



# Measuring the Concentration of Serum Syndecan-1 to Assess Vascular Endothelial Glycocalyx Injury During Hemodialysis

## OPEN ACCESS

### Edited by:

Tengis Pavlov,  
Henry Ford Health System,  
United States

### Reviewed by:

Nirupama Ramkumar,  
The University of Utah, United States  
Sherif Abdelaziz Ibrahim,  
Cairo University, Egypt

### \*Correspondence:

Hideshi Okada  
hideshi@gifu-u.ac.jp

†These authors have contributed  
equally to this work

### Specialty section:

This article was submitted to  
Nephrology,  
a section of the journal  
Frontiers in Medicine

Received: 08 October 2021

Accepted: 07 December 2021

Published: 23 December 2021

### Citation:

Kusuzawa K, Suzuki K, Okada H,  
Suzuki K, Takada C, Nagaya S,  
Yasuda R, Okamoto H, Ishihara T,  
Tomita H, Kawasaki Y, Minamiyama T,  
Nishio A, Fukuda H, Shimada T,  
Tamaoki Y, Yoshida T, Nakashima Y,  
Chiba N, Yoshimura G, Kamidani R,  
Miura T, Oiwa H, Yamaji F, Mizuno Y,  
Miyake T, Kitagawa Y, Fukuta T, Doi T,  
Suzuki A, Yoshida T, Tetsuka N,  
Yoshida S and Ogura S (2021)  
Measuring the Concentration of  
Serum Syndecan-1 to Assess  
Vascular Endothelial Glycocalyx Injury  
During Hemodialysis.  
Front. Med. 8:791309.  
doi: 10.3389/fmed.2021.791309

Keigo Kusuzawa<sup>1†</sup>, Keiko Suzuki<sup>2†</sup>, Hideshi Okada<sup>1\*†</sup>, Kodai Suzuki<sup>1</sup>, Chihiro Takada<sup>1</sup>, Soichiro Nagaya<sup>1</sup>, Ryu Yasuda<sup>1</sup>, Haruka Okamoto<sup>1</sup>, Takuma Ishihara<sup>3</sup>, Hiroyuki Tomita<sup>4</sup>, Yuki Kawasaki<sup>1</sup>, Toru Minamiyama<sup>1</sup>, Ayane Nishio<sup>1</sup>, Hirotsugu Fukuda<sup>1</sup>, Takuto Shimada<sup>1</sup>, Yuto Tamaoki<sup>1</sup>, Tomoki Yoshida<sup>1</sup>, Yusuke Nakashima<sup>1</sup>, Naokazu Chiba<sup>1</sup>, Genki Yoshimura<sup>1</sup>, Ryo Kamidani<sup>1</sup>, Tomotaka Miura<sup>1,2</sup>, Hideaki Oiwa<sup>1,5</sup>, Fuminori Yamaji<sup>1</sup>, Yosuke Mizuno<sup>1</sup>, Takahito Miyake<sup>1</sup>, Yuichiro Kitagawa<sup>1</sup>, Tetsuya Fukuta<sup>1</sup>, Tomoaki Doi<sup>1</sup>, Akio Suzuki<sup>6</sup>, Takahiro Yoshida<sup>1</sup>, Nobuyuki Tetsuka<sup>2</sup>, Shozo Yoshida<sup>1,5</sup> and Shinji Ogura<sup>1</sup>

<sup>1</sup> Department of Emergency and Disaster Medicine, Gifu University Graduate School of Medicine, Gifu, Japan, <sup>2</sup> Department of Infection Control, Gifu University Graduate School of Medicine, Gifu, Japan, <sup>3</sup> Innovative and Clinical Research Promotion Center, Gifu University Hospital, Gifu, Japan, <sup>4</sup> Department of Tumor Pathology, Gifu University Graduate School of Medicine, Gifu, Japan, <sup>5</sup> Abuse Prevention Center, Gifu University Graduate School of Medicine, Gifu, Japan, <sup>6</sup> Department of Pharmacy, Gifu University Hospital, Gifu, Japan

Glycocalyx is present on the surface of healthy endothelium, and the concentration of serum syndecan-1 can serve as an injury marker. This study aimed to assess endothelial injury using serum syndecan-1 as a marker of endothelial glycocalyx injury in patients who underwent hemodialysis. In this single-center, retrospective, observational study, 145 patients who underwent hemodialysis at the Gifu University Hospital between March 2017 and December 2019 were enrolled. The median dialysis period and time were 63 months and 3.7 h, respectively. The serum syndecan-1 concentration significantly increased from  $124.6 \pm 107.8$  ng/ml before hemodialysis to  $229.0 \pm 138.1$  ng/ml after hemodialysis ( $P < 0.001$ ). Treatment with anticoagulant nafamostat mesylate inhibited hemodialysis-induced increase in the levels of serum syndecan-1 in comparison to unfractionated heparin. Dialysis time and the change in the syndecan-1 concentration were positively correlated. Conversely, the amount of body fluid removed and the changes in the syndecan-1 concentration were not significantly correlated. The reduction in the amount of body fluid removed and dialysis time inhibited the change in the syndecan-1 levels before and after hemodialysis. In conclusion, quantitative assessment of the endothelial glycocalyx injury during hemodialysis can be performed by measuring the serum syndecan-1 concentration, which may aid in the selection of appropriate anticoagulants, reduction of hemodialysis time, and the amount of body fluid removed.

