

The *EIF2AK4*/rs4594236 AG/GG Genotype Is a Hazard Factor of Immunoglobulin Therapy Resistance in Southern Chinese Kawasaki Disease Patients

Hongyan Yu^{1†}, Fucheng Liu^{2†}, Kaining Chen¹, Yufen Xu¹, Yishuai Wang¹, Lanyan Fu¹, Huazhong Zhou¹, Lei Pi¹, Di Che¹, Hehong Li^{3*} and Xiaoqiong Gu^{1,4*}

OPEN ACCESS

Edited by:

Tieliu Shi, Hunan University of Arts and Science, China

Reviewed by:

Fatma Savran Oguz, Istanbul University, Turkey Hiromichi Hamada, Chiba University, Japan

*Correspondence:

Hehong Li hornc@163.com Xiaoqiong Gu guxiaoqiong@gwcmc.org

[†]These authors have contributed equally to this work

Specialty section:

This article was submitted to Genetics of Common and Rare Diseases, a section of the journal Frontiers in Genetics

Received: 02 February 2022 Accepted: 16 May 2022 Published: 22 June 2022

Citation:

Yu H, Liu F, Chen K, Xu Y, Wang Y, Fu L, Zhou H, Pi L, Che D, Li H and Gu X (2022) The EIF2AK4/rs4594236 AG/GG Genotype Is a Hazard Factor of Immunoglobulin Therapy Resistance in Southern Chinese Kawasaki Disease Patients. Front. Genet. 13:868159. doi: 10.3389/fgene.2022.868159 ¹Department of Clinical Biological Resource Bank, Guangdong Provincial Key Laboratory of Research in Structural Birth Defect Disease, Guangzhou Institute of Pediatrics, Guangzhou Women and Children's Medical Center, Guangzhou Medical University, Guangzhou, China, ²Department of Cardiology, The First Affiliated Hospital of Jinan University, Guangzhou, China, ³Department of Radiology, Guangzhou Women and Children's Medical Center, Guangzhou Medical University, Guangzhou, China, ⁴Department of Blood Transfusion and Clinical Lab, Guangzhou Institute of Pediatrics, Guangzhou Women and Children's Medical Center, Guangzhou Medical University, Guangzhou, China

Background: Kawasaki disease (KD) is an acute, self-limited vasculitis disorder of unknown etiology in children. Immunologic abnormalities were detected during the acute phase of KD, which reflected that the effect cells of the activated immune markedlv increased cytokine production. High-dose svstem intravenous immunoglobulin (IVIG) therapy is effective in resolving inflammation from KD and reducing occurrence of coronary artery abnormalities. However, 10%-20% of KD patients have no response to IVIG therapy, who were defined as IVIG resistance. Furthermore, these patients have persistent inflammation and increased risk of developing coronary artery aneurysm (CAA). EIF2AK4 is a stress sensor gene and can be activated by pathogen infection. In addition, the polymorphisms of EIF2AK4 were associated with various blood vessel disorders. However, it remains unclear whether the *EIF2AK4* gene polymorphisms were related to IVIG therapy outcome in KD patients.

Methods: *EIF2AK4*/rs4594236 polymorphism was genotyped in 795 IVIG response KD patients and 234 IVIG resistant KD patients through TaqMan, a real-time polymerase chain reaction. The odds ratios (ORs) and 95% confidence intervals (CIs) were calculated to assess the strength of association between *EIF2AK4*/rs4594236 polymorphism and IVIG therapeutic effects.

Results: Our results showed that the *EIF2AK4*/rs4594236 AG/GG genotype was significantly associated with increased risk to IVIG resistance compared to the AA genotype (AG vs. AA: adjusted ORs = 1.71, 95% Cls = 1.17-2.51, and p = 0.0061; GG vs. AA: adjusted ORs = 2.09, 95% Cls = 1.36-3.23, and p = 0.0009; AG/GG vs. AA: adjusted ORs = 1.27-2.63, and p = 0.0013; and GG vs. AA/AG: adjusted ORs = 1.45, 95% Cl = 1.04-2.02, and p = 0.0306). Furthermore, the stratified analysis of

1

age and gender in the KD cohort indicated that male patients carrying the rs4594236 AG/ GG genotype tends to be more resistant to IVIG therapy than female patients.

Conclusion: These results suggested that *EIF2AK4*/rs4594236 polymorphism might be associated with increased risk of IVIG resistance in southern Chinese KD patients.

Keywords: Kawasaki disease, IVIG resistance, polymorphism, EIF2AK4/rs4594236, intravenous immunoglobulin

INTRODUCTION

Kawasaki disease (KD) is an acute, self-limited vasculitis disease in children aged from 6 months to 5 years (Burns and Glodé, 2004). Immunologic abnormalities were detected during the acute phase of KD, which reflected that the effect cells of the activated immune system markedly increased cytokine production (Burns and Glodé, 2004). The etiology of KD is unknown, while several epidemiological and clinical reports have suggested that KD might be triggered by infectious agents or viruses (Sharma et al., 2021). This was evidenced by the fact that proinflammatory cytokines (IL-6, IL-10, TNFa, and IFNy) were increased significantly in the acute stage of KD (Wang et al., 2013). Since lots of cytokines and activated immune cells attacked medium-sized arteries, especially coronary arteries, 20%-25% of untreated patients will develop coronary artery aneurysm (CAA) (Gersony, 2009), which has made KD the leading cause of acquired heart disease among children in developed countries (Kato et al., 1975).

