

Research Paper

Quantitative assessment of insertion sequence impact on bacterial genome architecture

Mark D. Adams, Brian Bishop and Meredith S. Wright

J. Craig Venter Institute, 4120 Capricorn Lane, La Jolla, CA 92037, USA

Correspondence: Mark D. Adams (madams@jcvi.org) DOI: 10.1099/mgen.0.000062

Insertion sequence (IS) elements are important mediators of genome plasticity and can lead to phenotypic changes with evolutionary significance. In multidrug-resistant *Acinetobacter baumannii* and *Klebsiella pneumoniae*, IS elements have contributed significantly to the mobilization of genes that encode resistance to antimicrobial drugs. A systematic analysis of IS elements is needed for a more comprehensive understanding of their evolutionary impact. We developed a computational approach (ISseeker) to annotate IS elements in draft genome assemblies and applied the method to analysis of IS elements in all publicly available *A. baumannii*(>1000) and *K. pneumoniae*(>800) genome sequences, in a phylogenetic context. Most IS elements in *A. baumannii*(penomes are species-specific IS*Aba* elements, whereas *K. pneumoniae*genomes contain significant numbers of both IS*Kpn* elements and elements that are found throughout the Enterobacteriaceae. *A. baumannii*genomes have a higher density of IS elements than *K. pneumoniae*, averaging ~33 vs ~27 copies per genome. In *K. pneumoniae*, several insertion sites are shared by most genomes in the ST258 clade, whereas in *A. baumannii*, different IS elements are abundant in different phylogenetic groups, even among closely related Global Clone 2 strains. IS elements differ in the distribution of insertion locations relative to genes, with some more likely to disrupt genes and others predominantly in intergenic regions. Several genes and intergenic regions had multiple independent insertion locations revealed that IS elements have been active contributors to genome- and taxon-wide characterization of insertion locations revealed that IS elements have been active contributors to genome diversity in both species.

Keywords: insertion sequence; mobile genetic element; genome-wide analysis; Acinetobacter baumannii; Klebsiella pneumoniae; antibiotic resistance.

Abbreviations: IS, insertion sequence; SNV, single nucleotide variant; MLST, multi-locus sequence type.

Data statement: All supporting data, code and protocols have been provided within the article or through supplementary data files.

Data Summary

- 1. The ISseeker software has been deposited in Github at http://github.com/JCVI-VIRIFX/ISseeker
- 2. Tables S3 and S4 provide the GenBank accession numbers to all genome sequences analysed and a URL to retrieve each sequence from the GenBank database.

Introduction

Insertion sequences (IS) are mobile genetic elements smaller than \sim 2 kbp that encode only a transposase. Once acquired, IS elements can spread in a genome by transposition, creating genetic variation and playing important roles in adaptation (Bennett, 2004; Siguier *et al.*, 2014). The density of coding content in bacterial genomes means that most random insertions occur in functional genome regions. Intragenic insertions can cause loss-of-function mutations, while intergenic insertions may disrupt promoter function or can result in up-regulation of adjacent genes in cases where the IS element encodes an outwardfacing promoter. Most insertions are presumed to be

Received 1 February 2016; Accepted 18 April 2016

© 2016 The Authors. Published by Microbiology Society

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0/).

deleterious, but some may confer a selective advantage. For example, in Acinetobacter baumannii an ISAba1 insertion upstream of the chromosomal *ampC* gene results in overexpression of the Acinetobacter-derived cephalosporinase (ADC) beta-lactamase and resistance to extended-spectrum cephalosporins (Corvec et al., 2003; Heritier et al., 2006; Turton et al., 2006). In addition to disrupting a gene and modifying gene expression, pairs of IS elements can act as a transposon, mobilizing new genetic material via lateral gene transfer such as the ISAba1-flanked bla_{OXA-23} termed Tn2006 that confers carbapenem resistance (Mugnier et al., 2009). The bla_{KPC} carbapenemase is bracketed by ISKpn7 and ISKpn6 in Tn4401a (Naas et al., 2008) and the ISAba125 element was involved in the emergence of bla_{NDM-1} (Poirel et al., 2012, 2011). Several IS elements have been reported that drive mobilization and expression of bla_{OXA-58} (Poirel & Nordmann, 2006) and bla_{RTG} (Potron et al., 2009; Bonnin et al., 2012).

Despite the importance of these elements in the evolution of antimicrobial resistance, few studies have addressed their genome-wide distribution across a diverse set of strains. Gaffé et al. (2011) found that IS elements contributed significantly to adaptive evolution of Escherichia coli under controlled growth conditions in continuous culture. They examined the distribution of eight IS elements in 120 Escherichia coli genomes following long-term growth in chemostats, and identified new IS locations that altered the global regulatory program. A study of eight clinical isolates of E. coli O157 found that IS629 and ISEc8 caused frequent small-size structural polymorphisms and suggested that IS elements may play a role in the inactivation of incoming phage and plasmids (Ooka et al., 2009). Open questions remain regarding the genome-wide impact of IS elements, the relative abundance and diversity across evolutionary lineages, and the extent to which IS elements may be reshaping the genomes of clinically important pathogens.

In draft genome assemblies, multi-copy IS elements are typically collapsed into a single contig that represents the full-length IS element sequence. Each IS copy cannot be placed in its correct genome location during assembly unless long reads, mate pairs, or some other long-range linking strategy is employed. Typically, contigs are broken at IS locations, and sequence reads that span the junction from chromosomal sequence to IS element sequence extend several bases into the IS sequence (Fig. 1a). This extension or 'stub' is often approximately half the read length when using the Velvet (Zerbino & Birney, 2008) or SPAdes (Bankevich et al., 2012) assemblers. Three software programs have been described for mapping transposable element locations: ISmapper (Hawkey et al., 2015), TIF (Nakagome et al., 2014), and breseq (Barrick et al., 2014). Each of these programs relies on primary read data rather than sequence assemblies, making them effective at defining junctions, but difficult to apply to large surveys involving hundreds of genomes given the very large input datasets. We developed the ISseeker software to identify flanks of IS elements in genome assemblies - both full

Impact Statement

Mobile genetic elements are well recognized for the role they have played in the dissemination of antimicrobial resistance genes in Gram-negative bacteria and in the rise of multi-drug resistance in several human pathogens. With large collections of genome sequences available for many bacterial species, it is now possible to quantify the abundance and distribution of these elements and assess the role they have played in genome evolution. Genome-wide surveys of the locations of insertion sequence (IS) elements in Acinetobacter baumannii and Klebsiella pneumoniae showed that several different IS elements are common within each species, and that IS elements have made significant contributions to the evolution of genome structure and variation in both species.

length copies in long contiguous sequences and stubs at contig edges – extract the flanking sequences, and align those flanks to a common reference to enable comparison of IS locations across many strains.

