub>-gated cells percentage (ROS-positive cells) being significantly reduced in USP2a<sup>WT</sup> compared with USP2a<sup>MUT</sup> and empty vector cells, at both 1 and 6 h post treatment. By extending the time-window of the analysis up to 12 h, USP2a<sup>WT</sup> cells are further demonstrated to be protected from the H<sub>2</sub>O<sub>2</sub>-driven cytotoxic effect (Figure 3b). As the main master regulator of cellular redox homeostasis is the glutathione (GSH), we measured the intracellular GSH content in iPrEC transfectants (Figure 3c) and showed that USP2a<sup>WT</sup> buffers the H<sub>2</sub>O<sub>2</sub>-induced oxidative stress by implementing the GSH pool in prostate cells. In iPrEC-USP2a<sup>WT</sup> cells, GSH content is indeed increased by almost 30% if compared with USP2a<sup>MUT</sup> (Figure 3c) and empty vector control clone.

In support of these findings, deregulated USP2a expression is able to significantly affect GSH concentration also in the LNCaP experimental model (Supplementary Figure 4a).



**Figure 3** USP2a increases GSH content and scavenges ROS. (a) Analysis of ROS production performed in the iPrEC cells exposed to 100  $\mu$ M H<sub>2</sub>O<sub>2</sub> for 1 and 6 h. In the table below, the percentage of the M<sub>1</sub>-gated (ROS positive) cells is reported. (b) Apoptotic evaluation and (c) GSH intracellular content analysis performed in iPrEC cells treated with 100  $\mu$ M H<sub>2</sub>O<sub>2</sub> up to 12 h. Asterisks indicate the statistically significant differences, calculated in iPrEC\_USP2a<sup>WT</sup> versus EV (\* $P$  < 0.05, # $P$  < 0.01)

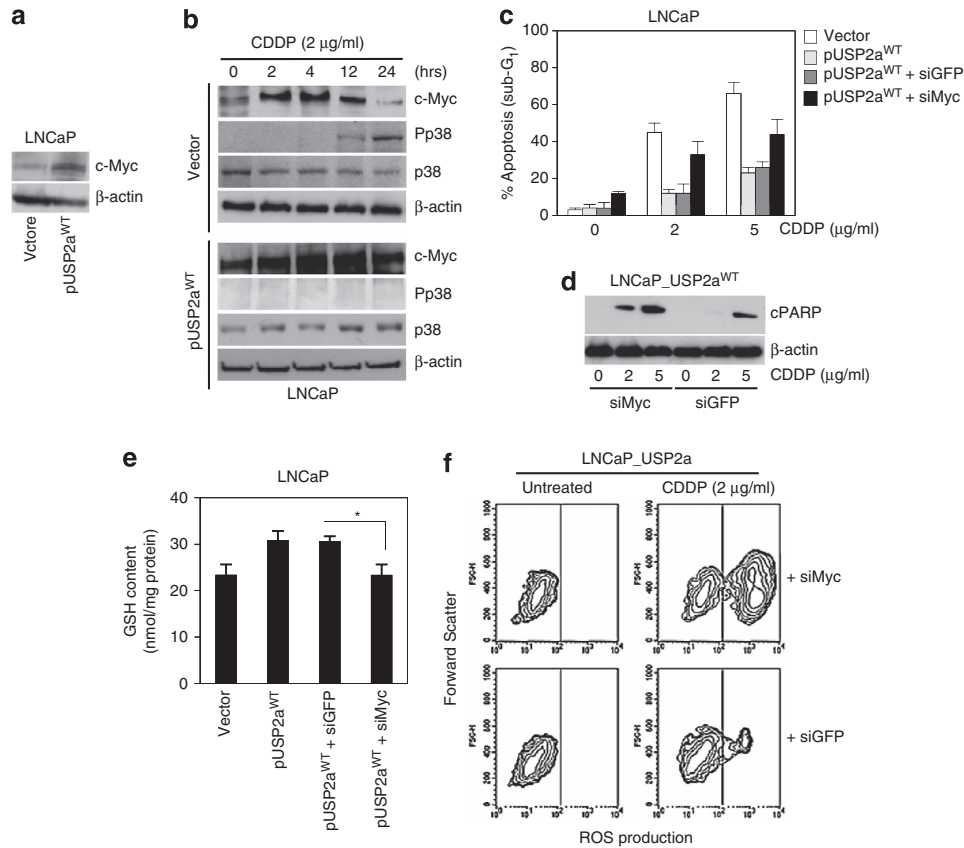
### miR-dependent c-Myc regulation is involved in the USP2a antioxidant ability.