**Keywords:** hemodialysis, glycocalyx, syndecan-1, body fluid removal, nafamostat mesylate

## INTRODUCTION

Malnutrition, inflammation, and atherosclerosis are strongly associated with each other in chronic kidney disease (1), and both a malnourished state and atherosclerosis can be caused by inflammation. Moreover, chronic microinflammation is observed in patients who undergo hemodialysis (2). Several factors, such as uremia, activation of free radicals and adhesion molecules, and hemodialysis membrane, can lead to microinflammation in patients who undergo hemodialysis (3–5). Uremic substances and the hemodialysis membrane promote the production of free radicals and cytokines by stimulating neutrophils, and the resulting inflammation further causes endothelial injury. However, there is no method to quantitatively assess endothelial cell injury.

The vascular endothelium is composed of a thin monolayer of endothelial cells, and this lines the entire circulatory system, particularly the parts that are exposed to the circulating blood. The healthy endothelium is covered by a glycoprotein called glycocalyx (6–10), which plays pivotal roles in vascular homeostasis (11, 12). The endothelial glycocalyx is degraded by several factors, such as sepsis, major surgery, trauma, ischemia/reperfusion, and prolonged hyperglycemia. Persistent and diffuse alterations in the glycocalyx are associated with widespread endothelial dysfunction, changed permeability, and impaired oxygen and nutrient delivery to the cells (11, 13, 14). Several studies have revealed the relationship between endothelial glycocalyx injury and serious diseases, such as cardiovascular disease, acute kidney injury, and chronic kidney disease (15, 16). Moreover, the structure of the endothelial glycocalyx is degraded in chronic diseases, such as aging (17), diabetes (18), and hypertriglyceridemia (19).

The glycocalyx comprises cell-bound proteoglycans, glycosaminoglycan side chains, and sialoproteins (13, 20).

The proteoglycans consist of a core protein, such as a member of the syndecan protein family, to which glycosaminoglycan molecules are linked (21). Syndecan-1 is the core protein in heparan sulfate proteoglycan that is observed in the glycocalyx. It is released from the endothelium when the glycocalyx is injured, causing an increase in its concentration in circulation (22). Moreover, serum syndecan-1 has been used as an endothelial injury marker in recent clinical studies in critically ill patients (23–26).

Therefore, the present study aimed to assess endothelial injury using the serum syndecan-1 level as a marker of endothelial glycocalyx injury in patients who underwent hemodialysis. Additionally, we examined the medication type and factors that influence the concentration of syndecan-1.