Intravenous immunoglobulin (IVIG) contains pooled immunoglobulin G (IgG) from the plasma of over thousand blood donors and is widely used in people with weakened immune systems or other diseases to fight off infections (Lünemann et al., 2015). High-dose IVIG therapy is effective in resolving inflammation from KD and reducing occurrence of CAA. However, 10%-20% of KD patients will develop IVIG resistance, defined by recrudescent or persistent fever for over 36 h after the end of the IVIG infusion primary therapy. In addition, these patients have persistent inflammation and increased risk of developing CAA (McCrindle et al., 2017). Therefore, uncovering the mechanism of IVIG resistance in KD is urgently needed. While the mechanism of IVIG action is complicated and how it works on KD is still confused and unknown, at present, several studies have shown that genetic polymorphisms, especially some immune functional genes, are associated with IVIG resistance, such as inositol 1.4.5-trisposhate 3-kinase C (ITPKC), Fcy IgG receptor 2A (FCGR2A), CD40, and interferon-gamma (IFN-y) (Onouchi et al., 2008; Khor et al., 2011; Lee et al., 2012; Onouchi et al., 2013; Huang et al., 2016). These studies suggested that genetic factors might be involved in IVIG resistance.

Eukaryotic translation initiation factor 2-alpha kinase 4 (*EIF2AK4*, also known as *GCN2*) is a member of the kinase family that phosphorylates the alpha subunit of eukaryotic translation initiation factor-2 (eIF2a)(Wang et al., 2019). EIF2AK4 phosphorylates eIF2a on the serine 51 site and reduces GDP/GTP exchange activity subsequently. In addition, this resulted in mRNA translation changes and subsequently modulated cellular physiological activities (McGaha et al., 2012).

EIF2AK4 mutation was found in patients classified as having idiopathic and heritable pulmonary arterial hypertension (Hadinnapola et al., 2017). Histopathology of EIF2AK4 mutation carriers in pulmonary veno-occlusive disease (PVOD) patients was distinctive from noncarriers regarding arterial remodeling, with significantly more severe intimal fibrosis and less severe medial hypertrophy (Nossent et al., 2018). Furthermore, under nutrientdeprived conditions, EIF2AK4 could promote angiogensis of endothelial cells by increasing VEGF expression (Longchamp et al., 2018). These literature studies showed that EIF2AK4 could modulate vascular remodeling and angiogensis, which are closely associated with coronary arterial lesions (CALs) of KD (Takahashi et al., 2013). What's more, EIF2AK4 was also found to regulate cytokine production and macrophage function in several infectious diseases. Eif2ak4 knockout mice challenged with lipopolysaccharide (LPS) showed reduced inflammatory response, including decreased IL-6 and IL-12 expression, as compared to wild-type mice (Liu et al., 2014). Interestingly, IL-6 and IL-10 were at a high level in the IVIG resistant group compared to the IVIG response group after IVIG treatment (Wang et al., 2013). In inflammatory kidney disease, IFNy-activated EIF2AK4 could suppress proinflammatory cytokine production in glomeruli and reduce macrophage recruitment to the kidneys (Chaudhary et al., 2015), while Ravindran et al. (2016) showed that EIF2AK4 also controlled intestinal inflammation inhibiting inflammasome activation through and IL-1B production. In conclusion, these studies indicate that EIF2AK4 may be associated with KD.

Our team has worked on the area of the etiology and therapy effect of KD for many years (Che et al., 2018; Lin et al., 2021; Wang et al., 2021). We found that the single-nucleotide polymorphism (SNP) of immune and/or cardiovascular-related genes were usually related to IVIG therapy outcome of KD, such as *IL-1β* (Fu et al., 2019), *PLA2G7* (Gu et al., 2020), *P2RY12* (Wang et al., 2020), and *MRP4* (Wang et al., 2021). However, evidence regarding the polymorphisms of *EIF2AK4* and IVIG resistance of KD is very scarce. Based on this background, we performed this epidemiology study to investigate whether *EIF2AK4* is related to IVIG resistance of KD by examining the association between *EIF2AK4* polymorphism (rs4594236) and the risk of IVIG resistance of KD.

MATERIALS AND METHODS

Study Subjects

A total of 1,029 KD patients from the Guangzhou Women and Children's Medical Center between January 2014 and December 2019 were enrolled in this study. All individuals with KD were diagnosed by pediatricians based on the criteria of the American Heart Association (Newburger et al., 2004; McCrindle et al., 2017). IVIG resistance was defined as persistent or recrudescent fever (temperature $\geq 38.0^{\circ}$ C, measured axilla or orally) for over 36 h, but for a period of less than 7 days, after completion of the first IVIG infusion (2 g/kg).

Polymorphism Genotyping and DNA Extraction

Peripheral blood was collected from KD patients after treatment completion. Genomic DNA was extracted with a TIANamp Blood DNA Kit (DP318, TIANGEN Biotech, Beijing) following the guidance of the manufacturer's instructions (Wu et al., 2020). Specific fluorescent allele probes for rs4594236 were purchased from ABI (Thermo Fisher Scientific, United States). PCR was performed in 384-well plates with an ABI-Q6 Sequence Detection System machine (Thermo Fisher Scientific).

The genotyping of the SNP was conducted using a TaqMan SNP genotyping assay (Lin et al., 2021). Laboratory technicians were blind to the sample information, including the identities of the replicate aliquots. 10% of the samples from both groups were arbitrarily chosen to repeat the genotyping results. A concordance rate of 100% was obtained.

Statistical Analysis

Statistical analysis of this study was performed by using SAS software (version 9.4; SAS Institute, Cary, NC). Pearson's chi square test was used to evaluate the significant differences between IVIG response and IVIG resistant cases in the distribution of demographic variables and genotype frequency. Odds ratios (ORs) and 95% confidence intervals (CIs) were calculated by logistic regression analysis for measuring the association between the EIF2AK4/rs4594236 polymorphism and the risk of IVIG treatment resistance in KD patients. Furthermore, stratification analysis was performed, classified by age and gender. We also performed the eQTL analysis using the GTEx Portal web site (https://www. gtexportal.org/home/) to predict potential associations between the SNP and gene-expression levels (Consortium, 2013). A p-value of less than 0.05 was regarded as statistically significant.