In search of a molecular effector underlying the USP2a antioxidant ability, we evaluated the involvement of c-Myc oncoprotein in the USP2a chemoprotection, as it is a well-known drug resistance factor in cancer,<sup>34,37,38</sup> and is upregulated by USP2a via miR-34b/c impairment in prostate cells.<sup>19</sup>

To this purpose, we first verified whether c-Myc expression was induced by drug treatment in the USP2a<sup>WT</sup> and control LNCaP cells (Figures 4a and b). In response to CDDP, LNCaP cells attempt to react to drug-induced damage by increasing c-Myc protein as early as 2 h after drug administration (Figure 4b). c-Myc expression level is maintained for the following 12 h, returning progressively to basal level by 24 h. Interestingly, activation of p38 is inversely correlated to c-Myc induction, the kinase being progressively phosphorylated with the disappearance of c-Myc. Through upregulation of basal c-Myc level (Figure 4a), USP2a<sup>WT</sup> confers resistance to CDDP by enabling LNCaP cells to quickly respond to drug damage. In fact, c-Myc protein expression is kept higher than

in empty vector transfectants, and no phosphorylation of p38 kinase can be triggered by drug in USP2a<sup>WT</sup> cells. The ability of c-Myc to specifically protect cells from CDDP-induced damage was confirmed by evaluating the drug response in c-Myc-overexpressing prostate cancer cells. c-Myc stimulation in LNCaP cells (Supplementary Figure 5a) abrogates CDDP-induced p38 phosphorylation (Supplementary Figure 5b), reduces the apoptotic subG<sub>1</sub> percentage in a dose-dependent manner, even more efficiently than what reported for USP2a<sup>WT</sup> overexpression (Supplementary Figure 5c), and inhibits PARP cleavage (Supplementary Figure 5d). In accordance, c-Myc interference in LNCaP\_USP2a<sup>WT</sup>-overexpressing cells partially reverts CDDP resistance. As shown in Figure 4c, siMyc stimulates apoptosis in response to cisplatin and leads to a concomitant appearance of cleaved PARP at the lowest dose reported (Figure 4d), although the drug susceptibility is never completely restored to parental level.

c-Myc-driven protection from CDDP in USP2a<sup>WT</sup> cells proceeds via GSH modulation. As reported in Figure 4e,