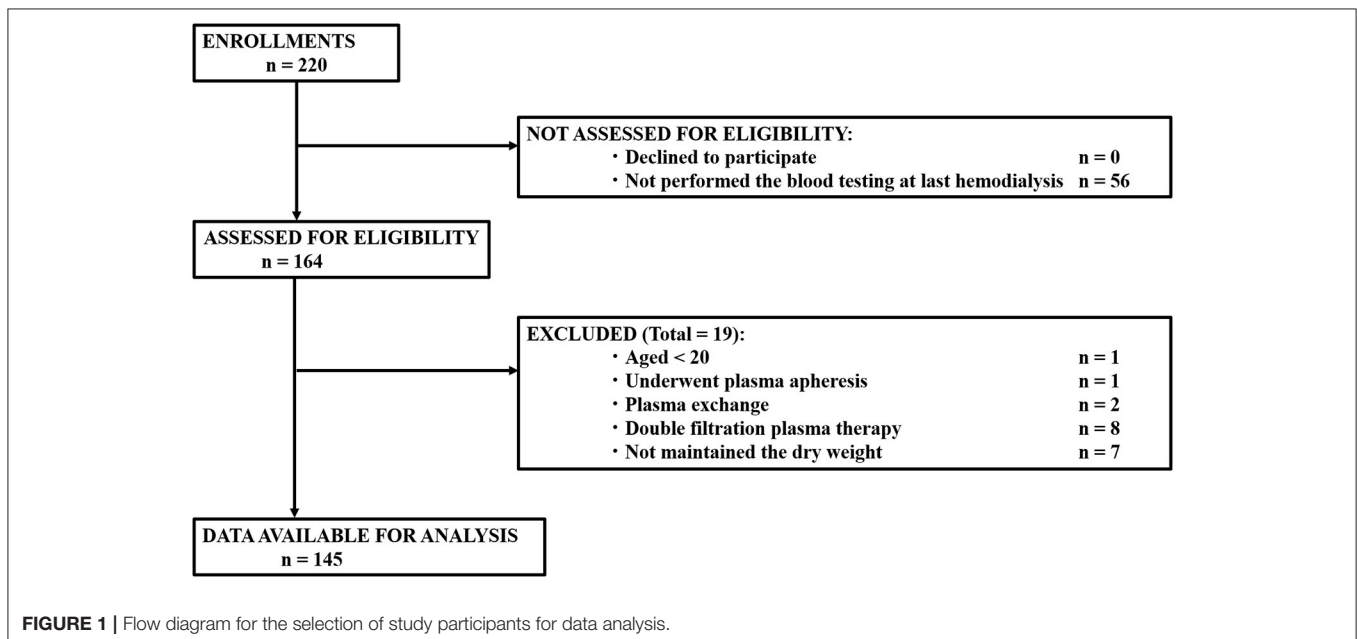
## METHODS

### Patients

This was a single-center, retrospective, observational study conducted at the Gifu University Hospital, affiliated to Gifu University, Gifu, Japan. Patients, who underwent hemodialysis at the Gifu University Hospital between March 2017 and December 2019 and whose dry weight remained unchanged in the last three examinations, were enrolled. Patients aged <20 years, who underwent plasma apheresis, plasma exchange, and double filtration plasma therapy, and had not maintained their dry weight were excluded. Finally, data from 145 patients were obtained and analyzed (Figure 1).

### Ethics Approval and Consent to Participate

The study conformed to the principles outlined in the Declaration of Helsinki (43). Ethics approval was obtained from the Medical Ethics Committee of the Gifu University Graduate



School of Medicine, Gifu, Japan (approval no.: 2021-A005). The need for informed consent from participants was waived by the medical ethics committee because of the retrospective nature of the study. Before initiation, the study was registered in the UMIN Clinical Trials Registry (registry number: UMIN000 051415).

## Data Collection and Study Design

Blood was routinely sampled before and after hemodialysis from eligible patients at the time of the last hemodialysis before being discharged from Gifu University Hospital, and data from these blood samples were used in the present study. All laboratory data (except serum syndecan-1), dry weight, and other patient demographics were extracted from the hospital's electronic medical records. The concentration of serum syndecan-1 was measured using an enzyme-linked immunosorbent assay (950.640.192; Diaclone, Besancon, Cedex, France). The data were retrospectively analyzed. As an index of the efficiency of dialysis, Kt/V was calculated as described previously (44).

## Statistical Analysis

The sample size in this study was calculated to obtain sufficient amount of data and conditions to avoid overfitting in multivariable regression analysis. It was necessary to collect at least 90 patients to estimate the parameters of the six covariates, including the interaction term in the multivariable regression model (45). Patients' baseline characteristics are presented as medians and interquartile ranges for continuous variables, and as frequencies and proportions for categorical variables. For the primary analysis, a mixed effect model was used to assess the change in the syndecan-1 levels with hemodialysis. The difference in the syndecan-1 levels before and after hemodialysis in the anticoagulant subgroup was confirmed using paired *t*-test. A multivariable linear regression analysis was performed to compare the change in the syndecan-1 levels before/after hemodialysis and treatment with anticoagulants. The covariates in the regression model were age, sex, dry weight, and dialysis period (46). These variables were selected *a priori* based on previous studies. In another model, dry weight and dialysis period were simultaneously incorporated into the linear regression model to evaluate the effect of factors during dialysis on the levels of syndecan-1. An interaction term was included in the model to confirm the effect of modification of dry weight and dialysis period on changes in the levels of syndecan-1. If the interaction term was statistically significant, the effect of the dialysis period (or dry weight) on syndecan-1 was determined to be modified by dry weight (or the dialysis period). There were no missing values in the data used in the analyses. A two-sided *P*-value < 0.05 was considered to be statistically significant. The study was exploratory and there were concerns concerning the low statistical power; therefore, the interaction was evaluated with a two-sided *P* < 0.1. R version 4.1.0 was used for statistical analyses (R Foundation for Statistical Computing, Vienna, Austria).

**TABLE 1 |** Patient demographics.

Characteristics	Median (range) or number
Number of cases, <i>n</i>	145
Age (years), mean (IQR)	68 (60–77)
Sex (female/male), <i>n</i> (%)	44 (30.3)/101 (69.7)
Dialysis period (months), median (IQR)	20.0 (1.0–87.0)
Dialysis time (h), median (IQR)	4.0 (3.0–4.0)
Systolic blood pressure before dialysis (mmHg), median (IQR)	136.0 (121.0–157.0)
Diastolic blood pressure before dialysis (mmHg), median (IQR)	69.0 (59.0–79.0)
Primary illness, <i>n</i> (%)	
Chronic glomerulonephritis	29 (20.0)
Rapidly progressive glomerulonephritis	4 (2.8)
Polycystic kidney disease	8 (5.5)
Nephrosclerosis	8 (5.5)
Diabetic nephropathy	37 (25.5)
Nephritis with autoimmune disease	6 (4.1)
Renal/urological tumor	5 (3.4)
Obstructive urinary tract/urination disorders	1 (0.7)
Paraproteinemia (myeloma)	1 (0.7)
Acute kidney injury	10 (6.9)
Congenital anomalies of the kidney and urinary tract	1 (0.7)
Unknown	33 (22.8)
Others	2 (1.4)
Hemodialysis type, <i>n</i> (%)	
HD	133 (91.7)
HDF	12 (8.3)
Vascular access type, <i>n</i> (%)	
Arteriovenous fistula	107 (73.8)
Arteriovenous graft	8 (5.5)
Temporary vascular catheter	23 (15.9)
Permanent vascular catheter	7 (4.8)
Dialysis efficiency	
Kt/V	1.20 (0.06–1.86)
Medication, <i>n</i> (%)	
Unfractionated heparin	101 (69.7)
Low-molecular-weight heparin	24 (16.6)
Nafamostat mesylate	20 (13.8)
Dialysis membrane, <i>n</i> (%)	
Polyethersulfone	110 (75.9)
Polysulfone	34 (23.4)
Asymmetric triacetate	1 (0.7)

HD, hemodialysis; HDF, hemodiafiltration; IQR, interquartile range.