RESULTS

Population Characteristics

The characteristic distribution of 795 IVIG therapy response KD patients and 234 IVIG therapy resistant KD patients is shown in **Table 1**. The average age of the IVIG response group was 25.14 ± 20.33 months (rang 1–131 months), and it was 26.08 ± 21.80 months (rang 2–132 months) for the IVIG resistant group. 67.17% of the KD patients who were responsive to IVIG therapy were men, and the male ratio was 72.65% in KD patients who were resistant to IVIG therapy. The proportion of women was 32.83% and 27.35%, respectively, while there were no significant difference in

age (p = 0.3750) and gender (p = 0.1096) between the IVIG response group and the resistant group.

Analysis of the Association Between *EIF2AK4*/rs4594236 Polymorphism and Intravenous Immunoglobulin Resistance

The genotype frequency distribution of EIF2AK4/rs4594236 polymorphism in the KD IVIG resistant group and response group is described in Table 2. To explore the association between EIF2AK4/rs4594236 polymorphism and the risk to IVIG therapy resistance, we performed χ^2 test analysis. We found that EIF2AK4/rs4594236 polymorphism was significantly associated with increased IVIG therapy resistance risk in KD patients (AG vs. AA: adjusted OR = 1.71, 95% confidence interval (CI) = 1.17–2.51, and *p* = 0.0061; GG vs. AA: adjusted OR = 2.09, 95% confidence interval (CI) = 1.36-3.23, and p = 0.0009; AG/GG vs. AA: adjusted OR = 1.82, 95% CI = 1.27–2.63, and *p* = 0.0013; GG vs. AG/AA: adjusted OR = 1.45, 95% confidence interval (CI) = 1.04–2.02, and p = 0.0306). The results indicated that patients with a GG/AG genotype had a higher risk of suffering IVIG therapy resistance than patients with an AA genotype, suggesting the resistive effect of this SNP against IVIG therapy.

Analysis of the Association Between *EIF2AK4*/rs4594236 Polymorphism and Coronary Arterial Lesions

It is well known that *EIF2AK4* is involved in vascular remodeling (Nossent et al., 2018), which is the critical step for CAL formation. Therefore, the association between *EIF2AK4*/rs4594236 polymorphism and CAL formation was explored. Patients with KD were then divided into the CAL group and the NCAL group depending on whether they had CAL or not, and *EIF2AK4*/rs4594236 genotyping was performed on the two groups (**Table 3**). As shown in **Table 3**, *EIF2AK4*/rs4594236 was not associated with CAL formation. We then analyzed the relation between *EIF2AK4*/rs4594236 polymorphism and CAA formation (the serious lesions of the coronary artery) of KD. However, there was no significant association observed between *EIF2AK4*/rs4594236 and CAA (**Table 4**).

Stratification Analysis

We further explored the association between *EIF2AK4*/rs4594236 polymorphism and the risk effect of IVIG resistance on certain subgroups classified by age and gender (**Table 5**). Compared with the rs4594236 AA genotype, the risk effect of the rs4594236 GG/ AG genotype was more prominent in male patients of all ages (adjust OR = 1.91, 95% CI = 1.23–2.95, and p = 0.0039).

Expression Quantitative Trait Loci Analyses

To assess the putative functional relevance of rs4594236 polymorphism affecting *EIF2AK4* mRNA expression, we used the data released from Genotype-Tissue Expression (GTEx) website. It was found that individuals carrying the rs4594236 G allele displayed significantly higher *EIF2AK4* mRNA levels in the artery of the aorta and tibia, the atrial

TABLE 1 | Characteristic distribution in the IVIG therapy resistant group and the response group of KD patients.

Variables	IVIG resistance ($n = 234$)		IVIG respo	pª	
	No.	%	No.	%	
Total	234	100	795	100	
Age range, month	2-132		1-		
Mean ± SD	26.08 ± 21.80		25.14		
≤60	219	93.59	756	95.09	0.3750
>60	15	6.41	39	4.91	
Gender					
Male	170	72.65	534	67.17	0.1096
Female	64	27.35	261	32.83	

^aTwo-sided χ^2 test for distributions between KD patients with the IVIG therapy–resistant and –response group.

TABLE 2 | Genotype distribution frequency of EIF2AK4/rs4594236 polymorphism in the IVIG therapy-resistant group and -response group of KD patients.

Genotype	IVIG resistance (N = 234)	IVIG response (N = 795)	<i>p</i> -value ^a	OR (95% CI)	p-value	Adjusted OR (95% CI)	<i>p</i> -value ^b
AA	43 (18.38)	231 (29.06)	0.0020	1		1	
AG	126 (53.85)	397 (49.94)		1.71 (1.16–2.50)	0.0062	1.71 (1.17–2.51)	0.0061
GG	65 (27.78)	167 (21.01)		2.09 (1.36-3.23)	0.0009	2.09 (1.36-3.23)	0.0009
Additive				1.43 (1.16-1.77)	0.0009	1.43 (1.16–1.77)	0.0009
Dominant	191 (81.62)	564 (70.94)	0.0008	1.82 (1.26-2.63)	0.0013	1.82 (1.27-2.63)	0.0013
Recessive	169 (72.22)	628 (78.99)	0.0322	1.45 (1.04–2.02)	0.0299	1.45 (1.04–2.02)	0.0306

^aTwo-sided χ^2 test for distributions between the IVIG therapy–resistant group and –response group of KD patients.

^bAdjusted for age and gender status in logistic regression models.