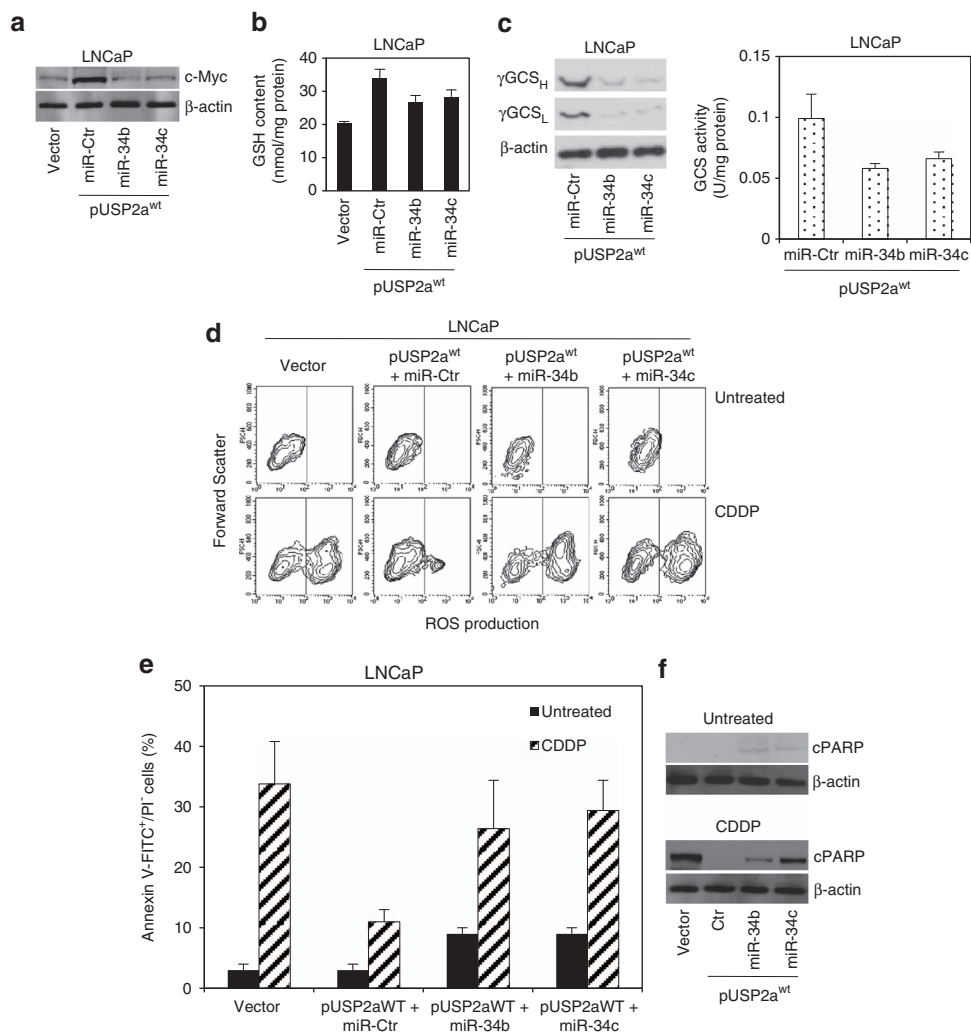
**Figure 4** c-Myc is the downstream mediator of the USP2a antioxidant ability in response to drugs. (a, b) Western blot analysis of c-Myc, basal and phosphorylated p38 (Pp38) expressions carried out in LNCaP empty vector and pUSP2a<sup>WT</sup> cells, following treatment with CDDP (2 μg/ml). (c) Cytofluorimetric evaluation of apoptotic subG<sub>1</sub> percentage and (d) cleaved PARP (cPARP) performed in the following CDDP-treated LNCaP transfectants: empty vector, pUSP2a<sup>WT</sup>, GFP-silenced (siGFP) and c-Myc-silenced (siMyc) pUSP2a<sup>WT</sup> cells. Evaluation of (e) GSH content and (f) CDDP-induced ROS production carried out in the indicated LNCaP\_USP2a<sup>WT</sup> transfectants. Asterisks indicate the statistically significant differences, calculated in siMyc versus siGFP LNCaP\_USP2a<sup>WT</sup> transfectants (\**P* < 0.05)

siMyc depletes GSH content in the LNCaP\_USP2a<sup>WT</sup>, down to basal levels, thus interfering with the ROS scavenging properties gained with USP2a<sup>WT</sup> overexpression (Figure 4f). We previously demonstrated that c-Myc transcription factor regulates the  $\gamma$ -glutamyl-cysteine synthetase ( $\gamma$ -GCS), the rate-limiting enzyme catalyzing GSH biosynthesis, by directly binding and activating the promoters of both heavy ( $\gamma$ -GCS<sub>H</sub>) and light ( $\gamma$ -GCS<sub>L</sub>) subunits.<sup>34,39</sup> We therefore verified whether the GSH modulation may proceed via a Myc-dependent regulation of  $\gamma$ -GCS expression in our USP2a<sup>WT</sup> prostate experimental model. As reported in Supplementary Figure 6, USP2a<sup>WT</sup> stimulates both  $\gamma$ -GCS subunit expression in LNCaP cells, in terms of mRNA, protein, and enzyme activity rise. Consistently, siUSP2a-LNCaP cells are characterized by a significant decrease in  $\gamma$ -GCS activity (Supplementary Figure 4b). This expression control is clearly USP2a<sup>WT</sup>-specific, as both LNCaP\_USP2a<sup>MUT</sup> and iPREC\_USP2a<sup>MUT</sup> cells resemble what is observed in the empty vector clones (Supplementary Figure 4b).

As hypothesized, c-Myc is the master regulator in the USP2a-driven  $\gamma$ -GCS regulation, as c-Myc silencing reverts the expression and activity stimulation elicited by USP2a<sup>WT</sup> (Supplementary Figures 6a–d).

We also demonstrated that c-Myc regulation by USP2a occurs via miR-34b/c inhibition.<sup>19</sup> We next moved to verify whether GSH-dependent drug resistance to pro-oxidant agents might also be reverted by the synthetic miR molecules. To this end, LNCaP\_USP2a<sup>WT</sup> cells have been preloaded with either control or mimic 34b and 34c molecules before undergoing drug treatment (Figure 5). As expected, both miR-34b and miR-34c are able to reduce c-Myc protein expression in prostate cells, whereas the control mimic does not modify its level (Figure 5a). Interestingly, exogenous miR-34b/c loading does not completely counteract c-Myc induction upon CDDP treatment (Supplementary Figure 7), suggesting the occurrence of alternative pathways controlling Myc induction and stabilization by the drug-induced stress.