## RESULTS

### Characteristics of Patients

We finally enrolled 145 patients with a median age of 66 years (Figure 1; Table 1). The median dialysis period and time were 63 months and 3 h and 45 min, respectively. The most common primary illness was diabetic nephropathy, which was observed in 37 patients (25.5%).

**TABLE 2** | Serum SDC-1 concentration before and after hemodialysis.

Status	SDC-1 concentration (ng/ml)	P-value
Before HD	124.6 ± 107.8	<0.001
After HD	229.0 ± 138.1	

SDC-1, syndecan-1; HD, hemodialysis. P-values were obtained from a mixed-effects model.

**TABLE 3** | Serum SDC-1 concentration and anticoagulants.

Anticoagulants	Before HD (ng/ml)	After HD (ng/ml)	P-value
Unfractionated heparin	112.0 ± 79.8	235.4 ± 126.8*	<0.001
Low-molecular-weight heparin	144.1 ± 135.8	248.5 ± 174.1*	<0.001
Nafamostat mesylate	164.2 ± 171.1	173.3 ± 141.3	0.459

SDC-1, syndecan-1; HD, hemodialysis.

\*Statistically significant ( $P < 0.05$ ).

The number of patients who underwent hemodialysis and hemodiafiltration was 133 (91.7%) and 12 (8.3%), respectively. Anticoagulation agents, such as unfractionated heparin, low-molecular-weight heparin, and nafamostat mesylate, were administered to 101, 24, and 20 patients, respectively. The median Kt/V-value, an index of dialysis efficiency, was 1.20.

## Concentration of Serum Syndecan-1 and Hemodialysis

The concentrations of serum syndecan-1 before and after hemodialysis were  $124.6 \pm 107.8$  and  $229.0 \pm 138.1$  ng/ml, respectively; this indicated that the serum syndecan-1 concentration significantly increased ( $P < 0.001$ ) after hemodialysis (Table 2).

The concentration of serum syndecan-1 significantly increased after hemodialysis in patients who received unfractionated heparin and low-molecular-weight heparin; however, the concentration of syndecan-1 was not significantly different before and after hemodialysis in those who received nafamostat mesylate (Table 3).

Additionally, according to the multivariable regression analysis after adjusting for age, sex, dry weight, and dialysis period, the treatment with nafamostat mesylate inhibited the increase in the concentration of serum syndecan-1 during hemodialysis compared to treatment with unfractionated heparin and low-molecular-weight heparin (Table 4).

Interestingly, there was no strong relationship between the syndecan-1 levels and blood pressure (Supplementary Table 1), vascular access (Supplementary Table 2), cardiovascular disease (Supplementary Table 3), and primary disease (Supplementary Table 4) before and after dialysis.

## Association of Concentration of Serum Syndecan-1 With Dialysis Time and Body Fluid Removal

The relationship between the concentration variability of syndecan-1 and the dialysis condition, including the dialysis time

and the amount of body fluid removed, was confirmed. The amount of body fluid removal was corrected by the dry weight. The dialysis time and change in concentration of syndecan-1 showed a positive correlation ( $P = 0.033$ ), but there was no significant association ( $P = 0.111$ ) between the amount of body fluid removed and the syndecan-1 concentration changes (Table 5).

Next, we examined the modifying effect of the amount of body fluid removed on the association between the change in the concentration of syndecan-1 and dialysis time. The change in the syndecan-1 concentration before and after hemodialysis increased with respect to enhanced removal of body fluids and prolonged dialysis time ( $P$  for interaction = 0.063, Figure 2). However, the change in the syndecan-1 concentration before and after hemodialysis decreased with respect to a decrease in the amount of body fluid removed and shortened dialysis time.

## DISCUSSION

The present study revealed the following: (a) the endothelial glycocalyx may be injured during hemodialysis; (b) endothelial glycocalyx injury may be attenuated by administering nafamostat mesylate as an anticoagulant; and (c) endothelial glycocalyx injury may be aggravated by an increase in the amount of body fluid removed and prolonged dialysis time.

A previous study reported that the serum syndecan-1 concentration was approximately 20 ng/ml in healthy individuals (19); in contrast, in this study, it was found to be  $124.6 \pm 107.8$  ng/ml in patients who underwent hemodialysis. This result confirmed that the endothelial glycocalyx sustained injuries in patients who underwent hemodialysis, consistent with the findings of a previous report (27). This indicates that endothelial glycocalyx injury may be aggravated by an increasing fluid volume.

Moreover, the serum syndecan-1 concentration increased after hemodialysis compared to the corresponding value before hemodialysis. This finding suggested that the endothelial glycocalyx was injured during hemodialysis, probably because of the production of free radicals and cytokines during this procedure.