Thirty-six Acinetobacter baumannii species-specific ISAba elements have been registered with the ISfinder database (https://www-is.biotoul.fr/; Siguier et al., 2006). Twenty-five ISKpn elements are in the ISfinder database. Several of these elements were initially described in genome sequencing projects, while others were identified based on their participation in antibiotic resistance gene mobilization (Tables 1 and 2). Klebsiella pneumoniae strains also have elements that are commonly found throughout the Enterobacteriaceae. Other IS elements that have been described in both genomes were included in the analysis as well. ISseeker was used to define the location of IS elements in over 1000 A. baumannii genome sequences and in over 800 K. pneumoniae genomes. The resulting patterns of IS distribution show that several elements are abundant in both species and that IS elements have played a significant role in genome evolution.

Methods

ISseeker was written in perl to annotate the locations of a range of IS elements in complete and draft genome sequences. Search results are output in a text file log, a comma-separated values file and as SQL statements that can be loaded to a MySQL database to facilitate complex queries. The outline of the program is illustrated in Fig. 1. ISseeker identifies complete and partial IS matches in a query genome using blastn, with a user-specifiable percent identity threshold (default 97 %). Using contig length information, matches are classified as embedded in a contig (and either full-length or partial), consisting of an entire contig (this is common in draft assemblies), or representing the edge of the IS element matching the edge of a contig. Full-length embedded matches and valid edge matches are selected for annotation.



Fig. 1. Outline of ISseeker process. (a) Illustration of four possible alignments between an IS element and a query genome. IS element sequences are represented by blue arrows and contig sequences as a solid line. (b) Process diagram for determining IS locations based on alignment of IS flanks with a reference genome. Inputs are in blue boxes and outputs in green boxes. ISseeker program steps are shown in yellow boxes. White boxes depict intermediate results that determine next steps in program execution. Arrows depict the flow of logic in the program.

A 500 bp sequence region adjacent to the IS element is extracted from the contig and searched against the reference genome using blastn. Matches are evaluated using userdefined thresholds for percent identity (default 97 %) and length, and those passing the threshold are reported. The location relative to adjacent genes in the reference genome is reported. The program attempts to link matches into pairs representing the start and end of the IS element that map to equivalent sites in the reference genome and thus correspond to a single insertion event. In practice, this is incomplete because it appears that deletions are common near IS elements and it is not obvious whether a single event is represented. Flanks that do not match the reference are also included in the output as 'unannotated' flanks. Output is saved in a log file and as SQL statements for bulk import into a MySQL database, which facilitates complex queries. When evaluating the IS locations, we found many instances of IS sites clustered within a few bases of one another. These could represent independent insertion events or alignment artifacts. Manual review suggested that the latter was common, so for the purpose of reporting the number of distinct insertion sites, we bundled annotated locations within 10 bases of each other as a single event. Locations relative to genes were inferred based on the GenBank annotation for the reference genome, with location outside of annotated coding regions designated as intergenic and those inside coding regions designated intragenic. The ISseeker software and the MySQL schema are available at https://github.com/JCVI-VIRIFX/ ISseeker.

A user-specified reference genome is required for ISseeker analysis. The TYTH-1 genome sequence [GenBank accession no. CP003856.1 (Liou et al., 2012)] was selected as the reference A. baumannii genome after consideration of several completed genome sequences. TYTH-1 was isolated in Taiwan in 2008 and is a GC2 strain (Nemec et al., 2004), as are a majority of strains with genome sequences in the Gen-Bank database. NJST258_1 [GenBank accession no. CP006923.1 (Deleo et al., 2014)] was selected as the K. pneumoniae reference genome. NJST258_1 is a KPC-positive ST258 strain isolated in New Jersey, USA, in 2010. All completed and draft A. baumannii and K. pneumoniae genomes available in the GenBank database as of 1 August 2015 were downloaded. Genome assemblies that were highly fragmented (>300 contigs), or were assembled with newbler, or represented non-baumannii Acinetobacter strains or non-pneumoniae Klebsiella strains were excluded. 1035 complete and draft A. baumannii genomes and 807 complete and draft K. pneumoniae genomes were analysed.

All species-specific IS elements cataloged in the ISfinder database (Siguier *et al.*, 2006) were downloaded and compared against the full genome set for each species. In addition, several complete genome sequences for each species were searched against the ISfinder database by BLAST to