The glutathione pool undergoes a significant depletion upon mimic miR addition (Figure 5b), and it is in turn responsible for the marked reduced ROS scavenging ability observed in CDDP-treated LNCaP\_USP2a<sup>WT</sup> cells (Figure 5d). miR-dependent GSH depletion is triggered by the impairment of the  $\gamma$ -GCS expression and enzyme activity (Figure 5c) in the USP2a<sup>WT</sup> cells. On the basis of an *in silico* prediction, carried out by interrogating the public databases and algorithms available on the web (Sanger miRbase, PicTar, Target Scan, miRanda, DIANA miRGen, miRNA Map),<sup>31</sup> we excluded that



**Figure 5** miR-34b/c are involved in the GSH-mediated chemo-resistance driven by USP2a. (a) Western blot analysis of c-Myc. (b) GSH content evaluation, and (c) analysis of the  $\gamma$ -GCS protein expression and enzyme activity carried out in LNCaP empty vector and pUSP2a<sup>WT</sup> transfectant. The latter cells have also been incubated with control miR mimic (miR-Ctr), miR-34b and miR-34c mimic molecules. Analysis of (d) ROS generation, (e) apoptosis (Annexin V/PI staining) and (f) PARP cleavage (cPARP) performed in the above detailed miR mimic-incubated pUSP2a<sup>WT</sup> transfectants, following CDDP exposure

the inhibition of  $\gamma$ -GCS expression in our experimental model is due to a direct targeting by miR-34b/c (data not shown).

The combination of USP2a<sup>WT</sup> and control mimic molecule still protects cells from CDDP-induced apoptosis, if compared with control clone, as evidenced by the percentage of cells exposing the early Annexin V marker (Figure 5d), and PARP cleavage (Figure 5e). In contrast, pre-treatment with either miR-34b or miR-34c makes USP2a<sup>WT</sup> cells unable to trigger drug resistance. Actually, miR analogs *per se* are responsible for activating a very small amount of apoptosis in proliferating USP2a<sup>WT</sup> cells, as demonstrated by the slight increase in Annexin V<sup>+</sup>/propidium iodide (PI)<sup>-</sup> percentage and by the faint appearance of PARP cleavage in the untreated cells (Figures 5d and e). Moreover, each specific miR analog abolishes the protective effect elicited by USP2a<sup>WT</sup> in response to CDDP, the percentage of Annexin V<sup>+</sup>/PI<sup>-</sup> cells being comparable to vector-transfected LNCaP cells

(Figure 5d). Analysis of PARP cleavage confirms that both miR-34b and miR-34c are able to revert cisplatin resistance, although at different extent, the relative protein amount being slightly different from parental CDDP-treated cells (Figure 5e).

## Discussion

Despite the critical role of the ubiquitin–proteasome pathway in the regulation of cellular processes and cancer transformation,<sup>3,40,41</sup> surprisingly little is known about the function specificity of individual DUB enzymes. We have recently identified a novel molecular pathway of oncogenic regulation driven by the USP2a deubiquitinating enzyme through its ability to recruit specific miRNAs in prostate cells.<sup>19</sup> Here we demonstrate that USP2a also triggers prostate chemo-resistance by acting as an antioxidant molecule via the miR-34-mediated Myc and GSH stimulation. To our

knowledge, this is the first report disclosing the cooperation between a DUB enzyme and the miR regulatory machinery specifically in response to oxidative stress.

We here report that the ectopic overexpression of USP2a<sup>WT</sup> drives chemo-resistance in both iPrEC and prostate cancer LNCaP cells, in accordance to that previously suggested in prostate histotype by our group.<sup>11</sup> This oncogenic property also applies to other cancer cell lines, as seen in bladder cancer and in testicular embryonal carcinoma cells, where USP2a deregulation affects response to cisplatin.<sup>14,42</sup> These findings are suggestive of a possible clinical application of USP2a inhibitors to cisplatin-resistant patient population to improve clinical benefit.