During hemodialysis, unfractionated heparin, low-molecular-weight heparin, and nafamostat mesylate were used as anticoagulating agents. Low-molecular-weight heparin and nafamostat were administered to patients that had any disease that was associated with bleeding tendencies. Our results suggested that an increase in the syndecan-1 concentration was attenuated in patients who received nafamostat mesylate.

Nafamostat mesylate, a synthetic serine protease inhibitor, is a short-acting anticoagulant (28), and is also used during hemodialysis to prevent proteolysis of fibrinogen into fibrin (29). It is a slow, tight-binding substrate that traps the target protein in the acyl-enzyme intermediate form and inhibits enzyme activity (30, 31). It was previously reported that nafamostat mesylate can inhibit the kallikrein-kinin system, which promoted vascular permeability via the produced bradykinin (32–34). In addition, nafamostat has been recently identified as a potential therapy

**TABLE 4** | Results of multivariable regression analysis between anticoagulants.

Anticoagulants	Coefficient*	95% LCL	95% UCL	P-value
Low-molecular-weight heparin vs. unfractionated heparin	-18.07	-56.776	20.637	0.358
Nafamostat mesylate vs. unfractionated heparin	-116.473	-158.442	-74.504	<0.001
Nafamostat mesylate vs. low-molecular-weight heparin	-98.403	-150.482	-45.324	<0.001

LCL, lower confidence limit; UCL, upper confidence limit.

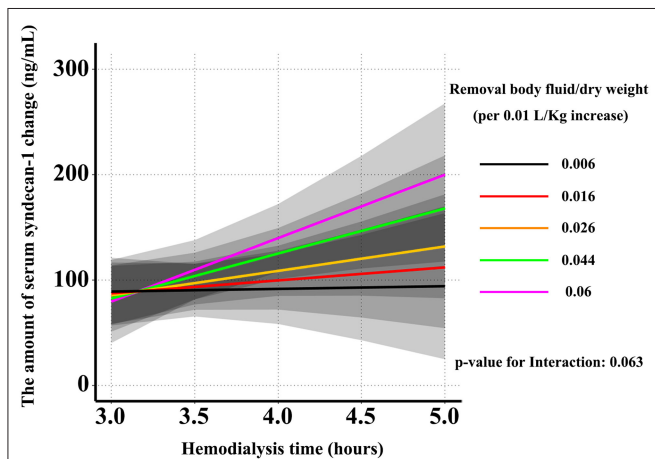
\*Coefficients from the multivariable linear regression model adjusted for age, sex, dry weight, and dialysis period, shown as differences in the serum syndecan-1 concentration for low-molecular-weight heparin vs. unfractionated heparin, nafamostat mesylate vs. unfractionated heparin, and nafamostat mesylate vs. low-molecular-weight heparin, respectively.

**TABLE 5** | Relationship between syndecan-1 concentration variability and dialysis conditions.

Factors	Coefficient*	95% LCL	95% UCL	P-value
Body fluid removed/dry weight (per 0.01 L/kg increase)	9.107	0.144	18.07	0.111
Dialysis time (per 1 min increase)	23.349	-8.836	55.533	0.033

LCL, lower confidence limit; UCL, upper confidence limit.

\*Coefficients from the multivariable linear regression model adjusted for age, sex, and dialysis period, shown as increment in the serum syndecan-1 concentration for a unit change in factors.



**FIGURE 2** | Effect of dialysis time and the amount of body fluid removed on the change in the serum syndecan-1 concentration. The change in the concentration of syndecan-1 before and after hemodialysis increased with respect to the enhanced body fluid removal and prolonged dialysis time. However, the change in the concentration of syndecan-1 before and after hemodialysis decreased with respect to the decreased amount of body fluid removal and the shortened dialysis time.

against the coronavirus disease (35). Infection with severe acute respiratory syndrome coronavirus 2 induces endotheliitis due to viral involvement and inflammatory response of the host and, thus, it is associated with endothelial glycocalyx injury (21). Therefore, nafamostat mesylate may have a beneficial effect on the endothelial glycocalyx, although this is supported only by circumstantial evidence.

Extension of dialysis time is a strategy to improve prognosis (36); however, it remains controversial (36, 37). The present study identified that changes in the serum syndecan-1 levels are small in patients who have prolonged dialysis time and

slow removal of body fluid. Therefore, these two strategies could prevent endothelial glycocalyx injury. Several reports have also revealed that prolonged hemodialysis was associated with improved blood pressure and fluid management (38–40). Additionally, rapid removal of body fluid is associated with a greater risk of mortality and cardiovascular events (41). Moreover, hypotension during hemodialysis is also associated with higher mortality (42).

These mechanisms may explain how lower ultrafiltration rates with prolonged hemodialysis and slow removal of body fluids may ameliorate endothelial vascular permeability via attenuation of endothelial glycocalyx injury. Therefore, we propose that slow removal of body fluids with prolonged hemodialysis can reduce hypotension during hemodialysis.