TABLE 3 Genotype distribution frequency of EIF2AK4/rs4594236 polymorphism in the NCAL group and CAL group of KD patients.								
Genotype	CAL (N = 408)	NCAL (N = 621)	p-value ^a	OR (95% CI)	p-value	Adjusted OR (95% CI)	p-value ^b	
AA	107 (26.23)	167 (26.89)	0.8297	1.000		1.000		
AG	212 (51.96)	311 (50.08)		1.064 (0.789–1.435)	0.6846	1.041 (0.767-1.414)	0.7945	
GG	89 (21.81)	143 (23.03)		0.971 (0.678–1.391)	0.874	0.941 (0.652-1.358)	0.7447	
Additive				0.989 (0.827-1.182)	0.9024	0.973 (0.811–1.168)	0.7686	
Dominant	301 (73.77)	454 (73.11)	0.8128	1.035 (0.780-1.373)	0.8132	1.010 (0.756-1.349)	0.9482	
Recessive	319 (78.19)	478 (76.97)	0.6481	0.933 (0.691–1.259)	0.65	0.916 (0.674-1.245)	0.5751	

^aTwo-sided χ^2 tests were used to determine differences in genotype distributions between KD with and without CAL.

^bAdjusted for age and gender status in logistic regression models.

TABLE 4 Genotype and allele frequencies of EIF2AK4 in KD Patients with (CAA) or without CAA (NCAA).								
Genotype	CAA (<i>N</i> = 216)	NCAA (N = 813)	p-value ^a	OR (95% CI)	p-value	Adjusted OR (95% CI)	<i>p</i> -value ^b	
AA	55 (25.46)	219 (26.94)	0.8108	1.000		1.000		
AG	114 (52.78)	409 (50.31)		1.110 (0.773–1.593)	0.5717	1.088 (0.754–1.571)	0.6506	
GG	47 (21.76)	185 (22.76)		1.012 (0.654-1.564)	0.9586	0.983 (0.632-1.530)	0.9389	
Additive				1.010 (0.815-1.251)	0.9289	0.995 (0.800-1.237)	0.9648	
Dominant	161 (74.54)	594 (73.06)	0.6619	1.079 (0.766-1.521)	0.6631	1.055 (0.745-1.495)	0.7615	
Recessive	169 (78.24)	628 (77.24)	0.7548	0.944 (0.657–1.356)	0.7556	0.929 (0.643–1.342)	0.6946	

^aTwo-sided χ^2 tests were used to determine differences in genotype distributions between KD with and without CAA.

^bAdjusted for age and gender status in logistic regression models.

appendage, and the left ventricle of the heart than those with the rs4594236 A allele (**Figure 1**). Furthermore, we evaluated the impact of the rs4594236 polymorphism on the mRNA level of the neighboring genes in the above-mentioned tissues and found that

signal recognition particle 14 (SRP14) (or divergent transcript (SRP14-AS1)) mRNA levels in tissues with the rs4594236 G allele were significantly lower than those with the rs4594236 A allele (**Figure 2**).

Variables	rs4594236 (IVIG resistance/IVIG		<i>p</i> -value ^a	OR (95%Cl)	p-value	Adjusted OR (95% Cl)	Adjust <i>p</i> -value ^b
	AA	AG/GG					
Age, months	40/016	177/540	0.0044	1 60 /1 16 0 44)	0.0050	1.67 (1.15, 0.40)	0.0068
≤60 >60	1/15	14/24	0.0044	8.75 (1.04–73.54)	0.0059	9.43 (1.08–82.13)	0.0088
Gender	30/154	140/380	0 0029	1 80 (1 22-2 03)	0.0042	1 01 (1 23-2 05)	0.0039
Female	13/77	51/184	0.1314	1.64 (0.85–3.19)	0.1437	1.62 (0.83–3.15)	0.1551

TABLE 5 | Stratification analysis of EIF2AK4/rs4594236 polymorphism in the IVIG therapy resistant group and response group of KD patients.

^aTwo-sided χ^2 test for distributions between the IVIG therapy resistant group and response group of KD patients.

^bAdjusted for gender/age status in logistic regression models.

DISCUSSION

IVIG has been the optimal and effective treatment for KD to reduce the prevalence of coronary-artery abnormalities and systemic inflammation until now (Newburger et al., 1986). Although the molecular and cellular basis of IVIG function is complicated and remains unknown, some evidence indicated that genetic factors played an important role in IVIG treatment activities. Taking the fact that several genes were associated with the susceptibility of KD and the rates of IVIG resistant patients differ among different ethnic groups (Kashef et al., 2005; Uehara et al., 2008; Tremoulet et al., 2011), some hot genetic factors, especially immune functional genes, were examined to be associated with IVIG resistance, such as FcyR2C and FcyR3B (Makowsky et al., 2013). Furthermore, Weng et al. (2010) had found that patients with IL-1 β (-511 TT) and IL- 1β (-31 CC) genotypes had increased risk of IVIG resistance and were associated with initial IVIG treatment failure based on a study of 156 KD patients (136 with and 20 without response to IVIG treatment) among Taiwanese children .

Herein, we demonstrated a potentially contributing role of *EIF2AK4*/rs4594236 polymorphism in IVIG resistance in KD and for the first time reported that *EIF2AK4*/rs4594236 polymorphism could predispose to IVIG resistance in southern Chinese KD children.

EIF2AK4 is a high molecular weight protein kinase activated by uncharged tRNA (Wek et al., 1990; Nakamura et al., 2018; Schmidt et al., 2019). Activated *EIF2AK4* can phosphorylate eIF2a to upregulate ATF4 translation, which in turn increases amino acid biosynthetic and activated transport pathways (Harding et al., 2000; Harding et al., 2003). The physiological functions of *EIF2AK4* currently remain poorly understood, but its function in human diseases has recently been emphasized.