Interestingly, in our prostate experimental model, USP2a is able to specifically impair the death response to the pro-oxidant agents, such as CDDP and Doxo, and to microtubules polymerization inhibitors, such as DTX and paclitaxel.<sup>11</sup> In accordance to our findings, siUSP2a bladder cancer cells are reported to display a higher apoptotic rate in response to CDDP and nocodazole, an antineoplastic agent specifically interfering with the polymerization of microtubules.<sup>42</sup>

This USP2a function can be attributable to its deubiquitinating activity, as the response elicited by the catalytically inactive mutant (C276A and H549R) resembles that reported in control cells. Thus, USP2a might specifically address the chemo-response by deubiquitinating and stabilizing different proteins according to the nature of the stressor. Besides, its action may depend on the cellular balance with other death-regulating effectors. This is particularly evident following the recent publication by Mahul-Mellier on HeLa and MCF7 breast cancer cells, where downregulation of USP2a by siRNA promotes NF- $\kappa$ B activation and protects cells against TNF-induced cell death, as a consequence of the impairment of the USP2a-TRAF protein ratio.<sup>43,44</sup>

Cisplatin has been extensively used in prostate cancer therapy, and, at molecular level, it is known to trigger apoptosis through mitochondria-mediated oxidative stress.<sup>35,36</sup> The cytotoxicity of anthracyclines is also associated with the generation of reactive oxygen species (ROS) and reactive nitrogen species (RNS). Besides, Doxo can bind to the DNA and uncoil the double-stranded helix with generation of free radicals and DNA damage.<sup>45,46</sup> Interestingly, although taxanes are mainly mitotic inhibitors, through disruption of microtubule function, recent data on mitochondrial ROS formation showed that they can display direct mitochondrial effect, induce mitochondrial permeability transition and ROS formation.<sup>29–31</sup>

We here demonstrate that ROS generation is a key upstream event responsible for CDDP cytotoxicity in prostate, and that USP2a behaves as a ROS scavenging molecule, even if p38 phosphorylation may occur independently of oxidative stress at higher doses. Both Doxo and DTX also elicit ROS generation, that is efficiently scavenged by USP2a<sup>WT</sup>, although, according to our data, chemo-resistance to taxanes proceeds via multiple pathways, and USP2a antioxidant function is just one of the involved mechanism. On the basis of what already published, resistance to taxanes might be explained in terms of the proteins stabilized by USP2a-deubiquitinating function, especially those involved in apoptosis, such as FASN, Mdm2, MdmX and AIF, or by the

multiple miR molecules regulated in the prostate experimental model. Future intensive experiments are required to exhaustively address these issues.

Pro-oxidant chemo-therapy appears particularly efficient in prostate cells. Over the last decade, epidemiological, experimental and clinical studies have in fact implicated oxidative stress in the development and progression of prostate cancer.<sup>47</sup> A possible explanation identifies in the oxidative stress the key factor responsible for the activation of androgen receptor signaling and for the pro-survival and antiapoptotic effects in response to androgen-deprivation therapy, as well as to cytotoxic and tumor-suppressive interventions.<sup>48</sup>

In the light of these evidences, it is comprehensible that the impact of USP2a overexpression in a prostate lesion, as, in response to CDDP, it displays an antioxidant effect. To our knowledge, this is the first experimental evidence demonstrating a ROS-scavenging role for USP2a and, more important, identifying in the intracellular GSH neo-synthesis the possible mediator of this protective effect. This finding is surprisingly in accordance to what has recently been published by Selvendiran *et al*.<sup>49</sup> They report that the synthetic compound HO-3867, known for its potent antioxidant activity, exhibits anticancer effects in many histotypes, including the breast, colon, head and neck, liver, lung, ovarian and prostate, by downregulating USP2a to ultimately trigger apoptosis.<sup>49</sup>

There is an emerging evidence supporting the role of miRNAs in modulating sensitivity to anticancer therapy, and miR-34a and miR-34b/c family in particular has been associated to prostate cancer survival and drug sensitivity.<sup>50–53</sup> We recently demonstrated that USP2a switches on Myc expression via miR-34b/c regulation.<sup>19</sup> The ability to affect c-Myc level by miR has its functional relevance in the regulation of apoptosis and response to chemotherapy. c-Myc is in fact known to be a drug resistance factor.<sup>34,37,38</sup> Different c-Myc-targeted antisense therapies have been so far developed and tested as reliable strategies to improve chemotherapy in solid tumors, strongly identifying in c-Myc a key factor of drug resistance and stress response. In prostate, *c-myc* is often amplified in advanced cancer specimens<sup>54</sup>, so that targeted inhibition of its overexpression has been demonstrated as a promising therapeutic strategy for clinical management of prostate cancer.<sup>55</sup> Moreover, therapeutic use of *c-myc* antisense renders hormone-refractory prostate cancers responsive to chemotherapy in the preclinical model.<sup>56</sup>