IS	No. of genc	omes with:			Distinct sit	es*				Total site	\$÷		Diversity	Linked genes
element	Annotated sites	Un- annotated sites	No. of sites	In genes	Between genes	% In genes	% Between genes	No. of sites	In genes	Between genes	% In genes	% Between genes	ratio‡	
IS26	776	30	265	208	57	78.5	21.5	3087	1612	1475	52.2	47.8	0.086	
ISAba1	815	20	1229	735	497	59.8	40.4	14510	7383	7127	50.9	49.1	0.085	
ISAba2	46	31	57	43	14	75.4	24.6	185	160	25	86.5	13.5	0.308	
ISAba3	4	12	14	6	5	64.3	35.7	20	13	7	65.0	35.0	0.700	OXA-58
ISAba4	0													
ISAba5	19	31	41	25	16	61.0	39.0	59	36	23	61.0	39.0	0.695	
ISA ba6	1													
ISA ba7	1													
ISAba8	1	4	4	3	1	75.0	25.0	4	33	1	75.0	25.0	1.000	
ISAba9	2		2		2	0.0	100.0	2		2	0.0	100.0	1.000	RTG-4
ISAba10	38	12	76	59	38	60.8	39.2	241	143	98	59.3	40.7	0.402	OXA-23
ISAba11	12	19	69	39	30	56.5	43.5	85	48	37	56.5	43.5	0.812	LpxA/C
ISAba12	142	122	357	209	148	58.5	41.5	066	592	398	59.8	40.2	0.361	
ISAba13	393	80	572	309	263	54.0	46.0	3639	1865	1774	51.3	48.7	0.157	
ISAba14	13	15	13	6	4	69.2	30.8	25	20	5	80.0	20.0	0.520	RTG-6
ISAba15	0													LpxD
ISAba16	140	15	258	172	86	66.7	33.3	1032	661	371	64.1	35.9	0.250	OXA-51
ISAba17	276	12	98	60	38	61.2	38.8	2007	1119	888	55.8	44.2	0.049	
ISAba18	46	25	60	46	14	76.7	23.3	199	172	27	86.4	13.6	0.302	
ISAba19	62	25	163	132	31	81.0	19.0	387	328	59	84.8	15.2	0.421	
ISAba20	3	1	5	4	1	80.0	20.0	5	4	1	80.0	20.0	1.000	
ISAba21	3	20	12	10	2	83.3	16.7	12	10	2	83.3	16.7	1.000	RTG-5
ISAba22	63	301	71	49	22	69.0	31.0	112	89	23	79.5	20.5	0.634	AadB
ISAba23	0													
ISAba24	200	1	6	7	2	77.8	22.2	240	49	191	20.4	79.6	0.038	
ISAba25	157	19	340	219	121	64.4	35.6	1391	884	507	63.6	36.4	0.244	
ISAba26	381	39	190	136	54	71.6	28.4	652	255	397	39.1	6.09	0.291	
ISAba27	46	20	343	116	227	33.8	66.2	818	258	560	31.5	68.5	0.419	
ISAba28	1	1												
ISAba29	56	25	128	102	26	79.7	20.3	308	263	45	85.4	14.6	0.416	
ISAba30	0													
ISAba31	29	51	93	34	60	36.6	64.5	122	39	83	32.0	68.0	0.762	
ISAba32	0	16												
ISAba33	29	20	64	33	31	51.6	48.4	94	41	53	43.6	56.4	0.681	

Table 1. IS Elements Surveyed in A. baumannii genomes

	JUI.													
SI	No. of gen	omes with:			Distinct sit	es*				Total site	s†		Diversity	Linked genes
element	Annotated sites	Un- annotated sites	No. of sites	In genes	Between genes	% In genes	% Between genes	No. of sites	In genes	Between genes	% In genes	% Between genes	ratio‡	
ISAba36	42	-	253	172	81	68.0	32.0	712	446	266	62.6	37.4	0.355	
ISAba125	402	160	528	338	190	64.0	36.0	1595	1054	541	66.1	33.9	0.331	OXA-58
ISAba825	3	2	9	3	33	50.0	50.0	9	3	3	50.0	50.0	1.000	OXA-58
*Distinct site †Total sites i ‡(Number of	s is the number n the number of f distinct insertic	of unique anno f annotated and m sites)/(Total i	ntated loca unannota insertions	tions on t ted sites : in all gen	the reference summed for omes).	genome. all genome	şi							

identify species non-specific elements. Eighteen additional (non-IS*Kpn*) IS elements found in *K. pneumoniae* genomes were analysed and seven additional (non-IS*Aba*) IS elements were analysed in *A. baumannii* genomes. Results are included in Tables 1 and 2 for those elements that were present in more than five genomes.

ISseeker was compared with ISmapper using a set of genomes for which both a finished genome sequence (e.g. a 'gold standard') and Illumina short reads were available. Illumina read sets were downloaded from NCBI's Sequence Read Archive (SRA) using the SRA Toolkit utility fastqdump. ISmapper was run on each read set using default parameters. Each Illumina read set was assembled using SPAdes (Bankevich et al., 2012). ISseeker was run on the finished genome sequence and on the Illumina assembly. The performance of ISseeker was evaluated by performing runs against the full set of sequences for each species using varying values for percent identity of matches to the IS element and of IS-flanking sequences to the reference, and using an alternative reference genome. It should be noted that the newbler assembler (Miller et al., 2010) suppresses these stubs so newbler assemblies cannot be used by ISseeker.

A core phylogeny based on single-nucleotide variants (SNVs; 278 322 SNVs for *A. baumannii*, 332 571 SNVs for *K. pneumoniae*) was inferred using SNVs identified by NASP (Sahl *et al.*, 2016) and constructed using FastTree 2 (Price *et al.*, 2010). Genome positions with allele calls in at least 80 % of strains were included in the analysis. Fig. 2 was prepared using the graphics tools available through the interactive Tree of Life (iTOL) web service (Letunic & Bork, 2011).

The statistical significance of comparisons of IS element composition between strain sets was assessed using Student's t-test.

Results

Description and evaluation of the ISseeker program

Four classes of IS alignment are considered by the ISseeker program (Fig. 1a): contigs that are comprised entirely of IS sequence, IS element matches that are full-length and embedded in a long genomic contig, matches to the beginning or end of an IS element at the start of end of a contig, and partial matches internal to a contig. Contig sequences flanking each IS element are extracted and compared to a reference genome (Fig. 1b). By mapping all IS/genome junctions to a single reference, it is possible to compare IS locations across strains.

The performance of ISseeker was evaluated from two perspectives: 1) comparison with ISmapper, using IS locations in completely sequenced reference genomes as a gold standard, and 2) to determine the impact of alternative run parameters on the detection of IS element locations. Two other programs that can identify IS elements in short read