Several studies demonstrated that *EIF2AK4* was associated with vascular remodeling (Lu et al., 2014; Nossent et al., 2018; Chen et al., 2021). One possible mechanism was that *EIF2AK4* dysfunction enhanced collagen I gene transcription via the ATF3/ p38 pathway, which led to increased collagen deposition in the pulmonary artery (Chen et al., 2021). As vascular remodeling is critical for CAL formation of KD, we also investigated the association between *EIF2AK4*/rs4594236 polymorphism and CAL or CAA formation of KD. However, we found that EIF2AK4/rs4594236 polymorphism was not associated with either CAL or CAA formation. The possible reason may be that the EIF2AK4 expression level was decreased significantly in EIF2AK4 mutation PVOD patients who had undergone vascular modeling. While the data from GTEx showed that individuals carrying the rs4594236 G allele displayed significantly higher EIF2AK4 mRNA levels in the artery of the aorta and tibia, the atrial appendage, and the left ventricle of the heart than those with the rs4594236 A allele (Figure 1), it is indicated that the patients with an rs4594236 G allele have a higher risk of IVIG therapy resistance and the EIF2AK4 expression level. In other words, the patients with higher EIF2AK4 expression tend to have a higher IVIG resistant incidence rate. Hence, different expression levels of EIF2AK4 stimulated diverse downstream signals to regulate cell physiological functions differently.

On the other hand, McGaha et al. found loss of EIF2AK4enhanced inflammatory macrophage transcription with a crowd of proinflammatory cytokine expression and production. In addition, the activated regulatory macrophage was attenuated with a decrease in the Arg1 and CCL22 mRNA expression and IL-10 protein level at the same time. Mechanistically, EIF2AK4 altered the myeloid function by activating the CREB-2/ATF4 signal pathway, which was required for maturation and polarization of macrophages in both mice and humans (Halaby et al., 2019). Interestingly, IVIG treatment promotes tumor-associated macrophages from M2 to M1 polarization, and the IVIG effect was dependent on the activation/polarization state of macrophages (Domínguez-Soto et al., 2014). It is possible that the immunomodulatory effect of IVIG observed in other autoimmune diseases such as KD follows a similar pattern. In addition, our results showed the IVIG resistant risk of KD patients may be linked to the upregulated expression levels of the EIF2AK4 gene (Figure 1); thus, we deduced the hypothesis that IVIG treatment promoted macrophage M1 polarization while the immunomodulatory function of the M1 macrophage was inhibited by upregulated EIF2AK4, which caused persistent inflammation. However, more functional experiments need to be carried out to support the possible mechanism.

To date, studies have been conducted regarding the epidemiologic assessment of *EIF2AK4* gene SNPs. Deng et al.



ventricle of the heart were searched on the public database GTEx Portal. $p = 3.4 \times 10^{-8}$. (D) The genotype of rs4594236 and expression of the *EIF2AK4* gene in the atrial appendage of the heart were searched on the public database GTEx Portal. $p = 2.1 \times 10^{-8}$.

carried out a genome-wide association study of body mass index (BMI) from a cohort containing 597 northern Chinese patients and reported 281,533 SNPs. They found that two adjacent SNPs (rs4432245 and rs711906) of *EIF2AK4* were significantly associated with BMI (Yang et al., 2014). Given the critical role of *EIF2AK4* in immunity reactions, it is necessary to investigate the association between *EIF2AK4* gene SNPs and IVIG resistance of KD. The current study revealed that the SNP rs4594236 polymorphism in the *EIF2AK4* gene was associated with increased risk to IVIG resistance of KD in southern Chinese population. Compared with the rs4594236 AA genotype, the AG/GG genotype increased the IVIG resistant risk significantly, especially in male KD patients. We then explored the potential mechanism for the risk role of

rs4594236 polymorphism in IVIG resistance. The results from eQTLs analysis indicated that IVIG resistant risk of KD was associated with upregulated expression levels of the *EIF2AK4* gene (**Figure 1**). We also evaluated the impact of rs4594236 polymorphism on the mRNA level of the neighboring genes. We found that the *SRP14* (or *SRP14-AS1*) mRNA level with the rs4594236 G genotype was significantly lower than that in cells with the rs4594236 A genotype (**Figure 2**). *SRP14* is a universal ribonucleoprotein, and combined with SRP9 as a heterodimer, it can recognize the RNA UGUNR motif to regulate target gene translation (Bovia et al., 1995; Hasler and Strub, 2006). Thus, we hypothesized that SRP14 combined with the *EIF2AK4* mRNA translation level



to trigger downstream physiological reactions to inhibit IVIG therapy response.

The major strength of this study is its novelty; as we know, this is the first study that focuses on the *EIF2AK4* function in IVIG therapy resistance of KD at present. However, our study still has some limitations. First, the enrolled patients of this study were mainly from the southern Chinese population; the study needs multicenter subjects from other geographic populations to support and evaluate the applicability of the findings to other ethnicities. Second, only one functional SNP in the *EIF2AK4* gene was included in this study; more potentially functional SNPs of *EIF2AK4* need to be investigated in the future. Last but not least, the exact biological mechanism of *EIF2AK4* in IVIG resistance of KD is worthy of further investigation.

DATA AVAILABILITY STATEMENT

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

ETHICS STATEMENT

This study was approved by the Ethics Review Committee of the Guangzhou Women and Children's Medical Center (2014073009). All studies had obtained the approval of the local hospital institutional review board, and all the participants were informed of the details of this study and signed the informed consent form.

AUTHOR CONTRIBUTIONS

HY, HL, and XG conceived and designed the study. HY, FL, KC, and YX analyzed and interpreted the data. YW, LF, and HZ provided assistance in performing the study. LP, DC, and HL revised the manuscript. HY, FL, and XG wrote the paper with feedback from all authors. All the authors read and approved the final manuscript.