Here we demonstrate that USP2a<sup>WT</sup> confers drug resistance by upregulating c-Myc expression and by implementing the GSH content. This is directly due to the ability of c-Myc transcription factor to stimulate the expression of  $\gamma$ -GCS, the rate-limiting enzyme catalyzing the GSH biosynthesis. Moreover, miR-34b/c-mediated c-Myc stimulation is set upstream the GSH neo-synthesis, as both miR mimic administration and Myc silencing in LNCaP-USP2a<sup>WT</sup> cells are responsible for  $\gamma$ -GCS inhibition and consequent GSH depletion, with further chemo-sensitization to pro-oxidant agents. This is consistent with our previously reported data demonstrating a direct antioxidant role of c-Myc through regulation of glutathione synthesis,<sup>34,39</sup> also corroborated by Gao *et al* who disclosed c-Myc involvement in energy and reactive oxygen species homeostasis.<sup>57</sup> These data might

strengthen the hypothesis of a therapeutic approach in USP2a-overexpressing prostate lesions based on the combination of conventional chemotherapy with either miR synthetic drugs or siMyc.

On the basis of our data, we propose a molecular model where USP2a is the upstream master regulator, driving the response to ROS-generating drugs via miR-34b/c-Myc-GSH in prostate cells (Figure 6). In the scheme, we also highlight those miRNAs that we previously identified as USP2a targets, such as let-7, miR-98 and miR-17-5p.<sup>19</sup> According to published evidence, they are known to interfere with the expression of specific targets (BIM, IL-6/STAT3, HMG2 and Bcl-2), specifically involved in the apoptotic response to pro-oxidant agents and antineoplastic drugs.<sup>58–60</sup> In our hypothesis, prostate chemo-resistance might be regulated by USP2a through multiple pathways, according to the dose and the nature of the stressor. Further studies in this direction will help in better defining the involvement of each specific molecular mediator in the complex web of interaction woven by USP2a in prostate cancer cells.

## Materials and Methods

**Chemicals, antibodies and plasmids.** Clinical-grade cisplatin and doxorubicin were obtained from Pharmacia (Milan, Italy). Docetaxel was purchased from Sigma (Sigma-Aldrich, St. Louis, MO, USA) and Glaxo Wellcome (Verona, Italy), respectively. Drug dilutions were freshly prepared before each experiment.

NAC, H<sub>2</sub>O<sub>2</sub> (30% w/w) and PI were from Sigma. DHE (dihydroethidium), JC-1 (5',6,6'-tetrachloro-1,1',3,3'-tetraethylbenzimidazolylcarbocyanine iodide) and the Annexin V-FITC *versus* PI kit (Vibrant apoptosis assay, V-13242) were purchased from Molecular Probes (Life Technologies, Grand Island, NY, USA).

For western blot analysis, the following primary antibodies have been used: anti-USP2a (N-term and C-term, Abgent, San Diego, CA, USA), anti-c-Myc (clone 9E10, Santa Cruz, CA, USA), anti-procaspase-3 and cleaved caspase-3 (Upstate Biotechnology, New York, NY), anti-cleaved PARP (cPARP, Roche Applied Sciences, Indianapolis, IN, USA), anti-p38 and phospho-p38 (Cell Signaling

Technology, Danvers, MA, USA), anti- $\gamma$ GCS<sub>H</sub> and anti- $\gamma$ GCS<sub>L</sub> (Santa Cruz), anti-GAPDH (6C5, Santa Cruz) and anti-beta actin (AC-74, Sigma).