IS	No. of g	enomes with:			Distinct site	es*				Total sites	+		Diversity	Linked
element	Annotated sites	Un-annotated sites	No. of sites	In genes	Between genes	% In genes	% Between genes	No. of sites	In genes	Between genes	% In genes	% Between genes	ratio#	genes
IS1294	40	16	127	67	30	76.4	23.6	325	227	86	69.8	30.2	0.391	
ISIF	452	49	87	52	35	59.8	40.2	1030	75	955	7.3	92.7	0.084	
IS26	494	69	211	160	51	75.9	24.2	2064	1577	487	76.4	23.6	0.102	
IS4321R	159	33	21	13	8	61.9	38.1	376	141	235	37.5	62.5	0.056	
IS5	19	23	44	22	22	50.0	50.0	53	26	27	49.1	50.9	0.830	
IS5075	318	26	36	22	14	61.1	38.9	883	292	591	33.1	6.99	0.041	
IS6100	430	30	15	11	4	73.3	26.7	937	140	797	14.9	85.1	0.016	
IS903B	491	73	194	119	75	61.3	38.7	1403	467	936	33.3	66.7	0.138	
ISEcp1B	06	16	51	35	16	68.6	31.4	136	66	37	72.8	27.2	0.375	
ISKpn1	583	4	146	27	119	18.5	81.5	2967	57	2910	1.9	98.1	0.049	
ISKpn2	0	35												
ISKpn3	0													
ISKpn4	0													
ISKpn6	413	1	3		3	0.0	100.0	823		823	0.0	100.0	0.004	KPC
ISKpn7	414	1	3		3	0.0	100.0	824		824	0.0	100.0	0.004	KPC
ISKpn8	24	5	8	1	7	12.5	87.5	32	3	29	9.4	90.6	0.250	
ISKpn9	0													
ISKpn10	0													
ISKpn11	0	6												
ISKpn12	0	12												
ISKpn13	0													
ISKpn14	64	142	58	33	25	56.9	43.1	98	51	47	52.0	48.0	0.592	
ISKpn15	0													
ISKpn18	387	1	85	76	6	89.4	10.6	1188	1153	35	97.1	2.9	0.072	
ISKpn19	12	16	11	6	2	81.8	18.2	12	10	2	83.3	16.7	0.917	
ISKpn20	9	Э												
ISKpn21	47	36	19	6	10	47.4	52.6	80	43	37	53.8	46.3	0.238	
ISKpn23	0													
ISKpn24	235	19	29	20	6	69.0	31.0	458	37	421	8.1	91.9	0.063	
ISKpn25	71	76	13	9	7	46.2	53.8	76	9	70	7.9	92.1	0.171	
ISKpn26	486	33	621	379	242	61.0	39.0	4238	1627	2611	38.4	61.6	0.147	
ISKpn27	0													
ISKpn28	452	25	49	20	29	40.8	59.2	855	166	689	19.4	80.6	0.057	
ISKpn31	0	10												
*Distinct s.	ites is the num	ber of unique ann	iotated loca	ntions on t	he reference §	genome.								
†Total site +(Number	s in the numbe: مf distinct inse	er of annotated an	d unannot: ' insertions	ated sites { in all genu	summed for a	ll genomes.								
12 11 11 1 1 + 1 +	OI MIDINI INTINO	TILUUT STUDY / / CONTO TILUUT		111 an 5~11	UIIIVoj.									

Microbial Genomics

Table 2. IS Elements Surveyed in K. pneumoniae genomes

data were not included in the evaluation because they are not strictly comparable. Breseq was designed for mutation-finding in long-term culture experiments and is best suited to comparing very closely related genomes to a sequenced reference. TIF uses the unix grep command to identify IS-matching reads and is thus unable to identify non-exact matches.

Results from ISseeker and ISmapper were compared on four K. pneumoniae genomes and four A. baumannii genomes for which Illumina reads, Illumina assemblies, and finished genome sequences were available (Table S1, available in the online Supplementary Material). Across these eight genomes, there were 74 insertion sites for the test IS elements ISAba1 or ISKpn26. ISseeker found all 74 sites when run using both the finished sequences and the draft genome assemblies, while ISmapper missed 20 sites for a sensitivity of 73 %. This is lower than the value reported by Hawkey et al. (2015). Further analysis showed that most missed sites were in genomes with low read coverage (<80x) or at locations with structural variation relative to the reference. ISseeker reports every IS-flanking sequence, including locations that cannot be annotated in the reference genome and those that are not in valid pairs matching both the IS element beginning and end sequences at a common reference location. ISmapper is more conservative in reporting only valid IS edge pairs in the primary output, with some additional information in ancillary output files. One interesting case identified by ISseeker, but not ISmapper, involved the creation and mobilization of a compound transposon comprised of inverted repeat copies of ISAba1 in the ORAB01 genome (Fig. S1). ISseeker recognized that there were two copies and the correct location of both, but the details of the structure were only apparent in the finished genome sequence.

The sensitivity to alteration in run parameters was evaluated for an abundant IS element in each species - ISAba1 and ISKpn26 - across the full set of assemblies for each species (Table S2). Reduction of the minimum percent identity of the matches (IS edge detection and flank alignment to the reference) from the default of 97 % to 95 % resulted in annotation of 3–4 % more sites. Upon manual review, some of these were determined to be spurious, so the more conservative threshold was retained. Increasing the stringency of the flank alignment to require a full 500 bp match reduced the number of annotated sites by 13 % (*A. baumannii*) and 8 % (*K. pneumoniae*). Use of an alternative *A. baumannii* reference -a GC1 strain rather than a GC2 strain - also resulted in a loss of about 7 % of the annotated sites.

Analysis of IS elements in *A. baumannii* and *K. pneumoniae*

Each ISAba and ISKpn element was compared against the corresponding full set of complete and draft genomes (Tables 1, 2, S3 and S4). Several non-species-specific IS elements were also included, based on elements present in a sampling of genomes from each species. 89 % of A. baumannii genomes and 94 % of K. pneumoniae genomes had

at least one IS element detected and several elements were detected in hundreds of genomes (Fig. 2). The overall numbers of insertions and chromosomal locations of IS elements were greater in *A. baumannii* than in *K. pneumoniae*. In *K. pneumoniae*, 18 869 total IS copies were found across 782 genomes. In *A. baumannii*, 32 539 copies were found across 976 genomes. On average, *A. baumannii* genomes contained 33 copies of IS elements, while *K. pneumoniae* genomes contained 27 copies (*p*<0.001).

A strong pattern of similar IS content among phylogenetically related strains is apparent, suggesting that many insertions are conserved. The number of genomes that share a set of IS insertion locations for the most abundant elements is shown in Fig. 3. ISAba1 and ISKpn26 are the only elements with >10 shared sites in a substantial number of strains. Four additional IS elements have >10 copies per genome in some *A. baumannii* strains. IS26 and ISKpn1 have 2 and 6 copies in shared locations per genome, respectively, in 300 strains, corresponding to ST258 strains.