FUNDING

This study was funded by the Guangdong Natural Science Fund, China (Grant numbers: 2019A1515012061 and 2021A1515011207), the Guangzhou Science and Technology Program Project, China (Grant numbers: 201904010486 and 202102010197), the Guangzhou Institute of Pediatrics/ Guangzhou Women and Children's Medical Center Fund, China (Grant numbers: GCP-2019-003, GCP-2019-006, and

REFERENCES

- Bovia, F., Fornallaz, M., Leffers, H., and Strub, K. (1995). The SRP9/14 Subunit of the Signal Recognition Particle (SRP) Is Present in More Than 20-fold Excess over SRP in Primate Cells and Exists Primarily Free but Also in Complex with Small Cytoplasmic Alu RNAs. *MBoC* 6 (4), 471–484. doi:10.1091/mbc.6.4.471
- Burns, J. C., and Glodé, M. P. (2004). Kawasaki Syndrome. Lancet 364 (9433), 533–544. doi:10.1016/s0140-6736(04)16814-1
- Chaudhary, K., Shinde, R., Liu, H., Gnana-Prakasam, J. P., Veeranan-Karmegam, R., Huang, L., et al. (2015). Amino Acid Metabolism Inhibits Antibody-Driven Kidney Injury by Inducing Autophagy. J. I. 194 (12), 5713–5724. doi:10.4049/ jimmunol.1500277
- Che, D., Pi, L., Fang, Z., Xu, Y., Cai, M., Fu, L., et al. (2018). ABCC4 Variants Modify Susceptibility to Kawasaki Disease in a Southern Chinese Population. *Dis. Markers* 2018, 8638096. doi:10.1155/2018/8638096
- Chen, Z., Zhang, J., Wei, D., Chen, J., and Yang, J. (2021). GCN2 Regulates ATF3-P38 MAPK Signaling Transduction in Pulmonary Veno-Occlusive Disease. J. Cardiovasc Pharmacol. Ther. 26 (6), 677–689. doi:10.1177/ 10742484211015535
- Consortium, G. T. (2013). The Genotype-Tissue Expression (GTEx) Project. Nat. Genet. 45 (6), 580–585. doi:10.1038/ng.2653
- Domínguez-Soto, A., de las Casas-Engel, M., Bragado, R., Medina-Echeverz, J., Aragoneses-Fenoll, L., Martín-Gayo, E., et al. (2014). Intravenous Immunoglobulin Promotes Antitumor Responses by Modulating Macrophage Polarization. J. I. 193 (10), 5181–5189. doi:10.4049/jimmunol.1303375
- Fu, L. Y., Qiu, X., Deng, Q. L., Huang, P., Pi, L., Xu, Y., et al. (2019). The IL-1B Gene Polymorphisms Rs16944 and Rs1143627 Contribute to an Increased Risk of Coronary Artery Lesions in Southern Chinese Children with Kawasaki Disease. J. Immunol. Res. 2019, 4730507. doi:10.1155/2019/4730507
- Gersony, W. M. (2009). The Adult after Kawasaki Disease. J. Am. Coll. Cardiol. 54 (21), 1921–1923. doi:10.1016/j.jacc.2009.06.057
- Gu, X., Lin, W., Xu, Y., Che, D., Tan, Y., Lu, Z., et al. (2020). The Rs1051931 G>A Polymorphism in the PLA2G7 Gene Confers Resistance to Immunoglobulin Therapy in Kawasaki Disease in a Southern Chinese Population. *Front. Pediatr.* 8, 338. doi:10.3389/fped.2020.00338
- Hadinnapola, C., Bleda, M., Haimel, M., Screaton, N., Swift, A., Dorfmüller, P., et al. (2017). Phenotypic Characterization of EIF2AK4 Mutation Carriers in a Large Cohort of Patients Diagnosed Clinically with Pulmonary Arterial Hypertension. *Circulation* 136 (21), 2022–2033. doi:10.1161/ CIRCULATIONAHA.117.028351
- Halaby, M. J., Hezaveh, K., Lamorte, S., Ciudad, M. T., Kloetgen, A., MacLeod, B. L., et al. (2019). GCN2 Drives Macrophage and MDSC Function and Immunosuppression in the Tumor Microenvironment. *Sci. Immunol.* 4 (42), 8189. doi:10.1126/sciimmunol.aax8189

YIP-2019-050), and the Postdoctoral Research Initiation Fund from Guangzhou Institute of Pediatrics, Guangzhou Women and Children's Medical Center (Grant number 3001162).

ACKNOWLEDGMENTS

The authors would like to thank the Clinical Biological Resource Bank of the Guangzhou Women and Children's Medical Center for providing all the clinical samples and greatly appreciate all the patients who donated the samples.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fgene.2022.868159/full#supplementary-material