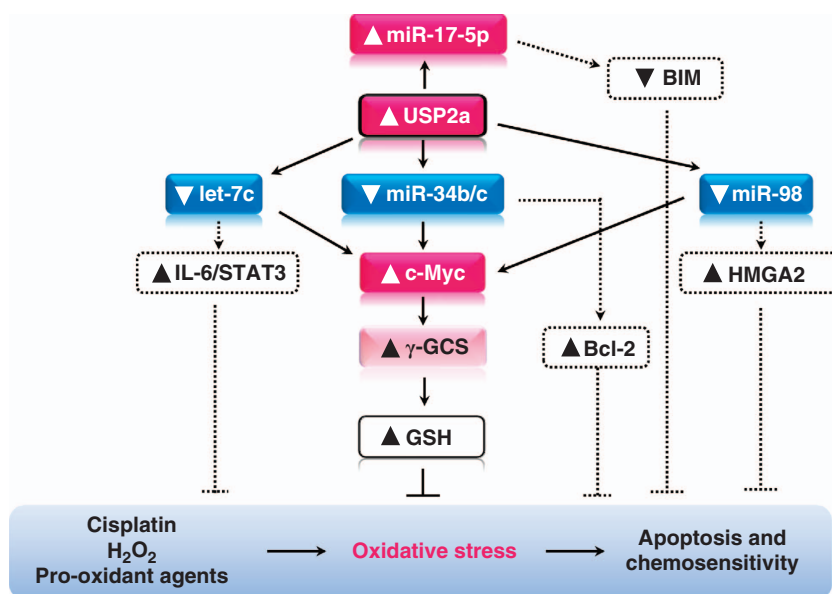
Overexpression experiments have been performed by transfecting cells with pCDNA3 empty vector (Invitrogen, Life Technologies), pCDNA3-USP2a<sup>WT</sup> (pUSP2a<sup>WT</sup>) and pCDNA3-USP2a<sup>MUT</sup> (pUSP2a<sup>MUT</sup>), as previously described.<sup>11,19</sup> Silencing oligonucleotides (siRNA) for knockdown experiments have been purchased from MWG (Huntsville, AL, USA), as previously detailed.<sup>19</sup> Synthetic (Mimic) miRNA sequences (control, miR-34b and 34c) were purchased from Dharmacon (Thermo Scientific, Pittsburgh PA, USA). Lyophilized molecules have been dissolved in diethylpyrocarbonate (DEPC) water, stocked and freshly diluted before experiments, according to the manufacturer's instructions.

**Cell culture and transfection.** Empty vector (Vector), wild-type (USP2a<sup>WT</sup>) and mutant (USP2a<sup>MUT</sup>) stable clones (previously established by infecting immortalized androgen receptor-expressing prostate epithelial cells (iPrEC)<sup>11,19</sup> were grown in specific PrEBM medium (Cambrex, East Rutherford, NJ, USA) and selected in 1.6  $\mu$ g/ml puromycin. Human prostate adenocarcinoma LNCaP cells were obtained from the American Type Culture Collection (ATCC, Manassas, VA, USA) and grown in RPMI-1640 medium (Invitrogen) containing 10% fetal bovine serum (GIBCO-Invitrogen) and 1% penicillin-streptomycin (Invitrogen).

Transient transfection experiments were carried out by seeding cells in 60-mm Petri dishes in complete medium ( $2 \times 10^5$  cells/plate). Transfection was performed 24 h after plating using Lipofectamine 2000 reagent (Invitrogen) in Optimem medium (Invitrogen) for siRNA oligonucleotides (100–500 nM) and synthetic miR molecules (10 nM), and by JetPei reagent (PloyPlus-Transfection, New York, NY, USA) in complete medium for expression vectors (0.5–5  $\mu$ g). Evaluation of protein, mRNA and miRNA expression levels was performed by harvesting cells at 24 h intervals (24–120 h) after transfection.

**Treatment with antineoplastic agents and H<sub>2</sub>O<sub>2</sub>.** For drug treatment, cells were seeded in 60-mm Petri dishes at a density of  $2 \times 10^5$  cells/plate. After 24 h, cells were exposed for 2–24 h to different doses of CDDP (ranging from 0.5 to 10  $\mu$ g/ml), DTX (ranging from 1 to 25 nM) and Doxo (ranging from 0.1 to 1  $\mu$ M). In the experiments with NAC antioxidant, cells were incubated with 5 mM NAC (dose with no toxic effect on cell survival) 6 h before drug administration.

In the experiments with H<sub>2</sub>O<sub>2</sub>, iPrEC cells ( $2 \times 10^5$  cells/plate) were incubated with 100  $\mu$ M of the pro-oxidant agent for 1–12 h and immediately analyzed for apoptosis, GSH content and ROS generation.