This study had some limitations. First, the hemodialysis time in most patients was <4 h. Therefore, an accurate examination of prolonged hemodialysis could not be performed. Second, less types of dialyzer were used in the present study. Third, in this study, other biomarkers of glycocalyx injury, such as the serum hyaluronan and hyaluronidase levels, were not measured. Although further studies are required, measuring the concentration of serum syndecan-1 may help assessing endothelial injury under low blood flow (e.g., chronic hemodiafiltration) and membrane compatibility by using a different type of dialyzer. Moreover, the serum syndecan-1 level is proposed to be a useful biomarker for daily monitoring of organ dysfunction, and may be an important risk factor for mortality in critically ill patients (25).

In conclusion, the study presented a method for the quantitative assessment of endothelial glycocalyx injury by measuring the concentration of serum syndecan-1 during hemodialysis. Although hemodialysis causes endothelial glycocalyx injury, it may be mitigated by maintenance of hemodialysis duration and by modulation of the amount of body fluid removed via the quantitative assessment of the serum syndecan-1 level.

## DATA AVAILABILITY STATEMENT

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

## ETHICS STATEMENT

Ethics approval was obtained from the Medical Ethics Committee of the Gifu University Graduate School of Medicine, Gifu, Japan (record no.: 2021-A005). Written informed consent for participation was not required for this study in accordance with the national legislation and the institutional requirements.

## AUTHOR CONTRIBUTIONS

KK and HOkad wrote the manuscript. KK, KoS, SN, HOkam, GY, RK, and TMiu collected the blood samples. FY, HOi, CT, YKa, TMin, AN, HF, TS, YT, ToY, YN, and NC measured the syndecan-1 concentration using ELISA. KeS and RY created a database. TI performed a statistical analysis. YM, TMiy, YKi, TF, TD, and TaY treated the patients. HT, SY, and SO supervised the study. HOkad and AS revised and edited the

manuscript. All authors contributed to the article and approved the submitted version.

## FUNDING

This work was supported by the Ministry of Education, Science and Culture of Japan grants-in-aid for scientific research (grant numbers 19H03756; HOkad, 19K18347; TF, 19K09410; TD, and 18K16511; SY).

## ACKNOWLEDGMENTS

The authors would like to thank the paramedical crew who shared the data obtained by them with us and allowed us to use the data for the writing of this report. The authors also thank Editage ([www.editage.com](http://www.editage.com)) for English language editing.

## SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fmed.2021.791309/full#supplementary-material>

## REFERENCES

- Stenvinkel P, Heimbürger O, Paultre F, Diczfalussy U, Wang T, Berglund L, et al. Strong association between malnutrition, inflammation, and atherosclerosis in chronic renal failure. *Kidney Int.* (1999) 55:1899–911. doi: 10.1046/j.1523-1755.1999.00422.x
- Zimmermann J, Herrlinger S, Pruy A, Metzger T, Wanner C. Inflammation enhances cardiovascular risk and mortality in hemodialysis patients. *Kidney Int.* (1999) 55:648–58. doi: 10.1046/j.1523-1755.1999.00273.x
- Kultz D. Hyperosmolality triggers oxidative damage in kidney cells. *Proc Natl Acad Sci USA.* (2004) 101:9177–8. doi: 10.1073/pnas.0403241101
- Rashid G, Benchetrit S, Fishman D, Bernheim J. Effect of advanced glycation end-products on gene expression and synthesis of TNF-alpha and endothelial nitric oxide synthase by endothelial cells. *Kidney Int.* (2004) 66:1099–106. doi: 10.1111/j.1523-1755.2004.00860.x
- Kawabata K, Nakai S, Miwa M, Sugiura T, Otsuka Y, Shinzato T, et al. CD31 expression on leukocytes is downregulated *in vivo* during hemodialysis. *Nephron.* (2001) 89:153–60. doi: 10.1159/000046062
- Ando Y, Okada H, Takemura G, Suzuki K, Takada C, Tomita H, et al. Brain-specific ultrastructure of capillary endothelial glycocalyx and its possible contribution for blood brain barrier. *Sci Rep.* (2018) 8:17523. doi: 10.1038/s41598-018-35976-2
- Becker BF, Chappell D, Jacob M. Endothelial glycocalyx and coronary vascular permeability: the fringe benefit. *Basic Res Cardiol.* (2010) 105:687–701. doi: 10.1007/s00395-010-0118-z
- Inagawa R, Okada H, Takemura G, Suzuki K, Takada C, Yano H, et al. Ultrastructural alteration of pulmonary capillary endothelial glycocalyx during endotoxemia. *Chest.* (2018) 154:317–25. doi: 10.1016/j.chest.2018.03.003
- Okada H, Takemura G, Suzuki K, Oda K, Takada C, Hotta Y, et al. Three-dimensional ultrastructure of capillary endothelial glycocalyx under normal and experimental endotoxemic conditions. *Crit Care.* (2017) 21:261. doi: 10.1186/s13054-017-1841-8
- Rehm M, Zahler S, Lötsch M, Welsch U, Conzen P, Jacob M, et al. Endothelial glycocalyx as an additional barrier determining extravasation of 6% hydroxyethyl starch or 5% albumin solutions in the coronary vascular bed. *Anesthesiology.* (2004) 100:1211–23. doi: 10.1097/0000542-200405000-00025
- Chelazzi C, Villa G, Mancinelli P, De Gaudio AR, Adembri C. Glycocalyx and sepsis-induced alterations in vascular permeability. *Crit Care.* (2015) 19:26. doi: 10.1186/s13054-015-0741-z
- Woodcock TE, Woodcock TM. Revised Starling equation and the glycocalyx model of transvascular fluid exchange: an improved paradigm for prescribing intravenous fluid therapy. *Br J Anaesth.* (2012) 108:384–94. doi: 10.1093/bja/aer515
- Salmon AH, Satchell SC. Endothelial glycocalyx dysfunction in disease: albuminuria and increased microvascular permeability. *J Pathol.* (2012) 226:562–74. doi: 10.1002/path.3964
- Henrich M, Gruss M, Weigand MA. Sepsis-induced degradation of endothelial glycocalyx. *ScientificWorldJournal.* (2010) 10:917–23. doi: 10.1100/tsw.2010.88
- Liborio AB, Braz MB, Seguro AC, Meneses GC, Neves FM, Pedrosa DC, et al. Endothelial glycocalyx damage is associated with leptospirosis acute kidney injury. *Am J Trop Med Hyg.* (2015) 92:611–6. doi: 10.4269/ajtmh.14-0232
- Padberg JS, Wiesinger A, di Marco GS, Reuter S, Grabner A, Kentrup D, et al. Damage of the endothelial glycocalyx in chronic kidney disease. *Atherosclerosis.* (2014) 234:335–43. doi: 10.1016/j.atherosclerosis.2014.03.016
- Machin DR, Bloom SI, Campbell RA, Phuong TTT, Gates PE, Lesniewski LA, et al. *Advanced age results in a diminished endothelial glycocalyx Am J Physiol Heart Circ Physiol.* (2018) 315:H531–9. doi: 10.1152/ajpheart.00104.2018
- Nieuwdorp M, van Haften TW, Gouverneur MC, Mooij HL, van Lieshout MH, Levi M, et al. Loss of endothelial glycocalyx during acute hyperglycemia coincides with endothelial dysfunction and coagulation activation *in vivo*. *Diabetes.* (2006) 55:480–6. doi: 10.2337/diabetes.55.02.06.db05-1103
- Oda K, Okada H, Suzuki A, Tomita H, Kobayashi R, Sumi K, et al. Factors enhancing serum syndecan-1 concentrations: a large-scale comprehensive medical examination. *J Clin Med.* (2019) 8:1320. doi: 10.3390/jcm8091320
- Paulus P, Jennewein C, Zacharowski K. Biomarkers of endothelial dysfunction: can they help us deciphering systemic inflammation and sepsis? *Biomarkers.* (2011) 16:S11–21. doi: 10.3109/1354750X.2011.587893
- Okada H, Yoshida S, Hara A, Ogura S, Tomita H. Vascular endothelial injury exacerbates coronavirus disease 2019: the role of endothelial glycocalyx protection. *Microcirculation.* (2021) 28:e12654. doi: 10.1111/micc.12654
- Rehm M, Bruegger D, Christ F, Conzen P, Thiel M, Jacob M, et al. Shedding of the endothelial glycocalyx in patients undergoing major vascular