There are more distinct IS element insertions in the examined A. baumannii genomes compared to K. pneumoniae. With respect to distinct sites mapped to each reference genome (TYTH-1 for A. baumannii and NJST258_1 for K. pneumoniae), there were 1843 distinct K. pneumoniae genome locations with IS insertions and 5341 distinct A. baumannii locations. These distinct insertion sites represent the minimum number of insertion events that occurred over time because some insertion sites could not be mapped to the selected reference genomes, and because multiple independent insertions could have occurred at a given site. Twelve different IS elements have over 100 distinct insertion sites across the A. baumannii strain set, but only five IS elements have that many distinct insertion sites in K. pneumoniae genomes. There are many more IS insertion sites shared by up to 100 A. baumannii genomes than there are shared sites across similar numbers of K. pneumoniae genomes (Table 3). In contrast, K. pneumoniae genomes have more sites shared in >250 genomes reflecting the relatively homogenous IS patterns in the dominant ST258 clade in the dataset. In addition, there are many more strain-specific insertion events in A. baumannii (3194 vs 1234). Another view of the extent of shared insertion sites is given in Fig. S2 that depicts the number of genomes that share sites along the A. baumannii or K. pneumoniae reference chromosome. There are many more moderately abundant shared sites among A. baumannii strains than K. pneumoniae strains.

A majority of genomes in the datasets for both species belong to multidrug-resistant clonal groups that have recently expanded: 62 % of *A. baumannii* genomes belong to GC2 [i.e. multi-locus sequence type (MLST) ST2] and 53 % of *K. pneumoniae* genomes belong to MLST ST258. In these subsets of strains, IS elements are also much more frequent and their locations are more diverse in *A. baumannii* compared to *K. pneumoniae* (see Fig. 2). In *K. pneumoniae*, the common location of IS insertion sites shared by large numbers of ST258 strains regardless of geographic origin



Fig. 2. IS representation in a phylogenetic context. The most abundant IS elements (present in >100 genomes) are shown in the context of the *A. baumannii* (a) and *K. pneumoniae* (b) phylogeny based on SNP markers. Isolation locations for the strains from the largest collections have a color code in the inner circle. The height of each bar represents the number of copies of each element in each genome. Scale rings illustrate the height of the histograms on each tree diagram. In (a), strain groups are denoted with coloured branches: Global Clone 1 (pink), GC2 (green) and ST79 (orange). In (b), the two major sub-groups of ST258 are denoted as ST258A (orange) and ST258B (blue).

supports a model of recent expansion with limited strainor clade-specific IS gain or loss. Variability in IS element composition and abundance is higher in the *A. baumannii* GC2 set, with more clustering of IS patterns by geographic location and phylogenetic position. It has been hypothesized that the ST258 lineage of *K. pneumoniae* arose around 1995 (Bowers *et al.*, 2015), whereas the oldest known MDR GC2 strain of *A. baumannii* was isolated in 1982 (Diancourt *et al.*, 2010; Blackwell *et al.*, 2015).

A. baumannii IS elements

ISAba1 has had the largest impact on *A. baumannii* genomes, with copies detected in 815 of the *A. baumannii* genome assemblies and over 14 500 total insertions mapped in those strains. An ISAba1 insertion site is present upstream of the bla_{ADC} (*ampC*) gene in most of the genomes that have copies of this element (736 genomes). The second most common insertion site for ISAba1 is upstream of the other chromosomal β -lactamase gene, $bla_{OXA-51-like}$ (369 genomes). This insertion results in over-expression of the OXA-51-like carbapenemase and resistance to imipenem and meropenem (Nemec *et al.*, 2008). The median number of ISAba1 sites per genome was 19 and the maximum number in a single genome was 34.

Four other elements (ISAba125, ISAba13, ISAba26 and IS26) were present in over 350 strains each, and five additional elements were present in more than 100 strains (Table 1). In a few cases, it seems that an IS element has run amok in a genome, such as the ISAba6 and ISAba7 elements in *A. baumannii* strain SDF (Vallenet *et al.*, 2008). Most ST79 strains have 50–100 copies of ISAba27. Seven genomes have copies of 10 or more different IS elements and five genomes have more than 100 total IS copies.

ISAba4, ISAba15, ISAba23, ISAba30 and ISAba32 were not found in any of the sequenced genomes. ISAba6 and ISAba7 were found only in the SDF genome (Vallenet et al., 2008). ISAba8 and ISAba28 were also only found in one genome each. ISAba2, ISAba18, ISAba19 and ISAba29 are IS3-family elements and are 85-95 % identical to one another, making inference of their abundance and correct locations difficult in draft genomes. Likewise, ISAba16 and ISAba25 are 97 % identical to one another and many of their annotated sites overlap with one another and are thus ambiguous as to the specific element that is present at each location. ISAba12 and ISAba13 are 84.8 % identical, including regions of 100 % identity in the first 23 bases and last 21 bases, and are also difficult to discriminate in draft genomes. Of the non-ISAba elements examined, only IS26 was abundant enough to be included.

Genomes that are closely related to each other on the phylogenetic tree tended to have similar patterns of IS element composition (Fig. 2). There are a few large strain collections representing restricted geographic regions among the 1035 genomes, including 442 isolates from Maryland (NCBI Bio-Project PRJNA224116) and 174 from Ohio (Wright *et al.*, 2014, 2016). Many of these genomes are very similar, potentially representing clonal series, but differences in IS content are apparent within each group. Among the GC2 genomes, there are several interesting phylogenetic clusters, some of which correspond to geographically restricted strain collections. For example, some clusters of strains isolated in Maryland have copies of ISAba16/ISAba25 that are largely absent from other strains. Strains previously identified as Clade D (Wright *et al.*, 2014, 2016) are clearly distinct from other GC2 strains by having 7–31 copies of ISAba12. Most Ohio strains have one or two copies of ISAba22, ISAba24, and ISAba26 that are found in relatively few other strains. Most Maryland strains have 7–10 copies of ISAba13 and of ISAba17, elements that are not as abundant in other branches of the tree.

K. pneumoniae IS elements

Overall, there are fewer IS element copies in *K. pneumoniae* genomes than in *A. baumannii*. This difference is reflected across both the number of distinct insertion sites (reflecting historical independent insertion events) and in the total number of copies across the genome set (reflecting the success of strains carrying those elements) (Fig. 2, Tables 1 and 2). In *K. pneumoniae*, about 350 ST258 genomes share IS insertion locations for IS*Kpn*1, IS*Kpn*26 and IS*1F*. This suggests that ST258 genomes have spread rapidly worldwide with a reasonably stable repertoire of IS elements and only limited new IS mobilization activity.