**Figure 6** USP2a-activated pathways in response to oxidative stress. The schematized molecular model describes the multiple miRNA-target connections triggered by USP2a overexpression in prostate cells, in response to pro-oxidant agents. Solid lines represent that demonstrated by our present and recently published data.<sup>19</sup> Dotted lines suggest further potential USP2a-miRNA-target cross-talks, underlying the antioxidant and chemo-protective function elicited by USP2a in prostate



**Western blot.** Cell lysis was performed on ice for 30 min in RIPA buffer (50 mM Tris-HCl pH 7.4, 150 mM NaCl, 1% NP-40, 0.25% sodium deoxycholate, 1 mM EDTA) supplemented with phosphatase inhibitors (1 mM PMSF, 1  $\mu$ g/ml aprotinin, leupeptin, pepstatin). Equal amounts of total protein extracts (20–80  $\mu$ g) were resolved by 10%, 12% or 15% denaturing SDS polyacrylamide gel electrophoresis (SDS–PAGE) and transferred for 4 h to the polyvinylidene difluoride membrane. Membranes were blocked in 5% milk-PBS-0.05% Tween 20 for 1 h and incubated overnight with the specific primary antibodies. Secondary antibodies were horseradish peroxidase-conjugated (Santa Cruz), and ECL reagent (Amersham, GE Healthcare, Piscataway, NJ, USA) was employed for chemo-luminescence detection.

**GSH Determination.** Intracellular GSH content was measured as previously described<sup>19</sup>, using a colorimetric assay (Bioxytech GSH-400; Oxis International Inc., Beverly Hills, CA, USA), according to the manufacturer's instruction.

**$\gamma$ GCS enzyme activity.** Enzymatic activity was evaluated by using a coupled assay with pyruvate kinase and lactate dehydrogenase and assaying the rate of decrease in absorbance at 340 nm at 37 °C. Enzyme activity was expressed as mmol of NADH oxidized per minute (U) per mg protein.<sup>34</sup>

**Flow cytometric evaluation of apoptosis.** Apoptosis was detected by both PI staining of 80% ethanol-fixed cells and Annexin V-FITC *versus* PI assay performed in fresh cells, as previously described.<sup>19</sup>

Briefly, ethanol-fixed cells were washed in phosphate buffered saline (PBS), stained in a solution (containing RNase A (50  $\mu$ g/ml, Sigma), Triton 0.1%, EDTA (0.1 mM, Sigma) and PI (50  $\mu$ g/ml) in PBS) and incubated in the dark for at least 30 min. SubG<sub>1</sub> percentage has been determined by analyzing DNA content using the flow cytometer (Beckton and Dickinson, Franklin Lakes, NJ, USA). For the Annexin assay, adherent cells were harvested, suspended in the annexin-binding buffer (about 1  $\times$  10<sup>6</sup> cells/ml) and incubated with the Annexin V-FITC and PI for 15 min, at room temperature in the dark, then immediately analyzed using flow cytometry. These data are presented as bi-parametric dot plots showing the annexin V-FITC green fluorescence *versus* the PI red fluorescence. Analysis of cell death was performed from 0 to 96 h after the end of each drug treatment.

**Assessment of the mitochondrial membrane potential.**  $\Delta\Psi$  was assayed by staining cells with the JC-1 probe, a cationic dye that exhibits mitochondria potential-dependent accumulation, without being affected by the plasmalemma potential. JC-1 accumulates in the cytoplasm, where it produces green fluorescence and forms red fluorescent J-aggregates in the mitochondria. Mitochondria depolarization is indicated by a decrease in the red/green fluorescence intensity ratio. Adherent cells (about 5  $\times$  10<sup>5</sup>) were first assayed for viability and then loaded with 10 mM JC-1 in complete medium, for 30 min at 37 °C in the dark. After incubation, cells were washed twice and suspended in PBS, then immediately analyzed using flow cytometry (Beckton and Dickinson). These data are presented as bi-parametric panels with the green J-monomers fluorescence plotted *versus* the red J-aggregates fluorescence. Analysis of  $\Delta\Psi$  was performed from 0 to 72 h following the end of drug administration.

**Evaluation of the reactive oxygen species.** The evaluation of ROS production was performed as previously described.<sup>19</sup> Briefly, adherent cells (about 5  $\times$  10<sup>5</sup>) were first assayed for viability and then incubated with 4 mM DHE for 45 min at 37 °C in PBS. After incubation, cells were immediately analyzed using flow cytometry. These data are presented as both mono-parametric histograms or bi-parametric panels with the red DHE fluorescence intensity plotted *versus* the forward scatter. Analysis of ROS generation was performed from 0 to 72 h following the end of each drug treatment.

**Statistical analysis.** All data are presented as mean  $\pm$  S.D., calculated in  $\geq$  three replicates. Statistical analyses were performed using an unpaired two-tailed *t* test using SPSS Software Version One. *P*-values <0.05 were taken as statistically significant.

### Conflict of Interest

The authors declare no conflict of interest.

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