- surgery with global and regional ischemia. *Circulation*. (2007) 116:1896–906. doi: 10.1161/CIRCULATIONAHA.106.684852
23. Ostrowski SR, Haase N, Müller RB, Möller MH, Pott FC, Perner A, et al. Association between biomarkers of endothelial injury and hypocoagulability in patients with severe sepsis: a prospective study. *Crit Care*. (2015) 19:191. doi: 10.1186/s13054-015-0918-5
  24. Puskarich MA, Cornelius DC, Tharp J, Nandi U, Jones AE. Plasma syndecan-1 levels identify a cohort of patients with severe sepsis at high risk for intubation after large-volume intravenous fluid resuscitation. *J Crit Care*. (2016) 36:125–9. doi: 10.1016/j.jcrc.2016.06.027
  25. Suzuki K, Okada H, Sumi K, Tomita H, Kobayashi R, Ishihara T, et al. Serum syndecan-1 reflects organ dysfunction in critically ill patients. *Sci Rep*. (2021) 11:8864. doi: 10.1038/s41598-021-88303-7
  26. Suzuki K, Okada H, Tomita H, Sumi K, Kakino Y, Yasuda R, et al. Possible involvement of Syndecan-1 in the state of COVID-19 related to endothelial injury. *Thromb J*. (2021) 19:5. doi: 10.1186/s12959-021-00258-x
  27. Berg S, Golster M, Lisander B. Albumin extravasation and tissue washout of hyaluronan after plasma volume expansion with crystalloid or hypooncotic colloid solutions. *Acta Anaesthesiol Scand*. (2002) 46:166–72. doi: 10.1034/j.1399-6576.2002.460207.x
  28. Al-Horani RA, Desai UR. Recent advances on plasmin inhibitors for the treatment of fibrinolysis-related disorders. *Med Res Rev*. (2014) 34:1168–216. doi: 10.1002/med.21315
  29. Sadahiro T, Yuzawa H, Kimura T, Oguchi M, Morito T, Mizushima S, et al. Current practices in acute blood purification therapy in Japan and topics for further study. *Contrib Nephrol*. (2018) 196:209–14. doi: 10.1159/000485724
  30. Ramjee MK, Henderson IM, McLoughlin SB, Padova A. The kinetic and structural characterization of the reaction of nafamostat with bovine pancreatic trypsin. *Thromb Res*. (2000) 98:559–69. doi: 10.1016/S0049-3848(00)00206-1
  31. Ramjee MK, Patel S. Continuous-flow injection microfluidic thrombin assays: the effect of binding kinetics on observed enzyme inhibition. *Anal Biochem*. (2017) 528:38–46. doi: 10.1016/j.ab.2017.04.016
  32. Fujii S, Hitomi Y. New synthetic inhibitors of C1r, C1 esterase, thrombin, plasmin, kallikrein and trypsin. *Biochim Biophys Acta*. (1981) 661:342–5. doi: 10.1016/0005-2744(81)90023-1
  33. Hitomi Y, Ikari N, Fujii S. Inhibitory effect of a new synthetic protease inhibitor (FUT-175) on the coagulation system. *Haemostasis*. (1985) 15:164–8. doi: 10.1159/000215139
  34. Paques EP, Romisch J. Comparative study on the *in vitro* effectiveness of antithrombotic agents. *Thromb Res*. (1991) 64:11–21. doi: 10.1016/0049-3848(91)90201-7
  35. Wang M, Cao R, Zhang L, Yang X, Liu J, Xu M, et al. Remdesivir and chloroquine effectively inhibit the recently emerged novel coronavirus (2019-nCoV) *in vitro*. *Cell Res*. (2020) 30:269–71. doi: 10.1038/s41422-020-0282-0
  36. Chertow GM, Levin NW, Beck GJ, Daugirdas JT, Eggers PW, Klinger AS, et al. Long-term effects of frequent in-center hemodialysis. *J Am Soc Nephrol*. (2016) 27:1830–6. doi: 10.1681/ASN.2015040426
  37. Rocco MV, Daugirdas JT, Greene T, Lockridge RS, Chan C, Pierratos A, et al. Long-term effects of frequent nocturnal hemodialysis on mortality: the Frequent Hemodialysis Network (FHN) nocturnal trial. *Am J Kidney Dis*. (2015) 66:459–68. doi: 10.1053/j.ajkd.2015.02.331
  38. Burton JO, Jefferies HJ, Selby NM, McIntyre CW. Hemodialysis-induced repetitive myocardial injury results in global and segmental reduction in systolic cardiac function. *Clin J Am Soc Nephrol*. (2009) 4:1925–31. doi: 10.2215/CJN.04470709
  39. Lacson E Jr, Xu J, Suri RS, Nesrallah G, Lindsay R, Garg AX, et al. Survival with three-times weekly in-center nocturnal versus conventional hemodialysis. *J Am Soc Nephrol*. (2012) 23:687–95. doi: 10.1681/ASN.2011070674
  40. Ok E, Duman S, Ascı G, Tumuklu M, Onen Sertoz O, Kayikcioglu M, et al. Comparison of 4- and 8-h dialysis sessions in thrice-weekly in-centre haemodialysis: a prospective, case-controlled study. *Nephrol Dial Transplant*. (2011) 26:1287–96. doi: 10.1093/ndt/gfq724
  41. Flythe JE, Kimmel SE, Brunelli SM. Rapid fluid removal during dialysis is associated with cardiovascular morbidity and mortality. *Kidney Int*. (2011) 79:250–7. doi: 10.1038/ki.2010.383
  42. Flythe JE, Xue H, Lynch KE, Curhan GC, Brunelli SM. Association of mortality risk with various definitions of intradialytic hypotension. *J Am Soc Nephrol*. (2015) 26:724–34. doi: 10.1681/ASN.2014020222
  43. Rickham PP. Human experimentation. Code of ethics of the World Medical Association Declaration of Helsinki. *Br Med J*. (1964) 2:177.
  44. Daugirdas JT. Second generation logarithmic estimates of single-pool variable volume Kt/V: an analysis of error. *J Am Soc Nephrol*. (1993) 4:1205–13. doi: 10.1681/ASN.V451205
  45. Harrell FE. *Regression Modeling Strategies: With Applications to Linear Models, Logistic and Ordinal Regression, and Survival Analysis*. New York, NY: Springer. (2015). doi: 10.1007/978-3-319-19425-7
  46. Koch J, Idzerda NMA, Ettema EM, Kuipers J, Dam W, van den Born J, et al. An acute rise of plasma Na(+) concentration associates with syndecan-1 shedding during hemodialysis. *Am J Physiol Renal Physiol*. (2020) 319:F171–7. doi: 10.1152/ajprenal.00005.2020

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

**Publisher's Note:** All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2021 Kusuzawa, Suzuki, Okada, Suzuki, Takada, Nagaya, Yasuda, Okamoto, Ishihara, Tomita, Kawasaki, Minamiyama, Nishio, Fukuda, Shimada, Tamaoki, Yoshida, Nakashima, Chiba, Yoshimura, Kamidani, Miura, Oiwa, Yamaji, Mizuno, Miyake, Kitagawa, Fukuta, Doi, Suzuki, Yoshida, Tetsuka, Yoshida and Ogura. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.