Thirteen IS elements were present in >100 K. pneumoniae genomes (Table 2, Fig. 2). ISKpn6 and ISKpn7 are present on Tn4401 that carries the $bla_{\rm KPC}$ gene; the presence of those two elements corresponded closely with the presence of the $bla_{\rm KPC}$ gene. Both of those elements were only observed in the Tn4401 context and so appear not to have mobilized to other sites in the K. pneumoniae genomes. ISKpn24 is also present on the pNJST258N2 plasmid that carries Tn4401,and copies in most ST258 genomes map to that plasmid. Unlike ISKpn6 and ISKpn7, ISKpn24 was observed at several other sites in a subset of genomes.

Eight additional IS elements were found in more than 200 genomes (ISKpn1, ISKpn18, ISKpn26, ISKpn28, IS26, IS903B, IS1F, and IS6100). Of these, ISKpn18 is almost entirely restricted to ST258 strains, but the other IS elements are found throughout the phylogenetic tree. ISKpn1 was the most broadly distributed, appearing in 583 genomes. Most ST258 genomes share five common ISKpn1 insertion sites. The number of copies of ISKpn26 is more variable across genomes, indicating more active mobilization than other elements. NJST258_1 has seven chromosomal copies of ISKpn26 and one on pNJST258N1. Copies at equivalent positions are present in other ST258 strains. ISKpn28 mapped to another large plasmid (pNJST258N1) in the reference genome, and one copy was present in the chromosome in most ST258 strains.

ST258 genomes have more than three times as many IS copies than non-ST258 genomes (average 34.5 vs 10; p<0.01). Unlike in *A. baumannii*, there are no large expansions in IS



Fig. 3. Distribution of conserved IS clusters for the most common IS elements. The number of strains sharing a set of IS element locations is plotted for the five IS elements with the largest number of copies (>1500 total copies for each element) in *A. baumannii* and for IS*Aba*27 that is greatly expanded in certain genomes (a), and for six *K. pneumoniae* IS elements with the largest number of copies (>1000 total copies) (b).

copy number in any of the *K. pneumoniae* genomes: the maximum number of copies of a single element was 22 copies of IS*Kpn*26 and only 18 genomes have \geq 50 total IS insertions, compared to 189 *A. baumannii* genomes with \geq 50 elements. Eight IS*Kpn* elements were not found in any sequenced genome.

Distribution relative to genes

The examined IS elements vary in their insertion locations relative to coding regions. By definition, an element may insert within a gene or between genes. Intragenic insertions have the potential to act as gene knockouts. Intergenic insertions may have no effect on adjacent genes or could either positively or negatively affect expression, depending on the precise location relative to promoters. ISAba1 and ISAba125 have strong outward-facing promoters and can up-regulate the expression of adjacent genes (Lopes &

10

Amyes, 2012); other elements have not been characterized for promoter activity. We considered the fraction of intragenic insertions for each element from two perspectives the total number of sites across all genomes, and the number of distinct sites (Tables 1 and 2). The former measure incorporates the abundance (number of genomes carrying each insertion) while the latter more accurately reflects the number of insertion events and is not biased by repeated sampling of closely related genomes. The two measures are closely correlated for IS elements with more than 15 distinct insertion sites. In A. baumannii, the proportion of intragenic insertions varies from approximately 30 % (ISAba31 and ISAba27) to over 70 % (ISAba19 and ISAba22). In K. pneumoniae, the intragenic proportion ranges less than 20 % (ISKpn1) to over 80 % (ISKpn18). A low proportion of intragenic insertions could be due to the fact that gene-disrupting insertions are more likely to be selected against than

Number of genomes sharing a site	Total	copies	Distin	ict sites
	A. baumannii	K. pneumoniae	A. baumannii	K. pneumoniae
>500	736	0	1	0
250-499	6975	12789	25	36
150–249	3794	3385	18	18
100–149	2561	1136	21	9
50–99	1559	911	21	14
20–49	4985	950	172	32
10–19	2642	612	195	42
2–9	5590	1634	1694	471
1	3194	1234	3194	1234

Table 3. Shared IS element sites

intergenic insertions. Alternatively, there may have been strong positive selection for certain intergenic events that has resulted in their high frequency.

Another indirect measure of the potential adaptive effects of IS insertion is the diversity of sites, which we calculated as the 'diversity ratio': the number of distinct sites divided by the total number of observed insertions in all genomes for each IS element. A high ratio means that most insertions are strain-specific, while a low ratio means that a few IS locations are shared by most strains carrying that element, with few additional strain-specific insertions. The latter group is more likely to represent positively selected insertions. In A. baumannii, the diversity ratio ranged from <10 % (ISAba1, ISAba17, IS26) to >75 % (ISAba11, ISAba31) (Table 1). As an example of a low diversity ratio, 190 of the 200 genomes that contain ISAba24 have an insertion between the genes encoding hypothetical proteins M3Q_2649 and M3Q_2651 in TYTH-1. On the other hand, among the 29 genomes with ISAba31, 72 of the 93 insertion sites are strain-specific. In K. pneumoniae, the abundant IS elements have diversity ratios less than 0.4, except ISKpn14 and IS5.

Multiple independent insertions by the same or different IS elements in the same genomic region may also indicate that those insertions convey a selective advantage. In K. pneumoniae, there are fewer than two dozen genes or intergeneic regions with insertions by more than two different IS elements. In A. baumannii, however, there are 320 genes with three or more different IS elements inserted in them across the strain set (Table S5). An additional 185 intergenic locations have three or more different IS elements (Table S6). Among these, there is a strong bias for insertions between genes that are oriented so as to be up-regulated by an ISencoded promoter. Only 18 (10 %) of the intergenic insertions are between genes oriented toward the IS insertion site; the remaining 89 % of insertion sites, a gene is oriented so as to be up-regulated by the adjacent IS element. One genome segment with multiple insertions is the four-gene region M3Q_2685-M3Q_2688 encoding the type I pilus proteins CsuA/B and their regulators, which has dozens of independent insertions by thirteen different IS elements. Twelve different IS elements were found in the 176 bp region between M3Q_2382 and M3Q_2383 in 389 strains. The repeated insertions at this locus suggest that these genes may encode important functions, although each encodes a hypothetical protein with no functionally characterized domains.