- Harding, H. P., Novoa, I., Zhang, Y., Zeng, H., Wek, R., Schapira, M., et al. (2000). Regulated Translation Initiation Controls Stress-Induced Gene Expression in Mammalian Cells. *Mol. Cell* 6 (5), 1099–1108. doi:10.1016/s1097-2765(00) 00108-8
- Harding, H. P., Zhang, Y., Zeng, H., Novoa, I., Lu, P. D., Calfon, M., et al. (2003). An Integrated Stress Response Regulates Amino Acid Metabolism and Resistance to Oxidative Stress. *Mol. Cell* 11 (3), 619–633. doi:10.1016/s1097-2765(03)00105-9
- Hasler, J., and Strub, K. (2006). Alu RNP and Alu RNA Regulate Translation Initiation In Vitro. Nucleic Acids Res. 34 (8), 2374–2385. doi:10.1093/nar/gkl246
- Huang, Y.-H., Hsu, Y.-W., Lu, H.-F., Wong, H. S.-C., Yu, H.-R., Kuo, H.-C., et al. (2016). Interferon-gamma Genetic Polymorphism and Expression in Kawasaki Disease. *Med. Baltim.* 95 (17), e3501. doi:10.1097/md.00000000003501
- Kashef, S., Safari, M., and Amin, R. (2005). Initial Intravenous γ-Globulin Treatment Failure in Iranian Children with Kawasaki Disease. *Kaohsiung J. Med. Sci.* 21 (9), 401–404. doi:10.1016/s1607-551x(09)70141-x
- Kato, H., Koike, S., Yamamoto, M., Ito, Y., and Yano, E. (1975). Coronary Aneurysms in Infants and Young Children with Acute Febrile Mucocutaneous Lymph Node Syndrome. J. Pediatr. 86 (6), 892–898. doi:10.1016/s0022-3476(75)80220-4
- Khor, C. C., Davila, S., Davila, S., Breunis, W. B., Lee, Y.-C., Shimizu, C., et al. (2011). Genome-wide Association Study Identifies FCGR2A as a Susceptibility Locus for Kawasaki Disease. *Nat. Genet.* 43 (12), 1241–1246. doi:10.1038/ng.981
- Lee, Y.-C., Kuo, H.-C., Chang, J.-S., Chang, L.-Y., Huang, L.-M., Chen, M.-R., et al. (2012). Two New Susceptibility Loci for Kawasaki Disease Identified through Genome-wide Association Analysis. *Nat. Genet.* 44 (5), 522–525. doi:10.1038/ng. 2227
- Lin, K., Zhang, L., Wang, Y., Li, J., Xu, Y., Che, D., et al. (2021). FNDC1 Polymorphism (Rs3003174 C > T) Increased the Incidence of Coronary Artery Aneurysm in Patients with Kawasaki Disease in a Southern Chinese Population. *Jir* 14, 2633–2640. doi:10.2147/jir.s311956
- Liu, H., Huang, L., Bradley, J., Liu, K., Bardhan, K., Ron, D., et al. (2014). GCN2dependent Metabolic Stress Is Essential for Endotoxemic Cytokine Induction and Pathology. *Mol. Cell Biol.* 34 (3), 428–438. doi:10.1128/mcb.00946-13
- Longchamp, A., Mirabella, T., Arduini, A., MacArthur, M. R., Das, A., Treviño-Villarreal, J. H., et al. (2018). Amino Acid Restriction Triggers Angiogenesis via GCN2/ATF4 Regulation of VEGF and H2S Production. *Cell* 173 (1), 117–129. doi:10.1016/j.cell.2018.03.001
- Lu, Z., Xu, X., Fassett, J., Kwak, D., Liu, X., Hu, X., et al. (2014). Loss of the Eukaryotic Initiation Factor 2α Kinase General Control Nonderepressible 2 Protects Mice from Pressure Overload-Induced Congestive Heart Failure without Affecting Ventricular Hypertrophy. *Hypertension* 63 (1), 128–135. doi:10.1161/hypertensionaha.113.02313
- Lünemann, J. D., Nimmerjahn, F., and Dalakas, M. C. (2015). Intravenous Immunoglobulin in Neurology-Mode of Action and Clinical Efficacy. *Nat. Rev. Neurol.* 11 (2), 80–89. doi:10.1038/nrneurol.2014.253

- Makowsky, R., Wiener, H. W., Ptacek, T. S., Silva, M., Shendre, A., Edberg, J. C., et al. (2013). FcγR Gene Copy Number in Kawasaki Disease and Intravenous Immunoglobulin Treatment Response. *Pharmacogenet Genomics* 23 (9), 455–462. doi:10.1097/fpc.0b013e328363686e
- McCrindle, B. W., Rowley, A. H., Newburger, J. W., Burns, J. C., Bolger, A. F., Gewitz, M., et al. (2017). Diagnosis, Treatment, and Long-Term Management of Kawasaki Disease: A Scientific Statement for Health Professionals from the American Heart Association. *Circulation* 135 (17), e927–e99. doi:10.1161/CIR. 000000000000484
- McGaha, T. L., Huang, L., Lemos, H., Metz, R., Mautino, M., Prendergast, G. C., et al. (2012). Amino Acid Catabolism: a Pivotal Regulator of Innate and Adaptive Immunity. *Immunol. Rev.* 249 (1), 135–157. doi:10.1111/j.1600-065x.2012.01149.x
- Nakamura, A., Nambu, T., Ebara, S., Hasegawa, Y., Toyoshima, K., Tsuchiya, Y., et al. (2018). Inhibition of GCN2 Sensitizes ASNS-Low Cancer Cells to Asparaginase by Disrupting the Amino Acid Response. *Proc. Natl. Acad. Sci. U. S. A.* 115 (33), E7776–E85. doi:10.1073/pnas.1805523115
- Newburger, J. W., Takahashi, M., Burns, J. C., Beiser, A. S., Chung, K. J., Duffy, C. E., et al. (1986). The Treatment of Kawasaki Syndrome with Intravenous Gamma Globulin. N. Engl. J. Med. 315 (6), 341–347. doi:10.1056/ nejm198608073150601
- Newburger, J. W., Takahashi, M., Gerber, M. A., Gewitz, M. H., Tani, L. Y., Burns, J. C., et al. (2004). Diagnosis, Treatment, and Long-Term Management of Kawasaki Disease. *Circulation* 110 (17), 2747–2771. doi:10.1161/01.cir. 0000145143.19711.78
- Nossent, E. J., Antigny, F., Montani, D., Bogaard, H. J., Ghigna, M. R., Lambert, M., et al. (2018). Pulmonary Vascular Remodeling Patterns and Expression of General Control Nonderepressible 2 (GCN2) in Pulmonary Veno-Occlusive Disease. J. Heart Lung Transplant. 37 (5), 647–655. doi:10.1016/j.healun.2017.09.022
- Onouchi, Y., Gunji, T., Burns, J. C., Shimizu, C., Newburger, J. W., Yashiro, M., et al. (2008). ITPKC Functional Polymorphism Associated with Kawasaki Disease Susceptibility and Formation of Coronary Artery Aneurysms. *Nat. Genet.* 40 (1), 35–42. doi:10.1038/ng.2007.59
- Onouchi, Y., Suzuki, Y., Suzuki, H., Terai, M., Yasukawa, K., Hamada, H., et al. (2013). ITPKC and CASP3 Polymorphisms and Risks for IVIG Unresponsiveness and Coronary Artery Lesion Formation in Kawasaki Disease. *Pharmacogenomics J.* 13 (1), 52–59. doi:10.1038/tpj.2011.45
- Ravindran, R., Loebbermann, J., Nakaya, H. I., Khan, N., Ma, H., Gama, L., et al. (2016). The Amino Acid Sensor GCN2 Controls Gut Inflammation by Inhibiting Inflammasome Activation. *Nature* 531 (7595), 523–527. doi:10. 1038/nature17186
- Schmidt, S., Gay, D., Uthe, F. W., Denk, S., Paauwe, M., Matthes, N., et al. (2019). A MYC-GCN2-eIF2a Negative Feedback Loop Limits Protein Synthesis to Prevent MYC-dependent Apoptosis in Colorectal Cancer. *Nat. Cell Biol.* 21 (11), 1413–1424. doi:10.1038/s41556-019-0408-0
- Sharma, K., Vignesh, P., Srivastava, P., Sharma, J., Chaudhary, H., Mondal, S., et al. (2021). Epigenetics in Kawasaki Disease. *Front. Pediatr.* 9, 673294. doi:10.3389/ fped.2021.673294
- Takahashi, K., Oharaseki, T., Yokouchi, Y., Naoe, S., and Saji, T. (2013). Kawasaki Disease: Basic and Pathological Findings. *Clin. Exp. Nephrol.* 17 (5), 690–693. doi:10.1007/s10157-012-0734-z
- Tremoulet, A. H., Devera, G., Best, B. M., Jimenez-Fernandez, S., Sun, X., Jain, S., et al. (2011). Increased Incidence and Severity of Kawasaki Disease Among