Discussion

After correcting for the larger number of *A. baumannii* genomes in the analysis, there were about 40 % more total insertions observed in *A. baumannii* genomes than in *K. pneumoniae* genomes, and more than twice as many distinct insertion sites. Considering that *A. baumannii* genomes (~4 Mbp) are about 30 % smaller than *K. pneumoniae* genomes (~5.6 Mbp), the IS element density is even greater, with about one IS element every ~109 kbp in *A. baumannii*, compared with every ~185 kbp in *K. pneumoniae*. As can be seen in Table 3, there are many more IS locations that are shared in up to 10 % of the *A. baumannii* strains, while *K. pneumoniae* genomes have more sites that are shared by about half of the genomes, reflecting the large proportion of very similar ST258 strains in the dataset and the greater diversity of IS location patterns among the *A. baumannii* genomes.

There are several potential explanations for the more diverse set of IS locations in A. baumannii than in K. pneumoniae. The most straightforward may be that the A. baumannii strains represent a more diverse evolutionary history than the K. pneumoniae strains, and thus more time for IS elements to move and accumulate. There are more genomes on long branches of the A. baumannii phylogenetic tree than the K. pneumoniae tree. Although difficult to discern in Fig. 2, this is also true for the most abundant MLST groups: the sum of the branch lengths of GC2 A. baumannii strains is about four times longer than the sum of the ST258 K. pneumoniae strains. However, in both species, IS elements are more abundant in the recently emerged lineages than in the more diverse strains, so divergence time alone cannot explain the abundance differences. It appears that strong selection for a founder ST258 strain carrying the *bla*_{KPC} gene

resulted in a rapid expansion of the this lineage (Bowers *et al.*, 2015) that contains a shared set of IS insertion sites that were present in the founder. Only the IS*Aba*1 site upstream of the bla_{ADC} gene is common to most GC2 strains, and it has been argued that this is due to multiple independent insertions, rather than a single shared ancestor (Hamidian & Hall, 2013).

Hawkey *et al.* (2015) have described a program that uses primary reads to identify the locations of insertion sites in *A. baumannii*. Their approach has some advantages over ours: by relying on primary sequence reads rather than assemblies, the impact of variation in sequencing and assembly methods and efficacy are reduced. However, publicly available read sets are much more difficult to work with than assemblies, requiring generally 50–150 times as much disk space and computing time during analysis. For example, it takes 20– 120 min to download the reads from a single genome from SRA using the SRA Toolkit program fastq-dump. In the same length of time, contig sequences can be downloaded from 1000 genomes from the WGS division of GenBank and searched for IS content using ISseeker.

Other limitations to mapping IS locations in draft genomes include sequence variation in the IS element, differences between the query genome and the reference, and assembly artifacts that tend to occur near repetitive genome regions. Use of alternative run parameters resulted in differences in detection rate (Table S2), and suggest that for any particular insertion site, additional computational analysis may be needed to determine the insertion status in strains of interest.

Acknowledgements

M. D. A. is supported by grants from the National Institute of General Medical Sciences of the National Institutes of Health (NIH) through award number R01GM094403 and from the National Institute of Allergy and Infectious Diseases of the NIH through award number U19AI110819. The funders had no role in study design, data collection and interpretation, or the decision to submit the work for publication.

References

Bankevich, A., Nurk, S., Antipov, D., Gurevich, A. A., Dvorkin, M., Kulikov, A. S., Lesin, V. M., Nikolenko, S. I., Pham, S.& other authors (2012). SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19, 455–477.

Barrick, J. E., Colburn, G., Deatherage, D. E., Traverse, C. C., Strand, M. D., Borges, J. J., Knoester, D. B., Reba, A. & Meyer, A. G. (2014). Identifying structural variation in haploid microbial genomes from short-read resequencing data using breseq. *BMC Genomics* 15, 1039.

Bennett, P. M. (2004). Genome plasticity: insertion sequence elements, transposons and integrons, and DNA rearrangement. *Methods Mol Biol* **266**, 71–113.

Blackwell, G. A., Nigro, S. J. & Hall, R. M. (2015). Evolution of AbGRI2-0, the progenitor of the AbGRI2 resistance island in Global Clone 2 of *Acinetobacter baumannii*. *Antimicrob Agents Chemother* **60**, 1421–1429.

Bonnin, R. A., Poirel, L. & Nordmann, P. (2012). A novel and hybrid composite transposon at the origin of acquisition of bla(RTG-5) in *Acinetobacter baumannii. Int J Antimicrob Agents* 40, 257–259.

Bowers, J. R., Kitchel, B., Driebe, E. M., MacCannell, D. R., Roe, C., Lemmer, D., de Man, T., Rasheed, J. K., Engelthaler, D. M. & other authors (2015). Genomic analysis of the emergence and rapid global dissemination of the clonal group 258 *Klebsiella pneumoniae* pandemic. *PLoS One* 10, e0133727.

Corvec, S., Caroff, N., Espaze, E., Giraudeau, C., Drugeon, H. & Reynaud, A. (2003). AmpC cephalosporinase hyperproduction in *Acinetobacter baumannii* clinical strains. *J Antimicrob Chemother* 52, 629–635.

Deleo, F. R., Chen, L., Porcella, S. F., Martens, C. A., Kobayashi, S. D., Porter, A. R., Chavda, K. D., Jacobs, M. R., Mathema, B. & other authors (2014). Molecular dissection of the evolution of carbapenem-resistant multilocus sequence type 258 *Klebsiella pneumoniae*. *Proc Natl Acad Sci U S A* 111, 4988–4993.

Diancourt, L., Passet, V., Nemec, A., Dijkshoorn, L. & Brisse, S. (2010). The population structure of *Acinetobacter baumannii*: expanding multiresistant clones from an ancestral susceptible genetic pool. *PLoS One* **5**, e10034.

Gaffé, J., McKenzie, C., Maharjan, R. P., Coursange, E., Ferenci, T. & Schneider, D. (2011). Insertion sequence-driven evolution of *Escherichia coli* in chemostats. *J Mol Evol* 72, 398–412.

Hamidian, M. & Hall, R. M. (2013). ISAba1 targets a specific position upstream of the intrinsic *ampC* gene of *Acinetobacter baumannii* leading to cephalosporin resistance. *J Antimicrob Chemother* 68, 2682–2683.

Hawkey, J., Hamidian, M., Wick, R. R., Edwards, D. J., Billman-Jacobe, H., Hall, R. M. & Holt, K. E. (2015). ISMapper: identifying transposase insertion sites in bacterial genomes from short read sequence data. *BMC Genomics* 16, 667.

Héritier, C., Poirel, L. & Nordmann, P. (2006). Cephalosporinase overexpression resulting from insertion of ISAba1 in Acinetobacter baumannii. Clin Microbiol Infect 12, 123–130.

Lemmer, D., Travis, J., Schupp, J., Gillece, J., Aziz, M., Driebe, E. M., Drees, K., Hicks, N. & Williamson, C. (2016). The Northern Arizona SNP Pipeline (NASP): accurate, flexible, and rapid identification of SNPs in WGS datasets. *bioRxiv*.

Letunic, I. & Bork, P. (2011). Interactive Tree of Life v2: online annotation and display of phylogenetic trees made easy. *Nucleic Acids Res* 39, 475–478.

Liou, M. L., Liu, C. C., Lu, C. W., Hsieh, M. F., Chang, K. C., Kuo, H. Y., Lee, C. C., Chang, C. T., Yang, C. Y. & Yang, C. Y. (2012). Genome sequence of *Acinetobacter baumannii* TYTH-1. *J Bacteriol* 194, 6974.

Lopes, B. S. & Amyes, S. G. (2012). Role of ISAba1 and ISAba125 in governing the expression of blaADC in clinically relevant Acinetobacter baumannii strains resistant to cephalosporins. J Med Microbiol 61, 1103–1108.

Miller, J. R., Koren, S. & Sutton, G. (2010). Assembly algorithms for next-generation sequencing data. *Genomics* 95, 315–327.

Mugnier, P. D., Poirel, L. & Nordmann, P. (2009). Functional analysis of insertion sequence ISAba1, responsible for genomic plasticity of Acinetobacter baumannii. J Bacteriol 191, 2414–2418.

Naas, T., Cuzon, G., Villegas, M. V., Lartigue, M. F., Quinn, J. P. & Nordmann, P. (2008). Genetic structures at the origin of acquisition of the beta-lactamase *bla*KPC gene. *Antimicrob Agents Chemother* 52, 1257–1263.

Nakagome, M., Solovieva, E., Takahashi, A., Yasue, H., Hirochika, H. & Miyao, A. (2014). Transposon insertion finder (TIF): a novel program for detection of de novo transpositions of transposable elements. *BMC Bioinformatics* 15, 71.

Nemec, A., Dijkshoorn, L. & van der Reijden, T. J. (2004). Long-term predominance of two pan-European clones among multi-resistant *Acinetobacter baumannii* strains in the Czech Republic. *J Med Microbiol* **53**, 147–153.

Nemec, A., Krízová, L., Maixnerová, M., Diancourt, L., van der Reijden, T. J., Brisse, S., van den Broek, P. & Dijkshoorn, L. (2008). Emergence of carbapenem resistance in *Acinetobacter baumannii* in the Czech Republic is associated with the spread of multidrug-resistant strains of European clone II. *J Antimicrob Chemother* 62, 484– 489.

Ooka, T., Ogura, Y., Asadulghani, M., Ohnishi, M., Nakayama, K., Terajima, J., Watanabe, H. & Hayashi, T. (2009). Inference of the impact of insertion sequence (IS) elements on bacterial genome diversification through analysis of small-size structural polymorphisms in *Escherichia coli* O157 genomes. *Genome Res* **19**, 1809– 1816.

Poirel, L. & Nordmann, P. (2006). Genetic structures at the origin of acquisition and expression of the carbapenem-hydrolyzing oxacillinase gene *bla*Oxa-58 in *Acinetobacter baumannii. Antimicrob Agents Chemother* **50**, 1442–1448.

Poirel, L., Bonnin, R. A. & Nordmann, P. (2011). Analysis of the resistome of a multidrug-resistant Ndm-1-producing *Escherichia coli* strain by high-throughput genome sequencing. *Antimicrob Agents Chemother (Bethesda)* 55, 4224–4229.

Poirel, L., Bonnin, R. A., Boulanger, A., Schrenzel, J., Kaase, M. & Nordmann, P. (2012). Tn125-related acquisition of blaNdm-like genes in *Acinetobacter baumannii. Antimicrob Agents Chemother* 56, 1087–1089.

Potron, A., Poirel, L., Croize, J., Chanteperdrix, V. & Nordmann, P. (2009). Genetic and biochemical characterization of the first extended-spectrum Carb-type ss-lactamase, RTG-4, from *Acineto-bacter baumannii*. *Antimicrob Agents Chemother* **53**, 3010–3016.

Price, M. N., Dehal, P. S. & Arkin, A. P. (2010). FastTree 2–approximately maximum-likelihood trees for large alignments. *PLoS One* 5, e9490.

Siguier, P., Gourbeyre, E. & Chandler, M. (2014). Bacterial insertion sequences: their genomic impact and diversity. *FEMS Microbiol Rev* 38, 865–891.

Siguier, P., Perochn, J., Lestrade, L., Mahillon, J. & Chandler, M. (2006). ISfinder: the reference centre for bacterial insertion sequences. *Nucleic Acids Res* 34, 32–36.

Turton, J. F., Ward, M. E., Woodford, N., Kaufmann, M. E., Pike, R., Livermore, D. M. & Pitt, T. L. (2006). The role of ISAba1 in expression of OXA carbapenemase genes in *Acinetobacter baumannii*. *FEMS Microbiol Lett* 258, 72–77.

Vallenet, D., Nordmann, P., Barbe, V., Poirel, L., Mangenot, S., Bataille, E., Dossat, C., Gas, S., Kreimeyer, A. & other authors (2008). Comparative analysis of Acinetobacters: three genomes for three lifestyles. *PLoS One* **3**, e1805.

Wright, M. S., Haft, D. H., Harkins, D. M., Perez, F., Hujer, K. M., Bajaksouzian, S., Benard, M. F., Jacobs, M. R. & Bonomo, R. A. (2014). New insights into dissemination and variation of the health care-associated pathogen *Acinetobacter baumannii* from genomic analysis. *MBio* 5, e00963–13.

Wright, M. S., Iovleva, A., Jacobs, M. R., Bonomo, R. A. & Adams, M. D. (2016). Genome dynamics of multidrug-resistant Acinetobacter baumannii during infection and treatment. *Genome Med* **8**, 26.

Zerbino, D. R. & Birney, E. (2008). Velvet: algorithms for de novo short read assembly using de Bruijn graphs. *Genome Res* 18, 821–829.

Data Bibliography

1. Adams, M.D., Bishop, B., Wright, M.S. The IS seeker software http://github.com/Jcvi-Virifx/ISseeker (2016).