Filipino-Americans in San Diego County. Pediatr. Infect. Dis. J. 30 (10), 909-911. doi:10.1097/inf.0b013e31821e52c6

- Uehara, R., Belay, E. D., Maddox, R. A., Holman, R. C., Nakamura, Y., Yashiro, M., et al. (2008). Analysis of Potential Risk Factors Associated with Nonresponse to Initial Intravenous Immunoglobulin Treatment Among Kawasaki Disease Patients in Japan. *Pediatr. Infect. Dis. J.* 27 (2), 155–160. doi:10.1097/inf. 0b013e31815922b5
- Wang, K., Cao, Y., Rong, Y., Ning, Q., Jia, P., Huang, Y., et al. (2019). A Novel SNP in EIF2AK4 Gene Is Associated with Thermal Tolerance Traits in Chinese Cattle. *Anim. (Basel)* 9 (6), 375. doi:10.3390/ani9060375
- Wang, Y., Wang, W., Gong, F., Fu, S., Zhang, Q., Hu, J., et al. (2013). Evaluation of Intravenous Immunoglobulin Resistance and Coronary Artery Lesions in Relation to Th1/Th2 Cytokine Profiles in Patients with Kawasaki Disease. *Arthritis & Rheumatism* 65 (3), 805–814. doi:10.1002/art.37815
- Wang, Y., Xu, Y., Huang, P., Che, D., Wang, Z., Huang, X., et al. (2021). Homozygous of MRP4 Gene Rs1751034 C Allele Is Related to Increased Risk of Intravenous Immunoglobulin Resistance in Kawasaki Disease. *Front. Genet.* 12, 510350. doi:10.3389/fgene.2021.510350
- Wang, Z., Xu, Y., Zhou, H., Wang, Y., Li, W., Lu, Z., et al. (2020). Association between P2RY12 Gene Polymorphisms and IVIG Resistance in Kawasaki Patients. *Cardiovasc Ther.* 2020, 3568608. doi:10.1155/2020/3568608
- Wek, R. C., Ramirez, M., Jackson, B. M., and Hinnebusch, A. G. (1990). Identification of Positive-Acting Domains in GCN2 Protein Kinase Required for Translational Activation of GCN4 Expression. *Mol. Cell Biol.* 10 (6), 2820–2831. doi:10.1128/mcb.10.6.2820-2831.1990
- Weng, K.-P., Hsieh, K.-S., Ho, T.-Y., Huang, S.-H., Lai, C.-R., Chiu, Y.-T., et al. (2010). IL-1B Polymorphism in Association with Initial Intravenous Immunoglobulin Treatment Failure in Taiwanese Children with Kawasaki Disease. Circ. J. 74 (3), 544–551. doi:10.1253/circj.cj-09-0664
- Wu, Z., Yu, Y., Fu, L., Mai, H., Huang, L., Che, D., et al. (2020). LncRNA SOX2OT Rs9839776 Polymorphism Reduces Sepsis Susceptibility in Southern Chinese Children. Jir 13, 1095–1101. doi:10.2147/jir.s281760
- Yang, F., Chen, X. D., Tan, L. J., Shen, J., Li, D. Y., Zhang, F., et al. (2014). Genome Wide Association Study: Searching for Genes Underlying Body Mass Index in the Chinese. *Biomed. Environ. Sci.* 27 (5), 360–370. doi:10.3967/bes2014.061

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors, and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2022 Yu, Liu, Chen, Xu, Wang, Fu, Zhou, Pi, Che, Li and Gu. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.